

1 **Title:**

2 Short-term evolution under copper stress increases probability of plasmid uptake

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4 **Author list:**

5 Uli Klümper<sup>1,2,3,\*</sup>, Arnaud Maillard<sup>1,4,5</sup>, Elze Hesse<sup>1,3</sup>, Florian Bayer<sup>1,3</sup>, Stineke van Houte<sup>1,3</sup>, Ben Longdon<sup>1</sup>,  
6 Will Gaze<sup>2,3</sup>, Angus Buckling<sup>1,3</sup>

7 <sup>1</sup> College of Life and Environmental Sciences, University of Exeter, Penryn, Cornwall, United Kingdom

8 <sup>2</sup> European Centre for Environment and Human Health, University of Exeter Medical School, Truro,  
9 Cornwall, United Kingdom

10 <sup>3</sup> Environment & Sustainability Institute, University of Exeter, Penryn, Cornwall, United Kingdom

11 <sup>4</sup> Ecole Normale Supérieure, ENS Ulm, Paris, France

12 <sup>5</sup> UPMC, Sorbonne Universities, Paris, France

13

14 \* corresponding author:

15 Uli Klümper

16 College of Life and Environmental Sciences

17 ESI Building

18 University of Exeter

19 TR109FE Penryn, Cornwall

20 United Kingdom

21 Email: [u.klumper@exeter.ac.uk](mailto:u.klumper@exeter.ac.uk)

22 Phone: (+44)7497497338

23 ORCID: 0000-0002-4169-6548

24

25 **Abstract:**

26 Understanding plasmid transfer dynamics remains a key knowledge gap in the mitigation of antibiotic  
27 resistance gene spread. Direct effects of exposure to stressors on plasmid uptake are well monitored.  
28 However, it remains untested whether evolution of strains under stress conditions modulates subsequent  
29 plasmid uptake. Here, we evolved a compost derived microbial community for six weeks under copper  
30 stress and non-exposed control conditions. We then tested the ability of isolated clones from both  
31 treatments to take up the broad host range plasmid pJK5 from an *E.coli* donor strain. Clones pre-adapted  
32 to copper displayed a significantly increased probability to be permissive towards the plasmid compared to  
33 those isolated from the control treatment. Further, increased phylogenetic distance to the donor strain was  
34 significantly and negatively correlated with plasmid uptake probabilities across both treatments.

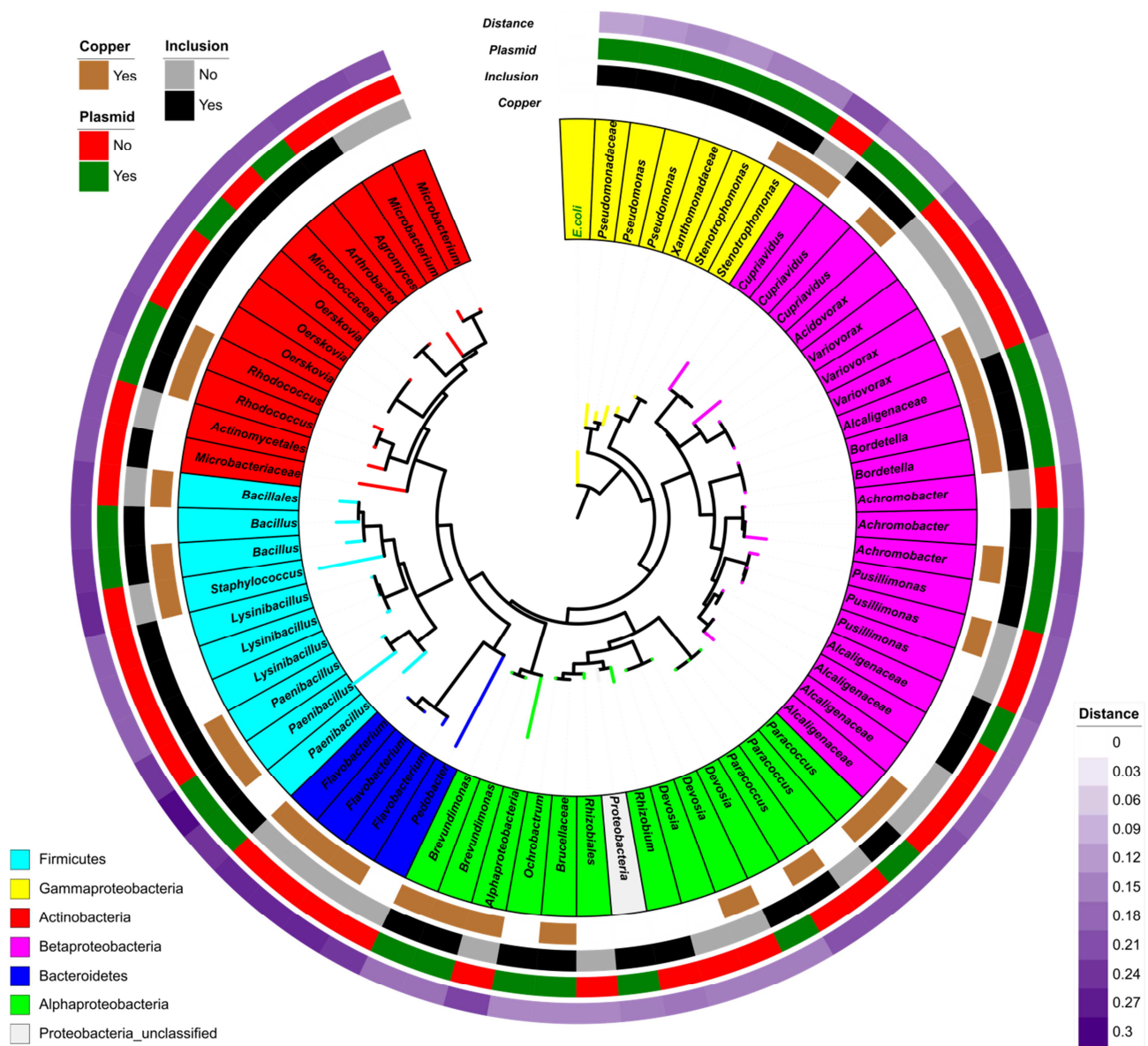
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36 **Text**

37 Conjugal plasmids serve as main means of bacterial evolutionary adaptation to environmental stressors  
38 (Norman et al., 2009). The spread of plasmids encoding antibiotic resistance is considered a major threat to  
39 human health (WHO, 2014). Crucially, understanding plasmid spread dynamics remains a key knowledge  
40 gap (Smalla et al., 2018). Direct exposure to environmental stressors such as antibiotics (Slager et al., 2014),  
41 non-antibiotic pharmaceuticals (Wang et al., 2018) or metals (Klümper et al., 2017) can modulate  
42 immediate plasmid uptake in single strains and across microbial communities. This effect can either  
43 originate from direct selection or as a by-product of a general stress response. While immediate stress  
44 effects on plasmid uptake are well monitored, it remains untested whether ecological or evolutionary  
45 selection under stress conditions results in phenotypes with intrinsically higher plasmid permissiveness.  
46 There is evidence that stress or other environmental change can select for increased mutation (Pal et al.,  
47 2007) and recombination (Cooper, 2007) rate in bacteria. We therefore hypothesize that more permissive  
48 bacteria might also be favoured, as a result of horizontally acquired adaptive stress resistance.

49 Here, we tested if evolution in a microbial community exposed to metal stress has an effect on the plasmid  
50 uptake ability of individual clones. To infer a causal relationship between exposure to copper and  
51 subsequent plasmid uptake, we set up experimental compost communities in sterile compost following the  
52 protocol of Hesse et al. (2018). Hence, all treatments started off with the same community and level of  
53 permissiveness. Microcosms were incubated (75% humidity, 26°C), and twice (after 1 and 21 days)  
54 supplemented with either 2 ml filter-sterilized 0.25M CuSO<sub>4</sub> or ddH<sub>2</sub>O. We here tested a total of 66 clones  
55 (27 copper, 39 control) that were isolated following 6 weeks of evolution. Copper tolerance increased  
56 significantly in clones pre-adapted to copper (Hesse et al., 2018). Clones were 16S sequenced using the 27F  
57 primer and a phylogenetic tree was constructed using mothur v1.41.1 (Schloss et al., 2009). Isolates  
58 belonged to 4 different phyla: Actinobacteria, Bacteroidetes, Firmicutes and three classes of Proteobacteria

59 ( $\alpha$ ,  $\beta$  &  $\gamma$ ) (Figure 1). Further, no significant difference in the distribution of isolates between metal and  
60 control treatment in the phylogenetic tree was detected (P-test,  $p=0.163$ , P-score=18).  
61

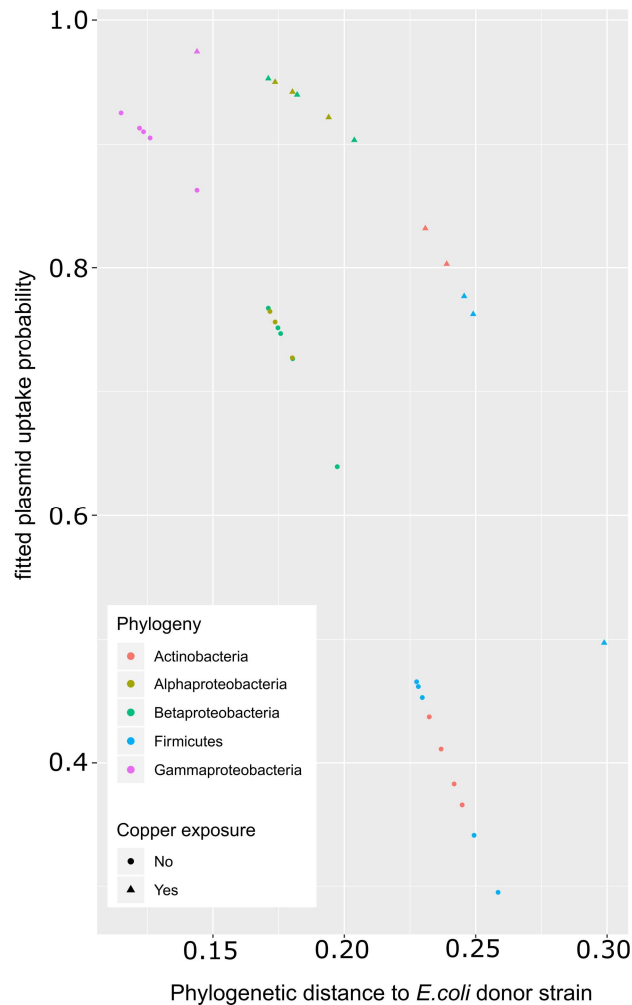


62

63 **Fig. 1: Phylogenetic tree of the 66 isolates and the donor strain *E.coli*.** Clone labels are color-coded based on  
64 phylogenetic identification. The 4 heatmap rings around the tree display: A) Isolation from the copper treatment  
65 (brown) or the control treatment (white). B) Inclusion (black) and exclusion (grey) from the study based on ability to  
66 grow on citrate while displaying susceptibility to tetracycline. C) Ability to take up plasmid pKJK5 in the experimental  
67 setup (Yes = green; No = red). D) Phylogenetic distance from the *E.coli* donor strain.  
68

69 To test permissiveness towards broad host range plasmid pKJK5 (Klümper et al., 2015; Li et al., 2018) each  
70 strain was mixed at 1:1 ratio with donor strain *Escherichia coli* MG1655::*Km<sup>R</sup>-Lpp-mCherry* hosting the IncP-  
71 1 $\epsilon$  conjugative plasmid pKJK5::*gfpmut3b* (Bahl et al., 2007; Klümper et al., 2014). Experiments were  
72 carried out in LB medium in absence of any selective pressure, centrifuged for 2 minutes at 10000xg to  
73 ensure cell-to-cell contact and incubated (24 h; 28°C). Cells were harvested, resuspended and inoculated on  
74 M9 minimal media plates supplemented with 10 mM citrate and 10  $\mu$ g/mL tetracycline. Citrate as the single  
75 carbon source counter-selects against the *E.coli* donor strain, while tetracycline selects for acquisition of  
76 the tetracycline resistance encoding plasmid. Upon successful growth, green fluorescence, repressed in the  
77 donor strain but expressed upon transfer in transconjugants, was confirmed using fluorescence  
78 microscopy. Out of the 66 strains 42 were able to grow on citrate medium and displayed susceptibility to  
79 tetracycline. These were included in subsequent analysis with 71.4% permissive to plasmid pKJK5 (Figure 1).  
80 However, permissiveness differed strongly across phyla. Out of 25 proteobacterial strains, belonging to the  
81 same phylum as the *E.coli* donor, 22 (88%) took up the plasmid, while only 47% of gram positive strains  
82 (8/17) were permissive.

83 We subsequently fitted a logistic regression model (Figure 2) with the isolates evolutionary background and  
84 phylogenetic distance to the donor strain as explanatory variables to predict plasmid uptake probability.  
85 Both copper background (ANOVA  $\chi^2$ -test,  $p=0.0416$ ,  $dF=39$ ) and phylogenetic distance (ANOVA  $\chi^2$ -test,  
86  $p=0.0033$ ,  $dF=40$ ) proved statistically significant in predicting plasmid uptake. However, while increasing  
87 distance to the donor strain had a negative effect on plasmid uptake probabilities, strains pre-adapted to  
88 copper were more likely to take up pKJK5 compared to non-adapted strains (Figure 2).



89

90 **Fig. 2: Predictive modelling of plasmid uptake probability based on phylogenetic distance to donor strain *E.coli* and**  
91 **previous metal exposure.** Fitted values of logistic regression for the 42 included strains based on the model  $p_{ij} =$   
92  $\alpha_i + \beta \times d$  ( $p_{ij}$ : the probability of pJK5 plasmid uptake in our experimental setup;  $\alpha_i$ : the effect associated with  
93 evolution under metal stress conditions;  $\beta$ : weighing parameter of the distance to the donor strain *E.coli*;  $d$ : distance  
94 to donor strain *E.coli*). Symbols are colour coded based on phylogenetic identification. Isolates from copper exposure  
95 are shown as triangles, isolates from the control treatment as circles.

96

97 Applying strong selection, such as high concentrations of metals, can have major effects on bacterial  
98 ecology and evolution (Giller et al., 1998). Here, under copper exposure phenotypes with increased  
99 likelihood to be permissive either evolved or were positively selected for. However, these two very  
100 different underpinning mechanisms cannot be distinguished at this point, as evolved communities different  
101 in their composition as a result of ecological species sorting (Hesse et al., 2018).

102 Subsequent work on single tractable species will explore the exact mechanisms underlying increased  
103 permissiveness. If, as a direct consequence of stress, more highly permissive bacteria were to survive  
104 better, they should also intrinsically host more plasmids, consequentially making them less permissive due  
105 to plasmid incompatibility or entry exclusion (Garcillán-Barcia and de la Cruz, 2008). This suggests that  
106 higher permissiveness might likely be an indirect by-product due to genetic changes in for example stress  
107 response, membrane permeability or pilus expression. Further, immunity towards plasmids could be  
108 evolutionary lost; CRISPR-Cas or less specific abortive infection systems can be lost under conditions when  
109 they bear immunity to horizontally transferred, potentially beneficial genes (Jiang et al., 2013).

110 Therefore, the conjugative host range can be increased under metal stress. The conjugative host range is  
111 generally assumed as much broader than the persistence host range (De Gelder et al., 2007). Consequently,  
112 the genomic signature of IncP-type plasmids suggests Proteobacteria as their main evolutionary hosts  
113 (Suzuki et al., 2010), which we also found to display far higher probabilities of plasmid uptake.

114 In strains with a higher degree of phylogenetic distance to the donor stability of pKJK5 might be very low  
115 and thus lost within few generations. However, long-term metal stress has been proven to elevate the  
116 retention of plasmids, even for those not coding for metal resistance (Smets et al., 2003). Even though  
117 plasmid-host co-evolution might reach an epidemiological dead end in some, likely more distant strains,  
118 such spill over remains crucial for propagation of antibiotic resistance. Unique events of recombination  
119 with the chromosome could happen before plasmid loss, especially since pKJK5 like many plasmids hosts a  
120 highly recombinative integrative element (Bahl et al., 2007). An increase of plasmid uptake ability and  
121 potentially retention time under metal stress conditions would consequently increase the likelihood and  
122 extent of such recombination events and foster the spread of antibiotic resistance.

123

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129

130 **Competing interests**

131 The authors declare no competing interests.

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133 **Author contributions**

134 UK, AM, SvH, WG and AB conceived the study and designed experiments; AM performed permissiveness  
135 assays with support from UK; EH performed evolution experiment and strain isolation; FB performed  
136 molecular work and sequencing; UK, AM, EH, BL analysed data; UK and AB wrote the manuscript.

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138 **Competing interests**

139 The authors declare no competing interests.

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141 **Materials & Correspondence**

142 All correspondence and material requests should be addressed to UK.



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