

1 The SONATA Data Format for Efficient 2 Description of Large-Scale Network Models

3 Kael Dai¹, Juan Hernando², Yazan N. Billeh¹, Sergey L. Gratiy¹, Judit Planas², Andrew P.
4 Davison³, Salvador Dura-Bernal^{4,5}, Pdraig Gleeson⁶, Adrien Devresse², Benjamin K. Dichter^{7,8},
5 Michael Gevaert², James G. King², Werner A. H. Van Geit², Arseny V. Povolotsky², Eilif
6 Muller², Jean-Denis Courcol², Anton Arkhipov^{1,9*}

7
8 ¹ Allen Institute for Brain Science, Seattle, WA, USA

9 ² Blue Brain Project, École Polytechnique Fédérale de Lausanne, Lausanne, Switzerland

10 ³ Paris-Saclay Institute of Neuroscience UMR 9197, Centre National de la Recherche
11 Scientifique/Université Paris Sud, France

12 ⁴ State University of New York Downstate Medical Center, Brooklyn, NY, USA

13 ⁵ Nathan Kline Institute for Psychiatric Research, Orangeburg, NY, USA

14 ⁶ Department of Neuroscience, Physiology and Pharmacology, University College London, UK

15 ⁷ Department of Neurosurgery, Stanford University, Stanford, CA, USA

16 ⁸ Biological Systems and Engineering, Lawrence Berkeley National Laboratory, Berkeley, CA,
17 USA

18 ⁹ Lead contact

19 * Correspondence: antona@alleninstitute.org

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22 Abstract

23 Increasing availability of comprehensive experimental datasets and of high-performance
24 computing resources are driving rapid growth in scale, complexity, and biological realism of
25 computational models in neuroscience. To support construction and simulation, as well as
26 sharing of such large-scale models, a broadly applicable, flexible, and high-performance data
27 format is necessary. To address this need, we have developed the Scalable Open Network
28 Architecture Template (SONATA) data format. It is designed for memory and computational
29 efficiency and works across multiple platforms. The format represents neuronal circuits and
30 simulation inputs and outputs via standardized files and provides much flexibility for adding new
31 conventions or extensions. SONATA is used in multiple modeling and visualization tools, and
32 we also provide reference Application Programming Interfaces and model examples to catalyze
33 further adoption. SONATA format is free and open for the community to use and build upon
34 with the goal of enabling efficient model building, sharing, and reproducibility.

35 Introduction

36 Modern systems neuroscience faces ever-widening streams of data on composition, connectivity,
37 and *in vivo* activity of brain networks (e.g., (Gouwens et al., 2018a; Jiang et al., 2015; Kasthuri et
38 al., 2015; Lee et al., 2016; Markov et al., 2012; Oh et al., 2014; Tasic et al., 2018; de Vries et al.,
39 2018)), supported by major funding initiatives around the world (Amunts et al., 2016; Bouchard
40 et al., 2016; Hawrylycz et al., 2016; Koch and Jones, 2016; Martin and Chun, 2016; Vogelstein
41 et al., 2016). Turning these complex data into knowledge is a challenging task requiring
42 systematic analysis and modeling. Detailed, data-driven modeling in particular will be essential
43 to integrate the experimentally observed hundreds of cell types, intricate connectivity rules, and
44 complex patterns of neuronal dynamics into predictive computational frameworks (Einevoll et
45 al., 2019).

46
47 For this task, scientists need tools that are up to the challenge. Simulation engines, such as
48 NEURON (Carnevale and Hines, 2006), NEST (Gewaltig and Diesmann, 2007), Brian
49 (Goodman and Brette, 2008), GENESIS (Bower and Beeman, 1997), MOOSE (Ray and Bhalla,
50 2008), Xolotl (Gorur-Shandilya et al., 2018), and others offer high computational performance,
51 and recently a number of software interfaces (e.g., neuroConstruct (Gleeson et al., 2007), PyNN
52 (Davison et al., 2009), NetPyNE (Dura-Bernal et al., 2019), Open Source Brain (Gleeson et al.,
53 2018), and the Allen Institute's Brain Modeling ToolKit (BMTK,
54 <https://alleninstitute.github.io/bmtk/>; (Gratiy et al., 2018)) have been developed that allow users
55 to interact with these engines without mastering the underlying programming environments.
56 However, the utility of these tools is limited without a broadly applicable, flexible, and high-
57 performance modeling data format. The current evolution of typical workstyles towards
58 collaborative team projects demands standardized formats for model sharing and reproducibility,
59 as well as for interoperability between tools. Meanwhile, high computational performance of
60 such formats becomes increasingly important to enable efficient representation of growing
61 biological complexity of models.

62
63 While existing solutions, such as the XML-based data format NeuroML (Cannon et al., 2014;
64 Gleeson et al., 2010), the PyNN language (Davison et al., 2009), and the NSDF standard for
65 simulator output (Ray et al., 2016), have proven useful, major challenges remain and are felt
66 acutely in the case of large data-driven models. One problem is a performance bottleneck:
67 storing data about thousands of neurons or millions of synapses in verbose text-based files
68 produces a large disk space footprint and may be challenging for reading/writing in parallel
69 compute environments. Another is that existing formats describe either static models or
70 simulation outputs, but not both. And, for broad adoption of a modeling data format, it needs to
71 be flexible enough to represent a variety of model types (point neuron, biophysically detailed,
72 etc.) and compatible with more specialized formats (e.g., SWC for neuronal morphologies
73 (Cannon et al., 1998)), without compromising computational performance.

74
75 Notably, similar challenges exist in experimental neuroscience (see, e.g., (Koch and Reid,
76 2012)). The situation is improving due to initiatives for experimental data formats, such as
77 NWB:N (Ruebel et al., 2019), BIDS (Gorgolewski et al., 2016), Loom
78 (<https://linnarssonlab.org/loompy>), or spacetx-starfish (<https://github.com/spacetx/starfish>), but
79 for many types of experimental data the community is still far from a widespread adoption of
80 universally agreed-upon formats. These challenges contribute to difficulties in closing the
81 virtuous experiment/modeling loop and to the overall reproducibility crisis (Baker, 2016;
82 Goodman et al., 2016; Koch and Jones, 2016)).
83
84 Here we present the SONATA (Scalable Open Network Architecture TemplAte) data format,
85 which provides an open-source framework for representing neuronal circuits, simulation
86 configurations, and simulation outputs. The format has been jointly developed by the Allen
87 Institute and the Blue Brain Project to facilitate exchange of their large scale cortical models
88 (e.g., (Arkhipov et al., 2018; Billeh et al., 2019; Markram et al., 2015)) and is supported by these
89 organizations' software tools, such as BMTK (<https://alleninstitute.github.io/bmtk/>; (Gratiy et al.,
90 2018)). Support for the format has also been added by other simulation tools -- pyNeuroML
91 (Cannon et al., 2014; Gleeson et al., 2010), PyNN (Davison et al. 2009), and NetPyNE (Dura-
92 Bernal et al., 2019) -- and an interface between SONATA and the NWB:N format (Ruebel et al.,
93 2019) for neurophysiological data has been developed.
94
95 As described below, SONATA utilizes computationally efficient binary formats for storing large
96 datasets while also offering text-based formats for easy editing of less data-rich model
97 components. SONATA represents all aspects of models and simulations, from network structure,
98 to simulation parameters, to input and output activity. It provides much flexibility for describing
99 models at different levels of resolution, including hybrid models. Importantly, because SONATA
100 is already supported by a number of widely used tools and applications, users can get all of the
101 benefits of the format with no extra work on their part. Full specification of the format can be
102 found at the SONATA GitHub page (<https://github.com/AllenInstitute/sonata>), along with the
103 open-source reference application programming interfaces (APIs). To enable broad applications
104 in the field, SONATA is freely available and open to the community.

105

106 Results

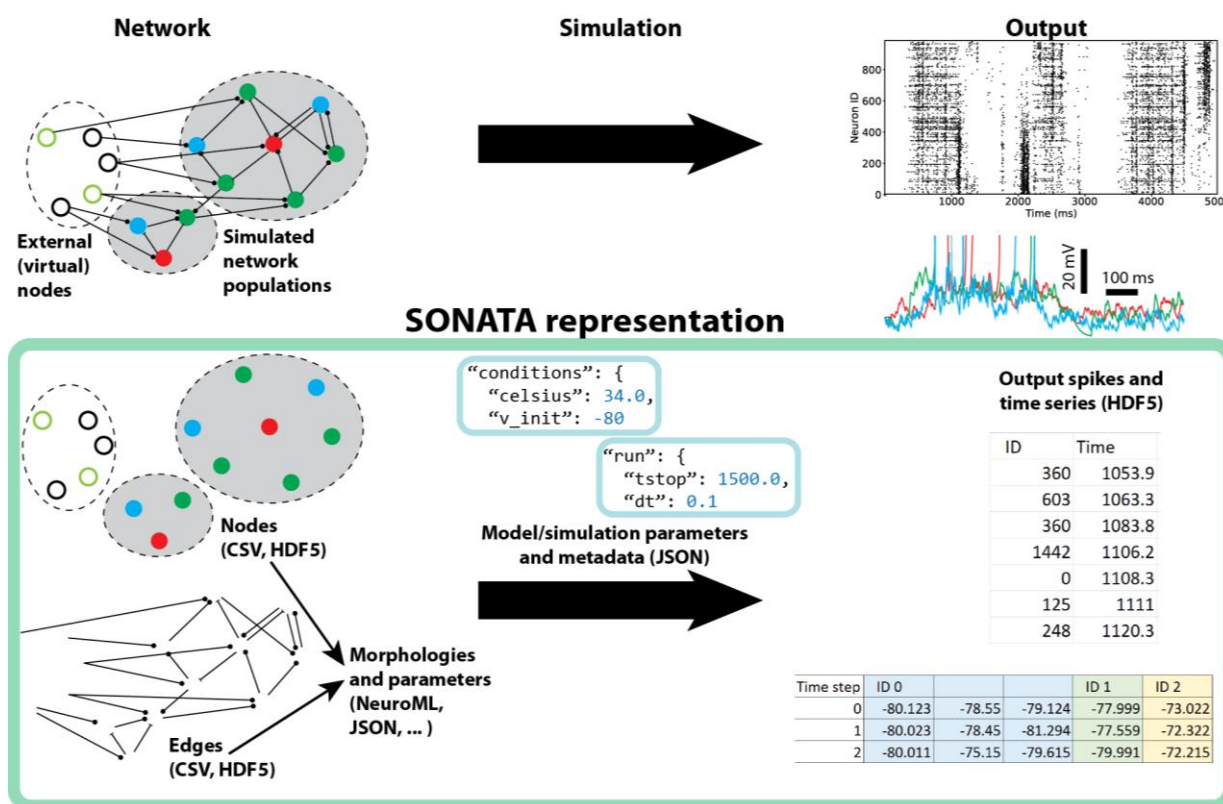
107 Overview of the SONATA format

108 The major object in SONATA is the model network (**Fig. 1**), which consists of **nodes** of two
109 types: explicitly simulated nodes and virtual nodes (the latter only providing inputs to the
110 simulated system). In both cases, nodes are grouped in one or more **populations** for convenience.
111 Nodes within and between populations are connected via **edges**. Simulations of model networks

112 are performed by applications that load SONATA files. Locations of these files and also
 113 parameters of simulation (e.g., the time step and temperature) are stored in the SONATA
 114 configuration (“**config**”) files. Finally, SONATA also provides specifications to store the
 115 incoming activity or simulation output, in the form of events (spikes) or time series.
 116

117 SONATA relies on existing file formats, HDF5, CSV, and JSON (see Methods), which ensures
 118 that files can be read/written by existing libraries and applications and used on all major
 119 operating systems. The SONATA specification on top of these formats accommodates multiple
 120 cell and synapse model types and is designed to optimally handle a heterogeneous network. To
 121 achieve flexibility in defining models, SONATA provides recipes for storing arbitrary attributes,
 122 with some attribute names being reserved for basic standardization.
 123

124 Below, we describe the details of these elements of the SONATA format. A more complete
 125 description is given in the **Online Documentation**
 126 (https://github.com/AllenInstitute/sonata/blob/master/docs/SONATA_DEVELOPER_GUIDE.md).
 127



128 **Figure 1. Overview of the SONATA data format.** (Top) A simulated model consists of one or
 129 more explicitly simulated network populations and external sources (virtual nodes) that provide
 130 inputs into the simulated populations. During and after simulation, output is created
 131 characterizing dynamics in the simulated model. (Bottom) The SONATA data format reflects the
 132 major components of simulation in dedicated file structures. Information about the model is
 133 stored in files (CSV and HDF5) describing nodes and edges of the network (left). Model
 134 metadata (e.g., path relations between files on disk) and information about simulation are stored
 135

136 in JSON configuration files (middle). The spiking and time series output is stored in a tabular
137 format, taking advantage of the HDF5 technology (right). In the case of time series (bottom
138 right), multiple variables can be stored for individual nodes (in this example, node ID 0 has three
139 variables stored), which can correspond, e.g., to multiple compartments of a neuron.
140

141 **Node and edge types**

142 Both nodes and edges can have **attributes** describing biological details (e.g. cell or synapse
143 properties). One major benefit of the SONATA format is its flexibility: while a small number of
144 attributes are reserved, users can create their own attributes for nodes or edges. Furthermore,
145 attributes can be described either individually for each node or more globally for whole subsets
146 of nodes (same for edges), due to the concepts of **node types** and **edge types**. It is up to the user
147 to decide which attributes are stored on a per-type basis and which should be stored individually
148 for each node or edge. Since the number of node/edge types in a network model is usually much
149 smaller than the number of nodes or edges, the node/edge type files are stored in the plain-text
150 CSV tabular format. This makes it easy for modelers to change and update the network *en-masse*
151 through a text editor. For example, **Table 1** shows five different node types, three of which
152 (node_type_id 100, 101, and 102) are biophysically detailed models and two (node_type_id 103
153 and 104) are much simpler, point neuron models. Whereas the total number of nodes in this
154 network can be many thousands, the five entries in **Table 1** succinctly describe many attributes
155 of the nodes.
156

157 The lists of attributes and instructions for constructing individual nodes are determined by each
158 node type's "model_type" (**Table 1**). The reserved values are "biophysical",
159 "single_compartment", "point_neuron", or "virtual". The mechanisms required for cell models
160 are described by "model_template", with possible values including references to a NeuroML2
161 file or a NEURON hoc template. The reserved "morphology" attribute references a morphology
162 file (e.g., in the widely used SWC format) and the "dynamics_params" references files that can
163 be optionally used to initialize or overwrite electrophysiological attribute values defined by the
164 template. In **Table 1**, node types 100 and 101 are built using hoc templates from the Allen Cell
165 Types Database (<http://celltypes.brain-map.org>), which take parameter values from the JSON
166 files in "dynamics_params". Node type 102 uses a NeuroML template file; dynamics_params =
167 NONE means that default values from the NeuroML model_template are used. Node types 103
168 and 104 are NEURON built-in IntFire1 point processes taking parameter values from the JSON
169 files under "dynamics_params".
170

171 Edge types are described in similar ways (**Table 1**). The "model_template" attribute determines
172 the synaptic model via a template file or a synaptic type defined in a particular simulator, e.g.,
173 NEURON's exp2syn, whereas the optional "dynamics_params" initializes or overwrites the
174 parameters of the synaptic mechanisms, e.g., time of rise and decay of synaptic conductance.

175 Other reserved attributes include synaptic weight, delay, and the afferent and efferent locations of
 176 synapses (only the delays are shown in **Table 1**).

177

178 **Table 1: Examples of “node types” and “edge types”.** In a network model, all individual
 179 nodes belonging to a particular node type share the respective attributes, and likewise all edges
 180 belonging to the same edge type share attributes of that type.

Node types				
node_type_id	model_type	model_template	morphology	dynamics_params
100	biophysical	ctdb:Biophys1.hoc	scnn1a_m.swc	472363762_fit.json
101	biophysical	ctdb:Biophys1.hoc	rorb_m.swc	473863510_fit.json
102	biophysical	nml:PV1.nml.xml	pv1_m.swc	NONE
103	point_neuron	nrn:IntFire1	NONE	if1_exc.json
104	point_neuron	nrn:IntFire1	NONE	if1_inh.json
Edge types				
edge_type_id	model_template	dynamics_params		delay
100	exp2syn	biophys_exc.json		2.0
101	exp2syn	biophys_inh.json		2.0
102	NONE	Instantaneous_exc.json		2.0
103	NONE	Instantaneous_inh.json		2.0

181

182 Nodes

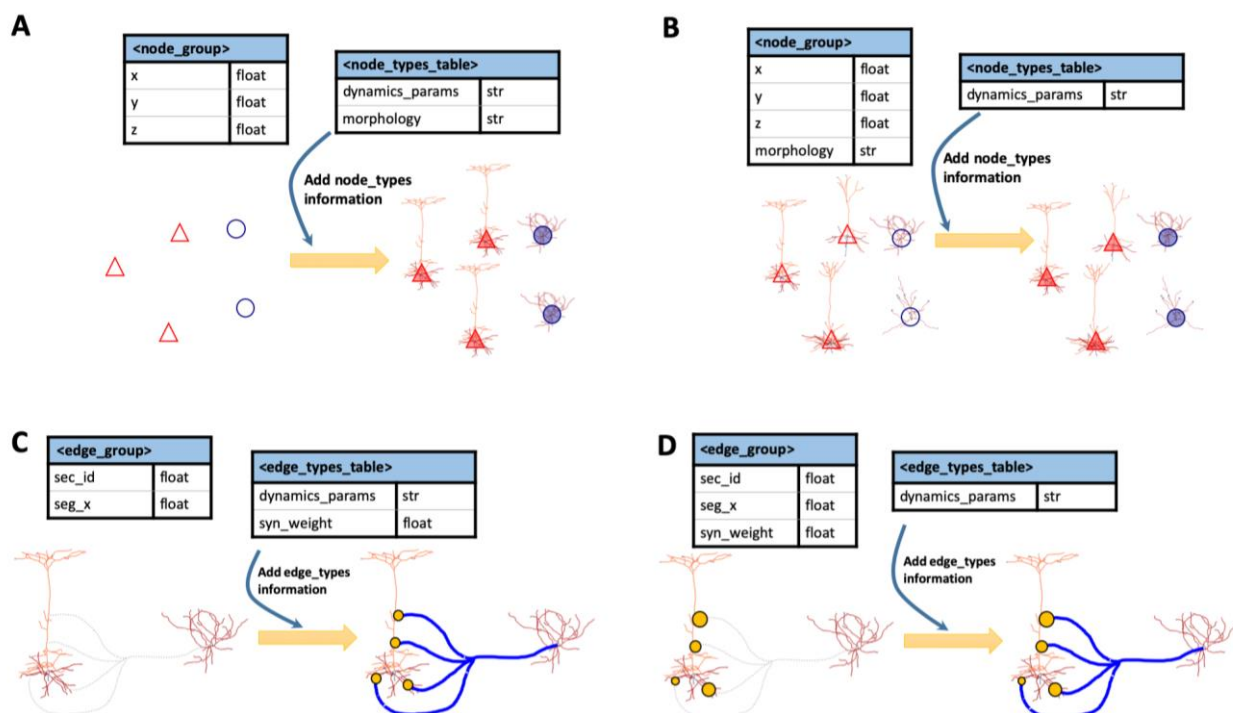
183 Individual attributes of nodes are listed in “node tables”, stored as HDF5 files. As discussed,
 184 users decide which attributes to store in node-type CSV and which in node table HDF5. For
 185 example (**Fig. 2A**), one can store only the coordinates of neurons (x, y, z locations) in the node
 186 table with a pointer (the *node_type_id*) to the node types table for repeated information such as
 187 morphology (see example in **Table 1**). Alternatively, each neuron may have its own unique
 188 morphology (**Fig. 2B**), and in that case the node table contains both the coordinates and the
 189 morphology attribute.

190

191 SONATA allows for nodes to be hierarchically organized into **populations** and **groups**.

192 Different populations may be stored in different files, allowing modelers to mix and reuse
 193 populations between simulations. For example, one may study one brain region -- say, visual area
 194 V1 -- in one simulation and visual area V2 in another simulation, and then build a simulation of

195 V1 and V2 together using the two populations without the need to create new nodes files. Within
196 a population, there is one or more node groups, each group using a homogeneous collection of
197 node attributes. This is useful for hybrid simulations. For example, compartmental neuron
198 models often have many more (and radically different) attributes than integrate-and-fire models.
199 Thus, for mixed populations it is practical to store attributes of compartmental and integrate-and-
200 fire nodes in different groups. Note that nodes of multiple types may be stored in each group, as
201 long as all the nodes in the group have the same lists of attributes. The SONATA implementation
202 of populations and groups utilizes HDF5 groups and datasets (see **Online Documentation**).
203



204
205 **Figure 2. Nodes and edges in SONATA format.** (A, B) Two examples are shown that
206 demonstrate how for each node one can find its model attributes in either the node_group (for
207 individually unique attributes) or the node_types table (for globally shared attributes). In (A), the
208 unique attributes are only the node locations (x, y, z), indicated by empty triangles and circles on
209 the left. Morphology and dynamic parameters are shared among multiple nodes within a type.
210 Hence, all red triangles share the same morphology, as do blue circles (right). In (B), the
211 morphology is unique for each node. The dynamics_params is the only attribute specified at the
212 type level; it is assigned to each node, as indicated by the triangles and circles being filled with
213 color on the right. (C, D) Same for edges. In (C), the synapse locations are stored individually
214 for each edge, whereas synaptic weights and dynamics_params attributes are stored at the edge
215 type level, as indicated by the uniform circle size and colored connections on the right.
216 (“dynamics_params” attributes here determine the dynamical properties of synapses, such as the
217 time of rise and time of decay of synaptic conductance). (D) The synapse locations as well as
218 synaptic weights are stored individually (hence different circle sizes), whereas the
219 dynamics_params attributes are stored at the edge type level.
220

221 Edges

222 An edge typically represents a synapse from one neuron to another. Like for nodes, shared
223 attributes of many edges can be stored in CSV edge type files and individual attributes in HDF5
224 edge tables files (**Fig. 2C, D**). Users decide which attributes belong to edge types and which to
225 edge tables. In the edge tables, edges are grouped together into **edge populations**. Each edge
226 population contains directed connections between nodes in one node population to nodes in
227 another population (the target and source populations can be the same). Each edge identifies the
228 node_id of the source node and the node_id of the target. There may be multiple edges for a
229 single source/target pair. As with nodes, each edge population consists of one or more edge
230 groups. One edge group contains edges with the same list of attributes.

231
232 Continuing our example of a model of V1 and V2 above, one can use one edge population for all
233 connections from V1 to V2, another for V2 to V1, another for V1 to V1, and one more for V2 to
234 V2. The specific partition is again up to users, but has to be consistent with the partition of nodes
235 into populations. Within the V1-to-V1 edge population, one may need to have two edge groups.
236 One edge group would be used for connections to biophysically detailed cell models, containing,
237 for example, attributes of synapse location on the dendritic tree of the target cell, local synapse
238 strength, time delay specific to that particular edge, and many others. The other edge group
239 would be used for connections to point-neuron models, perhaps containing only the synaptic
240 weight.

241
242 For technical details and benchmark examples of SONATA representation of edges, see
243 Methods.

244

245 Simulation configuration

246 SONATA provides a framework for storing the information about the location of the files
247 describing the model, as well as parameters of the simulation and metadata. This information is
248 stored in the **config** files that tie all the network, circuit, and output components together (**Fig. 1**).
249 The SONATA configuration files, the **primary config**, the **circuit config**, and the **simulation**
250 **config**, are JSON files containing key/value pairs. **Table 2** lists the keys required in each of these
251 files (see **Online Documentation** for details).

252

253 The circuit config contains pointers to the files with the information about nodes and edges that
254 describe the network being simulated. The simulation config describes properties unique to a
255 specific simulation run, such as the inputs the network receives, the simulation parameters (for
256 example, duration, time-step), optional parameters such as the temperature, the outputs to be
257 recorded (for instance spike times, membrane potentials, internal calcium concentrations, etc.),
258 paths to writing the outputs, and others. Both the simulation config and the circuit config may
259 contain a manifest block that defines the paths to be used/reused throughout the JSON file. The
260 primary config simply points to the simulation and circuit configs.

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Separating of config files in this manner provides flexibility to mix and match models and simulations. For example, one can use a single circuit config and multiple simulation configs to run many simulations of one model under different conditions, or alternatively use multiple circuit configs with one simulation config to study multiple circuits under identical conditions.

Table 2. Summary of the *config* files. Representative components are listed; additional entries can be used as described in the **Online Documentation**.

Primary config: Defines relative location of each part of a network simulation	
Key	Description
network	Defines the network config file
simulation	Defines the simulation config file
Circuit config: Defines relative locations of circuit components	
Key	Description
components	Directories for neuron morphologies, synaptic models, mechanisms, and neuron models
network/nodes	Specifies CSV file describing node types (key: node_types_file) and HDF5 file containing individual nodes (key: nodes_file)
network/edges	Specifies CSV file describing edge types (key: edge_types_file) and HDF5 file containing individual edges (key: edges_file)
Simulation config: Defines simulation conditions and inputs for the circuit	
Key	Description
manifest	Convenient handle on setting variables that point to base paths
run	Specifies global parameters of the simulation run, such as total duration
conditions	Specifies optional global parameters with reserved meaning associated with manipulation
node_sets	Contains subsets of nodes that act as targets for different reports or stimulations, or can also be used to name and define the target subpopulation to simulate
inputs	Specifies the inputs to the network with a different block for every input (if more than one)

output	Configures the location where output reports should be written, and if output should be overwritten
reports	Defines the specifications of the output variables

269

270 Input and output activity

271 In addition to representing models, SONATA also describes dynamical variables such as spikes
272 and time series, which is necessary for representing incoming activity or output of simulations.
273 For these types of data, SONATA's format is in many ways similar and consistent to the
274 experimental neurophysiology format NWB:N (Ruebel et al., 2019), the two formats having been
275 developed approximately simultaneously and with mutual influences due to interactions between
276 the two developer communities. Both are designed to be optimal for large-scale recordings or
277 simulations. At present, the SONATA output format and NWB:N are maintained in separate
278 projects, but conversion between the two is straightforward and is achieved by a tool described
279 below (see **Ecosystem support**). In the future, it may be desirable to achieve full integration
280 between NWB:N and SONATA.

281 Activity format design

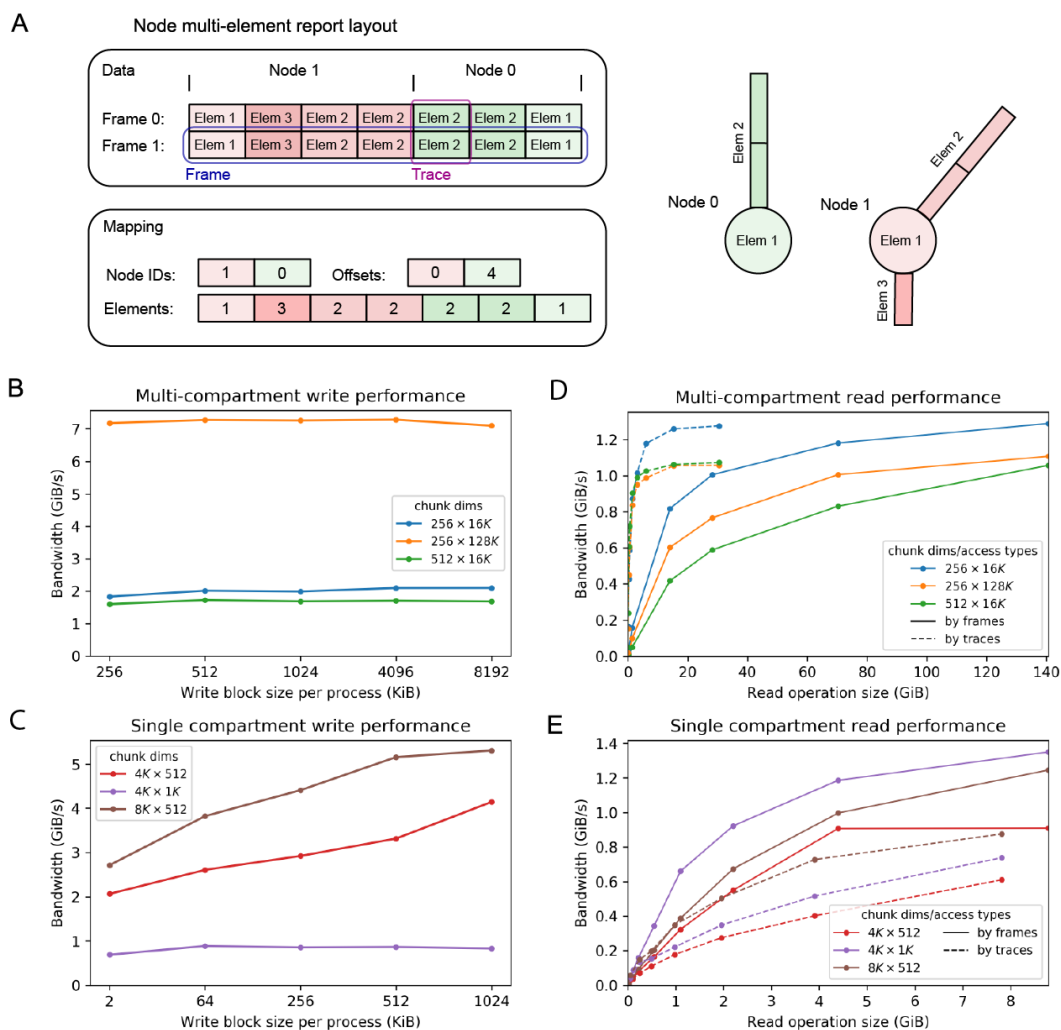
282 The SONATA activity format (also referred to as **reports**) is designed to efficiently support three
283 types of data: spike trains, time series for node elements (e.g., membrane voltage or Ca^{2+}
284 concentration in cell compartments) and time series that are not associated with specific node
285 elements (such as voltages recorded with extracellular probes). The file formats are based on
286 HDF5.

287
288 The data stored in a spike train report consists of a series of node identifiers and spike times,
289 stored in separate HDF5 datasets. For maximum flexibility, the standard allows the datasets to be
290 sorted according to three different criteria: by node ID, by spike time, or unsorted.

291
292 A node element report consists of a set of variables which are sampled at a fixed rate for some
293 elements of interest from a selected set of cells. Typically, the elements are electrical
294 compartments, but other elements can be used as well, such as individual synapses. The time
295 series associated with each element can be membrane voltage, synaptic current, or any other
296 variable. In the report, a simulation **frame** is the set of all values reported at a given timestamp
297 and a **trace** is the full time series of all values associated with one element (**Fig. 3A**). The
298 requirements we followed in designing the node element report were: (i) support for large data
299 sets both in total size (terabytes) and number of elements (millions of cells using multi-
300 compartment models), (ii) random read access to specific frames and elements within a frame,

301 (iii) high performance for different read access patterns (especially full frames and full cell
 302 traces) and (iv) high performance sequential parallel writing of full frames.

303
 304 In the resulting design, data are stored in a single NXM matrix dataset, with rows being frames
 305 and columns being traces, whereas extra metadata provides a *mapping* between (cell, element)
 306 identifiers and columns within the frame (Fig. 3A). The format provides substantial flexibility,
 307 in particular permitting one to save different types and amounts of information for different cells.
 308 For example, one can choose to save membrane voltage and synaptic currents for all
 309 compartments and all synapses for a few cells, only somatic membrane voltage for several other
 310 cells, and nothing at all for all the other cells. This design also readily represents non-cell-
 311 element time series reports. In this case, instead of the cell elements, each row represents a
 312 channel storing a particular time series -- for example, an electrode at which the extracellular
 313 voltage is recorded.



314
 315 **Figure 3. Recordings of activity in SONATA format.** (A) Layout of a multi-compartment
 316 report. The dataset is a matrix where each frame (set of values at one point in time) is a row and
 317 columns represent traces (the time series of all values associated with one element). All the

318 elements of a node are contiguous within a frame, but nodes may not appear sorted by GID. The
319 position of the first element of each node is indicated by the offset array. Node elements can
320 appear multiple times (e.g. morphological sections with multiple electrical compartments). (B-E)
321 Examples of read/write performance (see **Methods**). Write performance (B, C) and read
322 performance (D, E) of multi-compartment reports (B and D) and single compartment reports (C
323 and E) is measured as bandwidth (amount of data written/read per time unit). Three different
324 HDF5 chunk dimensions (specified in the legend, note that the K suffix indicates multiplication
325 by 1024) were evaluated to demonstrate that high effective bandwidth can be obtained. In the
326 reading evaluation, data was read by frames (continuous lines) and by traces (dotted lines) in
327 single operations of different sizes to demonstrate the flexibility and high performance of the
328 SONATA format; in the writing evaluation, data was only written by frames (continuous lines),
329 which imitates the way most simulators generate data.

330 Performance benchmarks

331 **Fig. 3B-E** illustrates the effective I/O bandwidth (amount of useful data read/written per time
332 unit) of SONATA multi-compartment and single-compartment reports, using 26,576 neurons
333 (41,389,269 reported cell elements) with 1,000 time steps for the former and 217,000 neurons
334 with 130,000 time steps for the latter (see **Methods**). We considered (i) the amount of data
335 read/written, (ii) HDF5 chunk dimensions, (iii) only for write benchmarks — the amount of data
336 written at each write operation (block size per process), and (iv) only for read benchmarks — the
337 direction in which data is accessed (by frames or by traces). We did not consider the latter option
338 in the write benchmark because simulators typically generate data which is ordered temporally,
339 i.e. in frames.

340
341 Note that HDF5 provides a storage layout in which the dataset is split into fixed size “chunks”
342 (see **Methods**). Chunking is essential for obtaining good performance with arbitrary access
343 patterns, and for that reason is supported in SONATA. However, SONATA does not prescribe
344 specific chunking, and taking advantage of chunking to optimize read/write performance for
345 specific applications is up to the specific software implementations that use SONATA.

346
347 The benchmarks in **Fig. 3B-E** show that SONATA supports high read and write performance.
348 The write performance reaches several GiB/s. In the case of multi-compartment reports, the
349 HDF5 chunk size is the main determinant of the effective write performance (**Fig. 3B**). This is
350 due to the overhead caused when using smaller HDF5 chunk dimensions, as the increase in
351 absolute number of HDF5 chunks makes the support data structures in the file larger. On the
352 contrary, in single-compartment reports (**Fig. 3C**) the amount of data written by each process at
353 each write operation affects performance, since writing data in small block sizes is not efficient.
354 Here the performance is also affected by the fact that, in some cases, multiple processes write to
355 the same HDF5 chunk, which leads to lower effective bandwidth (compare $4K \times 512$ vs $4K \times$
356 $1K$). The read performance tests (**Fig. 3D, E**) were run on a single-node, single-thread

357 configuration, because this is the typical scenario of analysis and visualization use cases. In all
358 cases, read bandwidth improves as the number of contiguous cells per operation increases and
359 reaches 1 GiB/s and above.

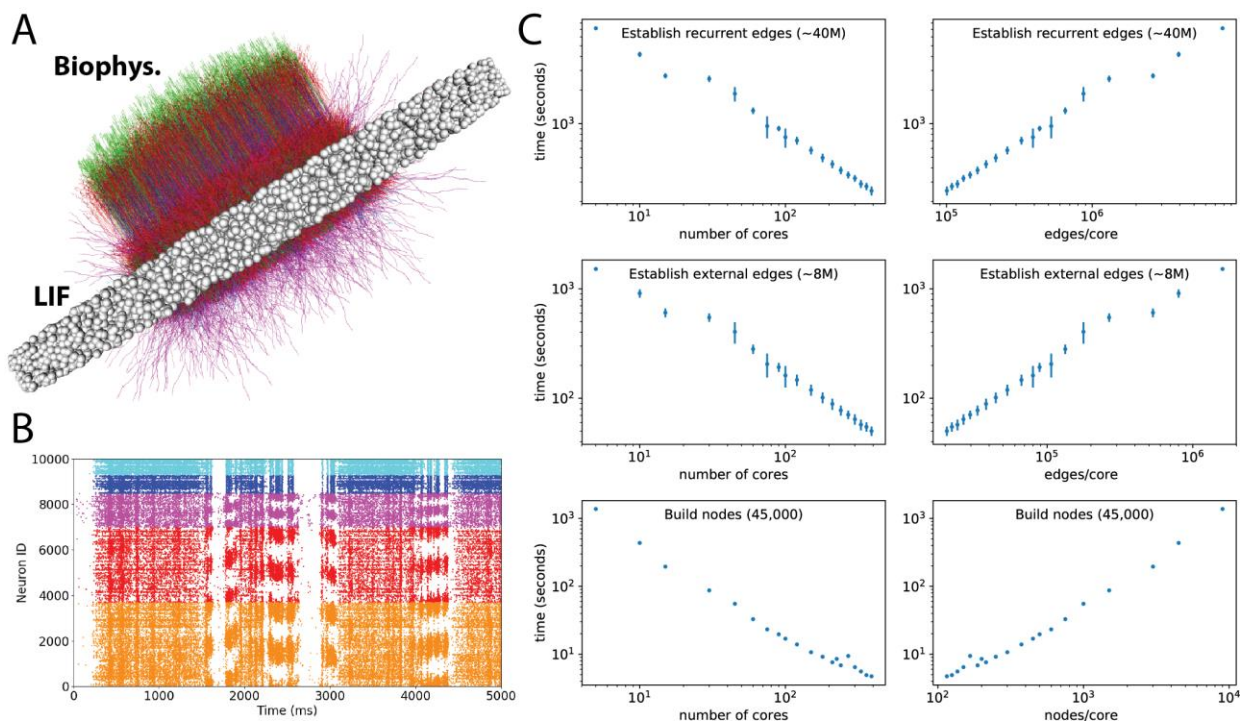
360 **An example of a large-scale model: a network model of the layer 4 of** 361 **mouse cortical area V1**

362 To provide a realistic example of handling large-scale biologically detailed networks with
363 SONATA, we consider the recently published network model of the layer 4 of the mouse
364 primary visual cortex (area V1) (Arkhipov et al., 2018). The model consists of 45,000 neurons
365 (representing more than half of layer 4 neurons in V1) and employs realistic patterns of highly
366 recurrent connectivity. The central portion of the model (**Fig. 4A**) consists of 10,000 neurons
367 modeled using a biophysically detailed, compartmental approach, whereas the remaining 35,000
368 neurons are modeled using a much simpler point-neuron, leaky integrate-and-fire (LIF) approach
369 and serve mainly to prevent boundary artifacts. This hybrid model contains ~40 million edges
370 for connections between explicitly modeled nodes and another ~8 million edges from ~10,000
371 external virtual nodes providing external spiking inputs. In the original study, the model was
372 subjected to a battery of visual stimuli (movies), and the results were compared to published
373 work and new in vivo experiments (Arkhipov et al., 2018) (see an example of spiking output in
374 **Fig. 4B**).

375
376 **Fig. 4C** shows benchmarks for loading the layer 4 model in SONATA format for simulation in
377 NEURON (Carnevale and Hines, 2006) using the BMTK's BioNet module (Gratiy et al., 2018),
378 performed on cluster partitions from 5 to 390 CPU cores. The times required to build the nodes,
379 establish edges from the external virtual nodes, and establish edges among the explicitly
380 simulated, recurrently connected nodes are shown (note that these times include both reading the
381 files and instantiating NEURON objects). Two views of the same data are presented: (i) scaling
382 with the number of cores and (ii) scaling with the number of edges or nodes per core. The
383 scaling is approximately linear (with a slope close to 1) starting at about 32 cores. The overall
384 simulation setup time is dominated by the recurrent connections, which are about 5 times more
385 numerous than the virtual input connections and take about 5 times longer to set up.

386
387 For a typical use case of hundreds of CPU cores, the 45,000-neuron hybrid layer 4 network
388 model requires <10 s for instantiating nodes, <50 s for external edges, and ~4 minutes for
389 recurrent edges, resulting in ~5-minute setup time total. Using uncompressed HDF5 files, the
390 total size of network files, including recurrent and feedforward network connections, is ~2.4 GB
391 (see <http://portal.brain-map.org/explore/models/l4-mv1>). Thus, for this considerably large and
392 detailed model, SONATA supports modest loading times and storage space footprint. We also
393 previously demonstrated good scaling of simulation time for this model (Gratiy et al., 2018).

394



395
 396 **Figure 4. A 45,000-neuron hybrid network model of the layer 4 of mouse cortical area V1.**
 397 (A) Visualization of the network model, which consists of 10,000 biophysically detailed neurons
 398 (colored morphologies) in the center and 35,000 point neurons (white balls) forming an annulus
 399 around the biophysical neurons to prevent boundary artifacts. (B) An example raster plot output
 400 from a simulation of the layer 4 model. Shown are the spikes of 10,000 biophysical neurons in
 401 response to a clip from a natural movie. Colors indicate the five types of neurons: excitatory
 402 Scnn1a (orange), Rorb (red), Nr5a1 (magenta) and inhibitory PV1 (blue) and PV2 (cyan). See
 403 details in (Arkhipov et al., 2018). (C) Benchmarks for instantiating different parts of the layer 4
 404 model. The left and right column show the same data: against the number of CPU cores used for
 405 simulation on the left and against the number of edges or nodes per core on the right.

406 Ecosystem support

407 SONATA is a free format open for community development. Anyone wishing to add SONATA
 408 support to a Python based application may utilize the PySONATA Python API hosted at GitHub
 409 and developed jointly by the Allen Institute and Blue Brain Project (BBP). Multiple tools from
 410 these two organizations and other modeling and standardization initiatives already implement
 411 SONATA support (Fig. 5).

412
 413 Below we briefly describe examples of using these tools to construct, read, write, visualize, and
 414 simulate network models in SONATA format. Note that, in general, when different simulators
 415 load one SONATA model for simulation, bitwise agreement between their outputs is not
 416 guaranteed. The reasons for that are non-standardized processing of certain data in simulation

417 software packages, different approaches for instantiating initial conditions, etc. For a real-life
418 example, consider that loading SWC morphologies in NEURON can be done using different
419 functions (e.g., hoc or Python), which employ different numerical precisions; as a result,
420 simulation outputs will not be bitwise identical, but will be only statistically the same to the level
421 permitted by the precision discrepancy in morphologies. Nevertheless, SONATA constrains a
422 vast variety of important degrees of freedom in network simulations, enabling statistically similar
423 results between simulators and bitwise reproducibility within a simulator with fixed software
424 code.

425
426 Although SONATA has been originally developed to support very large and biologically
427 complex simulations, it is fully consistent with more typical smaller-scale and less complex
428 applications. For example, it is rather common for modelers to use conceptual rules implemented
429 in a few lines of code to generate model geometries and connections. These approaches are fully
430 supported by BMTK, Brion/Brain, pyNeuroML, PyNN, and NetPyNE described below -- in
431 addition to the advanced capabilities of these tools to build and carry out very sophisticated, data-
432 driven, large-scale network simulations. Each of these software packages can generate models
433 using such high-level conceptual definitions, and in fact the examples illustrated in **Fig. 5** were
434 generated in such a simple way using the BMTK's model building module. The important new
435 contribution that SONATA makes is a standardized, efficient format for exchanging generated
436 network structures, as well as simulation results, between these applications. That is showcased
437 in **Fig. 5**, where the BMTK-generated models are simulated using several other tools.
438 Furthermore, it is important to note that large scale biologically realistic models (e.g., (Arhipov
439 et al., 2018; Markram et al., 2015)) often require as much or even more time to build than to run
440 a single simulation, and then saving model instantiations becomes very important, whereas for
441 small models this may be simply unnecessary. However, for sharing models with the community,
442 and especially across simulator platforms, the ability to save all instantiated parameters of
443 models and simulations systematically -- as provided by SONATA -- becomes important for
444 large and small models alike. The examples in **Fig. 5** are relatively small, 300-neuron models,
445 illustrating use cases that are more common in the field than the very large simulations with tens
446 of thousands of neurons (**Fig. 4**).

447
448 Currently, SONATA is not natively supported by the widely used simulation engines NEURON
449 and NEST, but the tools described below provide convenient interfaces to NEURON and NEST
450 and enable simulations with SONATA using these two engines. In the future, implementation of
451 native support in NEURON and NEST could be useful for systematic conversion of older,
452 existing models (which are typically stored as software code) to SONATA format by
453 instantiating these models in NEURON or NEST environment from the original code and then
454 saving as SONATA files.

455 PySONATA

456 PySONATA is a Python based API for reading SONATA files, open-sourced under a BSD
457 license and maintained as an official tool of the SONATA working group
458 (<https://github.com/AllenInstitute/sonata>). Users wishing to begin integrating the SONATA
459 format into their own software are encouraged to use the PySONATA Python modules.
460 Examples of how to use the module can be found at
461 [https://github.com/AllenInstitute/sonata/blob/master/src/pysonata/docs/Tutorial%20-](https://github.com/AllenInstitute/sonata/blob/master/src/pysonata/docs/Tutorial%20-%20pySONATA.ipynb)
462 [%20pySONATA.ipynb](https://github.com/AllenInstitute/sonata/blob/master/src/pysonata/docs/Tutorial%20-%20pySONATA.ipynb).

463 The Brain Modeling Toolkit

464 The Brain Modeling Toolkit (BMTK; <https://github.com/AllenInstitute/bmtk>) is a Python based
465 package for building, simulating and analyzing large-scale neural networks across different
466 levels of resolution. The BMTK is open-sourced under a BSD-3 license and has full support for
467 generating and reading the SONATA data format (**Fig. 5**). Modelers can use the BMTK Builder
468 submodule to create their own SONATA based networks from scratch. It supports cell template
469 files, electrophysiological parameters, and morphology from the Allen Cell Types Database
470 (<http://celltypes.brain-map.org/>) (Gouwens et al., 2018b; Teeter et al., 2018) as well as other cell
471 model formats, including NeuroML2 (Cannon et al., 2014; Gleeson et al., 2010), NEURON hoc
472 files (Carnevale and Hines, 2006), or even user defined Python functions. For simulations,
473 BMTK relies on an increasing array of simulation engines (NEURON (Carnevale and Hines,
474 2006), NEST (Gewaltig and Diesmann, 2007), Dipde (Cain et al., 2016), etc.), which allow users
475 to run simulations of SONATA networks using either multi-compartment, point, or population
476 based representations. The results of these simulations, regardless of the underlying simulator
477 used to run them, are transformed into SONATA output format, allowing networks built and run
478 with BMTK to be analyzed and visualized by any third-party software that supports SONATA.
479 **Fig. 5B** and **5C** show a network with 300 biophysically detailed cells, in SONATA format,
480 generated using BMTK and visualized with RTNeuron and NetPyNE, respectively. The results
481 of simulations of this network using BMTK and NetPyNE are shown in **Fig. 5D**. **Fig 5E** shows
482 simulations of a network of 300 integrate and fire neurons created with BMTK and simulated
483 using BMTK, PyNN, and pyNeuroML.

484 Brion/Brain

485 The Blue Brain's C++ libraries for handling large scale data and simulation setup, Brion/Brain
486 (<https://github.com/BlueBrain/Brion>), provide partial support for SONATA. Currently Brion
487 provides a low level API to read circuit and simulation JSON configurations, spike and multi-
488 compartment simulation outputs, SWC morphologies and query nodes in HDF5 files. It also
489 provides a single threaded writer for multi-compartment simulation output reports. Brain
490 provides a higher level API that makes it easier to work with full networks. All this functionality
491 is also available in Python through the associated Python wrapping module.

492 libSONATA

493 Blue Brain's libSONATA (<https://github.com/BlueBrain/libsonata>) is a library that provides
494 support to read SONATA files. The library is open-sourced under a LGPLv3 license and offers
495 an API for both Python and C++ applications. Currently libSONATA supports reading circuit
496 files, including nodes and edges populations.

497 RTNeuron

498 Blue Brain's RTNeuron (Hernando et al., 2013) is a framework for visualizing detailed neuronal
499 network models and simulations. As it relies on Brion/Brain for data access, it currently provides
500 basic support to visualize SONATA circuits and simulations. For instance, **Fig. 5B** illustrates the
501 RTNeuron visualization of a model of 300 biophysically detailed neurons, provided as an
502 example in the SONATA specification GitHub repository
503 (https://github.com/AllenInstitute/sonata/tree/master/examples/300_cells). Here, one can see
504 neuronal morphologies and the distribution of membrane voltage across the electrical
505 compartments comprising these morphologies as the simulation evolves over time.

506 pyNeuroML

507 NeuroML (Cannon et al., 2014; Gleeson et al., 2010) is a standardized format based on XML for
508 declaratively describing models of neurons and networks in computational neuroscience. Cellular
509 models which can be described range from simple point neurons (e.g. leaky integrate and fire) to
510 multicompartmental neuron models with multiple active conductances. Networks of these cells
511 can be specified, detailing the 3D positions or populations, connectivity between them and
512 stimulus applied to drive the network activity.

513
514 Multiple libraries have been created to support user adoption of the NeuroML language,
515 including jNeuroML (<https://github.com/NeuroML/jNeuroML>) in the Java language and
516 pyNeuroML (<https://github.com/NeuroML/pyNeuroML>) in Python. The latter package also gives
517 access to all of the functionality of jNeuroML (including the ability to convert NeuroML to
518 simulator specific code, e.g. for NEURON) through Python scripts, by bundling a binary copy of
519 the library. PyNeuroML has recently added support for importing networks and simulations
520 specified in the SONATA format and converting them to NeuroML. A related package currently
521 under development, NeuroMLite (<https://github.com/NeuroML/NeuroMLlite>) allows compact
522 description of networks and can export the generated structures to SONATA. **Fig. 5E** shows a
523 simulation of 300 integrate and fire cells in SONATA which has been imported by pyNeuroML,
524 converted to NeuroML and executed in the NEURON simulator.

525 PyNN

526 PyNN is a simulator-agnostic Python API for describing network models of point neurons, and
527 simulation experiments with such models (Davison et al. 2009). A reference implementation of

528 the API for the NEURON, NEST and Brian simulators is available
529 (<http://neuralensemble.org/PyNN>), and a number of other simulation tools, including
530 neuromorphic hardware systems, have implemented the API (Brüderle et al., 2011; Rhodes et al.,
531 2018). PyNN models can be converted to and from the NeuroML and SONATA formats with a
532 single function call. **Fig. 5E** illustrates an example where a model in SONATA format was
533 loaded using the PyNN “serialization” module, a simulation was carried out using the PyNN
534 NEST backend, and simulation output was saved in the SONATA format.

535 NetPyNE

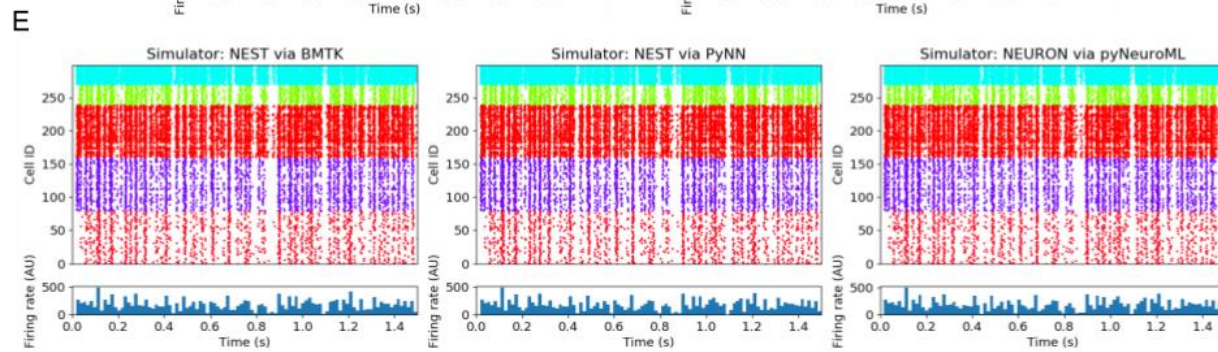
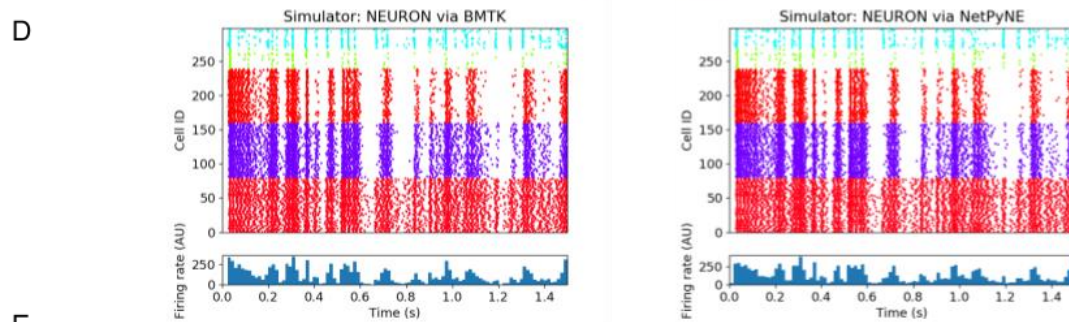
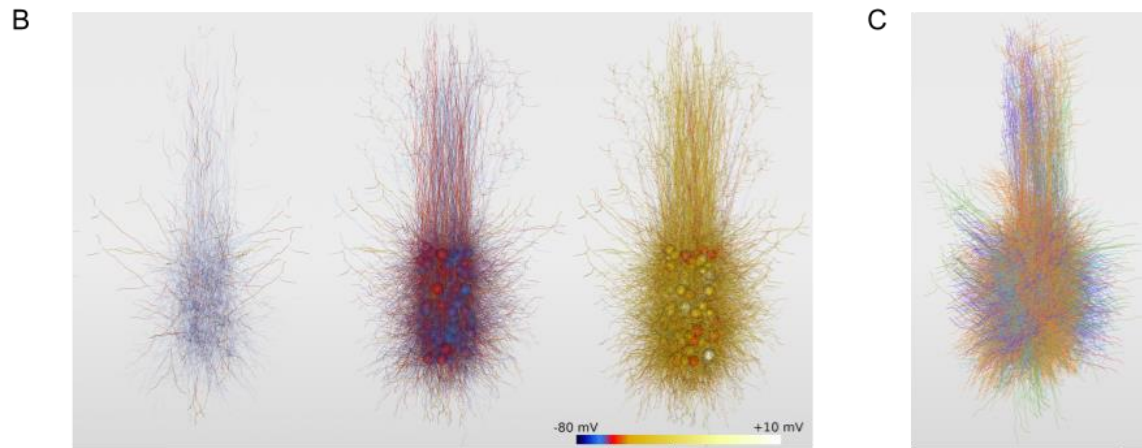
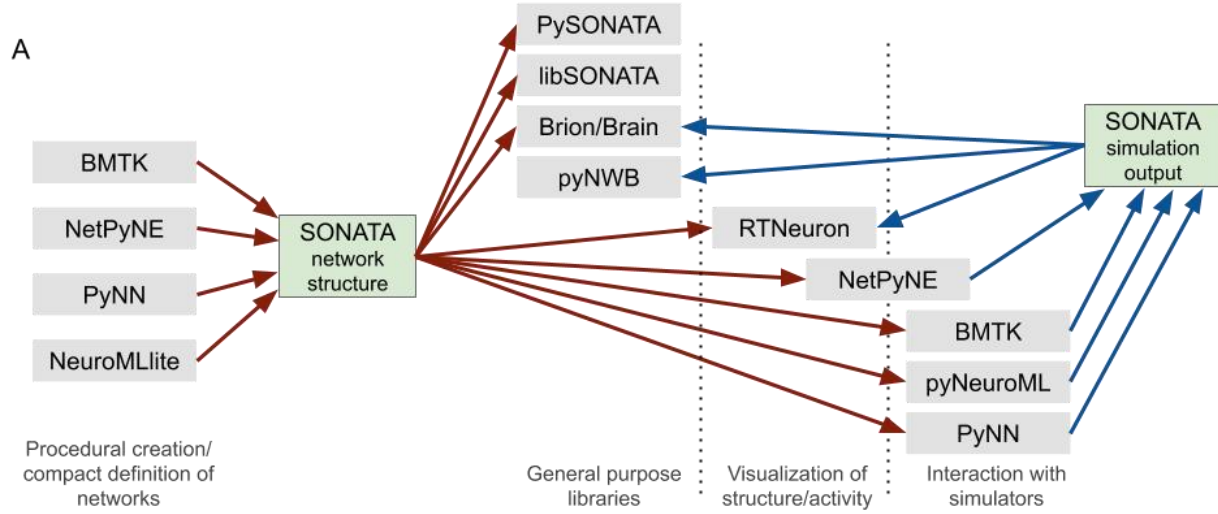
536 NetPyNE (www.netpyne.org; (Dura-Bernal et al., 2019)) is a package developed in Python and
537 building on the NEURON simulator (Carnevale and Hines, 2006). It provides both
538 programmatic and graphical interfaces that facilitate the definition, parallel simulation, and
539 analysis of data-driven multiscale models. Users can provide specifications at a high level via its
540 standardized declarative language. NetPyNE supports both point neurons and biophysically-
541 detailed multi-compartment neurons, as well as NEURON's Reaction-Diffusion (RxD)
542 molecular-level descriptions. The tool includes built-in functions to visualize and analyze the
543 model, including connectivity matrices, voltage traces, raster plot, local field potential (LFP)
544 plots and information transfer measures. Additionally, it facilitates parameter exploration and
545 optimization by automating the submission of batch parallel simulation on multicore machines
546 and supercomputers.

547 NetPyNE network model instantiations can be converted to and from the NeuroML and
548 SONATA formats. SONATA complements NetPyNE by providing a standardized and efficient
549 format to store and exchange large network models. This enables using other simulation tools to
550 run and explore models developed with NetPyNE, and vice versa. As an example, we imported
551 the 300-cell SONATA example with multicompartment cells into NetPyNE, visualized it using
552 the NetPyNE GUI (**Fig. 5C**), and carried out a NetPyNE simulation (**Fig. 5D**).

553 Neurodata Without Borders: Neurophysiology 2.0

554 Neurodata Without Borders: Neurophysiology (NWB:N) 2.0 is a data format for standardizing
555 experimental data across systems neuroscience. We developed an extension for NWB:N 2.0 to
556 accommodate large-scale simulation data, and developed a conversion script from SONATA to
557 NWB:N 2.0 (<https://github.com/ben-dichter-consulting/ndx-simulation-output>) (**Fig. 5A**). This
558 allows stimulated data to be stored side-by-side with experimental data and facilitates comparative
559 analysis between simulation and electrophysiology or calcium imaging experiments.

560



562 **Figure 5. Support for SONATA in simulators and libraries.** (A) Overview of applications
563 which can generate SONATA files (containing either a description of a network structure or
564 simulation output) and the various categories of applications which can read SONATA, including
565 general purpose libraries, visualization tools, and simulation packages. The software packages
566 BMTK, NetPyNE, PyNN, and pyNeuroML can read SONATA network descriptions for
567 execution in the simulation engines NEURON and NEST. The pyNWB package provides a
568 programming interface for reading and writing neurophysiology data (either from experiments or
569 from simulations) in the NWB:N 2.0 format. (B) RTNeuron visualization. Sample renderings at 3
570 simulation timesteps of an example network with 300 biophysically detailed cells, with somatic
571 and dendritic compartments colored according to the simulated membrane potential. The
572 biophysical 300-cell network, as well as its point-neuron counterpart, were created via the model-
573 building scripting interface in BMTK and saved using SONATA. These two models are used in
574 all subsequent panels here. (C) Rendering of the same model as in (B) using the NetPyNE GUI.
575 Each cell is colored according to which of the 5 node types it belongs. (D) The 300-cell
576 biophysically detailed example from (B) and (C) simulated in NEURON using BMTK (left) and
577 NetPyNE (right). (E) A network with 300 integrate and fire neurons generated by BMTK, and
578 simulated in NEST via BMTK (left), NEST after importing the SONATA files into PyNN
579 (middle) and NEURON after conversion of the network to NeuroML by pyNeuroML. Each
580 raster plot in (D) and (E) is accompanied by a panel underneath showing population firing rate
581 (arbitrary units).
582

583 Discussion

584 We have described SONATA, an open-source data format developed to answer the challenges of
585 modern computational neuroscience, especially those inherent in large-scale data-driven
586 modeling of brain networks. It is designed for memory and computational efficiency, as well as
587 for working across multiple platforms, and at the same time enabling as much flexibility as
588 possible for diverse applications. To achieve this, SONATA relies on commonly used data
589 formats such as CSV, HDF5, and JSON, which can be used across platforms, can be read and
590 written by many existing libraries in various programming languages, and (especially in the case
591 of HDF5) have been proven to work efficiently in parallel computations with very large datasets.
592 The SONATA specifications include network descriptions, simulation configuration, and input
593 or output activity. Close cooperation with existing standardisation and simulator independent
594 specification initiatives like NeuroML, PyNN, and NWB:N has helped to increase synergy with
595 existing formats, and has ensured compatibility with languages and tools already in use in the
596 community.

597
598 The flexibility of the SONATA specification is ensured by several design criteria. First, the
599 design leaves it up to users to decide which attributes are shared within node or edge type vs.
600 which are unique to specific nodes or edges. Second, it allows limitless creation of user-defined
601 attributes and maintains only a small number of reserved fields. And third, via a hierarchy of
602 types, populations, and groups of nodes/edges, it permits specification of hybrid models that may

603 include biophysically detailed neurons, point neurons, and many other model types, all in one
604 network model.

605
606 While SONATA offers computationally efficient solutions for storing many model properties,
607 we did not attempt to reinvent file formats for all properties. For example, SONATA utilizes the
608 well established ASCII-based SWC format for neuronal morphologies. We did not develop a
609 computationally optimized binary format for morphologies because their footprint in terms of
610 storage or computational demand is typically small. In the case of the Layer 4 model (**Fig. 4**),
611 loading SWC morphologies takes ~60% of the time of building nodes, but that expense is
612 dwarfed by the time it takes to establish connections (~300 s for external and recurrent
613 connections vs. ~5 s for nodes). Thus, we opted to develop efficient binary solutions only for
614 computationally demanding model properties, otherwise relying on widely used formats such as
615 SWC.

616
617 The SONATA community and ecosystem include multiple groups with diverse interests and are
618 growing due to the open-source design. Initially developed jointly by the Allen Institute and the
619 Blue Brain Project, SONATA is now supported by tools from many teams. As described above,
620 tools such as BMTK (Gratiy et al., 2018), RTNeuron (Hernando et al., 2013), PyNN (Davison et
621 al., 2009), NeuroML (Cannon et al., 2014; Gleeson et al., 2010), and NetPyNE (Dura-Bernal et
622 al., 2019) include SONATA support. Functionality for conversion between SONATA and
623 NWB:N (Ruebel et al., 2019) also exists. The SONATA data format and framework are reflected
624 in the free and open-source PySONATA project hosted on GitHub
625 (<https://github.com/AllenInstitute/sonata>), which is intended as a key resource for those wishing
626 to add support for SONATA to their applications and includes specification documentation,
627 open-source reference application programming interfaces, and model and simulation output
628 examples.

629
630 As an open living format, SONATA may be extended in the future to reflect developments in
631 modeling and in experimental neuroscience. In turn, we invite experimentalist colleagues to
632 explore SONATA's applicability to their circumstances, as the SONATA framework provides an
633 efficient description for a variety of network properties. Such cross-pollination will help
634 improve reproducibility and facilitate collaboration between experimental and computational
635 neuroscientists.

636

637 **Methods**

638 **JSON, CSV, and HDF5**

639 **JSON**

640 JSON (JavaScript Object Notation) is a data exchange format that is easy for both humans and
641 machines to read and write. Being text based, JSON is platform and language independent. Data
642 organization is based on two common structures: key-value pairs and ordered lists, which have
643 equivalents in almost all programming languages.

644 **CSV**

645 CSV stands for “*comma-separated values*” and it is a very common way of laying out tabular
646 data in text files. CSV is not a standard *per se*; the choices that have been made for SONATA are
647 described in the official specification. It should be noted that, although the CSV abbreviation
648 suggests comma as a separator, CSV files can use many types of separator, and, in fact,
649 SONATA format specifies spaces as preferred separators for CSV.

650 **HDF5**

651 HDF5 (Hierarchical Data Format version 5) is a technology designed for storing very large
652 heterogeneous data collections and their metadata in a single container file. HDF5 defines a
653 binary container file format for which the HDF Group provides an implementation in C.
654 Bindings for several other languages exist as well. Basic concepts of HDF5 include groups,
655 datasets and attributes. Making an analogy to filesystems, groups are similar to directories and
656 datasets to files. The main differences between HDF5 and a general purpose filesystem are that
657 a) a dataset is not a stream of bytes like a file, but consists of a multidimensional array with a
658 single data type for all values and that b) groups and datasets can be annotated by means of
659 attributes. HDF5 defines some basic data types common to most programming languages:
660 integers, floats, strings. Data can be stored linearly (the elements of a dataset are stored in
661 increasing order, according to their index and dimension) or in “chunks” for computational
662 efficiency (the order in how dataset elements is interleaved according to their index and
663 dimension; for details, see <https://support.hdfgroup.org/HDF5/doc/Advanced/Chunking/>).

664 **Benchmarking**

665 **Edge file benchmarks**

666 The performance of navigating through an edge file in SONATA format is illustrated in **Fig. 6**,
667 which shows the results of selecting 1000 neurons and accessing one arbitrary property of all the

668 edges of the selected neurons in the 45,000-cell recurrently connected model of Layer 4 of mouse
669 V1(Arkipov et al., 2018) On average each cell receives input from 438.8 neighbors with the
670 number and strength of synapses between any two cells being determined by source and target cell
671 types. The network file contains over 39.2 million unique synapses partitioned into two groups,
672 those synapses that target multicompartment neurons and those that target point points.
673 Connections that target point neurons only require synaptic strength variable, while those that
674 target multi-compartment neurons also require information about section number and segment
675 distance for each synapse. The HDF5 edge file is 1.9 GB in size.

676 The benchmarks were conducted on an HPE SGI 8600 supercomputer. Each compute node had
677 two Intel Xeon Gold 6140 CPUs (each with 18 cores at 2.30 GHz) and 768 GB of DRAM. Nodes
678 were connected through a Mellanox Infiniband (IB) EDR fabric to two GS14K storage racks
679 with a total storage capacity of 4 PB. The computing system was running Linux 3.10.0 and the
680 filesystem was GPFS 4.2.3-6, configured with 4 MiB block size. The storage system did not have
681 dedicated metadata drivers. The software components used and their versions are the following:
682 glibc 2.25-49, gcc 6.4, boost 1.58, HDF5 1.10.1, Python 2.7, numpy 1.13.3 and MPI 2.16
683 provided by HPE.

684
685 For reference, the maximum average read bandwidth obtained in pure I/O benchmark
686 experiments with IOR (<https://ior.readthedocs.io/en/latest/>) in this machine is 5.6 GiB/s using 1
687 single core accessing a 1 GiB file in 4 MiB blocks. The maximum average write bandwidth
688 measured is 9.5 GiB/s using 8 cores from 1 node writing 1 GiB per core in 4 MiB operations to a
689 shared file. POSIX I/O was used to obtain both measurements.

690
691 To illustrate SONATA's performance and flexibility, we use examples of ordering the edges data
692 in two different ways (**Fig. 6A**): target-major (**Fig. 6B**), where data is sorted according to the ID
693 of the target neuron (increasing), and hybrid ordering (**Fig. 6C**), where the connectivity matrix is
694 divided in blocks, and edges inside each block are enumerated, alternating (from block to block)
695 between source-major and target major orderings. We also compare the impact of selecting 1000
696 neurons randomly or sequentially.

697
698 Note that SONATA supports arbitrary ordering of edges, and the two variants tested in the
699 benchmarks are only for demonstration purposes.

700
701 A target-major sort is more efficient for instance in the case of a simulator creating the synapses
702 on the target cell when instantiating the network. A source-major sort (data sorted according to
703 the ID of the source neuron increasing) is favorable to analysis of efferent connectivity of large
704 network. The hybrid ordering is a compromise between the target-major and source-major
705 ordering.

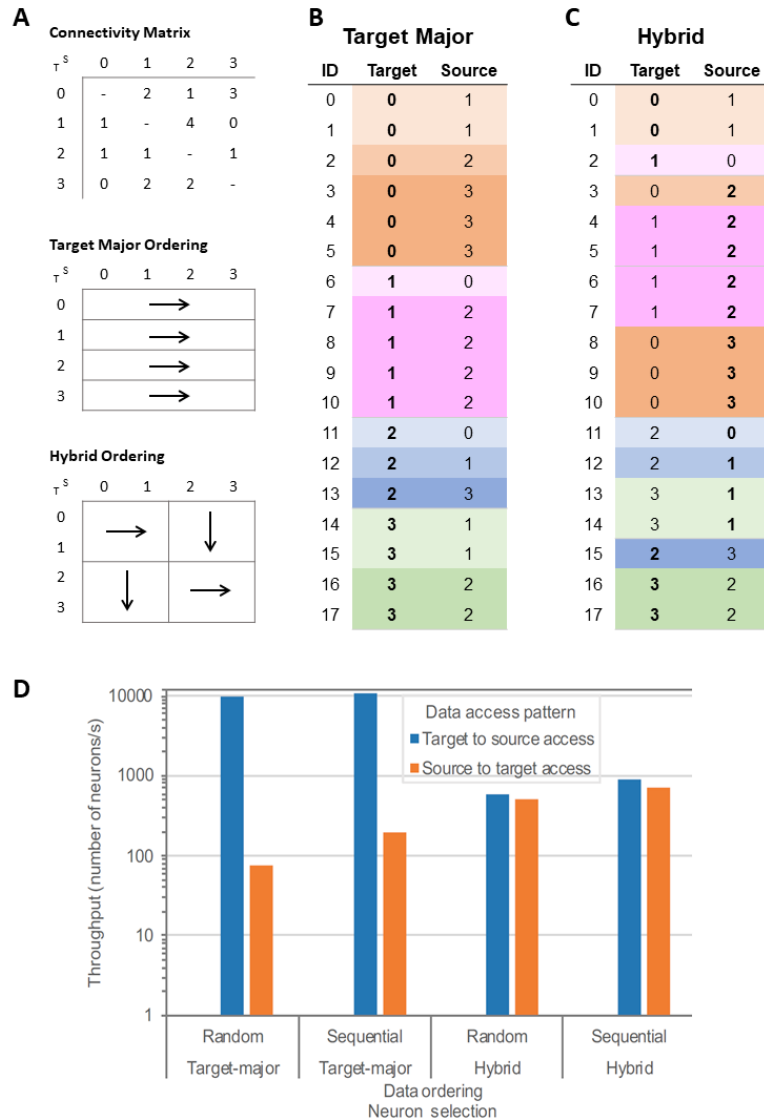
706

707 **Fig. 6D** shows that ordering has an impact on the performance of data access (whereas selecting
708 neurons randomly or sequentially does not impact performance substantially). By using target-
709 major ordering (or its symmetric source-major ordering) one can achieve optimal performance
710 when accessing data in the same access pattern as the ordering, but accessing data in the opposite
711 direction is much less efficient, by a factor of ~100. Ordering data in a hybrid manner is a
712 compromise to get balanced performance between the source-to-target and target-to-source
713 access patterns, but in this case the performance is not as good as the optimal performance for
714 non-hybrid ordering. Due to such large discrepancies, the SONATA format specification leaves
715 the choice of ordering open to users. Note that source-target pairs for each edge are always
716 defined in the edge files in the same way; it is the indexing of these edges that may differ
717 depending on user requirements. This means that the edges can always be read, but reading speed
718 for a particular application will depend on the choice of indexing, and this choice should be made
719 based on the desired application. Examples in **Fig. 6D** indicate that a rather high performance can
720 be achieved (close to 10,000 neurons processed per second for their edge attributes) in optimal
721 cases, but users should take advantage of the flexibility of SONATA specification to use edge
722 ordering that is most suitable for their needs. In situations where high performance for various
723 access patterns is essential, solutions may include two or more copies of edge files with different
724 orderings for different use cases.

725

726

727



728
 729 **Figure 6. Target major and hybrid ordering of edges.** (A) A simple example of connectivity
 730 matrix (the number within each matrix element indicates the number of edges -- i.e., synapses --
 731 between the two nodes) and schematics of target major and hybrid orderings. (B) and (C) Edge
 732 lists representing edges from the connectivity matrix in (A), sorted according to target major (B)
 733 or hybrid (C) ordering. (D) Throughput of accessing edge information for target major or hybrid
 734 ordering of edges in the SONATA files in a 45,000-cell model of Layer 4 of mouse V1
 735 (Arkhipov et al., 2018). The target-to-source and source-to-target access patterns are illustrated
 736 with either random or sequential selection of target or source neurons.

737 Simulation output benchmarks

738 The simulation output benchmarks (**Fig. 3**) were run on the aforementioned HPE SGI 8600
 739 system. Since most simulators can run in parallel (multi-thread and/or multi-process), the
 740 benchmarking of the report generation was also done in parallel, on 16 nodes and 36 processes

741 per node (using 1 core per process). All processes were periodically dumping data to a single,
742 shared HDF5 file in the SONATA format. At each write operation, each process was writing
743 several columns at its designated frame/trace region. The amount of data written at each
744 operation is presented as the “Write block size per process” illustrated in the performance plots
745 (the write block size applies for each process and for each write operation).

746
747 Write benchmarks made use of the Neuromapp library
748 (<https://github.com/BlueBrain/neuromapp>, revision f03d3ea) (Ewart et al., 2017), which uses
749 parallel HDF5 and MPI underneath. Read benchmarks were implemented using the Python
750 binding of Brion/Brain (revision c16a694), the testing and plotting code can be found in the
751 SONATA github repository in the benchmarks branch.

752 Loading of simulation data

753 Benchmarks for loading simulation data (**Fig. 4C**) were obtained for the full simulation of the
754 45,000-neuron recurrently connected model of Layer 4 of mouse V1 (Arkhipov et al., 2018).
755 Figure **Fig. 4C** shows the amount of time required to parse through the SONATA network files
756 and instantiate the in-memory cell and synaptic objects to run a full NEURON (Carnevale and
757 Hines, 2006) simulation. Each simulation was instantiated with a computing cluster of Intel Xeon
758 E5 processors (each core either 2.1 or 2.2 GHz), using a minimum of 5 cores and a maximum of
759 390 cores. The network was built using the Brain Modeling Toolkit with Python 3.6 and
760 NEURON 7.5 with Python bindings.

761

762 Acknowledgements

763 This project/research has received funding from the European Union’s Horizon 2020 Framework
764 Programme for Research and Innovation under the Specific Grant Agreement No. 785907
765 (Human Brain Project SGA2) and from the EPFL Blue Brain Project (funded by the Swiss
766 government’s ETH Board of the Swiss Federal Institutes of Technology). SD-B was funded by
767 NIH grants U01EB017695, R01EB022903 and 2R01DC012947-06A1, and New York State
768 grant DOH01-C32250GG-3450000. PG was supported by the Wellcome Trust (212941,
769 101445). This work benefited from interactions between the authors as part of the International
770 Neuroinformatics Coordinating Facility (INCF) Special Interest Group on Standardised
771 Representations of Network Structures. We are grateful to Christof Koch for inspiring and
772 supporting this work and to Michael Hines for many helpful discussions and suggestions. We
773 wish to thank the Allen Institute founder, Paul G. Allen, for his vision, encouragement, and
774 support.

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