Geothermal stickleback populations prefer cool water

despite multigenerational exposure to a warm environment

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ABSTRACT

Given the threat of climate change to biodiversity, a growing number of studies are

investigating the potential for organisms to adapt to rising temperatures. Earlier work has

predicted that physiological adaptation to climate change will be accompanied by a shift

in temperature preferences, but empirical evidence for this is lacking. Here, we test

whether exposure to different thermal environments has led to changes in preferred

temperatures in the wild. Our study takes advantage of a 'natural experiment' in Iceland,

where freshwater populations of threespine sticklebacks (*Gasterosteus aculeatus*) are found

in waters warmed by geothermal activity year-round (warm habitats), adjacent to

populations in ambient-temperature lakes (cold habitats). We used a shuttle-box

approach to measure temperature preferences of wild-caught sticklebacks from three

warm-cold population pairs. Our prediction was that fish from warm habitats would

prefer higher water temperatures than those from cold habitats. We found no support for

this, as fish from both warm and cold habitats had an average preferred temperature of

13°C. Thus, our results challenge the assumption that there will be a shift in ectotherm

temperature preferences in response to climate change. In addition, since warm-habitat

fish can persist at relatively high temperatures despite a lower temperature preference,

this suggests that preferred temperature alone may be a poor indicator of a population's

adaptive potential to a novel thermal environment.

INTRODUCTION

Climate change poses a substantial threat to biodiversity, as rising temperatures are altering abiotic and biotic conditions and, in turn, imposing novel selection pressures on organisms (Crozier & Hutchings 2014). In light of this, there is now a pressing need to understand the capacity of populations to respond and adapt to increasing temperatures (Crozier & Hutchings 2014). Due to their lability, behavioural responses can play a role in facilitating or hindering adaptation to new thermal environments, depending on certain attributes of the species and environmental conditions (Sih et al. 2011, Tuomainen & Candolin 2011, van Jaarsveld et al. 2021).

Thermoregulatory behaviour could be especially important in the context of climate change, as it allows animals to buffer the effects of temperature changes by choosing suitable microhabitats (Fangue et al. 2009, Kearney et al. 2009, Huey et al. 2012, Fey et al. 2019). Animals usually seek temperatures that coincide with their optimal physiological performance and growth, which tends to be determined by their thermal evolutionary history (Jobling 1981, Kellogg & Gift 1983, Diaz et al. 2007, Pörtner & Farrell 2008, Köhler et al. 2011; but see Huey & Bennett 1987, Martin & Huey 2008, Buckley et al. 2022). Thermoregulatory behaviour is particularly important in ectotherms, because ambient temperature directly influences their body temperature, making them vulnerable to temperature changes (Zuo et al. 2011). Thus, it has been proposed that ectotherms should adapt to increasing environmental temperatures through shifts in temperature preferences that accompany evolutionary changes in their physiology (Kearney et al. 2009, Gilbert & Miles 2017, Logan et al. 2018, Catullo et al. 2019). Yet, it is still unclear whether temperature preference has the capacity to evolve in response to long-term changes in the thermal environment (Paranjpe et al. 2013, Logan et al. 2018).

We addressed this by testing whether exposure to a warm environment over

multiple generations in the wild leads to the evolution of higher preferred temperatures.

We used a unique 'natural experiment' in Iceland, where freshwater populations of

threespine sticklebacks (*Gasterosteus aculeatus*) are found in waters warmed by geothermal

activity (warm habitats) adjacent to populations in ambient-temperature lakes (cold

habitats). This study system provides repeated examples of populations experiencing

long-term contrasting thermal environments over a small geographic scale, thereby

avoiding the confounding factors associated with latitudinal or elevational comparisons.

There is evidence for strong divergence in physiology of sticklebacks from these habitats,

with warm populations showing a lower standard metabolic rate than cold populations

when compared at the same temperature (Pilakouta et al. 2020). In addition, we have

demonstrated heritable divergence in morphology, with sticklebacks from warm habitats

being more deep-bodied with shorter jaws (Pilakouta et al. *in press*). Warm-habitat

sticklebacks also tend to be less social (Pilakouta et al. 2022). Together, the consistency

of these changes across populations suggests that they are undergoing adaptive

divergence.

Given that standard metabolic rate and temperature preference have previously

been shown to be negatively correlated (Killen 2014), we predicted that fish from warm

habitats would prefer higher water temperatures than fish from cold habitats. We also

expected that this potential divergence in temperature preference may be more

pronounced in populations that have been exposed to warm water for more generations.

Lastly, we examined individual activity to determine whether inter-individual variation

in activity can explain differences in temperature preferences (Killen 2014).

METHODS

Study animals

We collected adult threespine sticklebacks from six freshwater populations in Iceland in May–June 2016 (Table 1, Figure 1). Two of these populations were allopatric, meaning that the warm and cold habitats were in neighbouring but separate water bodies with little to no potential for gene flow (Table 1). Because these populations were both located in close proximity to the marine habitat (and were thus more likely to be directly invaded by a common marine ancestor), we considered them a comparable warm-cold 'population pair'. We also sampled two sympatric warm-cold population pairs, where the warm and cold habitats were in the same water body with no physical barriers between them (Table 1). Warm- and cold-habitat sticklebacks differ in their morphology and physiology both in sympatry and allopatry (Pilakouta et al. 2020, Pilakouta et al. *in press*).

The cold habitats have likely existed for thousands of years, since the last glacial period (Einarsson et al. 2004), but there is some variation in the age of the warm habitats (Table 1). The 'Mývatn warm' and Grettislaug sites have been naturally heated by geothermal activity for over 2000 years (Hight 1965, Einarsson 1982). In contrast, the 'Áshildarholtsvatn warm' population originated only 50–70 years ago, fed by excess hot water runoff from nearby residences using geothermal heating. Since the generation time for threespine sticklebacks is about 1 year, the age of the warm habitats corresponds to the maximum number of generations each population pair may have been separated (Table 1). These different timescales made it possible to qualitatively infer whether populations exposed to higher temperatures for a relatively short time have diverged in temperature preference to the same extent as much older populations.

Animal husbandry

We placed 15 individuals from each population in 10-litre tanks in a common

recirculation system. They were fed *ad libitum* twice a day with a mixture of frozen

bloodworms, *Mysis* shrimp, and *Daphnia*. Before the experiment, all fish were

anaesthetised using benzocaine and marked with visible implant elastomer tags

(Northwest Marine Technology Inc) to allow individual identification. They were kept at

15°C and a 12h:12h daylight cycle for at least one month before being used in the

experiment. The acclimation temperature of 15°C was chosen because it is an

intermediate temperature for the warm and cold populations (Pilakouta et al. 2020). It is

close to the maximum temperature experienced by cold populations in the summer and

the minimum temperature experienced by warm populations in the winter (Table 1).

Experimental set up

We tested each individual's temperature preference using a classical shuttle-box

approach in which the animal is allowed to behaviourally adjust the temperature of its

surroundings (McCauley 1977, Schurmann & Steffensen 1992, Westhoff & Rosenberger

2016, Macnaughton et al. 2018). Our shuttle-box apparatus (Loligo Systems) consisted

of two circular 40-cm diameter choice chambers joined by a 10-cm passage way (Killen

2014). The left chamber was designated as the warm chamber and the right one as the

cold chamber. Each choice chamber was filled with water to a depth of 7 cm and was

attached to its own external buffer tank. The set-up also included a heating reservoir,

kept at 30°C using aquarium heaters, and a cooling reservoir, kept at 4°C by an external

chilling unit. To adjust the temperature within each buffer tank, water was pumped from

the buffer tanks through steel coils in the heating and cooling reservoirs. The water

temperature in the two choice chambers was continually monitored using in-line

temperature probes connected to a computer-driven temperature controller and data

acquisition system (DAQ-M, Loligo Systems). In turn, the temperature within each

chamber was controlled by software (Shuttlesoft, Loligo Systems), which adjusted the

flow from reservoir tanks to change the temperature in the choice chambers as required.

We used two ways of adjusting temperatures in the shuttle-box: a static mode and a

dynamic mode. In the static mode, there was a constant temperature in each choice

chamber with a 2°C differential between them. In the dynamic mode, the warm and cold

chamber temperatures changed depending on the location of the focal fish but always

maintained a 2°C differential. When a fish moved into the warm chamber, the

temperature increased at a rate of $2°C\ h^{-1}$ in both chambers, whereas when a fish moved

into the cold chamber, the temperature decreased at a rate of $2°C\ h^{-1}$ in both chambers.

Thus, by moving between the warm and cold chambers in response to changing

temperatures, fish could regulate the water temperature they experienced (Schurmann &

Steffensen 1992, Killen 2014). Fish movements were tracked using a camera (uEye,

Imaging Development Systems GmbH) mounted above the shuttle-box.

Experimental protocol

All experimental fish were nonbreeding, uninfected adults with their mass ranging from

0.74 to 3.02 g (mean ± SD = 1.82 ± 0.54). Our sample size was $n = 10$ for each

population (total $n = 60$). Because feeding history can influence preferred temperature,

all fish were fasted for the same amount of time (36 h) before the temperature preference

test (Killen 2014).

For the test, a single fish was placed in the shuttle-box at static mode, with the cold

chamber at 14°C and the warm chamber at 16°C. The fish was left to acclimate overnight

with the lights off. The following morning, we turned on the lights at 9:00 and allowed

the fish to acclimate for one additional hour. From 10:00 to 18:00, the system was set to

dynamic mode, and the fish was allowed to select its preferred temperature. During this

time, we also collected data on each individual's activity (total distance moved).

Core body temperature (T_b) was calculated using the following equation (Killen

2014): $T_b = T_a + (T_i - T_a)*e^{-kt}$. Here, T_a is the current water temperature experienced by

the fish, T_i is the previous temperature it experienced, t is the time elapsed since

experiencing that previous water temperature, and k is the rate coefficient for thermal

equilibration. The rate coefficient k varies with body size and is defined as the

instantaneous rate of change in body temperature in relation to the difference between T_a

and T_b (Pépino et al. 2015). We calculated k using the equation $k = 3.69*m^{-0.574}$, where m

denotes mass (Stevens and Fry 1974). The final preferred temperature of each individual

was calculated as the mean core body temperature experienced during the final two

hours of the test when the system was in dynamic mode (Killen 2014).

Data analysis

R version 3.5.1 was used for all statistical tests (R Core Team 2018), and the ggplot2

package was used for generating figures (Wickham 2009). To test for differences in

temperature preference and activity, we used linear models with the following

explanatory variables: body mass, population pair (allopatric populations, sympatric

population 1, or sympatric population 2; as defined in Table 1), thermal habitat at

population of origin (warm or cold), and the interaction between population pair and

thermal habitat. Statistical results reported below are the values from the full models for

temperature preference and activity. We also tested for a correlation between the activity

level and preferred temperature of each individual (Killen 2014).

RESULTS

There was considerable variation in preferred temperature among individuals, ranging

from 8.0°C to 15.8°C (Figure 2). However, there was no difference in mean preferred

temperature between sticklebacks collected from warm versus cold habitats ($F_{1,54}$=1.03,

P=0.31). This finding is unlikely to be due to low statistical power given that the absolute

differences in mean preferred temperature between thermal habitats were very small

(Figure 2). There was also no evidence for a greater divergence in temperature preference

in populations exposed to warm water for more generations (Figure 2). Lastly,

temperature preference was not influenced by body mass ($F_{1,54}$=1.29, P=0.26), population

pair ($F_{2,54}$=0.03, P=0.97), or the interaction between population pair and thermal habitat

($F_{2,54}$=2.87, P=0.07).

Similarly, we found no differences in activity between fish from warm and cold

habitats ($F_{1,54}$=0.152, P=0.70; Figure 3) or between fish from different population pairs

($F_{2,54}$=0.66, P=0.52; Figure 3). Activity also did not depend on body mass ($F_{1,54}$=0.003,

P=0.95) or the interaction between population pair and thermal habitat ($F_{2,54}$=2.37,

P=0.10). Despite a positive trend, there was no statistically significant relationship

between individual activity and temperature preference (t_{58}=1.81, r=0.23, P=0.076;

Figure 4).

DISCUSSION

Contrary to our hypothesis, fish from warm habitats did not prefer higher water

temperatures than fish from cold habitats when acclimated to a common temperature.

There was also no evidence for a larger difference in temperature preference between

warm- and cold-habitat fish from old populations than from young populations. Average

preferred temperature was the same for warm- and cold-habitat fish from allopatric and

sympatric populations. Thus, the absence of a difference in sympatric populations is not

necessarily due to gene flow constraining adaptation. Lastly, we found no differences in

activity between fish from warm and cold habitats, and there was no relationship

between activity and temperature preference.

In this study system, populations in the cold habitats likely represent the ancestral

populations for those in the warm habitats. Therefore, we considered the average

preferred temperature of the cold-habitat fish (13°C) to be the ancestral state. On this

basis, our results suggest that temperature preference has not diverged in warm-habitat

populations even after being exposed to elevated temperatures for hundreds to thousands

of years. This finding is in line with a recent study showing that frogs inhabiting

Japanese hot springs prefer cooler water when given the option (Komaki et al. 2020). A

similar compensatory response has been observed in the social spider *Stegodyphus*

dumicola (Malmos et al. 2021). One explanation for these patterns is that there is a lack of

genetic variation in traits associated with temperature preference. Although this may

seem surprising, it is consistent with prior work showing low to no heritability of

temperature preferences. For example, there is evidence that non-genetic factors may

play a more important role in some species. In side-blotched lizards (*Uta stansburiana*),

maternal effects but not additive genetic variation influence the offspring's temperature

preference (Paranjpe et al. 2013). Similarly, a study on brown anole lizards (*Anolis sagrei*)

found no differences in and no heritability of thermoregulatory behaviour in two

populations from contrasting thermal environments (Logan et al. 2018).

Interestingly, the average preferred temperature we observed in this study (13°C)

matches the temperature corresponding to peak immune activity in some of these

Icelandic populations (Franke et al. 2017, Franke et al. 2019). Previous work using

sticklebacks from Lake Mývatn (sympatric population 2) showed that both warm- and

cold-habitat sticklebacks had improved acquired immunity at 13°C (Franke et al. 2017,

Franke et al. 2019). Such immunity benefits of lower temperatures may also apply to

other species, as warmer environments are associated with increased disease risk,

virulence, and parasite development (Harvell et al. 2002, Macnab & Barber 2011, Altizer

et al. 2013, Franke et al. 2017).

Given that preferred temperature often coincides with optimum temperature for

growth, survival, and offspring performance (Jobling 1981, Kellogg & Gift 1983, Diaz et

al. 2007, Paranjpe et al. 2013; but see Huey & Bennett 1987, MacLean et al. 2019, Clark

et al. 2022), future work should investigate whether temperature preference reflects

optimal physiological performance in these populations (Köhler et al. 2011, Artacho et

al. 2015, Gilbert & Miles 2017). However, even if performance peaks at a relatively low

temperature (as is the case for immune activity), sticklebacks inhabiting warm habitats

are clearly able to persist at much higher temperatures during the summer months (Table

1). Hence, we suggest that preferred temperature alone may be a poor indicator of

adaptation and evolutionary potential in relation to the thermal environment.

Our results also raise the question of why warm-habitat fish from sympatric

populations are found at non-preferred temperatures in the summer, even though they

are not restricted by geographic barriers. A plausible explanation is that although only

water temperature differed between the two chambers in our experimental set-up,

temperature is not the only difference between warm and cold habitats in the wild.

Habitat choice is based on many biotic and abiotic factors, which can lead to trade-offs.

For example, predation risk may be lower in warm habitats because large piscivorous

salmonids are less able to cope with high temperatures (Eliason et al. 2011). Moreover,

warm habitats may provide a longer breeding season, leading to a higher reproductive

output each annual cycle (Hovel et al. 2016). Previous work has also found differences in

prey type availability between these warm and cold habitats in Iceland (Kreiling et al.

2018). Lastly, it is possible that warm-habitat sticklebacks are found at these sites due to social interactions, such as competition or social inertia, rather than active habitat choice (Stodola & Ward 2017, Jacob et al. 2018).

A potential caveat of our study is that all experimental fish were acclimated to a common temperature before the test, which could have led to a plastic response for a similar preferred temperature (MacLean et al. 2019). Nevertheless, we believe this is unlikely, because we still observed substantial among-individual variation in temperature preferences, ranging from 8°C to 16°C (Figure 1). In addition, we have previously shown that metabolic rate differences between warm- and cold-habitat sticklebacks persist after similar long-term acclimation to a common temperature in the laboratory; this is the case for both allopatric and sympatric population pairs (Pilakouta et al. 2020). Prior work in many fish species, including sticklebacks, has also shown that acclimation temperature does not influence final preferred temperature (Røed 1979, Kelsch & Neill 1990, Pérez et al. 2003, Diaz et al. 2007, Habary et al. 2017, Tabin et al. 2018, but see Crawshaw 1975). For example, Røed (1979) showed that sticklebacks acclimated to different temperatures for different amounts of time all showed similar preferred temperatures. Similarly, a study on a coral reef fish (*Chromis viridis*) found that preferences for cooler water persist after prolonged acclimation to high temperatures, due to an inability to acclimate at the level of the metabolic rate (Habary et al. 2017).

In sum, we find no evidence for a divergence in temperature preference between natural populations that have been exposed to contrasting thermal regimes for many generations. Our findings highlight the need to re-evaluate the common assumption that temperature preferences in ectotherms will readily evolve in response to climate change (Kearney et al. 2009, Huey et al. 2012, Gilbert & Miles 2017, Fey et al. 2019). Furthermore, the fact that warm-habitat fish can persist at high temperatures despite a

lower temperature preference suggests that preferred temperature alone may be a poor

indicator of a population's adaptive potential to a novel thermal environment.

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DATA AVAILABILITY STATEMENT

All relevant data have been uploaded to the Dryad Digital Repository:

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ETHICAL STATEMENT

Our study adheres to the ASAB/ABS Guidelines for the Use of Animals in Research,

the institutional guidelines at University of Glasgow, and the legal requirements of the

UK Home Office (Project License P89482164).

COMPETING INTERESTS

The authors declare that they have no competing interests.

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Table 1. Sampling locations of warm- and cold-habitat sticklebacks collected in May–June 2016. Distance refers to how far apart the

warm-habitat and cold-habitat sample sites are for each warm-cold pair. All cold habitats have existed since the last glacial period and are

therefore approximately 10,000 years old, whereas warm habitats can be classified as either young (<100 years old) or old (>1,000 years

old). The summer and winter temperatures listed are the average water temperatures recorded at each sampling location during the

summer and winter, respectively.

Population pair	Water body	Thermal habitat	Age of warm habitat	Distance (km)	Summer temperature (ºC)	Winter temperature (ºC)	Sample size
allopatric populations	Grettislaug	warm	old	21.04	24.9	13.5	10
	Garðsvatn	cold			14.6	2.2	10
sympatric population 1	Áshildarholtsvatn	warm	young	0.05	24.1	12.5	10
		cold			12.2	3.4	10
sympatric population 2	Mývatn	warm	old	3.18	22.8	22.0	10
		cold			11.5	1.0	10

 FIGURE LEGENDS

Figure 1. Map of Iceland showing the sampling locations of warm and cold-habitat

sticklebacks we collected for this study. Each of the three population pairs is indicated by

a different colour.

Figure 2. Boxplots of final preferred temperatures ($^{\circ}$C) for warm-habitat (red) and cold-

habitat (blue) sticklebacks from three population pairs (n=10 for sampling location).

Filled circles represent individual data points. 'Allopatric populations' refers to

Grettislaug and Garðsvatn, 'sympatric population 1' refers to Áshildarholtsvatn, and

'sympatric population 2' refers to Mývatn.

Figure 3. Boxplots of individual activity level (distance travelled in m/hr) of warm-

habitat (red) and cold-habitat (blue) sticklebacks from three population pairs (n=10 for

each sampling location). Filled circles represent individual data points. 'Allopatric

populations' refers to Grettislaug and Garðsvatn, 'sympatric population 1' refers to

Áshildarholtsvatn, and 'sympatric population 2' refers to Mývatn.

Figure 4. Scatterplot and regression line for the relationship between temperature

preference ($^{\circ}$C) and activity level (distance travelled in m/hr) of warm-habitat (red) and

cold-habitat (blue) sticklebacks. The grey shaded area shows the 95% confidence

interval.

Figure 1. Map of Iceland showing the sampling locations of warmand cold-habitat sticklebacks we collected for this study. Each of the three population pairs is indicated by a different colour.