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2 **A meta-analysis on the benefits and costs of hosting secondary endosymbionts in sap-sucking**
3 **insects**

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9

10 **Abstract**

11 Herbivorous insects host various bacteria that help them to feed, grow, and survive. Sap-sucking insects,
12 in particular, feed on a nutrient-poor resource and have evolved obligate symbioses with nutritional
13 bacteria for survival. Additionally, sap-sucking insects have formed facultative associations with bacterial
14 symbionts that are not essential for growth and survival but assumed to confer some benefits, such as
15 resistance to natural enemies. Several recent reviews have highlighted the importance of these
16 symbionts in understanding their hosts' biology, but currently there is a lack of a quantitative and
17 systematic analysis of the published evidences exploring whether the different endosymbionts are
18 actually beneficial or not. In this meta-analysis we explored the potential costs and benefits associated
19 with hosting facultative endosymbionts in sap-sucking insects. Our first result is that most of the empirical
20 experimental data information is limited to a few species of aphid and one species of whiteflies. Through
21 the meta-analysis we showed that hosting symbionts generally leads to costs through increased
22 development time, reduced longevity, and reduced fecundity, and benefits via increased resistance to
23 parasitic wasps in sap-sucking insects. However, the impact of these costs and benefits was strongly
24 insect and symbiont species dependent. Many of the insects studied are agricultural pests, and
25 understanding the impact of bacterial symbionts on their hosts across different environments can benefit
26 sustainable management of greenhouses and agricultural land.

27

28

29 Introduction

30 Insect associations with mutualistic microbes are widespread, and it is nowadays widely accepted that
31 these symbionts, and bacteria in particular, play key roles in the biology of their hosts (Brownlie &
32 Johnson 2009; Feldhaar 2011). Thanks to important innovations in molecular techniques, the last two
33 decades have provided deep insights into these diverse and often intricate interactions. The guts of most
34 insects that feed on plant leaves, for instance, are colonised by complex communities of bacteria and
35 fungi (Dillon & Dillon 2004). The role played by most of these microbes has yet to be understood, but
36 evidence suggests that they are an important component of the hosts immune system, and that they
37 often assist in the digestive process. Relative to leaf-feeding insects, the nutritional services provided by
38 bacterial symbionts in sap-sucking insects are better understood (Moran, McCutcheon & Nakabachi
39 2008). This lifestyle has evolved multiple times among Hemiptera and includes most Sternorrhyncha
40 (whiteflies, mealybugs, aphids and psyllids), many Auchenorrhyncha (planthoppers and leafhoppers) and
41 most herbivorous Heteroptera (lygaeids, pentatomids, and coreids) (Dolling 1991).

42 Hemipterans are the most diverse group of hemimetabolous insects, with more than 100,000 species
43 described (Stork 2018) and a large majority that have adopted sap-sucking life histories. Insects
44 belonging to this guild feed on impoverished diets so that they all rely on mutualistic bacteria that live
45 within or inside insect cells (they are termed endosymbionts) to synthesise essential nutrients that the
46 insect cannot acquire directly from the diet (Douglas 1998). These primary, or obligate, symbionts are
47 more like an organelle than an independent organism and are found in almost all aphids (they carry
48 *Buchnera aphidicola*), psyllids (*Carsonella ruddii*), whiteflies (*Portiera aleyrodidarum*) and mealybugs
49 (*Tremblaya princeps*), among others (Moran, McCutcheon & Nakabachi 2008). Such endosymbionts are
50 required for insect survival and reproduction, transmitted from mother to offspring with high fidelity, and
51 thought to be at the core of the diversification process of their insect hosts (Moran, McCutcheon &
52 Nakabachi 2008). In addition to these obligate symbionts, sap-sucking insects have diverse associations
53 with facultative (also, secondary) endosymbionts. Some of these symbionts are known to increase their
54 hosts' tolerance to environmental stressors like heat shocks, or to protect them against natural enemies
55 like fungal pathogens or parasitic wasps (Oliver, Smith & Russell 2014; Guo *et al.* 2017). Although these
56 symbionts are not required for host survival, they are nowadays recognised as fundamental to
57 understand the ecology of sap-sucking insects, including the niche they occupy, their distribution ranges
58 and their interactions with other members in the community. Opposite to obligate symbionts, facultative

59 endosymbionts of sap-sucking insects are often transmitted horizontally (Caspi-Fluger *et al.* 2012;
60 Ahmed *et al.* 2013; Chrostek *et al.* 2017) thus being mobile elements that can release herbivore
61 populations from their natural enemies, or allow them to colonise previously inhospitable habitats. The
62 role of facultative endosymbionts in host plant use, adaptation to abiotic conditions and interactions with
63 natural enemies has been studied for more than a decade, particularly in well-established model systems
64 like aphids and whiteflies (Moran, McCutcheon & Nakabachi 2008; Brownlie & Johnson 2009; Frago,
65 Dicke & Godfray 2012; Oliver, Smith & Russell 2014; Zchori-Fein, Lahav & Freilich 2014; Zytynska &
66 Weisser 2016; Guo *et al.* 2017; Vorburger 2018; Zytynska & Meyer 2019a).

67 A question that has yet to be resolved is why facultative endosymbionts are often found in only a fraction
68 of the individuals of a population, or why they are more abundant in some populations throughout the
69 distribution range of a given species (Zytynska & Weisser 2016). One explanation is that in the absence
70 of the environmental stress or of natural enemy pressure, facultative symbionts can incur fitness costs
71 (Russell & Moran 2006; Vorburger & Gouskov 2011; Vorburger, Ganesanandamoorthy & Kwiatkowski
72 2013). Experimental work on the potential fitness costs and benefits of hosting bacterial symbionts
73 indicate that these effects are variable across symbiont species, host species, and also strains or
74 genotypes within these species (Russell & Moran 2006; Rouchet & Vorburger 2012). Variation in the
75 strength of costs and benefits across different environments can lead to the differences in infection rates
76 we see in natural systems (Zytynska & Weisser 2016; Zytynska & Venturino 2018). Here, we present
77 results from a meta-analysis study to understand the generality of the costs and benefits of hosting
78 symbionts across sap-sucking insects. The majority of work in this area has been on aphids and their
79 nine common facultative bacterial symbionts *Hamiltonella defensa*, *Regiella insecticola*, *Serratia*
80 *symbiotica*, *Rickettsia*, *Rickettsiella*, *Spiroplasma*, X-type (PAXS), *Arsenophonus*, and *Wolbachia*. A
81 number of recent reviews describe the roles these symbionts can have on their hosts, and therefore we
82 will not provide this detail here (Oliver, Smith & Russell 2014; Zytynska & Weisser 2016; Guo *et al.* 2017;
83 Vorburger 2018). Understanding the benefits different endosymbiont species confer to insect herbivores,
84 and the costs they impose is thus crucial to understand one of the most diverse communities of animals
85 in terrestrial ecosystems.

86 Several recent reviews have highlighted the importance of these symbionts in understanding their hosts'
87 biology, but currently there is a lack of a quantitative and systemic analysis of the published evidences
88 exploring whether the different endosymbionts are actually beneficial or not. Although most research on

89 this topic has been limited to few model systems and symbiont species, we believe the amount of data
90 available is large enough to perform a meta-analysis. Such a systemic analysis will provide insights into
91 the role of these microbial symbionts in different sap-sucking insect groups and will identify knowledge
92 gaps at the level of both research questions and insect (or symbiont) taxa.

93 In this review we address the following questions:

- 94 1. Symbionts are widespread in sap-sucking insects, but which groups of species have been
95 sufficiently experimentally examined for the effects of symbionts on the host?
- 96 2. Many researchers make the assumption that symbionts must incur a cost to the host (otherwise
97 why don't all individuals co-host multiple beneficial symbionts?); are these costs general across
98 host and symbiont species?
- 99 3. Benefits of hosting symbionts must outweigh any costs for associations to persist, but do we have
100 sufficient experimental data to show that symbionts confer general benefits across host and
101 symbiont species?
- 102 4. The cost-benefit trade-off for hosting symbionts can be mediated by the immediate environment
103 (biotic and abiotic) depending on the benefit conferred. Sap-sucking insect communities are
104 dependent on their plant hosts, so to what extent can their host-plant species affect the costs and
105 benefits of hosting symbionts?

106

107 **Methods**

108 The majority of experimental work has used aphid systems, but there is growing popularity to study this
109 in whitefly. Literature and data pertaining to other phloem-feeders was searched for, but with little to no
110 papers relevant to our meta-analyses. We detail the individual search terms and methods below for each
111 group.

112 **Whitefly meta-analysis**

113 We searched for relevant literature using keyword searches in Web of Science finding papers published
114 until the end of March 2018. We used the terms: whitefl* OR bemisia* OR siphoninus* OR Trialeurodes
115 OR Aleurodicus OR Aleuronudus OR Dialeurodicus OR Metaleurodicus OR Palaealeurodicus OR
116 Paraleyrodes AND symbio*. Resulting in 220 potential papers. To be included in the meta-analysis,
117 papers had to satisfy the following inclusion criteria: (1) data on at least one whitefly species; (2) an

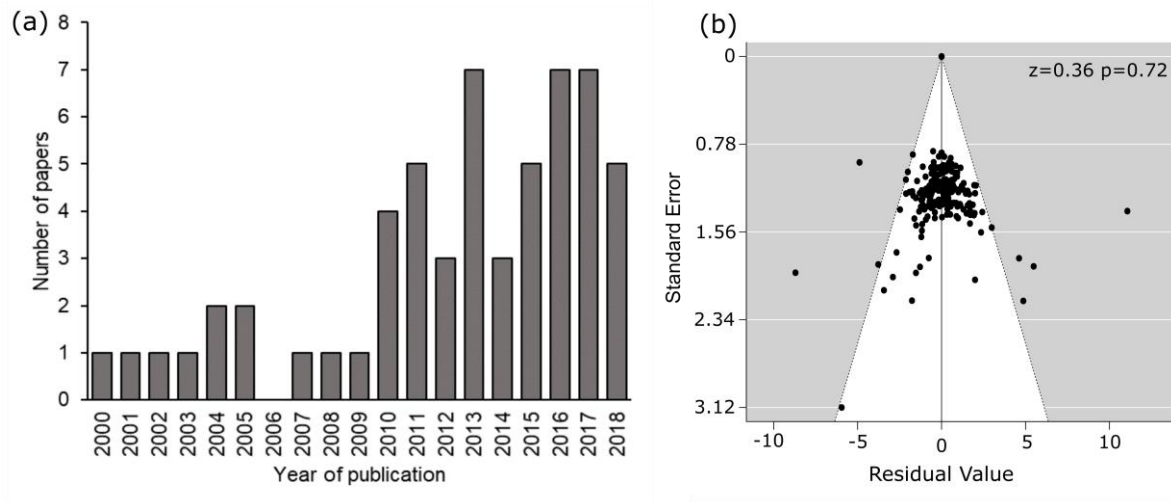
118 experimental test of symbiont effects on traits, either experimentally cured, a comparison of field-
119 collected infected and uninfected whitefly, or introgression of symbiont via crossing; (3) any of the
120 following types of variables tested: any behaviour, growth, fecundity, survival, or parasitism related
121 variable; and, (4) data on means, an estimation of variation and sample size. For the whitefly data, two
122 studies were eliminated because the antibiotic treatment also eliminated the primary symbiont. The final
123 dataset consisted of 9 papers (from 1993-2017, see Appendix 1 for list of included papers), and the only
124 variables of interest with sufficient data were growth rate (development time, growth rate and longevity),
125 fecundity, adult mass, and survival. To calculate overall symbiont effect on growth the variables
126 development time, growth rate and longevity were pooled together. For growth rate and longevity, larger
127 is better for the insect, but the opposite is true for development time (based on the slow-growth-high-
128 mortality hypothesis). In the overall test for growth, y_i was thus multiplied by -1.

129 **Aphid meta-analysis**

130 We searched for relevant literature using keyword searches in Web of Science finding papers published
131 until the end of 2018. We used the terms: ("aphid*" AND ("Hamiltonella" OR "Regiella" OR "Serratia" OR
132 "Rickettsia" OR "Rickettsiella" OR "Spiroplasma" OR "Arsenophonus" OR "Wolbachia" OR "X-type" OR
133 "PAXS")). This resulted in the extraction of 459 papers. To be included in the meta-analysis, papers had
134 to satisfy the following inclusion criteria: (1) data on at least one aphid species (Hemiptera: Aphididae);
135 (2) an experimental test of symbiont effects on aphids traits, either experimentally cured (Experimental)
136 or a comparison of field-collected infected and uninfected aphids (Natural); (3) any of the following types
137 of variables tested: aphid behaviour, growth, fecundity, survival, or parasitism-related; and, (4) data on
138 means, an estimation of variation and sample size. This resulted in 75 potential papers, but we then
139 included an additional inclusion criterion to include only traits with independent data points across at
140 least three independent studies, to reduce bias from single-lab results. Further, only data where aphids
141 hosted single symbionts were able to be used, as there was not sufficient data on multiple infections (10
142 studies, 37 data points [35 from *A. pisum* aphids] across 8 different symbiont combinations). In our final
143 57 papers (1997-2018; Figure 1a, see Appendix 2 for list of included papers) we were able to analyse the
144 effect of bacterial symbionts on: Development (age at first reproduction), Lifespan (longevity), Mass
145 (fresh weight), Fecundity (number of offspring), and Parasitism (proportion of aphids parasitized).

146 To account for differences between studies that artificially cured/inoculated aphid lines, and those
147 comparing naturally-collected infected to naturally uninfected aphid lines, we pooled data within studies

148 across aphid genotypes and symbiont strains (i.e. removing effects of within-species genetic variation);
149 this was necessary since no cross-comparison of common aphid or symbiont genetic lines was possible.
150 Data was still separated within a study for aphid species, symbiont species, and host-plant (experimental
151 plant, and plant of collection where possible).
152



153
154 **Figure 1. Summary of aphid meta-analysis data** (a) Number of aphid symbiont papers across year of
155 publication for the 57 papers used in the meta-analysis, and (b) funnel plot showing no strong publication
156 bias across these papers
157

158 The meta-analysis was conducted in R v3.5.1 (R Core Team 2019) in RStudio v1.1.463 (RStudio Team
159 2018) using the package metafor (Viechtbauer 2010). The standardised mean difference was used with
160 unbiased estimates of the sampling variances (SMDH, giving Hedges' g). This measure was used since
161 it gives a direct effect size comparison of the treated (infected with a symbiont) to untreated (no symbiont
162 control) data. We used a meta-analytic linear mixed effects model (rma) to test the effect of hosting
163 symbionts on the different aphid traits. 'Study' was included as random effect to account for multiple data
164 point across aphid and symbiont species within individual studies. Publication bias was assessed by
165 testing the funnel-plot asymmetry (Figure 1b for overall data, Figure S1 for data subsets). Data were also
166 subset into those where aphid lines had been directly compared through experimental curing/infecting
167 (Experimental) or a comparison of field-collected infected and uninfected aphids (Natural), and analyses
168 run as above on each separate dataset. The mean effect size and 95% confidence intervals are
169 presented; the mean effect size was considered significantly different from 0 if its 95% CI did not include
170 zero, and level of significant given from model outputs.

171 Further, we subset the data by aphid trait (i.e. one model for each trait) and explored differences across
172 aphid or symbiont species within these using meta-analytic linear mixed effects models by including
173 'aphid species' and 'symbiont species' as moderators (equivalent to main effects in standard linear
174 models). The interaction term was considered but in no case was there sufficient data for this to be a
175 meaningful term to include. We used model comparisons to estimate the effect of symbiont species and
176 aphid species using a LRT (likelihood ratio test) giving Chi-square and associated p-values. While the
177 overall effects of different aphid and symbionts species were analysed and relevant results presented in
178 the main text, for visual representation we present results from additional analyses that separated the
179 aphid species data into two categories: (a) *Acyrtosiphon pisum* aphid data (the commonly-used model
180 pea aphid species) and, (b) all other aphids (often representing less than half of the total data points).
181 Only aphid-symbiont combinations with at least two data points are presented in the figures.

182 **Results**

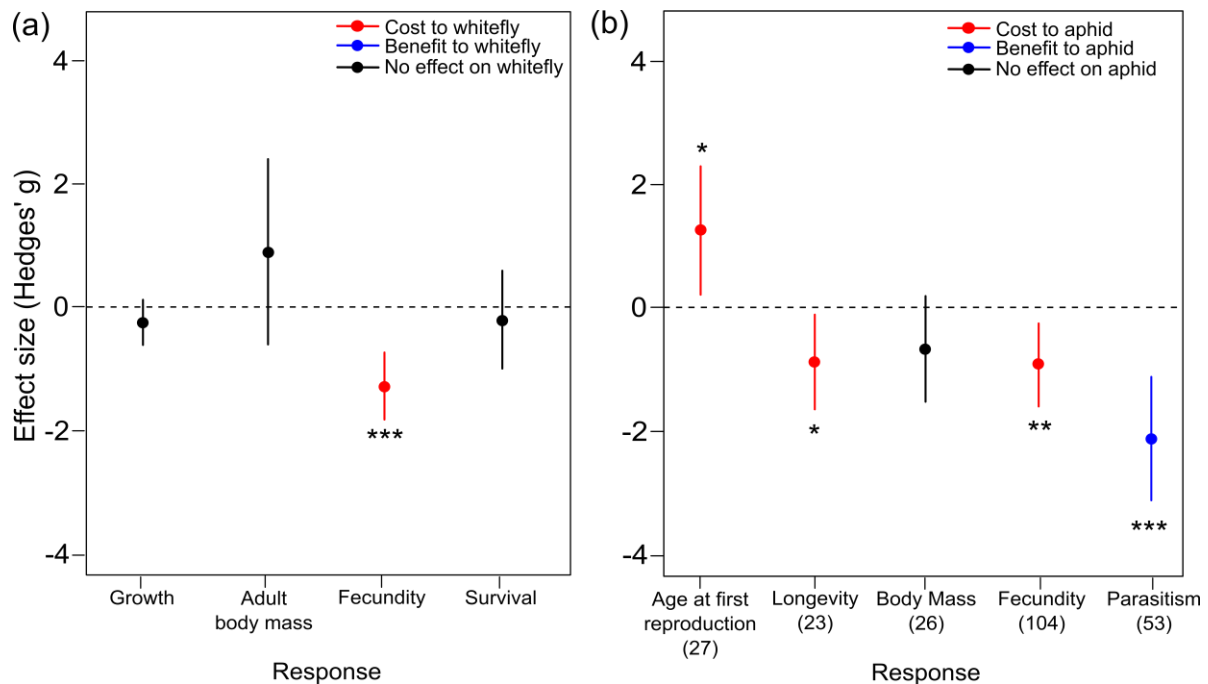
183 ***Whitefly traits***

184 The included studies are limited to a single species of whitefly (*B. tabaci*) and include some of its
185 biotypes (Q, B, among others) and three symbionts: *Hamiltonella*, *Wolbachia* and *Rickettsia*. Tests are
186 performed on different host plants and symbionts usually removed with antibiotics or via introgression
187 (Asiimwe, Kelly & Hunter 2014). Across all symbiont species, there was a cost of carrying symbionts in
188 *B. tabaci* through reduced fecundity (number of offspring). The following traits were not significantly
189 affected by symbiont carrying: growth (development time (days), growth rate, longevity (days)), adult
190 mass (body length (mm)) and survival (from egg to adult) (Figure 2a).

191 ***Aphid traits***

192 Our meta-analysis was based on data from 57 papers (2000-2018; Figure 1a, see Appendix 2 for list of
193 included papers) yielding 233 data points from 11 aphid species: *Acyrtosiphon pisum* (n=123), *Aphis*
194 *fabae* (n=32), *Sitobion avenae* (n=32), *Aphis craccivora* (n=13), *Acyrtosiphon kondoi* (n=12),
195 *Macrosiphum euphorbiae* (n=5), *Megoura crassicauda* (n=5), *Aphis glycines* (n=4), *Myzus persicae*
196 (n=3), *Rhopalosiphum padi* (n=3), *Obtusicauda frigida* (n=1). We compared aphid traits of aphids
197 infected with a symbiont to uninfected aphids (data were pooled within a study across aphid genotypes
198 and symbiont strains, but separated by aphid species, symbiont species, and host-plant). The data were
199 robust against publication bias as measured through funnel-plot asymmetry ($z = 0.363$, $p = 0.717$; Figure
200 1b).

201 The number of data points within our final dataset for the different response variables were: Age at first
 202 reproduction (N=27: Experimental n=26, Natural n=1; from 14 papers), Longevity (N=23: Experimental
 203 n=20, Natural n=3; from 12 papers), Mass (N=26: Experimental n=18, Natural n=8; from 15 papers),
 204 Fecundity (N=104: Experimental n=87, Natural n=17; from 40 papers), Parasitism (N=53: Experimental
 205 n=36, Natural n=17; from 28 papers).



206
 207 **Figure 2. Overall effect of endosymbionts** on (a) whitefly *Bemisia tabaci* across four different *B. tabaci*
 208 life history traits, and (b) aphids (Hemiptera: Aphididae) across five different variables, including four
 209 related to aphid life history traits and the last related to resistance to parasitism (proportion of aphids
 210 parasitized).

211 * P<0.05, ** P<0.01, *** P<0.001

212

213 Effects across all aphids and symbionts

214 Across all aphid and symbiont species, there was a cost to aphids through increased development time
 215 (age at first reproduction), reduced longevity (days), reduced fecundity (number of offspring) (Figure 2b).
 216 However, there was also a strong benefit to aphids of hosting endosymbionts that conferred resistance
 217 against attacks by parasitic wasps (reduced proportion of aphids with a symbiont are parasitized; Figure
 218 2b). The measures for age at first reproduction, longevity, and parasitism, were relatively consistent
 219 across studies and therefore we are able to present mean values for these effects. We found that age at
 220 first reproduction for the aphids was increased from 8.75 ± 1.21 days (uninfected controls) to 9.06 ± 1.21

221 days when hosting a symbiont, and longevity was decreased by five days when hosting a symbiont
222 (control: 30.31 ± 5.02 days, symbiont: 24.33 ± 4.95 days). The proportion of aphids parasitized reduced
223 from 0.54 ± 0.25 (uninfected controls) to 0.36 ± 0.29 when hosting a symbiont. The measures of body mass
224 and fecundity varied across studies, thereby reducing our ability to provide reliable mean values for these
225 traits.

226 **Experimental lines versus naturally-collected lines of aphid**

227 When the data were compared between those aphid lines that had been experimentally infected/cured
228 and those that were collected from the field as infected or uninfected, we see that there is substantial
229 variation in the results (Figure S2). The unequal distribution of data points (higher number of data points
230 for experimental lines compared to natural lines) needs to be noted here and results interpreted with this
231 potential strong bias in mind. The results indicate that the effects are stronger and less variable within
232 experimental studies, with a lack of overall significant results and greater range of data for 'natural' aphid
233 lines collected from the field. In some cases, e.g. fecundity, the data shows a potential for a change in
234 the direction of the result; however, further exploration of the data suggests that this may be influenced
235 by a few individual data points owing to the lack of data across multiple species, symbionts, and
236 laboratories.

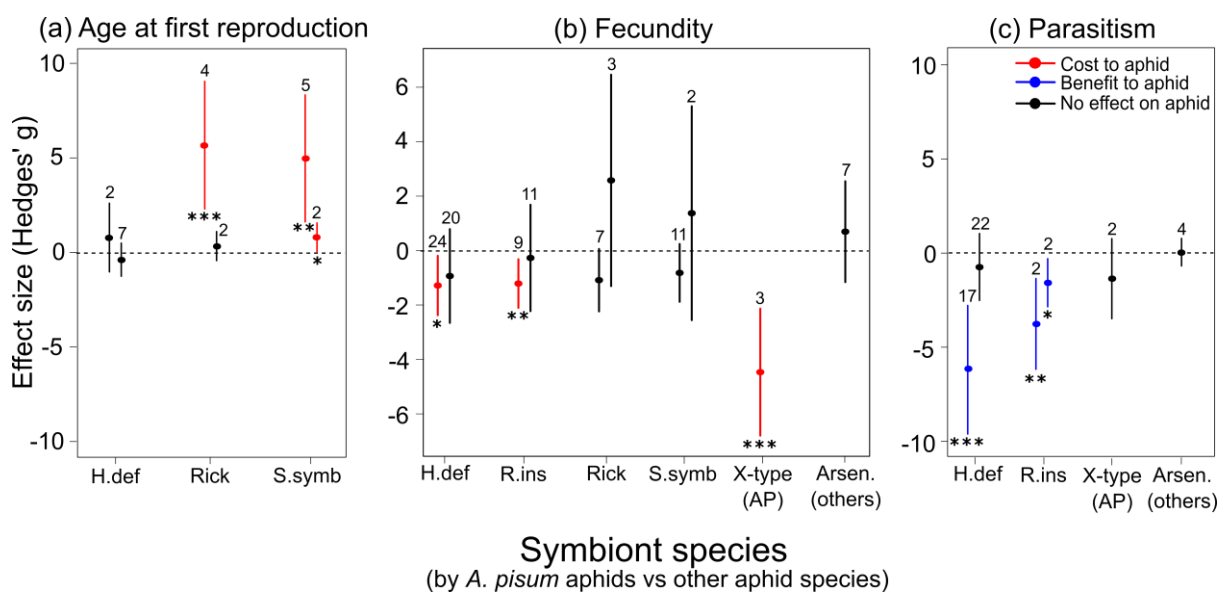
237 **Effects within aphid traits by symbiont and aphid species**

238 The age at first reproduction (days) in the aphid was increased by symbionts for all aphid species, but
239 the magnitude varied across aphid species ($X^2=18.54$, $df=6$, $P<0.001$), and the effect varied across the
240 different symbiont species ($X^2=12.07$, $df=7$, $P=0.007$). *Hamitonella defensa* symbionts did not increase
241 aphid age at first reproduction, while *Rickettsia* sp. increased only for *A. pisum* aphids and *Serratia*
242 *symbiotica* symbionts for all aphids (Figure 3a).

243 Overall, aphid fecundity was reduced for aphids hosting a symbiont (Figure 2b) but the effect size was
244 dependent on the aphid species ($X^2=13.79$, $df=9$, $P=0.003$) and, in part, the symbiont ($X^2=12.44$, $df=7$,
245 $P=0.088$). Both *H. defensa* and *Regiella insecticola* reduced the fecundity of *A. pisum* aphids but there
246 was no evidence for this to affect other aphid species (Figure 3b). *Rickettsia*, *S. symbiotica*, and
247 *Arsenophonus* sp. had no effect on aphid fecundity, while the *X-type* symbiont decreased *A. pisum* aphid
248 fecundity (not studied in other aphids) (Figure 3b).

249 Symbiont-conferred resistance to parasitic wasps was variable across symbiont ($X^2=18.57$, $df=7$,
 250 $P=0.002$) and aphid species ($X^2=3.73$, $df=7$, $P=0.053$). *Regiella insecticola* symbionts reduced the
 251 proportion of aphids parasitized for all aphids studied, whereas *H. defensa* only reduced this for *A. pisum*
 252 aphids, no other symbionts showed a significant effect (Figure 3c).

253 The influence of aphid or symbiont species was not presented for the aphid traits of body mass or
 254 longevity due to strong effects of publication bias within these traits (Figure S2), potentially leading to
 255 biased conclusions; however, analysis indicated little variation in the effect of symbiont hosting across
 256 the different aphid and symbiont species tested.



257
 258 **Figure 3. Variation in effect size of hosting symbionts for *A. pisum* pea aphids compared to other**
 259 **aphid species** on (a) aphid age at first reproduction (days) where a positive effect indicates a
 260 lengthened development time when hosting the symbiont; (b) fecundity, where a negative effect indicates
 261 fewer offspring produced when hosting the symbiont; and, (c) parasitism, where a negative effect
 262 indicates a reduced proportion of aphids were parasitized by a parasitic wasp when hosting the
 263 symbiont. The clustered lines show results from *A. pisum* pea-aphids (left lines, AP) compared to all
 264 other aphid species combined (right lines, others). Data points are the mean effect size with 95%
 265 confidence intervals, numbers above the lines indicates the number of data points in the meta-analysis.
 266

267 Discussion

268 In this meta-analysis we have explored the benefits and potential costs associated with carrying
 269 facultative endosymbionts in phloem-feeding insects. This is the first study that explores this question
 270 quantitatively across phloem-feeding Hemiptera [i.e. whiteflies, mealybugs, aphids, psyllids,
 271 planthoppers, leafhoppers, lygaeids, pentatomids and coreids (Dolling 1991)] and Thysanoptera. Our first
 272 highlight is that, although these two Orders comprise almost 110,000 described species (Stork 2018),
 273 most of them sap-suckers, information on the costs and benefits of facultative symbiont infection is

274 limited to aphids and whiteflies. Within these two groups, we found a strong bias towards a few well-
275 studied species, such as the model pea aphid (*A. pisum*), the black-bean aphid (*Aphis fabae*), the cereal
276 aphid (*Sitobion avenae*), and the whitefly *Bemisia tabaci*. This bias towards a few model species is likely
277 due to their importance as agricultural pest species and long history of ecological studies involving these
278 species. However, there are many other agriculturally-important sap-feeders, particularly aphid species,
279 that have been little used in symbiont studies. Other than lack of research effort, one reason for this bias
280 might be due to the difficulty of artificially removing symbionts in some insect species. In aphids using
281 antibiotics to "cure" them from secondary symbionts is simple and well-documented, albeit time-
282 consuming (Simon *et al.* 2007); however, this technique does not work for the potato aphid *Macrosiphum*
283 *euphorbiae* because the antibiotic treatment eliminates both facultative and the obligatory *Buchnera*
284 symbionts resulting in aphid death (Hackett, Karley & Bennett 2013). One work around, as done by the
285 authors working on *M. euphorbiae*, is by testing symbiont effects in various field-collected genotypes with
286 and without the bacterium (termed 'Natural' aphid lines in this meta-analysis), or creating infected lines
287 via introgression (as done in whiteflies, e.g. (Asiimwe, Kelly & Hunter 2014)).

288 We find support for an overall fitness cost of hosting bacterial symbionts in the aphid and whitefly species
289 studied; yet, the impact of these is strongly insect and symbiont species dependent. The general costs to
290 the aphid occur through increased time until the first reproduction, reduced fecundity and reduced
291 longevity. Thus, symbionts decrease aphid fitness by delaying development, reducing lifespan, and
292 reducing offspring production during this time. Hosts that carry costly symbionts but that do not confer
293 any benefit are expected to be lost in populations via purifying selection. We could show that there are
294 general benefits of symbionts through decreased parasitism rates, such that certain symbionts protect
295 certain aphid species from attack by specialist parasitic wasps (first shown by Oliver *et al.* 2003).

296 Although other benefits, such as resistance to entomopathogenic fungi or heat stress have been
297 highlighted in reviews on aphid symbiont effects (Oliver, Smith & Russell 2014; Zytynska & Weisser
298 2016; Guo *et al.* 2017; Vorburger 2018), these traits lack sufficient data across multiple aphid species
299 and symbionts to be included in a meta-analysis. In whiteflies, facultative symbionts were found to be
300 costly only through reduced fecundity, but as far as we are aware any benefits such as symbiont-
301 mediated resistance to natural enemies have never been tested in this insect group. Despite this, costly
302 symbionts like *Hamiltonella* are highly prevalent in whiteflies (Gueguen *et al.* 2010; Zchori-Fein, Lahav &
303 Freilich 2014), which suggests that the benefits associated to these bacteria are yet to be discovered.

304 The species-specific costs and benefits we identified in this meta-analysis have the potential to
305 contribute to the variation in symbiont-hosting frequencies observed in the field within and among
306 populations (Zytynska & Weisser 2016). All the experiments included in this meta-analysis compared
307 infected with uninfected aphid lines, and in the field both infected and uninfected aphids coexist
308 (Zytynska & Weisser 2016). The reduced fitness of aphids hosting a symbiont means they will be
309 outcompeted by the uninfected aphids when there is no benefit (e.g. through resistance), confirming the
310 'only helpful when required' statement of Vorburger and Gouskov (2011). Yet, the magnitude of the costs
311 and benefits will determine the impact on individual populations.

312 In this meta-analysis, we compared experiments that either directly assessed symbiont effects using
313 lines of aphid that had been artificially cured or infected, or compared lines of naturally-infected to
314 naturally-uninfected aphids collected to the field. We found that this separation resulted in strong
315 differences in the effects. A greater amount of variation with reduced effects on the aphid were seen for
316 those aphid lines that were collected from the field. Since this data comparison was potentially highly
317 biased, with small sample sizes for the naturally-collected aphids, we must interpret this carefully.
318 However, it may suggest that 'successful' aphid-symbiont combinations incur fewer costs, but perhaps
319 also reduced benefits. Or, this increased variation in effect size might indicate it is strongly dependent on
320 the particular combination of aphid (species, genotype), symbiont (species, strain), and surrounding
321 challenges (e.g. microclimate, host-plant, parasitism rate). Perhaps important benefits can only be
322 appreciated under more natural conditions. A field study in the US, for example, revealed that the
323 prevalence of the defensive symbiont *H. defensa* in *A. pisum* aphids increased throughout the season in
324 response to increased densities of parasitic wasps (Smith *et al.* 2015). This correlation, however, was
325 only significant in one of the two sites studied, a result that may reflect that parasitic wasps are not the
326 only natural enemies dictating the fate of symbiont-carrying insects. In addition, most studies have been
327 done on a restricted set of host plants, while only a small proportion of sap-sucking insects are
328 monophagous (most feed on more than one plant species). Data were collected on 'experimental host-
329 plant' and 'host-plant of aphid collection' but was insufficient for inclusion in the meta-analysis, indicating
330 the need for more empirical studies in this area.

331 Bringing the plant layer into account will certainly change our understanding of the cost-benefit balance
332 of symbiont infection in phloem feeders, with work suggesting the surrounding plant community can have
333 strong impacts on aphid endosymbiont communities (Zytynska *et al.* 2016; Zytynska & Meyer 2019a).

334 Moreover, in the aphids *A. pisum* and *C. cedri*, the facultative symbiont *S. symbiotica* can assist the
335 obligatory symbiont at the nutritional level, potentially enabling host-feeding on a wider selection of plants
336 (Koga, Tsuchida & Fukatsu 2003; Lamelas *et al.* 2011). In whiteflies (Su *et al.* 2015) and aphids (Frago
337 *et al.* 2017) recent studies also revealed that symbionts are able to help their hosts circumvent plant
338 defences that are triggered upon insect attack. Altogether, this means that symbionts could have a much
339 wider impact on aphid populations than is currently empirically tested. Thus, to better understand the
340 balance between costs and benefits of symbiont infection, and ultimately their prevalence in natural
341 populations, a wider community perspective is necessary. A recent review discusses the importance of
342 the immediate surrounding biotic community (plant diversity, natural enemy diversity) in combination with
343 the abiotic environment on mediating aphid-symbiont interactions (Zytynska & Meyer 2019a).

344 Complex community interactions also occur among symbionts inside their hosts, with potential
345 implications for the benefits that these bacteria provide (Ferrari & Vavre 2011). Our meta-analysis is
346 based only on insects with single symbiont infections since we lacked sufficient data on the role of
347 multiple symbiont infections. In the field it is estimated that aphids, for instance, host 0-4 symbionts per
348 individual (Ferrari *et al.* 2012; Russell *et al.* 2013; Smith *et al.* 2015; Zytynska *et al.* 2016), and that
349 multiple infections are particularly common in some genus like *Macrosiphum* (Henry *et al.* 2015). More
350 importantly, where symbiont co-infections occur strong fitness costs are often observed (Oliver, Moran &
351 Hunter 2006; Guay *et al.* 2009; Leclair *et al.* 2017; McLean *et al.* 2018). It would be very interesting to
352 test this across different aphid species, asking whether the proportion of multiple infections correlates
353 negatively with the fitness costs they impose. To better understand how multiple infections arise, more
354 work on how facultative symbionts are horizontally transferred within populations of the same species or
355 among species is also needed. While aphid symbionts are predominantly vertically transmitted from
356 mother to offspring, there is also evidence of horizontal transfer of symbionts among aphids during
357 sexual reproduction (Moran & Dunbar 2006), by parasitoids when ovipositing eggs into aphids (Gehrer &
358 Vorburger 2012), or even through infected honeydew (Darby & Douglas 2003). Based on a few
359 laboratory studies, aphid hosts impose little constraints to symbiont acquisition even if the host already
360 carries a facultative symbiont, demonstrated by successful microinjecting of different symbiont species
361 and strains into pea aphids (Leclair *et al.* 2017; McLean *et al.* 2018). A recent paper used mathematical
362 modelling to further show the importance of horizontal transmission of symbionts among aphids, focusing
363 on the potential of parasitoid wasps to transmit protective symbionts among aphids (Zytynska &

364 Venturino 2018). A low rate of horizontal transmission led to coexistence of uninfected aphids, infected
365 aphids, and parasitoid wasps, with the percentage of infected aphids ranging from 30-70% which is in
366 agreement with infection rates observed in field surveys (Zytynska & Weisser 2016; Zytynska &
367 Venturino 2018).

368
369 Many of these symbionts protect agricultural insect pests from their natural enemies, which is
370 counteractive to the aims of biological control programs (Vorburger 2018). Understanding how different
371 insect species interact with the common symbionts species, how symbionts are transmitted between
372 insect individuals, and the likely costs and benefits, all help to devise management programs to reduce
373 the impact of these symbionts. In a closed greenhouse system, the spread of a protective symbiont can
374 hinder biological control efforts, yet strategies to increase natural enemy diversity could mitigate this
375 impact (Vorburger 2018). In the field, if natural enemy density is low, as might occur in a monoculture
376 crop field, then insects with no symbionts (higher fitness) would outcompete insects with symbionts
377 (Zytynska & Meyer 2019b). Coupled with pesticide resistance evolution (e.g. seen in cereal aphids in
378 Europe; Malloch *et al.* (2016)), this can lead to increased risks of pest outbreaks.

379

380 **Conclusion**

381 Current molecular methods allow us to study the intricate ways insects establish mutualistic symbioses
382 with microbial partners. We used meta-analysis techniques to show the general costs (through increased
383 development time, reduced longevity and reduced fecundity) and benefits (increases resistance to
384 parasitic wasps) of hosting bacterial symbionts in sap-sucking insects. Current data is strongly biased
385 towards a few species of aphid and whitefly, and that there is a large variation of effects among insect as
386 well as symbiont species. Thus, the results cannot reliably be extrapolated to other phloem-feeding taxa,
387 and not even to other aphid or whitefly species. The impact of cost-benefit trade-offs in natural systems
388 are still to be uncovered, but an appreciation of the diversity of potential outcomes due to the species or
389 genetics of the insect/symbiont and the surrounding environment (plant diversity, natural enemy diversity,
390 microclimate) will benefit the design of future studies. While many of the studied insect species are
391 agricultural pests, studies in which the phenotypic consequences of facultative symbiont infection are
392 measured in non-model species are urgently needed. For agricultural systems, the spread of protective
393 symbionts in sap-sucking insects can hinder biological control efforts while reduced densities of natural

394 enemies might select for uninfected aphids with higher reproductive fitness. In both cases, this can lead
395 to pest outbreaks.

396

397 **Data availability**

398 All papers used in the meta-analysis are detailed in the appendices. Data will be made readily available
399 through contact with the corresponding author until final publication of the paper when data will be made
400 publically available.

401

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408

409 **References**

- 410 Ahmed, M.Z., De Barro, P.J., Ren, S.-X., Greeff, J.M. & Qiu, B.-L. (2013) Evidence for horizontal
411 transmission of secondary endosymbionts in the *Bemisia tabaci* cryptic species complex. *PLoS*
412 *ONE*, **8**, e53084.
- 413 Asimwe, P., Kelly, S.E. & Hunter, M.S. (2014) Symbiont infection affects whitefly dynamics in the field.
414 *Basic and Applied Ecology*, **15**, 507-515.
- 415 Brownlie, J.C. & Johnson, K.N.J.T.i.m. (2009) Symbiont-mediated protection in insect hosts. **17**, 348-354.
- 416 Caspi-Fluger, A., Inbar, M., Mozes-Daube, N., Katzir, N., Portnoy, V., Belausov, E., Hunter, M.S. &
417 Zchori-Fein, E. (2012) Horizontal transmission of the insect symbiont *Rickettsia* is plant-
418 mediated. *Proceedings of the Royal Society of London B: Biological Sciences*, **279**, 1791-1796.
- 419 Chrostek, E., Pelz-Stelinski, K., Hurst, G.D. & Hughes, G.L.J.F.i.m. (2017) Horizontal transmission of
420 intracellular insect symbionts via plants. **8**, 2237.
- 421 Darby, A. & Douglas, A. (2003) Elucidation of the transmission patterns of an insect-borne bacterium.
422 *Applied and Environmental Microbiology*, **69**, 4403-4407.
- 423 Dillon, R. & Dillon, V.J.A.R.i.E. (2004) The gut bacteria of insects: nonpathogenic interactions. **49**, 71-92.

- 424 Dolling, W.R. (1991) *Hemiptera*. Oxford University Press.
- 425 Douglas, A. (1998) Nutritional interactions in insect-microbial symbioses: aphids and their symbiotic
426 bacteria Buchnera. *Annual Review of Entomology*, **43**, 17-37.
- 427 Feldhaar, H. (2011) Bacterial symbionts as mediators of ecologically important traits of insect hosts.
428 *Ecological Entomology*, **36**, 533-543.
- 429 Ferrari, J. & Vavre, F. (2011) Bacterial symbionts in insects or the story of communities affecting
430 communities. *Phil Trans R Soc B*, **366**, 1389-1400.
- 431 Frago, E., Dicke, M. & Godfray, H.C.J. (2012) Insect symbionts as hidden players in insect-plant
432 interactions. *Trends in ecology & Evolution*, **27**, 705-711.
- 433 Frago, E., Mala, M., Weldegergis, B.T., Yang, C., McLean, A., Godfray, H.C.J., Gols, R. & Dicke,
434 M.J.N.c. (2017) Symbionts protect aphids from parasitic wasps by attenuating herbivore-induced
435 plant volatiles. **8**, 1860.
- 436 Gehrer, L. & Vorburger, C. (2012) Parasitoids as vectors of facultative bacterial endosymbionts in aphids.
437 *Biology letters*, **8**, 613-615.
- 438 Guay, J.F., Boudreault, S., Michaud, D. & Cloutier, C. (2009) Impact of environmental stress on aphid
439 clonal resistance to parasitoids: Role of *Hamiltonella defensa* bacterial symbiosis in association
440 with a new facultative symbiont of the pea aphid. *Journal of Insect Physiology*, **55**, 919-926.
- 441 Gueguen, G., Vavre, F., Gnankine, O., Peterschmitt, M., Charif, D., Chiel, E., Gottlieb, Y., Ghanim, M.,
442 Zchori-Fein, E. & Fleury, F. (2010) Endosymbiont metacommunities, mtDNA diversity and the
443 evolution of the *Bemisia tabaci* (Hemiptera: Aleyrodidae) species complex. *Molecular Ecology*,
444 **19**, 4365-4376.
- 445 Guo, J., Hatt, S., He, K., Chen, J., Francis, F. & Wang, Z. (2017) Nine facultative endosymbionts in
446 aphids. A review. *Journal of Asia-Pacific Entomology*.
- 447 Hackett, S.C., Karley, A.J. & Bennett, A.E. (2013) Unpredicted impacts of insect endosymbionts on
448 interactions between soil organisms, plants and aphids. *Proceedings of the Royal Society B-*
449 *Biological Sciences*, **280**.
- 450 Henry, L.M., Maiden, M.C., Ferrari, J. & Godfray, H.C. (2015) Insect life history and the evolution of
451 bacterial mutualism. *Ecology letters*, **18**, 516-525.
- 452 Koga, R., Tsuchida, T. & Fukatsu, T. (2003) Changing partners in an obligate symbiosis: a facultative
453 endosymbiont can compensate for loss of the essential endosymbiont Buchnera in an aphid.
454 *Proceedings of the Royal Society B-Biological Sciences*, **270**, 2543-2550.

- 455 Lamelas, A., Gosalbes, M.J., Manzano-Marin, A., Pereto, J., Moya, A. & Latorre, A. (2011) *Serratia*
456 *symbiotica* from the aphid *Cinara cedri*: a missing link from facultative to obligate insect
457 endosymbiont. *PLoS Genet*, **7**, e1002357.
- 458 Leclair, M., Polin, S., Jousseau, T., Simon, J.C., Sugio, A., Morliere, S., Fukatsu, T., Tsuchida, T. &
459 Outreman, Y. (2017) Consequences of coinfection with protective symbionts on the host
460 phenotype and symbiont titres in the pea aphid system. *Insect Science*, **24**, 798-808.
- 461 Malloch, G., Pickup, J., Hight, F., Foster, S., Williamson, M. & Fenton, B. (2016) Assessment of the
462 spread of pyrethroid resistant *Sitobion avenae* in the UK and an update on changes in the
463 population structure of *Myzus persicae* in Scotland. *Proceedings Crop Protection in Northern*
464 *Britain*, 223-228.
- 465 McLean, A.H.C., Parker, B.J., Hrcek, J., Kavanagh, J.C., Wellham, P.A.D. & Godfray, H.C.J. (2018)
466 Consequences of symbiont co-infections for insect host phenotypes. *Journal of Animal Ecology*,
467 **87**, 478-488.
- 468 Moran, N.A. & Dunbar, H.E. (2006) Sexual acquisition of beneficial symbionts in aphids. *Proceedings of*
469 *the National Academy of Sciences*, **103**, 12803-12806.
- 470 Moran, N.A., McCutcheon, J.P. & Nakabachi, A. (2008) Genomics and evolution of heritable bacterial
471 symbionts. *Annual review of genetics*, **42**, 165-190.
- 472 Oliver, K.M., Moran, N.A. & Hunter, M.S. (2006) Costs and benefits of a superinfection of facultative
473 symbionts in aphids. *Proceedings of the Royal Society B: Biological Sciences*, **273**, 1273-1280.
- 474 Oliver, K.M., Russell, J.A., Moran, N.A. & Hunter, M.S. (2003) Facultative bacterial symbionts in aphids
475 confer resistance to parasitic wasps. *PNAS*, **100**, 1803.
- 476 Oliver, K.M., Smith, A.H. & Russell, J.A. (2014) Defensive symbiosis in the real world – advancing
477 ecological studies of heritable, protective bacteria in aphids and beyond. *Functional Ecology*, **28**,
478 341-355.
- 479 R Core Team (2019) R: A Language and Environment for Statistical Computing. R Foundation for
480 Statistical Computing, Vienna, Austria.
- 481 Rouchet, R. & Vorburger, C. (2012) Strong specificity in the interaction between parasitoids and
482 symbiont-protected hosts. *Journal of Evolutionary Biology*, **25**, 2369-2375.
- 483 RStudio Team (2018) RStudio: Integrated Development for R. RStudio, Inc., Boston, MA.

- 484 Russell, J.A. & Moran, N.A. (2006) Costs and benefits of symbiont infection in aphids: variation among
485 symbionts and across temperatures. *Proceedings of the Royal Society B: Biological Sciences*,
486 **273**, 603-610.
- 487 Simon, J.C., Sakurai, M., Bonhomme, J., Suchida, T., Koga, R. & Fukatsu, T. (2007) Elimination of a
488 specialised facultative symbiont does not affect the reproductive mode of its aphid host. *Ecol*
489 *Entomology*, **32**, 296-301.
- 490 Smith, A.H., Lukasik, P., O'Connor, M.P., Lee, A., Mayo, G., Drott, M.T., Doll, S., Tuttle, R., Disciullo,
491 R.A., Messina, A., Oliver, K.M. & Russell, J.A. (2015) Patterns, causes and consequences of
492 defensive microbiome dynamics across multiple scales. *Mol Ecol*, **24**, 1135-1149.
- 493 Stork, N.E. (2018) How many species of insects and other terrestrial arthropods are there on Earth?
494 *Annual Review of Entomology*, **63**, 31-45.
- 495 Su, Q., Oliver, K.M., Xie, W., Wu, Q.J., Wang, S.L. & Zhang, Y.J. (2015) The whitefly-associated
496 facultative symbiont *Hamiltonella defensa* suppresses induced plant defences in tomato.
497 *Functional Ecology*, **29**, 1007-1018.
- 498 Viechtbauer, W. (2010) Conducting meta-analyses in R with the metafor package. *Journal of Statistical*
499 *Software*, **36**, 1-48.
- 500 Vorburger, C. (2018) Symbiont-conferred resistance to parasitoids in aphids – Challenges for biological
501 control. *Biological Control*, **116**, 17-26.
- 502 Vorburger, C., Ganesanandamoorthy, P. & Kwiatkowski, M. (2013) Comparing constitutive and induced
503 costs of symbiont-conferred resistance to parasitoids in aphids. *Ecol Evol*, **3**, 706-713.
- 504 Vorburger, C. & Gouskov, A. (2011) Only helpful when required: a longevity cost of harbouring defensive
505 symbionts. *Journal of Evolutionary Biology*, **24**, 1611-1617.
- 506 Zchori-Fein, E., Lahav, T. & Freilich, S. (2014) Variations in the identity and complexity of endosymbiont
507 combinations in whitefly hosts. *Front Microbiol*, **5**, 310.
- 508 Zytynska, S.E. & Meyer, S.T. (2019a) Effects of biodiversity in agricultural landscapes on the protective
509 microbiome of insects - a review. *Entomol Exp Appl*, **167**, 2-13.
- 510 Zytynska, S.E. & Meyer, S.T. (2019b) Effects of biodiversity in agricultural landscapes on the protective
511 microbiome of insects - a review. *Entomologia Experimentalis et Applicata*, **167**, 2-13.
- 512 Zytynska, S.E., Meyer, S.T., Sturm, S., Ullmann, W., Mehrparvar, M. & Weisser, W.W. (2016) Secondary
513 bacterial symbiont community in aphids responds to plant diversity. *Oecologia*, **180**, 735-747.

- 514 Zytynska, S.E. & Venturino, E. (2018) Modelling the role of vector transmission of aphid bacterial
515 endosymbionts and the protection against parasitoid wasps. *18th International Symposium on*
516 *Mathematical and Computational Biology*. Springer, Morocco.
- 517 Zytynska, S.E. & Weisser, W.W. (2016) The natural occurrence of secondary bacterial symbionts in
518 aphids. *Ecol Entomology*, **41**, 13-26.
- 519