

1 Title of the article: Anti-HIV-1 Activity of *Crocodylus mindorensis* (Philippine crocodile) serum in cell-free
2 and cell-associated virus interactions to human peripheral blood mononuclear cells

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11 Abstract:

12 Highly-Active Antiretroviral Therapy (HAART) is the recommended treatment and management strategy for
13 HIV infection. Although the existing antiretroviral drugs are indispensably significant in improving the
14 quality of lives of HIV/AIDS individuals, the drugs still have many limitations including resistance, production
15 of toxicity, and their limited availability. These limitations continue to open new opportunities in the use of
16 ethnomedicine for the management of HIV/AIDS. With this, few researchers have made an effort to test the
17 inhibitory activity of crocodile serum as it has a unique and diverse molecular activity in preventing HIV-1
18 replication. In this study, a cell culture-based assay was utilized coupled with colorimetric enzyme
19 immunoassay to determine the HIV-1 reverse transcriptase activity. One HIV-1 seropositive serum was
20 processed for Peripheral Blood Mononuclear Cells (PBMC) co-culture from which HIV-1 isolates were obtained.
21 The HIV-1 reverse transcriptase activity after 21 days was 0.5928 pg/well. Moreover, a baseline Philippine
22 crocodile serum concentration of 0.5% vol/vol was used based on the previous study conducted by Hinay and
23 Sarol (2018) and the cell viability results showed no cell reduction of mononuclear cells after 72 hours
24 incubation. The inhibitory activity of the Philippine crocodile serum at 0.5% and 0.25% vol/vol concentrations
25 inhibited 65.68±2.93% and 69.92±0.45% respectively in post-infection interactions. In addition, the Philippine
26 crocodile serum in pre-infection interaction at 0.5% and 0.25% vol/vol concentrations inhibited 68.61±1.67%
27 and 69.95±2.24% respectively. As has been noted, the inhibitory actions of the Philippine crocodile serum
28 effectively regulate the HIV-1 replication in both pre- and post-infection interactions.

29

30 **Keywords:** Philippine freshwater crocodile, HIV-1 co-culture, HIV-1 inhibition, HIV-1 reverse transcriptase

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32

33 **Introduction:**

34 Reptiles such as alligators exhibit remarkable ability to heal rapidly considering the septic environment in
35 which they live. Merchant and colleagues provided observation on the broad range of antiviral activity of
36 alligator serum (Merchant et. al, 2003, 2004). Merchant's recent studies also showed that these activities are
37 due to a potent and broad-acting serum complement system (Merchant et. al, 2004). In the Philippines, there
38 are two species of crocodile - the *Crocodylus porosus* locally known as Philippine saltwater crocodile and the
39 *Crocodylus mindorensis*, the endemic Mindoro crocodile. The relatedness of *Alligator mississippiensis*
40 (*American alligator*) described by Merchant and colleagues and the Philippine crocodile *Crocodylus*
41 *mindorensis* based on phylogeny shows that they belong to the same phylum (Crocodylia) but from different
42 Families (Casey, Gardner, & Farke, 2012).

43 Crocodiles may acquire serious injuries when they hunt food and may encounter higher order animals to
44 compete for survival. However, the crocodile's immune system responds well to these injuries and the
45 crocodiles generally survive and do not show signs of illness. Moreover, crocodylians have been shown to have
46 a strong immune system with a remarkably higher biological and pharmacological activity than others
47 animals, and also humans, which make them a good platform for drug discovery (Dzik, 2010). These
48 immunological effector mechanisms necessary for the efficient control of infectious agents such Human
49 Immunodeficiency Virus Type-1 that are dependent on distinct defense strategies. Some components
50 involved in these routes of the defense system have been identified and characterized which probably
51 include serum complement cascades (Merchant et al., 2003; Dzik, 2010), serum Mannose-Binding Protein
52 (Ezekowitz, Kuhlman, Groopman, & Byrn, 1989) and conglutinin-like protein (Ushijima, et al., 1992). All
53 these features may contribute to the antimicrobial and antiviral properties of crocodile serum.

54 The antiviral activity of *Alligator mississippiensis* (*American alligator*) serum described by Merchant, 2005
55 conducted using cell culture-based method provided the evidence that it contains anti-HIV-1 activity with
56 half maximal inhibitory activity of 0.9% (Merchant et. al, 2003, 2004). In the Philippines, particularly in
57 Davao, crocodiles which are characteristically similar to alligators are not only abundant but are
58 scientifically cultured and propagated. With the availability of Philippine crocodile, a scientific research that
59 will be a foothold and basis that *Crocodylus mindorensis* serum could be used as a potential inhibitor of
60 HIV/AIDS infection was investigated.

61

62

63 **Subjects and Methods:**

64 **Specimen Collection**

65 A minimum of 2 mL serum sample from one (1) purposively selected adult- male *Crocodylus mindorensis* was
66 collected at the Animal Clinic of the Davao Crocodile Park facility. The freshly collected sera were separated
67 from whole blood samples collected by a veterinarian as part of the health check of the crocodile.
68 Specifically, 5 mL of whole blood were allowed to clot at room temperature for approximately three (3)
69 hours. The serum was separated by centrifugation at 3,000 x g for 15 minutes and processed immediately.

70

71 **Ethical Considerations**

72 **Blood Sampling Protocol**

73 Peripheral blood mononuclear cells were obtained from two different individual sources. First from a healthy
74 non-HIV-1 reactive individual and second from a HIV-1 reactive individual. An inform consent were given to
75 both participants before blood collection.

76 A volunteer HIV-1 reactive individual with a signed inform consent was the source for HIV-1 co-culture
77 method. A three (3) 5 mL EDTA tube was used to collect the blood using venepuncture technique with strict
78 compliance to Occupational Safety and Health Administration 1910.1030 Bloodborne Pathogens guidelines
79 (OSHA 1910.1030). After collection, the sample was processed immediately. The same protocol was followed
80 for healthy non-HIV-1 reactive volunteer.

81

82 **Biosafety Considerations**

83 Upon arrival of crocodile serum sample, proper protocol was observed including serum separation to anti-
84 HIV-1 activity assessment. At the end of the experiment, the used materials such as microplate and washing
85 basin containing 5.25% sodium hypochlorite were decontaminated at 121°C at 15psi for 30 minutes including
86 all crocodile serum received.

87

88 **HIV-1 Isolation**

89 Isolation of HIV-1 from one sero-positive sample was done using a PBMC micro-co-culture assay (Dahake *et.al*
90 2013). Briefly, one healthy, HIV-sero-negative donor PBMCs were stimulated with the mitogen
91 phytohemagglutinin-P (PHA-P; at a final concentration of 5.0 µg/mL), in the presence of human interleukin 2
92 for 24-72 hours before use to promote blast formation and replication of T-cells. The cells were counted and

93 1×10^6 PHA-stimulated donor cells and 1×10^6 PBMCs from sero-positive HIV-1 individual was added in duplicate
94 wells of a 24-well tissue culture plate. The final volume was adjusted to 2.0 mL with growth media
95 containing RPMI1640 with 20% FBS and 10.0 U/mL IL-2. The plate was incubated at 37°C with 5% CO₂ in a
96 CO₂ incubator. On day 7 and 14, the cultures were replenished with fresh growth media containing 5×10^5 PHA
97 stimulated donor cells (feeder cells). The culture was continued until day 21 and then terminated.
98 Supernatant fractions from duplicate wells from day 14 and 21 were saved separately, and stored at -70°C
99 until analysis for HIV-1 RT by ELISA. Cultures were considered positive only when day 21 supernatants showed
100 an increase in HIV-1 RT from day 14 supernatants.

101

102 **Cell viability assay**

103 Before using the Philippine crocodile serum sample for antiviral assay, it was necessary to assess
104 their toxicity against Peripheral Blood Mononuclear Cells (PBMC). Briefly, a 100 μ L PBMC was suspended into
105 a 0.5 mL microcentrifuge tube and added 90 μ L of Safranin stain. The PBMC were counted to 1×10^6 cells.
106 After determining the PBMC count, cell viability assay was performed. Briefly, a 100 μ L 1×10^6 PBMC were
107 suspended into a 0.5 mL microcentrifuge tube and added 100 μ L of 0.5% and 0.25% vol/vol Philippine
108 crocodile serum. The interactions were allowed up to 72 hours and cell viability was counted.

109 **Post-infection interaction (cell-associated HIV)**

110 The procedure was based on the study of Dahake, R (2013). Briefly, HIV-1 virions from co-cultured were first
111 allowed to infect PBMCs and the Philippine crocodile serum were added to the suspension after one hour. A
112 volume of 100 μ L of HIV-1 virions were added to 1 mL of PHA-stimulated PBMCs and incubated for 2 hours in
113 a CO₂ incubator. After incubation, the cells were washed carefully and supernatants were aspirated set to
114 leave 50 μ L/well. After washing, 100 μ L of infected cells were plated into wells and 100 μ L of Philippine
115 crocodile serum were added. The plates were incubated for 72 hours at 37°C in the CO₂ incubator. The
116 contents of each well were transferred to microfuge tubes, cells were pelleted at 13,000 RPM for 5 minutes
117 and the supernatants used for determining the HIV-1 RT levels.

118

119 **Pre-infection interaction (cell-free HIV)**

120 The procedure was based on the study of Dahake, R (2013). Briefly, the HIV-1 virions from co-cultured were
121 first interacted with the Philippine crocodile serum for two (2) hour and then allowed to infect PBMCs. A
122 volume of 10 μ L of HIV-1 virions were added to 10 μ L of Philippine crocodile serum (Human serum as a

123 negative control) incubated for 2 hours at 37°C in a CO₂ incubator. After 2 hours of interaction the
124 experimental moiety-virus suspension was added to 100µL of PHA stimulated PBMCs and incubated for 2
125 hours in the CO₂. After incubation, the cells were washed carefully and supernatants were aspirated set to
126 leave 50 ul/well After washing, 100µL of infected cells were plated into wells and 100µL of Philippine
127 crocodile serum were added. The plates were incubated for 72 hours at 37°C in the CO₂ incubator. The
128 contents of each well were transferred to microfuge tubes, cells were pelleted at 13,000 RPM for 5 minutes
129 and the supernatants used for determining the HIV-1 RT levels.

130

131 **Data Management**

132 The results of pre-infection and post-infection assays were expressed as means ±SD in triplicates. The

133 Percent (%) reduction of HIV-1 activity was computed using the following mathematic formula:

134 % inhibition = $\frac{\text{Absorbance of Negative control}^* - \text{Absorbance of test sample}^{**}}{\text{Absorbance of Negative Control}} \times 100$

135

136 *Negative control -without crocodile serum

137 **Test sample - with Crocodile Serum

138

139 **Results and Discussion:**

140

141 **HIV-1 Co-culture and Cell viability**

142 The HIV-1 RT activity after 21 days was 0.5928 ng/well. On the other hand, a baseline Philippine crocodile
143 serum concentration of 0.5% vol/vol was used based on the study conducted by Hinay and Sarol (2018) and
144 the cell viability results showed no cell reduction of mononuclear cells after 72 hours incubation.

145

146 **Post-infection interaction (cell-free HIV)**

147 Table 1 shows the inhibitory activity of the Philippine crocodile serum. HIV-1 replication inhibition at 0.5%
148 and 0.25% vol/vol concentrations were $65.68 \pm 2.93\%$ and $69.92 \pm 0.45\%$ respectively. This is also supported with
149 the HIV-1 Reverse transcriptase activity of the peripheral blood mononuclear cells (Figure 1). The PBMCs
150 interacted with crocodile serum concentration of 0.5% and 0.25% vol/vol shows a decreased HIV-1 reverse
151 transcriptase activity of 0.1216 and 0.0954 ng/well respectively.

152 Unexpectedly, an inverse relationship was observed between crocodile serum concentration and the anti-
153 HIV-1 Reverse Transcriptase activity. This means that the inhibitory activity decreases as the Philippine
154 crocodile serum concentration increases. This is similar to the previous study of Hinay and Sarol (2018), were
155 the result could be explained by the presence of an “inhibitor of the inhibitor” in the crocodile serum. The
156 best candidate inhibitor present in the crocodile serum with putative inhibition activity to HIV-1 is the
157 Antimicrobial Peptides (AMPs) (Finger & Isberg, 2012). This is also supported by the study of Hinay and Sarol
158 (2018) where Philippine crocodile serum concentration showed *in vitro* inhibition of HIV-1 Reverse
159 transcriptase by high as $92.93 \pm 0.72\%$ at 0.5 vol/vol % (Hinay and Sarol, 2018). Moreover, another putative
160 component of the Philippine crocodile serum as described by Okada and colleagues provided a good support
161 that the complement fragment C5a, an anaphylotoxin and a small peptidase product during complement
162 activation, has a 30% HIV-1 Reverse transcriptase inhibitory activity (Okada, et al., 2011). The inhibition of
163 the HIV-1 virion interacted with the Philippine crocodile serum indicates that the inhibition occurs either
164 direct interaction to viral enzymes reverse transcriptase and or protease which are key viral enzymes of HIV-
165 1 replication.

166

167 **Pre-infection interaction (cell-free HIV)**

168 The potential of the Philippine crocodile serum as inhibitor against HIV-1 replication was determined
169 and the findings demonstrated a significant inhibition of HIV-1 replication prior to interaction to PBMCs.

170 Table 1 shows the inhibitory activity of the Philippine crocodile serum in pre-infection interaction. Reverse
171 Transcriptase inhibition at 0.5% and 0.25% vol/vol concentrations inhibited $68.61 \pm 1.67\%$ and $69.95 \pm 2.24\%$
172 respectively. This is also supported with the HIV-1 Reverse transcriptase activity of the peripheral blood
173 mononuclear cells (Figure 2). The PBMCs interacted with crocodile serum concentration of 0.5% and 0.25%
174 vol/vol with an HIV-1 reverse transcriptase activity of 0.1005 and 0.10054 ng/well respectively. With this the
175 Philippine crocodile serum have both direct virucidal effects to HIV-1 virion or the crocodile serum inhibits
176 the viral entry subsequently results through inhibition of fusion of the viral envelope with the host cell
177 membrane (Ray and Doms, 2006). Possible crocodile serum component such as Alpha-1 antitrypsin (AAT)
178 which is an acute phase reactant and the most abundant circulating serine protease inhibitor can be a
179 putative HIV-1 inhibitor (Shapiro, Pott, & Ralston, 2001). Shapiro and colleagues (2001) describe Alpha-1
180 antitrypsin (AAT) in their study as a natural HIV-1 antagonist. In their study the inhibitory activity of the
181 Alpha-1 antitrypsin was observed in the induction of latent virus from chronically infected cells, viral
182 infection and replication in PBMC, attachment CCR-5 -cell assay and infection and replication in whole blood.
183 Merchant ME and colleagues (2005) described the effectiveness of alligator serum against the laboratory
184 strain of Human immunodeficiency virus type-1 using a cell culture-based assay. The results exhibited potent
185 anti-HIV activity at 20- and 64-fold dilutions with 100 and 89% reduction in cytopathic effects, respectively.
186 The study also showed that the higher concentration of the serum was difficult to assess due to inherent
187 toxicity to the cell culture used in the study (Merchant, et al., 2005). In addition, the results suggest that
188 both the antiHIV-1 activity and cell toxicity were mediated by serum complement activity as heat treatment
189 of the serum destroyed both effects. Like human complement activity, alligator serum complement is
190 sensitive to heat inactivation (Merchant, Thibodeaux, Loubser, & Elsey, 2004).
191 These findings are quite significant since the Philippine crocodile serum concentration of 0.5% and 0.25%
192 vol/vol seems to work in both pre- and post-infection interactions and hence suggest that the effect may be
193 mediated by a direct action on the virus particle. Perhaps it blocks binding or uptake to cells or causes
194 immediate particle lysis.

195

196 **CONCLUSION**

197 The results showed that the HIV-1 Reverse Transcriptase activity was inhibited by $65.68 \pm 2.93\%$ and
198 $69.92 \pm 0.45\%$ in Philippine crocodile serum concentration of 0.5% and 0.25% vol/vol respectively in post-
199 infection interaction and $68.61 \pm 1.67\%$ and $69.95 \pm 2.24\%$ in Philippine crocodile serum concentration of 0.5%
200 and 0.25% vol/vol respectively in pre-infection interaction. As has been noted, the 0.5% and 0.25% vol/vol

201 concentration of the Philippine crocodile serum effectively regulates the HIV-1 replication in both pre- and
202 post-infection interactions. With this, the study recommends to identify and characterize the active
203 compounds in the Philippine crocodile serum and develop a novel source of HIV-1 replication inhibitors.

204

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208

209 **CONFLICT OF INTEREST**

210 No conflict of interests is declared by authors for the contents in this manuscript.

211

212 **AUTHORS CONTRIBUTION**

213 Alfredo A. Hinay Jr., Nelyn Mae T. Cadotdot and Marilou V. Tablizo designed and carried out the experiment
214 and Alfredo A. Hinay Jr prepared the manuscript.

215

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- 255

Table 1. Inhibitory Activity of the Philippine Crocodile Serum in Post-infection interaction (cell-associated HIV)

Concentration % vol/vol	Post-infection Mean±SD	Post-infection Mean±SD
0.50	65.68±2.93	68.61±1.67
0.25	69.32±0.45	69.95±2.24

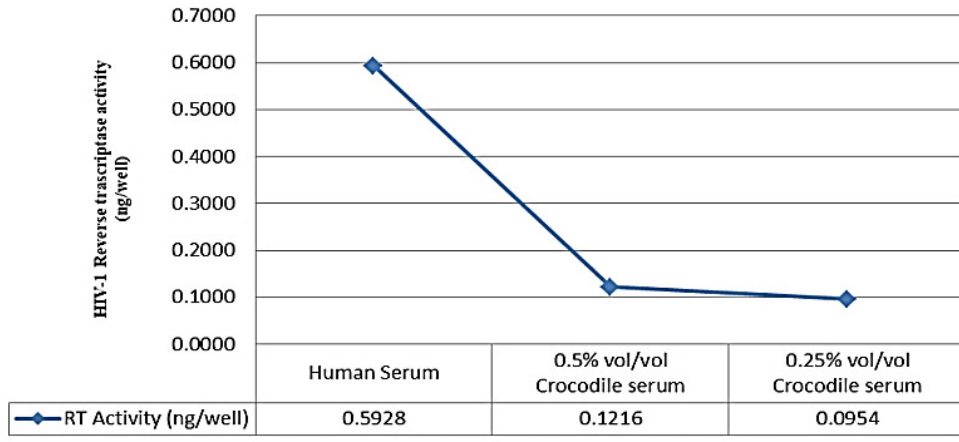


Figure 1. Reverse Transcriptase Activity in Post-infection interaction (cell-associated HIV)

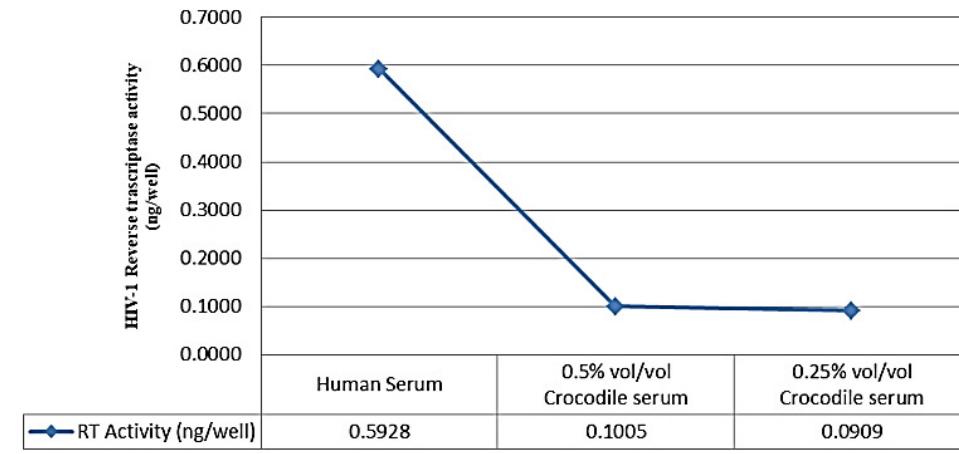


Figure 2. Reverse Transcriptase Activity in Pre-infection interaction (cell-free HIV)