

1 **Antimicrobial susceptibility and serotype distribution of *Streptococcus agalactiae* recto-**
2 **vaginal colonizing isolates from pregnant women at a tertiary hospital in Pretoria, South**
3 **Africa: an observational descriptive study**

4 **Short Title: *Streptococcus agalactiae* antimicrobial susceptibilities and serotype**
5 **distribution**

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24 **Abstract**

25 ***Introduction***

26 *Streptococcus agalactiae* or Group B Streptococcus (GBS) is a significant cause of neonatal
27 sepsis. Intrapartum antibiotic prophylaxis is recommended for pregnant women identified
28 to be recto-vaginally colonised between 34-37 weeks gestational age to decrease the risk of
29 invasive disease in their newborns. The aim of this study was to investigate serotype
30 distribution and antimicrobial susceptibility patterns of GBS isolates cultured from recto-
31 vaginal specimens during pregnancy.

32 ***Methods***

33 Sixty-nine archived maternal colonizing isolates were tested against penicillin, erythromycin,
34 clindamycin, vancomycin and levofloxacin. Minimum Inhibitory Concentration (MIC) testing
35 was performed using the E-test method. Serotyping was performed by latex agglutination
36 method.

37 ***Results***

38 The most common serotypes detected were Ia (54%), III (20%), V (16%), II (6%), IV (2%) and
39 Ib (1%), respectively. All isolates were fully susceptible to penicillin, vancomycin and
40 levofloxacin. Eight (11%) and 50 (56%) isolates showed intermediate resistance to
41 erythromycin and clindamycin respectively, and one isolate was resistant to erythromycin.
42 MLS_B phenomenon was noted in 3 (4%) of the isolates.

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44 **Conclusion**

45 GBS colonizing isolates remain susceptible to penicillin and remains the drug of choice for
46 intrapartum antibiotic prophylaxis and treatment of invasive disease in newborns.
47 Macrolides should only be used if clinically indicated due to the high prevalence of
48 intermediate resistance. A hexavalent GBS vaccine currently under development would
49 provide coverage for 100% of the isolates identified in this study.

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59 **Keywords**

60 *Streptococcus agalactiae*

61 Antibiotics susceptibility

62 Prophylaxis

63 Serotypes

64 Vaccine

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67 **Introduction**

68 *Streptococcus agalactiae* or Group B Streptococcus (GBS) remains a significant cause of
69 early-onset (<7 days age; EOD) and late-onset (7-89 days age; LOD) invasive disease [1]. The
70 incidence of EOD has declined significantly in countries where universal screening of
71 pregnant women for GBS colonization is undertaken between 34-37 weeks of gestational
72 age and intrapartum antibiotic prophylaxis (IAP) during labour is provided to colonized
73 women [2].

74 Penicillin remains the drug of choice for IAP and for the treatment of GBS-EOD and LOD.
75 Women with a history of penicillin allergy but at low risk for penicillin anaphylaxis should
76 receive alternative treatment with a cephalosporin such as cefazolin instead of
77 erythromycin or clindamycin [2]. This is due to an increasing resistance of GBS to
78 clindamycin and erythromycin. Reported rates of resistance of GBS to erythromycin range
79 from 25-32% and to clindamycin from 13-20% [2]. Vancomycin is an appropriate alternative
80 for patients with a history of anaphylaxis to penicillin and when an isolate is resistant to
81 clindamycin.

82 An effective GBS vaccine may prevent a broad scope of GBS associated diseases, such as
83 GBS-EOD, GBS-LOD, spontaneous abortions, stillbirth and maternal bacteraemia [2,4]. One
84 approach of vaccine development is to target the capsular polysaccharide (CPS) of GBS. GBS

85 serological grouping is based on the polysaccharide capsule. There are currently 10
86 serotypes i.e. Ia, Ib and II-IX. The distribution of the five most common GBS serotypes in
87 South Africa causing invasive disease are III-55.4%, Ia-28.2%, V-7.9%, II- 3.6% and Ib-3.4%, II-
88 5% [5]. This compares similarly to the global distribution [6]. Seven to thirty percent of GBS
89 isolates are serologically non-serotypeable [7].

90 The aim of this study was to determine the serotype distribution of recto-vaginal colonizing
91 isolates from pregnant women and the antimicrobial susceptibility patterns thereof.

92 **Materials and methods**

93 **Study Design**

94 This was a laboratory based observational study examining 69 archived isolates from a study
95 done in 2014 which investigated the prevalence of GBS colonisation in pregnant women
96 between 26 and 37 weeks gestation [8]. In that study, 284 pregnant women were enrolled
97 from an antenatal clinic and tested for GBS colonisation by Xpert GBS and culture. The
98 colonisation rate was found to be 25% by culture and 24% by Xpert GBS [8]. The GBS
99 isolates were stored in trypticase soy broth with 5% glycerol.

100 The women had been enrolled and microbiology testing done at the Tshwane Academic
101 Division Microbiology laboratory of the National Health Laboratory Services (NHLS). The
102 serotyping of the isolates was conducted at the Respiratory and Meningeal Pathogens
103 Research Unit (RMPRU, Johannesburg.)

104 **Specimen processing**

105 The stored isolates were sub-cultured on 5% sheep blood agar and incubated for 24 hours in
106 5% CO₂. Beta-haemolytic colonies were then lawned onto Mueller Hinton agar with 5%
107 sheep blood for Minimum Inhibitory Concentration (MIC) testing using Etest (bioMeriueux,
108 France) strips. Five antibiotics were tested for each isolate viz. penicillin, vancomycin,
109 erythromycin, clindamycin and levofloxacin. Plates were incubated for 24 hours in 5% CO₂
110 at 35-37°C . The MIC's were determined using the latest CLSI breakpoints (2015) and the
111 quantitative variables obtained were classified as susceptible, non-susceptible, intermediate
112 and resistant. The MIC's of GBS isolates which tested non-susceptible or resistant for any of
113 the 5 antibiotics, were repeated and the results confirmed. Furthermore, two observers
114 read the MIC values of all the isolates to minimise inter-observer variability or any form of
115 bias.

116 As per CLSI guidelines for beta-haemolytic streptococci, MLS_B testing was performed on
117 each isolate to test for inducible clindamycin resistance. The isolates were plated on Mueller
118 Hinton plus 5% sheep blood agar, after which erythromycin and clindamycin discs were
119 placed next to each other, 12mm apart. The plates were incubated at 35-37°C in 5% CO₂ for
120 18-24 hours. A "D-zone" on the side of the clindamycin disc facing the erythromycin disc
121 was taken as positive for the MLS_B resistance phenotype.

122 Serotyping was performed using the latex agglutination method as described by Kwatra et al
123 [9].

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125 **Ethics Approval**

126 Ethical approval for this study was obtained from the University of Pretoria Faculty of Health
127 Sciences Research Ethics Committee. The ethics reference number is 393/2013.

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131 **Results**

132 The serotype distribution of the 69 isolates were 54% Ia (n=37), 20% III (n=14), 16% V
133 (n=11), 6% II (n=4), 3% IV (n=2) and 1% Ib (n=1).

134 The antimicrobial susceptibility testing showed 69 (100%) isolates were susceptible to
135 penicillin (MIC range = 0.032-0.125 µg/ml). All isolates were susceptible to vancomycin with
136 13 (18%) isolates having an MIC at the breakpoint (1µg/ml) (MIC range = 0.38-1µg/ml). Sixty
137 (83%) isolates were sensitive to erythromycin, 8 (11%) isolates were intermediate and 1
138 (1%) was resistant (MIC range = 0.094-3µg/ml). The erythromycin intermediate isolates
139 belonged to serotypes Ia (3); III (2); IV (1) and V (2).

140 Thirty (42%) isolates were found to be fully susceptible to clindamycin while 40 (56%) were
141 intermediate-susceptible and no resistant isolates were detected (MIC range = 0.19-0.75
142 µg/ml). The clindamycin intermediate isolates belonged to serotypes Ia (23), II (1), III (8), IV
143 (1) and V (7). Only 3 (4%; serotypes Ia, III and V) of our isolates displayed a positive MLS_B
144 phenotype. All isolates were sensitive to levofloxacin (range = 0.38-1.5 µg/ml).

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153 **Table 1:** Antimicrobial susceptibility of 69 GBS isolates to 5 antimicrobial agents

Antimicrobial agent	MIC ₅₀ (µg/ml)	MIC ₉₀ (µg/ml)	Range (µg/ml)
Penicillin	0.047	0.064	0.032-0.125
Erythromycin	0.19	0.25	0.094-3
Clindamycin	0.25	0.5	0.19-0.75
Vancomycin	0.75	0.5	0.38 -1
Levofloxacin	0.5	0.75	0.38-1.5

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157 Discussion

158 The current study characterized the antimicrobial resistance patterns in GBS isolates from
159 pregnant women. In this study, sixty-nine (100%) isolates were fully susceptible to penicillin.

160 In a recent Chinese study looking at colonising GBS isolates from pregnant women 100% of
161 isolates were sensitive to penicillin, ceftriaxone, linezolid and vancomycin [10]. Longtin et al.

162 described a case of GBS with reduced susceptibility to penicillin emerging after long term
163 suppressive oral penicillin therapy for a prosthetic joint infection [11].

164 All isolates in this study were susceptible to vancomycin. There is a paucity of data on
165 vancomycin resistance in GBS isolates, with 2 case reports. These cases involved 2 patients
166 with invasive GBS infection with significant co-morbidities including diabetes, hypertension,
167 congestive cardiac failure, hypercholestromaemia in 1 patient and end stage renal disease,
168 obesity, cor-pulmonale and chronic osteomyelitis in the 2nd patient [12]. Only one of these
169 patients had previous prolonged exposure to vancomycin. Both isolates were characterised
170 as belonging to serotype II. The vancomycin MIC in both cases were 4ug/ml.

171 Macrolides are often regarded as alternative therapy for penicillin sensitive patients to treat
172 GBS infections, however resistance to macrolides has increased during recent years in
173 several countries with reported geographical variations [13]. In the Japanese study by
174 Matsubara et al (2001) the researchers found much lower rates of resistance to
175 erythromycin and clindamycin, 3% and 1% respectively [14]. In a Malaysian study, 23.3% of
176 isolates were resistant to erythromycin and 17.5% to clindamycin [15]. The prevalence of
177 resistance among invasive GBS isolates in the United States ranged from 25%-32% for
178 erythromycin and 13%-20% for clindamycin in reports published during 2006-2009 [2]. Our
179 data suggests a lower level of resistance to these two agents than those observed in the US
180 and is closer to the Japanese data [2,14].

181 The CLSI recommends MLS_B testing for beta-haemolytic streptococci which tests for
182 inducible clindamycin resistance. It was found to be the main mechanism of resistance in
183 GBS isolates isolated from the vagina as well as gastric fluid and ear specimens in a Tunisian

184 study performed by Hraoui and colleagues [13]. This phenomenon was only noted in 3
185 (4%) of our isolates. The serotype distribution of these isolates were Ia, III and V.

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187 All isolates in this study were susceptible to levofloxacin. However, there have been reports
188 of fluoroquinolone resistant GBS strains that have emerged in the past decade especially in
189 Asia, including China, Japan and Korea [16]. A study by Wu et al (2017) had confirmed that
190 respiratory samples and elderly patients are two independent risk factors associated with
191 levofloxacin resistance in GBS [16]. In addition the study found that levofloxacin-resistant
192 GBS isolates belonged mainly to the ST19/serotype III serogroup [16].

193 In low income settings, safe administration of intravenous antibiotics may not always be
194 affordable or feasible, particularly for settings where births do not occur in hospitals. In
195 addition, IAP has not proven to be effective in preventing LOD [1]. Therefore, new strategies
196 for prevention of GBS disease in neonates needs to be considered. Vaccination targeting
197 pregnant women to subsequently protect neonates against GBS infection is a potential
198 option.

199 Information regarding serotype distribution of GBS strains could guide the development of
200 vaccine candidates. Vaccinating pregnant women against GBS may protect infants from
201 developing invasive GBS disease. Universal screening programs for maternal GBS
202 colonisation followed by IAP in colonised mothers have shown to decrease the incidence of
203 EOD [2]. However, it is thought to have a minimal role in the prevention of LOD. GBS
204 maternal vaccination has the potential to decrease EOD as well impact on LOD.

205 This study showed that serotypes Ia (54%), III (20%) and V (16%) were the predominant
206 serotypes which is in concordance with other studies conducted among pregnant women in
207 South Africa [5]. Serotypes Ia and III together accounted for 74% of the colonised population
208 in our study, whilst the 3 dominant serotypes accounted for 90% of all cases. These results
209 are in keeping with another South African study which showed that serotype III is the
210 commonest cause of EOD in South Africa, accounting for 41.4% of all cases, whilst serotype
211 Ia accounted for 34.7% of cases [5]. The majority of invasive disease was caused by
212 serotypes Ia, III and V [5]. These 3 serotypes are included in a pentavalent polysaccharide
213 protein conjugate vaccine currently being developed and is in a phase 1 trial [17].

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218 **Conclusions**

219 GBS isolates remain susceptible to penicillin and vancomycin, however, surveillance for
220 resistance needs to be ongoing. Macrolides should only be used once susceptibility results
221 are available as significant rates of intermediate resistance have been detected in these
222 isolates. Ninety percent of colonizing isolates belong to 3 serotypes, viz. Ia, III and V.

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226 Acknowledgements

227 I would like to acknowledge Dr Alex Sihlabela from the Department of Obstetrics and
228 Gynaecology at Kalafong Hospital who collected the initial swabs from pregnant women at
229 Kalafong antenatal clinic.

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