1   **Title:** Speciation and gene flow across an elevational gradient in New Guinea kingfishers

2   **Running Title:** Speciation in New Guinea kingfishers

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Abstract: Theory suggests speciation without geographic isolation is plausible if

divergent natural selection is strong enough to counteract gene flow. Tropical mountains provide

strong and temporally stable environmental gradients that can promote local adaptation and

population genetic structure. Pairs of closely related species with adjacent but divergent

elevational ranges are common in these environments, a pattern that suggests parapatric

speciation (i.e., speciation with moderate gene flow), but evidence for this process is scarce.

Here we use genomic data from modern and historical museum specimens to investigate

speciation in a carefully chosen pair of sister New Guinea kingfisher species that segregate

ranges by elevation. We find that the lowland species *Syma torotoro* and montane species *S.*

megarhyncha form discrete genotypic clusters with bimodal variance in phenotypic traits.

Phylogenetic relationships among lineages are discordant between mitochondrial and nuclear

genomes, which *D*-tests and demographic inference indicate is partly driven by interspecific

gene flow after an initial period of isolation. Summary statistics reveal differentiation is

concentrated in a handful of small regions of the genome. These data underscore the apparent

scarcity of convincing cases of parapatric speciation. However, they also suggest selection across

elevational gradients can maintain species boundaries without strong intrinsic reproductive

isolation, a mechanism contributing to high tropical biodiversity.

Key words: parapatric speciation, speciation genomics, ecological speciation, Syma

Introduction: Adaptation across environmental gradients is ubiquitous in nature

(Turesson, 1925; McCormack & Smith, 2008; Cheviron & Brumfield, 2009), but its role in

promoting speciation remains contentious (Coyne & Orr, 2004; Mallet, 2005; Fitzpatrick,

Fordyce, & Gavrilets, 2008; Mallet, 2008). Disruptive natural selection can lead to local

adaptation that restricts gene flow between populations in different environments if it becomes

paired with a mechanism to promote nonrandom mating such as a pleiotropic "magic trait,"

linkage disequilibrium between separate loci involved with local adaptation and assortative

mating, or habitat preference leading to ecogeographic isolation (Rundle & Nosil, 2005; Via,

2009; Nosil, 2012). A robust body of theory suggests this process is possible under a range of

circumstances (Smith, 1966; Endler, 1977; Kirkpatrick & Ravigné, 2002; Doebeli & Dieckmann.

2003; Doebeli, Dieckmann, Metz, & Tautz, 2005; Hua, 2016). However, evolutionary biologists

have traditionally dismissed its relevance in natural systems given the obvious efficiency of

speciation without gene flow, and because there are few clear empirical examples (Coyne & Orr,

2004).

Tropical mountainsides generate strong and temporally stable environmental gradients,

providing a useful setting for testing the role of local adaptation in generating species richness in

nature. While allopatric divergence following dispersal or vicariance is doubtless an important

factor in the origin of species in the tropics (Smith et al., 2014), the importance of abiotic and

biotic ecological variables in driving evolutionary processes agnostic to geography is

increasingly recognized (Polato et al., 2018). Ecologists have long known that reduced seasonal

variation in temperature drives strong thermal stratification across tropical mountainsides (Polato

et al., 2018; Janzen, 1967; Sheldon, Huey, Kaspari, & Sanders, 2018), a pattern correlated with

high beta diversity (Jankowski, Ciecka, Meyer, & Rabenold, 2009; Cadena et al., 2012),

especially among closely related species (Terborgh & Weske, 1975; Freeman & Freeman, 2014).

Because many tropical taxa are residents that appear to show low rates of dispersal and have a

"slow pace of life" (Wiersma, Muñoz-Garcia, Walker, & Williams, 2007; Harvey, Aleixo, Ribas,

& Brumfield, 2017; Smith et al., 2017), strong disruptive selection could plausibly counteract the

homogenizing effects of gene flow to drive adaptive divergence.

Local adaptation across tropical elevational gradients has been documented by clines in

functional genes and intraspecific population genetic structure (Cheviron & Brumfield, 2009;

DuBay & Witt, 2014; Funk et al., 2016; Gadek et al., 2018). This has been shown theoretically to

"scale up" to speciation under a scenario of niche expansion (Hua, 2016), but empirical evidence

remains mixed. Phylogenetic and phylogeographic comparative studies suggest this process has

occurred in *Ithioma* butterflies (Elias et al., 2009) and Andean amphibians and reptiles (Arteaga

et al., 2016). In Andean birds, a broad consensus holds that elevational replacements primarily

form through divergence in allopatry followed by secondary contact and range displacement

(Cadena et al., 2012; DuBay & Witt, 2014; Caro, Caycedo-Rosales, Bowie, & Cadena, 2013;

Cadena & Céspedes, 2019), despite equivocal results in some tests. Yet we are aware of only one

study that has used population genomics to evaluate speciation across elevational gradients,

which found strong evidence that adaptation to altitude drives speciation in Senecio ragwort

plants in temperate Italy (Chapman, Hiscock, & Filatov, 2013; Osborne, Batstone, Hiscock, &

Filatov, 2013). The question of the relative contribution of adaptation in the presence of gene

flow across elevational gradients to speciation and broader patterns of tropical biodiversity

remains open.

The Yellow-billed and Mountain Kingfishers *Syma torotoro* and *S. megarhyncha* (Aves:

Alcedinidae) are putative sister taxa that segregate by elevation and vary only subtly in

phenotype (Mayr, 1942; Pratt & Beehler, 2015; Diamond, 1972). Several aspects of their

distribution and ecology make the pair unusually plausible candidates for speciation driven by

divergent natural selection in the absence of significant geographic isolation (Rand, 1936). First,

S. megarhyncha is only found in the highlands of New Guinea, where it replaces *S. torotoro*

above ~1000 m throughout most of the island (Pratt & Beehler, 2015). A scenario of divergence

in allopatry is therefore less parsimonious than divergence in parapatry, as it requires substantial

historical range shifts. Second, in the Arfak and Tamrau mountains of western New Guinea

where *S. megarhyncha* is absent, *S. torotoro* does not expand its elevational range upslope,

suggesting the two species are not partitioning a shared fundamental niche (Beehler & Pratt,

2016). This hypothesis is strengthened by evidence of morphological adaptation to a cooler

climate in the larger-bodied *S. megarhyncha* (Beehler & Pratt, 2016). Third, *Syma* are territorial

and sedentary, making it plausible that dispersal was sufficiently reduced to be overcome by

divergent selection across the steep environmental gradient they inhabit (Endler, 1977).

However, species limits and range-wide variation have never been quantitatively assessed and

observed differences might instead reflect phenotypic plasticity or clinal variation of a single

widespread lineage (Caro et al., 2013).

Here we use genome-wide DNA sequences, bioacoustic data, and morphometric analyses

to investigate processes of speciation in *Syma* kingfishers. We assessed species limits and

phylogenetic relationships, tested for introgression and inferred demographic history, and

described the landscape of genomic divergence. If speciation was driven by local adaptation in

the absence of geographic isolation, we expected *S. torotoro* and *S. megarhyncha* to be

reciprocally monophyletic taxa showing evidence of gene flow throughout their history of

divergence. If speciation occurred in allopatry followed by range expansion and elevational

displacement, we expected to see little evidence of past or present gene flow, as this scenario

would require sufficient reproductive isolation to develop to permit coexistence without genetic

homogenization and the fusion of lineages on secondary contact. If speciation occurred in

allopatric driven by adaptation to altitude and subsequent habitat isolation but few postzygotic

barriers, we expected to see support for a model of secondary contact and evidence of recent

gene flow.

Materials and Methods: *Study system.* Yellow-billed Kingfisher *Syma torotoro* and

Mountain Kingfisher *Syma megarhyncha* (Aves: Alcedinidae) are the sole members of their

genus. Members of the tree kingfisher subfamily Halcyoninae, they are endemic to New Guinea,

its satellite islands, and the Cape York Peninsula of Australia. *S. torotoro* is found in tropical

lowland forest and savannah from sea level to ∼500 m elevation, or less commonly to ∼1100 m

(Pratt & Beehler, 2015). *S. megarhyncha* is found from ∼600 m to 2700 m or higher (Pratt &

Beehler, 2015). Though cited as a classic example of congeneric elevational replacements

occurring in parapatry (Diamond, 1972), their elevational ranges have also been reported to

either overlap (Gregory, 2017) or be separated by a substantial gap (Freeman & Freeman, 2014;

Sam, Koane, & Novotny, 2014). Both species are omnivorous territorial interior forest residents

and differ only in *S. megarhyncha's* larger body size, deeper call, and the extent of black on the

top of its bill in one subspecies (Pratt & Beehler, 2015). Insular *S. torotoro* subspecies *S. t.*

ochracea differs substantially from its conspecifics and *S. megarhyncha* in call, and is

intermediate in size, leading some authors to propose it represents a distinct species (Beehler &

Pratt, 2016).

Morphological and bioacoustics data. We measured bill length, bill depth, tarsus, wing

chord, and tail length from 72 museum specimens of *Syma torotoro (n*=40) and *S. megarhyncha*

(*n*=32) at the American Museum of Natural History, collected from 1894-1965 and including

only individuals of known sex as originally identified by the preparator. Using these data, we

performed principal component analyses in R (R Core Team 2018) with normalized variables

and used PC1 to build mixture models using the R package mclust v. 5.4.1, which we evaluated

with a maximum likelihood classification approach (Scrucca, Fop, Murphy, & Rafferty, 2016).

We downloaded all available vocalizations from *S. torotoro* (*n*=34) and *S. megarhyncha* (*n*=14)

from xeno-canto and Cornell's Macaulay Library. We filtered these data for quality and

quantified 36 standard bioacoustic variables using the warbleR package v. 1.1.14 in R (Araya-

Salas & Smith-Vidaurre, 2017), analyzing 278 distinct vocalizations from *S. torotoro* and 106

from *S. megarhyncha* in total. We ran PCA with normalized variables on the output and used

these data to alternate species delimitation models using the same approach as with our

morphological data.

Sampling, library preparation, and DNA sequencing. We extracted DNA from fresh

tissues (*n*=6) and toepad samples from historical museum specimens (*n*=34) from 30 individuals

of *S. torotoro* (*n*=30) and 10 individuals of *S. megarhyncha* (*n*=10). Though only partially

overlapping with samples used in morphometric analyses, these individuals represented the full

extent of both species' ranges in New Guinea and Australia (Table 1) and included all described

subspecies. We extracted DNA from fresh tissues using a Qiagen DNAeasy kit and the

manufacturer's recommended protocol. For historical toepad samples (collected 1877-1973), we

extracted DNA using either a using a phenol-chloroform and centrifugal dialysis method

(Dumbacher & Fleischer, 2001) (for reduced representation sequencing) or a standard salt

extraction protocol (for whole genome sequencing). Due to constraints of cost and time, we

employed two complementary sequencing approaches. On a subset of samples (n=20), we

performed reduced representation genome sequencing using a hybridization capture with RAD

probes (hyRAD) approach, described in detail elsewhere (Linck, Hanna, Sellas, & Dumbacher,

2017). We sent the remaining samples to the UC Berkeley's Vincent J. Coates Genomic

Sequencing Laboratory, where laboratory staff prepared genomic libraries for low coverage

whole genome sequencing (WGS) using Illumina TruSeq Nano kits and a modified protocol that

skipped sonication and enzymatically repaired fragments with RNAse. They then pooled (n=20)

and sequenced these samples with 150 base pair paired end reads on a single lane of an Illumina

HiSeq 4000.

Sequence assembly and variant calling. We processed demultiplexed reads from both

sequencing strategies together with a custom bioinformatic pipeline optimized for handling

degraded DNA data and available at https://github.com/elinck/syma_speciation. Briefly, we

trimmed raw reads for adapters and low-quality bases using bbduk from BBTools version 38.06

suite of bioinformatics tools. We aligned these reads to an unpublished draft genome of

Woodland Kingfisher *Halcyon senegalensis* from the Bird 10K Genome Project using bbmap

with a k-mer value of 12, a maximum indel length of 200 bp, and a minimum sequence identity

of 0.65. These highly sensitive alignment parameters were necessary given the clade containing

Syma diverged from the clade containing *Halcyon* approximately 15 mya (Andersen,

McCullough, Mauck, Smith, & Moyle, 2018). We used PicardTools v. 2.17.8 and GATK v. 3.6.0

(McKenna et al., 2010) to append read groups and perform local realignment on .bam files. We

then used mapDamage 2.0.9 to account for postmortem damage to DNA from historical museum

specimens by rescaling quality scores (Jónsson, Ginolhac, Schubert, Johnson, & Orlando, 2013).

We performed multisample variant calling using the UnifiedGenotyper tool in GATK v. 3.6.0,

and filtered our variant calls for missing data, coverage, and quality with VCFtools 0.1.16

(Danecek et al., 2011). We generated multiple SNP datasets by implementing different filters to

suit the requirements and goals of different analyses; these settings and their underlying rationale

are described in detail below. To complement our nuclear DNA sequences, we assembled near-

complete and fully annotated mitochondrial genomes from a majority of individuals using

mitofinder v. 1.0.2 and a complete mtDNA genome from close relative *Todiramphus sanctus* as

a reference (Andersen et al., 2015). We then extracted NADH dehydrogenase 2 (ND2) from

these genomes and performed multiple sequence alignment using MAFFT v7.407 under its "–

auto" parameter setting package (Katoh & Standley, 2013).

Population structure inference. We evaluated population genetic structure within and

between species using a nonparametric clustering approach. We performed principal component

analysis of genotypes (PCA) and identified putative genetic clusters for K=2 through K=4 using

adegenet v. 2.1.1 (Jombart, 2008) and a 95% complete dataset with 66,917 SNPs from all

individuals in both datasets with a minimum depth of coverage of 3x per individual, a maximum

depth of coverage of 120x per individual, a minimum quality score of 30, and a minimum minor

allele frequency of 0.05. These relatively lenient filtering parameters were chosen to permit the

inclusion of samples from both sequencing strategies in our initial assessment of probable

species limits. We evaluated the best-fit clustering model using the Bayesian Information

Criterion, again implemented in adegenet.

Phylogenetic inference. We evaluated phylogenetic relationships among lineages with

three complementary approaches. Using our mitochondrial DNA (ND2) alignment, we inferred a

time-calibrated phylogeny in BEAST 2.6.0 (Bouckaert et al., 2019) including all samples and

Sacred Kingfisher *Todiramphus sanctus* as an outgroup (Andersen et al., 2015). We used a strict

molecular clock with a rate of 2.9×10^{-8} following Lerner, Meyer, James, Hofreiter, & Fleischer

(2011), a GTR+GAMMA model of nucleotide evolution, and a Yule species tree prior. We ran

BEAST for 50 million generations, storing trees every 1000 generations, and assessed mixing,

likelihood stationarity and adequate effective sample sizes (ESS) above 200 for all estimated

parameters using Tracer v.1.7.1 (Rambaut, Drummond, Xie, Baele, & Suchard, 2018). We then

generated a maximum clade credibility tree using TreeAnotator v.2.6.0, discarding the first 10%

of trees as burnin (Bouckaert et al., 2019). We next inferred a nuclear DNA phylogeny using

SVDquartets (as implemented in PAUP v.4.a165) without *a priori* species assignments

(Chiffman & Kubatko, 2014). To reduce the influence of sequencing errors, we used a

stringently filtered SNP dataset, including only whole genome sequencing data, sites with a

minimum minor allele count of 3, no more than 10% missing data, a minimum depth of 5x and a

maximum depth of 120x per individual, and a minimum quality score of 30. We converted this

diploid alignment to a single consensus IUPAC ambiguity code per individual using vcf2phylip

v2.0 (Ortiz, 2019) and included the reference sequence from *H. senegalensis* as an outgroup. We

ran SVDquartets with 2000 quartets and 100 bootstrap replicates. Lastly, we inferred a species

tree under a multispecies coalescent model with SVDquartets and the same SNP dataset as

above, using taxon partitions concordant with k-means clustering results and ND2 lineages and

the speciesTree=Yes parameter setting.

Demographic inference. We inferred the demographic history of diverging lineages using

moments v.1.0.0, which uses ordinary differential equations to model the evolution of allele

frequencies (Jouganous, Long, Ragsdale, & Gravel, 2017). We used the joint_sfs_folded()

function in Python package scikit-allel v.1.2.1 (Miles, Ralph, Rae, & Pisupati, 2019) to calculate

a folded joint site frequency spectrum (JSFS) using a SNP dataset that included only whole

genome sequencing data, a minimum minor allele count of 1, no more than 10% missing data per

site, a minimum depth of 6x per individual, and a minimum quality score of 30. We chose these

filtering parameters to reduce the influence of sequencing error without strongly biasing the

JSFS. This dataset did not include *S. t. ochracea,* and had been further thinned to include only

variants in approximate linkage equilibrium (defined as sites where $r^2 < 0.1$) using scikit-allel's

locate_unlinked() function. To account for the remaining missing data, we projected the JSFS

down to an effective sample size of 16 x 12 chromosomes for *S. megarhyncha* and *S. torotoro*

respectively.

We then specified four basic demographic models chosen to represent plausible

speciation scenarios in *Syma,* which differed by level and timing of gene flow. These were: an

isolation-with-migration model (IM), which permitted gene flow throughout the history of

diverging lineages; an isolation-with-ancestral-migration model (AM), which featured an initial

period of gene flow followed by a period of isolation; a model of secondary contact (SC), which

featured an initial period of isolation followed by a period of gene flow; and a model of strict

isolation (SI), which allowed no gene flow following initial divergence. Next, we specified four

additional models which differed from each of the above only by allowing exponential

population growth in the most recent time period: an isolation-with-migration-and-growth model

(IMg), an isolation-with-ancestral-migration-and-growth model (AMg), a secondary contact and

growth model (SCg), and a strict isolation and growth model (SIg).

After initially optimizing parameters for fitting each model using the "optimize log"

method and a maximum of 3 iterations, we ran 9 additional optimizations, randomly sampling

starting values from a uniform distribution within the bounds of 1×10^{-4} and 10. We converted

parameter values to real units using a genome-wide mutation rate of 2.3×10^{-9} (Smeds,

Qvarnström, & Ellegren, 2016), an effective sequence length scaled to reflect our LD-thinned

SNP dataset, and a generation time estimate of two years. We generated parameter uncertainty

estimates for the best-fit demographic model (selected based on AIC values) by fitting it to 200

bootstrapped site frequency spectra generated using the fs.sample() function on our data, an

approach which is appropriate when sites are unlinked (Jouganous et al., 2017).

Introgression. We explicitly tested for introgression between lineages using the four

taxon *D*-test (Green et al., 2010), which relies on asymmetries in discordant site patterns to

distinguish incomplete lineage sorting from introgression. We randomly selected one

representative sequence for *S. torotoro*, *S. megarhyncha*, and *S. t. ochracea* from the nuclear

DNA SNP alignment used in our phylogenetic analyses, and again used the reference sequence

for *H. senegalensis* as an outgroup. We then calculated *D* using a custom python script (available

at https://github.com/elinck/syma_speciation/), estimating uncertainty using a block jacknife

with 100 replicates.

Genome scans. We assessed levels of divergence across the genome by calculating

Wright's F_{ST} and D_{XY} in 50 kbp windows with a 10 kbp step size using scripts written by Simon

Martin (https://github.com/simonhmartin/genomics_general), using a VCF file generated from

our whole genome sequencing data that included invariant sites and was filtered for a minimum

Q score of 30 and a minimum depth of 3x per individual, for a total of 1,858,764 SNPs and

102,606,487 total sites. We permitted a lower depth of coverage threshold in this analysis as our

goal was to describe the overall landscape of differentiation as completely as possible given the

constraints of our data. To assign chromosome identity to scaffolds and windows, we aligned the

H. senegalensis draft genome to the chromosome-level genome assembly of *Taenopygia guttata*

(Warren et al 2010) using NUCmer in MUMmer v. 3.1 (Delcher et al., 1999), allowing a

maximum gap of 1000 bp and using a minimum sequence identity threshold of 10 kbp per

contig. We evaluated correlations among summary statistics using simple linear models

implemented in R (R Core Team, 2018).

Results: *DNA sequencing from historical specimens.* We extracted DNA and generated

genome-wide sequence data from all historic toepad samples across both species' relatively

inaccessible distributions, with collection dates ranging from 1896 to 1973 and including 3

individuals collected by Ernst Mayr himself in 1929. We present a detailed description of

reduced representation hyRAD data elsewhere (Linck et al., 2017). On average we were able to

align 83.3% of reads to the draft *Halcyon senegalensis* genome, ranging from 35.5% to 92.37%

across individuals; this broad range likely reflects high DNA degradation in a handful of

samples. Whole genome sequencing data had an average depth of coverage of 5.38x per

individual, ranging from 1.92x to 12.12x. Following variant calling and filtering for depth of

coverage and quality, a 95% complete data matrix including 37 individuals and both WGS and

hyRAD data had 66,917 SNPs, which was further reduced to 10,351 SNPs after thinning to 1 site

for every 50 kbp to reduce the influence of linkage disequilibrium. A second matrix of whole

genome sequencing data alone had 78,882,912 SNPs, which was reduced to 1,858,764 SNPs

after filtering for a minimum depth of coverage of 3x and a minimum quality score of 30.

Species limits and phylogeny. Analysis of genome-wide DNA sequence data,

morphometrics, and calls supported *Syma torotoro* and *S. megarhyncha* as distinct, assortatively

mating lineages (Figure 1). Principal component analysis of genotypes separated *S. torotoro* and

S. megarhyncha into discrete clusters (Figure 1A; Figure 1C) with little evidence of hybrid

genotypes along the primary axis of genetic differentiation, with PC1 explaining 17.02% of total

variance. PC2 explained 6.31% of total variance and separated insular subspecies *S. t. ochracea*

into a third discrete cluster. The best-fit model from a k-means clustering analysis (as identified

by BIC scores) perfectly recovered these groups (Figure 1C).

Analyses of phenotypic data formed discrete clusters concordant with genomic results

and consistent with morphological differentiation by elevation. Bill width and depth, tarsus, wing

chord, and tail length were significantly larger in *Syma megarhyncha* after correcting for

multiple comparisons in Welch's two sample T-tests (all comparisons $p<1\mathrm{x}10^{-7}$), and species

was a significant predictor of PC1 in a linear model ($p<1\mathrm{x}10^{-14}$). Following principal component

analysis, PC1 explained 81.47% of variance across all five traits. Bayesian Information Criterion

(BIC) selected two distinct normal distributions out of normal mixture models (NMMs) fit to

PC1 (log-likelihood=−378.0914) (Figure 1D). Calls of *S. torotoro* had a significantly higher

frequency ($p<1\mathrm{x}10^{-5}$) but did not differ in duration. Species was a significant predictor

($p<1\mathrm{x}10^{-5}$) of PC1 in a principal component analysis of the 36 bioacoustic variables quantified in

warbleR (Araya-Salas & Smith-Vidaurre, 2017) and explained 35.72% of total variance. Because

NMMs assume independence of observations, we did not perform formal model fitting for

bioacoustic data, but a frequency distribution strongly suggests values are bimodally distributed

by species identity (Figure 1E).

A time-calibrated phylogeny of the mitochondrial gene ND2 supported the reciprocal

monophyly of *S. megarhyncha* and *S. torotoro,* but unexpectedly recovered mainland *S. torotoro*

as sister with a clade containing *S. megarhyncha* and insular endemic subspecies *S. t. ochracea*

(Figure 1F). Within *S. torotoro,* a clade of four individuals with full bootstrap support from the

southeast peninsula of Papua New Guinea was separated from the remaining individuals, but

with little other apparent geographic structure across the tree. Based on a strict molecular clock

with a rate of 2.9x10−8, *S. megarhyncha* and *S. t. ochracea* initially diverged from *S. torotoro*

approximately 790,000 years ago (95% CI: 607,422-972,619 years); a subsequent split between

S. megarhyncha and *S. t. ochracea* occured approximately 377,997 years ago (95% CI: 243,262-

512,732 years), and a split between *S. torotoro* populations in the SE peninsula of Papua New

Guinea and the rest of the species occurred approximately 428,580 years ago (95% CI: 304,805-

552,355 years). In contrast, our nuclear DNA lineage phylogeny was more poorly resolved and

conflicted with both the ND2 tree and current taxonomy (Figure 1G). An initial divergence event

with full bootstrap support separated a clade containing *S. megarhyncha* individuals from far

western New Guinea and *S. t. ochracea*; a second fully supported split separated S. torotoro

populations in SE Papua New Guinea from an ingroup containing poorly resolved clades

containing the remainder of *S. torotoro* and *S. megarhyncha* individuals. Lastly (and again

conflicting with both other phylogenies), our nuclear DNA species tree from SVDquartets

grouped *S. torotoro* and *S. megarhyncha* together in a clade that was sister to *S. t. ochracea*, with

full bootstrap support at all internal nodes (Figure 1H)

Demographic history and introgression. D-tests indicated asymmetric genealogies among

lineages that were the product of introgression or ancestral population structure rather than

incomplete lineage sorting alone. For two of three possible species tree topologies, D differed

signficantly from 0 (one sample t-tests: $p<2x10^{-16}$ and $t=877.23$ given

(ochracea,megarhyncha),torororo); $p<2x10^{-16}$ and $t=837.36$ given

(ochracea,torotoro),megarhyncha)), with signs indicating introgression between *S. torotoro* and

S. megarhyncha, rather than either of these species and *S. t. ochracea* (Figure 1A). Bootstrapped

D-test values for the topology (megarhyncha,torororo),ochracea) also different significantly from

zero (one sample t-test: $p<2\times10^{-16}$ and $t=-19.416$), but with a range which encompassed it (-

0.023 to 0.003).

346   The best-fit demographic model involved secondary contact without recent exponential

population growth (AIC=1140.124; log-likelihood=-564.062), followed by a model of ancestral

migration (AIC=1192.047; log-likelihood=-590.062) (Table 2). Parameter estimates from the

secondary contact model provide additional evidence for introgression and suggest it has been

ongoing for long time periods (Figure 2B, 2C, 2D). After an initial period of approximately

458,469 years of geographic isolation following divergence (SD: 6,162 years), we inferred *Syma*

species have been exchanging genes for ~191,231 years (SD: 7,510 years) at a rate of 1.14

individuals per generation (SD: 0.023) from *S. torotoro* to *S. megarhyncha*, and a rate of 0.12

individuals per generation (SD: 0.006) from *S. megarhyncha* to *S. torotoro.* Summing the first

and second periods indicates an initial divergence event approximately 649,700 years ago—a

date consistent with the estimate from the mitochondrial DNA molecular clock analysis. We

inferred a population size for *S. torotoro* of ~1,909,985 individuals (SD: 26,028) and a

population size for *S. megarhyncha* of ~368,390 individuals (SD: 8,528).

359   *Genome scans.* Analysis of whole genome sequencing data suggests *S. torotoro* and S.

megarhyncha differ primarily in small regions of the genome (Figure 3A). Genome scans of

divergence in 50 kb sliding windows with a step size of 10 kb revealed globally low to moderate

genetic divergence (mean F_{ST}=0.0947) defined by 21 autosomal F_{ST} peaks (Figure 3A; Figure

3B). The Z chromosome showed low interspecific divergence, suggesting a limited role in

speciation (Irwin, 2018), though this may represent an artifact of high intraspecific diversity due

to misincorporation of reads from the W chromosome. Correlations between genome-wide

summary statistics suggested a role for both structural reductions in recombination and positive

selection in shaping F_{ST} peaks. F_{ST} was negatively but weakly correlated with D_{XY} ($p<1x10^{-15}$,

R^2=0.1271) (Figure 3A). Considering a subset of chromosomes with prominent F_{ST} peaks (Figure

3B), this correlation remained significant but was slightly reduced ($p<1x10^{-15}$, R^2=0.1044)

(Figure 3C).

Discussion: Speciation across environmental gradients is commonly invoked in

explanations of high tropical species richness and the latitudinal biodiversity gradient (Caro et

al., 2013; Schneider, Smith, Larison, & Moritz, 1999; Portik et al., 2017; Beheregaray, Cooke,

Chao, & Landguth, 2015). Tropical elevational gradients have high levels of temporally stable

thermal stratification (Janzen, 1967) that has been linked to selection for narrow thermal

physiologies (Sheldon et al., 2018), reduced upslope or downslope dispersal (Polato et al., 2018),

and high beta diversity (Jankowski et al., 2009). As a result, they have frequently been studied as

a stage for this process (Mayr, 1942; Cadena et al., 2012; Funk et al., 2016; Arteaga et al., 2016;

Caro et al., 2013; Moyle, Manthey, Hosner, Rahman, Lakim, & Sheldon, 2017). Yet previous

comparative phylogenetic studies have overwhelmingly supported models of divergence in

allopatry followed by secondary contact and range displacement without gene flow, particularly

in vertebrates (Arteaga et al., 2016; Caro et al., 2013; Moyle et al., 2017). Our finding of well-

defined genotypic and phenotypic clusters (Figure 1) in the face of recent introgression (Figure

2) suggests that selection across elevational gradients is sufficient to maintain species boundaries

with little evidence of strong postzygotic isolation. This result supports an intuitive and widely

cited but poorly buttressed theory: that selection across environmental gradients can serve as a

motor for tropical diversification, either as the primary driver of speciation (e.g., in ecological

speciation models) or as a force for reinforcement and disruptive selection after the build-up of

partial reproductive isolation in allopatry.

Though evidence from genotypic and phenotypic clustering analyses suggests *S. torotoro*

and *S. megarhyncha* are assortatively mating lineages that are largely match current taxonomy,

alternate approaches to phylogenetic inference provide conflicting data on species limits and

evolutionary relationships. A well-sampled phylogeny of the mitochondrial gene ND2 shows an

unexpected sister relationship between *S. megarhyncha* and insular *S. torotoro* subspecies

ochracea, previously hypothesized to represent a species-level lineage (Pratt & Beehler, 2015),

but also revealed the expected pattern of monophyly for *S. torotoro* and *S. megarhyncha*. In

contrast, a phylogeny from nuclear DNA is strongly discordant with both mitochondrial

relationships and the results of clustering analyses (Figure 1E). We propose this conflict can be

primarily explained by introgression between *S. torotoro* and *S. megarhyncha* in eastern New

Guinea, as supported by evidence from *D*-tests and demographic inference (Figure 2). Increased

structure in mitochondrial DNA relative to the nuclear genome may also be generated by with

reduced hybrid fitness in heterogametic females (in accordance with Haldane's rule), or male

biased dispersal (Toews & Brelsford, 2012)—both of which are plausibly contributing to

patterns in *Syma*.

Despite this phylogenetic uncertainty, we believe our data from *Syma* are consistent with

speciation with driven by niche expansion and disruptive selection across an elevational gradient

in allopatry followed by secondary contact and recent gene flow. Divergence in morphometric

traits, the most visible phenotypic difference between *S. torotoro* and *S. megarhyncha* (Figures

1D, 1E), is largely thought to reflect ecological adaptation in allopatric populations (Ricklefs &

Bermingham, 2007; Winger & Bates, 2015). While this may arise as a result of character

displacement from competition in populations experiencing secondary contact (Diamond, Pimm,

Gilpin, & LeCroy, 1989), it presumes sufficient reproductive isolation has developed to permit

coexistence without genetic homogenization and the fusion of lineages. In *Syma*, tests for

introgression and demographic inference indicate gene flow has occurred for nearly 200,000

years, rendering the presence of strong genetic incompatibilities unlikely. Though we cannot rule

out a role for ancestral population structure in generating significant *D*-test results, parameter

estimates from our best fit demographic model and reports of occasional hybridization in the

field (Beehler & Pratt, 2016) provide additional evidence for introgression. Additionally, the

failure of *S. torotoro* to expand its elevational range upslope in absence of *S. megarhyncha* in the

mountains of the Vogelkop Peninsula in West Papua supports a hypothesis of adaptation to

divergent elevational niches, rather than competitive segregation of a shared, broader realized

niche.

However, we emphasize that in conflict with distributional, ecological, and

morphological evidence, our genomic data fail to support a traditional model of parapatric

speciation—or what Endler (1977) termed "primary" intergradation. Given *Syma* appeared to be

an unusually plausible candidate for this process, our results underscore the apparent rarity of the

phenomenon in nature. While this is no doubt partly due to the stronger divergent selection

required to achieve the evolution of reproductive isolation in the face of gene flow compared to

in allopatry, we suggest the geographic complexity of landscapes and stochastic population size

changes overtime are additional factors rendering truly parapatric divergence unlikely. There is

some evidence that even the contemporary distribution of *Syma* is less continuous than it is

typically described: recent surveys on Mt. Wilhelm and Mt. Karimui in Papua New Guinea

found large gaps in the elevational ranges of *S. torotoro* and *S. megarhyncha* (Freeman &

Freeman, 2014; Sam et al., 2014), in contrast to earlier studies (Diamond, 1972). It is therefore

easy to imagine orogenic vicariance or a dispersal event into the highlands from the common

ancestor of *S. torotoro* and *S. megarhyncha* might have resulted in a substantial period of

geographic isolation despite relatively short horizontal distances between species' ranges.

Though we did not identify the targets of selection, our data are consistent with disruptive

positive selection leading to high divergence in small regions of the genome against a

background of long-term introgression. These genomic "islands" have become a hallmark of the

high throughput sequencing era and have been shown in wide range of nonmodel organisms

(Nosil, 2012; Bay & Ruegg 2017). Early interpretations of the pattern as a clear signature of

speciation with gene flow—where F_{ST} peaks correlate to genes for traits under disruptive

selection while low F_{ST} regions remain susceptible to introgression—have been complicated by

evidence numerous processes can produce similar distributions, including selective sweeps in

allopatry or linked selection paired with recombination rate variation (Cruickshank & Hahn,

2014; Bay & Ruegg 2017; Irwin et al., 2018; Via, 2012; Burri, 2017). Evidence from multiple

summary statistics across the same genomic windows can also help tease apart underlying

mechanisms in some cases (Irwin et al., 2018): for instance, low absolute differentiation (D_{XY}) in

genomic islands can indicate F_{ST} values are inflated by low intraspecific diversity, casting doubt

on models of divergence with gene flow (Cruickshank & Hahn, 2014).

Here, our data defy easy description, highlighting the limits of this approach. A handful

of major F_{ST} peaks stand out against a background of low F_{ST} windows, with a global average of

0.0947 (Figure 3A; Figure 3B). However, D_{XY} shows only a weak correlation with FST values

(Figure 3C), which is further reduced in outlier windows. Low in some F_{ST} peaks and high in

others, this relationship supports the likely hypothesis that recombination rate, selection against

hybrid ancestry, disruptive selection, and gene flow have interacted to form a complex mosaic in

the genomes of *S. torotoro* and *S. megarhyncha.* Given the limitations of short-read sequencing

data, however, we concede these patterns may be influenced by either alignment artifacts or

undetected structural rearrangements. In particular, we point to large peaks on chromosomes 5

and 11 that deserve further scrutiny as possible inversions contributing to reproductive isolation

(Hooper & Price, 2017) (Figure 3A and 3B).

As adaptive differentiation in morphology appears to have occurred during the initial

stages of lineage divergence, *Syma* is plausibly a case of ecological speciation *sensu* Nosil and

Schluter (Rundle & Nosil, 2005; Nosil, 2012; Schluter, 2009), who define the process agnostic to

the presence or absence of gene flow. While we lack strong evidence for a target of disruptive

selection or a specific mechanism to link it to reproductive isolation, we speculate selection on

body size may also affect mate choice. Morphometric data from *Syma* are consistent with

Bergmann's rule and its prediction of larger body size in the higher elevation cooler climate

species, *S. megarhyncha* (but see Freeman (2017)). As one of only three kingfisher species found

above 2000 m in New Guinea and the only regional kingfisher that is a high elevation specialist,

Syma megarhyncha is likely to have a thermal physiology under stronger selective constraint for

warm environments, requiring greater adaptive divergence in body size to colonize montane

environments. An increase in body size might directly lower frequency calls as a biproduct of

morphological divergence, and assortative mating might act on either trait individually or in

tandem (Derryberry et al., 2018; Slabbekoorn & Smith, 2002; Zhen et al., 2017).

What does *Syma* reveal about the origin of elevational series of congeners and by

extension their significant contribution to tropical montane biodiversity (Freeman & Freeman,

2014)? First, our results suggest disruptive selection across elevational gradients can effectively

maintain species limits in the absence of strong postzygotic reproductive isolation, a finding with

broad relevance to speciation in tropical mountains regardless of the geographic mode of

divergence or levels of gene flow. In a scenario of secondary contact without adaptation to

divergent elevational niches, *Syma* lineages would likely have fused. This suggests allopatric

divergence may not be sufficient in isolation of other processes to drive many speciation events

in tropical mountains and highlights the important role of habitat isolation in speciation

processes in taxa (like birds) that are slow to evolve reproductive incompatibilities. Yet at the

same time, our failure to find evidence for parapatric speciation in *Syma* in spite of its apparent

plausibility make the process seem likely to be vanishingly rare in birds, at least across tropical

elevation gradients. Furthermore, while New Guinea is large and geologically complex, it is

dwarfed by other tropical montane regions, which potentially have increased odds for speciation

in allopatry (Pratt & Beehler, 2015; Price, 2008). Still, we emphasize even if speciation across

elevational gradients remain a rare exception among birds, the conditions that make it possible in

Syma are more common in other taxa. Indeed, preliminary studies suggest it may be a much

more common mechanism in amphibians, insects, and plants (Funk et al., 2016; Elias et al.,

2009; Arteaga et al., 2016; Chapman et al., 2013). We highlight the importance of natural history

studies of poorly known tropical organisms in establishing candidates for further investigation

with genomic and experimental approaches.

Data availability: All code used in this study can be found at

https://github.com/elinck/syma_speciation Processed data are available from Dryad [doi:

pending], and sequence data are available from the NCBI SRA [accession: pending].

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Table 1. Sampling, sequencing, and analysis information. WP=Indonesian New Guinea; PNG=Papua New Guinea.

Specimen	Species	Subspecies	Locality	Tissue source	Date collected	Sequencing strategy	PCA	mtDNA phylogeny	nuDNA phylogeny	D-tests	Demographic inference	Genome scans
AMNH:Birds:293714	torotoro	torotoro	Ifaar, WP	toepad	1928	hyRAD	yes	yes	no	no	no	no
AMNH:Birds:293715	torotoro	torotoro	Kepaur, WP	toepad	1897	hyRAD	yes	yes	no	no	no	no
AMNH:Birds:300723	torotoro	torotoro	Kepaur, WP	toepad	1897	hyRAD	yes	yes	no	no	no	no
AMNH:Birds:300723	torotoro	torotoro	Misol Island, WP	toepad	1900	hyRAD	yes	yes	no	no	no	no
AMNH:Birds:329542	torotoro	torotoro	Wasior, WP	toepad	1928	hyRAD	yes	yes	no	no	no	no
AMNH:Birds:437798	torotoro	torotoro	Amberbaki, WP	toepad	1877	hyRAD	yes	yes	no	no	no	no
AMNH:Birds:637429	torotoro	torotoro	Humbolt Bay, WP	toepad	1928	hyRAD	yes	yes	no	no	no	no
AMNH:Birds:637441	torotoro	torotoro	Mt. Mori, WP	toepad	1899	hyRAD	yes	yes	no	no	no	no
AMNH:Birds:637445	torotoro	torotoro	East Sepik Province, PNG	toepad	2003	hyRAD	yes	yes	no	no	no	no
AMNH:Birds:637446	torotoro	torotoro	Waigeu Island, WP	toepad	1900	hyRAD	yes	yes	no	no	no	no
AMNH:Birds:637450	torotoro	tentelare	Aru Islands, WP	toepad	1896	hyRAD	yes	yes	no	no	no	no
AMNH:Birds:637464	torotoro	tentelare	Aru Islands, WP	toepad	1900	hyRAD	yes	yes	no	no	no	no
AMNH:Birds:637464	torotoro	ochracea	Normanby Island, PNG	toepad	1934	hyRAD	yes	yes	no	no	no	no
CAS:Birds:7131	torotoro	pseuestes	Gulf Province, PNG	muscle	2002	hyRAD	yes	yes	no	no	no	no
KU:Birds:5215	torotoro	pseuestes	Gulf Province, PNG	muscle	2003	hyRAD	yes	yes	no	no	no	no
KU:Birds:5464	torotoro	pseuestes	Gulf Province, PNG	muscle	2003	hyRAD	yes	yes	no	no	no	no
KU:Birds:626	torotoro	meeki	Varirata National Park, PNG	muscle	2011	hyRAD	yes	yes	no	no	no	no
KU:Birds:6927	torotoro	meeki	Mt. Suckling, PNG	muscle	2011	hyRAD	yes	yes	no	no	no	no
NHMUK:Birds:1911.12.20.822	torotoro	pseuestes	Satakwa River, WP	toepad	1911	hyRAD	yes	yes	no	no	no	no
NHMUK:Birds:1911.12.20.823	torotoro	pseuestes	Mimika River, WP	toepad	1913	hyRAD	yes	yes	no	no	no	no

AMNH:Birds:301861	megarhyncha	wellsi	Weylendgep Kunupi, WP	toepad	1931	WGS	yes	yes	yes	yes	yes	yes
AMNH:Birds:302859	megarhyncha	wellsi	Mt. Derimapa, WP	toepad	1930	WGS	yes	yes	yes	yes	yes	yes
AMNH:Birds:339957	megarhyncha	wellsi	Bernhard Camp, WP	toepad	1939	WGS	yes	yes	yes	yes	yes	yes
AMNH:Birds:808986	megarhyncha	megarhyncha	Okapa, PNG	toepad	1965	WGS	yes	yes	yes	yes	yes	yes
ANWC:Birds:B02192	megarhyncha	megrhyncha	Western Highlands, PNG	toepad	1963	WGS	yes	yes	yes	yes	yes	yes
ANWC:Birds:B04293	megarhyncha	megarhyncha	Morobe, PNG	toepad	1966	WGS	yes	yes	yes	yes	yes	yes
ANWC:Birds:B25307	megarhyncha	megarhyncha	Wagau, PNG	toepad	1973	WGS	yes	yes	yes	yes	yes	yes
ANWC:Birds:B25652	megarhyncha	sellamontis	Huon Peninsula, PNG	toepad	1973	WGS	yes	yes	yes	yes	yes	yes
ANWC:Birds:B26140	megarhyncha	sellamontis	Huon Peninsula, PNG	toepad	1973	WGS	yes	yes	yes	yes	yes	yes
YPM:Birds:91444	megarhyncha	sellamontis	Huon Peninsula, PNG	toepad	1969	WGS	yes	yes	yes	yes	yes	yes
AMNH:Birds:329539	torotoro	ochracea	Fergusson Island, PNG	toepad	1928	WGS	yes	yes	yes	yes	yes	yes
AMNH:Birds:329540	torotoro	ochracea	Fergusson Island, PNG	toepad	1928	WGS	yes	yes	yes	yes	yes	yes
AMNH:Birds:426095	torotoro	flavirostris	Fly River, PNG	toepad	1936	WGS	yes	yes	yes	yes	yes	yes
AMNH:Birds:426096	torotoro	flavirostris	Fly River, PNG	toepad	1936	WGS	yes	yes	yes	yes	yes	yes
AMNH:Birds:426121	torotoro	meeki	Wassi Kussi River, PNG	toepad	1937	WGS	yes	yes	yes	yes	yes	yes
AMNH:Birds:637460	torotoro	tentelare	Aru Islands, WP	toepad	1900	WGS	yes	yes	yes	yes	yes	yes
AMNH:Birds:637471	torotoro	meeki	Simbang, PNG	toepad	1899	WGS	yes	yes	yes	yes	yes	yes
AMNH:Birds:637511	torotoro	flavirostris	Cape York Peninsula, AU	toepad	1913	WGS	yes	yes	yes	yes	yes	yes
ANWC:Birds:B07293	torotoro	torotoro	East Sepik Province, PNG	toepad	1966	WGS	yes	yes	yes	yes	yes	yes
ANWC:Birds:B07499	torotoro	torotoro	East Sepik Province, PNG	toepad	1966	WGS	yes	yes	yes	yes	yes	yes

518   **Table 2.** Demographic model test results.

Model	Log-likelihood	AIC
SC	-564.062	1140.124
AM	-590.0233	1192.047
SIg	-611.0493	1228.099
IMg	-617.116	1244.232
IM	-631.8897	1273.779
SI	-637.185	1280.37
AMg	-653.5472	1319.094
SCg	-716.0587	1444.117

Figure 1. Genomic and phenotypic data provide evidence of assortative mating. A)
Sampling localities for *Syma* kingfishers across New Guinea and Australia, color-coded by
genotype PC1 and scaled by number of individuals. B) Illustration of *S. megarhyncha* (top)
and *S. torotoro* (bottom), by Kevin Epperly. C) Principal component analysis of genotypes,
clustered by the best fit *k*-means result (*K*=3) and color-coded by the mean PC1 value for all
individuals in a given cluster. D) The first principal component of bioacoustic parameters
measured from vocalizations is bimodally distributed by species. E) The first principal
component of morphological data is bimodally distributed by species. F) A time-calibrated ND2
phylogeny supports reciprocal monophyly of *S. torotoro* and a clade with *S. megarhyncha* and *S.*
t. ochracea. G) SVDquartets nuclear DNA phylogeny. H) SVDquartets species tree, using *k*-
means clustering results and mitochondrial DNA clades as taxa.

Fig. 2. Evidence for introgression between species. A) Bootstrapped results for D tests for introgression across all three possible species tree topologies. The dashed red line indicates 0, the expectation under incomplete lineage sorting alone. B) Inferred migrants per generation from the best-fit model of secondary contact in *moments*. C) Inferred timing in years of the original divergence event between *S. torotoro* and *S. megarhyncha* ("Tsplit"), the duration of the initial period of isolation ("T0"), and the period of secondary contact ("T1") from *moments*. D) Inferred census population size estimates from *moments*.

552 **Fig. 3. Heterogeneous genomic divergence.** A) Relative divergence (F_{ST}) in 50 kb sliding
553 windows across the genome with 10 kb overlap. Outliers at an arbitrary 0.005 significance level
554 are highlighted in red. B) The genomic landscape of divergence in four chromosomes with
555 strong F_{ST} peaks. C) The correlation between F_{ST} and D_{XY} in the same four chromosomes.

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