Scalable method for groupwise comparisons and its application to chromatin accessibility

among primates

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Abstract

Changes in transcriptional regulation are thought to be a major contributor to the evolution of

phenotypic traits. We developed a new method to identify DNase I Hypersensitive (DHS) sites

with differential chromatin accessibility between species. Our method replaces threshold-based

multiple pairwise comparisons with a single quantitative test, which detects subtle differences

while scaling well to multiple taxa. We applied this method to DHS sites in fibroblast cells from

five primates (human, chimpanzee, gorilla, orangutan, and rhesus macaque). We identified

approximately 90,000 DHS sites, of which 59% are non differential between species, 27% are

differential and likely due to a single evolutionary change, and 14% are differential due to

multiple evolutionary changes. We found that using closely related species in our analyses

allows us to distinguish between accessibility changes that are specific to a single species and

those that have experienced multiple changes in chromatin accessibility during primate

evolution. The non differential sites are enriched for nucleotide conservation, while the species-

specific changes are enriched for positive selection. Differential DHS sites with decreased

chromatin accessibility relative to rhesus macaque occur more commonly in proximal regulatory

elements, while those with increased chromatin accessibility relative to rhesus macaque occur

more commonly in distal regulatory elements. Differential DHS sites overlapping proximal

regulatory elements display less cell-type specificity than those overlapping distal regulatory

elements. Taken together, these results identify several classes of chromatin accessibility, each

with distinct characteristics of selection, genomic location, and cell-type specificity.

Key words: cis-regulatory evolution, comparative functional genomics, positive selection,

chromatin accessibility, transcriptional regulation

Introduction

It has long been hypothesized that phenotypic differences between species are more often

due to genetic variation in non-coding regulatory regions than in protein-coding regions (King

and Wilson 1975; Wray 2007; Wittkopp and Kalay 2011). The development of diverse genome-

wide assays, combined with the publication of primate reference genomes, has allowed

identification of inter-species differences in gene expression (Cáceres et al. 2003; Gilad et al.

2006; Blekhman et al. 2008; Brawand et al. 2011), DNA methylation (Pai et al. 2011; Zeng et al.

2012; Hernando-Herraez et al. 2013), histone modifications (Zhou et al. 2014; Villar et al. 2015),

transcription factor binding motifs (Dermitzakis and Clark 2002; Odom et al. 2007; Schmidt et

al. 2010), chromatin accessibility (Shibata et al. 2012; Gallego Romero et al. 2018), and

alternative splicing (Blekhman et al. 2010; Barbosa-Morais et al. 2012). These differences in

molecular function among primate species can provide valuable insights into species-specific

trait differences, including disease risk (Prabhakar et al. 2008; Boyd et al. 2015; Prescott et al.

2015).

Previous approaches to analyzing multi-species data have used multiple pairwise

comparisons to detect differences between species (Robinson et al. 2010; Love et al. 2014).

Applying these methods to comparisons between three or more groups becomes cumbersome

because it requires multiple pairwise comparisons followed by an overlap step that intersects the

results from those comparisons. The number of required pairwise comparisons increases

substantially as the number of groups is increased (e.g., 10 comparisons for 5 species).

Calculating p-values for multiple species comparisons is difficult because species are not

completely independent and indeed differ in their degree of dependence due to phylogenetic

relationships.

As an alternative to performing multiple pairwise comparisons, we developed a negative

binomial generalized linear model that jointly models data across multiple species and replicates,

and can be applied to any count-based data such as DNase-seq, RNA-seq, ChIP-seq, and ATAC-

seq. We applied this method to chromatin accessibility DNase-seq data from cultured skin

fibroblasts obtained from human, three recently diverged great apes (chimpanzee, gorilla, and

orangutan), and rhesus macaque for use as an outgroup. We identified a total of 89,744 DNase I

Hypersensitive (DHS) sites that are directly comparable between species, of which 36,666 (41%)

displayed a statistically significant difference between species. Using effect estimates from the

negative binomial generalized linear model, we inferred the direction of the change relative to

rhesus macaque and the possible branch or branches on which the changes occured.

Materials and Methods

DNase-seq Experiments and Sequencing

Fibroblast cell lines from 15 individuals comprising 3 biological replicates from each of

five primate species (human, chimpanzee, gorilla, orangutan, and rhesus macaque) were obtained

from Coriell (**Supplementary Table 1**). It is estimated that human and chimpanzee diverged 7

million years ago, gorilla diverged from the human-chimpanzee ancestor 10 million years ago,

orangutan diverged from the human-chimpanzee-gorilla ancestor 18 million years ago, and

rhesus macaque diverged from the human-chimpanzee-gorilla-orangutan ancestor 30 million

years ago (Schrago and Voloch 2013). DNase-seq experiments were performed as previously

described (Shibata et al. 2012). DNase-seq libraries were generated from 50 million cells and

sequenced on Illumina instruments (**Supplementary Table 1)**.

DNase-seq Read Mapping and Conversion to Human Genome

Due to the use of MmeI to generate DNase-seq libraries (Boyle et al. 2008), genomic

DNA fragments are only 20 bases long. Therefore, sequencing reads were trimmed to 20 bases

using a custom perl script. Reads for each species were mapped to its native genome: hg19 for

human, panTro4 for chimpanzee, gorGor3 for gorilla, ponAbe2 for orangutan, and rheMac3 for

rhesus macaque (Lander et al. 2001; Chimpanzee Sequencing and Analysis Consortium 2005;

Locke et al. 2011; Yan et al. 2011; Scally et al. 2012). Reads were mapped using Bowtie version

0.12.9 (Langmead et al. 2009) (parameters: -trim5 0 --trim3 0 -m 1 -l 20) as part of a custom

two-step pipeline. In the first step ("tier 1"), reads were required to match to a unique location

with no mismatches (parameter: -n 0). In the second step ("tier 2"), unmapped reads from step

one were re-mapped with a relaxed mismatch parameter of one mismatch (parameter: -n 1).

Reads that mapped to multiple locations or had more than one mismatch were discarded.

Samtools version 0.1.19-44428cd (Li et al. 2009) was used to convert the sam files from each

step to bam files, merge them into one file, and remove duplicate reads (defined as having the

same chromosomal coordinates). Bedtools v2.17.0 (Quinlan and Hall 2010) was used to convert

the bam files to bed files. Details on the number of reads in the input files and at each step are

included in **Supplementary Table 2**.

Reads from the non-human samples were converted from their native genomic

coordinates to hg19 coordinates using a three-step process that removed reads without a one-to-

one relationship between the genomes. In each step, read coordinates were converted from one

genome to the other using the UCSC liftOver software (Hinrichs et al. 2006) with a minMatch

parameter of 0.8, which requires that 80% of the read maps to the new genome. Note that this

parameter filters only on genomic coverage, not sequence identity. In the first step, read

coordinates were converted from their native genome to hg19. Read coordinates that successfully

lifted to hg19 were then lifted back to the native genome. Read coordinates that did not lift back

to the same coordinates on the native genome were removed. Reads that did lift back to the same

coordinates were lifted back to hg19 for further processing. An additional filtering step was

added to ensure the reads did not map to a duplicated region. In that step, overlapping reads on

the native genome were merged into a region, and that region was lifted to hg19. Regions that

failed to lift uniquely to hg19 were flagged and reads that overlapped them were removed.

Because some of the samples were from males and some were from females (**Supplementary**

Table 1), we removed reads that mapped or lifted over to the human X or Y chromosomes to

eliminate any sex-specific bias. Details on the number of reads lifted over and remaining after

removal of sex chromosomes are included in **Supplementary Table 2**. Phylogenetic trees were

drawn with ggtree (Yu et al., 2016).

DHS Site Identification and Filters

To avoid bias due to large differences in depth of library sequencing, 20 million reads

were randomly selected from samples with library sizes greater than 20 million reads. To

determine the appropriate subset size, we generated subsets of 10, 15, 20, and 25 million reads

and compared the average number of per-species DHS sites in each subset (**Supplementary**

Figure 1).

Peak calling was performed on each sample separately using the MACS2 callpeak

command with an FDR cutoff of 5% (Zhang et al. 2008) (version 2.1.0.20150420; parameters: --

nomodel --extsize 20 --qvalue .05). For each species, DHS sites were identified by taking the

union set of peaks that were found in at least 2 out of 3 biological replicates. We removed DHS

sites that overlapped the ENCODE blacklist

(http://hgdownload.cse.ucsc.edu/goldenPath/hg19/encodeDCC/wgEncodeMapability/wgEncode

DacMapabilityConsensusExcludable.bed.gz) (Rosenbloom et al. 2013). To generate a master set

of DHS sites to use for cross-species comparisons, we first took the union set of DHS sites

identified in each species, then applied two filters. In the first filter, we removed DHS sites

without at least 95% genomic coverage between the start and stop coordinates for each of the

species. Genomic coverage was determined using the Multiz Alignment MAF file

(http://hgdownload.cse.ucsc.edu/goldenPath/hg19/multiz100way/maf/) from the UCSC Genome

Browser Multiz Alignment of 100 Vertebrates (Blanchette et al. 2004), the Galaxy MAF

Coverage Stats tool at usegalaxy.org (Afgan et al. 2018), and custom perl scripts. Next, we

assigned read counts to each DHS site using bedtools v2.17.0 (Quinlan and Hall 2010). In the

second filter, we removed DHS sites without DNase-seq sequence reads in at least two biological

replicates from each species because they may be indicative of regions that cannot be uniquely

aligned. In other words, we expect at least some level of background DNase cutting across the

genome. See **Supplementary Table 3** for DHS site counts before and after each filtering step.

Principal Components Analysis

We performed a principal components analysis on the read counts from the 15 samples

(Supplementary Figure 4). We first normalized the counts by library size then ran the R

prcomp function with the center and scale parameters set to true. We also performed a principal

components analysis on the read counts from the 15 samples and 3 additional chimpanzee

samples from Pizzollo et al. 2018 **(Supplementary Figure 4)**. See **Supplementary Table 1** for

details on the samples.

Differential Site Identification and Classification

The read counts for each DHS site were used as input to a custom R script that identified and

classified differential DHS sites. To address the over-dispersion problem inherent in count-based

sequencing data, we used the R package DSS (Wu et al. 2013) to calculate a dispersion

parameter for each DHS site, as well as a normalization offset (based on total library size) for

each sample. For each DHS site, the read counts, dispersion parameter, and normalization offset

were fit using a negative binomial generalized linear model with a log link function. Specifically,

we fit two models: a species informed model and a null model in which species was not

predictive of normalized counts. The species informed model modelled the expected counts by

$log\ (\lambda_j) = \alpha_j + x_j{}^T\beta$, where λ represents expected counts and j indexes the sample. The design

vector x_j indicates to which species the j$^{\text{th}}$ sample belongs and $x_j{}^T$ denotes its transpose. This

vector has 4 elements comprised of indicator functions of whether the sample is human,

chimpanzee, gorilla, or orangutan. Because accessibility changes are relative to rhesus macaque,

it is used as the intercept in the models and does not have an indicator function. Specifically,

$x_j{}^T = (1\ 0\ 0\ 0)$ if the j$^{\text{th}}$ sample is human; $x_j{}^T = (0\ 1\ 0\ 0)$ if the j$^{\text{th}}$ sample is chimpanzee;

$x_j{}^T = (0\ 0\ 1\ 0)$ if the j$^{\text{th}}$ sample is gorilla; and $x_j{}^T = (0\ 0\ 0\ 1)$ if the j$^{\text{th}}$ sample is orangutan. The

vector $\beta = (\beta_h, \beta_c, \beta_g, \beta_o)^T$ parameterizes the change in expected counts between species and

α_j is a normalization offset for sample j. The null model assumes the vector β is zero and models

the expected counts by $log\ (\lambda_j) = \alpha_j$. These models were fit using the R package glm using

negative.binomial as the family. The DSS normalization offset value was used for the offset

parameter. The inverse of the DSS dispersion parameter value was used for the theta parameter.

The difference in deviances between these two models was used to form a likelihood ratio test of

whether the sites were differential. A Benjamini-Hochberg correction was performed using the R

function p.adjust. DHS sites with a corrected p-value of less than .01 were classified as

differential.

To determine which species (or combination of species) were different from rhesus

macaque, 15 contrasts were constructed using the β values estimated in the regression model

detailed above. Five contrasts were used to identify changes in a single species, six for changes

in two species, and four for changes in three species (**Supplementary Table 4**). Note that the

contrast for changes in rhesus macaque will also identify changes that occurred in the human-

chimpanzee-orangutan-gorilla internal branch. A value for each contrast c using constraint

matrix C and variance-covariance matrix $Var(C\beta)$ was calculated as

$c = [C\beta]^T [Var(C\beta)]^{-1} C\beta$, where $[Var(C\beta)]^{-1}$ denotes the matrix inverse of $Var(C\beta)$. A p-

value was calculated for each contrast using a chi-squared test and the contrast with the lowest p-

value was chosen. For 30 DHS sites, none of the p-values were less than .01, so the change was

marked as "other". The sign of the β was used to determine whether each change was an increase

(referred to as "increased accessibility") or decrease (referred to as "decreased accessibility")

relative to rhesus macaque.

Note that since we classified only the sites that were shown to be differential and we

controlled the type one error rate in the differential site identification step, there is no need to

further adjust our analyses for the multiple contrasts being considered here. In the statistical

literature this is referred to as a "gateway" procedure as the differential site identification is a

gatekeeper for site classification.

We have included supplementary files that contain the input data for the R script

(**glm_analysis.input_file.txt**), the results from all of the analyses discussed in this paper

(**glm_output_and_analsyis_results.txt**), and the field information for the input and output files

(**input_and_output_file_field_information.xlsx**). Raw fastq files and bed files containing hg19

coordinates for the reads are available under GEO accession GSE129034. The reads, DHS sites,

and differential analysis classifications are available in a UCSC Genome Browser session at

http://genome.ucsc.edu/s/ledsall/2019primate. All scripts for the data processing and analyses

described above are available at http://github.com/ledsall/primate.

Testing for Positive Selection and Determining Vertebrate Conservation

We performed selection analysis on both the differential and non differential DHS sites.

We tested for selection on the species branches for human, chimpanzee, gorilla, and orangutan,

and on the internal branches for human-chimpanzee and human-chimpanzee-gorilla.

As the stochasticity of the evolutionary process may be elevated in short alignments, we

expanded each DHS site that was smaller than 300 bases up to 300 bases, while maintaining the

size of any DHS site longer than 300 bases. We removed any sites that couldn't be expanded due

to gaps in the non-human genomes.

To investigate the extent of positive selection among the DHS sites, we used a branch-

specific method we first developed in 2007 (Haygood et al. 2007) and recently improved (Berrio

et al. under review). Briefly, the method uses a likelihood ratio test based on the maximum

likelihood estimates obtained from HyPhy (Pond et al. 2005). The branch of interest (e.g. human

species branch) is used as the foreground and the rest of the tree is used as the background. The

assumption for the background is the same for both the null and alternative models; specifically,

neutral evolution and negative (purifying) selection are permitted, but positive selection is not. In

the null model, the assumption for the foreground is the same as the one for the background. In

the alternative model, all three models of evolution are permitted (neutral evolution, negative

selection, and positive selection) in the foreground. This method is highly sensitive and specific

in that it can differentiate between positive selection and relaxation of constraint.

The method requires a 3kb reference alignment for each species that is used as a

putatively neutral proxy for computing substitution rates. To generate this alignment, we first

identified a set of functional regions on the human genome using annotations from the ENCODE

project at UCSC (http://genome.ucsc.edu/encode/downloads.html) (ENCODE Project

Consortium 2012) and annotations from the HoneyBadger2-intersect dataset from the ENCODE

and Roadmap Epigenomics projects

(https://personal.broadinstitute.org/meuleman/reg2map/HoneyBadger2-intersect_release/)

(Roadmap Epigenomics Consortium et al. 2015). We used the set of 56,893 putative promoter

regions, 1,598,323 putative enhancer regions, and 31,255 putative dyadic regions. We then

masked the genomes using those functional regions, along with 5' and 3' UTRs, coding and non-

coding RNAs, CpG repeats, microsatellite repeats, and simple repeats. Next, we extracted

windows of 300 bases and excluded those with substitution rates that are too high or slow

relative to the entire tree. Finally, we concatenated the set of these windows until we reached a

length of 3kb (Berrio et al. under review).

We used the PHAST library msa_split (Hubisz et al. 2011) to extract query regions from

the UCSC Genome Browser Multiz Alignment of 100 Vertebrates

(http://hgdownload.cse.ucsc.edu/goldenPath/hg19/multiz100way/maf/) (Blanchette et al. 2004)

for the human, chimpanzee, gorilla, orangutan, and rhesus macaque genomes.

For each DHS site (called a query site), we used HyPhy (Pond et al. 2005) to fit the null and

alternative models and generate maximum likelihood values. We used a custom R script to

compute the likelihood ratio, which was used as a test statistic for a chi-squared test with one

degree of freedom to calculate a p-value. We classified a DHS site as under positive selection if

the p-value was less than .05. We were unable to successfully run HyPhy on 12 sites due to

unknown reasons and removed these regions from analysis.

We found that the substitution rates of the orangutan branch and the internal branches for

human-chimpanzee and human-chimpanzee-gorilla were extremely variable and caused an

overestimation of selection in these branches and in the subset of non functional data. Therefore,

we excluded these branches from the subsequent analyses and focused on the human,

chimpanzee, and gorilla branches.

We then tested for significant enrichments of positive selection in different classes of

DHS sites. The classes we investigated are 1) human-specific accessibility increases and

decreases; 2) chimpanzee-specific accessibility increases and decreases; 3) gorilla-specific

accessibility increases and decreases; and 4) non differential sites. We performed Fisher's exact

tests using a test statistic of the number of DHS sites classified as under positive selection. For

the human-specific accessibility changes, we performed two tests; 1) the human species branch

as the foreground compared to the chimpanzee species branch as the foreground; and 2) the

human species branch as the foreground compared to the gorilla species branch as the

foreground. For the chimpanzee-specific accessibility changes, we performed two tests; 1) the

chimpanzee species branch as the foreground compared to the human species branch as the

foreground; and 2) the chimpanzee species branch as the foreground compared to the gorilla

species branch as the foreground. For the gorilla-specific accessibility changes, we performed

two tests; 1) the gorilla species branch as the foreground compared to the human species branch

as the foreground; and 2) the gorilla species branch as the foreground compared to the

chimpanzee species branch as the foreground. For the non differential sites, we performed three

tests; 1) the human species branch as the foreground compared to the chimpanzee species branch

as the foreground; 2) the human species branch as the foreground compared to the gorilla species

branch as the foreground; and 3) the chimpanzee species branch as the foreground compared to

the gorilla species branch as the foreground. We used a Bonferroni correction to adjust for the

multiple tests performed.

To visualize the strength of selection, we computed the statistic ζ(zeta), representing the

ratio of evolution, by calculating the ratio of the substitution rate in each query compared to the

reference alignment; we computed ζ for the human, chimpanzee, and gorilla species branches.

This parameter is homologous to ω (omega), the ratio of dN/dS, where a value of $\omega < 1$

indicates constraint or negative selection; a value of $\omega = 1$ indicates neutrality; and a value of

$\omega > 1$ indicates positive selection.

We then tested whether the distributions of ζ differed between classes of differential DHS

sites. The classes we investigated are 1) human-specific increased accessibility; 2) chimpanzee-

specific increased accessibility; 3) gorilla-specific increased accessibility; 4) orangutan-specific

increased accessibility; 5) human-chimpanzee increased accessibility; 6) human-chimpanzee-

gorilla increased accessibility; 7) human-specific decreased accessibility; 8) chimpanzee-specific

decreased accessibility; 9) gorilla-specific decreased accessibility; 10) orangutan-specific

decreased accessibility; 11) human-chimpanzee decreased accessibility; and 12) human-

chimpanzee-gorilla decreased accessibility. We performed Wilcoxon tests on human-specific

increased accessibility against the other classes of DHS sites with increased accessibility (classes

2-6) and used a Bonferroni correction to adjust for the multiple tests. Similarly, we performed

Wilcoxon tests on human-specific decreased accessibility against the other classes of DHS sites

with decreased accessibility (classes 8-12) and used a Bonferroni correction to adjust for the

multiple tests. Finally, we performed a Wilcoxon test on the non differential sites against the non

functional sites defined above.

To determine the amount of vertebrate conservation, we computed the median value of

PhastCons scores for each DHS site using bedops (Neph et al. 2012), the UCSC 100-way

PhastCons table (http://hgdownload.cse.ucsc.edu/goldenpath/hg19/phastCons100way) (Siepel et

al. 2005; Pollard et al. 2010), and custom scripts. The PhastCons score represents the probability

of a particular base being conserved. The values range from 0 to 1, with higher values

representing an increased probability of conservation. We classified a DHS site as constrained if

the median PhastCons score was above 0.9.

We then investigated whether the amount of conservation differed between differential

and non differential DHS sites. We used the percentage of constrained DHS sites as our test

statistic and performed a Fisher's exact test on non differential sites compared to three classes of

differential DHS sites; specifically, 1) human-specific accessibility changes; 2) chimpanzee-

specific accessibility changes; and 3) gorilla-specific accessibility changes. We used a

Bonferroni correction to adjust for the multiple tests.

Intersection with Human Putative Regulatory Annotations

We characterized each DHS site as a proximal element, distal element, or unannotated

region using the HoneyBadger2-intersect dataset from the ENCODE and Roadmap Epigenomics

projects (https://personal.broadinstitute.org/meuleman/reg2map/HoneyBadger2-

intersect_release/) (Roadmap Epigenomics Consortium et al. 2015). We used the putative

promoter and enhancer regions as above, but did not use the putative dyadic regions. We used

bedtools2 (Quinlan and Hall 2010) to identify DHS sites that overlapped the annotated promoters

(which we characterized as "proximal elements") and enhancers (which we characterized as

"distal elements"). DHS sites that didn't overlap promoters or enhancers were characterized as

unannotated regions.

Determining Cell-type Specificity

We characterized the cell-type specificity of each DHS site by intersecting it with DHS

sites from 125 human cell types and tissues (Thurman et al. 2012). We used bedtools2 (Quinlan

and Hall 2010) to find the overlap with the wgEncodeAwgDnaseMasterSites dataset from the

UCSC Genome Browser

(http://hgdownload.cse.ucsc.edu/goldenpath/hg19/encodeDCC/wgEncodeAwgDnaseMasterSites

/wgEncodeAwgDnaseMasterSites.bed.gz) . We assigned a score to each DHS site representing

its cell-type specificity. The score was calculated as $1 - \frac{N}{126}$, where N represents the number of

cell-types and tissues in which the DHS site is present (including the fibroblast cell line from this

study). The score ranges from 0 for a DHS site present in all tissues and cell types to $\frac{125}{126}$ (which

is approximately 0.99) for a DHS site present in only our dataset.

We then asked whether the distribution of cell-type specificity scores varied between

different classes of DHS sites. We subset the DHS sites into those overlapping proximal

elements and those overlapping distal elements. For each subset, we investigated the following

classes: 1) human-specific increased accessibility; 2) chimpanzee-specific increased

accessibility; 3) gorilla-specific increased accessibility; 4) orangutan-specific increased

accessibility; 5) human-chimpanzee increased accessibility; 6) human-chimpanzee-gorilla

increased accessibility; 7) human-specific decreased accessibility; 8) chimpanzee-specific

decreased accessibility; 9) gorilla-specific decreased accessibility; 10) orangutan-specific

decreased accessibility; 11) human-chimpanzee decreased accessibility; 12) human-chimpanzee-

gorilla decreased accessibility; and 13) non differential sites.

Results

Method Development to Identify and Classify Differential DNase I Hypersensitive Sites Across

Multiple Primate Species

We developed a negative binomial generalized linear model based method to allow us to

quantitatively compare DNase I hypersensitive (DHS) sites across five primate species with one

statistical test, rather than performing multiple pairwise comparisons. This greatly simplifies the

analysis, scales well for additional replicates and species, and has straightforward p-value

calculations. The output of this test indicates for each DHS site whether or not at least one

species displays a statistically significant difference in read counts. In addition, the output from

the model includes the vector $\beta = (\beta_h, \beta_c, \beta_g, \beta_o)^T$, representing the changes in expected counts

compared to rhesus macaque for human, chimpanzee, gorilla, and orangutan respectively. We

used both the value and the sign (positive vs. negative) of the β values to classify the type of

difference between a given species and the outgroup, rhesus macaque (see Materials and

Methods). DHS sites that are not differential have values near zero for all of the β values **(Figure**

1A). DHS sites that have increased accessibility in a particular species have a positive β for that

species with the other species having β values near zero (**Figure 1B** for human-specific

increased accessibility; **Supplementary Figure 2** for other species). DHS sites that have

decreased accessibility in a particular species have a negative β value for that species with the

other species having β values near zero (**Figure 1C** for human-specific decreased accessibility;

Supplementary Figure 2 for other species). We also detect regions that have altered β values in

more than one species (**Figure 1D**, **Supplementary Figures 2 and 3**).

Using this approach, we identified 89,744 total DHS sites that can be compared across all

5 species at 1:1:1:1:1 orthologous genomic regions (Materials and Methods). As a first step in

analyzing these data, we carried out a principal components analysis and found that the first

principal component separated the single old world monkey (Rhesus macaque) from the four

great apes, while the second principal component recapitulated the phylogeny of the great apes

(**Supplementary Figure 4**). Because we are drawing on the original data from Shibata et al.

2012 for three species and have generated new data for two species (**Supplementary Table 1**),

we also investigated whether batch effects would overwhelm the species signal by comparing

principal components analyses with and without three additional chimpanzee samples generated

more recently from Pizzollo et al. 2018 (**Supplementary Table 1**). As shown in **Supplementary**

Figure 4, the Pizzollo et al. chimpanzee samples cluster with the original Shibata et al.

chimpanzee samples across the first four principal components (cumulative proportion of

variance of 0.53), suggesting that biological signal is retained even when samples are prepared

and sequenced years apart.

Of the 89,744 total DHS sites, 53,078 (59%) are not statistically significantly different

between species, 23,926 (27%) display a difference that likely resulted from a single

evolutionary event, and 12,710 (14%) that display a difference due to multiple evolutionary

events **(Figure 1E, Table 1)**. Of the 23,926 DHS sites with changes that are likely due to a

single event, about half (11,879) appear to have changed more recently and are detected only in a

single species (human, chimpanzee, gorilla, or orangutan) **(Figure 1F, Table 1)**. About one

quarter of single event changes (5,738) appear to have occured in either the human-chimpanzee

internal branch or the human-chimpanzee-gorilla internal branch **(Figure 1F, Table 1)**. The

remaining single events (6,309) occurred either recently in rhesus macaque or longer ago in the

internal human-chimpanzee-gorilla-orangutan internal branch **(Figure 1F, Table 1)**. Because we

are using rhesus macaque as the outgroup, we are unable to differentiate between changes in the

rhesus macaque species branch and changes in the human-chimpanzee-gorilla-orangutan internal

branch (see Discussion). Consistent with our earlier study (Shibata et al. 2012), as well as studies

by other groups (Reilly et al. 2015; Villar et al. 2015; Emera et al. 2016), the majority of the

changes are increased accessibility rather than decreased accessibility (see Discussion). For

changes on the species branches, there are approximately 10x the number of DHS sites with

increased accessibility as DHS sites with decreased accessibility; 12:1 for human, 8:1 for

chimpanzee, 11:1 for gorilla, 6:1 for orangutan. For changes on the internal branches, the ratio of

DHS sites with increased accessibility to DHS sites with decreased accessibility is much less; 2:1

for the human-chimpanzee branch, and nearly equal for the human-chimpanzee-gorilla branch

(Figure 1G, Table 1).

Changes in Chromatin Accessibility Detected in a Single Species

Using the methods described above, we identified 10,574 DHS sites with increased

accessibility specific to a single species; 2,539 in human, 1,539 in chimpanzee, 2,737 in gorilla,

and 3,759 in orangutan **(Figure 2A, Table 1)**. We identified 1,305 DHS sites with decreased

accessibility specific to a single species; 209 in human, 196 in chimpanzee, 254 in gorilla, and

646 in orangutan **(Figure 2B, Table 1)**. Heatmap overviews **(Figure 2A, 2B)** of each class of

increased accessibility and decreased accessibility show that these differences are not binary, but

instead span the continuum from extremely large differences to those that represent more modest

changes. Representative screenshots of individual genomic loci are provided **(Figure 2A, 2B)**.

A complete list of coordinates for all DHS sites, β values, and p-values that allow regions to be

filtered at different cutoff stringencies are included in **Supplementary File**

glm_output_and_analsyis_results.txt.

Even though we can't classify rhesus macaque-specific changes, we can identify sites

where rhesus macaque is different from the other four species. We identified 4,992 sites that

have increased accessibility in rhesus macaque relative to human, chimpanzee, gorilla, and

orangutan **(Figure 2A, Table 1)**. We identified 1,317 sites that have decreased accessibility in

rhesus macaque compared to human, chimpanzee, gorilla, and orangutan **(Figure 2B, Table 1)**.

Changes in Chromatin Accessibility that Likely Occurred on Internal Branches

Our method allows us to identify ancient changes in chromatin accessibility that likely

occurred as a single change on internal branches. Using rhesus macaque as an outgroup, we can

classify whether these internal changes are accessibility increases or decreases. We identified

1,735 DHS sites with increased accessibility and 814 DHS sites with decreased accessibility that

likely occurred during the common lineage of human and chimpanzee **(Figure 3, Table 1)**. We

identified 1,736 DHS sites with increased accessibility and 1,453 DHS sites with decreased

accessibility that likely occurred before the split between human, chimpanzee, and gorilla

(Figure 3, Table 1).

Multiple Changes in Chromatin Accessibility

In addition to detecting likely single changes in chromatin accessibility on either species

(Figure 2) or internal branches **(Figure 3)**, we also identified changes in chromatin accessibility

that appear to have occurred multiple times, resulting in different combinations of chromatin

accessibility patterns between species. There are many possible ways these differences could

have happened and our method cannot determine if these changes resulted from multiple

increases in accessibility, multiple decreases in accessibility, or a combination of increases and

decreases (see Discussion).

We identified 6,339 DHS sites where two species display increased accessibility relative

to rhesus macaque; 997 in human and gorilla; 997 in human and orangutan; 1,568 in chimpanzee

and gorilla; 657 in chimpanzee and orangutan; and 2,120 in gorilla and orangutan **(Figure 4A,**

Table 1). We identified 1,801 DHS sites where two species display decreased accessibility

relative to rhesus macaque; 221 in human and gorilla; 424 in human and orangutan; 387 in

chimpanzee and gorilla; 252 in chimpanzee and orangutan; and 517 in gorilla and orangutan

(Figure 4B, Table 1).

There were 2,424 DHS sites where three species displayed increased chromatin

accessibility relative to rhesus macaque; 794 in human, chimpanzee, and orangutan; 691 in

human, gorilla, and orangutan; 939 in chimpanzee, gorilla, and orangutan **(Figure 4C, Table 1)**.

There were 2,146 sites where three species displayed decreased accessibility relative to rhesus

macaque; 906 in human, chimpanzee, and orangutan; 554 in human, gorilla, and orangutan; 686

in chimpanzee, gorilla, and orangutan **(Figure 4D, Table 1)**.

Comparison to Previous Study with Fewer Species

To test our new method for identifying differences in chromatin accessibility across five

species, we compared our results with those from our previous study that used individual

pairwise edgeR (Robinson et al. 2010) comparisons for human, chimpanzee, and rhesus macaque

(Shibata et al. 2012). We used the same raw fastq files for the human, chimpanzee, and rhesus

macaque samples. Due to updates in the analysis pipeline **(Supplementary Table 5)**, not all of

the DHS sites that were previously characterized were also identified as DHS sites in this study

(Supplementary Table 5).

Of the DHS sites that were previously characterized by Shibata et al. as being human-

specific increased accessibility, human-specific decreased accessibility, chimpanzee-specific

increased accessibility, or chimpanzee-specific decreased accessibility, we find that 88-98% of

our more recent set of DHS sites that overlap them displayed a consistent call **(Table 2)**. The

additional gorilla and orangutan DNase-seq data in this study allows us to fill in missing branch

data and gauge the accuracy of our previous classification of human-specific or chimpanzee-

specific changes. For 342 DHS sites that overlap sites previously characterized by Shibata et al.

as being human-specific increased accessibility, 245 (72%) are still characterized as human-

specific increased accessibility after including gorilla and orangutan, while 91 (27%) are now

characterized as increased accessibility in human and at least one other species **(Table 2)**. For

234 DHS sites that overlap previously called chimpanzee-specific increased accessibility, 105

(45%) are still characterized as chimpanzee-specific increased accessibility after including

gorilla and orangutan, while 114 (49%) are now characterized as increased accessibility in

chimpanzee and at least one other species **(Table 2)**.

A similar trend was detected for previously identified human-specific or chimpanzee-

specific decreased accessibility. For 146 DHS sites that overlap sites previously characterized as

human-specific decreased accessibility, 21 (14%) are still characterized as human-specific

decreased accessibility even after including orangutan and gorilla, while 109 (75%) are now

characterized as decreased accessibility in human and at least one other species **(Table 2)**. For 98

DHS sites that overlap sites previously called chimpanzee-specific decreased accessibility, 17

(17%) are still characterized as chimpanzee-specific decreased accessibility even after including

orangutan and gorilla, while 69 (70%) are now characterized as decreased accessibility in

chimpanzee and at least one other species **(Table 2)**.

For 1,154 DHS sites that overlap sites Shibata et al. (2012) characterized as common

between human, chimpanzee, and rhesus macaque, 95% (1,100) displayed a non-differential call

among these same species using our pipeline **(Table 2)**. After adding gorilla and orangutan, 90%

(1,043) of the DHS sites are still characterized as non-differential, while 5% (57) displayed

changes in accessibility on branches not considered by Shibata et al., namely gorilla, orangutan,

or both **(Table 2)**.

Together, this indicates that adding chromatin accessibility data from additional primate

species allows us to identify a substantial subset of DHS sites that have experienced changes in

chromatin accessibility across multiple species during evolution.

DHS Sites with Decreased Accessibility are Enriched for Proximal Elements and DHS Sites

with Increased Accessibility are Enriched for Distal Elements

After identifying and classifying DHS sites, we next determined their location in the

human genome relative to previously annotated proximal and distal elements. We used the

HoneyBadger2 annotations (see Materials and Methods), which are predicted promoters or

enhancers based on histone marks identified in human cells and tissues as part of the Roadmap

Epigenomics project (Roadmap Epigenomics Consortium et al. 2015). We overlapped these

annotations to characterize each DHS site identified in this study as a proximal element, distal

element, or unannotated region.

For DHS sites that are not differential between primate species, 22% (11,850) of these

regions overlap with proximal elements, 57% (30,371) overlap with distal elements, and the

remaining 20% (10,857) are unannotated **(Figure 5A, Supplementary Table 6)**. All DHS sites

with increased accessibility relative to rhesus macaque display a substantially depleted amount of

proximal element overlap compared to the non differential DHS sites (human: 2%; chimpanzee:

4%; gorilla: 9%; orangutan: 10%; human-chimpanzee: 3%; human-chimpanzee-gorilla: 5%)

(Figure 5A, Supplementary Table 6). Conversely, DHS sites with decreased accessibility

relative to rhesus macaque overlap proximal elements to a similar degree as non differential DHS

sites (human: 18%; chimpanzee: 21%; gorilla: 11%; orangutan: 20%; human-chimpanzee: 34%;

human-chimpanzee-gorilla: 10%) **(Figure 5A, Supplementary Table 6)**. These results indicate

that decreased accessibility changes are more likely to be associated with proximal elements,

while increased accessibility changes are more likely to be associated with distal elements. In

every category of accessibility changes, there are substantially more distal than proximal

elements, which is consistent with other studies (Schmidt et al. 2010; Villar et al. 2015).

We note that all of these proximal and distal annotations are from human tissues, which

allows us to make specific inferences about comparisons only to human. There is not yet a

similar Roadmap effort for non-human primate species. Categories of accessibility increases that

include human (human-specific, human-chimpanzee, and human-chimpanzee-gorilla) have the

lowest amount of overlap with unannotated regions of the genome. DHS sites with increased

accessibility specific to chimpanzee, gorilla, or orangutan all have much higher overlaps with

unannotated regions, with orangutan-specific increased accessibility showing the highest degree

of overlap with unannotated regions **(Figure 5A, Supplementary Table 6)**. This is expected

since orangutan is the most distantly related great ape species in our study. Similarly, we find

that DHS sites with decreased accessibility in human (human-specific, human-chimpanzee, and

human-chimpanzee-gorilla) have a higher overlap with unannotated regions compared to DHS

sites with decreased accessibility specific to chimpanzee, gorilla, and orangutan. This is also

expected since DHS sites with decreased accessibility in non-human primates will by definition

have higher chromatin accessibility signals in human fibroblasts.

Evolutionary Changes in Accessibility are Associated with Cell-type Specificity

We calculated cell-type specificity (see Materials and Methods) for the union set of DHS

sites detected in primate fibroblasts by comparing them to a much larger set of DHS sites

detected in 125 different human cell and tissue types (Thurman et al. 2012). A cell-type

specificity score close to 1 indicates the DHS site is present in only a few of the 125 tissues and

cell types, while a score near 0 indicates that the DHS site is present in almost all of the 125

tissues and cell types.

As with the proximal and distal annotations, we can make inferences about evolutionary

changes in chromatin accessibility only for DHS sites that overlap the human annotations. The

union set of the DHS sites we identified show a continuum of cell-type specificity scores with

DHS sites from different human cell types (**Figure 5B**). 1,914 (2%) of our DHS sites overlapped

DHS sites found in all 125 tissues and cell types, while 3,873 (4%) of our DHS sites were not

found in any of previously tested tissues and cell types.

We then analyzed the distribution of cell type specificity scores in distal and proximal

DHS sites that displayed changes in chromatin accessibility. In general, distal elements have

higher specificity scores than proximal elements **(Figure 5C,D)**, consistent with previous studies

(Thurman et al. 2012).

For proximal elements showing increases in accessibility, tissue specificity is higher on

all four species branches than on the two internal branches (one sided Wilcoxon test comparing

pooled distributions of external vs internal; $P = 2.03 \times 10^{-28}$) **(Figure 5C)**. The opposite pattern is

evident for decreases in accessibility (one sided Wilcoxon test comparing pooled distributions of

external vs internal; $P = 3.52 \times 10^{-32}$) **(Figure 5C)**. Since all changes on the internal branches are

more ancient than those on external branches, this result hints at the possibility that degree of

chromatin accessibility is positively correlated with broader utilization across cell types. One

possible explanation is that increases in chromatin accessibility raise the likelihood that a

proximal regulatory element is co-opted for use by another tissue. The same trends are observed

for distal elements, with the exception of distal sites having higher tissue specificity scores,

which is expected since distal chromatin accessible sites are more likely to be cell type specific

than proximal elements (**Figure 5D**).

For proximal elements showing changes in chromatin state, the human branch shows

lower cell type specificity compared to the three other species for increases (one sided Wilcoxon

test with Bonferroni correction; $P_{H:C} = 1.62 \times 10^{-5}$; $P_{H:G} = 1.11 \times 10^{-6}$; $P_{H:O} = 4.69 \times 10^{-11}$) and

higher cell type specificity for decreases (one sided Wilcoxon test with Bonferroni correction;

$P_{H:C} = 0.004$; $P_{H:G} = 0.036$; $P_{H:O} = 0.004$) **(Figure 5C)**. The same pattern is present for distal

elements, both for increases and decreases (one sided Wilcoxon test with Bonferroni correction;

Increases: $P_{H:C} = 4.55 \times 10^{-70}$; $P_{H:G} = 1.18 \times 10^{-44}$; $P_{H:O} = 3.47 \times 10^{-133}$; Decreases: $P_{H:C} = 1.11 \times 10^{-6}$;

$P_{H:G} = 2.54 \times 10^{-3}$; $P_{H:O} = 2.38 \times 10^{-8}$) **(Figure 5D)**. This may reflect an ascertainment bias arising

from relying on human tissue comparisons for the cell type specificity score.

Species-specific Chromatin Changes are Enriched in Positive Selection

To investigate the evolutionary significance of species-specific changes in chromatin

accessibility, we tested each DHS site for signatures of positive selection on the human,

chimpanzee, and gorilla branches separately (see Materials and Methods). Testing for positive

selection required additional filtering of DHS sites (see Materials and Methods), resulting in a

reduced set of 87,431 DHS sites used in this analysis. The figure of merit in these analyses is ζ

(zeta), the ratio of substitution rates within a DHS site on a given branch relative to the rest of the

tree in comparison to that ratio for a collection of proxy neutral sites (Wong and Nielsen 2004;

Haygood et al. 2007; Haygood et al. 2010). Similar to the analogous and more familiar ω

(omega), high values of ζ indicate positive selection, values near 1 indicate neutrality, and low

values indicate negative selection.

Putative non functional elements display a relatively tight distribution of ζ on the human

branch centered around 1 (**Figure 6A**), confirming they are a good proxy for neutral evolution in

non-coding regions of the genome. Non differential DHS sites (those where the degree of

chromatin accessibility does not change on any branch) have a distribution of ζ on the human

branch that is centered significantly below 1 (one sided Wilcoxon test; $P = 1.57 \times 10^{-283}$) (**Figure**

6A), consistent with ongoing negative selection. Additionally, the distribution of ζ values is

much broader for non differential DHS sites compared to putative non functional sites, with a

small fraction showing elevated substitution rates on the human branch that are consistent with

positive selection.

DHS sites that showed a change in chromatin accessibility on the human branch have

positively-shifted distributions of ζ on the human branch relative to non differential DHS sites

(**Figure 6A**). This suggests that both increases and decreases are accompanied by enrichment for

a combination of relaxed selection and positive selection on the same branch. As expected, this

enrichment is less pronounced when the accessibility change occurs on a different branch of the

phylogeny: the distributions of ζ on the human branch are higher when the chromatin state

change occurred on the human branch rather than the gorilla or orangutan branches, and this is

true for both increases and decreases (one sided Wilcoxon test with Bonferroni correction;

Increases: $P_{H:C} = 0.12$, $P_{H:G} = 2.17 \times 10^{-10}$, $P_{H:O} = 6.39 \times 10^{-6}$, $P_{H:H-C} = 9.36 \times 10^{-13}$, $P_{H:H-C-G} =$

1.40×10^{-10}; Decreases: $P_{H:C} = 0.006$, $P_{H:G} = 4.13 \times 10^{-5}$, $P_{H:O} = 2.26 \times 10^{-5}$, $P_{H:H-C} = 0.03$, $P_{H:H-C-G} =$

1.92×10^{-4}), although these differences are all modest in magnitude (**Figure 6A**).

For the human-specific accessibility changes, we tested for enrichment of positive

selection of these regions in the human branch relative to either the chimpanzee or gorilla

branches. We performed a similar comparison for the chimpanzee-specific accessibility changes

by testing for enrichment of positive selection on the chimpanzee branch relative to either the

human or gorilla branches. Finally, we tested for enrichment of gorilla-specific accessibility

changes on the gorilla branch relative to the human or chimpanzee branches (**Figure 6B**). None

of the Fisher's exact tests were significant after Bonferroni correction, even though the data

trends in the expected patterns (e.g., human-specific changes have more selection on the human

branch, etc.).

As a control, non differential DHS sites show no significant differences in positive

selection between branches (**Figure 6B**). Additionally, putatively non functional sites in the

genome do not display an enrichment of positive selection (**Figure 6B**). These results suggest

that evolutionary changes in chromatin accessibility between species are phylogenetically

correlated with an enrichment of positive selection.

Next, we investigated the converse: whether signatures of positive selection on individual

regulatory elements are generally limited to branches where the state change occurred. This is

clearly not the case: DHS sites with increased accessibility on the human branch show positive

selection on the chimpanzee branch almost as often as on the human branch (**Figure 6C**). The

same pattern is evident for increased accessibility on the chimpanzee branch and for decreased

accessibility on the human or chimpanzee branch (**Figure 6C**). These results suggest that

positive selection often acts on the DNA sequence of DHS sites in ways that do not affect

chromatin state. Interestingly, some DHS sites show evidence of positive selection on both

branches (**Figure 6C**). Given that 1.96% of DHS sites contain signatures of positive selection in

the human branch, this result suggests that positive selection is not distributed randomly across

the regulatory genome, but occurs repeatedly in a very small percentage of sites (0.12%).

On average, 5.4% of DHS sites that show species-specific accessibility changes for human,

chimpanzee, and gorilla are highly constrained in sequence evolution, with nucleotide

substitution rates that are significantly lower than the neutral expectation across vertebrates

(**Figure 6D**). In contrast, DHS sites that do not change state are enriched for highly constrained

sites in comparison to species-specific changes for human, chimpanzee, and gorilla (Fisher's

exact test, one-sided with Bonferroni adjust for 3 comparisons; $P_{ND:H} = 9.10 \times 10^{-6}$;

$P_{ND:C} = 2.42 \times 10^{-6}$; $P_{ND:G} = 3.18 \times 10^{-4}$) (**Figure 6D**). Together, these results suggest that positive

selection contributes to species-specific chromatin accessibility increases and decreases, while

purifying selection contributes to the conservation of non differential DHS sites.

Discussion

We developed a new method to identify regions of differential chromatin accessibility

among multiple species, based on a negative binomial generalized linear model. This method

does not rely on thresholding and is therefore able to detect subtle differences in degree of

chromatin accessibility that are obscured using conventional approaches. Furthermore, unlike

methods based on pairwise comparison, our method is readily scalable to multiple taxa. Here we

applied it to DNase-seq data from cultured skin fibroblasts obtained from five species. While the

majority of DHS sites were not quantitatively distinct between species, we identified over 35,000

DHS sites with significant differences in chromatin accessibility between human, chimpanzee,

gorilla, orangutan, and rhesus macaque. Of those, approximately 65% are likely the result of a

single evolutionary change in chromatin state that occured on either an internal or external

branch, while the remainder imply multiple evolutionary changes in state.

Our results are largely congruent with our earlier study (Shibata et al. 2012) that used a

threshold-based multiple pairwise comparison approach and considered three primate species

(human, chimpanzee, and rhesus macaque). Here, the use of five species provides additional

confidence in the identification of species-specific accessibility changes and also allows for the

identification of changes that likely occurred multiple times throughout evolution. For these

multiple events, the method we developed does not characterize how exactly they occurred. That

characterization will be the subject of future work using a likelihood analysis that incorporates

the phylogenetic information and models evolutionary processes (Felsenstein 1973; Hansen

1997; Felsenstein 2008; Paradis and Schliep 2019).

As mentioned in the results, we identified substantially more accessibility increases than

decreases. It seems in principle unlikely that increases and decreases in accessibility actually

occur at such different rates, since, if true, primate genomes would eventually become saturated

with open chromatin regions. The same asymmetry was observed previously by us (Shibata et al.

2012) and other groups (Villar et al. 2015; Reilly et al. 2015; Emera et al. 2016) using

conventional pairwise comparisons and thresholding, so the source is unlikely to lie in the

method used to identify chromatin state changes. Instead, it seems likely that the asymmetry is

an ascertainment bias that derives, in part, from unequal statistical power to call gains and losses,

though the exact basis of the bias is currently unclear.

Finding so many DHS site differences in non-human primates is a fascinating result with

implications for understanding the evolution of transcriptional regulation. Nevertheless, we also

suggest that these results describing cell type specificity should be interpreted carefully. One

non-biological possible scenario for such enrichment is that there is ascertainment bias in our

analyses due to the cell-type specificity score being based entirely on data from human, a

limitation imposed by lack of relevant data from other primate species. Additionally, many of

the chromatin accessibility datasets analyzed by Thurman et al. that we used to calculate the cell-

type specificity scores are from fibroblast-like cells. Some of the DHS sites that we identified as

being used in many cell types may in fact be used only in fibroblast-like tissues. Although the

patterns of positive selection that we detected are consistent with expectations, none of the tests

found statistically significant enrichment on the human, chimpanzee, and gorilla branches. This

may be due to our method of positive selection detection relying on human functional

annotations to identify proxy neutral regions, which may result in a loss of power with increasing

phylogenetic distance.

Interestingly, our results suggest that DHS sites are not homogenous from either a

functional or an evolutionary perspective. Those near transcription start sites (including likely

core promoter regions) differ from DHS sites that are distant (including classic enhancers and

other kinds of distal elements) in several regards. DHS sites that show no state changes across

the species surveyed are enriched in conserved nucleotides, consistent with greater functional

significance. Compared with proximal DHS sites, gains in chromatin accessibility in distal sites

are more likely to show same branch-specific signatures of positive selection, as might be

expected if these DHS sites are contributing to changes in gene regulation. These and other

trends we observed suggest that functional constraints and opportunities differ markedly among

classes of DHS sites. Additional studies will be needed to delineate these distinct classes of

likely regulatory elements and to understand how evolutionary mechanisms operate on their

chromatin state and underlying DNA sequence.

Functional characterization studies will be necessary to understand these regions and

their contribution to species-specific gene expression patterns and organismal traits. High-

throughput reporter assays such as MPRA (Klein et al. 2018) and population STARR-seq

(Vockley et al. 2015) can quantify the impact of these differentially utilized regulatory regions,

as well as variants within these regions. In addition, methods such as CRISPR (Diao et al. 2017;

Klann et al. 2017) can characterize the impact of these regions in their natural context, including

identifying the correct target gene(s) for these regulatory elements. Finally, additional replicates

from these species can provide characterization of variability within each species. While

obtaining data from additional tissues for primate species is not possible for most tissues,

generation of induced pluripotent stem cells (iPSCs) followed by differentiation (Gallego

Romero et al. 2018), will provide insights into how these differential chromatin signals translate

into different cell types across many species.

While we used our negative binomial generalized linear model method to identify

differences in chromatin accessibility between five species, we believe this strategy can be used

for quantitative comparisons across many tissues, cell types, and time-series experiments. In

addition to DNase-seq, we expect this method can be readily applied to any count-based data

type such as RNA-seq, ATAC-seq, and ChIP-seq. We also note that this method is scalable, as

increasing the number of groups or replicates requires simply changing the design matrix and

contrasts.

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Figure and table legends

FIG. 1.—Classification of DHS sites. (A-D) Density plots showing the beta values for human (black), chimpanzee (green), gorilla (blue), and orangutan (orange). Beta values near zero indicate the accessibility for that species is similar to rhesus macaque. Positive beta values indicate the accessibility for that species is increased relative to rhesus macaque. Negative beta values indicate the accessibility for that species is decreased relative to rhesus macaque. (A) Non differential sites. (B) Human-specific increases in accessibility. (C) Human-specific decreases in accessibility. (D) Accessibility increases in human and gorilla. (E) Pie chart showing the number and proportion of DHS sites that 1) are non differential; 2) have accessibility changes likely due to a single event; and 3) have accessibility changes that are due to multiple events. Percentages are of the total number of DHS sites. (F) Pie chart showing the number and proportion of each type of single event. The slice labeled "H-C-G-O and Rhesus macaque" contains changes that happened on either the human-chimpanzee-gorilla-orangutan inner branch or the rhesus macaque species branch. Percentages are of the number of DHS sites with a single change. (G) Bar chart showing the relative proportions of increases and decreases in accessibility. Numbers at the top are the total number of DHS sites in each category. Numbers in or just above the orange bar are the number of DHS sites with decreased accessibility. Numbers at the bottom of the green bar are the number of DHS sites with increased accessibility. H-C: Human-chimpanzee internal branch. H-C-G: Human-chimpanzee-gorilla internal branch. H-C-G-O: Human-chimpanzee-gorilla-orangutan internal branch.

FIG. 2.—Species-specific changes in chromatin accessibility. Phylogenetic tree with divergence dates (to scale). UCSC Genome Browser screenshots of representative DHS sites for species-

specific DHS sites for (A) increased accessibility and (B) decreased accessibility. Heatmaps of

normalized scores are rank-ordered DHS sites based on hierarchical clustering.

FIG. 3. —Internal branch changes in chromatin accessibility. Phylogenetic tree with divergence

dates (to scale). UCSC Genome Browser screenshot of a representative DHS site changes that

likely occurred before the human-chimpanzee split (top) and human-chimpanzee-gorilla split

(bottom). Heatmaps of normalized scores are rank-ordered DHS sites based on hierarchical

clustering.

FIG. 4. —Changes in chromatin accessibility due to multiple evolutionary events.

Heatmaps of normalized scores that are rank-ordered DHS sites based on hierarchical clustering.

(A) Two species have increased chromatin accessibility relative to rhesus macaque. (B) Two

species have decreased chromatin accessibility relative to rhesus macaque. (C) Three species

have increased chromatin accessibility relative to rhesus macaque. (D) Three species have

decreased chromatin accessibility relative to rhesus macaque.

FIG. 5. —Chromatin accessibility changes relative to proximal/distal location and cell-type

specificity. (A) The percentage of proximal elements, distal elements, and unannotated elements

for each category of DHS sites. (B) Histogram of specificity scores for DHS sites identified in

this study compared to DHS sites detected in 125 different cell types (Thurman et al). A high

specificity score indicates the DHS site is specific to a small number of cell types. A low

specificity score indicates the DHS site is shared across many cell types. The DHS site categories

are separated into proximal elements (C) and distal elements (D). H-C: human-chimpanzee

internal branch. H-C-G: human-chimpanzee-gorilla internal branch.

FIG. 6. —Effect of natural selection on increases and decreases in chromatin accessibility.

(A) Distribution of the ratio of evolution ζ (zeta) in the human branch for DHS sites. The dashed

green line depicts the critical value where the human zeta value becomes significant ($P < 0.05$).

Zeta values around 1 are expected to be neutral and below 1 are expected to be constrained. (B)

Percentages of DHS sites that are significant for positive selection ($P < 0.05$). Each DHS site was

tested with 3 different foregrounds: human, chimpanzee, and gorilla. (C) Scatterplots of zeta

values for DHS sites with significant positive selection on the human branch (purple diamond),

the chimpanzee branch (blue cross), or both the human and chimpanzee branches (black solid

circle). Zeta values for the human branch are on the x-axis and zeta values for the chimpanzee

branch are on the y-axis. The kernel density depicts non functional sites. (Top left) increased

accessibility in human. (Top right) decreased accessibility in human. (Bottom left) increased

accessibility in chimpanzee. (Bottom right) decreased accessibility in chimpanzee. (D)

Percentages of DHS sites that are highly constrained (median vertebrate PhastCons > 0.9). H-C:

human-chimpanzee internal branch. H-C-G: human-chimpanzee-gorilla internal branch. ***: p-

value < 0.001. **: p-value < 0.01.

Table 1

Number of DHS sites in each category

Table 2

Comparison to earlier study (Shibata et al., 2012, PLoS Genetics)

NOTE. —Increase refers to an increase in chromatin accessibility compared to rhesus macaque.

Decrease refers to a decrease in chromatin accessibility compared to rhesus macaque.

FIG. S1. —Average number of per-species DHS sites at each level of subsampling. There is a

large range in the number of mapped reads between the samples in this study (Supplementary

Table 2). We performed peak calling on random subsets of varying sizes (10 million, 15 million,

20 million, 25 million) for each sample. Trajectories are shown.

FIG. S2. —Distribution of beta values for chromatin accessibility changes due to a single

evolutionary event. Density plots showing the beta values for human (black), chimpanzee

(green), gorilla (blue), and orangutan (orange). Beta values near zero indicate the accessibility

for that species is similar to rhesus macaque. Positive beta values indicate the accessibility for

that species is increased relative to rhesus macaque. Negative beta values indicate the

accessibility for that species is decreased relative to rhesus macaque. Each plot is for a different

category of differential site.

FIG. S3. —Distribution of beta values for chromatin accessibility changes due to multiple

evolutionary events. Density plots showing the beta values for human (black), chimpanzee

(green), gorilla (blue), and orangutan (orange). Beta values near zero indicate the accessibility

for that species is similar to rhesus macaque. Positive beta values indicate the accessibility for

that species is increased relative to rhesus macaque. Negative beta values indicate the

accessibility for that species is decreased relative to rhesus macaque. Each plot is for a different

category of differential site.

FIG. S4. —Principal components analysis of the samples analyzed in this study with (right side)

and without (left side) three additional chimpanzee samples from Pizzollo et al. 2018. (Top)

Principal component 1 (x-axis) vs. principal component 2 (y-axis). (Middle) Principal

component 1 (x-axis) vs. principal component 3 (y-axis). (Bottom) Principal component 1 (x-

axis) vs. principal component 3 (y-axis). The original samples are represented by filled circles

and the Pizzollo et al. samples are represented by filled diamonds. Human samples are in black,

chimpanzee samples are in green, gorilla samples are in blue, orangutan samples are in orange,

and rhesus macaque samples are in purple.

Table S1

Sample information

Table S2

Mapping and LiftOver statistics

NOTE. —(a) Human replicates were not lifted over; the numbers are repeated for consistency.

Percentages of "After lift over" reads are of mapped reads. (b) Final counts are after reads

mapped to sex chromosomes are removed.

Table S3

DHS site counts at different steps in the analysis pipeline

Table S4

Constraint matrices

Table S5

Comparison to earlier study (Shibata et al., 2012, PLoS Genetics)

Table S6

Location of DHS sites

NOTE. —Percentages are of the total number of DHS sites. H-C: human-chimpanzee internal

branch. H-C-G: human-chimpanzee-gorilla internal branch.

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FIG. 1

FIG. 2

FIG. 3

FIG. 4

A

Human/Gorilla Increased Accessibility (n=997)

Human/Orangutan Increased Accessibility (n=997)

Chimpanzee/Gorilla Increased Accessibility (n=1,568)

Chimpanzee/Orangutan Increased Accessibility (n=657)

Gorilla/Orangutan Increased Accessibility (n=2,120)

B

Human/Gorilla Decreased Accessibility (n=221)

Human/Orangutan Decreased Accessibility (n=424)

Chimpanzee/Gorilla Decreased Accessibility (n=387)

Chimpanzee/Orangutan Decreased Accessibility (n=252)

Gorilla/Orangutan Decreased Accessibility (n=517)

C

Human/Chimpanzee/Orangutan Increased Accessibility (n=794)

Human/Gorilla/Orangutan Increased Accessibility (n=691)

Chimpanzee/Gorilla/Orangutan Increased Accessibility (n=939)

D

Human/Chimpanzee/Orangutan Decreased Accessibility (n=906)

Human/Gorilla/Orangutan Decreased Accessibility (n=554)

Chimpanzee/Gorilla/Orangutan Decreased Accessibility (n=686)

FIG. 5

FIG. 6

TABLE 1

ALL DHS SITES	Total	% of total
Non differential (no changes)	53,078	59%
Differential (one or more changes)	36,666	41%
TOTAL	**89,744**	

SINGLE CHANGE AFFECTING SINGLE SPECIES	Increases	Decreases	Total	% of differential
Human	2,539	209	2,748	7%
Chimpanzee	1,539	196	1,735	5%
Gorilla	2,737	254	2,991	8%
Orangutan	3,759	646	4,405	12%
TOTAL	**10,574**	**1,305**	**11,879**	**32%**

SINGLE CHANGE AFFECTING MULTIPLE SPECIES	Increases	Decreases	Total	% of differential
Human/chimpanzee	1,735	814	2,549	7%
Human/chimpanzee/gorilla	1,736	1,453	3,189	9%
Human/chimpanzee/gorilla/ orangutan	1,317	4,992	6,309	17%
TOTAL	**4,788**	**7,259**	**12,047**	**33%**

MULTIPLE CHANGES AFFECTING TWO SPECIES	Increases	Decreases	Total	% of differential
Human/gorilla	997	221	1,218	3%
Human/orangutan	997	424	1,421	4%
Chimpanzee/gorilla	1,568	387	1,955	5%
Chimpanzee/orangutan	657	252	909	2%
Gorilla/orangutan	2,120	517	2,637	7%
TOTAL	**6,339**	**1,801**	**8,140**	**22%**

MULTIPLE CHANGES AFFECTING THREE SPECIES	Increases	Decreases	Total	% of differential
Human/chimpanzee/orangutan	794	906	1,700	5%
Human/gorilla/orangutan	691	554	1,245	3%
Chimpanzee/gorilla/orangutan	939	686	1,625	4%
TOTAL	**2,424**	**2,146**	**4,570**	**12%**

TABLE 2

HUMAN AND CHIMPANZEE CHANGES							
	Consistent (species-specific)		Consistent (multi-species)		Not consistent		Total
Human increases	245	72%	91	27%	6	2%	342
Chimpanzee increases	105	45%	114	49%	15	6%	234
Human decreases	21	14%	109	75%	16	11%	146
Chimpanzee decreases	17	17%	69	70%	12	12%	98

COMMON SITES							
Consistent (non differential)		Consistent (differential only in gorilla, orangutan)		Not consistent		Total	
1,043	90%	57	5%	54	5%	1,154	

FIG. S1

FIG. S2

FIG. S3

FIG. S4

TABLE S1

Sample	Coriell ID	Technical Replicates	Library Date	Number of Sequencing Lanes	Sequencer Type (Machine Name)	Biopsy Source	Sex	Age at Sampling	Notes
Chimpanzee 1	S003649	1	8/6/08	4 (2 lanes per sequencer)	Genome Analyzer II (HWI-EAS215) (HWUSI-EAS465)	Unknown	Male	9 YR	from the Yerkes Primates name is Josh
Chimpanzee 2	S003624	1	11/21/08	3 (2 lanes in one run; 1 lane in another)	Genome Analyzer IIx (HWUSI-EAS1676)	Unknown	Male	14 YR	from the Yerkes Primates name is Tank
Chimpanzee 3	S007603	1	11/21/08	4	Genome Analyzer II (HWUSI-EAS465)	Unknown	Male	16 YR	from the Yerkes Primates name is Donald
Chimpanzee 4	PR00238	1	9/13/12	1	HiSeq 2000 (D3NH4HQ1)	Unknown	Male	24 YR	name is Clint
Chimpanzee 5	S006007	1	9/13/12	1	HiSeq 2000 (D3NH4HQ1)	Unknown	Male	22 YR	
Chimpanzee 6	S008975	1	9/13/12	1	HiSeq 2000 (D3NH4HQ1)	Unknown	Male	19 YR	
Gorilla 1	PR00107	1	10/17/12	1	HiSeq 2000 (HWI-ST1293)	Unknown	Male	19 YR	
Gorilla 2	PR00669	1	10/17/12	1	HiSeq 2000 (HWI-ST1293)	Unknown	Male	15 YR	
Gorilla 3	PR01103	1	10/17/12	1	HiSeq 2000 (HWI-ST1293)	Unknown	Male	16 YR	
Human 1	AG16409	1	8/6/08	4 (2 lanes per sequencer)	Genome Analyzer II (HWI-EAS215) (HWUSI-EAS465)	Unknown	Male	12 YR	Caucasian
Human 2	GM02185	1	12/16/08	4	Genome Analyzer II (HWUSI-EAS465)	Unknown	Male	36 YR	Caucasian
Human 3	GM05879	1	12/16/08	4	Genome Analyzer II (HWUSI-EAS465)	Arm	Female	48 YR	Caucasian
Rhesus macaque 1	AG06252	2	8/6/08	6 (3 lanes per technical replicate)	Genome Analyzer IIx (HWUSI-EAS407)	Arm	Female	5 YR	Indian strain
Rhesus macaque 2	AG08305	2	12/16/08	7 total *Technical replicate #1* 4 lanes (3 lanes in one run, 1 lane in another) *Technical replicate #2* 3 lanes (2 lanes in one run, 1 lane in another)	*Technical replicate #1* Genome Analyzer II (HWUSI-EAS465) *Technical replicate #2* Genome Analyzer IIx (HWUSI-EAS1676) for 1 lane Genome Analyzer II (HWI-EAS215) for 2 lanes	Arm	Male	1 YR	
Rhesus macaque 3	AG08308	1	11/21/08	4 (3 lanes in one run; 1 lane in another)	Genome Analyzer II (unknown)	Arm	Male	1 YR	
Orangutan 1	PR00054	1	8/3/12	1	HiSeq 2000 (D3NH4HQ1)	Unknown	Male	4 YR	Sumatran orangutan (Pongo pygmaeus)
Orangutan 2	PR01107	1	8/3/12	1	HiSeq 2000 (D3NH4HQ1)	Unknown	Male	11 YR	Sumatran orangutan (Pongo pygmaeus)
Orangutan 3	GM04272	1	8/24/12	1	HiSeq 2000 (D3NH4HQ1)	Unknown	Male	10 YR	Sumatran orangutan (Pongo pygmaeus)

TABLE S2

Replicate	Raw reads	Tier 1 (No mismatches)								Tier 2 (1 mismatch)					
		Unique hits		Multiple hits		Unaligned		Unique hits		Multiple hits		Unaligned			
chimpanzee 1	50,096,931	31,674,269	63%	13,476,106	27%	4,946,556	10%	1,612,581	3%	1,735,149	3%	1,598,826	3%
chimpanzee 2	111,972,567	72,746,554	65%	29,841,886	27%	9,384,127	8%	2,750,866	2%	3,307,427	3%	3,325,834	3%
chimpanzee 3	34,388,687	21,848,911	64%	9,120,646	27%	3,419,130	10%	1,169,139	3%	1,379,792	4%	870,199	3%
gorilla 1	200,394,049	113,954,996	57%	46,452,406	23%	39,986,647	20%	17,164,167	9%	15,177,562	8%	7,644,918	4%
gorilla 2	207,690,245	116,195,086	56%	46,244,395	22%	45,250,764	22%	18,247,477	9%	15,755,405	8%	11,247,882	5%
gorilla 3	218,321,295	139,735,317	64%	56,153,448	26%	22,432,530	10%	7,057,645	3%	8,422,152	4%	6,952,733	3%
human 1	50,938,241	32,879,743	65%	12,283,795	24%	5,774,703	11%	2,136,098	4%	2,347,034	5%	1,291,571	3%
human 2	33,630,278	22,180,616	66%	8,140,672	24%	3,308,990	10%	1,141,379	3%	1,654,187	5%	513,424	2%
human 3	23,844,246	15,548,666	65%	5,556,547	23%	2,739,033	11%	1,017,236	4%	1,263,353	5%	458,444	2%
macaque 1	70,897,729	34,129,807	48%	12,911,828	18%	23,856,094	34%	9,092,082	13%	11,503,154	16%	3,260,858	5%
macaque 2	69,917,208	42,438,629	61%	14,314,693	20%	13,163,886	19%	4,609,783	7%	4,988,172	7%	3,565,931	5%
macaque 3	29,133,743	18,596,481	64%	6,046,944	21%	4,490,318	15%	1,456,525	5%	1,666,868	6%	1,366,925	5%
orangutan 1	204,329,161	126,205,138	62%	44,601,400	22%	33,522,623	16%	9,849,034	5%	10,301,994	5%	13,371,595	7%
orangutan 2	223,997,352	136,190,707	61%	51,142,667	23%	36,663,978	16%	10,465,410	5%	12,391,751	6%	13,806,817	6%
orangutan 3	228,118,747	124,383,888	55%	46,679,518	20%	57,055,341	25%	23,616,339	10%	20,069,852	9%	13,369,150	6%

Replicate	Final mapping results						Final results	
	Unique hits		Multiple hits		Unaligned		Unique monoclonal	
chimpanzee 1	33,286,850	66%	15,211,255	30%	1,598,826	3%	11,245,529	22%
chimpanzee 2	75,497,420	67%	33,149,313	30%	3,325,834	3%	51,812,496	46%
chimpanzee 3	23,018,050	67%	10,500,438	31%	870,199	3%	15,660,736	46%
gorilla 1	131,119,163	65%	61,629,968	31%	7,644,918	4%	108,696,016	54%
gorilla 2	134,442,563	65%	61,999,800	30%	11,247,882	5%	95,844,959	46%
gorilla 3	146,792,962	67%	64,575,600	30%	6,952,733	3%	119,415,564	55%
human 1	35,015,841	69%	14,630,829	29%	1,291,571	3%	14,476,479	28%
human 2	23,321,995	69%	9,794,859	29%	513,424	2%	15,287,873	45%
human 3	16,565,902	69%	6,819,900	29%	458,444	2%	13,516,977	57%
macaque 1	43,221,889	61%	24,414,982	34%	3,260,858	5%	40,813,669	58%
macaque 2	47,048,412	67%	19,302,865	28%	3,565,931	5%	23,197,615	33%
macaque 3	20,053,006	69%	7,713,812	26%	1,366,925	5%	13,836,905	47%
orangutan 1	136,054,172	67%	54,903,394	27%	13,371,595	7%	99,830,828	49%
orangutan 2	146,656,117	65%	63,534,418	28%	13,806,817	6%	47,027,229	21%
orangutan 3	148,000,227	65%	66,749,370	29%	13,369,150	6%	108,387,926	48%

Replicate	Mapped	After lift over (a)		Final counts (b)
chimpanzee 1	11,245,529	10,861,657	97%	10,624,634
chimpanzee 2	47,421,354	45,614,631	96%	44,529,183
chimpanzee 3	15,660,736	15,129,473	97%	14,783,843
gorilla 1	104,413,179	101,111,374	97%	98,813,074
gorilla 2	91,500,123	88,494,541	97%	86,567,493
gorilla 3	114,551,049	110,815,063	97%	108,405,287
human 1	14,388,372	14,388,372	100%	14,014,031
human 2	15,204,571	15,204,571	100%	14,810,459
human 3	13,437,729	13,437,729	100%	13,105,259
macaque 1	40,813,669	35,868,548	88%	34,430,825
macaque 2	23,197,615	20,611,242	89%	20,140,263
macaque 3	13,836,905	12,332,942	89%	12,040,473
orangutan 1	99,830,828	89,033,545	89%	86,998,499
orangutan 2	47,027,229	43,114,002	92%	42,076,972
orangutan 3	108,387,926	98,211,154	91%	95,803,771

TABLE S3

Sample	DHS sites
Chimpanzee 1	163,806
Chimpanzee 2	113,653
Chimpanzee 3	104,601
Gorilla 1	107,869
Gorilla 2	122,317
Gorilla 3	86,317
Human 1	99,030
Human 2	84,421
Human 3	90,871
Macaque 1	119,191
Macaque 2	123,237
Macaque 3	85,305
Orangutan 1	161,994
Orangutan 2	180,239
Orangutan 3	113,979

Species	DHS sites
Chimpanzee	75,700
Gorilla	63,512
Human	62,017
Macaque	68,720
Orangutan	94,883

Step	DHS Sites
Before filtering	166,331
After filtering for genomic coverage	124,900
After filtering for zero-count	89,744

TABLE S4

	Human Chimpanzee Gorilla Orangutan
human	[1 -¼ -¼ -¼]
chimpanzee	[-¼ 1 -¼ -¼]
gorilla	[-¼ -¼ 1 -¼]
orangutan	[-¼ -¼ -¼ 1]
macaque	[-¼ -¼ -¼ -¼]
human/chimpanzee	[½ ½ -⅓ -⅓]
human/gorilla	[½ -⅓ ½ -⅓]
human/orangutan	[½ -⅓ -⅓ ½]
chimpanzee/gorilla	[-⅓ ½ ½ -⅓]
chimpanzee/orangutan	[-⅓ ½ -⅓ ½]
gorilla/orangutan	[-⅓ -⅓ ½ ½]
human/chimpanzee/gorilla	[⅓ ⅓ ⅓ -½]
human/chimpanzee/orangutan	[⅓ ⅓ -½ ⅓]
human/gorilla/orangutan	[⅓ -½ ⅓ ⅓]
chimpanzee/gorilla/orangutan	[-½ ⅓ ⅓ ⅓]

TABLE S5

Step	Type of Change	Shibata et al. Pipeline	This Pipeline
Mapping	Mapping software	BWA	Bowtie
Mapping	Mapping parameters	Up to 3 mismatches	Up to 1 mismatch
Mapping	Mapping parameters	Multiple locations allowed	Unique locations only
Mapping	Chimpanzee genome	panTro2	panTro4
Mapping	Macaque genome	rheMac2	rheMac3
DHS site identification	Peak calling program	F-Seq	MACS2
DHS site identification	Per-species DHS sites	Union set of top 100,000 peaks present in any replicate	Union set of all peaks present in at least two replicates
DHS site identification	DHS site width	DHS sites less than 300 bases were extended to 300 bases	DHS site widths unchanged
DHS site identification	Windowing	DHS sites were divided into overlapping windows of 300 bases	None
DHS site filtering	Genomic coverage	None	Genomic coverage of at least 95% of the bases in every species
DHS site filtering	Zero-count filtering	At least one replicate had to have non-zero counts	At least two replicates had to have non-zero counts

	Original sites	With overlapping reads		With 1+ replicate peaks		With 2+ replicate peaks		Passed genomic coverage filter		Passed zero-count filter and analyzed		Not analyzed	
Human Increases	836	831	99%	761	91%	606	72%	531	64%	**310**	**37%**	526	63%
Chimpanzee Increases	676	650	96%	562	83%	442	65%	399	59%	**215**	**32%**	461	68%
Human Decreases	286	281	98%	N/A	N/A	N/A	N/A	198	69%	**133**	**47%**	153	53%
Chimpanzee Decreases	211	194	92%	N/A	N/A	N/A	N/A	135	64%	**89**	**42%**	122	58%

TABLE S6

INCREASED ACCESSIBILITY COMPARED TO RHESUS MACAQUE

	Human		Chimpanzee		Gorilla		Orangutan		H-C		H-C-G	
Proximal	62	2%	54	4%	235	9%	364	10%	44	3%	88	5%
Distal	2,107	83%	1,007	65%	1,638	60%	1,944	52%	1,433	83%	1,353	78%
Unannotated	370	15%	478	31%	864	32%	1,451	39%	258	15%	295	17%
Total	**2,539**		**1,539**		**2,737**		**3,759**		**1,735**		**1,736**	

DECREASED ACCESSIBILITY COMPARED TO RHESUS MACAQUE

	Human		Chimpanzee		Gorilla		Orangutan		H-C		H-C-G	
Proximal	38	18%	41	21%	27	11%	126	20%	274	34%	151	10%
Distal	103	49%	124	63%	183	72%	398	62%	299	37%	806	55%
Unannotated	68	33%	31	16%	44	17%	122	19%	241	30%	496	34%
Total	**209**		**196**		**254**		**646**		**814**		**1,453**	

	Non Differential	
Proximal	11,850	22%
Distal	30,371	57%
Unannotated	10,857	20%
Total	**53,078**	