COMICS: A pipeline for the composite identification of selection across multiple genomic scans using Invariant Coordinate Selection in R

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Running Head: composite identification of selection signals

Abstract

Identifying loci that are under selection versus those that are evolving neutrally is a common

challenge in evolutionary genetics. Moreover, with the increase in sequence data, genomic

studies have begun to incorporate the use of multiple methods to identify candidate loci under

selection. Composite methods are usually implemented to transform the data into a multi-

dimensional scatter where outliers are identified using a distance metric, the most common being

Mahalanobis distance. However, studies have shown that the power of Mahalanobis distance

reduces as the number of dimensions increases. Because the number of methods for detecting

selection continue to grow, this is an undesirable feature of Mahalanobis distance. Other

composite methods such as invariant coordinate selection (ICS) have proven to be a robust

method for identifying outliers in multi-dimensional space; though, this method has not been

implemented for genomic data. Here we use simulated genomic data to test the performance of

ICS in identifying outlier loci from multiple selection scans and compare the results to the

performance of Mahalanobis distances. We show that the ICS out performed Mahalanobis

distance in all aspects including false positives, false negatives, and recall. Furthermore, ICS also

performed better when identifying loci with weaker selection coefficients. We also introduce a

pipeline in a R-Shiny smart wrapper environment that implements the ICS on multiple scans of

selection. Importantly, we show that the ICS is a robust method for identifying outliers in multi-

dimensional space and recommend its use for studies aimed at identifying loci under selection in

the genome.

Introduction

A main goal of evolutionary genetics and ecological genomics is to identify candidate regions of the genome that show patterns consistent with selection (Beaumont and Balding 2004; Campbell-Staton et al. 2016; Forester et al. 2016; Martins et al. 2016). Highly differentiated loci are often interpreted as regions of the genome under selection, potentially due to local adaptation of a population to a novel or changing environment (Beaumont and Balding 2004; Campbell-Staton et al. 2016; Bekkevold et al. 2016; Rellstab et al. 2016). With the improvement of sequencing methods, the quantity and quality of genetic data continues to increase, allowing a more rigorous analysis of highly differentiated loci and how they are distributed throughout the genome. Despite this increase in genetic data methods to identify putatively selected loci (Gunther and Coop 2013; Lotterhos and Whitlock 2015; Capblancq et al. 2018), identifying candidate loci under selection and differentiating between true outliers and false positives still proves to be a difficult task.

Many selection methods/scans include the analysis of single nucleotide polymorphisms (SNPs), where each SNP is treated as an independent hypothesis when tested against the rest of the genome (Nielsen et al. 2005; Lotterhos and Whitlock 2014; Lotterhos and Whitlock 2015). The increase in SNP data ultimately decreases our ability to differentiate between neutrally and selectively evolving loci, because of concomitant increase in false positives as the amount of data increases. This problem is also present when referring to methods that use discrete windows containing multiple SNPs to detect overall deviations in the pattern of variation in local regions when compared to the rest of the genome (Sabeti et al. 2002; Nielsen et al. 2005; Sabeti et al. 2007; Chen et al. 2010; Zhong et al. 2011; Alachiotis et al. 2012; Pavlidis et al. 2013). To circumvent this issue, some studies have implemented the use of multiple selection methods to

identify regions of interest and combine them in a meaningful way (*Anopheles gambiae* 1000

Genomes Consortium 2017; Zueva et al. 2018; Hodel et al. 2018). For example, a common

method is to only define genomic regions as under selection if they are identified as outliers by

multiple selection methods. Because of the nuances across selection methods and the differences

in the assumptions and types of signals, the number of intersections identified may underestimate

the true number of regions under selection, increasing the false negative rate and making this

approach very conservative. For instance, methods that detect selection using the site frequency

spectrum (SFS) identify a selective sweep as an area with higher than expected levels of allele

differentiation (Nielsen et al. 2005), while methods using linkage disequilibrium identify

selective sweeps as regions with larger than expected associations among SNPs (Alachiotis et al.

2012). When looking at the intersection between these methods, regions under moderate

selection near high levels of recombination would most likely be considered neutral.

In lieu of looking at the intersection across multiple selection methods to identify outlier

loci, composite methods should be implemented. A composite method includes the use of

multiple selection statistics to identify regions of the genome under selection (Verity et al. 2017;

Capblancq et al. 2018). In this case, each method/statistic will result in a distribution that is

described by its location and dispersion. A multi-dimensional transformation creates a cluster of

points, derived from the distributions of the selection/population statistics, where the location is

defined as the center of the cluster and the spread of the data are defined as the scatter (Verity et

al. 2017; Capblancq et al. 2018; Archimbaud, Aurore, Nordhausen, Ruiz-Gazen 2018a).

Moreover, multidimensional analyses create ample opportunity to take different selection metrics

and compare them linearly under one distribution (Verity et al. 2017). In the case of multi-

dimensional data, most outliers are identified by a distance metric, where they are expected to

have a larger than average distances from the central location of the cluster (Archimbaud,

Aurore, Nordhausen, Ruiz-Gazen 2018a).

A popular composite statistic recently proposed to identify outliers across multiple tests

of selection is the Mahalanobis distance, a measure of the number of standard deviations that a

given data point is from the multi-dimensional mean (Mahalanobis 1936; Gnanadesikan and

Kettenring 1972; De Maesschalck et al. 2000; Verity et al. 2017; Archimbaud, Aurore,

Nordhausen, Ruiz-Gazen 2018a). Though the Mahalanobis distance has been implemented for

identifying outlier loci in a multi-dimensional data set, recent studies have shown that an

increase in the number of dimensions (the number of attributes within a data set) results in an

inflation of false negatives and false positives (Leys et al. 2018; Archimbaud, Aurore,

Nordhausen, Ruiz-Gazen 2018a). Specifically, results from Archimbaud et al. (2018) show that

if outliers belong to a reduced dimension space, the probability that the Mahalanobis distance of

a neutral locus exceeds the distance of a true outlier is low; however, this is not the case when

dimension space is increased. In other words, as the number of selection methods used increases,

it becomes more difficult to differentiate between true positives and true-negatives. This

phenomenon is known as the "curse of dimensionality," (Trunk 1979; Bellman, Richard 2013;

Bellman, Richard E. 2015).

We propose the use of invariant coordinate selection (ICS) as an alternative method to

combine multiple selection scans and identify true outliers in growing multi-dimensional space.

Here, we also present a pipeline to easily implement the method. Specifically, ICS is a method

for identifying outlier data points in multi-dimensional space with coordinates derived from

eigenvalues and eigenvectors (Tyler et al. 2007; Archimbaud, Aurore, Nordhausen, Ruiz-Gazen

2018a). The ICS is analogous to a principal component analysis (PCA), however, it differs in

two ways. First, instead of using one scatter matrix (the covariance matrix commonly used in

PCA), ICS adds further constraints by solving inequality conditions on a second matrix with

either the third moment (skewness) or the fourth moment (kurtosis) (Tyler et al. 2007;

Archimbaud, Aurore, Nordhausen, Ruiz-Gazen 2018a). Second, the invariant components are

maintained in parallel while principal components are orthogonal (Tyler et al. 2007;

Archimbaud, Aurore, Nordhausen, Ruiz-Gazen 2018a). Once the univariate data are transformed

into a multi-dimensional data set, data points with the largest ICS distance (i.e., the Euclidean

distance) in the transformed space are deemed as outliers. Importantly, because of the

information drawn from the two different scatter matrices, ICS methods are less susceptible to

the inflation of distance with an increase in the number of dimensions (Tyler et al. 2007;

Archimbaud, Aurore, Nordhausen, Ruiz-Gazen 2018a).

ICS has previously been identified as a robust method for identifying outlier in

multidimensional space and has been implemented to detect subgrouping within populations

based on genotypic data, however, it has not been implemented for identifying outlier regions

based on the input of selection scans. (Archimbaud, A. et al. 2016; Fischer et al. 2017;

Archimbaud, Aurore, Nordhausen, Ruiz-Gazen 2018b). We use simulated genetic data to

compare the number of false positives, false negatives, and recall between Mahalanobis and ICS

distances. We also introduce an easy to use R-Shiny wrapper specifically designed for

performing ICS using genomic data. We call this app COMICS (**C**alling **O**utlier loci from **M**ulti-

dimensional data using **I**nvariant **C**oordinate **S**election) and make it available at:

https://github.com/JTNelsonWSU/COMICS.

Results

The COMICS R-package

COMICS was implemented with the R-Shiny environment, which is an open source R-

package that provides a framework for building web applications using R (Chang et al. 2015;

Team 2018). Specifically, the Shiny environment allows for the development of a graphical user

interface (GUI) that permits the user to operate the ICS proficiently. Using the COMICS

package, the user is able to analyze data in several different ways. First, COMICS has the option

to observe the data before the multi-dimensional transformation; this feature allows the user to

view the results from the different selection scans used to generate the multi-dimensional data in

individual plots. COMICS allows for the customization of the end results of the ICS

identification of outliers by modifying statistical cutoffs, and specific chromosome intervals.

COMICS allows the user to download the multi-dimensional data frame and corresponding

figures in csv and pdf formats. The pipeline is user friendly while maintaining flexibility in the

implementation of the method. You can find the COMICS app and example files at:

https://github.com/JTNelsonWSU/COMICS.

Data upload and input file format

COMICS requires two input data files. The first file is a multivariate data file containing

a list of each genomic position with the corresponding likelihood estimate, or effect size

estimate, from each scan of selection used. The first two columns contain the chromosome and

physical position along the chromosome (or equivalent). COMICS can handle SNP or window

analyses, as long as the windows are consistent across scans of selection; in the latter case, the

position will correspond to the midpoint of the window. The physical positions need to be

numerically organized relative to only the corresponding chromosome; COMICS is designed to

convert all physical positions to the length of the genome using data stored in the genome

configuration file. The second data file is known as the genome configuration file. Specifically,

the configuration file is used to define the number of chromosomes (or scaffolds) used, including

the total length of each chromosomes in base pairs. The main role of this file is to tell COMICS

how large the genome (or assembly) is and the order in which the positions align along the

genome; this is taken into account when COMICS creates genomic scans.

COMICS features and figure generation

In addition to customizing statistical thresholds (e.g., quantiles of interest), COMICS will

generate several different figures for both single statistics and ICS distances. For example, two

different figures are generated from the multivariate data prior to the analysis. The first is a

scatter plot generated by the ggplot and reshape R-packages (Wickham 2007; Wickham 2016),

consolidating all genome scans into one panel and color codes them by chromosome (Figure 1).

The second figure is user defined representation of one particular scan of selection at a time.

After analyses with ICS, COMICS will produce two histograms of the log ICS distance for 1) the

entire genome and 2) a user defined chromosome. Each of these has a vertical line that defines a

cutoff based on the quantile of interest. The main figure that COMICS produces is a genome

scan of the logged ICS Euclidean distance. Like the univariate genome scans, the ICS genome

scan is color coded by chromosome and has a horizontal line representing the statistical cutoff

(Figure 2.). Another feature of the COMICS package is the ability to download the data frame

generated from the ICS. Specifically, the output file contains six different columns, including the

chromosome, the SNP position (or midpoint position in a window), outlier status (Boolean

form), ICS Distance, logged ICS, and the position with respect to the chromosome. Importantly,

the downloadable content in COMICS allows for the flexibility of both downstream analyses of

multi-dimensional data and novel figure generation.

 Testing the ICS with genomic data

Since the development of the ICS, studies have shown that it is a robust method for

detecting outliers in a multidimensional space (Tyler et al. 2007; Archimbaud, Aurore,

Nordhausen, Ruiz-Gazen 2018a). However, none of these analyses have been geared toward

genomic data. To test the robustness of the ICS, we used simulated genomic data from

previously published analyses (Lotterhos and Whitlock 2014; Lotterhos and Whitlock 2015;

Verity et al. 2017). A comprehensive description of the previously generated data is available in

(Lotterhos and Whitlock 2014; Lotterhos and Whitlock 2015). Briefly, genetic data were

simulated under a two-refuge demographic expansion where allele frequencies were sampled in

each of the 30 populations consisting of 20 individuals. The simulated data also consisted of

10,000 total loci where 9,900 were evolving neutrally and the remaining 100 loci were under the

influence of natural selection with different selection coefficients: 12 loci where s = 0.1, 38 loci

where s = 0.01, and 50 loci where s = 0.005. For each locus, three different univariant statistics

were calculated from BAYENV2 and used for multi-dimension analyses; these statistics include

the log-Bayes factor, Spearman's rho, and X^TX (Gunther and Coop 2013).

To directly compare between Mahalanobis and ICS, we identified the number of true-

positives/negatives, false-positives/negatives, and recall (the number of true-positives over the

number of putative positives identified) as a measure of performance. We display this

information in the form of a confusion matrix (Figure 3B & 3D). A confusion matrix is a table

that is used to describe the performance of a defined model (ICS and Mahalanobis distances) on

a test data set of known values (simulated data). In our confusion matrices, there are four

different categories that are defined by a Boolean format: Actual Neutral, Identified Neural,

Actual Selection, and Identified Selection; where "Actual" is defined by the known values from

the simulated data and "Identified" is defined by the outputs of the ICS and Mahalanobis. For

example, the number of loci that were "Actual Neutral" and "Identified Neutral" are true-

negatives and those that were "Actual Selection" and "Identified Selection" are true-positives.

In the COMICS application, outliers are determined by a statistical cutoff. Statistical

cutoffs in COMICS are defined by the user and are picked out of the distribution of ICS

distances, usually in the form of quantiles (i.e., top 1% of ICS distances). For our comparative

analyses we used a 1% cutoff. Of the 10,000 simulated loci (9,900 neutral and 100 selected), the

ICS was able to separate most selected loci from the neutral background across loci under

varying selective coefficients, including those under a weak selective pressure (Figure 3A). Out

of the top 1% (the first 100 loci with the largest ICS distance), there was a total of 11 false-

positives, with the remaining 89 loci being true-positives. In terms of neutral loci, the ICS

identified a total of 9,889 loci as true-negatives with only 11 false-negatives (Figure 3B). Across

different regimes of selective coefficients, all loci with a large selective coefficient (s = 0.1) were

identified as an outlier, ~95% (36 of 38) of loci with intermediate selection coefficient (s = 0.01)

were correctly identified as an outlier, while 82% of weakly selected loci (s = 0.005) were

identified as outliers. The fraction of relevant outliers verses the total of relevant outliers (recall)

for ICS was 0.89; meaning that out of the top 100 ICS distances, 89% of loci were under

selection (Figure 3A). Analysis with Mahalanobis distances allowed us to correctly identify loci

with strong selection coefficients (s = 0.1) as outliers. 87% of loci with intermediate coefficients

(s = 0.01) were correctly identified as outliers, and only 52% of weakly selected (s = 0.005) loci

were correctly identified as outliers (Figure 3C). The total recall for selected loci under the

Mahalanobis distance was 0.69 (Figure 3D). When ICS results are compared with those obtained

with Mahalanobis distances, we found nearly a threefold decrease in false-positives (total false-

positives in Mahalanobis = 31, as opposed to 36 in ICS) as well as an increase in the true-

positives (true-positives in Mahalanobis = 69 as opposed to 89 in ICS, Figure 3B & 3D).

Discussion:

As the amount of genetic data increase, so does the number of SNPs (or windows) that

can be tested for neutral or selective evolution. Because each SNP (or window) is considered an

individual hypothesis, confounding effects on genetic analyses including the inflation of false-

positives and false-negatives will become more prominent. The inflation of type I and type II

errors brings further issues pertaining to the interpretation of putatively selected genetic markers

and their implication on downstream analyses. There is a growing need in developing methods to

combine methods of selection, as the number of methods keeps growing in numbers. Here we

developed, tested, and introduced an R-package (COMICS) using the Shiny smart wrapper that

incorporates the use ICS specifically for genomic data.

We show that ICS is a robust method for identifying outlier loci in multi-dimensional

space and propose its use over other similar existing methods like Mahalanobis distances.

Specifically, the ICS identified fewer false-positives/negatives, more true-positives/negatives and

had a 20% higher recall for true-positives. One of the more interesting aspects of the ICS was its

ability to detect a larger proportion of weakly selected loci. Across the genome and across a

geographic space, loci experience selective pressures at varying strengths. Across methods,

signatures of loci under strong selection are often identified as true-positives. As the strength of

selection acting on a locus is reduced, the chances of identifying it as a true outlier are also

reduced, even after combining multiple selection detection methods that vary in their ability to

detect signatures of selection. Compared to contemporary methods, the ICS offers an increase in

the recall for outlier loci even for weakly selected loci. We therefore recommend that future

studies using univariate statistics to identify loci under selection consider the ICS as means to

identify outliers in multi-dimensional space.

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Figure 1. A screenshot of the suite of simulated selection scans that can be generated with
COMICS. COMICS also allows the user to create individual selection scans of each univariate
statistic that contributes the multidimensional scatter. In this example, there are three different
selection methods that contribute to the ICS distance, Scan1, Scan2, and Scan3. Note that each
color change represents a different chromosome. For simplicity, this example figure contains
only ten 2kb chromosomes.

Figure 2. A screen shot of the from the COMICS GUI showing the final genome scan once transformed to the multi-dimensional scatter. The Y-axis is the ICS distance while the X-axis the position along the genome. The color difference between grey and black represents transitions from one chromosome to another. The black horizontal line is the statistical cutoff that is defined by the user. Depending on the statistical cutoff, COMICS will render all data points about the threshold red where all other loci will conform to the default chromosome color. Not that the chromosomes numbers and sizes are the same as Figure 1.

A

356   B

ICS	Identified: Neutral	Identified: Selection	
Actual: Neutral	9,889	11	Recall: 0.89
Actual: Selection	11	89	

C

D

Mahalanobis	Identified: Neutral	Identified: Selection	
Actual: Neutral	9,869	31	Recall: 0.69
Actual: Selection	31	69	

Figure 3. Selection scans and confusion matrices for the ICS (A & B) and Mahalanobis (C &D)
distances calculated using simulated data. The horizontal line represents our statistical cutoff for
outlier loci where all true-negatives are marked as grey and false-positives are purple. Loci that
were simulated with a selection coefficient are in shades of red where the darkest represent loci
with the largest selection coefficient. Importantly, loci with a selection coefficient that were
below the statistical cutoff were deemed false-negatives and those above the line were true-
positives. Within confusion matrices, the sum of the columns and the sum of the rows equals the
total number of loci used in the simulated data. The red lines under the false-negatives, true-
positives, and recall signifies which method performed better.

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