

1 **Efficient use of genomic information for sustainable genetic improvement in**
2 **small cattle populations**

3
4 J. Obšteter^{*}, J. Jenko^{*,†}, J. M. Hickey[‡] & G. Gorjanc^{‡,§}

5
6 ^{*}Department of Animal Science, Agricultural Institute of Slovenia, Hacquetova ulica 17, 1000
7 Ljubljana, Slovenia

8 [†]Geno Breeding and A.I. Association, Storhamargata 44, 2317 Hamar, Norway

9 [‡]The Roslin Institute and Royal (Dick) School of Veterinary Studies, University of Edinburgh,
10 Easter Bush, Midlothian, EH259RG, United Kingdom

11 [§]Biotechnical Faculty, University of Ljubljana, Jamnikarjeva 101, 1000 Ljubljana, Slovenia

12
13 Jana Obšteter, Agricultural Institute of Slovenia, Department of Animal Science, Hacquetova ulica
14 17, 1000 Ljubljana, Slovenia

15 +386 1 280 51 34

16 jana.obsteter@kis.si (Corresponding Author)

17

ABSTRACT

18 This paper compares genetic gain, genetic variation, and the efficiency of converting variation into
19 gain under different genomic selection scenarios with truncation or optimum contribution selection
20 in a small dairy population by simulation. Breeding programs have to maximize genetic gain but
21 also ensure sustainability by maintaining genetic variation. Numerous studies showed that genomic
22 selection increases genetic gain. Although genomic selection is a well-established method, small
23 populations still struggle with choosing the most sustainable strategy to adopt this type of selection.
24 We developed a simulator of a dairy population and simulated a model after the Slovenian Brown
25 Swiss population with ~10,500 cows. We compared different truncation selection scenarios by
26 varying i) the method of sire selection and their use on cows or bull-dams, and ii) selection intensity
27 and the number of years a sire is in use. Furthermore, we compared different optimum contribution
28 selection scenarios with optimization of sire selection and their usage. We compared the scenarios
29 in terms of genetic gain, selection accuracy, generation interval, genetic and genic variance, the rate
30 of coancestry, effective population size, and the conversion efficiency. The results show that early
31 use of genomically tested sires increased genetic gain compared to progeny testing as expected from
32 changes in selection accuracy and generation interval. A faster turnover of sires from year to year
33 and higher intensity increased the genetic gain even further but increased the loss of genetic
34 variation per year. While maximizing intensity gave the lowest conversion efficiency, a faster turn-
35 over of sires gave an intermediate conversion efficiency. The largest conversion efficiency was
36 achieved with the simultaneous use of genomically and progeny tested sires that were used over
37 several years. Compared to truncation selection optimizing sire selection and their usage increased
38 the conversion efficiency by either achieving comparable genetic gain for a smaller loss of genetic
39 variation or achieving higher genetic gain for a comparable loss of genetic variation. Our results
40 will help breeding organizations to implement sustainable genomic selection.

41 Key words: small population, sustainability, genomic selection, optimum contribution selection

42

INTRODUCTION

43 This paper compares genetic gain, genetic variation, and the efficiency of converting variation into
44 gain under different genomic selection scenarios in a small dairy cattle population with truncation
45 or optimum contribution selection by simulation. Genomic selection has profoundly changed dairy
46 cattle breeding programs (Schaeffer, 2006; Garcia-Ruiz et al., 2016; Wiggans et al., 2017). It has
47 doubled the rate of genetic gain through decreased generation interval, increased selection accuracy
48 for young animals, increased selection intensity, and identification and management of recessive
49 lethal alleles (Cole, 2015; Garcia-Ruiz et al., 2016; Wiggans et al., 2017). The prerequisite for these
50 gains is a large number of genotyped animals, which is an issue for small populations (Thomasen et
51 al., 2014; Jenko et al., 2017; Ducrocq et al., 2018), though this problem can be addressed with
52 international training populations (Jorjani, 2012; Liu, 2013; Vandenplas et al., 2017). An effective
53 implementation also requires an optimal use of genomic selection for different groups of animals
54 (Thomasen et al., 2014). Further, small populations struggle to maximize selection intensity due to a
55 limited number of animals and limited resources, but also due to genetic drift and related genetic
56 variation issues, which can be enhanced with intense and rapid genomic selection (Falconer and
57 Mackay, 1996; Gorjanc et al., 2018).

58 Breeding programs aim to maximize genetic gain. Previous studies compared the conventional
59 progeny testing with genomic pre-selection prior to progeny testing or direct genomic selection for
60 widespread use without progeny testing (de Roos et al., 2011; Lillehammer et al., 2011; Pryce et al.,
61 2010). These studies reported up to 30% increase in genetic gain with the genomic pre-selection and
62 up to 195% increase with the direct genomic selection. Thomasen et al. (2014) deterministically
63 evaluated hybrid schemes that use both progeny and young genomically tested sires in populations
64 of different size. They concluded that genomic selection gives higher genetic gain than conventional
65 progeny testing irrespective of population size, but that the hybrid schemes maximize annual
66 monetary genetic gain when a population is small and accuracy of genomic selection is low.

67 Breeding programs also have to maintain genetic variation to ensure long-term sustainability. This
68 is especially important for small populations, since they have to be competitive in the international
69 market to justify the national breeding program. While short-term success depends on the genetic
70 gain in the next few generations, long-term success depends also on maintenance of sufficient
71 genetic variation to ensure a stable rate of genetic gain (Woolliams et al., 2015). Studies on the
72 effect of genomic selection on genetic variation have had contradictory results. For example,
73 Lillehammer et al. (2011) and Pryce et al. (2010) reported a decreased rate of coancestry per year,
74 while de Roos et al. (2011) reported that it depends on the proportion of genetic variation captured
75 with markers and a breeding program design. Genomic selection has a potential to decrease the rate
76 of coancestry due to a more accurate estimation of Mendelian sampling terms for young animals,
77 which enables differentiation of sibs and avoidance of their co-selection (Daetwyler et al., 2007).
78 Balancing short- and long-term success can be further enhanced with the optimum contribution
79 selection (Woolliams et al., 2015).

80 Although genomic selection is a well-established method, small populations still struggle with
81 choosing a sustainable strategy. The right strategy should ensure short- and long-term success as
82 well as being economically and logistically viable. To address some of these issues this study
83 evaluates different genomic breeding program designs for a small dairy population with a focus on
84 selection and usage of sires and how this affects changes in genetic gain and genetic variation.

85

MATERIAL AND METHODS

86 We compared conventional and different genomic breeding program designs in a small dairy cattle
87 population with simulation. Altogether we compared twenty-two scenarios. In fifteen scenarios we
88 used truncation selection with five different selection criteria to choose sires for the insemination of
89 cows and bull-dams. Additionally, we tested each of the sire selection criterion scenarios within
90 three sire usage scenarios that varied the number of sires and the period of their usage. To maximize
91 genetic gain for a given loss in genetic variation we compared the truncation selection scenarios
92 with seven optimum contribution selection scenarios where we varied balance between genetic gain
93 and maintenance of genetic variation. We compared all the scenarios in terms of genetic gain,
94 genetic variation, and efficiency of converting genetic variation into gain.

95 *Simulation*

96 We developed a simulator of a realistic dairy population. The simulator is a Python wrapper around
97 the simulation program AlphaSim (Faux et al., 2016; Hickey and Gorjanc, 2012), the genetic
98 evaluation program blupf90 (Misztal et al., 2002), and the optimum contribution selection program
99 AlphaMate (Gorjanc and Hickey, 2018). The simulator is driven by a set of parameters describing a
100 dairy breeding program, including the percentage of animals selected at each stage and in each
101 selection path, age at selection, selection criterion (pedigree or genomic), the number of progeny
102 per sire, the number of years a group of animals is used, and the number of years a simulation is
103 run. These parameters allow the simulation of relevant dairy breeding programs. In each year the
104 simulator generates phenotypic data, estimates breeding values, culls, selects and mates animals,
105 and generates progeny - including their pedigree and genotypic data.

106 *Population*

107 The simulated population mimicked the Slovenian Brown Swiss population of ~30,000 animals of
108 which ~10,500 are cows. The simulation started with a coalescent process to generate whole-

109 genome sequence for ten cattle-like chromosomes with 10^8 base pairs per chromosome, mutation
110 rate of 2.5×10^{-8} , recombination rate of 1.0×10^{-8} and historical effective population size in line
111 with the estimates for dairy cattle (Villa-Angulo et al., 2009; Hickey and Gorjanc, 2012). We
112 randomly sampled segregating sequence variants to construct a set of 10,000 causal variants
113 (1,000 per chromosome) and two distinct sets of 20,000 marker variants (2,000 per chromosome).
114 We used the two sets of marker variants to create two SNP arrays, one was used for genomic
115 selection and the other for monitoring “neutral” diversity. We sampled the effects of causal variants
116 from a normal distribution with a variance that gave a trait with the heritability of 0.25 in the base
117 population. Subsequently we initiated a base population by randomly allocating simulated genomes
118 to animals, which were further allocated to different categories (male and female calves, cows, bull-
119 dams, young bulls, AI bulls and natural service bulls) to initiate a dairy breeding program. We have
120 then run a conventional breeding program with selection on phenotype based estimated breeding
121 values for 20 years, followed by a further 20 years of different scenarios described below. We
122 repeated simulation of the base population and each scenario 20 times.

123 We generated 4,320 female calves every year of which we removed a random 3.7% due to stillbirths
124 and early deaths, and a further 7.5% due to other losses, for example, reproductive issues (Figure
125 S1). The remaining heifers were inseminated in the second year and became cows in the third year.
126 In each subsequent lactation we culled 20% of the starting number of cows at random and all
127 remaining cows after the fourth lactation. This scheme totaled to about 10,500 active cows per year.
128 During the first lactation we assigned 43 cows with the highest estimated breeding values as bull-
129 dams and used them for three lactations, which gave us 129 active bull-dams per year (Figure S1).
130 Every year we inseminated the best 90 bull-dams with relevant sires to generate elite male selection
131 candidates.

132 We selected sires based on genomic or progeny tests. Every year 45 elite male calves were tested
133 following one of three scenarios: a) progeny test with a pre-selection based on pedigree prediction

134 (PT), b) progeny test with a pre-selection based on genomic test (GT-PT) or c) genomic test (GT).
135 With the PT scenario, 8 out of 27 calves were chosen for progeny test based on pedigree prediction
136 in their second year, while the remaining 19 calves were used in natural service (Figure S2). With
137 the GT-PT scenario 8 out of 45 calves were chosen for progeny test based on genomic test. With the
138 PT and GT-PT scenario 5 out of 8 progeny tested bulls were selected as sires based on estimated
139 breeding value in their sixth year (Figure S2). With the GT scenario 5 out of 45 genomically tested
140 calves were directly selected as sires and were used for insemination from their second year
141 onwards. Unselected genomically tested calves in all genomic scenarios were used as natural
142 service sires (Figure S3).

143 Bull dams were inseminated with selected AI bulls only. For the insemination of cows, AI sires
144 contributed 400 doses of semen per year when 5 sires were used for 5 years and 2,000 doses per
145 year when 5 sires were used for 1 year or 1 sire was used for 5 years; natural service sires
146 contributed 27 doses; and young bulls (where applicable) contributed 250 doses. The expected
147 proportion of offspring of natural service sires therefore ranged between 7 and 17%.

148 ***Breeding value estimation***

149 We estimated breeding values with the pedigree model (Henderson, 1984) or the single-step
150 genomic model (Legarra et al., 2009) using the blupf90 program with default options (Miszta et al.,
151 2002). In genomic breeding scenarios we assumed an initial reference population of about 11,000
152 cows and 100 progeny tested sires. This mimicked the availability of international genomic
153 evaluation in Brown Swiss (Jorjani, 2012). We updated the reference population each year by
154 replacing the oldest cows with about 2,000 new cows and elite male selection candidates. Variance
155 components were assumed known and set to simulated values.

156 ***Breeding scenarios***

157 We created different truncation selection scenarios by varying i) the method of sire selection and
158 their use on cows or bull-dams, and ii) selection intensity and the number of years a sire is in use.
159 Furthermore, we created different optimum contribution selection scenarios with optimization of
160 sire selection and their usage.

161 ***Truncation selection.*** The scenarios that varied the selection of sires in combination with
162 their use on cows or bull-dams were: i) PT scenario used PT sires for the insemination of cows and
163 bull-dams, ii) GT-PT scenario used GT-PT sires for the insemination of cows and bull-dams, iii)
164 GT-C scenario used GT sires for the insemination of cows and GT-PT sires for the insemination of
165 bull-dams, iv) GT-BD scenario used GT sires for the insemination of bull-dams and GT-PT sires for
166 the insemination of cows, and v) GT scenario used GT sires for the insemination of both cows and
167 bull-dams. The GT-C and GT-BD scenarios are also referred to as the hybrid scenarios.

168 The scenarios that varied selection intensity and the number of years a sire is in use were: i) select
169 five sires every year and keep them in use for five years (5 sires/year, use 5 years), ii) reduce
170 generation interval by using five sires for one year only (5 sires/year, use 1 year) and iii) maximize
171 selection intensity by selecting only one sire and use it for five years (1 sire/year, use 5 years).

172 ***Optimum contribution selection.*** We have optimized sire selection and usage with
173 optimum contribution selection (Woolliams et al., 2015) using the AlphaMate program (Gorjanc
174 and Hickey, 2018). Every year we have added the 45 genotyped elite male calves to the pool of sires
175 selected in the previous year with a limit of 5 years for sire usage. We then optimized their
176 contributions while fixing female (heifers' and cows') contributions to one progeny per female.
177 After optimization we randomly paired the optimized male contributions with the fixed female
178 contributions. Inputs for optimum contribution selection were estimated breeding values and a
179 coancestry matrix (Woolliams et al., 2015) from the genomic single-step model (Legarra et al.,

180 2009). We optimized contributions with different emphasis on genetic gain versus group coancestry
181 using the target degrees of the angle between the truncation selection solution and an optimum
182 contribution solution (Kinghorn, 2011). For example, target degrees of 0 maximize genetic gain by
183 selecting only one male, while target degrees of 90 solely minimize group coancestry. We evaluated
184 a range of target degrees and reported results for 45, 50, 55, 60, and 75 degrees.

185 *Analysis*

186 We compared the scenarios in terms of genetic gain, selection accuracy, generation interval, genetic
187 and genic variance, the rate of coancestry, effective population size, and the efficiency of converting
188 genetic variation into genetic gain. Genetic gain was expressed as a deviation from average true
189 breeding values of the individuals in the first year of comparison in the units of genetic standard
190 deviation. Selection accuracy was measured with the Pearson correlation between the true and
191 estimated breeding values. Calibration of estimated breeding values (bias) was measured with the
192 coefficient of regression of true breeding values on estimated breeding values. Generation interval
193 was computed as the average age of the parents at the birth of their selected offspring. Genetic
194 variance measured variance of true breeding values. Genic variance measured variance of true
195 breeding values under the assumption of no linkage between causal loci. The rate of coancestry per
196 year was calculated from pedigree or genomic information. The pedigree coancestry was computed
197 following Wright (1922) from which the rate of coancestry (ΔC_P) was estimated as one minus the
198 exponent of the coefficient of regression of $\log(C_{Pt})$ on year of birth (Pérez-Enciso, 1995). The
199 genomic coancestry was computed based on the direct link with heterozygosity, $Het_t = Het_0(1 - C_t)$
200 (Falconer and Mackay, 1996). We computed heterozygosity separately for causal, marker, and
201 neutral loci. We regressed $\log(C_t)$ on the year of birth to estimate the rate of coancestry for causal
202 loci (ΔC_Q), marker loci (ΔC_M), and neutral loci (ΔC_N). Effective population size (N_e) was estimated
203 for every measure of the rate of coancestry as $1/(2\Delta C)$. Finally, the conversion efficiency was
204 measured with the coefficient of regression of the achieved genetic gain on the loss of genic

205 standard deviation (Gorjanc et al., 2018). This metric quantifies the genetic gain achieved in units of
206 genic standard deviation when all variation is converted into gain or lost due to drift. Results are
207 presented as the mean of 20 replicates for each scenario on a per year or cumulative basis. The
208 progeny testing breeding program with 5 sires selected per year and used for 5 years was the
209 baseline for comparison.

210

RESULTS

211 The results compare different breeding scenarios for a small dairy cattle population in terms of
212 genetic gain, genetic variation, and the efficiency of converting genetic variation into genetic gain.
213 The early use of genomically tested sires increased genetic gain compared to progeny testing. A
214 faster turnover of sires from year to year and higher intensity increased the genetic gain even further
215 but increased the loss of genetic variation per year. The conversion efficiency increased with the
216 simultaneous use of genomically and progeny tested sires. Maximizing intensity resulted in the
217 lowest effective population size and the lowest conversion efficiency. A faster turn-over of sires
218 decreased the conversion efficiency to an intermediate degree. Compared to truncation selection
219 optimizing male contributions increased the conversion efficiency by either achieving comparable
220 genetic gain for a smaller loss of genetic variation or achieving higher genetic gain for a comparable
221 loss of genetic variation.

222 *Genetic gain*

223 Early use of genomically tested sires, their faster turn-over and higher intensity of selection
224 increased genetic gain. This is shown in Table 1, which presents genetic gain by breeding program
225 and by sire selection and their usage scenario. Genomic pre-selection for progeny testing increased
226 genetic gain by 36% compared to the baseline. Genomic selection of sires for a direct insemination
227 of cows or bull-dams increased genetic gain respectively by 62% or 68%, and by 94% when used
228 for both, cows and bull-dams. Reducing the use of the selected sires from 5 years to 1 year further
229 increased genetic gain, between 10% and 142% compared to the baseline. Reducing the number of
230 selected sires per year from 5 to 1 and using that sire for 5 years also increased genetic gain,
231 between 21% and 124% compared to the baseline, but not compared to the scenario where 5
232 selected sires per year were used for 1 year. These genetic gains were a direct function of realized
233 generation intervals (Table S1) and selection accuracies (Table S2). Table S2 also reports the
234 calibration (bias) of estimated breeding values.

235 *Genetic and genic standard deviation*

236 Early use of genomically tested sires, their faster turn-over and higher intensity of selection
237 decreased genetic variation. This is shown in Figure 1, which presents genic and genetic standard
238 deviation by breeding program and by sire selection and their usage scenario. The genic and genetic
239 standard deviations are expressed as the percentage change to the baseline that had in the final year
240 genic standard deviation of 0.97 and genetic standard deviation of 0.94. Genomic pre-selection for
241 progeny test did not significantly change genic standard deviation. Genomic selection of sires for a
242 direct insemination of cows or bull-dams reduced genic standard deviation between 1.3% and 2.5%.
243 Reducing the number of years sires were used from 5 to 1 further reduced genic standard deviation,
244 between 0.9% and 5.0% compared to the baseline. Increasing selection intensity, by selecting only 1
245 sire per year instead of 5, reduced genic standard deviation even further, between 3.0 and 10.3%.
246 We observed a similar trend in the reduction of genetic standard deviation as for genic standard
247 deviation, but the reductions were overall larger and had higher variation between simulation
248 replicates.

249 *Effective population size*

250 Early use of genomically tested sires and increased selection intensity decreased effective
251 population size. This is shown in Table 2, which presents effective population size at causal loci by
252 breeding program and by sire selection and their usage scenario. Genomic pre-selection for progeny
253 testing did not significantly change the effective population size. Inseminating cows, bull-dams or
254 both with young genomically tested sires decreased effective population size respectively by 25%,
255 31%, and 48%. Reducing the years the sires are used from 5 to 1 did not significantly change
256 effective population size for any of the corresponding breeding scenarios. In contrast, reducing the
257 number of sires selected per year from 5 to 1 and using that sire for 5 years decreased effective
258 population size for all scenarios. The decrease ranged from 42% (with genomic pre-selection for
259 progeny testing) to 79% (when both cows and bull-dams were inseminated with one genomically

260 tested sire) compared to the baseline. These results were qualitatively the same as results for the
261 effective population sizes at marker loci used for genomic selection or at “neutral” loci (results not
262 shown).

263 *Conversion efficiency*

264 The greatest efficiency of converting genetic variation into gain was achieved with the simultaneous
265 use of genomically and progeny tested sires that were used over several years. This is shown in
266 Table 3, which presents the conversion efficiency by breeding program and by sire selection and
267 their usage scenario. This measure indicates long-term genetic gain in standard deviation units when
268 all genic variance will be exhausted and is calculated by regressing the achieved genetic gain on the
269 lost genic variance over the 20 years of selection, which we graphically represent in Figure 2 to
270 complement the Table 3. Compared to the baseline, the introduction of genomic selection increased
271 the conversion efficiency. The highest increase, 31%, was achieved with the genomic pre-selection
272 for progeny testing. Genomic selection of sires for the insemination of cows or bull-dams increased
273 the conversion efficiency respectively by 28% or 22%. Genomic selection of sires for the
274 insemination of both cows and bull-dams did not significantly increase the conversion efficiency
275 compared to the baseline. Reducing the usage of sires from 5 years to 1 year decreased the
276 conversion efficiency, except for the two scenarios with the highest genetic gain, that is, when using
277 genomically tested sires for the insemination of bull-dams or all females. Reducing the number of
278 selected sires per year to 1 and using it for 5 years reduced conversion efficiency furthermore.

279 *Optimum contribution selection*

280 Optimization of male contributions increased the conversion efficiency compared to truncation
281 selection. This is shown in Table 4 and Figure 3, which compare scenarios with truncation selection
282 and optimum contribution selection. Optimization increased the conversion efficiency when we
283 increased emphasis on maintenance of genetic variation. Therefore, there was always an optimum

284 contribution selection scenario that either achieved comparable genetic gain as a truncation
285 selection scenario, but with a smaller loss in genetic variation, or achieved larger genetic gain than a
286 truncation selection scenario with a comparable loss in genetic variation. For example, optimum
287 contribution selection with the target degrees of 75 achieved 21% higher genetic gain with a slightly
288 lower rate of coancestry than the truncation selection scenario that used 5 progeny tested sires for 5
289 years, which taken together resulted in 121% higher conversion efficiency. Similarly, optimum
290 contribution selection with the target degrees of 55 and 60 degrees achieved comparable or even
291 higher genetic gain than the truncation selection scenario that used 5 genomically tested sires for 5
292 years on cows and bull-dams, but had slightly smaller rates of coancestry, which taken together
293 increased conversion efficiency by respectively 38 and 51%. On the other hand, optimum
294 contribution selection with the target degrees of 50 achieved a 26% higher genetic gain with a
295 comparable rate of coancestry as the truncation selection scenario that used 5 genomically tested
296 sires for 5 years. Further, optimum contribution selection with the target degrees of 45 and 50 had
297 comparable genetic gain as the truncation selection scenario that used 5 genomically tested sires for
298 1 year on both, cows and bull-dams. And while the conversion efficiency for optimization at 45
299 degrees was comparable to the specified truncation scenario, optimization at 50 degrees had a 16%
300 higher conversion efficiency. Increasing the emphasis on maintenance of genetic variation in
301 optimization increased the number of selected sires and their usage over time. The average number
302 of used sires ranged from 9.6 with the target degrees of 45 to 153.0 with the target degrees of 75.
303 The years of usage ranged from 1.6 to 4.9 for the same span of target degrees.

304

DISCUSSION

305 Selection dynamics in small populations differs from that of large populations. Small populations
306 can not perform very intensive selection due to limited resources that allow for testing of only
307 limited number of individuals. Further on, due to limited number of animals and progeny per sire,
308 small populations struggle with the accuracy of progeny and genomic testing. And last, limited
309 accuracy and limited number of animals could potentially affect genetic variation of the population.
310 Despite all this, small populations have to find a way to deliver both short- and long-term genetic
311 gain to stay competitive with larger populations and to justify domestic selection. The results show
312 that we can increase genetic gain in such populations by implementing the genomic selection of
313 sires, a faster turn-over of sires, and increasing the intensity of sire selection. However, these
314 strategies also increase the loss of genetic variation, though this loss has to be assessed against the
315 larger genetic gains. For this reason, we evaluated the efficiency of converting genetic variation into
316 genetic gain. The results show that in small dairy populations the conversion efficiency can be
317 improved by the simultaneous use of genomically and progeny tested sires. Optimization of male
318 contributions can further increase the conversion efficiency. Specifically, it can increase the genetic
319 gain of the truncation selection with a comparable loss of genetic variation or it can reduce the loss
320 of genetic variation with a comparable genetic gain. To address these main findings, we divided
321 discussion into four parts: i) how genomic truncation selection affects genetic gain in small
322 populations and how this compares to large populations; ii) how genomic truncation selection
323 affects the loss of genetic variation in small populations; iii) how optimum contribution selection
324 can increase the conversion efficiency, which has implications for small and large populations; and
325 iv) how small populations could further leverage the benefits of genomic selection.

326 *Genetic gain with genomic truncation selection*

327 As expected, genomic selection increased the genetic gain in all sire selection and usage scenarios.
328 This was due to a higher selection accuracy for young non-phenotyped animals and reduced

329 generation interval (Schaeffer, 2006). Using genomic prediction as the pre-selection step increased
330 genetic gain between 37% and 59% in different scenarios without reducing generation interval. This
331 is a larger increase than in studies of larger populations (Pryce et al., 2010) or larger progeny groups
332 (Lillehammer et al., 2011). In small populations additional benefit of genomic pre-selection comes
333 from the fact that progeny testing is not as accurate as in large populations due to smaller progeny
334 groups. Reducing the generation interval by using young genomically tested sires directly on cows
335 and bull-dams further increased genetic gain between up to 144% when we used 5 sires per year
336 and up to 126% when we maximized intensity and used only 1 sire per year. These results are
337 largely in concordance with Pryce et al. (2010), Lillehammer et al. (2011) and de Roos et al. (2011),
338 although these studies evaluated typical large cattle populations with about ten-times larger number
339 of selection candidates.

340 Thomasen et al. (2014) argued that the benefit of genomic selection in small dairy populations is
341 undermined by a limited selection accuracy for young non-phenotyped animals caused by a small
342 reference population. A small reference population will invariably lead to inaccurate genomic
343 predictions. In this study we achieved comparable accuracies of about 0.8 with limited progeny test
344 and with genomic prediction based on a reference population of about 11,000 cows and 100
345 progeny tested sires, that was updated each year. Recent drops in prices for genome-wide
346 genotyping should enable small dairy populations to build such reference populations. Further,
347 some phenotyping resources could be diverted to genotyping to maximize return on investment. A
348 comparable level of accuracy can be also achieved with international reference populations (Jorjani,
349 2012; Špehar et al., 2013) or a combination of national and international reference populations
350 (Vandenplas and Gengler, 2015; Vandenplas et al., 2017; Vandenplas et al., 2018). When this level
351 of accuracy is combined with a reduced generation interval, small populations can achieve
352 substantially larger genetic gains than with progeny testing. Finally, increasing the selection
353 intensity to the unrealistic use of just one sire, to come closer to the intensity of selection in large

354 populations, further increased genetic gain, but with a considerable loss in genetic variation that
355 started to limit genetic gain within the simulated 20 years.

356 *Loss of genetic variation with genomic truncation selection*

357 The results show that small populations can increase genetic gain without increasing the loss of
358 genetic variation by using genomic pre-selection of bulls for progeny testing. All other genomic
359 selection scenarios increased the loss of genetic variation compared to a conventional scenario with
360 progeny testing, although the accuracies of progeny and genomic tests were comparable and that we
361 selected the same number of sires per year. We observed this with genic and genetic variance as
362 well as effective population size (measured with pedigree and neutral, marker or causal loci). While
363 losses of genic and genetic variance in the simulated period of 20 years were not substantial (at
364 most 0.13 genic standard deviation), the changes in effective population size were substantial –
365 from about 175 with the conventional scenarios to about 80 with the full genomic scenarios, which
366 indicates reduced sustainability.

367 Our results for the rate of coancestry are not in concordance with what was observed in studies of
368 large populations (Pryce et al., 2010) or with higher selection intensity (Lillehammer et al., 2011).
369 which observed lower rates with genomic selection. However, lower intensity of selection in small
370 populations stems from fewer tested animals, and not more selected, which reduces a genetic pool
371 for selection. Our results are more in line with Doekes et al. (2018). They attribute the higher rates
372 of inbreeding with genomic selection to the fact, that the animals with a higher relatedness to the
373 reference population have more accurate genomic predictions and are more likely to deviate
374 substantially and therefore to be selected (Habier et al., 2007; Clark et al., 2012). Another
375 explanation for a larger loss of genetic variability with genomic selection is that shortening
376 generation interval increases the turnover of germplasm from year to year, which increases genetic
377 gain per unit of time, but also increases the loss of genetic variation per unit of time (Buch et al.,

378 2012; Boichard et al., 2015; Gorjanc et al., 2018). Further, studies mostly report the rate of
379 inbreeding, which measures increase in individual homozygosity (Pryce et al., 2010; Doekes et al.
380 2018), while we report the rate of coancestry, which measures increase in population homozygosity.
381 While these two measures are correlated, it is the rate of coancestry that determines the
382 sustainability of a breeding program.

383 To compare the simultaneous change in genetic gain and loss in genetic variation we compared
384 different scenarios with the efficiency of converting genetic variation into genetic gain. We
385 measured this with a linear regression of the achieved genetic gain on the lost genetic standard
386 deviation (Gorjanc et al., 2018). We found that in small cattle populations genomic pre-selection for
387 progeny test and hybrid scenarios achieved the highest conversion efficiencies. The two extremes –
388 conventional and complete genomic scenarios – were the least efficient. Despite their similar
389 conversion efficiencies, there are large differences between these scenarios – namely, the genomic
390 scenario almost doubled genetic gain. The conventional scenario had low conversion efficiency due
391 to a small genetic gain (caused by long generation intervals) although it retained most of genetic
392 variation. The low conversion efficiency of the conventional scenarios could be specific to small
393 populations, since the accuracy and selection intensity of progeny testing is smaller than in large
394 populations. The completely genomic scenario had low conversion efficiency despite a large genetic
395 gain (caused by short generation intervals) as it lost the most of genetic variation.

396 Increasing the turnover of the sires and increasing selection intensity have different consequences
397 on short and long-term success of selection. Although both of these scenarios increase genetic gain
398 (up to 125%), increasing the intensity also increased the loss of genetic variation and in turn
399 reduced conversion efficiency. Increased turn-over of sires from 5 to 1 year in this study achieved
400 higher genetic gain over the 20 years than reducing the number of sires from 5 to 1, because it did
401 not impact genetic variation so severely.

402 *Comparison of truncation and optimum contribution selection*

403 Optimization of male contributions increased the conversion efficiency of truncation selection
404 scenarios. The optimization involved all active males - the young calves with genomic prediction
405 and sires selected in previous years – either young sires with genomic test or older with progeny
406 test. Optimum contribution selection with genomic information has been tested before (e.g. Clark et
407 al., 2013) with the conclusion that there is not much scope for optimization with genomic
408 relationships unless there are very large full-sib families. Here we use optimal contribution selection
409 to optimize selection and usage of genomically and progeny tested bulls of different ages and
410 observe substantial differences over 20 years in a small dairy population. We achieved this by
411 optimizing male contributions with a range of emphasis on genetic gain versus maintenance of
412 genetic variation. In this we followed the multi-objective approach of Kinghorn (2011), where the
413 emphasis is measured with the angle between truncation selection solution and targeted optimum
414 contribution selection solution.

415 For every truncation selection scenario, we found an optimum contribution selection scenario that
416 increased conversion efficiency. This higher efficiency was either achieved with the same genetic
417 gain but smaller loss of genetic variation than truncation selection or with a higher genetic gain and
418 the same loss of genetic variation as truncation selection. This improvement was achieved by
419 optimized selection and usage of sires. For example, the average number of sires with the truncation
420 selection of 5 progeny tested sires that were used for 5 years was about 55 (this includes young,
421 natural service and proven bulls). Here the sires of the same age and the same status had an
422 approximately the same number of progeny. This scenario achieved genetic gain of 2.50 genetic
423 standard deviations, generation interval for sire-sire and sire-dam paths of 9.0 and 7.0 years,
424 effective population size of 172 and conversion efficiency of 77. A comparable number of sires (49)
425 was used with the optimization targeting 60 degrees, which involved mostly young sires (3 years in
426 use). Their optimized usage delivered genetic gain of 4.77 genetic standard deviations, generation

427 interval for sire-sire and sire-dam paths of 3.3 and 3.1 years, effective population size of 144 and
428 conversion efficiency of 126. The highest genetic gain was achieved with the targeted degrees
429 between 45 and 50. These targets drive optimization to achieve every year between 71% and 65%
430 of maximum possible genetic gain with truncation selection and between 71% and 77% minimum
431 possible group coancestry (Kinghorn, 2011; Gorjanc and Hickey, 2018). Further, although the
432 optimization could choose genomically and progeny tested bulls, we observed that it chose mostly
433 young genomically tested bulls, for example the maximum years in use was on average 4.9 when
434 we optimized for 75 target degrees. This is in contrast with truncation selection scenarios, where the
435 highest conversion efficiency was achieved with the simultaneous use of genomically and progeny
436 tested bulls.

437 The results have implications also for large populations, namely they show that genomic selection is
438 increasing turnover of germplasm per year with positive effect on genetic gain and negative effect
439 on genetic variation. This has been already indicated in real large populations (Doekes et al., 2018).
440 While our results are likely specific to small populations, combining these with the results from a
441 wheat simulation study (Gorjanc et al., 2018) that used a small or a large number of parents suggest
442 that both small and large populations can increase the conversion efficiency of genomic selection by
443 optimizing contributions.

444 *Further opportunities*

445 There are further opportunities with genomic selection for small populations that we have not
446 addressed in this study. We specifically highlight the increasing number of genotyped females and
447 the role of importation of external genetics. In this paper we have focused only on comparing male
448 selection and usage strategies that required minimal changes to a breeding program. However,
449 genotyping prices have decreased substantially in the recent years and it's likely that in future a
450 significant proportion of cows will be genotyped. This will increase accuracy of genetic evaluation

451 of cows early in their lives and enable even shorter generation intervals. It will also enable accurate
452 assessment of relationships amongst cows and bulls and open possibility for further optimization.
453 This has been partially realized in this study by combining genomic and pedigree relationships
454 through the single-step genetic evaluation method, which propagates all the genomic information
455 throughout a pedigree (Legarra et al., 2009).

456 Many dairy breeding programs, small and large, supplement their internal breeding activities with
457 importation of external genetics. Importation is of particular importance for small populations
458 because they struggle to be competitive due to limited financial resources for collecting data and
459 limited numbers of animals for collecting data and for use as selection candidates. Combining own
460 breeding and importation opens further possibilities for optimization as it expands the genetic pool
461 for breeding. Further, genomic selection now enables accurate genetic evaluation and relationship
462 of foreign animals to a local population. Such foreign animals could be added into the presented
463 optimization to exploit the expanded genetic pool and further increase sustainability of small
464 breeding populations.

465

CONCLUSION

466 This paper evaluated different genomic breeding programs in a small dairy cattle population with
467 truncation selection to quantify its short- and long-term success. Furthermore, it evaluated the value
468 of optimizing male contributions to increase efficiency of converting genetic variation into genetic
469 gain. We concluded that genomic selection increases short-term genetic gain but can also improve
470 long-term genetic gain when used in combination with conventional selection. We also showed that
471 optimum contribution selection improves conversion efficiency at a comparable genetic gain or
472 achieves higher genetic gain at a similar conversion efficiency. Our results will be of help to
473 breeding organization that aim to implement sustainable genomic selection.

474

ACKNOWLEDGMENTS

475 The authors acknowledge T. Perpar (Agricultural Institute of Slovenia), M. Rigler (Chamber of
476 Agriculture and Forestry of Slovenia), and K. Potočnik (Biotechnical Faculty, University of
477 Ljubljana) for their advice about the Brown-Swiss breeding scheme and Andres Legarra (INRA)
478 and Ivan Pocrnic (University of Georgia) for their advice about preparing the single-step coancestry
479 matrix. The first author also thanks the European Association for Animal Production (EAAP) for
480 scholarship to present preliminary results of this study at the 2018 annual meeting. G. Gorjanc and
481 J. M. Hickey acknowledge support from the BBSRC funding to The Roslin Institute
482 (BBS/E/D/30002275).

483

APPENDIX

Table S1: Generation interval by path of selection, by breeding program and by sire selection and their usage scenario.

| Breeding program | Sire selection and usage | | | | | |
|------------------|-------------------------------------|------------------------------------|------------------------------------|------------------------------------|------------------------------------|------------------------------------|
| | 5 sires/year, use 5 years | | 5 sires/year, use 1 year | | 1 sire/year, use 5 years | |
| | Sire of sires | Sire of dams | Sire of sires | Sire of dams | Sire of sires | Sire of dams |
| PT | 9.0 _{0.06} ^{ab,A} | 7.0 _{0.05} ^{a,A} | 7.1 _{0.00} ^{a,B} | 5.8 _{0.02} ^{a,B} | 9.1 _{0.07} ^{a,C} | 7.7 _{0.00} ^{a,C} |
| GT-PT | 9.0 _{0.06} ^{a,A} | 7.0 _{0.05} ^{a,A} | 7.1 _{0.00} ^{a,B} | 5.8 _{0.00} ^{a,B} | 9.1 _{0.07} ^{a,C} | 7.7 _{0.02} ^{a,C} |
| GT-C | 9.0 _{0.05} ^{b,A} | 4.1 _{0.04} ^{b,A} | 7.1 _{0.00} ^{a,B} | 2.5 _{0.00} ^{b,B} | 9.1 _{0.06} ^{a,C} | 4.1 _{0.00} ^{b,A} |
| GT-BD | 3.8 _{0.05} ^{c,A} | 7.0 _{0.05} ^{c,A} | 3.8 _{0.05} ^{b,A} | 5.7 _{0.00} ^{c,B} | 3.8 _{0.04} ^{b,A} | 7.6 _{0.00} ^{c,C} |
| GT | 4.2 _{0.05} ^{d,A} | 3.9 _{0.05} ^{d,A} | 2.3 _{0.05} ^{c,B} | 2.3 _{0.00} ^{d,B} | 4.2 _{0.07} ^{c,A} | 3.9 _{0.00} ^{d,C} |

484 PT = conventional progeny testing; GT-PT = genomic pre-selection of bulls for progeny testing, GT-C = genomic
 485 selection of sires for the insemination of cows; GT-BD = genomic selection of sires for the insemination of bull-dams;
 486 GT = genomic selection of sires for the insemination of cows and bull-dams. Subscript numbers indicate standard
 487 deviation across simulation replicates. Lower-case letters denote statistically significant differences between breeding
 488 programs and upper-case letters between sire selection and usage scenarios.

489

490 Table S2: Accuracy of selection and prediction bias by animal category, by breeding program and
 491 by sire selection and their usage scenario.

| Breeding program | Sire selection and usage | | | | | |
|------------------|---------------------------|----------------------|--------------------------|----------------------|--------------------------|----------------------|
| | 5 sires/year, use 5 years | | 5 sires/year, use 1 year | | 1 sire/year, use 5 years | |
| | Accuracy | Bias | Accuracy | Bias | Accuracy | Bias |
| PT | | | | | | |
| young bulls | 0.31 _{0.18} | 0.17 _{0.11} | 0.23 _{0.20} | 0.12 _{0.11} | 0.30 _{0.19} | 0.17 _{0.12} |
| sires | 0.88 _{0.10} | 0.83 _{0.18} | 0.87 _{0.10} | 0.82 _{0.18} | 0.82 _{0.17} | 0.76 _{0.22} |
| heifers | 0.35 _{0.05} | 0.22 _{0.03} | 0.32 _{0.06} | 0.19 _{0.05} | 0.36 _{0.06} | 0.22 _{0.05} |
| GT-PT | | | | | | |
| young bulls | 0.80 _{0.11} | 1.01 _{0.22} | 0.78 _{0.12} | 1.01 _{0.23} | 0.80 _{0.11} | 1.06 _{0.23} |
| sires | 0.82 _{0.16} | 0.92 _{0.27} | 0.83 _{0.15} | 0.92 _{0.25} | 0.78 _{0.20} | 0.88 _{0.31} |
| heifers | 0.47 _{0.04} | 0.34 _{0.05} | 0.44 _{0.05} | 0.30 _{0.05} | 0.46 _{0.06} | 0.33 _{0.06} |
| GT-C | | | | | | |
| young bulls | 0.81 _{0.11} | 1.02 _{0.21} | 0.80 _{0.11} | 1.03 _{0.22} | 0.82 _{0.12} | 1.07 _{0.23} |
| sires | 0.89 _{0.09} | 0.93 _{0.18} | 0.90 _{0.08} | 0.94 _{0.21} | 0.87 _{0.13} | 0.89 _{0.20} |
| heifers | 0.44 _{0.05} | 0.31 _{0.05} | 0.41 _{0.05} | 0.27 _{0.05} | 0.46 _{0.07} | 0.34 _{0.08} |
| GT-BD | | | | | | |
| young bulls | 0.77 _{0.11} | 1.00 _{0.22} | 0.78 _{0.12} | 1.00 _{0.23} | 0.77 _{0.11} | 0.98 _{0.22} |
| sires | 0.84 _{0.14} | 0.92 _{0.23} | 0.82 _{0.16} | 0.92 _{0.29} | 0.80 _{0.17} | 0.89 _{0.27} |
| heifers | 0.51 _{0.06} | 0.39 _{0.07} | 0.46 _{0.06} | 0.32 _{0.06} | 0.49 _{0.06} | 0.36 _{0.06} |
| GT | | | | | | |
| young bulls | 0.79 _{0.12} | 1.03 _{0.22} | 0.75 _{0.11} | 0.96 _{0.22} | 0.80 _{0.12} | 1.16 _{0.30} |
| heifers | 0.48 _{0.05} | 0.36 _{0.06} | 0.43 _{0.06} | 0.30 _{0.05} | 0.49 _{0.07} | 0.39 _{0.13} |

492 PT = conventional progeny testing; GT-PT = genomic pre-selection of bulls for progeny testing, GT-C = genomic
 493 selection of sires for the insemination of cows; GT-BD = genomic selection of sires for the insemination of bull-dams;
 494 GT = genomic selection of sires for the insemination of cows and bull-dams. Subscript numbers indicate standard
 495 deviation across simulation replicates.

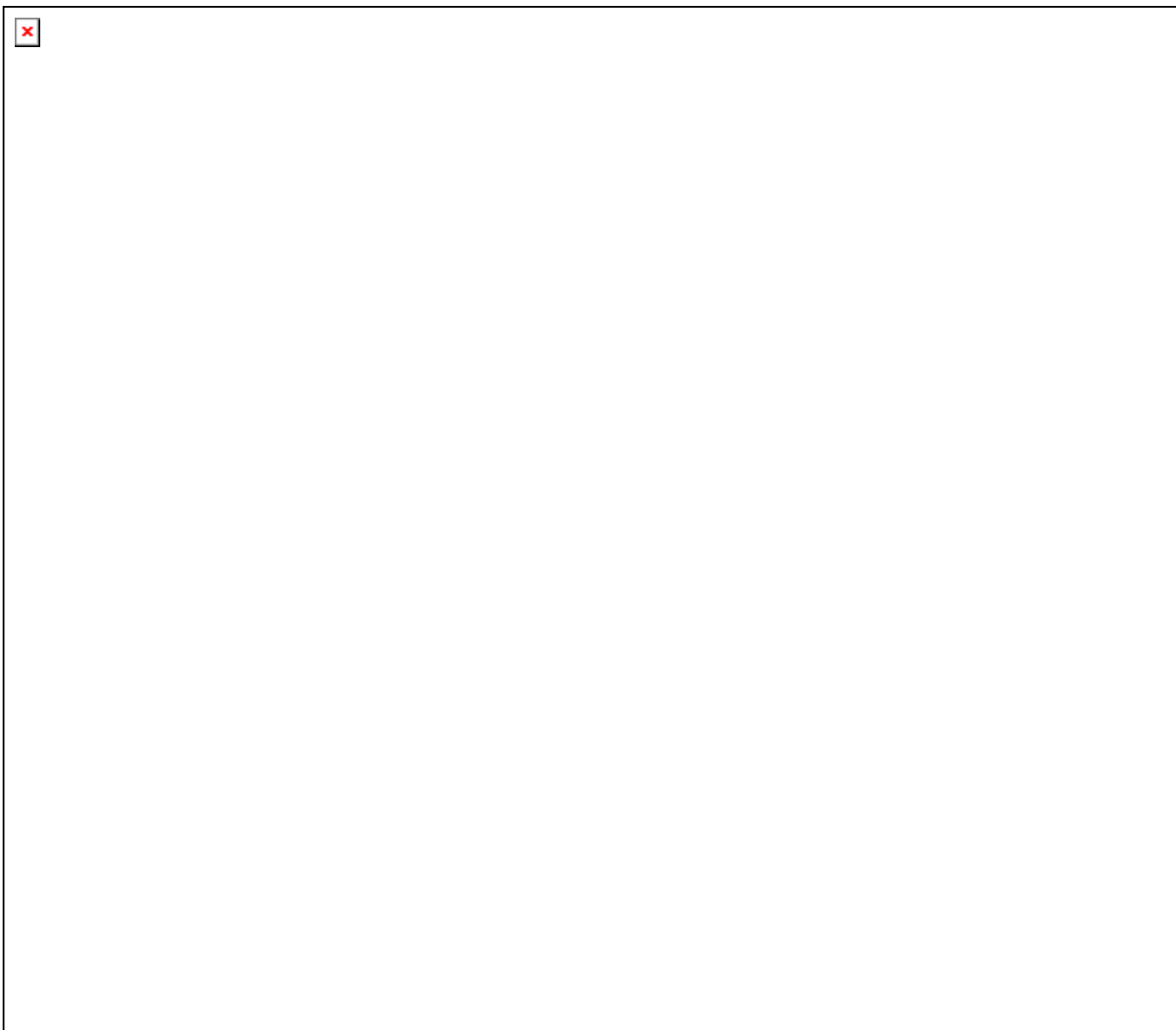
496 Figure S1: Description of the population structure and selection procedure of the simulated
497 population for females. The arrows represent selection decisions and the numbers in bold represent
498 the number of animals in each category.



499

500 *AI = artificial insemination

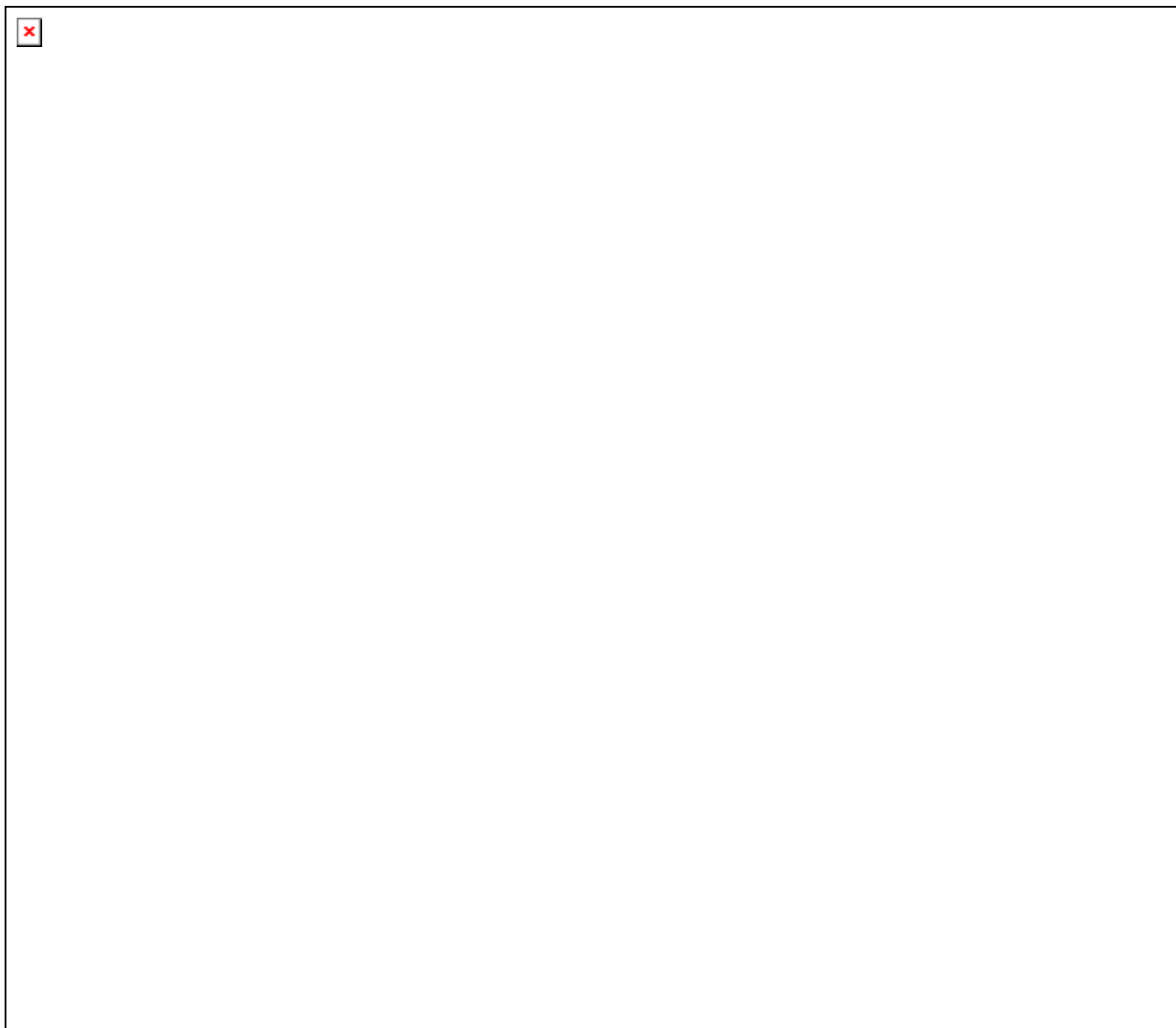
501 Figure S2: Description of the population structure and selection procedure of the simulated
502 population for progeny tested males. The arrows represent selection decisions and the numbers in
503 bold represent the number of animals in each category.



504
505

*AI = artificial insemination

506 Figure S3: Description of the population structure and selection procedure of the simulated
507 population for genomically tested males. The arrows represent selection decisions and the numbers
508 in bold represent the number of animals in each category.



509
510

*AI = artificial insemination

REFERENCES

- Boichard, D., V. Ducrocq, and S. Fritz. 2015. Sustainable dairy cattle selection in the genomic era. *J. Anim. Breed. Genet.* 132:135–143. <https://doi.org/10.1111/jbg.12150>
- Buch, L., M. Sørensen, P. Berg, L. Pedersen, and A. Sørensen. 2012. Genomic selection strategies in dairy cattle: Strong positive interaction between use of genotypic information and intensive use of young bulls on genetic gain. *J. Anim. Breed. Genet.* 129:138-151. doi:10.1111/j.1439-0388.2011.00947.x
- Clark, S. A., J. M. Hickey, H. D. Daetwyler, and J. H. van der Werf. 2012. The importance of information on relatives for the prediction of genomic breeding values and the implications for the makeup of reference data sets in livestock breeding schemes. *Genetics, selection, evolution : GSE*, 44: 4. doi:10.1186/1297-9686-44-4
- Clark, S. A., B. P. Kinghorn, J. M. Hickey and J. H. van der Werf. 2013. The effect of genomic information on optimal contribution selection in livestock breeding programs. *Genetics Selection Evolution.* 45:44. <https://doi.org/10.1186/1297-9686-45-44>
- Cole, J. B. 2015. A simple strategy for managing many recessive disorders in a dairy cattle breeding program. *Genet. Sel. Evol.* 47:94. <https://doi.org/10.1186/s12711-015-0174-9>
- Daetwyler, H. D., B. Villanueva, P. Bijma, and J. A. Woolliams. 2007. Inbreeding in genome-wide selection. *J. Anim. Breed. Genet.* 124:369–376.
- de Roos, A. P. W., C. Schrooten, R. F. Veerkamp, and J. A. M. van Arendonk. 2011. Effects of genomic selection on genetic improvement, inbreeding, and merit of young versus proven bulls. *J. Dairy Sci.* 94:1559–1567. <https://doi.org/10.3168/jds.2010-3354>
- Doekes, H. P., R. F. Veerkamp, P. Bijma, S. J. Hiemstra, and J. J. Windig, J.J. 2018. Trends in genome-wide and region-specific genetic diversity in the Dutch-Flemish Holstein–Friesian breeding program from 1986 to 2015. *Genet. Sel. Evol.* 50:15. <https://doi.org/10.1186/s12711-018-0385-y>

- Ducrocq, V., D. Laloe, M. Swaminathan, X. Rognon, M. Tixier-Boichard, and T. Zerjal. 2018. Genomics for Ruminants in Developing Countries: From Principles to Practice. *Front. Genet.* 9:251. <https://doi.org/10.3389/fgene.2018.00251>
- Falconer, D. S. and T. F. C. Mackay. 1996. *Introduction to Quantitative Genetics*. 4th ed. Longmans Green, Harlow, Essex, UK.
- Faux, A. M., G. Gorjanc, R. C. Gaynor, M. Battagin, S. M. Edwards, D. L. Wilson, S. J. Hearne, S. Gonen, and J. M. Hickey. 2016. AlphaSim: Software for Breeding Program Simulation. *Plant Genome* 9:1-14. <https://doi.org/10.3835/plantgenome2016.02.0013>
- García-Ruiz, A., J. B. Cole, P. M. VanRaden, G. R. Wiggans, F. J. Ruiz-López, and C. P. V. Tassell. 2016. Changes in genetic selection differentials and generation intervals in US Holstein dairy cattle as a result of genomic selection. *Proc. Natl. Acad. Sci.* 113:e3995–e4004. <https://doi.org/10.1073/pnas.1519061113>
- Gorjanc, G., R. C. Gaynor, and J. M. Hickey. 2018. Optimal cross selection for long-term genetic gain in two-part programs with rapid recurrent genomic selection. *Theor. Appl. Genet.* 131:1953–1966. <https://doi.org/10.1007/s00122-018-3125-3>
- Gorjanc, G., and J. M. Hickey. 2018. AlphaMate: a program for optimizing selection, maintenance of diversity and mate allocation in breeding programs. *Bioinformatics* 34:3408–3411. <https://doi.org/10.1093/bioinformatics/bty375>
- Habier, D., R. L. Fernando, and J. C. M. Dekkers. 2007. The impact of genetic relationship information on genome-assisted breeding values. *Genetics* 177:2389–2397. <https://doi.org/10.1534/genetics.107.081190>
- Henderson, C. R. 1984. *Applications of Linear Models in Animal Breeding*. University of Guelph, Ontario, Canada.

- Hickey, J. M., and G. Gorjanc. 2012. Simulated data for genomic selection and genome-wide association studies using a combination of coalescent and gene drop methods. *G3* (Bethesda, Md.), 2:425–427. doi:10.1534/g3.111.001297
- Jenko, J., G. R. Wiggans, T. A. Cooper, S. A. E. Eaglen, W. G. de L. Luff, M. Bichard, R. Pong-Wong, and J. A. Woolliams. 2017. Cow genotyping strategies for genomic selection in a small dairy cattle population. *J. Dairy Sci.* 100:439–452. <https://doi.org/10.3168/jds.2016-11479>
- Jorjani, H. 2012. Genomic evaluation of BSW populations, InterGenomics: Results and Deliverables. *Interbull Bull.* 43:5-8.
- Kinghorn, B. P. 2011. An algorithm for efficient constrained mate selection. *Genet. Sel. Evol.* 43:4. <https://doi.org/10.1186/1297-9686-43-4>
- Legarra, A., I. Aguilar, and I. Misztal. 2009. A relationship matrix including full pedigree and genomic information. *J. Dairy Sci.* 92:4656–4663. <https://doi.org/10.3168/jds.2009-2061>
- Lillehammer, M., T. H. E. Meuwissen, and A. K. Sonesson. 2011. A comparison of dairy cattle breeding designs that use genomic selection. *J. Dairy Sci.* 94:493–500. <https://doi.org/10.3168/jds.2010-3518>
- Misztal, I., S. Tsuruta, T. Strabel, B. Auvray, T. Druet, and D. H. Lee. 2002. BLUPF90 and related programs (BGF90). Pages 1-2 in Proc. 7th World Congress on Genetics Applied to Livestock Production, Montpellier, France. Australian Egg Corporation, Hurstville, NSW, Australia.
- Pérez-Enciso, M. 1995. Use of the uncertain relationship matrix to compute effective population size. *J. Anim. Breed. Genet.* 112:327–332. <https://doi.org/10.1111/j.1439-0388.1995.tb00574.x>

- Pryce, J. E., M. E. Goddard, H. W. Raadsma, and B. J. Hayes. 2010. Deterministic models of breeding scheme designs that incorporate genomic selection. *J. Dairy Sci.* 93:5455–5466. <https://doi.org/10.3168/jds.2010-3256>
- Schaeffer, L. R. 2006. Strategy for applying genome-wide selection in dairy cattle. *J. Anim. Breed. Genet.* 123:218–223. <https://doi.org/10.1111/j.1439-0388.2006.00595.x>
- Špehar, M., K. Potočnik, G. Gorjanc. 2013. Accuracy of genomic prediction for milk production traits with different approaches in a small population of Slovenian Brown bulls. *Livest. Sci.* 157:421–426. <https://doi.org/10.1016/j.livsci.2013.08.008>
- Thomasen, J. R., C. Egger-Danner, A. Willam, B. Guldbrandtsen, M. S. Lund, M.S., and A. C. Sørensen. 2014. Genomic selection strategies in a small dairy cattle population evaluated for genetic gain and profit. *J. Dairy Sci.* 97:458–470. <https://doi.org/10.3168/jds.2013-6599>
- Vandenplas, J., and N. Gengler. 2015. Strategies for comparing and combining different genetic and genomic evaluations: A review. *Livestock Science.* 181:121-130. <https://doi.org/10.1016/j.livsci.2015.09.012>.
- Vandenplas, J., M. Spehar, K. Potocnik, N. Gengler and G. Gorjanc. 2017. National single-step genomic method that integrates multi-national genomic information. *J Dairy Sci.* 100:465-478. doi: 10.3168/jds.2016-11733.
- Vandenplas, J., M. P. L. Calus and G. Gorjanc. 2018. Genomic Prediction Using Individual-Level Data and Summary Statistics from Multiple Populations. *Genetics.* 210:53-69. doi: 10.1534/genetics.118.301109.
- Wiggans, G. R., J. B. Cole, S. M. Hubbard, and T. S. Sonstegard. 2017. Genomic Selection in Dairy Cattle: The USDA Experience. *Annu. Rev. Anim. Biosci.* 5:309–327. <https://doi.org/10.1146/annurev-animal-021815-111422>
- Woolliams, J. A., P. Berg, B. S. Dagnachew, and T. H. E. Meuwissen. 2015. Genetic contributions and their optimization. *J. Anim. Breed. Genet.* 132:89–99. <https://doi.org/10.1111/jbg.12148>

Wright, S. 1922. Coefficients of Inbreeding and Relationship. *Am. Nat.* 56:330–338.

512 Table 1: Genetic gain in genetic standard deviation units by breeding program and by sire selection
 513 and their usage scenario.

| Breeding program | Sire selection and usage | | |
|------------------|--------------------------------------|--------------------------------------|--------------------------------------|
| | 5 sires/year, use 5 years | 5 sires/year, use 1 year | 1 sire/year, use 5 years |
| PT | 2.50 _{0.22} ^{a, A} | 2.75 _{0.19} ^{a, B} | 3.03 _{0.11} ^{a, C} |
| GT-PT | 3.41 _{0.14} ^{b, A} | 3.96 _{0.17} ^{b, B} | 3.84 _{0.13} ^{b, B} |
| GT-C | 4.05 _{0.15} ^{c, A} | 4.65 _{0.21} ^{c, B} | 4.82 _{0.21} ^{c, B} |
| GT-BD | 4.20 _{0.19} ^{c, A} | 4.56 _{0.25} ^{c, B} | 4.51 _{0.20} ^{d, B} |
| GT | 4.84 _{0.26} ^{d, A} | 6.04 _{0.27} ^{d, C} | 5.60 _{0.27} ^{e, B} |

514 PT = conventional progeny testing; GT-PT = genomic pre-selection of bulls for progeny testing, GT-C = genomic
 515 selection of sires for the insemination of cows; GT-BD = genomic selection of sires for the insemination of bull-dams;
 516 GT = genomic selection of sires for the insemination of cows and bull-dams. Subscript numbers indicate standard
 517 deviation across simulation replicates. Lower-case letters denote statistically significant differences between breeding
 518 programs and upper-case letters between sire selection and usage scenarios.

519 Table 2: Effective population size at causal loci by breeding program and by sire selection and their
 520 usage scenario.

| Breeding program | Sire selection and their usage | | |
|------------------|-----------------------------------|------------------------------------|----------------------------------|
| | 5 sires/year, use 5 years | 5 sires/year, use 1 year | 1 sire/year, use 5 years |
| PT | 172 ₄₈ ^{a, A} | 184 ₅₇ ^{a, A} | 96 ₂₀ ^{a, B} |
| GT-PT | 159 ₄₃ ^{a, A} | 146 ₄₀ ^{b, A} | 99 ₂₀ ^{a, B} |
| GT-C | 129 ₂₉ ^{b, A} | 124 ₃₂ ^{bc, A} | 64 ₁₁ ^{b, B} |
| GT-BD | 119 ₂₇ ^{b, A} | 113 ₂₄ ^{c, AB} | 93 ₂₄ ^{a, B} |
| GT | 90 ₁₄ ^{c, A} | 72 ₁₀ ^{d, A} | 38 ₆ ^{b, B} |

521 PT = conventional progeny testing; GT-PT = genomic pre-selection of bulls for progeny testing, GT-C = genomic
 522 selection of sires for the insemination of cows; GT-BD = genomic selection of sires for the insemination of bull-dams;
 523 GT = genomic selection of sires for the insemination of cows and bull-dams. Subscript numbers indicate standard
 524 deviation across simulation replicates. Lower-case letters denote statistically significant differences between breeding
 525 programs and upper-case letters between sire selection and usage scenarios.

526 Table 3: Efficiency of converting genetic variation into gain by breeding program and by sire
 527 selection and their usage scenario.

| Sire selection and their usage | | | |
|--------------------------------|-----------------------------------|----------------------------------|-----------------------------------|
| Breeding program | 5 sires/year, use 5 years | 5 sires/year, use 1 year | 1 sire/year, use 5 years |
| PT | 77 ₁₇ ^{a, A} | 61 ₉ ^{a, B} | 48 ₉ ^{ab, C} |
| GT-PT | 100 ₂₁ ^{b, A} | 84 ₁₇ ^{b, B} | 64 ₁₀ ^{cd, C} |
| GT-C | 98 ₂₀ ^{b, A} | 81 ₁₂ ^{b, B} | 54 ₁₀ ^{ac, C} |
| GT-BD | 93 ₁₉ ^{bc, A} | 87 ₁₅ ^{b, A} | 72 ₁₅ ^{d, B} |
| GT | 84 ₁₁ ^{ac, A} | 75 ₁₁ ^{b, A} | 42 ₅ ^{b, B} |

528 PT = conventional progeny testing; GT-PT = genomic pre-selection of bulls for progeny testing, GT-C = genomic
 529 selection of sires for the insemination of cows; GT-BD = genomic selection of sires for the insemination of bull-dams;
 530 GT = genomic selection of sires for the insemination of cows and bull-dams. Subscript numbers indicate standard
 531 deviation across simulation replicates. Lower-case letters denote statistically significant differences between breeding
 532 programs and upper-case letters between sire selection and usage scenarios.

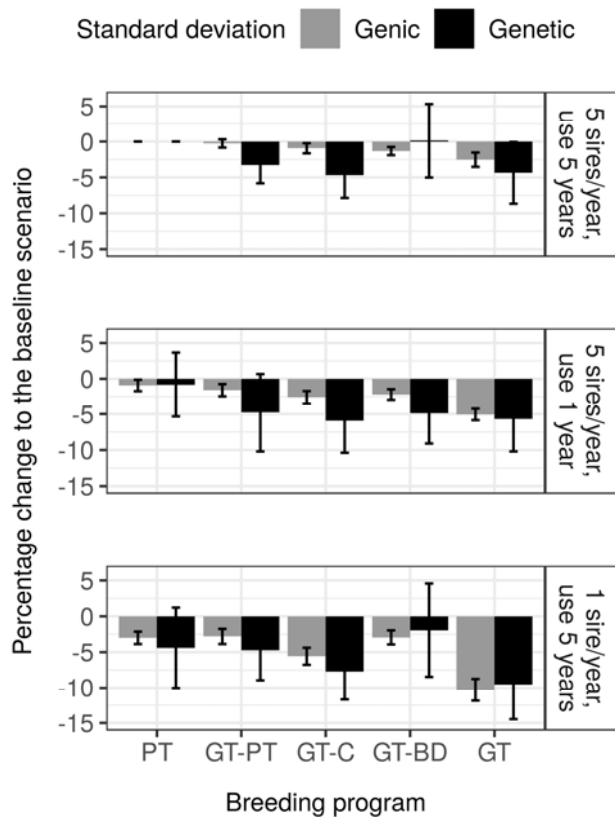
533 Table 4: Comparison of breeding programs that use truncation or optimum contribution selection.

| Breeding program | Genetic gain | Sire selection accuracy | No. sires | Years in use | Generation interval (sire-sire) | Generation interval (sire-dam) | Genic standard deviation | Rate of coancestry | Effective population size | Conversion efficiency |
|--------------------------------|-----------------------------------|-------------------------------------|-----------------------------------|----------------------------------|----------------------------------|----------------------------------|------------------------------------|--------------------------------------|---------------------------------|--------------------------------|
| Truncation selection | | | | | | | | | | |
| 5 sires/year, use 5 years, PT | 2.50 _{0.22} ^a | 0.88 _{0.10} ^a | 55 _{0.0} ^a | 2.7 _{0.0} ^a | 9.0 _{0.06} ^a | 7.0 _{0.05} ^a | 0.97 _{0.01} ^a | 0.003 _{0.001} ^a | 172 ₄₈ ^a | 77 ₁₇ ^a |
| 5 sires/year, use 5 years, GT | 4.84 _{0.26} ^b | 0.79 _{0.12} ^b | 56 _{0.1} ^a | 2.1 _{0.0} ^b | 4.2 _{0.05} ^b | 3.9 _{0.05} ^b | 0.94 _{0.01} ^b | 0.006 _{0.001} ^b | 90 ₁₄ ^{bc} | 84 ₁₁ ^a |
| 5 sires/year, use 1 year, GT | 6.04 _{0.27} ^c | 0.75 _{0.11} ^c | 36 _{0.0} ^b | 1.3 _{0.0} ^c | 2.3 _{0.05} ^c | 2.3 _{0.00} ^c | 0.92 _{0.01} ^{cd} | 0.007 _{0.001} ^c | 72 ₁₀ ^{bd} | 75 ₁₁ ^a |
| Optimum contribution selection | | | | | | | | | | |
| OCS _{45°} | 6.26 _{0.39} ^c | 0.77 _{0.02} ^{bc} | 9.6 _{0.6} ^c | 1.6 _{0.06} ^d | 2.8 _{0.07} ^d | 2.7 _{0.07} ^d | 0.91 _{0.01} ^c | 0.008 _{0.001} ^d | 61 ₁₀ ^d | 72 ₈ ^a |
| OCS _{50°} | 6.10 _{0.23} ^c | 0.79 _{0.02} ^{abc} | 14.3 _{0.9} ^c | 1.7 _{0.05} ^d | 2.9 _{0.07} ^e | 2.8 _{0.05} ^e | 0.93 _{0.01} ^d | 0.007 _{0.001} ^{bc} | 75 ₉ ^{bcd} | 87 ₁₁ ^a |
| OCS _{55°} | 5.27 _{0.28} ^d | 0.79 _{0.02} ^{abc} | 25.1 _{4.7} ^d | 2.1 _{0.17} ^b | 3.0 _{0.07} ^f | 2.9 _{0.06} ^f | 0.95 _{0.01} ^{be} | 0.005 _{0.001} ^e | 113 ₂₂ ^{ce} | 115 ₂₄ ^b |
| OCS _{60°} | 4.77 _{0.25} ^b | 0.81 _{0.02} ^{abc} | 49.0 _{8.9} ^e | 3.0 _{0.34} ^e | 3.3 _{0.10} ^g | 3.1 _{0.07} ^g | 0.96 _{0.01} ^{ae} | 0.004 _{0.001} ^{ae} | 144 ₂₄ ^e | 126 ₁₇ ^b |
| OCS _{75°} | 3.03 _{0.17} ^e | 0.82 _{0.01} ^{abc} | 153.0 _{9.1} ^f | 4.9 _{0.07} ^f | 4.2 _{0.08} ^b | 4.0 _{0.06} ^h | 0.98 _{0.00} ^f | 0.002 _{0.001} ^f | 276 ₄₃ ^f | 162 ₃₇ ^c |

534 PT = conventional progeny testing; GT = genomic selection of sires for the insemination of cows and bull-dams; OCS_{X°} = optimum contribution selection of sires for the
 535 insemination of cows and bull-dams with the target degrees of X°. Subscript numbers indicate standard deviation across simulation replicates. Lower-case letters denote statistically
 536 significant differences between breeding programs.

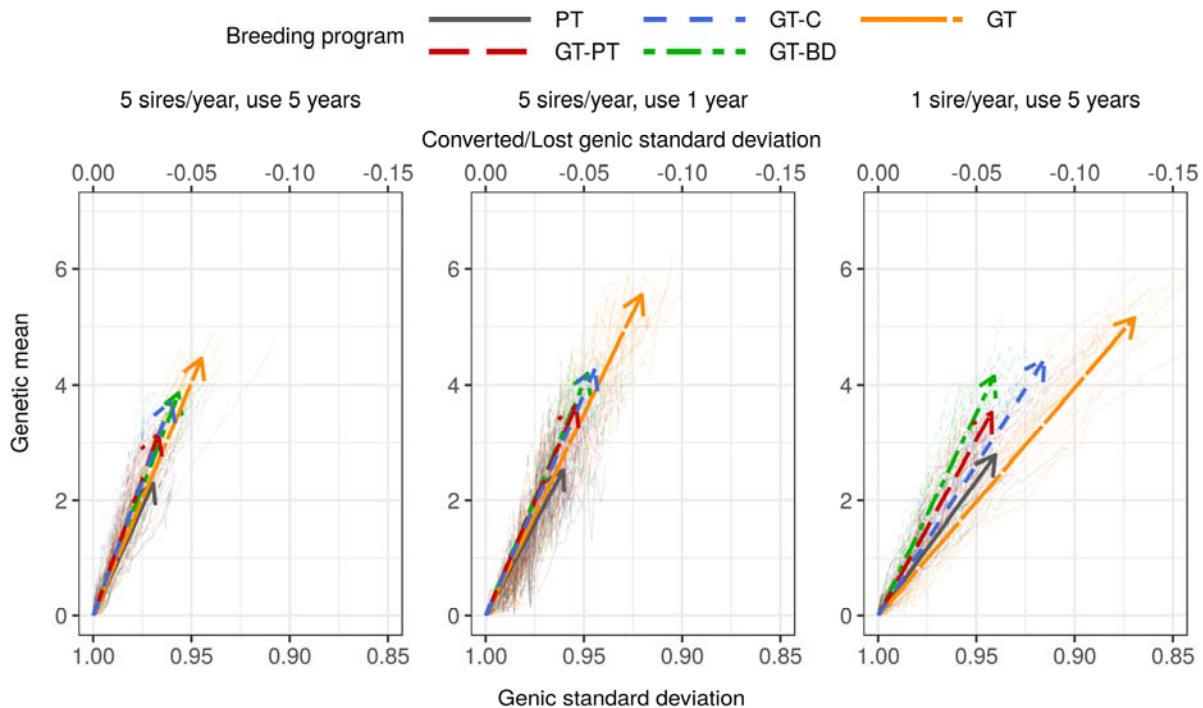
537

Obšteter et al., 1



538 Figure 1: Genic and genetic standard deviation by breeding program and by sire selection and their
539 usage scenario expressed as percentage change to the baseline that had in the final year genic
540 standard deviation of 0.97 and genetic standard deviation of 0.94. PT = conventional progeny testing; GT-
541 PT = genomic pre-selection of bulls for progeny testing, GT-C = genomic selection of sires for the insemination of
542 cows; GT-BD = genomic selection of sires for the insemination of bull-dams; GT = genomic selection of sires for the
543 insemination of cows and bull-dams.

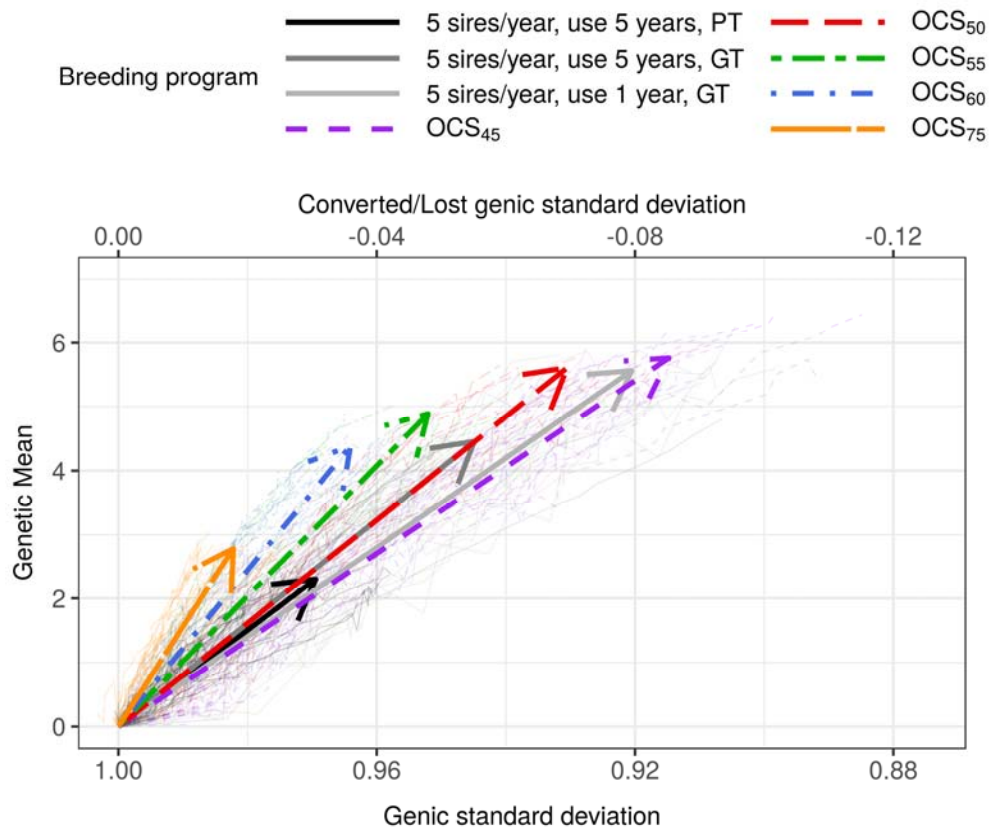
544 Obšteter et al., 2



545

546 Figure 2: Change of genetic mean and genic standard deviation over the 20 years of selection by
547 breeding program and by sire selection and their usage scenario. Thin lines represent individual
548 replicates, while thick lines represent average linear regression with arrows pointing in the direction
549 of change. PT = conventional progeny testing; GT-PT = genomic pre-selection of bulls for progeny testing, GT-C =
550 genomic selection of sires for the insemination of cows; GT-BD = genomic selection of sires for the insemination of
551 bull-dams; GT = genomic selection of sires for the insemination of cows and bull-dams.

552 Obšteter et al., 3



553

554 Figure 3: Change of genetic mean and genic standard deviation over the 20 years of selection for
555 fixed or optimized breeding programs. Thin lines represent individual replicates, while thick lines
556 represent average linear regression with arrows pointing in the direction of change. PT = conventional
557 progeny testing; GT = genomic selection of sires for the insemination of cows and bull-dams; OCS_{X°} = optimum
558 contribution selection of sires for the insemination of cows and bull-dams with the target degrees of X°.