A multi-omic cohort as a reference point for promoting a healthy human gut microbiome

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Summary

More than a decade of gut microbiome studies has a common goal for human health. As most of the disease studies sample the elderly or the middle-aged, a reference cohort for young healthy individuals has been lacking. It is also not clear what other omics data need to be measured to better understand the gut microbiome. Here we present high-depth metagenomic shotgun sequencing data for the fecal microbiome together with other omics data in a cohort of ~2183 adults. A number of vitamins, amino acids and trace elements leave their marks on the gut microbiome and cluster with T cell receptor types. Gender and hormones, especially aldosterone have their impacts. Effects of physical fitness, sleeping habits and dairy consumption are also demonstrated in this large multi-omic cohort. Thus, our comprehensive data are poised to advise future study designs and offer support for exercising and yogurt consumption.

INTRODUCTION

The gut microbiome has been implicated in a growing list of complex diseases,

showing great potential for the diagnosis and treatment of metabolic,

autoimmune and neurological diseases as well as cancer. While case-control

studies have been illuminating (Wang and Jia, 2016), recently published studies

have emphasized difficulty in extrapolating to natural cohorts due to location and

ethnicity (Deschasaux et al., 2018; He et al., 2018). So far only the LifeLines

Deep cohort (n=1135) from the Netherlands made use of low-depth

metagenomic sequencing (7.5 million reads per sample) instead of 16S rRNA

gene amplicon sequencing (Zhernakova et al., 2016). Metabolites are more or

less included in gut microbiome studies, but the conclusions usually did not go

beyond short-chain fatty acids (SCFA), amino acids, vitamin B complex or bile

acids. Levels of trace elements such as arsenic have been a health concern

(FDA; USGS), but are unexplored in the microbiome field. Biological sex is a

strong determent for the gut microbiome in mice and livestock (Xiao et al., 2015,

2016; Zhao et al., 2013), but the impact of hormones, especially non-gonadal

hormones on the human gut microbiome remains unclear.

Here we present metagenomic shotgun sequencing data sufficient for high-

resolution taxonomic and functional profiling (86.1 ±23.3 million reads per

sample) of the fecal microbiome in a cohort of 2183 adults, along with

questionnaire data, physical fitness tests, facial skin features, plasma

metabolome and immune repertoire. Trans-omics analyses uncover previously

overlooked measurements such as aldosterone, trace elements, vitamin A that

heavily influence the gut microbiome. This is also to our knowledge the largest

cohort with facial skin data and immune repertoire data.

A recent study casted doubt over the health benefits of yogurt consumption,

concluding that colonization of the yogurt strains was highly variable between

individuals (Zmora et al., 2018). Our large cohort unequivocally showed yogurt

strains, especially *Streptococcus thermophilus* in feces, along with beneficial

effects in cardiometabolic and mental health.

RESULTS

The gut microbiome as a relatively independent dimension for health

Fecal samples were collected during a physical examination, and 2183 samples

were subjected to metagenomic shotgun sequencing, yielding 81.20 ± 24.59

million high-quality non-human reads per sample (Table S1), ensuring accurate

taxonomic and functional profilings. The reads were mapped to a comprehensive

human gut microbiome reference gene catalog containing 9.9 million genes (with

a saturating mapping rate of 80.1 ± 4.9 %) and then assigned to 1507

Metagenomic Species (MGSs) (Li et al., 2014; Moll1 et al., 2019; Nielsen et al.,

2014) and 2981 metagenomic linkage groups (MLGs) (Qin et al, 2012, Jie et al,

2017).

Other omics data, including 104 plasma metabolites (3980 samples), 634

immune indices (VDJ usage and diversity, 4120 samples) from buffy coat, 72

basic medical data (body measurements and routine blood test, 2715 samples),

49 facial skin imaging scores (2049 samples), 18 entries from psychological

questionnaire (2039 samples), and 56 entries from lifestyle questionnaire (3820

samples) were collected from the same individuals (Table S2; S3).

An inter-omics prediction value between omics data was calculated using a 5-fold

cross-validated random forest model (Figure 1A). Basic medical data showed the

highest global systematic association with other omics data, the accuracy of

prediction from basic medical data to physical fitness and metabolites to basic

medical data reaching 75% quantile RFCV R = 0.461 and 0.399, respectively

(Figure 1A, B, Figure S1). Medical test and immune indices showed high

prediction accuracy to metabolites, especially serum creatinine, BMI, waist to hip

ratio, hematocrit, triglyceride in medical test data (Figure 1A, B, C). Metabolites

constituted the most important factor to immune indices (R= 0.292), in particular,

vitamin K made the greatest contribution to prediction, followed by selenium,

argininosuccinic acid, N5-methyltetrahydrofolate and glutamic acid (Figure 1 B,

C). Life style also showed high prediction accuracy to physical fitness conditions

(R= 0.255 Figure 1A, B, C). Most facial skin features can be predicted by or to

predict many omics data type, especially medical test, metabolites and physical

fitness factors (Figure 1B, C). Among the life style questionnaire, smoking,

drinking (especially low concentration alcohol), sports habits (especially

resistance training), high sugar and high fat food diet habit, going to bed late past

0.00 am can be predicted by multiple omics data type (Figure 1B).

A number of factors have been reported to explain gut microbial composition,

while the total percentage of variance explained remained in single digits (Falony

et al., 2016). In our cohort, defecation frequency and gender were the strongest

factors to explain gut microbiome composition, followed by serum uric acid level,

triglyceride, testosterone, estradiol, bust circumference, grip strength, as well as

psychological symptoms and pores and speckles on the cheek, each explaining

more than 0.002 of the variances (PERMANOVA test Figure S2). However, none

of these data explained more than 0.1 (10%) of the variances to the gut

microbiome composition (Figure S2**)**. Gut microbiome was not a strong factor for

other omics either.

These results were confirmed by another RFCV predict model which was a more

efficient model than global PERMANOVA in capturing the specific gut microbes

association with omics data factors (Figure 2). Gut microbe showed prediction

power for plasma levels of vitamins (VA, folic acid, VB5, VD), hormones

(testosterone, aldosterone) and trace elements (mercury, selenium, arsenic) and

amino acids (BCAA, glutamic acid, tryptophan, histidine) in metabolites. Grip

strength, vital capacity, facial skin porphyrins and staying up late past 0.00 am

can also be predicted by the gut microbiome. Influence from other lifestyle

questionnaire factors such as defecation, yogurt, age, gender, smoking, milk,

drinking alcohol, fruit and vegetables were also detected by the RFCV model.

Influence from ABO blood group was also detected (Figure 2), and specific

differences include Lachospiraceae bacterium 3_1_46FAA in blood type A (q =

6.12E-5), *Ruminococcus torques* in blood type B (q = 1.59E-2), unnamed

MGS209 in blood type AB (q = 1.59E-2) and *Megaspaera micronuciformis* in

blood type O (q = 1.69E-2).

Defecation, hormone and gender

Perhaps due to population differences and a younger age of this cohort (29.6 ±

5.5, average ± sev, Table S1), the gender (female 1016, male 1007) effect on the

gut microbiome is clearly evident (Benjamin Hochberg-adjusted p value < 0.05,

Figure S3). The females showed a greater microbial diversity (BH-adjusted

pvalue =1.52e-12). *Fusobacterium mortiferum, Eubacterium dolichum, Blautia*

hansenii, Prevotella stercorea and *Ruminococcus mgs0922* were significantly

more abundant in males (Figure S3), despite the absence of colorectal cancer in

this young cohort. *Bifidobacterium* and its five species (*B. longum*, *B.*

adolescentis, *B. breve*, *B. bifidum*, *B. animalis* and *B.catenulatum-*

B.pseudocatenulatum complex) were all significantly enriched in females (Figure

S3). GABA (γ-aminobutyric acid) production by bacteria including the female-

enriched *Bifidobacterium* and *Bacteroides* could impact on the brain and immune

functions in the gut (Strandwitz et al., 2018). The greater gut microbiome gene

richness in women could also approximate a larger number of cells per gram of

feces (Vandeputte et al., 2017), and influence defecation frequency and

regularity.

In the unstratified cohort, testosterone negatively correlated with *Clostridia*. Aldosterone, one of the major adrenal gland mineralcorticoid, positively correlated with three *Bacteroides* species (*B. intestinalis, B. cellulosilyticus, B. stercorirosoris)* and *Eubacterium eligens*. Estradiol/Estrogen positively correlated with *Alistipes obesi*; pyridoxine (Vitamin B6) positively correlated with *Eubacterium siraeum* and *Eubacterium eligens* (Table S5). In addition, plasma levels of the hormones progesterone, estradiol, and aldosterone positively associated with gut microbial enzymes for secondary bile acids (Figure S4). Genes for producing secondary bile acids have been found in bacteria such as *Lactobacillus*, *Streptococcus* and *Bifidobacterium* (Gu et al., 2017), and could affect defecation as well as metabolism (Figure S4).

Facial skin features reflect systemic health

Facial skin imaging data (percentile score relative to VISIA database population of the same gender and age) could be predicted by other omics such as basic medical test and lifestyle (Figure 1). We found that individuals with a higher level of HDLC and water content scored younger than one's actual age. Distinct patterns were seen for males and females (Figure 3). In males, four of the TCR VDJ usage TRBV6−5:TRBJ2−6,TRBV12−3:TRBJ1-1/TRBJ2−1, and TRBV11−1:TRBJ1−4, together with immune diversity score and serum dehydroepiandrosteron correlated with more porphyrins yet less UV stains and brown spots on the face. In females, dehydroepiandrosteron associated with not only porphyrins but also frontal and lateral pores. For females but not males, a

better score from the sleeping and diet assessment corresponded to less

porphyrins (Figure 3), providing a strong incentive for keeping healthy habits.

Both males and females appeared older than actual age with increased BMI,

systolic blood pressure and metabolites associated with cardiometabolic risk,

especially lead, glutamate, vitamin D, B1 and B5. However, higher selenium and

mercury corresponded to less UV stains in females. In general, the gut

microbiome did not have a lot of associations with facial skin features, except for

one association between an MGS from Oscillospiraceae and less red zone on

the cheeks for females.

The metabolome-immune-gut axis

Among the strongest associations between different omics is that between

immune repertoire and plasma metabolites (Figure 1). More strikingly, when we

plot the associations in detail, the clusters of metabolites corresponded either to

the same *TRBV* (T-cell receptor beta variable gene) or to the same *TRBJ* (T-cell

receptor beta joining gene) (Figure 4). Vitamin A, N5-methyltetrahydrofolicacid,

selenium and mercury and serum aldosterone all showed negative associations

with a few TRBJ1-4, TRBJ2-1 and positive associations with TRBJ2-4. Vitamin

B5, Vitamin E, phosphoserine, arginosuccinic acid and arsenic showed these

associations with TRBJ1-4, TRBJ2, TRBJ2-4, as well as positive associations

with TRBV20-1 and TRBV3-1. Glutamic acid and serine showed a largely

opposite pattern, except for positive associations with TRBV20-1.

When we looked into the gut microbiome, a number of associations could also be identified with the immune repertoire data (Figure 4). Vitamin A is central to a healthy immune system but is typically studied for its role in early development (Veldhoen and Ferreira, 2015). The Clostridiales species likely reduce intestinal level of vitamin A (Figure S5) through modulating expression of retinol dehydrogenase 7 and dampen antimicrobial response in the gut (Grizotte-Lake et al., 2018). Vitamin D derivatives did not show significant associations with fecal MGSs, yet largely followed the pattern of N5-methyltetrahydrofolicacid, which positively associated with the butyrate-producing and atherosclerotic cardiovascular disease-depleted bacterium *Eubacterium eligens* (Jie et al., 2017). N5-methyltetrahydrofolicacid also negatively associated with the formate or hydrogen-producing and obesity-related bacteria including *Dorea longicatena* and *Blautia* (Beaumont et al., 2016; Liu et al., 2017; Taras et al., 2002) (Figure 4), implying that folic acid supplementation might modulate fecal levels of these bacteria. Associations between the gut microbiome and plasma levels of trace elements including mercury, selenium and arsenic might be surprising (Figure 4). Selenium-containing rice is commercially promoted as anti-cancer (Stolz and Oremland, 1999), and we found that the association pattern largely followed arsenic, consistent with these two trace elements' similar function in anaerobic respiration. Selenium and mercury also correlated with disease-associated species such as *Clostridium bolteae*, *Streptococcus oralis* and *Ruminococcus gnavus* in the gut microbiome. Mercury, selenium and arsenic also associated with facial skin features, favoring UV spots versus porphyrins, while lead and

magnisium, along with glutamic acid, aminoadipic acid and vitamin B5 correlated

with red areas (Figure 3**).**

The cluster represented by vitamin B5 associated with the butyrate-producing

and arsenic-metabolizing bacterium, *Faecalibacterium prausnitzii* (Coryell et al.,

2018). *Bacteroides coprophilus*, a prevalent but not very abundant species from

the *Bacteroides* genus, showed the opposite pattern (Figure 4). The third cluster

represented by glutamic acid showed negative associations with previously

reported bacteria implicated with lower BMI such as *Alistipes shahii*, *Bacteroides*

cellulosilyticus, *Ruminococcus lactis* and *Eubacterium eligens* (Jie et al., 2017) in

this large cohort (Figure 4), consistent with higher glutamic acid in obese

individuals (Liu et al., 2017).

Biomarkers for cardiometabolic diseases and colorectal cancer

Hyperuricemia is common in the East Asian population, and urate is excreted in

urine or through the gastrointestinal tract. In our cohort, serum uric acid positively

correlated with vitamin A, B5, D3 and E, high sugar/high fat diet, amino acids

(glutamic acid and alanine) and trace elements (arsenic, mercury and selenium),

while negatively associated with defecation frequency, testosterone and

aldosterone, and beneficial gut bacteria such as *Faecalibacterium prausnitzii*,

Alistipes shahii, *Oscillospiraceae* and *Bacteroides intestinalis* (Figure 5A, Figure

S6A).

We next defined a score according to 8 blood parameters and 80 fecal microbiome features for cardiometabolic disease risk (see methods) and tested it in previously published case-control samples. Fecal metagenomic samples from Chinese patients with atherosclerotic cardiovascular disease (ACVD), liver cirrhosis, obesity and inflammatory bowel diseases all scored higher compared to control samples without the disease, while those from diseases such as colorectal cancer, rheumatoid arthritis and unstratified T2D did not (Figure 5B) (He et al., 2017; Jie et al., 2017; Liu et al., 2017; Qin et al., 2012, 2014; Yu et al., 2017; Zhang et al., 2015b).

This young multi-omic cohort also helps understand colorectal cancer (CRC). The butyrate-producing *E. eligens* positively associated with fruit and vegetable intake, plasma hydroxyproline, while negatively associated with plasma alanine (Figure 5A). Previously reported CRC-enriched bacteria (Feng et al., 2015; Wang and Jia, 2016; Yu et al., 2017; Zeller et al., 2014) *Peptostreptococcus stomatis* associated with leucine, phenylalanine, alanine, tyrosine in the plasma, *Solobacterium moorei* with leucine and tryptophan, *Fusobacterium mortiferum* with glutamic acid; Enterobacericeae including *Escherichia coli, Klebsiella pneumoniae, Enterobacter cloacae, Citrobacter freundii* positively associated with sarcosine, hydroxylysine, branched chain amino acids, tyrosine, tryptophan, 1–methylhistidine, hydroxyproline, and argininosuccinic acid (Figure 5A, Figure

S6B). *Flavonifractor plautii* and *Bilophila wadsworthia* also showed positive

correlation with leucine, 1-methylhistidine and aminobutyric acid (Figure S6B).

Physical fitness, exercising and sleeping reflected in the gut microbiome

Vital capacity, a commonly taken measure for lung function, explained a

considerable fraction of variance in the gut microbiome and associated with

bacteria such as *A. shahii*, *F. prausnitzii* and *Bifidobacterium adolescentis*, while

negatively correlated with cardiometabolic disease-related bacterium *Clostridium*

clostridioforme, *Ruminococcus gnavus* and *E. coli* (Figure 2, Figure 6). Grip

strength, a known protective factor for cardiovascular casualty, negatively

associated with *E. coli* (Figure 6). Vertical jump score negatively associated with

E. coli, while positively associated with *B. cellulosilyticus*, *B. intestinalis*,

Butyrisimonas virosa. Reassuringly, sports activities, intensity and time of

exercising correlated with better scores in such physical fitness tests. Gut

microbiome diversity (Shannon index) associated with favorable scores in most

of the fitness tests (Table S4). *Bacteroides cellulosilyticus* and *B. stercorirosoris*,

which associated with exercise intensity, even correlated with a faster reaction

time (Figure 6).

Besides, those who often stay up after midnight also showed negative

correlations with *Haemophilus parainfluenzae, Veillonella tobetsu*ensis and

hydroxy vitamin D3/D, while positively correlated with *Clostridium phoceensis*,

mercury, selenium, arsenic, vitamin A, hydroxyproline and phosphoserine (Figure

S3, 7). And selenium positively correlated with high level serum uric acid (Figure

S6A). Thus, we recommend going to bed earlier from an omics point of view.

A presumably benign species from yogurt in the healthy microbiome

Besides defecation frequency and gender, yogurt consumption explained a

considerable portion of variances in the gut microbiome (Figure 2). A recent

study casted doubt over the health benefits of yogurt consumption, concluding

that colonization of the yogurt strains was highly variable between individuals

(Zmora et al., 2018). In our large cohort, *Streptococcus thermophilus*, a species

included in commercial yogurt mainly for its thermal stability and metabolic

support for other strains, was consistently detected in yogurt eaters, and scaled

with self-reported frequency of yogurt consumption (Figure 7A, D).

Bifidobacterium animalis, probably representing the star strain from CHR

HANSEN, *B. animalis* subsp. lactis BB-12, was also enriched in yogurt eaters

(Figure 7A). In contrast, there was no significant increase in any *Lactobacillus*

strains. The increase in *S. thermophilus* and *B. animalis* by taking yogurt at least

reduced the amount of *Clostridium bolteae*, which was known to be elevated in a

number of cardiometabolic diseases (Wang and Jia, 2016).

A closer look into the host metadata revealed that yogurt consumption was

associated with a number of favorable measurements such as a higher HDL

(high-density lipoprotein) cholesterols, lower triglycerides, better elastic thinking,

less depression and lower serum vitamin A (Figure 7A). Consistently, fecal

relative abundance of *S. thermophilus* associated with a lower diastolic blood

pressure, lower indirect bilirubin, lower hematocrit (Figure 7C). Other intriguing

effects included facial skin improvements seen as less redness at the front and

less porphyrins; and metabolomic changes such as higher estradiol, proline,

lower glycine and ornithine (Figure 7A).

Regarding increasing *Bifidobacterium* in the gut microbiome, however, taking

milk was more effective than yogurt in this Chinese cohort (Figure 7A, B),

implying that some of the beneficial effects come from the exogenous strains in

yogurt. The changes in *Blautia* species due to milk consumption may affect

methane metabolism in the gut and susceptibility to inflammatory bowel diseases

or irritable bowel syndrome (Halkjær et al., 2018; Manor and Borenstein, 2017).

DISCUSSION

In summary, our trans-omic investigation of thousands volunteers establish an

unprecedented reference data set for the healthy gut microbiome. A number of

factors in circulation crosstalk with the gut microbiome, and then manifest on the

face, in the heads and in fitness tests. Levels of trace elements, such as heavy

metals, as important cofactors for bacteria respiration and other functions, should

be measured even in uncontaminated regions. The TCR usage in such a large

cohort is a great resource for discovering microbial antigens other than those

from traditional pathogens.

Potential influence of physical activity on the gut microbiota has been analyzed in

rugby athletes (Barton et al., 2017) and colorectal cancer (Feng et al., 2015).

Compliance to recordings such as Fitbit is notoriously bad in healthy individuals

(Price et al., 2017). Results from this large cohort at least suggest that exercising

might help improve cardio-pulmonary function (grip strength, vital capacity) to

decrease incidence of cardiometabolic diseases.

Our large-scale analyses provide substantial support for health benefits of yogurt

consumption. The universally present species were *Streptococcus thermophilus*

and to a lesser extend *Bifidobacterium animalis* instead of commonly tested

probiotics from *Lactobacillus*. Yet, all of these live or dead probiotics could

nonetheless exert functions on the immune system or even the brain. Faced with

skyrocketing incidences of metabolic diseases, we have yogurt as an

inexpensive option.

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AUTHOR CONTRIBUTIONS

J.W. initiated the overall health project. Y.Z., H.Z., X.X. organized the sample collection, with immune repertoire from X.L., W.Z, serum metabolomics from W.R., Y.L., D.Z., X.Q., X.C., metagenomic profile from Q.D, X.W., and J.Z., Q.D., S.T., Y.L., D.W. checked the host metadata and matched the omics data. Z.J. led the bioinformatic analyses, including S.L. and F.L. H.Z., Q.D., S.T., D.W. performed early analyses which did not make it into the manuscript. Z.J., H.J. interpreted the data. H.J., S.L., Z.J. wrote the manuscript. All authors contributed to finalizing this manuscript.

COMPETING FINANCIAL INTERESTS

The authors declare no competing financial interest.

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FIGURE LEGENDS

Figure 1. Overview of the interrelationship between omics.

(A) Global association strength between omics datasets. Each arrow is a 5-fold

cross-validation random forest (RFCV) prediction. The direction of the arrow

indicated the direction of prediction, used the source omics dataset to predict the

target dataset. The darkness and size of the arrow lines indicated 75% quantile

of spearman's correlation between measured value and 5-fold cross-validation

RFCV predicted value (RFCV R) (B) Detailed predict power of source omics for

each target omics. Tick label in x-axis is source omics. Title in top is target omics.

Each node in box is a target factor. The color of the node and box line indicated

the source omics data type. The fill color of box indicated blue means predict

using all samples, yellow means used female samples only, and purple means

used male samples only. Y-axis is the target factor RFCV R predicted from

source omics in x-axis tick label. (C) Common representative factors that could

be strongest predicted by multiple omics data type. Y-axis tick label is source

omics. Title is target omics. X-axis tick label is common representative factors

(target factors). The cell color in heat map indicated the RFCV R using the omics

data in y-axis to predict each factor in x-axis.

Figure 2. Factors associated with gut microbiome.

Top 45 factors with RFCV R > 0.1 in each type of omics that are predicted by gut

microbiome. The length of the bar indicated the rank RFCV R using all samples

and the color indicated the rank of max of RFCV R using male or female samples

independently.

Figure 3. The association of facial skin imaging.

The omics factors association with facial skin is sex specific using multi-omics

analyses method (see methods). The cell color of heat map indicated the

spearman correlation. +, BH-adjusted pvalue<0.1; *, BH-adjusted pvalue <0.05;

**, BH-adjusted pvalue <0.01.

Figure 4. The metabolome-immune-gut axis

Immune repertoire is clustered into 3 groups by fecal microbiome and plasma

metabolites. Factors are selected using multi-omics analyses method (see

methods). The cell color of triadic heatmap shows the partial spearman

correlation adjust age and gender. BH-adjusted p-value is denoted: +, q-

value<0.1; *, q-value<0.05; **, q-value<0.01.

Figure 5. Gut microbial markers for hyperuricemia and other

cardiometabolic diseases.

(A) Association map of the four-tiered analyses integrating the metabolites,

clinical indices, life style and the fecal microbiome. The color of heat map shows

the partial spearman correlation adjusted age and gender. BH-adjusted p-value

is denoted: +, q-value<0.1; *, q-value<0.05; **, q-value<0.01. (B) Microbial

metabolic syndrome risk index was computed for eight additional cardiometabolic

cohort. The box depicts the interquartile range (IQR) between the first and third

quartiles (25th and 75th percentiles, respectively) and the line inside denotes the

median, whereas the points represent the microbial metabolic syndrome risk

index in each sample

Figure 6. Gut microbiome associated with physical fitness and exercise.

The color of heat map shows the partial spearman correlation adjusted age, gender and BMI. BH adjusted p-value is denoted: **+**, q-value<0.1; *, q-value<0.05; **, q-value<0.01

Figure 7. Influence of yogurt and milk intake on omic.

(A-C) The top 5 factor in each omics data associated with yogurt, milk intake habit and the *Streptococcus thermophiles* abundance. The length of the bars represents partial spearman's correlation coefficient adjusted age, gender.

BH adjusted p-value is denoted: **+**, q-value<0.1; *, q-value<0.05; **, q-value<0.01; ***, q-value<0.001; ****, q-value<0.0001. (D) Fecal relative abundance of *S. thermophilus* in volunteers with increasing frequency of yogurt consumption.

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Study Cohort

All the >2000 volunteers were recruited between May 2017 and July 2017 during

physical examination in BGI-Shenzhen, China, the samples in each omics are

shown in Table S3. Baseline characteristics of the cohort are shown in Table S2.

The study was approved by the Institutional Review Boards (IRB) at BGI-

Shenzhen, and all participants provided written informed consent at enrolment.

METHODS DETAILS

Demographic Data Collection

The lifestyle questionnaire contained age, disease history, eating and exercise

habits and so on. The psychological questionnaire contained the evaluation of

irritability, dizziness, frustration, fear, appetite, self-confidence, resilience and so

on (Table S2).

Samples Collection

Fecal samples were self-collected by volunteers, using a kit containing a room

temperature stabilizing reagent to preserve the metagenome (Han et al., 2018).

The samples were frozen on the same day. The overnight fasting blood samples

were drawn from a cubital vein of volunteers by the doctors.

DNA extraction and metagenomics shotgun sequencing

DNA extraction of the stored fecal samples within the next few months was

performed as previously described (Qin et al., 2012). Metagenomic sequencing

was done on the BGISEQ-500 platform (100bp of singled-end reads for fecal

samples and four libraries were constructed for each lane) (Fang et al., 2018).

Amino Acid Measurements

40 µl plasma was deproteinized with 20 µl 10% (w/v) sulfosalicylic acid (Sigma)

containing internal standards, then 120 µl aqueous solution was added. After

centrifuged, the supernatant was used for analysis. The analysis was performed

by ultra high pressure liquid chromatography (UHPLC) coupled to an AB Sciex

Qtrap 5500 mass spectrometry (AB Sciex, US) with the electrospray ionization

(ESI) source in positive ion mode. A Waters ACQUITY UPLC HSS T3 column

(1.8 µm, 2.1 × 100 mm) was used for amino compound separation with a flow

rate at 0.5 ml/min and column temperature of 55 °C. The mobile phases were (A)

water containing 0.05% and 0.1% formic acid (v/v), (B) acetonitrile containing

0.05% and 0.1% formic acid (v/v). The gradient elution was 2% B kept for 0.5

min, then changed linearly to 10% B during 1 min, continued up to 35% B in 2

min, increased to 95% B in 0.1 min and maintained for 1.4 min. Multiple Reaction

Monitoring (MRM) was used to monitor all amino compounds. The mass

parameters were as follows, Curtain gas flow 35 L/min, Collision Gas (CAD) was

medium, Ion Source Gas 1 (GS 1) flow 60 l/min, Ion Source Gas 2 (GS 2) flow 60

l/min, IonSpray Voltage (IS) 5500V, temperature 600 °C. All amino compound

standards were purchased from sigma and Toronto research chemical (TRC).

Hormone Measurements

250 µl plasma was diluted with 205 µl aqueous solution, For SPE experiments, HLB (Waters, USA) was activated with 1.0 ml of dichloromethane, acetonitrile, methanol, respectively and was equilibrated with 1.0 ml of water. The pretreated plasma sample was loaded onto the cartridge and was extracted using gravity. Clean up was accomplished by washing the cartridges with 1.0 ml of 25% methanol in water. After drying under vacuum, samples on the cartridges were eluted with 1.0 ml of dichloromethane. The eluted extract was dried under nitrogen and the residual was dissolved with 25% methanol in water and was transferred to an autosampler vial prior to LC−MS/MS analysis. The analysis was performed by UHPLC coupled to an AB Sciex Qtrap 5500 mass spectrometry (AB Sciex, US) with the atmospheric pressure chemical ionization (APCI) source in positive ion mode. A Phenomone Kinetex C18 column (2.6 µm, 2.1 × 50 mm) was used for steroid hormone separation with a flow rate at 0.8 ml/min and column temperature of 55 °C. The mobile phases were (A) water containing 1mM Ammonium acetate, (B) Methanol containing 1mM Ammonium acetate. The gradient elution was 25% B kept for 0.9min, then changed linearly to 40% B during 0.9min, continued up to 70% B in 2 min, increased to 95% B in 0.1 min and maintained for 1.6 min. Multiple Reaction Monitoring (MRM) was used to monitor all steroid hormone compounds. The mass parameters were as follows, Curtain gas flow 35 l/min, Collision Gas (CAD) was medium, Ion Source Gas 1 (GS 1) flow 60 l/min, Ion Source Gas 2 (GS 2) flow 60 l/min, Nebulizer Current (NC) 5, temperature 500 °C. All steroid hormone profiling compound standards were purchased from sigma, Toronto research chemical (TRC), Cerilliant and DR. Ehrenstorfer.

Trace element Measurements

200 µl of whole blood were transferred into a 15 mL polyethylene tube and diluted 1:25

with a diluent solution consisting of 0.1% (v/v) Triton X-100, 0.1% (v/v) HNO_3,2mg/L AU,

and internal standards (20 µg/L). The mixture was sonicated for 10min before ICP-MS

analysis. Multi-element determination was performed on an Agilent 7700x ICP-MS

(Agilent Technologies, Tokyo, Japan) equipped with an octupole reaction system (ORS)

collision/reaction cell technology to minimize spectral interferences. The continuous

sample introduction system consisted of an autosampler, a quartz torch with a 2.5-

mmdiameter injector with a Shield Torch system, a Scott double-pass spray chamber

and nickel cones (Agilent Technologies, Tokyo, Japan). A glass concentric MicroMist

nebuliser (Agilent Technologies, Tokyo, Japan) was used for the analysis of diluted

samples.

Water-soluble Vitamins Measurements

200 µl plasma were deproteinized with 600 µl methanol (Merck), water, acetic acid

(9:1:0.1) containing internal standards, thiamine-(4-methyl-13C-thiazol-5-yl-13C3)

hydrochloride (Sigma-Aldrich), levomefolic acid-13C, d3, riboflavin-13C,15N2, 4-

pyridoxic acid-d3 and pantothenic acid-13C3,15N hemi calcium salt (Toronto Research

Chemicals). 500 µl supernatant were dried by nitrogen flow. 60 µl water were added to

the residues, vortexed, the mixture was centrifuged and the supernatant was for analysis.

The analysis was performed by UPLC coupled to a Waters Xevo TQ-S Triple Quad

mass spectrometry (Waters, USA) with the electrospray ionization (ESI) source in

positive ion mode. A Waters ACQUITY UPLC HSS T3 column (1.7 µm, 2.1 × 50 mm)

was used for water-soluble vitamins separation with a flow rate at 0.45 ml/min and

column temperature of 45 °C. The mobile phases were (A) 0.1 % formic acid in water, (B)

0.1% formic acid in methanol. The following elution gradient was used: 0–1 min,99.0%–

99.0% A; 1–1.5 min, 99.0% A–97.0% A; 1.5–2 min, 97.0% A–70.0% A,2–3.5 min, 70%

A–30% A; 3.5–4.0 min, 30%A–10.0%A; 4.0–4.8 min, 10%A–10.0%A; 4.9–6.0 min,

99.0%A–99.0%A. Multiple Reaction Monitoring (MRM) was used to monitor all water-

soluble vitamins. The mass parameters were as follows, the capillary voltages of 3000V

and source temperature of 150°C were adopted. The desolvation temperature was

500°C. The collision gas flow was set at 0.10 ml/min. The cone gas and desolvation gas

flow were 150 l/h and 1000 l/h, respectively. All water-soluble vitamins standards were

purchased from Sigma-Aldrich (USA).

Fat-soluble Vitamins Measurements

250 µl plasma were deproteinized with 1000 µl methanol and acetonitrile, (v/v,1:1)

(Fisher Chemical) containing internal standards, all-trans-Retinol-d5, 25-

HydroxyVitamin-D2-d6, 25-HydroxyVitamin-D3-d6, vitamin K1-d7, α-Tocopherol-d6

(Toronto Research Chemicals). 900 µl supernatant were dried by nitrogen flow. 80 µl 80%

acetonitrile were added to the residues, vortexed, the mixture was centrifuged, and the

supernatant was used for analysis. The analysis was performed by UPLC coupled to an

AB Sciex Qtrap 4500 mass spectrometry (AB Sciex, USA) with the atmospheric

pressure chemical ionization (APCI) source in positive ion mode. A Waters ACQUITY

UPLC BEH C18 column (1.7 µm, 2.1 × 50 mm) was used for fat-soluble vitamins

separation with a flow rate at 0.50 ml/min and column temperature of 45 °C. The mobile

phases were (A) 0.1 % formic acid in water, (B) 0.1% formic acid in acetonitrile. The

following elution gradient was used: 0–0.5 min,60.0%–60.0% A; 0.5–1.5 min, 60.0% A–

20.0% A; 1.5–2.5 min, 20.0% A–0% A,2.5–4.5 min, 0% A–0% A; 4.5–4.6 min, 0%A–

60.0%A; 4.6–5.0 min, 60.0%A–60.0%A. Multiple Reaction Monitoring (MRM) was used

to monitor all fat-soluble vitamins. The mass parameters were as follows, Curtain gas

flow 30 l/min, Collision Gas (CAD) was medium, Ion Source Gas 1 (GS 1) flow 40 l/min,

Ion Source Gas 2 (GS 2) flow 50 l/min, Nebulizer Current (NC) 5, temperature 400 °C.

All fat-soluble vitamins standards were purchased from Sigma-Aldrich (USA), Toronto

research chemical (TRC).

Immune indices Measurements

10 ml whole blood was centrifuged at 3,000 r/min for 10 min, then 165 µl buffy

coat were obtained to extract DNA using MagPure Buffy Coat DNA Midi KF Kit

(Magen, China). The DNA was sequenced on the BGISEQ- 500 platform using

200 bp singled-end reads. The data processing was performed using Immune

IMonitor(Zhang et al., 2015a).

Medical Parameters

All the volunteers were recruited during the physical examination. The medical

test including blood tests, urinalysis, routine examination of cervical secretion

and so on. All the medical parameters were measured by the physical

examination center and shown in Table S2.

Facial Skin feature

The volunteer's frontal face without makeup was photographed by VISIA-CR™

imaging system (Canfield Scientific, Fairfield, NJ, USA) equipped with chin

supports and forehead clamps that fix the face during the photographing process

and maintain a fixed distance between the volunteers and the camera at all times.

Eight indices were obtained including spots, pores, wrinkles, texture, UV spots,

porphyrins, brown spots and red area from the cheek and forehead, respectively

(Table S2). The percentile of index was calculated based on the index value

ranked in the age-matched database, the higher the better (Table S2).

Physical fitness test

8 kinds of tests were performed to evaluate volunteers' physical fitness condition

(Table S2). Vital capacity was measured by HK6800-FH (Hengkangjiaye, China).

Eye-closed and single-legged standing was measured by HK6800-ZL. Choice

reaction time was measured by HK6800-FY. Grip strength was measured by

HK6800-WL. Sit and reach was measured by HK6800-TQ. Sit-ups was

measured by HK6800-YW. Step index was measured by HK6800-TJ. Vertical

jump was measured by HK6800-ZT. We got a measure value from each test.

Then each measure value score was assigned grades A through E based on its

corresponding age-matched database.

QUANTIFICATION AND STATISTICAL ANALYSIS

Quality control, Taxonomic annotation and abundance calculation

The sequencing reads were quality-controlled as described previously(Fang et al.,

2018).

Taxonomic assignment of the high-quality fecal metagenomic data was

performed using the reference gene catalog comprising 9,879,896 genes(Li et

al., 2014). Taxonomy of the fecal MGSs/MLGs were then determined from their

constituent genes, as previously described(Moll1 et al., 2019; Nielsen et al., 2014;

Qin et al., 2012; Wang and Jia, 2016).

Gut microbiota composition explained by factors in each type of omics.

To assess the effect of each factor in each type of omics on gut microbiome, we performed PERMANOVA (Adonis) test on the MGS/MLG abundance profiles using euclidean distance and 999 permutations in the vegan R package. Factors with BH adjusted $P<0.05$ were considered to associate with MGS/MLG (Figure S2B).

The global effect size between fecal microbiome and omics data.

The cumulative contribution of metadata variables/omics data lists was determined by forward stepwise redundancy analysis on MGS profile with the forward.sel function in packfor R package with variables that was sub-selected by envfit function in vegan (Figure S2A).

The factors in each type of omics predicted by fecal microbiome

The factors in each type of omics were regressed against the relative abundances of mgs profile (found in at least 10% of the samples) in the fecal samples using default parameters in the RFCV function from randomForest package in R. All the categorical variables were converted into continue variables, and nominal variables were converted into dummy variables. RFCV R defined as spearman's correlation between measured value and 5-fold cross-validation predicted value was calculated, then rank the top 5 predictable factors in each omics type. The same prediction process was done between any two types of omics. Then ggplot package in R was used to boxplot predict power of target omics factors by all kinds of other predictor omics (Figure 1B). 75% quantile RFCV R between any two types omics (from a to b and from b to a) was used to

construct the bi-direction global omics correlation network using CytoScape

(Figure 1A). R package pheatmap and barplot was used to make heatmap plot

for some representative factors (Figure 1C, Figure 2, Figure S1).

Robust association network construction between any two omics data type
including fecal microbial MGSs

An rank average method(Marbach et al., 2012) was used to combine the results

of multiple inference methods to make a robust omics association network. In

our study, we consider to combine variance importance from randomForest that

was an all vs one non-linear regression model and spearman's correlation that

was a one vs one non-linear model.

Step 1: data prepare.

All categorical variables in other omics data were converted into continuous

variables, and nominal variables were converted into dummy variables. Missing

values were filled with median and variables with more than 70% missing were

dropped. The microbial species less than10% in all the samples were also

removed. Near zero variance variables was removed. Outliers were defined as

outside of the 95% quartiles and removed.

Step 2: multiple inference methods perform.

Random forest variable importance was used to identify the most important

predictor variables(Louppe et al., 2013). RFCV regression function from

randomForest package in R with default parameter was used to get the 5-fold

average variable importance. Spearman's correlation with the cor.test function in

base R software. Partial spearman's correlation with ppcor package.

Step 3: Robust networks construction.

The average rank (source factors -target factors edge from any two different

omics) was computed as sum of the ranks across the RFCV importance and

spearman rho. For the scales of variables, the RFCV importance was rank within

each target factor in each omics. For each omics type, we just kept the top 20

target factors with highest RFCV R. Second we kept top 30 average ranks for

each target variable and retained edges with spearman's correlation BH-adjusted

pvalue <0.05. For example, metabolites as target and gut microbe as source.

We regressed gut microbes against the metabolites and compute the 5-fold cross

validation predict power (RFCV R) for each metabolites and spearman

correlation between all gut microbes and all metabolites. 20 of 107 metabolites

with highest RFCV were kept. For each of the 20 select metabolites such as VA,

average ranks across RFCV and spearman were done. At last gut microbe

biomarker for VA was found with average rank top 30th and passed the spearman

BH-adjusted pvalue <0.05.

Step 4: network visualize.

Top N average ranks for each target as representative factors was used to make

barplots using ggplot2 package (Figure 2,7). The pheatmap package was used to

plot the common representative factors that could be strongest predicted by

multiple omics data type(Figure 1C). All the source-target factors pair RFCVR (a

as source, b as target and b as source, a as target) was boxplot (Figure1B)

using ggplot2. The ComplexHeatmap package in R was used to plot omics triadic

relation (Figure 3-6). CytoScape was also used to visualized the global omics

network (Figure 1A). R package igraph was used to plot omics factors network

that are all significant correlate to serum uric acid (Figure S6A).

Microbial metabolic syndrome risk index validation in cardiometabolic
cohort.

Using multi-omics analyses method, based on the current cohort we pick up 80

MGS significant correlate with one of the eight cardiometabolic risk factors

(Waist Hip Ratio, BMI, Triglyceride (mmol/L),High-Density-Lipoprotein

(mmol/L),Serum Uric Acid (μ mol/L), γ -Glutamyl Transpeptidase (U/L),Serum

Alanine Aminotransferase(U/L),Fasting Blood Sugar (mmol/L)). 12 of 80 MGS

was defined as bad cardiometabolic microbe for they were positively correlate

with BMI and Triglyceride. The rest 68 MGS was defined as good with

cardiometabolic microbe who are negatively correlate with BMI and Triglyceride.

For each MGS and each validation chinese disease cohort samples (ACVD, CD,

UC, HongKong CRC, Cirrhosis, Obesity, RA, T2D) we get the percentage rank

based onthe current cohort as reference. Then microbial metabolic syndrome risk

index for new sample computed as average bad cardiometabolic microbe

percentage rank minus average good cardiometabolic microbe percentage rank.

Rank sum test was done between the disease and healthy group to test the

microbial metabolic syndrome risk index performance in validation disease cohort.

DATA AND SOFTWARE AVAILABILITY
Metagenomic sequencing data for all samples have been deposited to the

(CNGB) database under the accession code CNPxx.

A Yogurt eating habits

B Milk Habits

C Streptococcus thermophilus LMD−9 MGS0852

D

Fecal microbiome | Medical test | Metabolites | Physical fitness test | Psychological questionnaire | Facial skin feature