1

Title: Ultra-local adaptation due to genetic accommodation

Syuan-Jyun Sun[1*], Andrew M. Catherall[1], Sonia Pascoal[1], Benjamin J. M. Jarrett[1,2], Sara E. Miller[3], Michael J. Sheehan[3], Rebecca M. Kilner[1]

[1] Department of Zoology, University of Cambridge, Downing Street, Cambridge, CB2 3EJ, UK

[2] Department of Entomology, Michigan State University, 288 Farm Lane Room 243, East Lansing, MI 48824, USA

[3] Department of Neurobiology and Behavior, Cornell University, 215 Tower Road, Ithaca, NY 14853, USA

*Corresponding author: Syuan-Jyun Sun,

Phone: 01223 (3)34466

e-mail: sjs243@cam.ac.uk

Keywords: interspecific competition, phenotypic plasticity, plasticity-led evolution, burying beetles, *Nicrophorus vespilloides,* niche expansion, local adaptation.

Statement of authorship: R.M.K. conceived the idea. S.-J.S., A.M.C. S.P and R.M.K designed the study. S.-J.S., A.M.C. and B.J.M.J. acquired the phenotypic data, which was analysed by S.-J.S. and A.M.C. M.J.S. and S.P designed the genetic's component, which was collected by S.P and analysed by M.J.S and S.E.M. All authors contributed to discussion of the results and writing the manuscript.

Abstract

Models of 'plasticity-first' evolution suggest that phenotypic plasticity precedes and facilitates the rapid evolution of new adaptations. Previous work has emphasised how new adaptations arise through the loss of plasticity, when populations split to occupy new environments and become reproductively isolated from the ancestral population. Here we analyse the role of 'plasticity-first' evolution in generating divergent new adaptations when populations are still connected by gene flow. We report that neighbouring populations of burying beetles *Nicrophorus vespilloides*, in different Cambridgeshire woodlands, have divergently adapted to breed on different-sized carrion despite ongoing gene flow. We show experimentally that this ultra-local adaptation is due to evolution of the reaction norm linking clutch size to carrion size, a process known as genetic accommodation. Plasticity is not lost in the derived population, but instead is fine-tuned by evolution to match the greater range in carrion available naturally. Accordingly, we find that loci associated with oogenesis have become differentiated in beetles from the different woodlands. We suggest that genetic accommodation can enable organisms to rapidly and specifically adapt to very fine-scale differences in their local environment, even while gene flow persists.

Introduction

A key challenge in evolutionary biology is to explain how natural populations evolve new adaptations. An old idea [1–3], which has recently been revitalized [4,5], is that pre-existing phenotypic plasticity can facilitate adaptive evolution in new environments ('plasticity-first evolution'). The extent of phenotypic plasticity in any trait is described by the reaction norm which relates environmental variation to the phenotype it induces. Plasticity-first evolution is possible because the shape, slope and elevation of a reaction norm each have genetic components, upon which selection can act [6]. Focusing mainly on traits that promote survival, such as melanic colouration or morphology, recent experimental work on natural populations has shown that novel adaptive phenotypes can indeed evolve as a consequence of plasticity-first evolution [7, 8]. In populations of toads and lizards, for example, selection in a new environment has greatly reduced the slope of the reaction norm, thus eliminating phenotypic plasticity and causing the constitutive expression of novel, canalised traits [7,8]. New adaptations can therefore evolve through 'genetic assimilation' (sensu [9]) of traits that were once induced environmentally.

However, genetic assimilation is predicted to occur only when there is little or no gene flow between populations that inhabit different local environments [10–12]. Consistent with

this prediction, the populations of toads and lizards that have evolved new traits through

genetic assimilation are apparently reproductively isolated [7,8]. While gene flow still

persists, theory predicts that selection will instead act to maintain plasticity in adaptive traits

[11,12]. Nevertheless, plasticity-first evolution can still facilitate local adaptation because

selection can fine-tune the slope or elevation of the reaction norm within each population to

match local conditions. This is the process of genetic accommodation [13]. In principle,

genetic accommodation of the reaction norm provides a mechanism for the ultra-local

adaptation seen in populations that still experience gene flow [14]. However, whether this

mechanism is at work in natural animal populations is unclear (but see [15] for evidence from

a plant species).

We tested for evidence of plasticity-first adaptation through the genetic accommodation

of a reaction norm, in wild populations of burying beetles *Nicrophorus vespilloides.* We

focused on populations occupying neighbouring woodlands in Cambridgeshire, UK, which

are c. 2.5 km apart: Gamlingay Wood (Latitude: 52.15555°; Longitude: −0.19286°), and

Waresley Wood (Latitude: 52.17487°; Longitude: −0.17354°). In previous work, we showed

that there is gene flow between the *N. vespilloides* populations inhabiting each woodland

because they cannot be differentiated at neutral genetic markers [16]. Here we show that the

burying beetle guilds differ between the two woodlands. We then test whether genetic

accommodation of traits related to fecundity has enabled each population of *N. vespilloides* to

become ultra-locally adapted to the contrasting levels of competition from other *Nicrophorus*

species.

Burying beetles (*Nicrophorus* spp.) breed on the dead body of small vertebrates, such as

rodents, and typically match their clutch size to the size of the carcass they opportunistically

secure for reproduction [17]. The adaptive clutch size therefore depends on the size of the

carrion that is routinely available for reproduction. This is the population's carrion niche. It is

common for several species of *Nicrophorus* beetle to exist in sympatry within the same

woodland and then the carrion niche is divided between them, according to their relative size

[18]. Burying beetles compete within and between species to obtain exclusive ownership of a

carcass to breeding upon, and larger beetles are more likely to win these fights [19,20].

Competition for carrion divides the carrion niche so that larger burying beetle species tend to

breed on larger carrion, while the smallest species (*N. vespilloides* in the UK) usually breeds

on the smaller carrion [18,21]. Larger carrion potentially yields more larvae than small

carcasses, and so is more intensely fought over [22].

In this study, we collected ecological, genomic and experimental data to test three of the

criteria for plasticity-first evolution, set out by Levis & Pfennig (2016). With ecological and

genomic data, we show that *N. vespilloides* from Gamlingay Wood is an ancestral-proxy

population while the *N. vespilloides* population from Waresley Wood represents a derived

condition. Through experimentation and analysis of ecological data, we find that *N.*

vespilloides from each woodland now occupies a different carrion niche. Gamlingay *N.*

vespilloides faces competition for carrion from three larger *Nicrophorus* species and has a

narrow carrion niche in which reproduction is specialised on small carcasses. By contrast,

Waresley *N. vespilloides* competes with only one rival *Nicrophorus* species, and has

expanded its carrion niche to breed successfully on carcasses that range more widely in size.

To test whether this niche expansion has evolved under genetic accommodation we

experimentally: (1) expose Gamlingay *N. vespilloides* to larger carrion and test whether this

induces an increase in clutch size; (2) characterise the equivalent reaction norm in Waresley

N. vespilloides to test whether the slope or elevation has evolved to be different; (3) test

whether any population divergence in the reaction norm is adaptive by quantifying how

effectively *N. vespilloides* from each population converts small and large carrion into

offspring. Finally, we use genomic analyses to test for evidence of divergence between the

populations at loci associated with oogenesis.

Results and Discussion:

Gamlingay and Waresley Woods are 'woodland islands' that have existed for 1000-4000

years

Our study focuses on the *Nicrophorus* populations that inhabit Gamlingay Wood and

Waresley Wood, in Cambridgeshire, England, UK. Five thousand years ago, England was

almost uniformly covered in ancient forest: the so-called 'Wild Wood' [23]. During the

Bronze Age, and especially from the Iron Age onwards (i.e. from 3000-4000 years ago), the

Wild Wood underwent extensive destruction [23]. We know from the Domesday Book (a

land survey commissioned almost 1000 years ago by William the Conqueror) that Gamlingay

Wood and Waresley Wood have existed as distinct woodlands since at least 1086, meaning

that they separated c.1000-4000 years ago. In common with other ancient woodlands created

at this time, these woods have stayed approximately the same size since then [24]. Gamlingay

Wood was acquired and managed by Merton College, Oxford for c. 800 years [25]. Its

ecology was described in detail in 1912 [25]. The modern history of Waresley Wood is less

well-known [24,26]. Both sites are now designated as 'ancient woodland' and are managed

by the Bedfordshire, Cambridgeshire and Northamptonshire Wildlife Trusts. They are

woodland islands of approximately the same size (c 50ha) in a landscape dominated by arable

farming (S1A Fig).

The Nicrophorus guild differs between Gamlingay and Waresley Woods

Each year from 2014-2017 we set five beetle traps per woodland, at exactly the same five locations within each wood, and checked the contents every 2-3 weeks from June until October (see Materials and Methods; S1A Fig). In general, we found that the two woodlands harboured a similar number of *Nicrophorus* beetles: we caught a total of 1873 *Nicrophorus* individuals in Gamlingay Wood over the four-year sampling period compared with 1806 *Nicrophorus* individuals in Waresley Wood. However, we discovered that whereas Gamlingay Wood routinely supports four *Nicrophorus* species, Waresley Wood is routinely inhabited by only two species (Fig 1A). The average abundance per trap of each *Nicrophorus* species differed between the two woods (species x woodland interaction: χ^2 = 142.68, d.f. = 4, $p < 0.001$). The smallest species (Fig 1B; S1 and S2 Tables), *N. vespilloides*, was by far the most abundant in each woodland; 80.6% (1510 individuals) of all *Nicrophorus* beetles trapped in Gamlingay Wood and 93.9% (1695 individuals) of those trapped in Waresley Wood were *N. vespilloides*. Both sites also contained stable populations of the largest species, *N. humator* (Fig 1B; S1 and S2 Tables) Only Gamlingay Wood contained populations of intermediate-sized *N. interruptus* and *N. investigator* in all four years of the study (Fig 1B; S1 and S2 Tables). Overall, each of these two species was significantly more abundant in Gamlingay than in Waresley Woods (Tukey HSD, $z = 7.90$, $p < 0.001$ and $z = 5.80$, $p < 0.001$, respectively). The *Nicrophorus* guild is therefore significantly different between the two woodlands (PERMANOVA (permutational multivariate analysis of variance) test $F = 0.024$, $p < 0.001$). A clear separation of the guild structure between the woods can be seen for each sampling time across the four years in NMDS (nonmetric multidimensional scaling) ordination (S1B Fig).

No evidence for divergence in the small mammal population between Gamlingay and Waresley Woods

In 2016, we sampled the small mammal population in Gamlingay and Waresley Woods to estimate the abundance and type of carrion that might be available to the burying beetles to breed upon (see Materials and Methods). In Gamlingay, 32 animals were caught across 50 trap sessions (23 new catches and 9 recaptures); in Waresley 41 animals were caught across 50 trap sessions (30 new catches and 11 recaptures). Across both woods, bank voles (*Myodes glareolus*; range: 15-40 g) and wood mice (*Apodemus sylvaticus*; range: 13-27 g) were the dominant species, constituting 53% and 43% of all trapped mammals respectively. One yellow-necked mouse (*Apodemus flavicollis*; range: 14-45 g) and one common shrew (*Sorex*

araneus; range: 5-14 g) were caught in Waresley. There was no difference in the mean body

mass of small mammals sampled between the two sites ($\chi^2 = 0.19$, d.f. $= 1$, $p = 0.661$; S2

Fig). We have no reason to think that the mortality of these rodent species should differ

between woodlands that are in such close proximity and that are subjected to similar levels of

ecological management.

Greater competition among Nicrophorus for carrion in Gamlingay versus Waresley Woods

We conclude from these data that there are approximately similar numbers of *Nicrophorus*

beetles within each woodland competing for an approximately similar size, abundance and

type of rodent carrion to breed upon. The key difference lies in the number of burying beetle

species in each wood. Evidence from other populations suggests that our measurements

reflect long-term differences because abundance measures of *Nicrophorus* are robust over

time, even when there are marked change in woodland management [25].

Tests of plasticity-led evolution critically depend on the ability to distinguish ancestral

populations from more recently evolved populations [6]. We infer from the ecological data

that the more speciose burying beetle guild in Gamlingay Wood more closely approximates

the ancestral burying beetle guild that was present in the 'Wild Wood', whereas the guild in

Waresley Wood represents a more recent, derived condition. This is because burying beetle

guilds in pristine ancient forests in North America are more speciose than those present in

degraded woodlands [27], in keeping with the general observation that there is a positive

relationship between habitat size and species richness [28]. Furthermore, smaller-bodied

generalist carrion beetles are more likely to survive in fragmented forests [27].

Niche expansion by N. vespilloides beetles in Waresley Wood

The ecological data additionally suggest that in Gamlingay Wood, *N. vespilloides* is more

likely to be confined to breeding only on smaller carrion, whereas in Waresley Wood this

species' carrion niche is likely to extend to include larger carrion because it very rarely faces

competition from *N. interruptus* and *N. investigator* for larger carcasses (see [18]). Although

the proportion of intermediate-sized *Nicrophorus* species is relatively small (Fig 1C), *N.*

vespilloides in Gamlingay Wood is more likely to face competition from congenerics for

rarer, larger carrion than *N. vespilloides* in Waresley Wood [18,22]. This inference is further

supported by previous experimental work on *Nicrophorus*. In continental forests, which are

presumably also rich in *Nicrophorus* species, Urbański & Baraniak (2015) found that *N.*

vespilloides was more than twice as likely to be found on small carcasses than on large

carcasses. In addition, larger species appear to be under selection to breed on larger carcasses
[29,30].
To test these inferences about the division of the carrion niche among *Nicrophorus*
species within each woodland, we measured the reproductive performance of *N. interruptus*
(from Gamlingay Wood), *N. investigator* (from Gamlingay Wood) and *N. vespilloides* beetles
(from both Gamlingay and Waresley Woods) in the laboratory (see Materials and Methods).
For each species, we experimentally varied the size of carrion available for reproduction,
presenting pairs with either a small mouse carcass (range: 12-20 g; mean ± S.E.M: 16.80 ±
0.55 g) or a large mouse carcass (range: 25-31 g; mean ± S.E.M: 28.27 ± 0.53 g; natural
carcass range = 8.5-41 g). To quantify reproductive performance on each carcass size, we
measured 'carcass use efficiency': this is the extent to which each species converted the
carcass resource into larvae, and was calculated by dividing the total brood mass at the end of
larval development (measured when larvae had dispersed from the carcass) by the size of the
carcass the brood was reared on. The rationale was that beetles should exhibit greatest
efficiency when breeding on the carcass size to which they are locally adapted in nature.
Therefore, if there is local adaptation, *N. interruptus* and *N. investigator* should each exhibit
greatest efficiency when breeding on a large carcass, while *N. vespilloides* from Gamlingay
Wood should exhibit greatest efficiency when breeding on a small carcass. *N. vespilloides*
from Waresley Wood should show similar levels of efficiency whether breeding on a small or
large carcass.
We found that the efficiency of converting the carcass into larvae varied with carcass
size, but in a different way for each *Nicrophorus* species (carcass size x *Nicrophorus* species
interaction term: $\chi^2 = 28.85$, d.f. = 3, $p < 0.001$; Fig 2 and S3 Table). Specifically, *N.
interruptus* had an equal efficiency when breeding on small and large carcasses ($t = 0.99$, $p =
0.325$), whereas the larger *N. investigator* exhibited a greater efficiency when breeding on
large carcasses than on small carcasses ($t = 3.51$, $p < 0.001$). *N. vespilloides* from Gamlingay
Wood exhibited greatest efficiency on small carcasses ($t = -4.16$ $p < 0.001$), whereas *N.
vespilloides* from Waresley had similar levels of efficiency on small and large carcasses ($t =
-0.27$, $p = 0.791$). Confining our analyses to reproductive efficiency on larger carcasses, we
found that Gamlingay *N. vespilloides* was less efficient than both *N. interruptus* ($t = -3.92$, $p
< 0.001$) and *N. investigator* ($t = -2.69$, $p = 0.038$), as well as *N. vespilloides* from Waresley ($t
= -3.11$, $p = 0.011$).
Therefore, we conclude that the population of *N. vespilloides* in Waresley Wood has
adaptively expanded its carrion niche to breed on larger carcasses, in response to the
reduction in competition from *N. interruptus* and *N. investigator* for larger carrion.

Niche expansion by Waresley N. vespilloides is due genetic accommodation

To test whether niche expansion was facilitated by genetic accommodation, we first
generated reaction norms relating carcass size to brood size for *N. vespilloides* from the two
woodland populations (see Materials and Methods). We predicted that the reaction norm for
N. vespilloides from Waresley Wood should be significantly steeper than the reaction norm
for *N. vespilloides* from Gamlingay Wood, and this was indeed the case (woodland x carcass
size interaction: $\chi^2 = 4.80$, d.f. = 1, $p = 0.029$; Fig 3; S4 Table). When given a larger carcass
for reproduction, *N. vespilloides* from Gamlingay produced only a marginally larger brood
than when breeding on a small carcass ($z = 1.82$, $p = 0.068$). By contrast, Waresley *N.
vespilloides* produced significantly larger broods on larger carcasses than on smaller
carcasses ($z = 5.53$, $p < 0.001$). Thus, on a large carcass, Waresley *N. vespilloides* produced
more larvae than Gamlingay *N. vespilloides* ($z = -1.97$, $p = 0.049$), whereas on a small
carcass the number of larvae produced did not differ between the two populations ($z = 0.76$, $p
= 0.447$). Whether they bred on small or large carcasses, the variance was similar for
Gamlingay and Waresley *N. vespilloides* in brood size (Bartlett's test, $p = 0.615$) and carcass
use efficiency (Bartlett's test, $p = 0.943$). Females produced heavier larvae at dispersal when
breeding on a large carcass than on a small carcass ($\chi^2 = 139.05$, d.f. = 1, $p < 0.001$; S4
Table), irrespective of their woodland of origin ($\chi^2 = 1.06$, d.f. = 1, $p = 0.304$; S4 Table). We
could find no difference in the size of adult *N. vespilloides* trapped in Gamlingay and
Waresley Woods, ($\chi^2 = 2.02$, d.f. = 1, $p = 0.156$; S3A Fig), even after controlling for sex ($\chi^2 =
1.41$, d.f. = 1, $p = 0.235$). Nor could we detect any differences in body size distribution
between Gamlingay and Waresley *N. vespilloides* ($D = 0.044$, $p = 0.410$; S3B Fig). Thus,
given an abundance of resources for reproduction, *N. vespilloides* allocates them to producing
more offspring rather than larger offspring.

The relationship between carcass size and brood size at dispersal depends partly on the
number of eggs laid, and partly on any mortality that occurs between egg-laying and the
dispersal of larvae away from the carcass at the end of their development to pupate. In further
analyses, we compared the reaction norms relating carcass size to clutch size for each
population. Again, we found that they differed. This time the difference between reaction
norms was in their intercept, rather than in their slope (Fig 4A). Female *N. vespilloides* from
both woodlands laid more eggs when given a larger carcass to breed upon (carcass size effect:
$\chi^2 = 10.97$, d.f. = 1, $p < 0.001$; S4 Table), but Waresley *N. vespilloides* consistently laid more
eggs than Gamlingay *N. vespilloides* (woodland effect: $\chi^2 = 21.07$, d.f. = 1, $p < 0.001$; S4
Table), irrespective of carcass size (carcass size x woodland interaction: $\chi^2 = 1.44$, d.f. = 1, p

= 0.231) or female size (woodland x female size interaction: $\chi^2 = 1.30$, d.f. = 1, $p = 0.254$).

Likewise, female *N. vespilloides* from both woodlands laid smaller eggs when given a larger

carcass to breed upon (carcass size effect: $\chi^2 = 5.90$, d.f. = 1, $p = 0.015$; Fig 4B), but Waresley

N. vespilloides consistently laid smaller eggs than Gamlingay *N. vespilloides* (woodland

effect: $\chi^2 = 4.59$, d.f. = 1, $p = 0.032$), irrespective of carcass size (carcass size x woodland

interaction: $\chi^2 = 0.33$, d.f. = 1, $p = 0.565$). Mean egg volume per clutch did not predict mean

larval mass per brood at dispersal ($\chi^2 = 0.28$, d.f. = 1, $p = 0.595$), regardless of the population

of origin. This suggests that any under-provisioning of eggs is compensated by the

over-abundance of resources available on the carcass after hatching [31,32]. The total volume

of all eggs in a clutch did not differ between populations ($\chi^2 = 2.16$, d.f. = 1, $p = 0.141$; S4

Table) or carcass size treatments ($\chi^2 = 2.38$, d.f. = 1, $p = 0.123$; S4 Table).

We conclude that the reaction norms relating carcass size to brood size differ in slope

between the two woodlands (Fig 3), partly because Waresley females routinely lay more eggs

when they breed (Fig 4A), and partly because Waresley broods on smaller carcasses exhibit

greater mortality between egg-laying and larvae dispersal. Niche expansion by Waresley *N.*

vespilloides is due to genetic accommodation of the reaction norm linking carrion size and

clutch size.

288 *Divergence at loci associated with oogenesis in N. vespilloides from Gamlingay v. Waresley*

289 *Woods*

To test whether these differences between woodlands really are due to evolutionary change,

and hence genetic change, we generated low-coverage whole genome sequences for 40

individuals collected from each wood ($n = 80$ chromosomes per population). Population

genetic summary statistics were similar for both populations (mean of 2 kb windows –

Gamlingay Pi = 0.0055 ± 1.02e-5, Tajima's D = -0.70 ± 0.002; Waresley Pi = 0.0056 ±

1.0e-5, Tajima's D = -0.67 ± 0.002). Consistent with previous microsatellite analyses [16],

we found little to no genetic differentiation between populations from Gamlingay and

Waresley Woods (unweighted *Fst* = 0.0069; weighted *Fst* = 0.013), strongly suggesting there

is ongoing gene flow between the two populations. There was minimal divergence across the

genome with no instances of extreme outliers (S4 Fig). That no loci were highly diverged

between the two populations indicates that the divergent traits are likely controlled by many

loci, as is typical for behavioural and life history traits. It is consistent with the predicted

quantitative genetic basis of genetic accommodation [6]. The highest *Fst*-windows in the

genome showed only modest absolute values of divergence. Nevertheless, they were extreme

outliers due to the otherwise consistently low pattern of *Fst* between populations. For

example, the top window of divergence between the two populations fell in *transmembrane*

protein 214 (Fst = 0.11, $zFst$ = 19.2, p = 7.2e-82). This gene is highly expressed in the

ovaries in *Drosophila melanogaster*, suggesting a potential candidate gene that may influence

differences in *N. vespilloides* egg laying behaviour between Gamlingay and Waresley

Woods.

Next, we investigated whether the divergent *Fst* scores could be attributed to shifts in

allele frequency in *N. vespilloides* from Waresley Wood, Gamlingay Wood, or both

woodland populations. Greater differentiation attributable to Waresley compared to

Gamlingay is the pattern we would expect if the Waresley *N. vespilloides* is the derived

population. By comparing differentiation between both woodlands and a third population, we

can polarise the divergence between populations. For these analyses, we generated

low-coverage whole genome sequences from 22 individuals trapped in woods near Swansea,

Wales, UK, approximately 300 km away from Gamlingay and Waresley Woods. We

calculated all pairwise *Fst* and assigned the relative divergence of each population using

population branch statistics (PBS) to understand which population was driving divergence

across the genome [33,34]. Not surprisingly, the distant Welsh population showed the highest

genome-wide PBS. Interestingly, the population from Waresley Wood showed higher

differentiation compared to the population from Gamlingay Wood (mean PBS: Waresley =

0.0074, Gamlingay = 0.0056, Wales = 0.0109). These analyses therefore support the

ecological data in indicating that the Gamlingay *N. vespilloides* represents the ancestral

condition, whereas the Waresley *N. vespilloides* are the derived population (S5 Fig).

To visualise the relative divergence between populations across the genome, we

generated a scatterplot of the PBS values for 2kb non-overlapping windows for each

population (Fig 5A). The analysis highlighted multiple potential candidate genes associated

with the differences in egg-laying behaviour, again consistent with this trait being controlled

by many loci of small effect. These genes showed elevated PBS in *N. vespilloides* from

Waresley Wood (PBS > 0.05), but not Gamlingay Wood, and they are linked to arthropod

oogenesis. For example, homologs of three of the highly differentiated genes in the Waresley

Wood population – *obg-like ATPase*, *transmembrane protein 214*, and *liprin-alpha* – show

elevated expression in the ovaries of fruit flies [35]. FlyAtlas: survey of adult and larval

expression. http://www.flyatlas.org/) and other arthropods [36], suggesting a plausible role in

regulating egg production. Another gene, *kekkon1*, is a transmembrane protein known to

regulate the activity of the epidermal growth factor receptor (EGFR) during oogenesis in

Drosophila [37,38]. A different portion of *kek1* is moderately differentiated in *N. vespilloides*

from Gamlingay Wood (Fig 5A), highlighting that differentiation related to oogenesis is not

limited to *N. vespilloides* from Waresley Wood. One of the stronger signals of differentiation

in Waresley Wood that may contribute to the phenotypic differences observed between

populations is found in a serotonin receptor (Fig 5B). Serotonin has been linked to

reproduction via effects on the production of ecdysteroids such as juvenile hormone in

multiple insects [39–41]. Serotonin has also been related to the intensity of aggressive

behaviour in contests across diverse insect groups [42–45].

Finally, we asked whether genes involved in oogenesis generally showed elevated levels

of differentiation in each population, in comparison with the rest of the genome. For each

population, we ranked genes by the highest PBS score in 500bp windows overlapping with

the gene-body and conducted a gene set enrichment analysis for each population. N.

vespilloides from both Waresley and Gamlingay Woods showed enrichment in multiple GO

terms associated with ovaries and oogenesis (Table 1). Consistent with the outlier analyses,

oogenesis-related GO-terms tended to have higher enrichment scores in *N. vespilloides* from

Waresley Wood compared to Gamlingay Wood. The gene enrichment analysis also revealed

local divergence between beetles from Waresley and Gamlingay Woods at genes associated

with other traits, including learning and memory and sensory systems (S1 Data).

Conclusions

Our analyses support three of the criteria required to show evidence for plasticity-first

evolution [6]. We show that the ancestral reaction norm linking carrion size to clutch size has

evolved to have a greater intercept in *N. vespilloides* from Waresley Wood. This change has

enabled Waresley N. *vespilloides* beetles to expand their carrion niche in the absence of rival

congenerics. It is adaptive because Waresley *N. vespilloides* are now more effective than

Gamlingay *N. vespilloides* at converting carrion that ranges widely in size into offspring.

Previous analyses of plasticity-first evolution in natural populations have emphasised

how new adaptations can evolve from the genetic assimilation of plastic traits, and are caused

by the loss of plasticity [6–8,46]. In these studies, there is little evidence of ongoing gene

flow between diverging populations [7,46]. The new adaptations presumably arise from a

loss-of-function mutation(s), that makes trait expression independent of wider environmental

conditions.

Fewer studies have explicitly shown how new adaptations can also evolve from the

genetic accommodation of previously plastic traits. Here, adaptation involves re-tuning the

reaction norm to change the extent of plasticity that is induced by an environmental cue

(Bock et al. (2018) [15] and this study, but see also [47,48]). The candidate loci for

evolutionary change are thus likely to function in gene regulation. Our data suggest that the

candidate loci that have re-tuned the fecundity reaction norm in *N. vespilloides* include genes

associated with the regulation of oogenesis. In both Bock et al. (2018) and this study,

plasticity-first evolution through genetic accommodation was possible despite gene flow

between the diverging populations. Ultra-local adaptations, between populations that are

separated by just a few kilometres and that exhibit ongoing gene flow, have now been

reported from diverse contexts (e.g. [14]). We suggest that these types of adaptation are likely

to have evolved by genetic accommodation as well.

The emerging empirical evidence therefore suggests that plasticity-first evolution could

frequently be the source of diverse new adaptations in natural populations. Where gene flow

is minimal, new adaptations have been shown to arise through a loss of plasticity. By

contrast, while gene flow continues, our study suggests that new adaptations are more likely

to arise through changes in the extent of plasticity, just as theory predicts. Through this

mechanism, organisms can rapidly and specifically adapt to very fine-scale differences in

their local environment.

Materials and Methods

Competition for carrion in Gamlingay and Waresley Woods

Burying beetle trapping

Surveys of burying beetle species in Gamlingay and Waresley Woods were conducted from

June to October in 2014-2017. Five carrion-baited soil-filled traps were placed in each site to

attract *Nicrophorus* beetles, with traps set at least 150 m apart from each other and at the

same location each year (S1A Fig). Every 2-3 weeks, we collected all the *Nicrophorus* spp.

within each trap, and rebaited the trap with fresh compost and a dead mouse. In the lab, we

identified each burying beetle to species-level, and measured the pronotum width (to the

nearest 0.01 mm) as an index of body size (see below).

We caught five species in total (in increasing order of size): *N. vespilloides*, *N.*

interruptus, *N. vespillo, N. investigator* and *N. humator*. However, *N. vespillo* were only

caught 5 and 3 times within the four years in Gamlingay and Waresley respectively,

indicating that there is no stable population of *N. vespillo* in either wood.

After removing any phoretic mites, beetles were retained and kept individually in

plastic boxes (12cm x 8cm x 2cm) filled with moist soil in a laboratory kept at 20°C and on a

16:8 light to dark cycle. Beetles were fed twice a week with minced beef. We kept all

field-caught individuals for at least two weeks before breeding to ensure that they were

sexually mature and to reduce any variation in nutritional status. We then maintained stock

populations of both Gamlingay and Waresley Woods by breeding pairs of unrelated

individuals on 8-16 g mice carcasses.

For the genomic analyses, we analysed *N. vespilloides* from three ancient woodlands

near Swansea in Wales, trapped by Dr Chris Cunningham in 2017. The three sites in Wales

(Park Wood (Latitude: 51.57258°; Longitude: −4.03096°); Clyne Valley Wood (Latitude:

51.61262°; Longitude: −4.02293°); and Caswell Bay Wood (Latitude: 51.57258°; Longitude:

−4.03096°)) are approximately 300 km away from our two study sites in Cambridgeshire.

Size distributions of the Nicrophorus spp.

Body size was measured for *Nicrophorus* spp. collected from 2014 to 2017 (except in 2016).

In total, 838 *N. vespilloides*, 41 *N. humator*, 127 *N. interruptus*, 54 *N. investigator*, and 5 *N.*

vespillo were measured for Gamlingay Wood, whereas 824 *N. vespilloides*, 51 *N. humator*,

25 *N. interruptus*, 4 *N.* investigator, and 6 *N. vespillo* were measured for Waresley Wood.

Mean body size of *Nicrophorus* spp. significantly varied among species (GLMM: $\chi^2 =$

2069.44, d.f. = 4, $p < 0.001$; S2 Table). Post-hoc comparisons revealed that *N. vespilloides*

was the smallest species compared to other *Nicrophorus* spp. (S1 and S2 Tables).

Kolmogorov-Smirnov test comparing pairwise differences in *Nicrophorus* spp. body size

frequency distribution also revealed similar patterns found in differences of mean body size

(Fig 1 and S5 Table).

Mark-recapture experiment

In 2014, we investigated the interconnectivity of populations of burying beetles between

Gamlingay and Waresley Woods using a mark-recapture survey. In total, 98 *N. vespilloides*,

9 *N. humator*, 17 *N. interruptus*, 9 *N. investigator,* and 2 *N. vespillo* were marked for

Gamlingay Wood, whereas 113 *N. vespilloides*, 5 *N. humator*, 1 *N. interruptus*, 1 *N.*

investigator were marked for Waresley Wood. Beetles were marked with a numbered plastic

bee tag on either the right or left elytra for those found in Gamlingay or Waresley Woods

respectively. To identify any previously caught beetles that lost their tags, we permanently

marked them by cutting a small portion of the elytra (less than 2%). All marked beetles from

each wood were released from a designated location at the geographic midpoint with the

minimum total distance to all trapping sites for Gamlingay (Latitude: 52.16294°; Longitude:

−0.18984°) and Waresley Woods (Latitude: 52.176508°; Longitude: −0.156776°). We found

no evidence of migration between the two sites. We recaught 8 of 98 marked *N. vespilloides*

from Gamlingay Wood in Gamlingay Wood and 8 out of 113 marked *N. vespilloides* from

Waresley Wood in Waresley Wood. None of the other marked *Nicrophorus* spp. was

recaptured in either wood.

Small mammal trapping

To assess the rodent carrion available for *Nicrophorus* spp. reproduction, we sampled the

small mammal communities in the two woodlands. In general, rodent populations peak in the

autumn, because breeding for the year has just ceased and there has yet to be any

winter-induced mortality [49]. Sampling at this time is therefore ideal for detecting which

species are present and for determining their relative abundance. We placed Longworth traps

in both woodlands in November 2016. Traps were baited with oats and blowfly maggots

(with hay provided as bedding) and set in pairs within 20 m of each original beetle trapping

site (S1A Fig), with 10 traps set per wood. We continuously trapped rodents for three days,

generating 50 trap sessions per woodland. Traps were checked daily at approximately 0830

and 1500 (generating a total of 30 trap sessions overnight and 20 trap sessions in daylight

hours). Trapped mammals were identified, weighed, sexed, marked by a fur clip on either the

right or left rear flank, and released *in situ*. Any recaptured mammal was recorded in

subsequent censuses. All traps were reset and rebaited immediately after checking.

Division of the carrion niche by Nicrophorus beetles in Gamlingay and Waresley Woods

During the field season from 2017 to 2018, pairs of wild-caught *N. vespilloides*, *N.*

interruptus, and *N. investigator* were bred on either small (12-20 g; 16.80 ± 0.55 g) or large

carcasses (25-31 g; 28.27 ± 0.53 g) within a breeding box (17 x 12 x 6 cm) filled with 2 cm

of moist soil. All field-caught beetles were kept for two weeks in the laboratory and fed twice

a week prior to breeding. In total, we established eight treatments: large ($n = 47$) and small (n

$= 48$) carcasses for Gamlingay *N. vespilloides*; large ($n = 42$) and small ($n = 33$) carcasses for

Waresley *N. vespilloides*; large ($n = 25$) and small ($n = 18$) carcasses for *N. interruptus*; large

($n = 9$) and small ($n = 13$) carcasses for *N. investigator*. *N. interruptus* and *N. investigator*

were both drawn from Gamlingay Wood as the populations of these species in Waresley were

too small to be used experimentally.

Approximately eight days after parents were given a carcass to breed on, larvae switch

from aggregating on the carcass to dispersing away into the soil to pupate. When one or more

larvae from each brood switched their behaviour we scored the whole brood as having

reached the dispersal stage. At this point, all larvae were counted and total brood mass was

weighed to the nearest 0.001 g. We also calculated average larval mass for each brood by

dividing total brood mass by number of larvae.

Niche expansion in *N. vespilloides* by genetic accommodation

The reaction norm relating carcass size to brood size in each woodland

This experiment was conducted in the laboratory, over two blocks in 2017, using the second
and third descendant generations of field-caught beetles from Gamlingay and Waresley
Woods. By rearing beetles from both woodlands in the lab in a common garden environment
for at least one generation prior to testing, we minimized any residual environmental effects
when quantifying the reaction norm for each population. To pair beetles for reproduction, we
began by haphazardly casting broods into dyads when larvae had matured into adults. Within
each dyad, we haphazardly chose four males from one brood, and paired them with four
haphazardly chosen females from the second brood. Two of these pairs were then given a
small mouse to breed upon (12-17 g; 15.03 ± 0.67 g), while the remaining two pairs were
given a large mouse to breed upon (26-31 g; 28.86 ± 0.67 g). By using sibships to generate
pairs in this way, we were able to compare how very similar genotypes responded to the
opportunity to breed on either a small or large mouse. Each pair, and their mouse, was housed
in a clear plastic box (17 x 12 x 6 cm), with 2 cm depth of Miracle-Gro compost. The box
was placed in a dark cupboard for eight days after pairing the beetles, at which point larvae
started to disperse away from the carcass. We measured the number of larvae present at
dispersal and weighed the whole brood.

The reaction norm relating carcass size to clutch size in each woodland

In the second block of the experiment, we also measured clutch size. Fifty-six hours after we
introduced the beetles to the carcass, we photographed the base of each transparent breeding
box. Using Digimizer ver. 5.1.0, we then counted the number of visible eggs, and also
measured the length and width of all eggs that were able to be measured accurately (i.e. those
that were fully visible and lying flat on the base of the box). All counting and measuring of
eggs was performed blind to the carcass size treatment and the population from whence the
breeding beetles came. Egg volume was then calculated using the formula $V=1/6 *\pi*w2**L$,
which assumes eggs to be a prolate spheroid (following [50]).

In total, 2518 eggs were counted across 132 breeding boxes, of which 1633 could be
measured. No eggs could be seen in four of these boxes, and these were excluded from
further analysis (hence $n = 128$). Both the total number of eggs counted (observed clutch
size) and the number of eggs which could accurately be measured correlated positively with
brood size (number of eggs counted: $\chi^2 = 27.18$, d.f. = 1, $p < 0.001$; number of eggs

measured: $\chi^2 = 24.26$, d.f. = 1, $p < 0.001$), indicating these counts capture biologically

meaningful information.

Divergence at loci associated with oogenesis in *N. vespilloides* from Gamlingay *v.*

Waresley Woods

We generated low-coverage whole genome sequences for three populations of *N. vespilloides*

from Waresely Wood, Gamlingay Wood and Swansea, Wales, UK. DNA was individually

extracted from beetle heads using the DNeasy Blood and Tissue kit (Qiagen) and

subsequently quantified and quality checked using NanoDrop and Qubit. DNA was then

shipped to Cornell University, where libraries were prepared using partial reactions of a

Nexterra kit by the Cornell Genomics Core. Libraries were subsequently sequenced by

Novogene (Davis, CA, USA) at an average depth of approximately 5x coverage. Paired-end

550 bp insert libraries were prepared for each sample with the Nextera library preparation kit.

Libraries were sequenced using the Illumina HiSeq (Novogene, Davis, CA) at an average

coverage of 3.4X. Trimmomatic (v0.36) was used to removed adaptors and poor-quality

sequence. Trimmed reads were mapped to the *N. vespilloides* reference genome using the

Burrows-Wheeler Aligner (v0.7.13) [51]. SNPs were identified using Picard (v2.8.2) and

GATK (v3.6) HaplotypeCaller following best practice recommendations [52]. After

alignment, SNPs were hard filtered using the parameters: QD < 2.0 || SOR > 3.0 || FS > 200.

We used VCFtools to calculate population genetic statistics for each population. To examine

population structure, we generated a thinned VCF file with one SNP per 5kb and used Tassel

(version 5) [53] to calculate generate an MDS plot. For calculation of *Fst* and PBS values,

aligned bam files were analysed in ANGSD (v 0.911) [54], which is specifically designed for

analysis of low-coverage genome sequencing data. Data for this project is available at the

NCBI Sequence Read Archive under Bioproject PRJNA530213.

Genes were assigned gene ontology (GO) terms using the BLAST2GO workflow

(v5.1.1). In brief, gene identity was determined based upon BLAST searches to the

Arthropod or Drosophila non-redundant protein databases and protein domains were

identified based on matches to Interpro database. GO terms were assigned to each gene

model based upon mapping results. GO terms were filtered with the "Filter Annotation by

GO Taxa" option to remove GO terms that are incompatible for Arthropods.

Statistical analyses

All statistical analyses were performed in R version 3.4.3 (R Development Core Team),

with NMDS in the package *vegan*, GLM and GLMM in the package *lme4* [55], and Tukey's

HSD post hoc comparisons in the package *lsmeans* [56].

Competition for carrion in Gamlingay and Waresley Woods: field data

To test for a difference in the *Nicrophorus* guild between Gamlingay and Waresley Woods, a

frequency table of beetle communities was analysed using permutational multivariate

analysis of variance (PERMANOVA) on two-dimensional nonmetric multidimensional

scaling (NMDS) based on Bray-Curtis distances [57], with the 'adonis' function (vegan

package). We tested the effect of study site on the composition of the beetle community,

using sampling year as strata. The analysis was based on 10000 permutations of the data. We

visualized the difference of beetle community between Gamlingay and Waresley population

in two dimensions of a NMDS plot. NMDS two-dimensional stress values (a measure of

goodness of fit) were below 0.1 (0.085), indicating the ordination provides a good fit to the

data [58].

We used general linear mixed models (GLMM) with a Poisson error structure to test for

differences across beetle species and sites on average abundance for each species per trap as

an independent variable and year as a random factor. Beetle species and population

(Gamlingay/Waresley) were included as fixed effects, whereas trap ID and sampling year

were included as random factors.

We assessed the significant difference in body size frequency distribution between

Gamlingay and Waresley *N. vespilloides* and among *Nicrophorus* spp. using the

Kolmogorov-Smirnov (K-S) two-sample test, which compares if the cumulative distributions

of two data sets are derived from the identical distribution. We also tested for significant

differences in mean body size between *N. vespilloides* in Gamlingay and Waresley, using a

GLMM that included population (Gamlingay/Waresley) and sex (male/female) as fixed

effects, and sampling year as a random factor. Differences of mean body size among

Nicrophorus spp. was assessed in a GLMM that included species and sex as fixed effects, and

sampling year as a random factor.

We assessed the significant difference in mean body mass of rodents in Gamlingay and

Waresley using a GLM by including rodent species and population (Gamlingay/Waresley) as

fixed effects.

Division of the carrion niche by Nicrophorus beetles in Gamlingay and Waresley Woods

To test for significant differences in reproductive performance between species, we

conducted a GLMM regression to analyse differences in efficiency (total brood mass divided

by carcass mass), which was logit transformed prior to analysis. Beetle species, carcass size

(small/large), and their interaction were included as explanatory variables. Sampling year was

included as a random factor. In this analysis, we included beetle species as *N. interruptus*, *N.*

investigator, Gamlingay *N. vespilloides* and Waresley *N. vespilloides* to fully compare

differences not only between *Nicrophorus* beetle species, but also *N. vespilloides* between

populations.

Niche expansion in N. vespilloides by genetic accommodation

For the reaction norm experiment, we used GLMMs to test the interacting effect of

population and carcass size on brood size and average larval mass, with dyad identity nested

within block and included as a random factor. In all models, brood size and average larval

mass were analysed with a Poisson and Gaussian error distribution, respectively. A similar

statistical approach was used for analyses of clutch traits to test for the significant differences

on clutch size and average egg volume in GLMMs with a Poisson and Gaussian error

distribution, respectively. The effect of population of origin, carcass size, and their

interaction were included as fixed effects, whereas dyad identity was included as a random

factor If a significant interaction was found, a Tukey's post-hoc test was performed to detect

significant effects using multiple pairwise comparisons.

Acknowledgements

We thank P. Pilbeam for carrying out small mammal trapping in the field. We thank A.

Attisano, C. Swannack, and R. Mashoodh for help with beetle trapping. S.-J.S. was supported

by the Taiwan Cambridge Scholarship from the Cambridge Commonwealth, European &

International Trust. A.M.C. was supported by a studentship from the National Environmental

Research Council's Earth System Sciences Doctoral Training Partnership at Cambridge

University and a Victoria Brahm Schild Internship Grant from Homerton College. R.M.K.

was supported by a European Research Council Consolidator's grant 301785

BALDWINIAN_BEETLES and a Wolfson Merit Award from the Royal Society.

References

1.   Baldwin M. A new factor in evolution. Am Nat. 1896;30: 441–451.

2.   Morgan CL. On modification and variation. Science. 1896;4: 733–740.

3.   Robinson BW, Dukas R. The influence of phenotypic modifications on evolution: the
Baldwin effect and modern perspectives. Oikos. 1999;85: 582. doi:10.2307/3546709

4.   West-Eberhard MJ. Developmental plasticity and evolution. 1st ed. Oxford University
Press; 2003.

5.   Pfennig DW, Wund MA, Snell-Rood EC, Cruickshank T, Schlichting CD, Moczek
AP. Phenotypic plasticity's impacts on diversification and speciation. Trends Ecol
Evol. 2010;25: 459–467. doi:10.1016/J.TREE.2010.05.006

6.   Levis NA, Pfennig DW. Evaluating "plasticity-first" evolution in nature: key criteria
and empirical approaches. Trends Ecol Evol. 2016;31: 563–574.
doi:10.1016/j.tree.2016.03.012

7.   Levis NA, Isdaner AJ, Pfennig DW. Morphological novelty emerges from pre-existing
phenotypic plasticity. Nat Ecol Evol. 2018;2: 1289–1297.
doi:10.1038/s41559-018-0601-8

8.   Corl A, Bi K, Luke C, Challa AS, Stern AJ, Sinervo B, et al. The genetic basis of
adaptation following plastic changes in coloration in a novel environment. Curr Biol.
2018;28: 2970–2977.e7. doi:10.1016/j.cub.2018.06.075

9.   Waddington CH. Genetic assimilation of an acquired character. Evolution. 1953;7:
118–126. doi:10.1111/j.1558-5646.1953.tb00070.x

10. Lenormand T. Gene flow and the limits to natural selection. Trends Ecol Evol.
2002;17: 183–189. doi:10.1016/S0169-5347(02)02497-7

11. Sultan SE, Spencer HG. Metapopulation structure favors plasticity over local
adaptation. Am Nat. The University of Chicago Press; 2002;160: 271–83.
doi:10.1086/341015

12. Crispo E. Modifying effects of phenotypic plasticity on interactions among natural

selection, adaptation and gene flow. J Evol Biol. 2008;21: 1460–1469.

doi:10.1111/j.1420-9101.2008.01592.x

13. West-Eberhard MJ. Developmental plasticity and the origin of species differences.

Proc Natl Acad Sci U S A. 2005;102 Suppl: 6543–9. doi:10.1073/pnas.0501844102

14. Thompson JN. Relentless evolution. Chicago: University of Chicago Press; 2013.

15. Bock DG, Kantar MB, Caseys C, Matthey-Doret R, Rieseberg LH. Evolution of

invasiveness by genetic accommodation. Nat Ecol Evol. 2018;2: 991–999.

doi:10.1038/s41559-018-0553-z

16. Pascoal S, Kilner RM. Development and application of 14 microsatellite markers in

the burying beetle *Nicrophorus vespilloides* reveals population genetic differentiation

at local spatial scales. PeerJ. 2017;5: e3278. doi:10.7717/peerj.3278

17. Müller JK, Eggert A-K, Furlkröger E. Clutch size regulation in the burying beetle

Necrophorus vespilloides Herbst (Coleoptera: Silphidae). J Insect Behav. 1990;3:

265–270.

18. Scott MP. The ecology and behavior of burying beetles. Annu Rev Entomol. 1998;43:

595–618. doi:10.1086/596276

19. Otronen M. The effects of body size on the outcome of fights in burying beetles

(*Nicrophorus*). Ann Zool Fennici. 1988;25: 191–201.

20. Safryn SA, Scott MP. Sizing up the competition: do burying beetles weigh or measure

their opponents? J Insect Behav. 2000;13: 291–297. doi:10.1023/A:1007700601095

21. Urbański A, Baraniak E. Differences in early seasonal activity of three burying beetle

species (Coleoptera: Silphidae: *Nicrophorus* F.) in Poland. Coleopt Bull. 2015;69:

283–292. doi:10.1649/0010-065X-69.2.283

22. Trumbo ST. Monogamy to communal breeding: exploitation of a variable resource

base in burying beetles (*Nicrophorus*). Ecol Entomol. 1992;17: 289–298.

23. Rackham O. The history of the countryside : the classic history of Britain's landscape,

flora and fauna. Phoenix Press; 2000.

24. Rackham O. Trees and woodland in the British landscape. London: Phoenix Giant;

1996.

25.   Adamson RS. An ecological study of a Cambridgeshire woodland. J Linn Soc London,
Bot. 1912;40: 339–387. doi:10.1111/j.1095-8339.1912.tb00613.x

26.   Darby HC. Domesday woodland. Econ Hist Rev. 1950;3: 21. doi:10.2307/2589941

27.   Gibbs JP, Stanton EJ. Habitat fragmentation and arthropod community change: carrion
Beetles, phoretic mites, and flies. Ecol Appl. 2001;11: 79. doi:10.2307/3061057

28.   Lomolino M V. Ecology's most general, yet protean 1 pattern: the species-area
relationship. J Biogeogr. 2000;27: 17–26. doi:10.1046/j.1365-2699.2000.00377.x

29.   Smith RJ, Heese B. Carcass selection in a high altitude population of the burying
beetle, *Nicrophorus investigator* (Silphidae). Southwest Nat. 1995;40: 50–55.

30.   Hopwood PE, Moore AJ, Tregenza T, Royle NJ. Niche variation and the maintenance
of variation in body size in a burying beetle. Ecol Entomol. 2016;41: 96–104.
doi:10.1111/een.12275

31.   Russell AF, Langmore NE, Cockburn A, Astheimer LB, Kilner RM. Reduced egg
investment can conceal helper effects in cooperatively breeding birds. Science.
2007;317: 941–944. doi:10.1126/science.1146037

32.   Smiseth PT, Andrews CP, Mattey SN, Mooney R. Phenotypic variation in resource
acquisition influences trade-off between number and mass of offspring in a burying
beetle. J Zool. 2014;293: 80–83. doi:10.1111/jzo.12115

33.   Yi X, Liang Y, Huerta-Sanchez E, Jin X, Cuo ZXP, Pool JE, et al. Sequencing of 50
human exomes reveals adaptation to high altitude. Science. 2010;329: 75–8.
doi:10.1126/science.1190371

34.   Fumagalli M, Moltke I, Grarup N, Racimo F, Bjerregaard P, Jorgensen ME, et al.
Greenlandic Inuit show genetic signatures of diet and climate adaptation. Science.
2015;349: 1343–1347. doi:10.1126/science.aab2319

35.   Chintapalli VR, Al Bratty M, Korzekwa D, Watson DG, Dow JAT. Mapping an atlas
of tissue-specific *Drosophila melanogaster* metabolomes by high resolution mass
spectrometry. Bundy JG, editor. PLoS One. 2013;8: e78066.

696   doi:10.1371/journal.pone.0078066

36. Cao X, Jiang H. An analysis of 67 RNA-seq datasets from various tissues at different
stages of a model insect, *Manduca sexta*. BMC Genomics. 2017;18: 796.
doi:10.1186/s12864-017-4147-y

37. Ghiglione C, Carraway KL, Amundadottir LT, Boswell RE, Perrimon N, Duffy JB.
The transmembrane molecule kekkon 1 acts in a feedback loop to negatively regulate
the activity of the *Drosophila* EGF receptor during oogenesis. Cell. 1999;96: 847–56.
Available: http://www.ncbi.nlm.nih.gov/pubmed/10102272

38. Wittes J, Schüpbach T. A gene expression screen in Drosophila melanogaster
identifies novel JAK/STAT and EGFR targets during oogenesis. G3 Genes, Genomes,
Genet. 2019;9: 47–60. doi:10.1534/g3.118.200786

39. Woodring J, Hoffmann KH. The effects of octopamine, dopamine and serotonin on
juvenile hormone synthesis, In vitro, in the cricket, *Gryllus bimaculatus*. J Insect
Physiol. 1994;40: 797–802. doi:10.1016/0022-1910(94)90009-4

40. Shimada-Niwa Y, Niwa R. Serotonergic neurons respond to nutrients and regulate the
timing of steroid hormone biosynthesis in *Drosophila*. Nat Commun. 2014;5: 5778.
doi:10.1038/ncomms6778

41. Cook CN, Lawson SP, Brent CS, Rehan SM. Biogenic amines shift during the
pre-reproductive to reproductive transition in the small carpenter bee, *Ceratina*
calcarata. Apidologie. 2019;50: 90–99. doi:10.1007/s13592-018-0624-9

42. Dierick HA, Greenspan RJ. Serotonin and neuropeptide F have opposite modulatory
effects on fly aggression. Nat Genet. 2007;39: 678–682. doi:10.1038/ng2029

43. Aleksandrowicz O, Komosiński K. On the fauna of carrion beetles (Coleoptera,
Silphidae) of Mazurian lakeland (north-eastern Poland). In: J. Skłodowski, S. Huruk,
A. Barševskis ST, editor. Protection of Coleoptera in the Baltic Sea Region. Warsaw
Agricultural University Press; 2005. pp. 147–153.

44. Bubak AN, Renner KJ, Swallow JG. Heightened serotonin influences contest outcome
and enhances expression of high-intensity aggressive behaviors. Behav Brain Res.
2014;259: 137–142. doi:10.1016/j.bbr.2013.10.050

45.   Stevenson PA, Rillich J. Controlling the decision to fight or flee: the roles of biogenic
726   amines and nitric oxide in the cricket. Curr Zool. 2016;62: 265–275.
727   doi:10.1093/cz/zow028

728   46.   Scoville AG, Pfrender ME. Phenotypic plasticity facilitates recurrent rapid adaptation
729   to introduced predators. Proc Natl Acad Sci U S A. 2010;107: 4260–3.
730   doi:10.1073/pnas.0912748107

731   47.   Postma E, van Noordwijk AJ. Genetic variation for clutch size in natural populations
732   of birds from a reaction norm perspective. Ecology. 2005;86: 2344–2357.
733   doi:10.1890/04-0348

734   48.   Charmantier A, McCleery RH, Cole LR, Perrins C, Kruuk LEB, Sheldon BC.
735   Adaptive phenotypic plasticity in response to climate change in a wild bird population.
736   Science. 2008;320: 800–3. doi:10.1126/science.1157174

737   49.   Jackson DM, Trayhurn P, Speakman JR. Associations between energetics and
738   over-winter survival in the short-tailed field vole *Microtus agrestis*. J Anim Ecol.
739   2001;70: 633–640. Available:
740   https://www.abdn.ac.uk/energetics-research/publications/pdf_docs/155.pdf

741   50.   Berrigan D. The allometry of egg size and number in insects. Oikos. 1991;60: 313.
742   doi:10.2307/3545073

743   51.   Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler
744   transform. Bioinformatics. 2009;25: 1754–1760. doi:10.1093/bioinformatics/btp324

745   52.   Van der Auwera GA, Carneiro MO, Hartl C, Poplin R, del Angel G, Levy-Moonshine
746   A, et al. From FastQ data to high confidence variant calls: the Genome Analysis
747   Toolkit best practices pipeline. Current Protocols in Bioinformatics. 2013. p.
748   11.10.1-11.10.33. doi:10.1002/0471250953.bi1110s43

749   53.   Bradbury PJ, Zhang Z, Kroon DE, Casstevens TM, Ramdoss Y, Buckler ES. TASSEL:
750   software for association mapping of complex traits in diverse samples. Bioinformatics.
751   2007;23: 2633–2635. doi:10.1093/bioinformatics/btm308

752   54.   Korneliussen TS, Albrechtsen A, Nielsen R. ANGSD: Analysis of Next Generation
753   Sequencing Data. BMC Bioinformatics. 2014;15: 356.

doi:10.1186/s12859-014-0356-4

55.   Bates D, Maechler M, Bolker B, Walker S. Fitting linear mixed-effects models using
lme4. In: R package version. 2015 p. 1.0-6.

56.   Lenth R V. Least-squares means: the R package lsmeans. J Stat Softw. 2016;69: 1–33.
doi:10.18637/jss.v069.i01

57.   Clarke KR. Non-parametric multivariate analyses of changes in community structure.
Aust J Ecol. 2016;18: 117–143. doi:10.1111/j.1442-9993.1993.tb00438.x

58.   Ramette A. Multivariate analyses in microbial ecology. FEMS Microbiol Ecol.
2007;62: 142–160. doi:10.1111/j.1574-6941.2007.00375.x

765
766

769 **Figure 1.** (A) Number of *Nicrophorus* beetles caught per trap from 2014-2017. (B) Body size
770 diversity and frequency distribution of field-caught *Nicrophorus* spp. as illustrated in
771 box-and-whisker plots and kernel density estimation. Outliers are depicted as points. (C) The
772 proportion of traps set at which *N. vespilloides* was trapped with another *Nicrophorus*
773 species, in the two woodlands. Error bars indicate mean ± S.E.M.

775
776 **Figure 2.** Efficiency (%) of carcass use (total brood mass divided by carcass mass) of *N.*
777 *interruptus*, *N. investigator*, and *N. vespilloides* from Gamlingay Wood and *N. vespilloides*
778 from Waresley Wood.
779
780

781
782

783
784
785 **Figure 3.** The effect of carcass size on the brood size (A) and average larval mass (B) of *N.*
786 *vespilloides* in the reaction norm experiment (n = 46 and 46 for small and large carcasses in
787 Gamlingay Wood, and n = 62 and 62 for small and large carcasses in Waresley Wood).
788 Values represent the mean ± S.E.M.
789
790

795 **Figure 4.** The effect of carcass size on the (A) clutch size and (B) average egg volume of *N.*
796 *vespilloides.* $n = 27$ for small and large carcasses in Gamlingay Wood, and $n = 37$ for small
797 and large carcasses in Waresley Wood. Values represent the mean ± S.E.M.
798

A

B

800
801
Figure 5. Differentiation at putative oogenesis genes. (A) A scatterplot of PBS values for
Waresely and Gamlingay in 2kb windows genome-wide. Loci in the lower right hand of the
graph show high differentiation in Waresely but Gamlingay. Loci with PBS scores greater
than 0.05 are highlight – Waresley = blue, Gamlingay = red, Both = purple. Notable genes
are highlighted. (B) Sliding window analysis (window = 500bp, slide = 100bp) of PBS values
at the *5HT receptor 2a-like* receptor. The peak PBS in Waresley (blue) falls in the last intron
of the gene.
809

810 **Table 1. Population differences in enrichment scores in multiple GO terms associated with ovaries and oogenesis.**

			Waresley		Gamling		Wales	
GO ID	GO Name	Size	NES	FDR q-val	NES	FDR q-val	NES	FDR q-val
GO:0030707	ovarian follicle cell development	345	3.36	0.00E+00	3.27	1.2E-05	2.52	1.35E-03
GO:0007297	ovarian follicle cell migration	129	3.32	0.00E+00	2.86	1.0E-04	3.09	4.10E-05
GO:1905879	regulation of oogenesis	66	2.22	7.31E-03	2.07	1.4E-02		
GO:0048599	oocyte development	165	2.16	9.70E-03	1.83	4.1E-02		
GO:0007304	chorion-containing eggshell formation	78	2.12	1.20E-02	2.41	2.0E-03		
GO:0007308	oocyte construction	157	2.07	1.55E-02	1.83	4.2E-02		
GO:0030703	eggshell formation	79	2.04	1.80E-02	2.50	1.3E-03		
GO:1905881	positive regulation of oogenesis	37	2.04	1.83E-02				
GO:0009994	oocyte differentiation	194	2.01	2.05E-02	2.25	5.1E-03		
GO:0007309	oocyte axis specification	145	1.90	3.44E-02	1.85	3.9E-02		
GO:0030728	ovulation	11	1.83	4.58E-02				
GO:0007306	eggshell chorion assembly	66			2.04	1.6E-02		
GO:0060281	regulation of oocyte development	29			1.79	4.9E-02		

    Supporting information

S1 Fig. Study sites and population differences in community structure. (A) Location of

traps in Gamlingay Wood (red points) and Waresley Wood (blue points). (B) NMDS

ordination of Gamlingay ($n = 141$) and Waresley ($n = 134$) community structure. Each point

represents the sum of the beetle community collected in each trap per sampling time.

S2 Fig. Mean body weight of small mammals. Gamlingay ($n = 21$, red) and Waresley

Woods ($n = 30$, blue). One bank vole and one wood mouse, each caught in in Gamlingay

Wood, escaped before they could be measured.

830 **S3 Fig. Differences in body size (A) and frequency distribution (B) of field-caught *N.*

831 *vespilloides* from Gamlingay and Waresley Woods.** Median values, inter-quartile range,

832 maximum, and minimum are as illustrated in box-and-whisker plots. Outliers are depicted as

833 points. Pronotum width was measured as an indicator of beetle body size of Gamlingay (*n* =

834 839) and Waresley Woods (*n* = 824).

S4 Fig. Density plot of *Fst* value between Gamlingay and Waresley Woods. The plot
shows the narrow distribution of low *Fst* values between the two woodland populations.
Though there is a long tail to the right, the extreme values are modest in absolute terms.

845 **S5 Fig. MDS plot of three burying beetle populations.** The plot shows the first two

846 dimensions of a multidimensional scaling analysis of genetic diversity among burying beetles

847 in three populations. The populations do not separate out, indicating little to no genetic

848 structure.

851 **S1 Table. Quantifying *Nicrophorus* spp. body size (mm).**

Species	N	Mean	SD	Minimum	Q1	Median	Q3	Maximum
N. vespilloides	1662	4.76	0.46	3.25	4.43	4.76	5.08	6.03
N. interruptus	152	5.59	0.64	4.28	5.10	5.60	6.10	7.07
N. investigator	58	6.19	0.61	4.89	5.80	6.18	6.58	7.59
N. humator	92	6.86	0.89	4.85	6.30	6.75	7.31	9.25
N. vespillo	11	5.74	0.59	4.86	5.44	5.76	5.98	6.93

852 Number of *Nicrophorus* spp. (*N*); Standard Deviation (SD); 25% Quantile (Q1); 75%

853 Quantile (Q3).

S2 Table. Post-hoc Tukey HSD comparisons for differences in mean body size between

Nicrophorus **spp.**

Species	*N. vespilloides*		*N. interruptus*		*N. investigator*		*N. humator*	
	t	*p*	*t*	*p*	*t*	*p*	*t*	*p*
N. interruptus	18.69	**<0.001**	-	-	-	-	-	-
N. investigator	20.62	**<0.001**	-7.61	**<0.001**	-	-	-	-
N. humator	38.15	**<0.001**	18.54	**<0.001**	7.69	**<0.001**	-	-
N. vespillo	6.31	**<0.001**	-0.93	0.268	2.68	0.058	6.85	**<0.001**

p value in bold indicate comparisons with significant difference, and dashes indicate

comparisons that were not made.

S3 Table. Results of the ANOVAs for division of carrion niche by *Nicrophorus* spp.

Dependent variable	Explanatory variables	χ^2	d.f.	*p* value
Efficiency (%)	Carcass size	17.26	1	**<0.001**
	Beetle species	20.42	3	**<0.001**
	Carcass size x Beetle species	28.85	3	**<0.001**

868 *p* value in bold indicate significant differences in GLMMs.

S4 Table. Results of the ANOVAs for reaction norm experiment.

Dependent variable	Explanatory variables	χ^2	d.f.	*p* value
Brood size	Carcass size	3.33	1	0.068
	Population	3.89	1	**0.048**
	Carcass size x Population	4.80	1	**0.029**
Average larval mass	Carcass size	139.05	1	**<0.001**
	Population	1.06	1	0.304
Clutch size	Carcass size	11.33	1	**<0.001**
	Population	21.07	1	**<0.001**
	Female size	4.26	1	**0.039**
Average egg volume	Carcass size	5.90	1	**0.015**
	Population	4.59	1	**0.032**
Total egg volume	Carcass size	2.38	1	0.123
	Population	2.16	1	0.141

p value in bold indicate significant differences in GLMMs.

S5 Table. Differences in body size frequency distribution between *Nicrophorus* spp.

Species	*N. vespilloides*		*N. interruptus*		*N. investigator*		*N. humator*	
	D	*p*	*D*	*p*	*D*	*p*	*D*	*p*
N. interruptus	0.54	**<0.001**	-	-	-	-	-	-
N. investigator	0.81	**<0.001**	0.41	**<0.001**	-	-	-	-
N. humator	0.91	**<0.001**	0.62	**<0.001**	0.38	**<0.001**	-	-
N. vespillo	0.70	**<0.001**	0.31	0.268	0.43	0.061	0.69	**<0.001**

D statistic and corresponding values of significant difference from Kolmogorov-Smirnov tests. *p* value in bold indicate comparisons with significant difference, and dashes indicate comparisons that were not made.

S1 Data. Spreadsheet file of gene set enrichment analysis results of the multiple GO terms for Gamlingay and Waresley *N. vespilloides*.