Data Descriptor

A DNA barcode reference library of the French Polynesian shore fishes

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Abstract

The emergence of DNA barcoding and metabarcoding opened new ways to study biological diversity, however, the completion of DNA barcode libraries is fundamental for such approaches to succeed. This dataset is a DNA barcode reference library (fragment of Cytochrome Oxydase I gene) for 2,190 specimens representing at least 540 species of shore fishes collected over 10 years at 154 sites across the four volcanic archipelagos of French Polynesia; the Austral, Gambier, Marquesas and Society Islands, a 5,000,000 km^2 area. At present, 65% of the known shore fish species of these archipelagoes possess a DNA barcode associated with preserved, photographed, tissue sampled and cataloged specimens, and extensive collection locality data. This dataset represents one of the most comprehensive DNA barcoding efforts for a vertebrate fauna to date. Considering the challenges associated with the conservation of coral reef fishes and the difficulties of accurately identifying species using morphological characters, this publicly available library is expected to be helpful for both authorities and academics in various fields.

Background & Summary

DNA barcoding aims to identify individual to the species level by using a short and standardized portion of a gene as a species tag[1]. This standardized procedure has revolutionized how biodiversity can be surveyed as the identification of a species then becomes independent of the level of taxonomic expertise of the collector[2], the life stage of the species[3,4] or the state of conservation of the specimen[5,6]. Due to its large spectrum of potential applications, DNA barcoding has been employed in a large array of scientific fields such as taxonomy[7], biogeography, biodiversity inventories[8] and ecology[9]; but see Hubert and Hanner for a review[10]. In the genomic era, this approach has been successfully applied to the simultaneous identification of multiple samples (*i.e.* the metabarcoding approach), extending its applications to surveys of whole ecological communities[11], but also to monitoring species diet[12,13], identifiying the presence of specific species in a region[14], or to studying changes in the community through time by sampling environmental DNA[15,16].

By design, DNA barcoding has proved to be fast and accurate, but its accuracy is highly dependent on the completeness of DNA barcode reference libraries. These libraries turn surveys of Operational Taxonomic Units (OTU) into species surveys through the assignment of species names to OTU[17,18], hence giving meaning to data for ecologists, evolutionists and decision-makers. Taxonomists increasingly provide DNA barcodes of new species they are describing; but thousands of species of shore fishes still lack this diagnostic molecular marker.

In the South Pacific, an early initiative led by the CRIOBE Laboratory was successfully carried out for French Polynesian coral reef fishes at the scale of one island, Moorea (Society Island)[19]. The fish fauna of Moorea's waters is one of the best known of the region given the historical operation of research laboratories and long term surveys[20,21]. Yet, the discovery of a high level of cryptic diversity[19] motivated the CRIOBE Laboratory to extend this biodiversity survey of shore fishes to the remaining islands of French Polynesia. French Polynesia (FP) is a 5,000,000 km^2 region located between 7° and 27° South Latitude with only 4,167 km^2 of emerged lands that constitutes a priority area to conduct such a barcoding survey, being a species rich region and hosting closely related species due to its position at the junction of several biogeographic areas with varying levels of endemism. The Marquesas Islands (northeastern FP) rank as the third highest region of endemism for coral reef fishes in the Indo-Pacific (13,7%[22]). The Austral Islands (southwestern FP) and Gambier Islands (southeastern FP) host numerous

southern subtropical endemic species[23–25]. Finally, the Society Islands (western FP) possess the highest species richness (877 species) and the highest number of widespread species in French Polynesia[26]. Here, we present the result of a large-scale effort to DNA barcode the shore fishes in French Polynesia. Conducted between 2008 and 2014, a total of 154 sites were inventoried across these four archipelagoes. Islands of varying ages and topographies were visited ranging from atolls to high islands surrounded by a barrier reef, or solely fringing reefs. Furthermore, inventories were conducted across different shore environments at each island (*i.e.* sand bank, coral reefs, rubble, rocky, etc.).

In total, 2,190 specimens were identified, preserved, photographed, tissue sampled, DNA barcoded and cataloged with extensive collection locality data to build a library representing at least 540 species, 232 genera and 61 families (Fig. 1). Merged with previous sampling efforts at Moorea, a total of 3,131 specimens now possess a DNA barcode representing at least 645 nominal species for a coverage of approximately 65% of the known species diversity of these four archipelagoes. These biodiversity surveys have already resulted in the publication of updated species checklists[22,26] and in the description of 17 new species[27–34]. This comprehensive library for French Polynesia shore fishes will certainly benefit a wide community of users with different interests, ranging from basic to applied science, and including fisheries management, functional ecology, taxonomy and conservation. Furthermore, many newly detected taxa for science are flagged here for which complete collection data and DNA barcodes are being made available, which should facilitate their formal descriptions as new species. While shedding new light on the species diversity of the Pacific region, this publicly available library is expected to fuel the development of DNA barcode libraries in the Pacific Ocean and to provide more accurate results for the growing number of studies using DNA metabarcoding in the Indo-Pacific region.

Methods

Sampling strategy

We explored a diversity of habitats across the four corners of French Polynesia with shallow and deep air SCUBA dives (down to 50–55 m) for a total of 154 sampled sites (Fig. 2, Table 1). A total of 2,190 specimens, representing at least 540 species, 232 genera and 61 families (Fig. 3a) have been collected across four archipelagos representing the four corners of French Polynesia (FP), through six campaigns: Marquesas Islands (1) in 2008 at Mohotani and (2) in 2011 at every island of the archipelago aboard the M.V. Braveheart (Clark Bank, Motu One, Hatutaa, Eiao, Motu Iti, Nuku-Hiva, Ua-Huka, Ua-Pou, Fatu-Huku, Hiva-Oa, Tahuata, Fatu-Hiva; 52 sites), (3) in 2010 at Gambier Islands aboard the M.V. Claymore (Mangareva, Taravai, Akamaru, and all along the barrier reef; 53 sites), (4) at Austral Islands in 2013 aboard the Golden Shadow (Raivavae, Tubuai, Rurutu, Rimatara, Maria Islands; 25 sites), (5) at westernmost atolls of the Society Islands in 2014 aboard the M.V. Braveheart (Manuae and Maupiha'a; 20 sites). A sixth campaign took place on Moorea's deep reefs in 2008 (Society Islands) as a small scale campaign that included the exploration and sampling of some of the deep reefs of Moorea (53 to 56 m depth; 4 sites) (Fig. 2).

Specimen collection

Specimens were captured using rotenone (powdered root of the Derris plant) and spear guns while SCUBA diving. These complementary sampling methods[35] allowed us to sample both the cryptic and small fish fauna as well as the larger specimens of species not susceptible to rotenone collecting. Four

individuals per species were collected on average. Fishes were sorted and identified onboard to the species level using identification keys and taxonomic references[23,36] and representative specimens of all species collected were photographed in a fish photo tank to capture fresh color patterns, labeled and tissue sampled for genetic analyses (fin clip or muscle biopsies preserved in 96% ethanol). The photographed/sampled voucher specimens were preserved in 10% formalin (3.7% formaldehyde solution) and later transferred into 75% ethanol for permanent archival storage. Preserved voucher specimens and tissues were deposited and cataloged into the fish collection at the Museum Support Center, National Museum of Natural History, Smithsonian Institution, Suitland, Maryland, USA. Nomenclature follows Randall (2005) and we followed recent taxonomic changes using the California Academy of Sciences Online Catalog of Fishes[37].

DNA barcode sequencing

We extracted whole genomic DNA using QIAxtractor (QIAGEN, Crawley) and Autogen AutoGenPrep 965 according to manufacturer's protocols. A 655bp fragment of the cytochrome oxidase I gene (COI) was amplified using Fish COI primers FISHCOILBC (TCAACYAATCAYAAAGATATYGGCAC) and FISHCOIHBC (ACTTCYGGGTGRCCRAARAATCA) and PCR and Sanger sequencing protocols as in Weigt et al.[38]. Sequences were edited using Sequencher 5.4 (Gene Codes) and aligned with Clustal W as implemented in BOLD. Alignments were unambiguous with no indels or frameshift mutations. A total of 2,190 DNA barcode have been generated (Data Citation 1 and 2).

Specimen identification

All morphological identifications were revised as needed after the specimens were deposited in the archival specimen collection to confirm initial identifications made in the field. Specimens of specific groups like Antennaridae, Bythitidae, Chlopsidae or Muraenidae were revised by additional taxonomist specialists (David Smith, John McCosker, Leslie W. Knapp, Werner Schwarzhans). After the morphological identification, we used the Taxon-ID Tree tool and Barcode Index Numbers (BIN) discordance tools as implemented in the Sequence Analysis module of BOLD to check every identification using the DNA barcodes generated. The Taxon-ID tool consists of the construction of a neighbor-joining (NJ) tree using K2P (Kimura 2 Parameter) distances by BOLD to provide a graphic representation of the species divergence[39] (Data Citation 2). The BIN discordance tool uses the Refined Single Linkage algorithm (RESL Ratnasingham & Hebert, 2013) to provide a total number of operational taxonomic units (OTUs) referenced as Barcode Index Numbers (BIN).

Technical Validation

To test the robustness of our library, we first computed the distribution of the interspecific and intraspecific variability for all the described species (Fig. 3b, c, d). We found that there is little to no overlap in the distribution of divergence within and between species for the vast majority of the species identified morphologically. The RESL algorithm identified more BINS (617) than nominal species identified morphologically (540). The reexamination of specimens in the light of these results suggest that 65 species could be new species for science awaiting a formal description (Table 2). Taxonomic paraphyly (*i.e.* potentially cryptic species) has been found for 18 additional species (Table 3) while incomplete lineage sorting or very shallow inter-species divergence (<2%) was observed for 17 species

(Table 4), mostly between Marquesan endemics and their closest relatives. In summary, 94% of the BINs match species identified using morphological characters, meaning that it is possible to successfully identify a species using DNA barcodes in 94% of the cases.

Data Records

This library is composed of three main components: (1) voucher specimens which are in the national fish collection at the Smithsonian Institution (Washington, DC) that were photographed in the field, (2) complete collection data associated with the voucher specimen, and (3) DNA barcodes (Fig. 1).

All photographs, voucher collection numbers, DNA barcodes and collection data are publicly available in the Barcode Of Life Datasystem (BOLD, http://www.boldsystems.org; Data Citation 1) in the Container INDOF "Fish of French Polynesia" or by sampling campaign ("AUSTR", "GAMBA", "MARQ", "MOH", "MOOP" and "SCILL") and in Figshare (Data Citation 2). DNA barcodes have also been made available in GenBank (Data Citation 3) and this database is accessible through the CRIOBE portal (Data Citation 4).

The library fulfill the BARCODE data standard which requires: 1) Species name, 2) Voucher data, 3) Collection data, 4) Identifier of the specimen, 5) COI sequence of at least 500 bp, 6) PCR primers used to generate the amplicon, 7) Trace files[41,42]. In BOLD, each record in a project represents a voucher specimen with its photographs, voucher collection numbers, sequences associated and extensive collection data related to (1) the Voucher: Sample ID, Field ID, Museum ID, Institution Storing; (2) the Taxonomy: Phylum, Class, Order, Family, Subfamily, Genus, species, Identifier, Identifier E-mail, Taxonomy Notes; (3) Specimen Details: Sex, Reproduction, Life Stage, FAO Zone, Notes such as sizes of the specimens, Voucher Status, and (4) Collection Data: Collectors, Collection Date, Continent, Country/Ocean*, State/Province, Region, Sector, Exact Site, GPS Coordinates, Elevation, Depth, Depth Precision, GPS Source, and Collection Notes (Data Citation 2).

Usage Notes

This Barcode release dataset is freely available to use in a barcoding or metabarcoding approach for specimen identifications. Several approaches can be considered:
(1) directly downloading the sequences in fasta format and working offline merging this dataset with an ongoing barcoding project;
(2) working online through the BOLD website (registration is free) merging the Container INDOF "Fish of French Polynesia" or parts of the sampling campaigns (Table 1) with an ongoing BOLD project;
(3) through online identification tools as data are indexed in both BOLD and Genbank databases. This library will be considered when any queries of molecular identification will be made through the identification engine of BOLD (http://www.boldsystems.org/index.php/IDS-OpenIdEngine) or the standard nucleotide Basic Local Alignment Search Tool (BLAST, https://blast.ncbi.nlm.nih.gov/). In the same manner, this dataset should also be indexed in the MIDORI database[43,44]. Composed of both endemic and widespread species, this library is expected to benefit a large community from academics to authorities who use molecular data to monitor and survey biodiversity.

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Author contributions

E.D.T. drafted the first manuscript, J.T.W., D.P., A.D., and N.H. provided extensive edits and comments. E.D.T, J.T.W., T.C., R.G., M.K., T.L.M., J.M., G.M.-T., V.P., P.P., P.S., G.S., N.T., M.V. and S.P. collected fish specimens. E.D.T., J.T.W., D.P., A.D., N.H., J.V., B.E., C. M., L.W. and S.P. produced DNA barcodes and cleaned the database. R.G. and S.P. financed the campaigns. C.M. and S.P. financed the sequencing. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Figure Legends

Figure 1. Overview of data generation. From collection of specimen to the validation of data generation.

Figure 2. Sampling localities across French Polynesia. In parenthesis the number of sites and number of specimens collected for each archipelago. Several sampling localities may be represented by a single dot due to the geographic scale of French Polynesia. Map data: Google, DigitalGlobe.

Figure 3. **Species diversity by family for the four archipelagoes sampled and distribution of genetic distance across the DNA barcode library.** (a) Species diversity by family for the four archipelagoes sampled; (b) Distribution of maximum intraspecific distances (K2P, percent); (c) Distribution of nearest neighbor distances (K2P, percent); (d) Relationship between maximum intraspecific and nearest neighbor distances. Points above the diagonal line indicate species with a barcode gap.

Figures

Figure 1

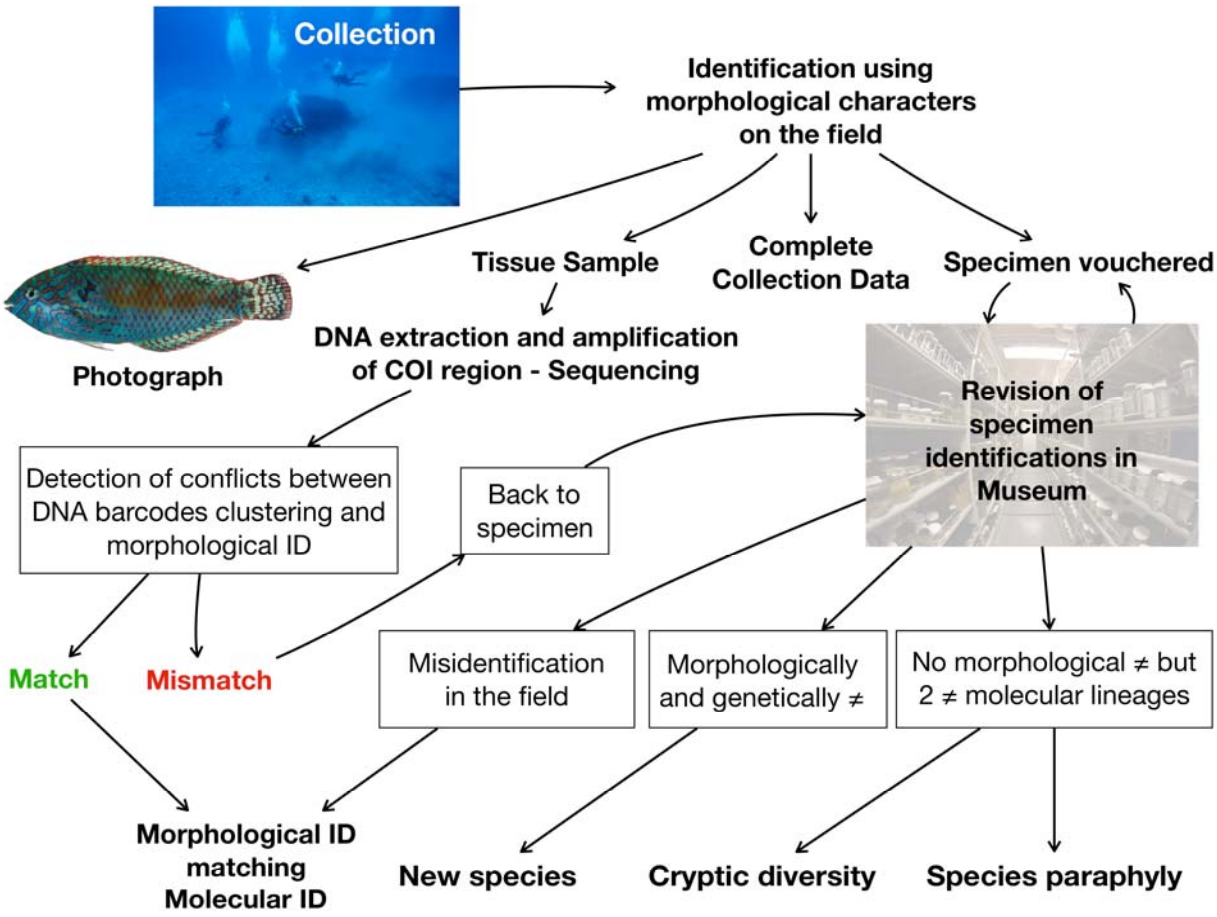
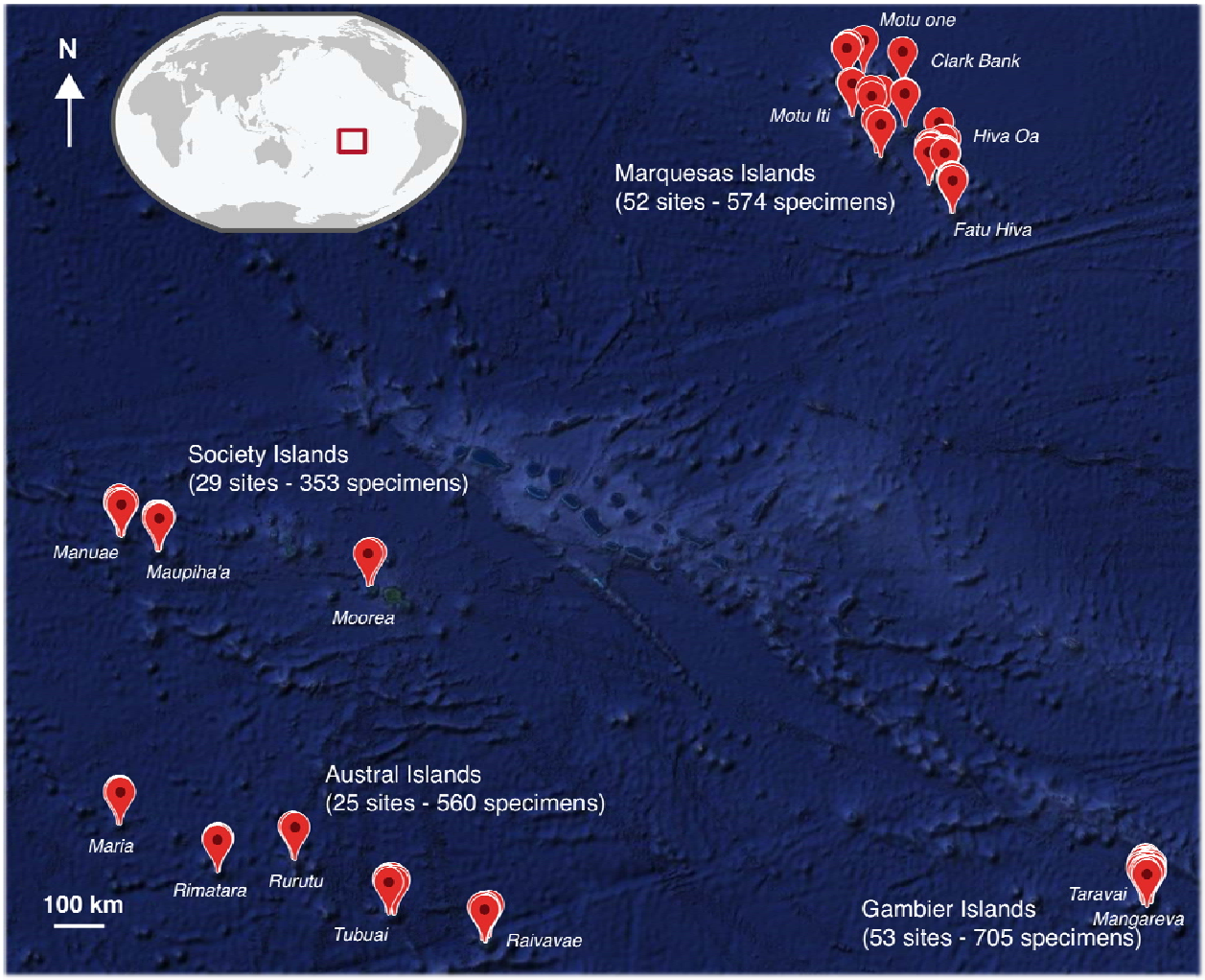
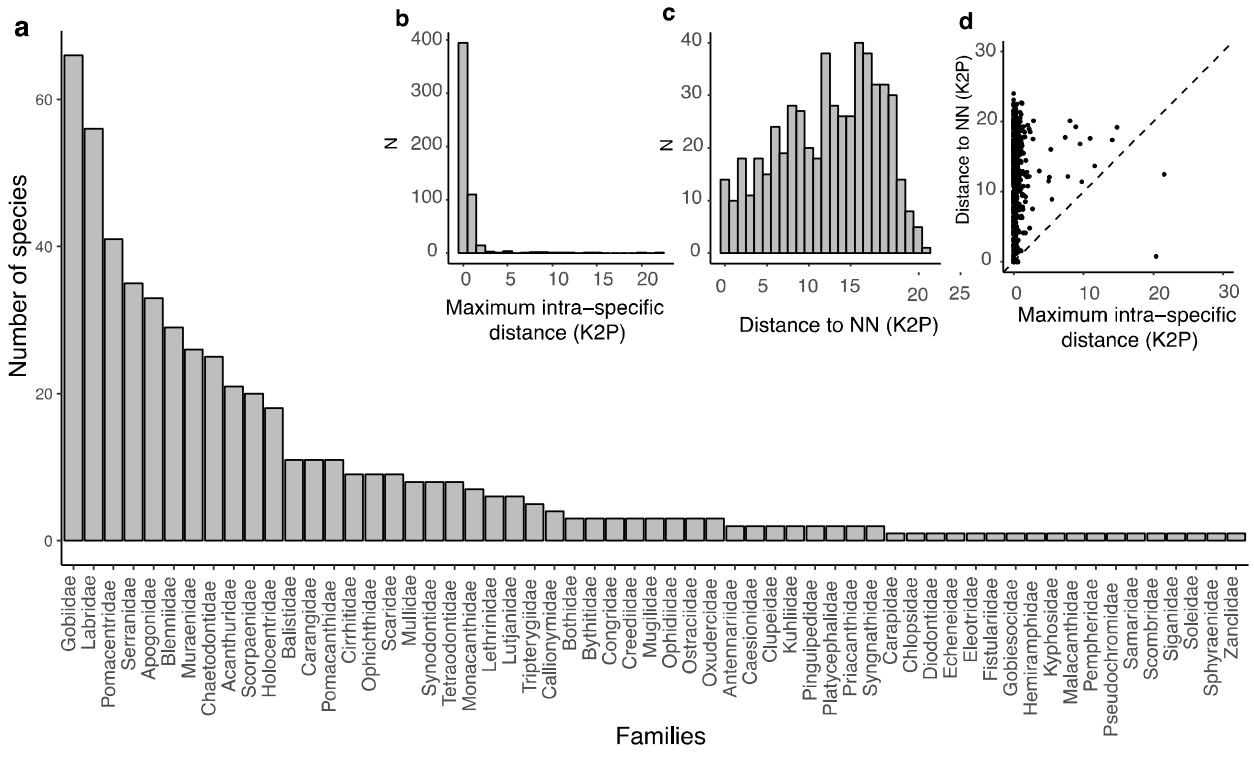


Figure 2


N
Motu one
Clark Bank
Motu Iti
Hiva Oa
Marquesas Islands
(52 sites - 574 specimens)
Fatu Hiva
Society Islands
(29 sites - 353 specimens)
Manuae
Maupiha'a
Moorea
Austral Islands
(25 sites - 560 specimens)
Maria
Rimatara
Rurutu
Tubuai
Raivavae
100 km
Taravai
Mangareva
Gambier Islands
(53 sites - 705 specimens)

253 **Figure 3**



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Tables Legends

Table 1. Overview of the dataset. Number of specimens and species collected for each sampling campaign.

Table 2. Potential new species detected in the dataset. Specimens which were identified only to the genus level and which represent potentially new species waiting to be described. Number of specimens included in each Barcode Index Number (BIN).

Table 3. Potential cryptic species. Species with number of specimens collected from the present survey presenting taxonomic paraphyly most likely representing undescribed cryptic species. Sample ID includes sampling location (AUST: Austral Islands, GAMB: Gambier Islands, MARQ and MOH: Marquesas Islands, SCIL and MOOP: Society Islands).

Table 4. Species displaying either incomplete lineage sorting or shallow inter-species divergence. Mean and Maximum intra-Species distances (Mean Intra-Sp and Max Intra-Sp), and Kimura 2 Parameter distances from the Nearest Neighbour (NN).

Tables

Table 1.

BOLD project	Geographical location	No. of specimen collected	No. of species collected
AUSTR	Austral Islands	560	263
GAMBA	Gambier Islands	705	290
MARQ	Marquesas Islands	386	182
MOH	Marquesas Islands	190	107
MOOP	Society Islands	42	27
SCIL	Society Islands	309	213

Table 2.

Sample ID	Family	Genus	Species	No. of specimens
AUST-419	Antennariidae	*Antennatus*	sp1	1
AUST-570, GAM-355	Antennariidae	*Antennatus*	sp2	2
MARQ-022, MOH-087	Antennariidae	*Antennatus*	sp3	2
SCIL-293	Antennariidae	*Antennatus*	sp4	1
MARQ-105, MARQ-106, MARQ-107	Apogonidae	*Fowleria*	sp	3
MARQ-380, MARQ-381, MOH-068	Apogonidae	*Gymnapogon*	sp	3
AUST-142, AUST-143	Apogonidae	*Pseudamiops*	sp	2
AUST-470, AUST-471	Apogonidae	*Siphamia*	sp	2
MARQ-177, MARQ-208, MOH-040	Blenniidae	*Blenniella*	sp	3
AUST-242, GAM-791, GAM-792, MARQ-139, MARQ-140	Blenniidae	*Cirripectes*	*sp*	5
GAM-278, GAM-279, SCIL-017, SCIL-021, SCIL-058, SCIL-288, SCIL-322	Blenniidae	*Enchelyurus*	*sp*	7
AUST-407, AUST-408, AUST-409	Blenniidae	*Entomacrodus*	sp	3
MARQ-184, MARQ-187, MARQ-378, MARQ-379	Blenniidae	*Rhabdoblennius*	sp	4
MOOP-028	Chlopsidae	*Kaupichthys*	sp	1
AUST-600	Congridae	*Ariosoma*	sp1	1
MARQ-318	Congridae	*Ariosoma*	sp2	1
MARQ-314, MARQ-315, MARQ-316	Congridae	*Gnathophis*	sp	3
MARQ-397, MOH-062	Creediidae	*Chalixodytes*	sp	1
AUST-427, AUST-538, AUST-539	Creediidae	*Crystallodytes*	*sp*	3
AUST-413	Gobiesocidae	*NA*	sp	1
AUST-159, AUST-305	Gobiesocidae	*Pherallodus*	sp1	2
AUST-532, AUST-533, AUST-534	Gobiesocidae	*Propherallodus*	sp	3
AUST-082	Gobiidae	*Bryaninops*	sp	1
GAM-374, GAM-375	Gobiidae	*Cabillus*	sp	2
MARQ-430, MOH-129, MOH-211	Gobiidae	*Callogobius*	sp	3
AUST-303, GAM-379	Gobiidae	*Eviota*	sp1	1
GAM-697	Gobiidae	*Eviota*	sp2	1

SCIL-038	Gobiidae	*Eviota*	sp3	1
AUST-346, AUST-347	Gobiidae	*Gobiodon*	sp	2
MARQ-097, MARQ-098	Gobiidae	*Gobiodon*	sp	2
SCIL-240	Gobiidae	*Gobiodon*	sp	1
AUST-566, AUST-567, AUST-568, GAM-364	Gobiidae	*Paragobiodon*	sp	4
MARQ-363, MARQ-364, MARQ-365	Gobiidae	*Pleurosicya*	sp1	3
MOOP-007	Gobiidae	*Pleurosicya*	sp2	1
SCIL-237	Gobiidae	*Priolepis*	sp	1
AUST-032	Gobiidae	*Silhouettea*	sp	1
MOOP-047	Gobiidae	*Sueviota*	sp	1
AUST-446	Gobiidae	*Trimma*	sp1	1
MARQ-435, MARQ-436	Gobiidae	*Trimma*	sp2	2
GAM-32	Gobiidae	*Trimmatom*	sp1	1
GAM-33	Gobiidae	*Trimmatom*	sp2	1
MOOP-002	Gobiidae	*Trimmatom*	sp3	1
AUST-423, AUST-424, AUST-425, AUST-544	Isonidae	*Iso*	sp1	4
GAM-521, GAM-522	Isonidae	*Iso*	sp2	2
MOH-079	Lethrinidae	*Gymnocranius*	sp	1
AUST-383, GAM-176, GAM-177, GAM-178, MOH-200, SCIL-299	Moringuidae	*Moringua*	sp1	6
SCIL-300	Moringuidae	*Moringua*	sp2	1
MARQ-505	Muraenidae	*Gymnothorax*	sp1	1
SCIL-335	Muraenidae	*Gymnothorax*	sp2	1
AUST-573, GAM-709	Ophidiidae	*Brotula*	sp	2
AUST-297, AUST-298, AUST-299, AUST-300, GAM-761	Pempheridae	*Pempheris*	sp1	5
MARQ-063, MARQ-165, MARQ-166, MARQ-167, MARQ-276, MARQ-382, MOH-178, MOH-179	Pempheridae	*Pempheris*	sp2	8
AUST-308, AUST-309, AUST-310, AUST-311	Pomacentridae	*Stegastes*	sp	4
MOOP-018, MOOP-019, MOOP-034, MOOP-035	Pseudochromidae	*Lubbockichthys*	sp	4
GAM-599, GAM-600	Scorpaenidae	*Scorpaenodes*	sp1	2
MOH-137, MOH-151	Scorpaenidae	*Scorpaenodes*	sp2	2
GAM-56, GAM-569, GAM-574, GAM-58	Scorpaenidae	*Sebastapistes*	sp1	4
MARQ-328, SCIL-114	Scorpaenidae	*Sebastapistes*	sp2	1

MARQ-046, MOH-027	Syngnathidae	*Doryrhamphus*	sp	1
MARQ-321	Synodontidae	*Synodus*	sp	1
AUST-011, AUST-048	Tripterygiidae	*Enneapterygius*	sp1	1
AUST-360, AUST-057, AUST-058, AUST-059	Tripterygiidae	*Enneapterygius*	sp2	4
GAM-68, GAM-69, GAM-123, GAM-124, GAM-125	Tripterygiidae	*Enneapterygius*	sp3	3
GAM-002	Tripterygiidae	*Enneapterygius*	sp4	1
SCIL-112, SCIL-133, SCIL-156	Tripterygiidae	*Enneapterygius*	sp5	3

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BIN	Taxa	No. of specimens
BOLD:AAF8427	*Apogon crassiceps*	2
BOLD:ABW7007	*Apogon crassiceps*	4
BOLD:ACE7901	*Apogon crassiceps*	1
BOLD:ACX1964	*Apogon doryssa*	1
BOLD:ABW8494	*Apogon doryssa*	2
BOLD:AAF5636	*Aporops bilinearis*	1
BOLD:AAF5637	*Aporops bilinearis*	4
BOLD:AAD2580	*Centropyge flavissima*	2
BOLD:AAD9019	*Centropyge flavissima*	6
BOLD:ACD1956	*Fusigobius duospilus*	5
BOLD:AAD1050	*Fusigobius duospilus*	1
BOLD:AAA6306	*Gnatholepis cauerensis*	9
BOLD:AAC6155	*Gnatholepis cauerensis*	5
BOLD:ACC5235	*Gymnothorax melatremus*	3
BOLD:AAC8364	*Gymnothorax melatremus*	5
BOLD:AAF0704	*Leiuranus semicinctus*	3
BOLD:AAL6561	*Leiuranus semicinctus*	2
BOLD:ACD1820	*Myrophis microchir*	1
BOLD:AAE0976	*Myrophis microchir*	2
BOLD:AAB3862	*Parupeneus multifasciatus*	6
BOLD:ACD1989	*Parupeneus multifasciatus*	3
BOLD:ACD1988	*Priolepis triops*	3
BOLD:AAX7961	*Priolepis triops*	1
BOLD:AAB4082	*Pristiapogon kallopterus*	1
BOLD:ABZ7996	*Pristiapogon kallopterus*	7
BOLD:ACC5180	*Pseudocheilinus octotaenia*	10
BOLD:AAD3038	*Pseudocheilinus octotaenia*	9
BOLD:AAB4821	*Pterocaesio tile*	4
BOLD:ACK9118	*Pterocaesio tile*	1
BOLD:ACP9778	*Scolecenchelys gymnota*	1
BOLD:AAJ8783	*Scolecenchelys gymnota*	2
BOLD:AAC7090	*Stegastes fasciolatus*	11
BOLD:ABZ0285	*Stegastes fasciolatus*	2
BOLD:ACC5053	*Uropterygius kamar*	1
BOLD:ACC5109	*Uropterygius kamar*	1
BOLD:ACD1642	*Uropterygius macrocephalus*	1
BOLD:AAU1965	*Uropterygius macrocephalus*	2

Table 4.

Family	Species	Mean Intra-Sp	Max Intra-Sp	Nearest Neighbour	Nearest Species	Distance to NN
Acanthuridae	*Acanthurus reversus*	0.08	0.15	AUSTR453-13	*Acanthurus olivaceus*	0
Holocentridae	*Myripristis earlei*	0.28	0.62	SCILL065-15	*Myripristis berndti*	0
Monacanthidae	*Pervagor marginalis*	0.36	0.62	SCILL083-15	*Pervagor aspricaudus*	0
Tetraodontidae	*Canthigaster criobe*	0	0	MOH030-16	*Canthigaster janthinoptera*	0
Mullidae	*Mulloidichthys mimicus*	0.52	0.52	AUSTR089-13	*Mulloidichthys vanicolensis*	0.17
Pomacentridae	*Chromis abrupta*	0	0	SCILL209-15	*Chromis margaritifer*	0.31
Labridae	*Coris marquesensis*	0	0	SCILL040-15	*Coris gaimard*	0.46
Apogonidae	*Ostorhinchus relativus*	N/A	0	SCILL142-15	*Ostorhinchus angustatus*	0.93
Tetraodontidae	*Canthigaster rapaensis*	0.21	0.31	MARQ456-12	*Canthigaster marquesensis*	1.1
Pomacentridae	*Abudefduf conformis*	0.15	0.15	GAMBA844-12	*Abudefduf sexfasciatus*	1.24
Monacanthidae	*Cantherhines nukuhiva*	0.15	0.31	GAMBA711-12	*Cantherhines sandwichiensis*	1.4
Pomacentridae	*Plectroglyphidodon sagmarius*	0.08	0.15	AUSTR222-13	*Plectroglyphidodon imparipennis*	1.56
Holocentridae	*Sargocentron caudimaculatum*	0.68	1.1	SCILL104-15	*Sargocentron tiere*	1.57
Acanthuridae	*Zebrasoma rostratum*	0	0	AUSTR376-13	*Zebrasoma scopas*	1.72
Apogonidae	*Apogon marquesensis*	0.23	0.31	GAMBA657-12	*Apogon susanae*	1.88
Chaetodontidae	*Chaetodon flavirostris*	0.08	0.15	SCILL269-15	*Chaetodon lunula*	1.88
Chaetodontidae	*Chaetodon lunula*	0.1	0.15	GAMBA555-12	*Chaetodon flavirostris*	1.88

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Data Citations

1. *BOLD DS-INDOF* (2019) (active when the ms will be published)
2. Delrieu-Trottin et al. (2019) Figshare https://doi.org/ 10.6084/m9.figshare.7923905 (2019) (active when the ms will be published).
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