1	
2	A platform for semi-automated voluntary training of common marmosets for behavioral
3	neuroscience
4	
5 6 7	Voluntary training of common marmosets
8	Jeffrey D. Walker ^{1,2*} , Friederice Pirschel ² , Nicholas Gidmark ³ , Jason N. MacLean ^{1,4} , Nicholas
9	G. Hatsopoulos ^{1,2}
10	¹ Committee on Computational Neuroscience, University of Chicago, Chicago, USA
11	² Department of Organismal Biology and Anatomy, University of Chicago, USA
12	³ Biology Department, Knox College, USA 61401
13	⁴ Department of Neurobiology, University of Chicago, Chicago, USA 60637
14	*Corresponding author: walkerjd@uchicago.edu
15	
16	Number of pages: 19
17	Number of figures: 5
18	Number of words for abstract: 123
19	Number of words for introduction: 521
20	Number of words for discussion: 408
21	
22	Conflict of interest: The authors declare no competing financial interests.
23	
24	Acknowledgements: We thank Marek Niekrasz and the veterinary staff at the University of
25	Chicago for their assistance with marmoset care and Callum Ross for assistance with the
26	XROMM system. This work was supported by NIH R01 NS104898, NSF MRI 1338066, NSF
27	IGERT, UChicago Big Vision Fund and the Tarrson Fund.
28	

29 ABSTRACT

30 In most cases, behavioral neuroscience studies of the common marmoset employ adaptations of 31 well-established methods used with macaque monkeys. However, in most cases these approaches 32 do not readily generalize to marmosets indicating a need for alternatives. Here we present the 33 development of one such alternate: a platform for semi-automated, voluntary in-home cage 34 behavioral training that allows for the study of naturalistic behaviors. We describe the design and 35 production of a modular behavioral training apparatus using CAD software and digital 36 fabrication. We demonstrate that this apparatus permits voluntary behavioral training and data 37 collection throughout the marmoset's waking hours with little experimenter intervention. 38 Further we demonstrate the use of this apparatus to reconstruct the kinematics of the marmoset's 39 upper limb movement during natural foraging behavior.

40

41 NEW AND NOTEWORTHY

42 The study of marmosets in neuroscience has grown rapidly and this model organism presents 43 challenges that are unique to this primate species. Here we address those challenges with an 44 innovative platform for semi-automated and voluntary training of common marmosets. The 45 platform allows marmosets to train throughout their waking hours with little to no experimenter 46 intervention. We describe the use of this platform to capture the kinematics of the upper limb 47 during natural foraging behavior and to expand the opportunities for behavioral training beyond 48 the limits of traditional behavioral training sessions. The platform is flexible and can be easily 49 extended to incorporate other motor tasks (e.g. visually cued reaching or manipulandum based 50 tasks) using CAD models and digital fabrication.

51 **INTRODUCTION**

52 Neurophysiological recordings of isolated single neurons in awake, behaving macaques began in 53 the late 1960's, whereas the first reports of single neuron recordings from awake marmosets did 54 not occur until the early 2000s (Evarts, 1968; Lu et al., 2001). Despite the relative recency of 55 broad adoption of the marmoset as a model species for systems neuroscience there is growing 56 interest, but the techniques for working with marmosets in this context are relatively new as 57 compared to those used with more standard model primate species (e.g. rhesus macaques) in 58 neuroscience research. Because of the success of the model, the approach to training a macaque 59 to perform an experimental task has remained, with few exceptions, relatively unchanged for 60 decades. In general, the monkey is restrained while engaging in a trained task for a few hours in exchange for water or juice. This method is popular because it generally yields hundreds to 61 62 thousands of repetitions of a given behavior over the course of a training session. However, our 63 experience and the early behavioral work indicate that this approach may be ill-suited for 64 working with marmosets. It yields far fewer trials and limits the expression of natural behavior 65 (Johnston et al. 2017; Prins et al., 2017; Eliades and Wang, 2003). To partly address these issues, 66 Wang and colleagues developed a technique for wireless neural recordings which allowed for the 67 study of sensorimotor processing in freely vocalizing marmosets (Roy and Wang, 2012). 68 However, there has not been a complimentary innovation in behavioral training paradigms to 69 increase trial counts.

70 Marmoset ethology and its implications for experimental design

Marmosets are obligate gum feeders and prey species. Field studies estimate that
marmosets spend about 30 percent of their waking hours feeding on exudates (Maier et al., 1982
in Sussman and Kinzey 1984) and spend 25-30% of their waking time foraging for insects

74 (Abreu et al., 2016; Stevenson and Rylands, 1988). In order to feed on exudates, marmosets must 75 gouge wounds into the trunks of trees to access the gum. They gouge new holes and revisit 76 previously gouged holes to feed on newly accumulated gum (Stevenson and Rylands, 1988). 77 These visits only last a few seconds (Stevenson and Rylands, 1988). Their daily behavioral repertoire generally does not involve them sitting in a single place engaging in repetitive 78 79 behaviors for multiple hours. With this in mind, we designed an approach to training marmosets 80 that would allow them to voluntarily engage in experimental behavior for short sessions 81 throughout their waking hours. In order to do so we sought to modify an approach successful 82 applied to rodents where rats voluntarily head-fixed themselves for in vivo calcium imaging 83 (Scott, Brody, and Tank 2013). To implement this approach, researchers designed a set of 84 custom elements to ensure stable imaging and slowly acclimated the rat to the apparatus, 85 gradually extending the duration of head fixation. Once the animal was trained, the process of 86 data collection could proceed with minimal experimenter involvement. This sort of voluntary 87 setup, that allowed the animal to engage in the experiment throughout the day as an expression 88 of its normal behavioral repertoire, seemed like a promising approach to behavioral training of 89 marmosets.

90 MATERIALS AND METHODS

91 Subjects

All work described were done with three common marmosets (Callithrix jacchus) (two females,
and one male, 375-410 g). All methods were approved by the Institutional Animal Care and Use
Committee of the University of Chicago.

95 Design criteria

96 Informed by field studies of the marmoset's natural behavioral repertoire (Stevenson and 97 Rylands, 1988; Sussman and Kinzey, 1984), early work with marmosets in neuroscience (Eliades 98 and Wang, 2003, 2005, 2008a), and the novel approach to training and in-vivo calcium imaging 99 developed by Scott, Brody, and Tank (2013), we developed a behavioral training apparatus that 100 attaches to the marmosets' home cage. This apparatus allows marmosets to voluntarily engage in 101 behavioral training throughout their waking hours.

102 The three primary design criteria for the final apparatus were 1) that it mounts to the 103 home cage to allow for voluntary engagement in training throughout the marmosets' waking 104 hours, 2) that it provides reliable positioning of marmosets and clear views of the upper limbs for 105 capturing the kinematics of reaching movements, and 3) it provides a flexible way to present 106 different experimental tasks. Additionally, to validate the effectiveness of the apparatus as a 107 training instrument, it had to have a way to monitor and record the marmosets' behavior within 108 it. Finally, to facilitate training using operant conditioning, the apparatus also had to include a 109 method for precisely timed reward delivery.

110 Hardware design and iteration

Inspired by the gum feeding behavior in which marmosets naturally engage, the first version of the apparatus trained the marmosets to assume the appropriate posture to receive a small volume of yogurt (Figure 1A). This posture placed them in front of a tray that contained foraging substrate. The next version of the apparatus removed the yogurt reward, and we found that marmosets would still engage in foraging behavior within the apparatus. After a series of iterations optimizing the form of the apparatus to multiple motion capture modalities, the current

117	version of the apparatus (Figure 1B) has allowed us to record the kinematics of this foraging
118	behavior to study sensorimotor cortical responses related to upper limb movement.
119	Designs of early versions of the behavioral training apparatus were done with 3D CAD
120	software called SketchUp, while later versions were designed using AutoDesk Fusion 360
121	(Figure 1). The core of the apparatus was constructed using 1/8" or 1/4" thick clear acrylic
122	sheets (continuous cast, McMaster Carr, Elmhurst, IL) that were cut into interlocking panels
123	using a laser cutter (Universal Laser Systems VLS4.60). These panels were then assembled to
124	achieve the form of the apparatus. To monitor the activity of the marmosets within the
125	apparatus, we designed a simple circuit (Figure 2A-B) that included two photocells (CdS -
126	photoresistor) and one infrared light based switch (IR switch comprising an IR phototransistor
127	and IR LED pair), a syringe pump (syringepump.com, NE-500) and a network-connected
128	microcontroller (Arduino YÚN). The sensors acted as triggers to log the marmosets entering and
129	leaving the gate, the belt, and the nosepiece of the apparatus. The sensor readings were logged to
130	an SD card within the microcontroller, and the network connection of the microcontroller
131	allowed remote operation of the apparatus. The apparatus sat on top of a round gate installed in
132	ceiling of the home cage (part # 1822K314, McMaster-Carr).

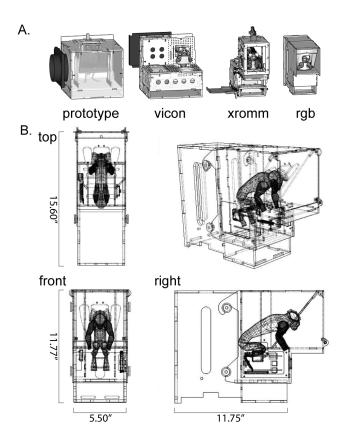


Figure 1. Developing a voluntary in-home cage approach to behavioral training with marmosets. A) Iterations of apparatus design optimized for different motion capture modalities. B) Drawing of current version of the behavioral training apparatus.

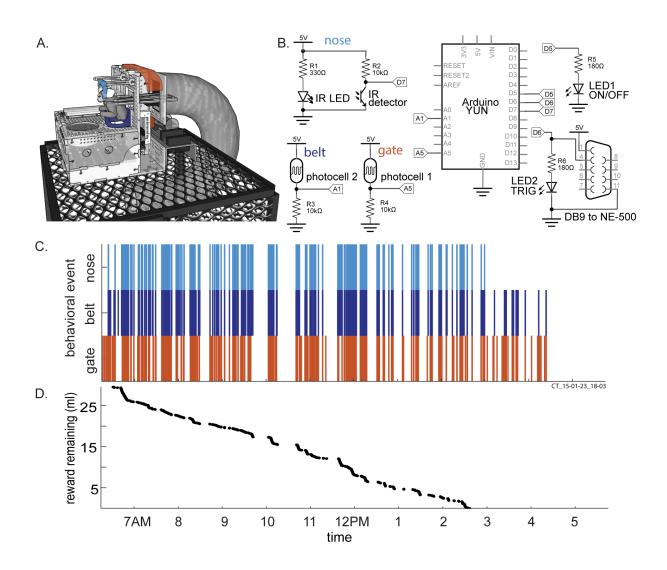


Figure 2. Hardware design and single day of behavior within the apparatus. A) Illustration of behavioral training apparatus with sensors embedded into the gate (orange), belt (dark blue) and nosepiece (light blue) to log behavior throughout the day. The foraging tray is placed in front of the belt. A syringe pump is connected to deliver reward. The whole assembly sits on top of the home cage. B) Circuit diagram detailing the circuit logging behavior and delivering reward. C and D) Results for single day of behavior within the apparatus. C) Vertical ticks indicate the time of trigger events for sensors within the gate, belt and nosepiece. For instance, an orange tick indicates the marmoset crossed the gate of the apparatus, a dark blue tick indicates the marmoset is within the nosepiece. When the marmoset stays within the nosepiece, 0.1 ml of yogurt is dispensed every 10 seconds as positive reinforcement for assuming the appropriate posture. D) Reward remaining as a function of time of day.

135 Software for automating and monitoring training

We wrote a library (C++) to coordinate logging activity within the apparatus, evaluate rewardconditions, deliver reward, and allow remote apparatus operation. The object-oriented design of
this library is meant to facilitate integration of future experimental tasks.

139

140 **RESULTS**

141 Foraging

142 We began by studying foraging since this was a behavior in which the marmosets readily 143 engaged. Foraging was coupled with the task of assuming an appropriate posture in exchange for 144 vogurt reward. Marmosets engaged in behavior within the training apparatus throughout the day. 145 and their engagement was sensitive to reward availability (Figure 2C-D). We measured each 146 time a marmoset entered and exited the belt of the apparatus, i.e. the start and end of a session, to 147 quantify the duration of these sessions. This measure allowed us to generate an estimate of how 148 much time marmosets would spend engaging in behavior within the apparatus and how that 149 behavior was distributed throughout their waking hours. Over the course multiple days, we 150 found that marmosets would spend up to an hour each day engaging in behavior within the 151 apparatus spanning 65 - 216 sessions (Figure 3A-B). Most of these sessions were not longer 152 than 20 seconds, but some lasted almost five minutes (n=1739 sessions over 11 days, mean = 153 17.00 sec, median = 9.36 sec, $Q_1 = 6.99$ sec, $Q_3 = 16.67$, max = 270 sec) (Figure 3C-D). 154

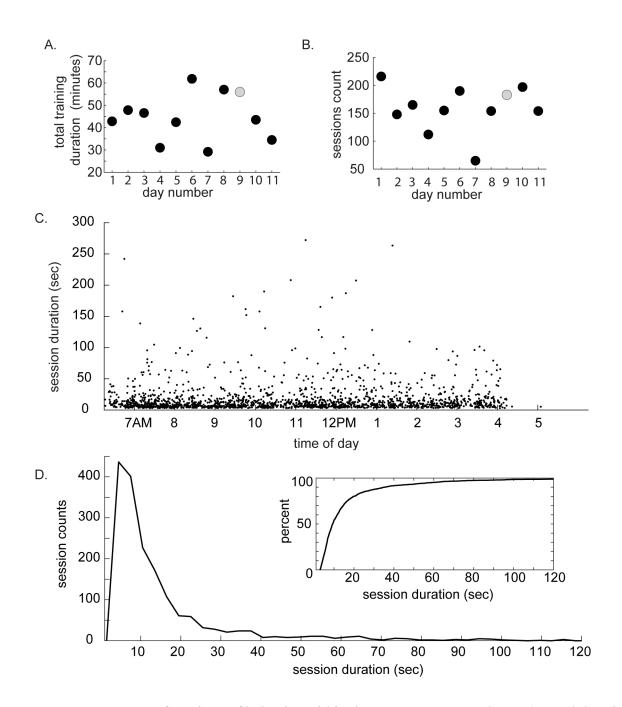


Figure 3. Summary of sessions of behavior within the apparatus across days. A) Total duration of all sessions within each day. B) Number of sessions of behavior within each day. Grey circle indicates data point corresponding to day illustrated in Figure 2. C) Session durations as a function of time of day. Data were pooled across all days. Each point represents a single session. D) Distribution of session durations. Inset: cumulative distribution of session durations.

156 After validating that marmosets engage in voluntary behavioral training and gaining a 157 sense of their attention span, we optimized the design of the apparatus to provide unobscured 158 views of the upper limb (Figure 1) and set out to characterize foraging behavior within the 159 apparatus. Using a custom-written algorithm (MATLAB) to define the video frame when the 160 animal started foraging, reaches were subsequently counted manually. Reaches with both hands 161 were counted over the course of a day (12 hours). When only the foraging mix was provided in 162 the apparatus, marmosets performed between 20 and 80 reaches while foraging each day (Figure 163 4A). In contrast, if their entire daily diet (i.e. foraging mix and normal diet) was provided within 164 the behavioral training apparatus, the marmosets performed 100-300 reaches per day (Figure 165 4B).

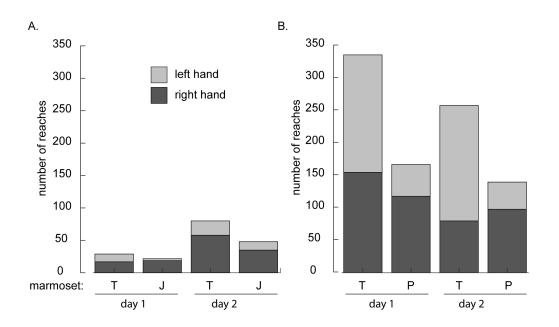


Figure 4. Reaches counted during foraging behavior within the apparatus. A) Counts of reaches for two marmosets (T and J) across two days when only foraging mix was provided in the apparatus. B) Counts of reaches for two marmosets (T and P) across two days when their entire daily diet was provided within the training apparatus.

167 *Recording upper limb kinematics during foraging with XROMM*

168 We next sought to record the kinematics of the upper limb during foraging. After

169 confirming that marmosets do not tolerate retro-reflective markers placed on their skin needed

- 170 for traditional near infrared based motion capture systems (e.g. VICON) (Takemi et al., 2014;
- 171 Young et al., 2016) we moved to using an x-ray based system called XROMM, or X-Ray
- 172 Reconstruction of Moving Morphology (Brainerd et al. 2010) (Figure 5A). Bi-planar x-ray

173 sources and image intensifiers (90 kV, 25 mA at 200fps) allowed us to reconstruct time varying

- 174 joint angles by tracking the 3D position of radio-opaque tantalum beads (0.5-1 mm, Bal-tec)
- placed within the soft tissue of the arm, hand and torso (Figure 5B). Using a set of tools
- developed at Brown University (Brainerd et al., 2010; Knörlein et al., 2016; Miranda et al.,

- 177 2011), and adaptations of joint coordinate systems for the upper limb (Baier and Gatesy, 2013;
- 178 Wu et al., 2005), we could translate the position of these markers into joint kinematics (Figure
- 179 5). We placed markers in the torso and upper limb subcutaneously using angiocatheters (16G,
- 180 Becton, Dickinson and Company) (Figure 5B). The marker set illustrated allowed reconstruction
- 181 of the seven degrees of freedom of the shoulder, elbow and wrist (Figure 5C).

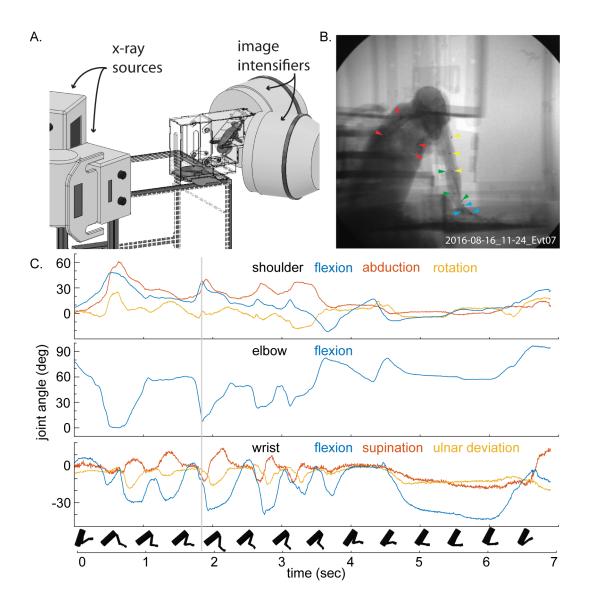


Figure 5. Capturing the kinematics of upper limb during foraging with XROMM. A) An illustration of the XROMM bi-planar x-ray motion capture system together with the behavioral training apparatus and marmoset in the capture volume. B) A single frame of x-ray video of a marmoset foraging within the apparatus. Note radio-opaque markers placed within the marmoset's torso (red), upper arm (yellow), forearm (green) and hand (blue). C) Seven degrees of freedom of upper limb movement reconstructed by tracking the movement of the radio-opaque markers seen in B). Grey line indicates timestamp of the frame in B). Rigid bodies represent kinematics of the torso, upper arm, forearm and hand over the course of the foraging sequence.

182 **DISCUSSION**

183 Here we present a method for in home-cage, semi-automated, and voluntary behavioral 184 training of marmosets that has in our experience been more prolific than adaptations of the more 185 traditional approaches. The method presented also allows for the training of multiple marmosets 186 in parallel. It provides a flexible platform for a variety of experimental tasks and liberates the 187 animals from excessive restraints and provides a platform for marmosets to self-initiate natural behavior in addition to engaging in more traditional operant paradigms. It allows for behavioral 188 189 engagement in short sessions throughout the marmosets waking hours rather than extended 190 sessions, which are limited by marmoset cooperation and satiation. This flexible approach should 191 allow us to contextualize results from constrained and over-trained experimental tasks within the 192 space of the marmoset's natural behavioral repertoire. Toward this end, we are in the process of 193 implementing an additional motor learning task and we are pairing this training approach with 194 wireless neural recordings.

195 It is clear that marmosets are well poised to contribute to our understanding of the 196 operating principles of neocortex as attested by their increasing prevalence in published systems 197 neuroscience reports (Miller, 2017). Moreover the structure of marmoset neocortex provides a 198 strong potential for targeted circuit manipulations (Belmonte et al., 2015; Sasaki et al., 2009). 199 Marmosets have been trained to perform experimental tasks such as eye fixation and a smooth 200 pursuit (Mitchell, Reynolds, and Miller 2014; Mitchell, Priebe, and Miller 2015) and basic 201 reaching and neuroprosthetic tasks (Ebina et al., 2018; Pohlmeyer et al. 2012, 2014) using 202 training procedures common in macaque studies. But the quantity of behavior marmosets 203 produce using these procedures is generally limited in comparison to that of macaques. In 204 contrast, the techniques we designed dramatically increased the time available for behavioral 205 training by eliminating the use of restraint and making the experimental training apparatus

available to the marmosets throughout their waking hours. With this paradigm, our initial

- 207 estimates suggest that we can minimally double and can often quadruple the quantity of
- 208 experimentally useful behavioral trials with the added benefit that this behavior is self-initiated
- 209 rather than generated through restriction. We would like to see if, with adaptations such as
- support for reliable eye positioning, this behavioral training approach could be useful to increase
- 211 the behavioral output of marmosets in studies of other systems. Finally, as we have argued
- 212 (Walker et al., 2017), natural behaviors in of themselves warrant study and this particular
- 213 experimental behavioral paradigm facilitates this class of study.
- 214

215 CODE AND DESIGN FILE ACCESSIBILITY

216 Both software written to coordinate training and design files used to fabricate the apparatus are

217 available from the authors upon request.

219

REFERENCES

- Abreu, F., De la Fuente, M.F.C., Schiel, N., and Souto, A. (2016). Feeding ecology and
- behavioral adjustments: flexibility of a small neotropical primate (Callithrix jacchus) to survive in a semiarid environment. Mammal Res. *61*, 221–229.
- Baier, D.B., and Gatesy, S.M. (2013). Three-dimensional skeletal kinematics of the shoulder girdle and forelimb in walking *Alligator*. J. Anat. 223, 462-473.
- 225 Belmonte, J.C.I., Callaway, E.M., Caddick, S.J., Churchland, P., Feng, G., Homanics, G.E., Lee,
- K.-F., Leopold, D.A., Miller, C.T., Mitchell, J.F., et al. (2015). Brains, Genes, and Primates.
 Neuron *86*, 617–631.
- 228 Brainerd, E.L., Baier, D.B., Gatesy, S.M., Hedrick, T.L., Metzger, K.A., Gilbert, S.L., and
- 229 Crisco, J.J. (2010). X-ray reconstruction of moving morphology (XROMM): precision, accuracy
- and applications in comparative biomechanics research. J. Exp. Zool. Part Ecol. Genet. Physiol.
- *313*, 262–279.
- Ebina, T., Masamizu, Y., Tanaka, Y.R., Watakabe, A., Hirakawa, R., Hirayama, Y., Hira, R.,
- 233 Terada, S.-I., Koketsu, D., Hikosaka, K., et al. (2018). Two-photon imaging of neuronal activity
- in motor cortex of marmosets during upper-limb movement tasks. Nat. Commun. 9.
- Eliades, S.J., and Wang, X. (2003). Sensory-Motor Interaction in the Primate Auditory Cortex
 During Self-Initiated Vocalizations. J. Neurophysiol. *89*, 2194–2207.
- Eliades, S.J., and Wang, X. (2005). Dynamics of Auditory–Vocal Interaction in Monkey
 Auditory Cortex. Cereb. Cortex *15*, 1510–1523.
- Eliades, S.J., and Wang, X. (2008a). Chronic multi-electrode neural recording in free-roaming
 monkeys. J. Neurosci. Methods *172*, 201–214.
- Eliades, S.J., and Wang, X. (2008b). Neural substrates of vocalization feedback monitoring in
 primate auditory cortex. Nature 453, 1102–1106.
- Evarts, E.V. (1968). Relation of Pyramidal Tract Activity to Force Exerted During Voluntary
 Movement. J Neurophysiol 1968 Jan *31*, 14–27.
- Johnston, K.D., Barker, K., Schaeffer, L., Schaeffer, D., and Everling, S. Methods for chair
 restraint and training of the common marmoset on oculomotor tasks. J Neurophysiol 11.
- 247 Knörlein, B.J., Baier, D.B., Gatesy, S.M., Laurence-Chasen, J.D., and Brainerd, E.L. (2016).
- 248 Validation of XMALab software for marker-based XROMM. J. Exp. Biol. 219, 3701–3711.
- Lu, T., Liang, L., and Wang, X. (2001). Neural representations of temporally asymmetric stimuli in the auditory cortex of awake primates. J. Neurophysiol. *85*, 2364–2380.

- 251 Maier, W., Alonso, C., and Langguth, A. (1982). Field observations on Callithrix jacchus. Z
- 252 Saugetier-Kunde 334–346.
- 253 Miller, C.T. (2017). Why marmosets?: Editorial. Dev. Neurobiol. 77, 237–243.
- 254 Miranda, D.L., Schwartz, J.B., Loomis, A.C., Brainerd, E.L., Fleming, B.C., and Crisco, J.J.
- 255 (2011). Static and dynamic error of a biplanar videoradiography system using marker-based and
- 256 markerless tracking techniques. J. Biomech. Eng. 133, 121002.
- Mitchell, J.F., Reynolds, J.H., and Miller, C.T. (2014). Active Vision in Marmosets: A Model System for Visual Neuroscience. J. Neurosci. *34*, 1183–1194.
- Mitchell, J.F., Priebe, N.J., and Miller, C.T. (2015). Motion dependence of smooth pursuit eye movements in the marmoset. J. Neurophysiol. *113*, 3954–3960.
- 261 Pohlmeyer, E.A., Mahmoudi, B., Geng, S., Prins, N., and Sanchez, J.C. (2012). Brain-machine
- 262 interface control of a robot arm using actor-critic rainforcement learning. In Engineering in
- 263 Medicine and Biology Society (EMBC), 2012 Annual International Conference of the IEEE, pp.
- **264 4108–4111**.
- 265 Pohlmeyer, E.A., Mahmoudi, B., Geng, S., Prins, N.W., and Sanchez, J.C. (2014). Using
- Reinforcement Learning to Provide Stable Brain-Machine Interface Control Despite Neural Input
 Reorganization. PLoS ONE 9, e87253.
- 268 Prins, N.W., Pohlmeyer, E.A., Debnath, S., Mylavarapu, R., Geng, S., Sanchez, J.C., Rothen, D.,
- and Prasad, A. (2017). Common marmoset (Callithrix jacchus) as a primate model for
 behavioral neuroscience studies. J. Neurosci. Methods 284, 35–46.
- Roy, S., and Wang, X. (2012). Wireless multi-channel single unit recording in freely moving and
 vocalizing primates. J. Neurosci. Methods 203, 28–40.
- 273 Sasaki, E., Suemizu, H., Shimada, A., Hanazawa, K., Oiwa, R., Kamioka, M., Tomioka, I.,
- Sotomaru, Y., Hirakawa, R., Eto, T., et al. (2009). Generation of transgenic non-human primates
 with germline transmission. Nature 459, 523–527.
- Scott, B.B., Brody, C.D., and Tank, D.W. (2013). Cellular Resolution Functional Imaging in
 Behaving Rats Using Voluntary Head Restraint. Neuron *80*, 371–384.
- Stevenson, M.F., and Rylands, A., B. (1988). The Marmosets, Genus Callithrix. In Ecology and
 Behavior of Neotropical Primates, (Washington, D.C.: World Wildlife Fund), pp. 131–222.
- Sussman, R.W., and Kinzey, W.G. (1984). The ecological role of the Callitrichidae: a review.
- 281 Am. J. Phys. Anthropol. *64*, 419–449.
- 282 Takemi, M., Kondo, T., Yoshino-Saito, K., Sekiguchi, T., Kosugi, A., Kasuga, S., Okano, H.J.,
- 283 Okano, H., and Ushiba, J. (2014). Three-dimensional motion analysis of arm-reaching
- movements in healthy and hemispinalized common marmosets. Behav. Brain Res. 275, 259–268.

- 285 Walker, J., MacLean, J., and Hatsopoulos, N.G. (2017). The marmoset as a model system for
- studying voluntary motor control: Studying Motor Control with Marmosets. Dev. Neurobiol. 77, 272–285
- 287 273–285.
- 288 Wu, G., van der Helm, F.C.T., (DirkJan) Veeger, H.E.J., Makhsous, M., Van Roy, P., Anglin, C.,
- 289 Nagels, J., Karduna, A.R., McQuade, K., Wang, X., et al. (2005). ISB recommendation on
- 290 definitions of joint coordinate systems of various joints for the reporting of human joint
- 291 motion—Part II: shoulder, elbow, wrist and hand. J. Biomech. 38, 981–992.
- 292 Young, J.W., Stricklen, B.M., and Chadwell, B.A. (2016). Effects of support diameter and
- compliance on common marmoset (*Callithrix jacchus*) gait kinematics. J. Exp. Biol. 219, 2659–
 2672.