

1 Title:

2 Gut Microbiome Functional and Taxonomic Diversity within an Amazonian semi-
3 nomadic hunter-gatherer group

4

5 Conteville, LC¹ ; Oliveira-Ferreira J² ; Vicente, AC¹

6

7 ¹Laboratory of Molecular Genetics of Microorganisms, Oswaldo Cruz
8 Institute/Fiocruz, Av. Brasil 4365, Manguinhos, Rio de Janeiro, Brazil

9 ²Laboratory of Immunoparasitology, Oswaldo Cruz Institute/Fiocruz, Av. Brasil
10 4365, Manguinhos, Rio de Janeiro, Brazil

11

12 Liliane Costa Conteville: lilianeconteville@gmail.com

13 Joseli Oliveira-Ferreira: lila@ioc.fiocruz.br

14 Ana Carolina Paulo Vicente: anapaulo@ioc.fiocruz.br

15

16 Corresponding author:

17 Liliane Costa Conteville: lilianeconteville@gmail.com

18

19

20

21

22

23 **Abstract**

24 **Background:** Human gut microbiome profiles have been associated with
25 human health and disease. These profiles have been defined based on
26 microbes' taxonomy and more recently, on their functionality. Human groups
27 that still maintain traditional modes of subsistence (hunter-gatherers and rural
28 agriculturalists) represent the groups non-impacted by urban-industrialized
29 lifestyles, and therefore study them provide the basis for understanding the
30 human microbiome evolution. The Yanomami is the largest semi-nomadic
31 hunter-gatherer group of the Americas, exploring different niches of the Amazon
32 rainforest in Brazil and Venezuela. In order to extend the analysis of this unique
33 and diverse group, we focused on the gut microbiome of the Yanomami from
34 Brazil and compared with those from Venezuela, and also with other traditional
35 groups from the Amazon, considering taxonomic and functional profiles.

36 **Results:** A diversity of taxonomic biomarkers were identified to each South
37 American traditional group studied, including the two Yanomami groups, despite
38 their overall similarity in the taxonomic gut microbiome profiles. Broader levels
39 of functional categories poorly discriminated traditional and urban-industrialized
40 groups. Interestingly, a diversity was observed with the stratification of these
41 categories, clearly segregating those groups. The Yanomami/Brazil gut
42 microbiome presented unique functional features, such as a higher abundance
43 of gene families involved in regulation/cell signaling, motility/chemotaxis, and
44 virulence, contrasting with the microbiomes from the Yanomami/Venezuela and
45 other groups.

46 **Conclusions:** Our study revealed biomarkers, taxonomic and functional
47 differences between the gut microbiome of Yanomami/Brazil and

48 Yanomami/Venezuela individuals. This intra-Yanomami group diversity was
49 accessed due to the increase number of individuals and group studied. These
50 differences may reflect their semi-nomadic behavior, as well as, the local and
51 seasonal diversity of the vast rainforest niche they explore, despite their shared
52 cultural and genetic background. Overall, their microbiome profiles are shared
53 with South American and African traditional groups, probably due to their
54 lifestyle. The unique features identified within the Yanomami highlight the bias
55 imposed by underrepresented sampling, and factors such as variations over
56 space and time (seasonality) that impact, mainly, the hunter-gatherers.
57 Therefore, to reach knowledge about human microbiome variations and their
58 implications in human health, it is essential to enlarge data concerning the
59 number of individuals, as well as the groups representing different lifestyles.

60

61 **Keywords**

62 Gut microbiome, hunter-gatherers, semi-nomadic, Yanomami, Amerindian,
63 westernization, biomarkers, functionality, taxonomic

64

65 **Background**

66 The transition of the traditional modes of subsistence to the current western
67 lifestyles that occurred with the advent of modern practices (urbanization and
68 industrialization) brought wide differences in diet and environment, factors
69 proposed to be the main determinants of the gut microbiome composition. In
70 fact, cross-population studies have demonstrated distinct taxonomic and
71 functional profiles between the gut microbiome of hunter-gatherers/rural

72 agriculturalists and urban-industrialized human groups. The main differences
73 among the gut microbiome of westernized and non-westernized groups are that
74 individuals from traditional communities harbor a more diverse gut microbiome,
75 with higher levels of fiber-degrading bacteria, and unique taxa that are depleted
76 in urban-industrialized populations [1-10]. The lifestyle aspects that
77 characterize most of the westernized groups, concerning diet, environment,
78 sedentary practices, among others, shape the gut microbiome, defining some
79 taxonomic profiles that have been associated with an increased risk of
80 metabolic and chronic disorders that affect modern populations [11]. Studies
81 focusing on the differences between traditional and urban-industrialized groups
82 may reveal diets and bacterial/archaeal taxa that can be helpful in the
83 development of prebiotics and probiotics for modern disorders prevention and
84 treatment. Considering that human groups that live in a non-western lifestyle
85 are in decline, the study of the remaining traditional groups constitutes an
86 extraordinary opportunity to explore and unravel the human gut microbiome
87 before modernization.

88 The Amazon region is the largest tropical wilderness area in the world,
89 covering ~ 7 million km². This region includes the most extensive and preserved
90 rainforest in the world (the Amazon Rainforest), vast areas of scrub-savannah
91 that dominate the headwaters of the Brazilian and Guyana shields, as well as
92 the Andes highlands, which are characterized by tundra-like grassy tussocks
93 called the Puna [12]. Moreover, this region presents elevations ranging from sea
94 level at the river's mouth to an altitude of 6,500 meters in the Andes [13].
95 Having such high variable geomorphology, climate and vegetation cover, and
96 harboring estimated 400-500 indigenous Amazonian Indian groups

97 (Amerindians), this region offers a unique scenario for microbiome studies.
98 These groups live in the same geographical region but explore distinct
99 ecological niches, present distinct dietary habits, culture, language and degrees
100 of isolation [14, 15]. This diversity may reflect in the gut microbiome composition
101 and functionality, expressing the adaptation to evolutionary and ecological
102 constraints of each site inhabited, despite being non-western populations.

103 The Yanomami is the largest indigenous semi-isolated group in the Amazon
104 to maintain a traditional system of production based on hunting, fishing,
105 gathering, and swidden horticulture [16]. They inhabit an area of 192,000 km² in
106 the Amazon region encompassing the Brazil and Venezuela border. Of the
107 estimated 40,000 Yanomami, approximately 26,000 live in the 37,260 m²
108 reserve in Brazil and another 16,000 in Venezuela. Even though they represent
109 a single semi-nomadic ethnic group of hunter-gatherers, they speak four
110 different languages of the same family and live in villages located at sea level
111 as well as on high mountains in a huge area in the Amazon [17]. Their diet is
112 low in fat and salt, and high in fruits, fiber, and sylvatic animals. Atherosclerosis
113 and obesity are virtually unknown among semi-isolated Yanomami, having low
114 blood pressure, with no apparent increase as they age [18].

115 A previous study with uncontacted Yanomami from Venezuela revealed
116 some aspects from the gut microbiome of this group [4]. In order to go deeper
117 into the characterization of gut microbiome among traditional subsistence
118 groups, we studied semi-isolated hunter-gatherers Yanomami individuals from
119 Brazil. For this, we generated and analyzed metagenomic data of Yanomami
120 from Brazil, and performed comparative analyses with those from Venezuela,
121 other traditional groups from the Amazon (the Matses and the Tunapuco), as

122 well as an urban-industrialized group (Figure 1). The Matses and the Tunapuco
123 inhabit the borders of Peru that comprise the Amazon Region, but their lifestyles
124 and environment are strikingly different. The Matses are traditional hunter-
125 gatherers living by the sea level, while the Tunapuco is a rural agriculturalist
126 community situated in the Andes highlands [6].

127 We hypothesized that, since the previous Yanomami group studied was
128 uncontacted and lived in a remote area in Venezuela [4], their gut microbiome
129 would present unique features in comparison with the large semi-isolated
130 Yanomami group living in the Brazilian Amazon. Even though they are hunter-
131 gatherers, their diet varies depending on the niche explored, since they live in
132 areas ranging from near rivers to frankly mountainous regions. Moreover, we
133 explored the taxonomy and functionality of these microbiomes, contributing to
134 the understanding of features that can affect the health outcomes observed in
135 modern populations.

136

137 **Results**

138

139 **Intra and Inter Individual Diversity of the Gut Microbiomes**

140

141 To unravel the gut microbiome diversity of Yanomami from Brazil
142 individuals (Yanomami/Brazil, $n = 15$), we performed alpha and beta diversity
143 analyses based on the bacterial genera profile identified by Kraken. For these
144 analyses, we also reanalyzed and compared gut microbiome data gathered
145 from other South American traditional communities: the Yanomami from the
146 Venezuelan Amazon (Yanomami/Venezuela, $n = 8$) [4], the Matses from the

147 Peruvian Amazon (n= 24) [6], the Tunapuco from the Andean highlands (n= 12)
148 [6]; and a representative group of urban individuals from United States (US, n=
149 44) [19,20].

150 There was no statistically difference between the Yanomami/Brazil and
151 Yanomami/Venezuela regarding intra and inter diversity (alpha- and beta-
152 diversity, respectively) of gut microbiome, although the group from Brazil had
153 the lowest alpha-diversity values. However, the Yanomami individuals showed
154 the lowest bacterial alpha-diversity among the traditional groups and all the
155 traditional human groups presented higher bacterial diversity compared to the
156 urban individuals (Figure 2a). Regarding the beta-diversity, the Yanomami/Brazil
157 presented the highest interpersonal variation, and the urbans presented the
158 lowest, although there was no significant difference with the
159 Yanomami/Venezuela group (Figure 2b). A clear segregation was observed
160 among the semi-isolated and westernized individuals (PERMANOVA, $P=0.001$)
161 based on Principal Coordinate Analysis (PcoA) generated with Bray-Curtis
162 distances. In addition, a higher dispersion of Yanomami/Brazil and
163 Yanomami/Venezuela samples was observed, stressing their higher
164 interpersonal variation (Figure 2c).

165

166 **Microbiomes Taxonomic Characterization**

167

168 In order identify which bacterial and archaeal taxa differentiate the
169 traditional groups from the urban group, the microbiomes were compared at
170 both phylum and genus scales. Thirty-two bacterial phyla were identified, with
171 16 phyla having significant differences in the relative abundances among the

172 groups (Kruskal-Wallis test: $P < 0.0001$). Considering the traditional and urban
173 groups, a clear difference at the phylum level was observed, with the former
174 having a higher biodiversity characterized by *Firmicutes*, *Proteobacteria*,
175 *Bacteroidetes* and *Spirochaetes*, while the urban group is mainly characterized
176 by *Bacteroidetes* (Figure 3a).

177 The Yanomami/Brazil as well as the other traditionalists individuals follow a
178 trend in which they have higher *Firmicutes* and lower *Bacteroidetes* levels, while
179 the opposite was observed in the urban individuals (Figure 3b). Even though,
180 the *Firmicutes* in each traditional group was characterized by distinct genera, no
181 prevalent genus was consistently observed in the groups. In fact, all traditional
182 groups presented different genera from the *Firmicutes* phylum as biomarkers
183 (Figure 3c). Genera from *Bacteroidetes* phylum were demonstrated to be the
184 biomarkers of the urban group (Figure 3c).

185 Distinctly from the other groups, *Proteobacteria* was the most prevalent
186 phylum among the Yanomami individuals, despite their geographic origin (Brazil
187 and Venezuela). The most abundant genera of this phylum in the traditional
188 groups were *Escherichia* and *Klebsiella*, however, there is a contrasting higher
189 abundance of *Escherichia* and *Ralstonia* genera in the Yanomami/Brazil, and
190 therefore, they were defined as Yanomami/Brazil biomarkers. On the other
191 hand, *Neisseria* and *Desulfovibrio* were defined as Yanomami/Venezuela
192 biomarkers, while *Klebsiella* was the biomarker of the Matses group. It is
193 noteworthy that *Cutibacterium* from the Actinobacteria phylum and *Akkermansia*
194 from the Verrucomicrobia phylum were also deemed as the biomarkers of the
195 Yanomami/Brazil (Figure 3c). Besides that, the Yanomami/Brazil, similarly with
196 the other semi-isolated, present *Treponema* and *Brachyspira*, two genera from

197 the Spirochaetes that were not detected in the urban group.

198 With respect to Archaea, we observed that the most abundant genus in the
199 traditional groups was *Methanobrevibacter*, comprising ~70% of all archaea
200 classified reads, while in the urban population, there were a high abundance of
201 *Methanoculleus* and *Methanothermobacter*, all methane-producers
202 archaea (Figure 3d).

203

204 **Microbiomes Functional Characterization**

205

206 For functional characterization, the metagenomic reads of all groups were
207 assigned to gene families from the SEED database, and were categorized for
208 their functional roles in subsystems with 3 levels of resolution, in which level 1
209 represent the broader category. We observed a segregation between the
210 traditional and urban groups regarding the abundance of functions at the level 3
211 subsystems. Interestingly, among the traditional groups, the Matses and
212 Yanomami/Brazil individuals exhibits a clear segregation, however the
213 Yanomami/Brazil present a more disperse pattern, indicating a quite diverse
214 functional characteristic concerning functions at the level 3 subsystem (Figure
215 4a).

216 The most abundant metabolic functions present in the microbiome of all
217 groups at the level 1 were the metabolism of carbohydrates and proteins, while
218 the membrane transport was also a main function in the urban group, but was
219 depleted in the traditional groups (Figure 4b). We observed differences in the
220 carbohydrate metabolic functions between the traditional and the urban groups.
221 At level 2, the main carbohydrate metabolic functions in the traditional groups

222 belonged to the central carbohydrate metabolism, with functions as pyruvate
223 metabolism and glycolysis/gluconeogenesis being the most predominant; while
224 in the urbans, the major functions were the monosaccharides and di-
225 /oligosaccharides metabolism (Figure 4b). In monosaccharides metabolism,
226 there was also differences between the groups: D-galacturonate/D-glucuronate
227 and xylose use were the most abundant functions in the traditional groups,
228 while mannose metabolism was the most abundant in the urban group. The di-
229 /oligosaccharides microbiome metabolism of US individuals is mainly driven by
230 functions associated with lactose utilization, which was depleted in the
231 microbiome of traditional groups. Regarding protein metabolism, there was no
232 difference among the groups' microbiome, with the most abundant functions
233 being those associated with protein biosynthesis and degradation. We also
234 observed differences in metabolic pathways related to cofactors, vitamins,
235 prosthetic groups and pigments among the groups, the major gene families
236 found in the microbiome of the Yanomami/Brazil and Yanomami/Venezuela
237 were associated with folate/pterines, in the Matses and Tunapuco was
238 riboflavin, and in the US group was tetrapyrroles (Figure 4b).

239 Interestingly, at level 1 subsystems, the microbiome of the Yanomami/Brazil
240 was distinct from the other groups due to its significant higher abundance of
241 gene families involved in regulation/cell signaling, motility/chemotaxis, and
242 virulence (Figure 4b). The regulation/cell signaling functions in the
243 Yanomami/Brazil is driven by abundance of programmed cell death and toxin-
244 antitoxin systems, while the motility/chemotaxis function is driven by the
245 presence of genes involved in flagellar motility in prokaryotes. The most
246 abundant virulence function at subsystems 3 in the Yanomami/Brazil is cobalt,

247 zinc and cadmium resistance.

248

249 **Discussion**

250

251 The gut microbiome is a diverse ecosystem with multiple metabolic and
252 immune functions associated with the diet and lifestyle of the host [11,21].
253 Therefore, considering the current variety of lifestyles and diets in human
254 society, many aspects concerning the gut microbiome composition and
255 functionality are yet to be accessed and explored to understand the influence of
256 these factors in the gut microbiome. The study of human groups that still
257 maintain traditional modes of subsistence (hunter-gatherers and rural
258 agriculturalists) provides valuable information regarding the ancestral
259 microbiome that existed before the urbanization and industrialization impacted
260 human diet and lifestyle. So far, few studies explored worldwide traditional
261 groups gut microbiome, and it is essential to enlarge data concerning the
262 number of individuals, as well as different groups.

263 Therefore, in the present study, we characterized the gut microbiome of 15
264 semi-isolated Yanomami individuals from Brazil, and compared with other South
265 American traditional groups (uncontacted Yanomami from Venezuela, the
266 Matsigenka and the Tunapuco) as well as an urban-industrialized group (US)
267 [4,6,19,20], enlarging the number of Yanomami individuals analyzed, as well as
268 of hunter-gatherers. These traditional groups explore different Amazonian
269 niches and contrast with the US group, which lives in a densely populated
270 urbanized and industrialized society with access to medical care and high
271 hygiene standards.

272 Consistent with previous studies [1-10, 22], our analysis point to a higher
273 bacterial diversity in the traditional groups, with diverse taxonomic and
274 functional features that distinguish them from urban-industrialized individuals.
275 Microbiomes harboring a high diversity showed a positive association with
276 health, as consequence of the presence of a higher global metabolic potential,
277 providing the host with a wide range of health-relevant metabolites [23, 24].
278 Besides that, the microbiomes of the traditional South American groups share
279 features with traditional African groups (Western, Central and Eastern Africa):
280 they are also more diverse than the urbans, are enriched in *Proteobacteria*, with
281 the presence of some *Spirochaetes* that are depleted in industrialized
282 populations (*Treponema* and *Brachyspira*) [1-10, 22]. Despite the South
283 American and the African groups being in different continents and having
284 distinct genetic origin, they maintain a traditional mode of subsistence and do
285 not have access to processed and refined food in their daily diet [21]. This
286 corroborates that population lifestyle and diet are the major determinants of the
287 gut microbiome composition and diversity, overruling genetic backgrounds and
288 geographic origin.

289 The taxonomic analysis of the South-American traditional groups
290 demonstrated a common profile at bacterial genera level, even though each
291 group presented a specific set of biomarkers. Interestingly, some biomarkers
292 converge in their functional profile, e.g. *Roseburia*, *Anaerostipes*, *Eubacterium*,
293 *Flavonifactor*, biomarkers of the Yanomami/Brazil, Yanomami/Venezuela,
294 Matses and Tunapuco, respectively, are butyrate-producing bacteria. Butyrate is
295 an anti-inflammatory short chain fatty acid (SCFA) that induces mucin synthesis,
296 contributing to colon health and gut integrity [25, 26]. In contrast, the urban-

297 industrialized biomarkers produce SCFAs other than butyrate, such as
298 propionate, acetate, and succinate, which, in high proportions, may increase gut
299 permeability, leading to a further unhealthy status [27]. Other biomarker of the
300 Yanomami/Brazil group is Akkermansia, a mucin degrader, which has been
301 associated with healthier metabolic status and better clinical outcomes [28,29].

302 Broader levels of functional categories poorly discriminated traditional and
303 urban-industrialized groups. Interestingly, the stratification of these categories
304 clearly segregated those groups. Differences were identified at level 3 of
305 monosaccharides metabolism, where the main functions in the traditionalists
306 and urbans were xylose and mannose metabolism, respectively. Xylans and
307 Mannans, polysaccharides of xylose and mannose, are the two major classes of
308 hemicelluloses that accumulate in plant secondary walls [30]. Interestingly,
309 recent studies with mouse models revealed that mannose increased the
310 Bacteroidetes to Firmicutes ratio in the gut, a characteristic observed in urban-
311 industrialized groups [31]. On the other hand, Treponema, a prevalent genus in
312 traditional populations that consume polysaccharide-rich diets, is a key xylan-
313 degrader [32]. Another difference observed in the US versus traditional groups
314 was the lactose utilization, which is enriched in the former and depleted in the
315 latter group. This difference may be related to the lack of intake of dairy in the
316 traditional groups [6]. Within the traditional groups, there was differences at
317 level 3 of biosynthesis of vitamins: both Yanomami groups presented an
318 enrichment in folate biosynthesis while the Matses and Tunapuco presented an
319 enrichment in riboflavin biosynthesis, as well as in the US group. Riboflavin is
320 the most commonly synthesized vitamin in the gut [33], and has been
321 associated with the immune response through the activation of T-cells [34].

322 Folate is associated with high-fiber and low-fat diets [35], which agrees with
323 Yanomami diet from the present study.

324 The microbiome of the Yanomami/Brazil is unique concerning the presence
325 of higher levels of functions associated with virulence, driven by the cobalt, zinc
326 and cadmium resistance. Cobalt is commonly distributed in nature and has a
327 biological role as metal constituent of the vitamin B12, however, excessive
328 exposure induces adverse health effects [36]. Zinc is an essential nutrient and
329 play a role in gene expression, biomolecular activity and structural DNA
330 stabilization [37]. Cadmium is a non-essential element, representing an
331 environmental hazard to human health when contaminates the food chain,
332 causing cumulative toxic effects in diverse human organs [38]. Cadmium and
333 Zinc are present in mine discharges, which disperses into air, water and soils,
334 contaminating areas nearby mines [39]. However, in Yanomami/Brazil area,
335 Cadmium contamination may occur as consequence of the continuous
336 discharge of batteries anywhere by the Yanomami along decades.

337

338 **Conclusions**

339

340 Exploring the gut microbiome of traditional groups is challenging, mainly
341 due the difficult to access them. These groups are important, since they
342 represent living representatives of ancestral behaviors/dietary long lost for a
343 long time in the westernized groups. Our study revealed that even within very
344 close and related traditional groups (as Yanomami/Brazil and
345 Yanomami/Venezuela), there are taxonomic differences that distinguish their gut
346 microbiome. These variations may reflect their nomadic behavior, as well as,

347 the local and seasonal diversity of the vast rainforest niche they explore, despite
348 their shared cultural and genetic background. Overall, their microbiome profiles
349 are shared with South American and African traditional groups, probably due to
350 their diet and lifestyle. This highlight the need to characterize larger sampling of
351 human microbiomes, considering not only distinct lifestyle but also a broad
352 population representing a particular lifestyle. Thus, we expect novel insights into
353 the diverse factors that are associated with microbiome composition and human
354 health.

355

356 **Methods**

357

358 **Study Participants and Sample Collection**

359

360 The protocol of this study was reviewed and approved by Oswaldo Cruz
361 Foundation's Ethics Research Committee N ° 638/11 and by the National Ethics
362 the Committee in Research – CONEP N° 16907. Before participating in the
363 study, a bilingual interpreter (a Yanomami native who spoke Portuguese)
364 explained the leaders and/or Indigenous representatives, the purpose and
365 importance of the study, the procedures to be carried out and finally requested
366 permission by fingerprint consent of each participant. Participants were
367 requested to provide a morning faecal sample and a labelled screw-capped
368 plastic container was provided. A single stool sample was collected from each
369 subject on the following day and samples were stored in separate sterile feces
370 containers. At the time of the collection, age and sex information of the
371 individuals were also acquired. These details are summarized in Table S1.

372 **DNA extraction, library preparation, and sequencing**

373

374 Total DNA was extracted from 15 stool samples with FastDNA® SPIN Kit
375 (MP Biomedicals), following the manufacturer's instructions. DNA concentration
376 were evaluated using Qubit® 2.0 Fluorometer (Life Technologies). Metagenomic
377 libraries were constructed with TruSeq DNA Sample Preparation v2 Kit following
378 the standard protocols. Purified libraries were sequenced on a HiSeq® 2500
379 sequencer (Oswaldo Cruz Foundation High-throughput sequencing Platform) in
380 two batches, producing a total of ~ 219 million reads, with an average of ~ 14
381 million reads per sample.

382

383 **Bioinformatic Processing**

384

385 Raw reads were trimmed and filtered (phred quality < 20 , length < 30)
386 using Trimmomatic [40]. The remaining reads (~ 206 million reads) were
387 mapped to a human reference genome (Hg38) using Bowtie2 [41]. Non-host
388 reads (~ 198 million reads) were used in further analysis. Besides the
389 metagenomes generated in this study, we also analyzed shotgun metagenomic
390 data from previously published studies: two hunter-gatherer communities
391 (Yanomami from Venezuela, n=8 [4] ; Matses, n=24 [6]), a rural agricultural
392 community (Tunapuco, n=12 [6]) and urban populations (USA, n=44 [19,20]).
393 These datasets were sequenced on Illumina platforms and bioinformatic
394 processing was performed in parallel with the data generated in this study.

395 Taxonomic classification was performed by Kraken [42], using a database
396 of whole genomes of bacteria and archaea from NCBI. Functional classification

397 were classified by SUPER-FOCUS [43] based on the genes families from the
398 SEED database. Linear discriminant analysis (LDAs) were performed using
399 LEfSe [44] to detect bacterial genera that characterize the differences between
400 the groups (LDA score of > 4.0).

401 For general data manipulation and statistical analysis we employed the
402 vegan [45] and phyloseq [46] packages in R. Shannon index of alpha-diversity
403 was estimated for each metagenome, with pairwise Wilcoxon test being used
404 for statistical difference evaluation. Beta diversity was estimated using Bray–
405 Curtis dissimilarity and permutational multivariate analysis of variance
406 (PERMANOVA) were performed with 999 permutations to estimate a P-value for
407 differences among traditional and westernized groups.

408

409 **List of abbreviations**

410

411 **LDA:** Linear discriminant analysis

412 **LefSe:** Linear discriminant analysis effect size

413 **PcoA:** Principal coordinates analysis

414 **PERMANOVA:** Permutational multivariate analysis of variance

415 **SCFA:** Short chain fatty acid

416

417

418 **Declarations**

419 **Ethics approval and consent to participate**

420 The protocol of this study was reviewed and approved by Oswaldo Cruz
421 Foundation's Ethics Research Committee N^o 638/11 and by the National Ethics
422 the Committee in Research – CONEP N^o 16907.

423

424 **Consent for publication**

425 Not applicable.

426

427 **Availability of data and materials**

428 The quality-filtered metagenomic sequences are available on the NCBI under
429 the BioProject PRJNA527208.

430

431 **Competing interests**

432 The authors declare that they have no competing interests.

433

434 **Funding**

435 This study was financed in part by the Coordenação de Aperfeiçoamento de
436 Pessoal de Nível Superior (CAPES) – Finance code 001, CNPQ, and PAEF
437 (IOC-023-FIO-18-2-47).

438

439 **Authors' contributions**

440 JO collected the samples. LCC processed the samples and analyzed the data.
441 LCC and ACPV interpreted the data and drafted the manuscript. All authors
442 revised it. All authors approve the final version to be published and agree to be
443 accountable for the work.

444

445 **Acknowledgements**

446 We are especially grateful to the Yanomami people and we also thank the
447 health personnel of the Distrito Sanitário Especial Indígena Yanomami for
448 overall support during field work. We are particularly thankful to Dr. Edson
449 Delatorre for the discussion of the manuscript and help with a figure.

450

451 **References**

452 1. De Filippo C, Cavalieri D, Di Paola M, Ramazzotti M, Poullet JB, Massart S,
453 et al. Impact of diet in shaping gut microbiota revealed by a comparative study
454 in children from Europe and rural Africa. *Proc Natl Acad Sci U S A*.
455 2010;107(33):14691-6.

456 2. Yatsunencko T, Rey FE, Manary MJ, Trehan I, Dominguez-Bello MG,
457 Contreras M, et al. Human gut microbiome viewed across age and geography.
458 *Nature*. 2012;486(7402):222-7

459 3. Schnorr SL, Candela M, Rampelli S, Centanni M, Consolandi C, Basaglia G,
460 et al. Gut microbiome of the Hadza hunter-gatherers. *Nat Commun*.
461 2014;5:3654.

- 462 4. Clemente JC, Pehrsson EC, Blaser MJ, Sandhu K, Gao Z, Wang B, et al. The
463 microbiome of uncontacted Amerindians. *Sci Adv.* 2015;3;1(3).
- 464 5 . Martínez I, Stegen JC, Maldonado-Gómez MX, Eren AM, Siba PM, Greenhill
465 AR, et al. The gut microbiota of rural papua new guineans: composition,
466 diversity patterns, and ecological processes. *Cell Rep.* 2015;28;11(4):527-38.
- 467 6 . Obregon-Tito AJ, Tito RY, Metcalf J, Sankaranarayanan K, Clemente JC,
468 Ursell LK, et al. Subsistence strategies in traditional societies distinguish gut
469 microbiomes. *Nat Commun.* 2015;6:6505.
- 470 7. Rampelli S, Schnorr SL, Consolandi C, Turrioni S, Severgnini M, Peano C, et
471 al. Metagenome Sequencing of the Hadza Hunter-Gatherer Gut Microbiota.
472 *Curr Biol.* 2015;25(13):1682-93.
- 473 8. Gomez A, Petrzekova KJ, Burns MB, Yeoman CJ, Amato KR, Vlckova K, et
474 al. Gut Microbiome of Coexisting BaAka Pygmies and Bantu Reflects Gradients
475 of Traditional Subsistence Patterns. *Cell Rep.* 2016;14(9):2142-2153.
- 476 9 . De Filippo C, Di Paola M, Ramazzotti M, Albanese D, Pieraccini G, Banci
477 E, et al. Diet, Environments, and Gut Microbiota. A Preliminary Investigation in
478 Children Living in Rural and Urban Burkina Faso and Italy. *Front Microbiol.*
479 2017;8:1979.
- 480 10. Rothschild D, Weissbrod O, Barkan E, Kurilshikov A, Korem T, Zeevi D, et
481 al. Environment dominates over host genetics in shaping human gut microbiota.
482 *Nature.* 2018;555(7695):210-215
- 483 11. Zinöcker MK, Lindseth IA. The Western Diet-Microbiome-Host Interaction
484 and Its Role in Metabolic Disease. *Nutrients.* 2018;10(3):365.
- 485 12. GIWA. 2004. *Amazon Basin*. GIWA Regional assessment 40b. Global
486 International Waters Assessment. Barthem RB, Charvet-Almeida P, Montag

- 487 LFA, Lanna AE. University of Kalmar, Kalmar, Sweden. Global International
488 Waters Assessment
- 489 13. Braga B, Varella P, Gonçalves H. Transboundary Water Management of the
490 Amazon Basin. *International Journal of Water Resources Development*. 2011;
491 27:3;477-496.
- 492 14. Sioli H. The Amazon and its main affluents: hydrography, morphology of the
493 river courses and river types. In: Sioli H, editor. *The Amazon: limnology and*
494 *landscape ecology of a mighty tropical river and its basin*. Dordrecht: Dr. W.
495 Junk Publishers. 1984;127–166
- 496 15. Capobianco JPR, Veríssimo A, Moreira A, Sawyer D, Santos I, Pinto LP.
497 Biodiversidade na Amazônia Brasileira: avaliação e ações prioritárias para
498 conservação. Editora Estação Liberdade/Instituto Socioambiental, São
499 Paulo. (2001)
- 500 16. Albert B, LeTourneau FM. Ethnogeography and Resource Use among the
501 Yanomami: Toward a Model of “Reticular Space.” *Current Anthropology*.
502 2007;48:1–19.
- 503 17. Pithan OA, Confalonieri UEC, Morgado AF. A situação de saúde dos índios
504 Yanomámi: diagnóstico a partir da casa do índio de Boa Vista, Roraima, 1987 -
505 1989. *Cad Saude Publica*. Escola Nacional de Saúde Pública, Fundação
506 Oswaldo Cruz; 1991 Dec;7(4):563–80.
- 507 18. Mueller NT, Noya-Alarcon O, Contreras M, Appel LJ, Dominguez-Bello MG.
508 Association of Age With Blood Pressure Across the Lifespan in Isolated
509 Yanomami and Yekwana Villages. *JAMA Cardiol*. 2018. doi:
510 10.1001/jamacardio.2018.3676.

- 511 19. Human Microbiome Project (HMP) Consortium. Structure, function and
512 diversity of the healthy human microbiome. *Nature*. 2012;486(7402):207-14.
- 513 20. Lloyd-Price J, Mahurkar A, Rahnavard G, Crabtree J, Orvis J, Hall AB, et al.
514 Strains, functions and dynamics in the expanded Human Microbiome Project.
515 *Nature*. 2017;550(7674):61-66.
- 516 21. Gupta VK, Paul S, Dutta C. Geography, Ethnicity or Subsistence-Specific
517 Variations in Human Microbiome Composition and Diversity. *Front Microbiol*.
518 2017;8:1162.
- 519 22. Mancabelli L, Milani C, Lugli GA, Turrone F, Ferrario C, van Sinderen D, et
520 al. Meta-analysis of the human gut microbiome from urbanized and pre-
521 agricultural populations. *Environ Microbiol*. 2017;19(4):1379-1390.
- 522 23. Larsen OFA, Claassen E. The mechanistic link between health and gut
523 microbiota diversity. *Sci Rep*. 2018;8(1):2183.
- 524 24. Heinken A, Thiele I. Systematic prediction of health-relevant human-
525 microbial co-metabolism through a computational framework. *Gut Microbes*.
526 2015;6(2):120-30.
- 527 25. Burger-van Paassen N, Vincent A, Puiman PJ, van der Sluis M, Bouma J,
528 Boehm G, et al. The regulation of intestinal mucin MUC2 expression by short-
529 chain fatty acids: implications for epithelial protection. *Biochem J*. 2009;420:
530 211–219.
- 531 26. Rivièrè A, Selak M, Lantin D, Leroy F, De Vuyst L. Bifidobacteria and
532 Butyrate-Producing Colon Bacteria: Importance and Strategies for Their
533 Stimulation in the Human Gut. *Front Microbiol*. 2016;7:979.
- 534 27. Brown CT, Davis-Richardson AG, Giongo A, Gano KA, Crabb DB,
535 Mukherjee N, et al. Gut microbiome metagenomics analysis suggests a

- 536 functional model for the development of autoimmunity for type 1 diabetes. *PLoS*
537 *One*. 2011;6(10):e25792.
- 538 28. Png CW, Lindén SK, Gilshenan KS, Zoetendal EG, McSweeney CS, Sly LI,
539 et al. Mucolytic bacteria with increased prevalence in IBD mucosa augment in
540 vitro utilization of mucin by other bacteria. *Am J Gastroenterol*.
541 2010;105(11):2420-8.
- 542 29. Dao MC, Everard A, Aron-Wisnewsky J, Sokolovska N, Prifti E, Verger EO,
543 et al. *Akkermansia muciniphila* and improved metabolic health during a dietary
544 intervention in obesity: relationship with gut microbiome richness and ecology.
545 *Gut*. 2016;65(3):426-36.
- 546 30. Scheller HV, Ulvskov P. Hemicelluloses. *Annu Rev Plant Biol*. 2010;61:263-
547 89.
- 548 31. Sharma V, Smolin J, Nayak J, Ayala JE, Scott DA, Peterson SN, et al.
549 Mannose Alters Gut Microbiome, Prevents Diet-Induced Obesity, and Improves
550 Host Metabolism. *Cell Rep*. 2018;24(12):3087-3098.
- 551 32. Flint HJ, Bayer EA, Rincon MT, Lamed R, White BA. Polysaccharide
552 utilization by gut bacteria: potential for new insights from genomic analysis. *Nat*
553 *Rev Microbiol*. 2008 Feb;6(2):121-31.
- 554 33. Magnúsdóttir S, Ravcheev D, de Crécy-Lagard V, Thiele I. Systematic
555 genome assessment of B-vitamin biosynthesis suggests co-operation among
556 gut microbes. *Front Genet*. 2015;6:148.
- 557 34. Kjer-Nielsen L, Patel O, Corbett AJ, Le Nours J, Meehan B, Liu L, et al. MR1
558 presents microbial vitamin B metabolites to MAIT cells. *Nature*. 2012
559 ;491(7426):717-23.

- 560 35. Chan YM, Aufreiter S, O'Keefe SJ, O'Connor DL. Switching to a fibre-rich
561 and low-fat diet increases colonic folate contents among African Americans.
562 *Appl Physiol Nutr Metab.* 2019;44(2):127-132.
- 563 36. Leysens L, Vinck B, Van Der Straeten C, Wuyts F, Maes L. Cobalt toxicity
564 in humans-A review of the potential sources and systemic health effects.
565 *Toxicology.* 2017;387:43-56.
- 566 37. Bruins MR, Kapil S, Oehme FW. Microbial resistance to metals in the
567 environment. *Ecotoxicol Environ Saf.* 2000;45(3):198-207.
- 568 38. Hyder O, Chung M, Cosgrove D, Herman JM, Li Z, Firoozmand A, et al.
569 Cadmium exposure and liver disease among US adults. *J Gastrointest Surg.*
570 2013;17(7):1265-73.
- 571 39. Xue S, Shi L, Wu C, Wu H, Qin Y, Pan W, et al. Cadmium, lead, and arsenic
572 contamination in paddy soils of a mining area and their exposure effects on
573 human HEPG2 and keratinocyte cell-lines. *Environ Res.* 2017;156:23-30.
- 574 40. Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for Illumina
575 sequence data. *Bioinformatics.* 2014;30(15):2114–120
- 576 41. Langmead B, Salzberg SL. Fast gapped-read alignment with Bowtie 2.
577 *Nature methods.* 2012;9(4):357-359.
- 578 42. Wood DE, Salzberg SL. Kraken: ultrafast metagenomic sequence
579 classification using exact alignments. *Genome Biol.* 2014;15(3):R46.
- 580 43. Silva GG, Green KT, Dutilh BE, Edwards RA. SUPER-FOCUS: a tool for
581 agile functional analysis of shotgun metagenomic data. *Bioinformatics.*
582 2016;32(3):354-61.

583 44. Segata N, Waldron L, Ballarini A, Narasimhan V, Jousson O, Huttenhower
584 C. Metagenomic microbial community profiling using unique clade-specific
585 marker genes. Nat Methods. 2012;9(8):811-4.

586 45. Oksanen J, Blanchet FG, Friendly M, Kindt R, Legendre P, McGlinn D, et al.
587 vegan: Community Ecology PackageR package version 2.0-10.2013

588 46. McMurdie P, Holmes S. Phyloseq: an R package for reproducible interactive
589 analysis and graphics of microbiome census data. PLoS ONE. 2013;8:e61217

590

591

592

593

594

595

596

597

598

599

600

601

602

603

604

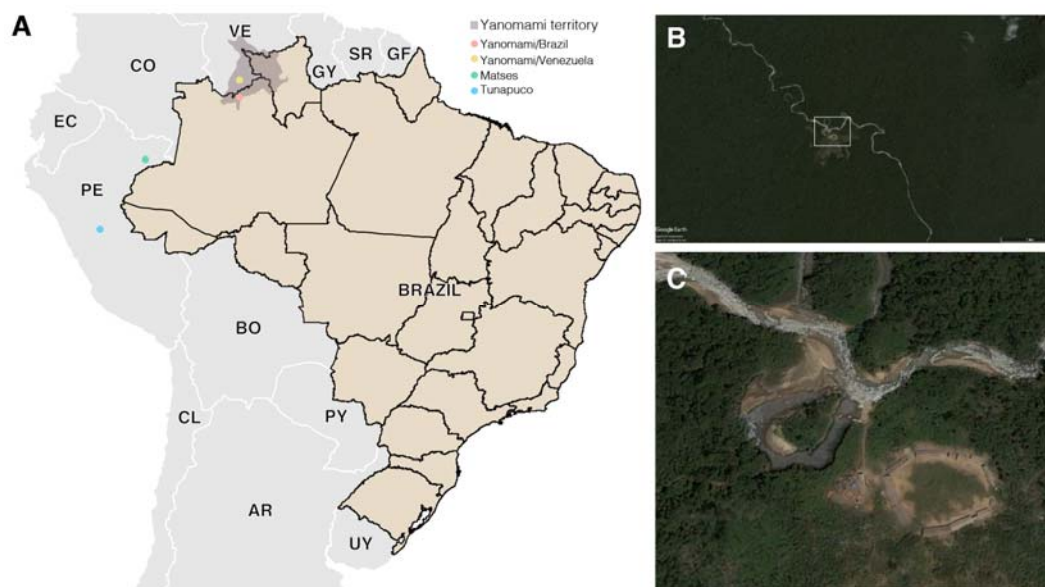
605

606

607

608 **Figures**

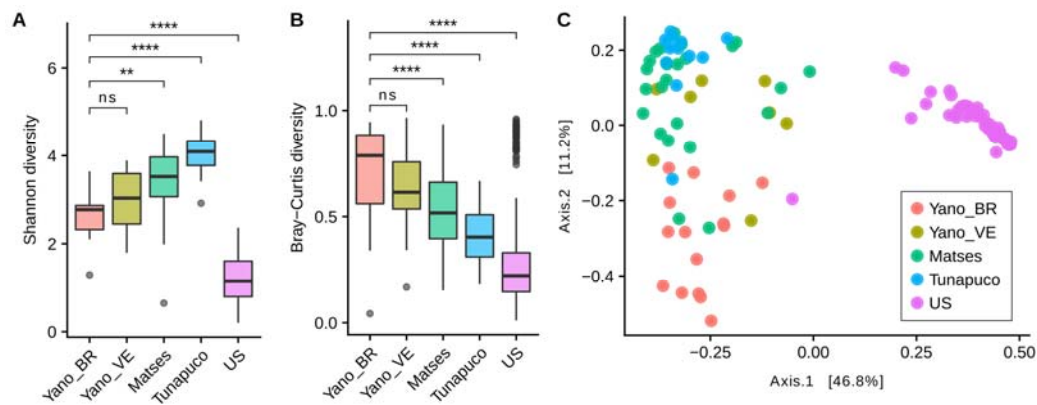
609



610

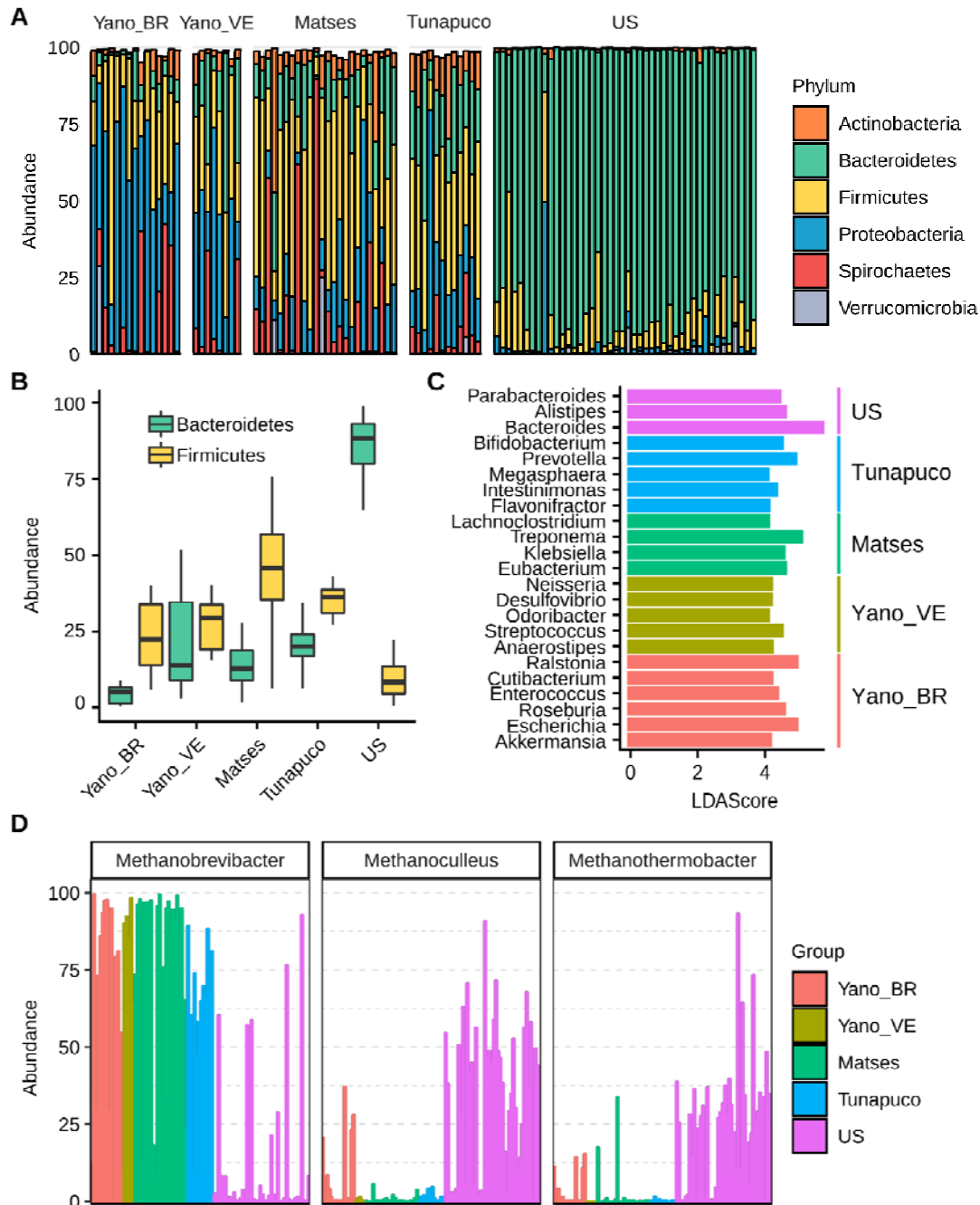
611 **Figure 1:** (A) Geographic locations of the South American traditional groups (B and C)
612 Satellite image of a Yanomami village in the Brazilian Amazon. Source: Google Earth
613 and Instituto Socioambiental (<https://acervo.socioambiental.org/>)

614



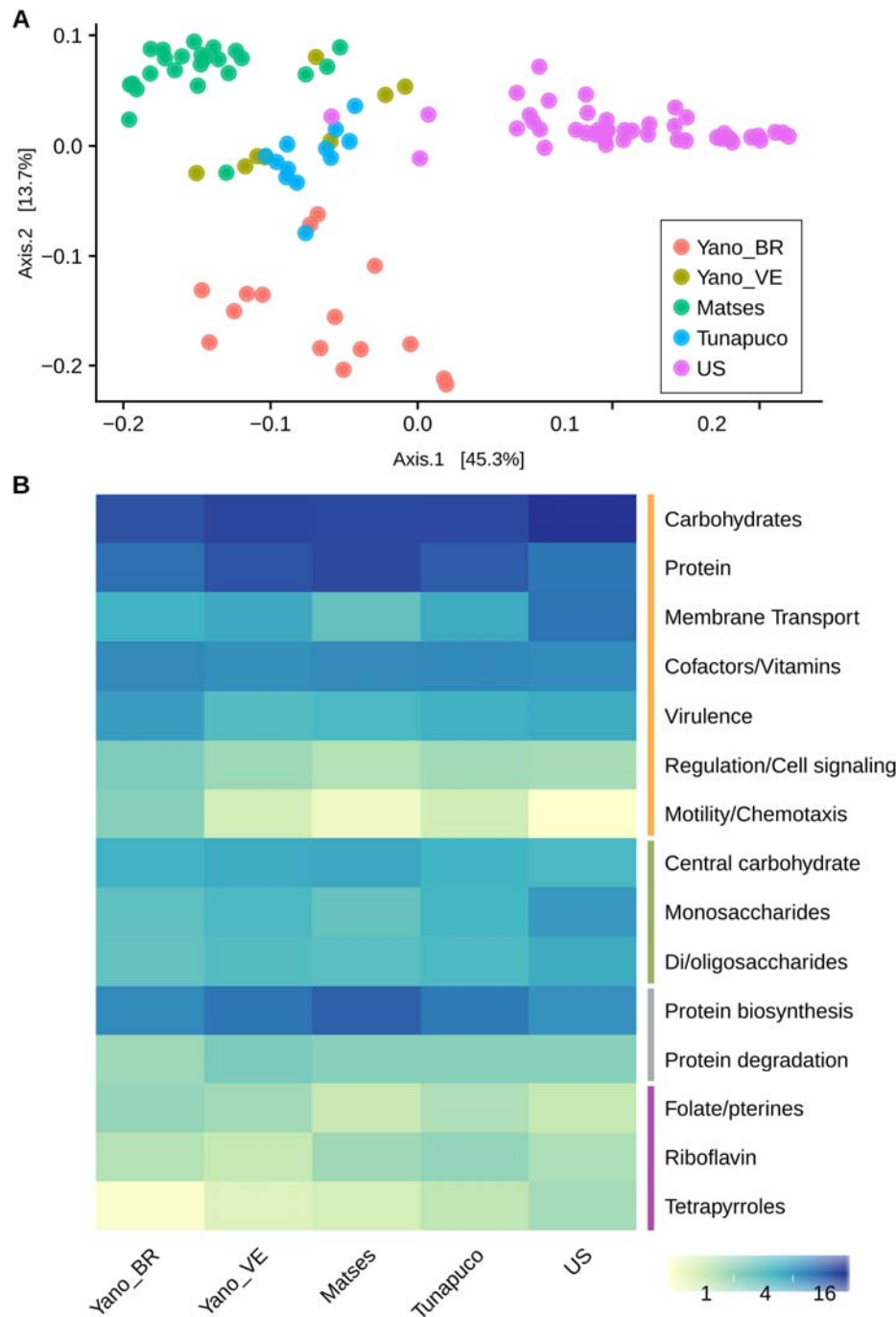
615

616 **Figure 2:** Alpha and beta-diversity comparisons of the gut microbiomes of each group.
617 Analyses were performed on genus-level taxa tables. ns = not significant, ** $P < 0.01$,
618 **** $P < 0.0001$ (Wilcoxon test) (A) Boxplot of the Shannon diversity of each group (B)
619 Bray-Curtis distances within each group (C) Principal coordinate analysis of Bray-Curtis
620 distances. The colors of the boxplots and dots represent the different groups analyzed
621 according to the legend. Yano_BR, Yanomami/Brazil; Yano_VE, Yanomami/Venezuela;
622 US, US individuals.



623

624 **Figure 3:** Bacterial and archaeal taxa differences among traditional and urban groups.
 625 (A) Barplot representing the relative abundance (percentage) of the most frequent
 626 phyla (B) Boxplots showing the *Bacteroidetes* and *Firmicutes* abundance (percentage)
 627 in each group (C) Bar chart showing the LDA scores > 4 of bacterial genera found to be
 628 significantly associated with each group (D) Relative abundance of the most prevalent
 629 archaeas identified in the groups. Yano_BR, Yanomami/Brazil; Yano_VE,
 630 Yanomami/Venezuela; US, US individuals.



631

632 **Figure 4:** Functional metabolic characteristics of the traditional and urbanized
633 microbiomes. (A) Principal coordinate analysis of Bray-Curtis distances based on
634 functions at the level 3 subsystems. (B) Heatmap showing the main functions at level 1
635 (orange bar) and level 2 regarding carbohydrates metabolism (green bar), protein
636 metabolism (gray bar) and cofactors, vitamins, prosthetic groups and pigments (purple
637 bar). Yano_BR, Yanomami/Brazil; Yano_VE, Yanomami/Venezuela; US, US individuals.