

1 **Identification of microRNA-target interactions for antioxidant defense response under**
2 **drought stress by high-throughput sequencing in *Zanthoxylum bungeanum* Maxim.**

3 Xitong Fei^{1,2}, Haichao Hu^{1,2}, Jingmiao Li^{1,2}, Yulin Liu^{1,2}, Anzhi Wei^{1,2*}

4
5 Xitong Fei^{1,2}, Haichao Hu^{1,2}, Jingmiao Li^{1,2}, Yulin Liu^{1,2}, Anzhi Wei^{1,2*}

6
7 ¹ College of Forestry, Northwest A&F University, Yangling, Shaanxi, China;

8 ² Research Centre for Engineering and Technology of *Zanthoxylum* State Forestry
9 Administration, Yangling, Xianyang 712100, China;

10 * Correspondence: weianzhi@nwafu.edu.cn; Tel.: +86-29-8708-2211

11
12 **ABSTRACT**

13 When the plant is in an unfavorable environment such as drought or high temperature, it will
14 accumulate a large amount of active oxygen, which will seriously affect the normal growth and
15 development of the plant. The antioxidant system can remove the reactive oxygen species
16 produced under drought conditions and so mitigate oxidative damage. We examined the trends of
17 antioxidant enzymes, miRNAs and their target genes in *Zanthoxylum bungeanum* under drought
18 stress. According to the changes of antioxidant enzymes, miRNAs and their target genes
19 expression patterns of *Zanthoxylum bungeanum* under drought stress, an interaction model was
20 constructed to provide a reference for further understanding of plant antioxidant mechanism. The
21 results indicate that under drought stress, POD, CAT, APX, proline, MDA and related genes all
22 show positive responses to drought, while SOD and its genes showed a negative response. It is

23 speculated that in the antioxidant process of *Zanthoxylum bungeanum*, POD, CAT, and APX
24 play a major role, and SOD plays a supporting role. In addition, the expression levels of miRNAs
25 and their target genes were basically negatively correlated, indicating that miRNAs are involved
26 in the regulation of the antioxidant system of *Zanthoxylum bungeanum*.

27 **Keywords:** antioxidant system; *Zanthoxylum bungeanum* Maxim.; reactive oxygen species;
28 drought stress; miRNA-Target gene

29

30 INTRODUCTION

31 *Zanthoxylum bungeanum* Maxim. (common name Chinese prickly ash, family *Rutaceae*) is
32 widely distributed in Asia (Yang et al., 2013) where it is an important economic crop. Evolution
33 and natural selection have led the epidermis of *Z. bungeanum* to bear prickles. This species also
34 has strong drought adaptability. The skin of *Z. bungeanum* is the source of one of the eight
35 traditional Chinese condiments, so this plant plays a very important role in Chinese food culture.
36 Because of its unique numbing taste, *Z. bungeanum* is difficult to replace with other seasonings
37 (Zhang et al., 2014). It has become an important component of the diet in various parts of Asia,
38 especially in China. *Zanthoxylum bungeanum* and pepper become best companions and are
39 together an important part of the ‘hot pot’ culture. In addition to its use as a food seasoning, the
40 skin of *Z. bungeanum* also contains chemical components showing proven medicinal properties,
41 including bactericidal (Zhang et al., 2016b), insecticidal (Zhang et al., 2016a), antioxidant
42 (Zhang et al., 2014) and topical anesthetic (Rong et al., 2016).

43 Drought stress can cause a series of physiological and molecular reactions in plants, which
44 seriously affect normal growth. Thus, drought can cause imbalances in cellular reactive oxygen

45 species (ROS), it can also upset cell membrane lipid peroxidation and it can damage cell and
46 organelle membranes. Excessive ROS have toxic effects on plants. Irrigated agriculture is not yet
47 general, so drought remains one of the most important unfavorable factors affecting both the
48 yield and quality of most commercial crops. Plants have many protective responses to maintain
49 metabolic stability and so continue life under environmental stress. Their antioxidant systems are
50 able to produce a variety of antioxidant enzymes - including superoxide dismutase, peroxidase,
51 antioxidant enzymes, ascorbate peroxidase - to combat the ROS produced under drought stress
52 (Gill & Tuteja, 2010, N. & K., 1977). For example, catalase can decompose H₂O₂ produced in
53 plants to form water and oxygen, reducing or eliminating damage by this ROS (Chance. &
54 Maehly., 1955). The antioxidant system also maintains organelle stability, preventing damage to
55 the chloroplast membrane and so stabilizing the PSII system (Lima et al., 2018). In addition,
56 stomata are important gas exchange organs of plants, playing irreplaceable roles in the regulation
57 of photosynthesis, respiration, transpiration and temperature (Li et al., 2017, Martin-StPaul et al.,
58 2017). The stomata are also the water-regulating organs of plants. Under drought, water
59 conservation becomes the decisive factor for plant survival. The response of stomata to drought
60 is also a way for plants to protect themselves. Under drought stress, plants reduce water loss by
61 stomatal closure and so increase their ability to resist drought (García-Mata. & Lamattina.,
62 2001, Cornic., 2000).

63 The impact of drought on the yield and quality of *Z. bungeanum* is huge and seriously
64 hinders the development of this industry. miRNAs and their target genes for antioxidant defense
65 response under drought stress remain unclear. Hence a study of their behavior under drought
66 stress, will have significance for better understating this species' drought-resistance mechanisms.
67 It can also provide a basis for drought-resistant breeding of *Z. bungeanum* and of related species.

68 MATERIALS AND METHODS

69 Materials

70 The *Z. bungeanum* seeds were collected from the Fengxian Chinese prickly ash Experimental
71 Station of Northwest Agriculture and Forestry University. They were germinated and cultured in
72 an artificial climate chamber at $25\pm 2^{\circ}\text{C}$. Air humidity was set to 80% and the photoperiod to 16:8
73 h (light:dark). Three-month-old seedlings were used as experimental material. *Zanthoxylum*
74 *bungeanum* seedlings were then cultured in half-strength Murashige and Skoog (MS) liquid
75 medium containing 20% PEG6000. Leaf samples were collected and stored in liquid nitrogen
76 after periods of 0, 3, 6, 12, 24, 36 and 48 h.

77 Methods

78 Physiological index determination

79 The leaf samples after different periods of drought stress were used to determine antioxidant
80 enzyme activity and malondialdehyde (MDA) and proline contents. Superoxide dismutase
81 (SOD) activity was determined by the nitroblue tetrazolium method (N. & K., 1977). Peroxidase
82 (POD) activity was determined by the guaiacol method and catalase (CAT) activity was
83 determined by the hydrogen peroxide ultraviolet method (Chance. & Maehly., 1955). For APX
84 (L-ascorbate peroxidase) activity we used the method of Panchuk et al (Panchuk et al., 2002).
85 The MDA content was determined by the thiobarbituric acid (TBA) method (Fu. & Huang.,
86 2001). Proline was determined by ninhydrin colorimetry (Bates et al., 1973).

87 **Total RNA extraction**

88 Total RNA of the *Z. bungeanum* samples was extracted using the TaKaRa MiniBEST Plant RNA
89 Extraction Kit (TaKaRa, Beijing, China) following the manufacturer's instructions. The purity
90 and concentration of the RNA obtained were measured using NanoDrop 20000 (Thermo
91 Scientific, Pittsburgh, PA, USA). Only samples where the OD260/280 value was 1.8-2.0 and the
92 OD260/230 value was higher than 2.0 were used for cDNA synthesis.

93 **Quantitative Real-time PCR**

94 Primer 7.0 software (Premier, Palo Alto, CA, USA) was used to design RT-qPCT primers (see
95 Table 1). The qRT-PCR assays were carried out on a CFX96 Real-Time PCR Detection System
96 (Bio-Rad, Hercules, CA, USA). The reaction system was of 10 μ l, containing 5 μ l of 2 \times SYBR
97 Premix Ex Taq II (TaKaRa), 1 μ l of cDNA, 1 μ l of each of the upstream and downstream
98 primers and 2 μ l of ddH₂O. *ZbUBQ* and *Zb α -EF* were used for the reference genes to correct the
99 RT-qPCR data (Fei et al., 2018). The RT-qPCT reaction system of miRNAs is identical to
100 mRNA, with a reaction system of 10 μ L, containing 5 μ L of 2 \times SYBR Premix Ex Taq II
101 (TaKaRa, Beijing, China), 1 μ L of cDNA, 1 μ L of each of the forward and universal reverse
102 primer and 2 μ L of ddH₂O. U6 was used for the correction of relative expression levels of
103 miRNAs (Zhang et al., 2018).

104 **Table 1.** RT-qPCR primers.

Name	Description	Forward primer sequence (5'-3')	Reverse primer sequence (5'-3')
<i>SOD1</i>	Superoxide dismutase [Mn] 1, mitochondrial	CAGCGTCTCACATCATTTCATTT	TCTTCAGTCCACGTAGCCCTAGT

<i>SOD2</i>	Superoxide dismutase [Fe], chloroplastic-like isoform X2	GGGATTACTCACCTCTCCTTACC	TCCTCTTCTCTTTTCCTCCTTTC
<i>PRX2E</i>	Peroxiredoxin-2E, chloroplastic (POD)	AATCAAAAGGCATCGACACCA	ACTCCCCATTCCCATCAGACA
<i>CAT</i>	Catalase isozyme 1 (CAT)	TTTCCCTGTCTTCTTCATCCGT	TCCTTCCATGTGCCTGTAATCT
<i>APX3</i>	L-ascorbate peroxidase 3	GCCTTCCAGATGCCAAACAA	CCCCCTGAGAGTGCCACTAT
<i>GPXI</i>	Phospholipid hydroperoxide glutathione peroxidase 1, chloroplastic (GPX1)	CAAGTGCTGGGGGATTTTTA	ATGGTGATGTTGTTGGTGGA
<i>P5CS</i>	Delta-1-pyrroline-5-carboxylate synthase. Key enzyme for the synthesis of proline	AGCAAAACACCAAGCAGAAATA A	AATAACAGGGATACCAGCATAA G
<i>JARI</i>	Jasmonic acid-amido synthetase JAR1, participate in the synthesis of jasmonic acid	CTCGGAAGCAGCAGCCAAACT	AGCAAAGGAGCAATCCAAACA
<i>ABI</i>	ABSCISIC ACID-INSENSITIVE 5-like protein 5. Key nodes and inhibitors of the abscisic acid (ABA) signaling pathway regulate a variety of ABA responses, such as stomatal closure, plasma membrane permeability and water permeability.	TGTCTCCAGTTCCTTACATGTTT	CTTGCTGCTGACTCTCTATTCTT

<i>MAPK1</i>	Mitogen-activated protein kinase 1	CTAACTCTAACCCCTCCAGCCCAG	TTTCGTTCCACATCATTTCCTT
<i>PDI52</i>	Protein disulfide-isomerase 5-2 isoform X1	AGAGAAGGAAGAACCGAAAA	GAAGTGCCAACACTGAGAGG
<i>RBOHC</i>	Respiratory burst oxidase homolog protein C [Citrus sinensis]	GGCACCCATTTTCAATAA	GCTCTGAGGAGTCCACTT
<i>NRX1</i>	Probable nucleoredoxin 1	TGAAGCCATCGAAGAACAC	CCCCTAAAACCAAACAGA
<i>TCTP</i>	Translationally-controlled tumor protein homolog; Involved in the regulation of abscisic acid- and calcium-mediated stomatal closure	TCTCTCAGACTCGTTTCCCTAC	CTCCTTGAACAACCCACTTTCC
<i>ZbUBQ</i>	<i>Zanthoxylum bungeanum</i> ubiquitin extension protein, reference gene	TCGAAGATGGCCGTACATTG	TCCTCTAAGCCTCAGCACCA
<i>Zba-EF</i>	<i>Zanthoxylum bungeanum</i> Elongation factor 1-alpha, reference gene	GTGCTTGACTGCCACACCTC	TTCCGGCATCTCCATTCTTC
ath-miR396a-5p	Target gene is SOD1	TTCCACAGCTTTCTTGAAGT	–
ath-miR834	Target gene is SOD2	TGGTAGCAGTAGCGGTGGTAA	–
ath-miR167a-3p	Target gene is PRX2E	GATCATGTTTCGCAGTTTCACC	–
ath-miR169b-3p	Target gene is CAT	GGCAAGTTGTCCTTCGGCTACA	–
ath-miR447a-3p	Target gene is APX3	TTGGGGACGAGATGTTTTGTTG	–
ath-miR773b-3p	Target gene is GPX1	TTTGATTCCAGCTTTTGTCTC	–
ath-miR397b	Target gene is P5CS	TCATTGAGTGCATCGTTGATG	–

ath-miR397b	Target gene is JAR1	TCATTGAGTGCATCGTTGATG	–
ath-miR859	Target gene is ABI	TCTCTCTGTTGTGAAGTCAAA	–
ath-miR5632-5p	Target gene is MAPK1	TTGATTCTCTTATCCAAGTGT	–
ath-miR1888a	Target gene is PDI52	TAAGTTAAGATTTGTGAAGAA	–
ath-miR5638a	Target gene is RBOHC	ATACCAAACTCTCTCACTTT	–
ath-miR398a-3p	Target gene is NRX1	TGTGTTCTCAGGTCACCCCTT	–
ath-miR3434-3p	Target gene is TCTP	TCAGAGTATCAGCCATGTGA	–
U6	miRNAs expression level reference gene	TTGGACCATTCTCGATTTGTGC	CCTTAGGGGACATCCGATAAAA TTG

105 miRNA prediction

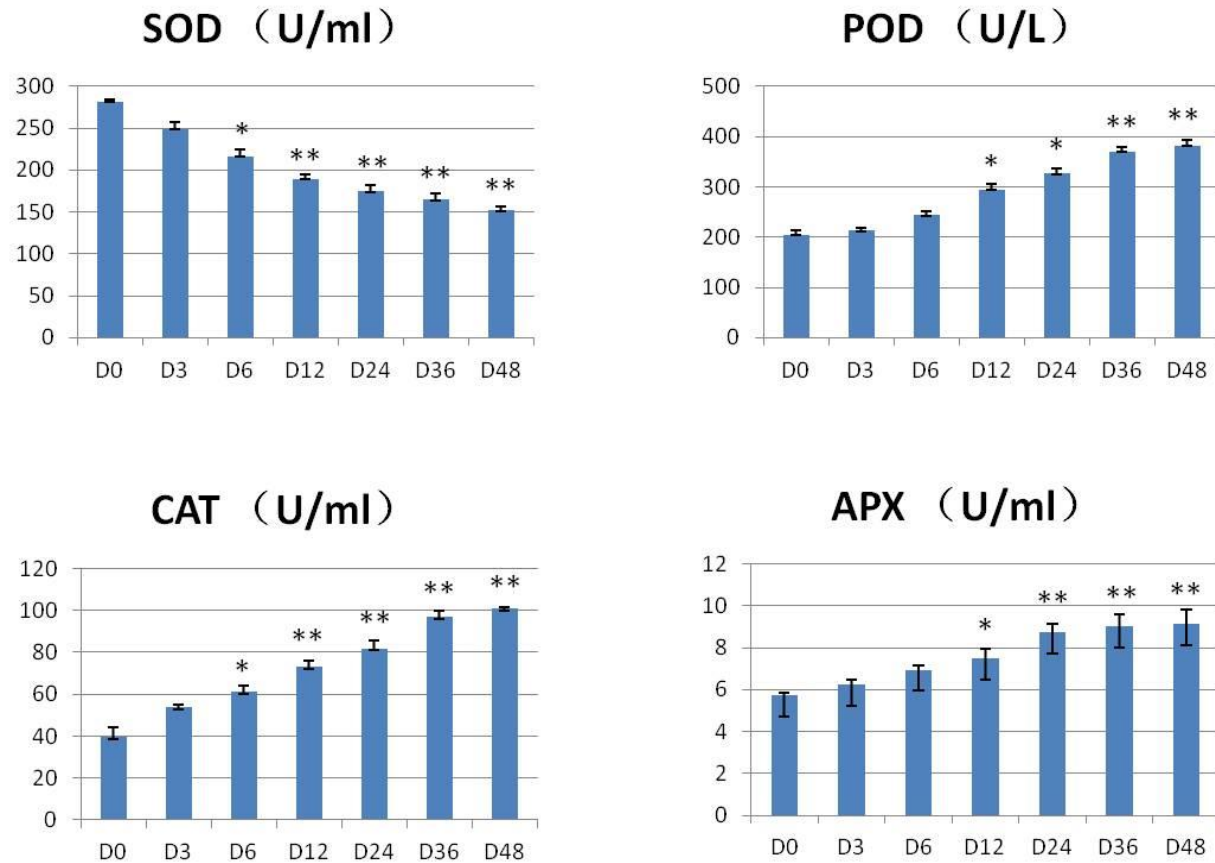
106 miRNAs that interact with mRNA are predicted in the Arabidopsis miRNA database using the
 107 psRNATarget website (http://plantgrn.noble.org/v1_psRNATarget/).

108 RESULTS

109 Effect of Drought Stress on Antioxidant Enzyme Activity of *Zanthoxylum bungeanum*

110 Antioxidant enzymes are important roles in plant antioxidant systems. They can eliminate
 111 reactive oxygen species in plants and avoid damage to plant plasma membranes. They are also
 112 important indicators for evaluating plant antioxidant capacity. The activities of four antioxidant
 113 enzymes, SOD, POD, CAT and APX, were determined under drought stress for 7 periods. The
 114 results showed that the activity of SOD decreased slowly with the prolongation of drought stress,
 115 while POD, CAT and APX decreased. It rose slowly within 6 hours of drought stress, and then

116 increased rapidly between 6h and 24h, and remained at a higher activity level after 24h (Figure
117 1).

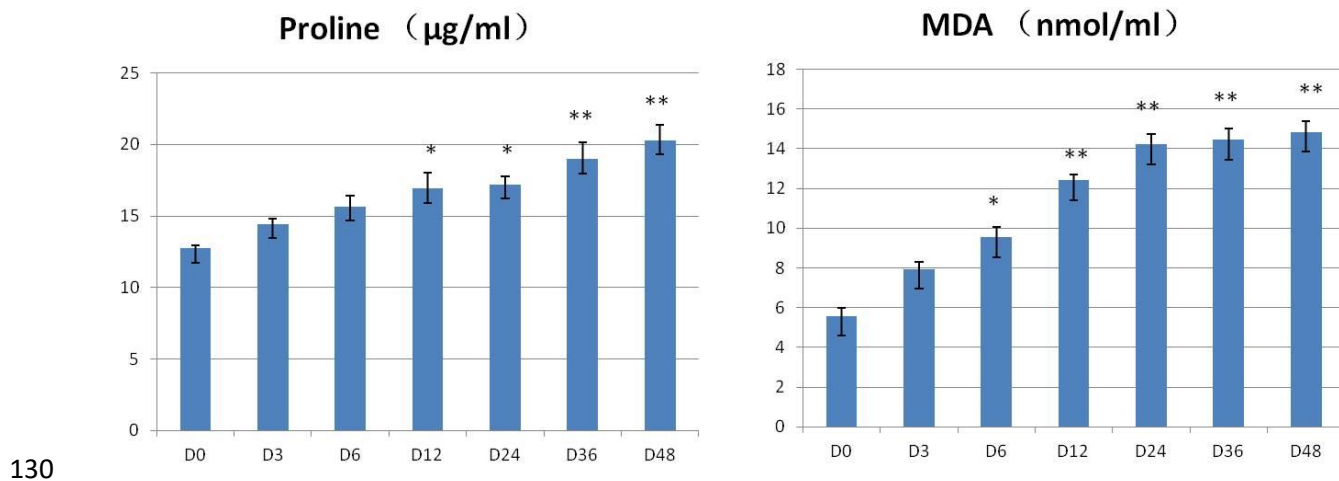


118

119 **Figure 1.** Antioxidant system enzyme activity. * P<0.05; ** P<0.01

120 The changes in the activity of the four enzymes under drought stress indicate that the
121 response of antioxidant enzymes to drought requires reaction time, and it may be necessary to
122 reach a certain amount of signal to activate the activities of antioxidant enzymes and synthetic
123 pathways. In addition, once activated, antioxidant enzymes are not endlessly synthesized, but
124 remain in a range after a period of drought stress.

125 Proline is one of the important protective substances for plants to resist stress. Usually,
126 when plants are subjected to abiotic stress, they will synthesize proline in large quantities to cope
127 with adverse environment. Malondialdehyde (MDA) is produced by the peroxidation of
128 membrane lipids in tissues or organs of plants when they are aging or under drought. Therefore,
129 the content of proline and MDA is an important indicator for evaluating plant stress resistance.



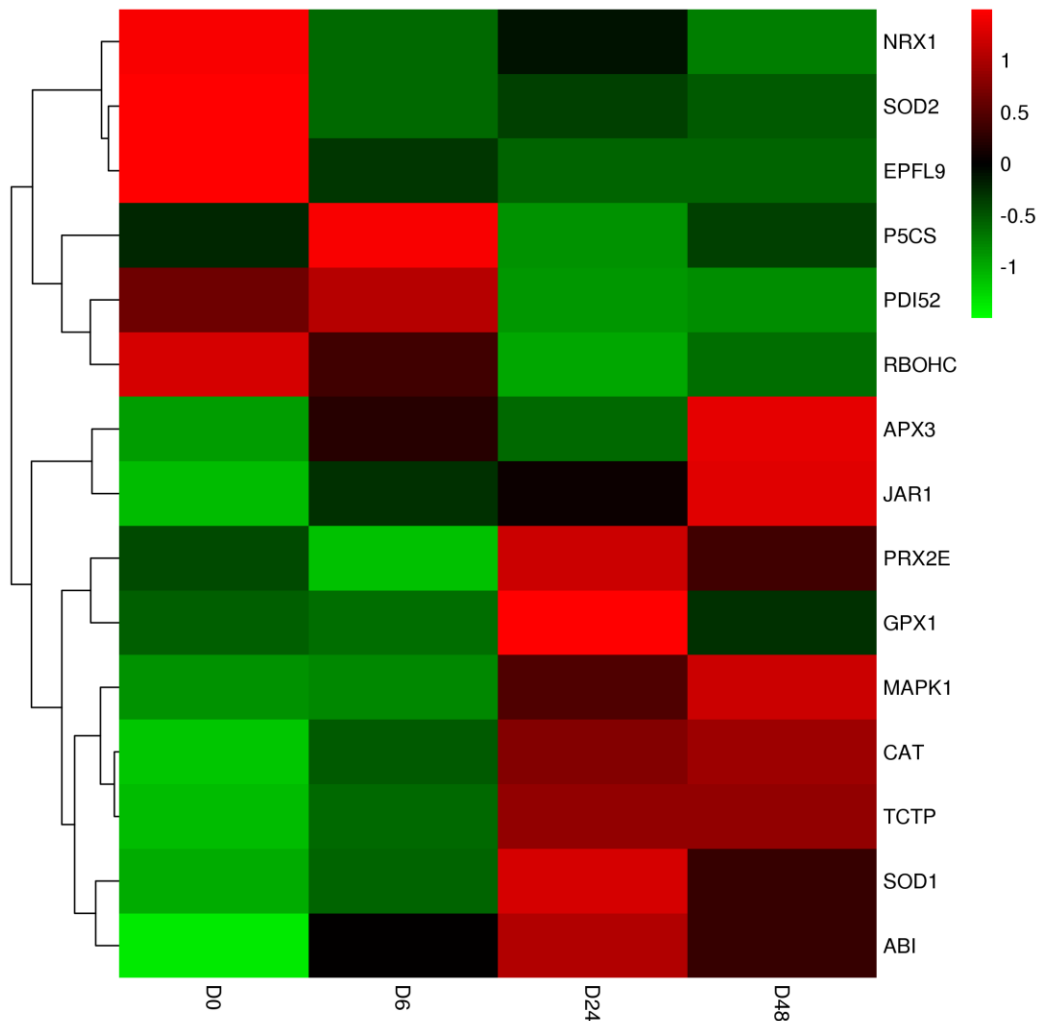
130

131 **Figure 2.** Proline and MDA contents. * P<0.05; ** P<0.01

132 The proline content in *Zanthoxylum bungeanum* was gradually increased from 13 µg/ml at
133 the initial stage of drought stress to 20 µg/ml at 48 h. The content of MDA changed more
134 vigorously, and it quickly exceeded 12 nmol/ml within 12 hours, which was more than twice that
135 of D0 period, and then remained above 14 nmol/ml for 24h to 48h (Figure 2). The increase of
136 proline and MDA content indicates that in the process of drought stress of *Zanthoxylum*
137 *bungeanum*, on the one hand, the peroxidation of organ membrane lipids affects the
138 physiological function of plants, on the other hand, the protective substances are also synthesized
139 to slow down the damage of plants caused by adverse environment.

140 **Expression pattern of miRNAs and their target genes under drought stress**

141 The expression patterns of genes related to the antioxidant system were essentially consistent
142 with the changes in the related substances. Genes such as *PRX2E*, *CAT*, *APX3*, *P5CS* and *GPX1*
143 showed a significant increase under drought stress (Figure 3).



144

145 **Figure 3.** Heat map of gene expressions relating to the antioxidant system.

146 miRNAs interacting with mRNAs were predicted via the psRNATarget website (Table 2).

147 The predicted results indicate that ath-miR396a-5p binds to *SOD1* and inhibits its transcription.

148 Ath-miR167a-3p interacts with *PRX2E*, and *CAT* is the target gene of ath-miR169b-3p. In
149 addition, we also predicted miRNAs interacting with other genes on the drought stress-related
150 signaling pathway, involving jasmonic acid signaling pathway, ABA signaling pathway, MAPK
151 signaling pathway and proline synthesis pathway. Furthermore, the relative expression levels of
152 miRNAs regulating the genes involved in the synthesis of antioxidant enzymes were detected.
153 The results showed that the expression trends of miRNAs and their target genes were basically
154 negatively correlated (Figure 4). It can be stated that these miRNAs are important regulators
155 involved in the antioxidant system of *Z. bungeanum*.

156 **Table 2.** miRNAs and their target genes.

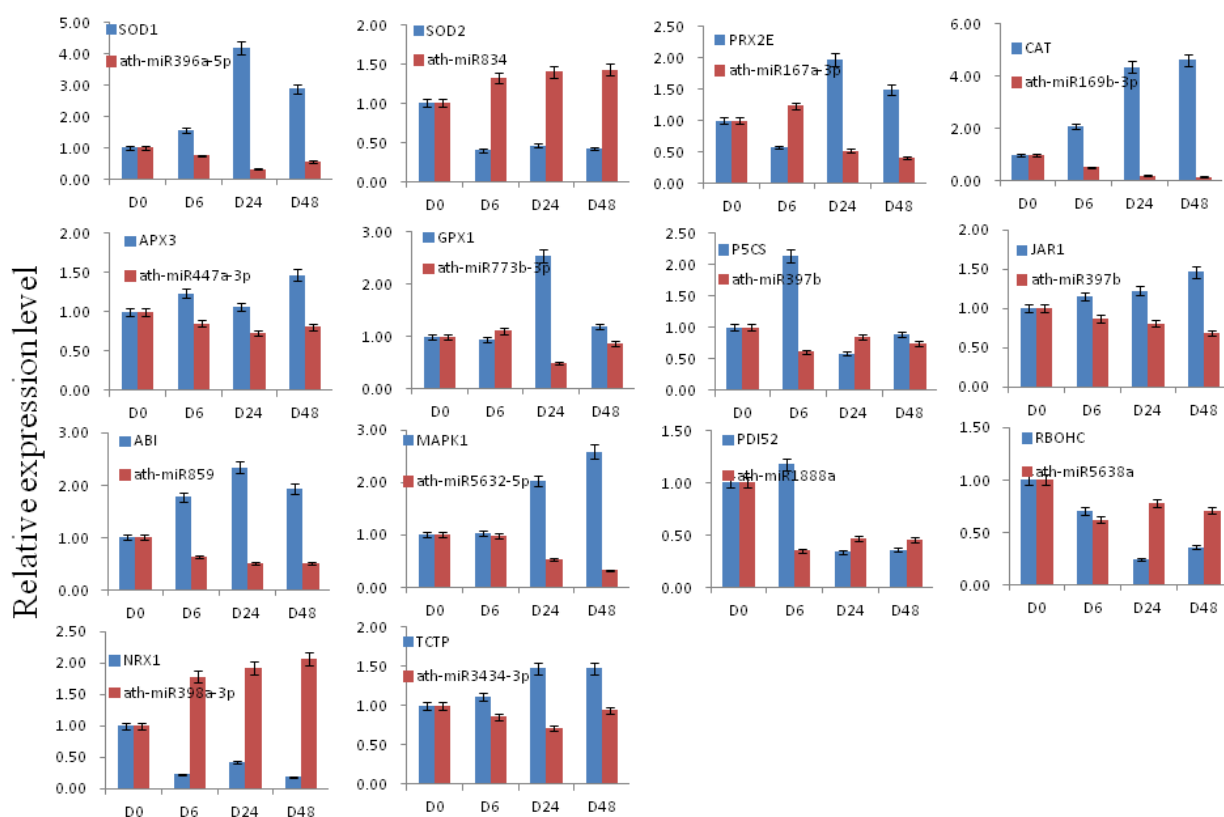
miRNA	Mutant	Target gene
ath-miR396a-5p	UUCCACAGCUUUCUUGAACUG	<i>SOD1</i>
ath-miR834	UGGUAGCAGUAGCGGUGGUAA	<i>SOD2</i>
ath-miR167a-3p	GAUCAUGUUCGCAGUUUCACC	<i>PRX2E</i>
ath-miR169b-3p	GGCAAGUUGUCCUUCGGCUACA	<i>CAT</i>
ath-miR447a-3p	UUGGGGACGAGAUGUUUUGUUG	<i>APX3</i>
ath-miR773b-3p	UUUGAUUCCAGCUUUUGUCUC	<i>GPX1</i>
ath-miR397b	UCAUUGAGUGCAUCGUUGAUG	<i>P5CS</i>
ath-miR397b	UCAUUGAGUGCAUCGUUGAUG	<i>JAR1</i>
ath-miR859	UCUCUCUGUUGUGAAGUCAAA	<i>ABI</i>

ath-miR5632-5p	UUGAUUCUCUUAUCCAACUGU	<i>MAPK1</i>
ath-miR1888a	UAAGUUAAGAUUUGUGAAGAA	<i>PDI52</i>
ath-miR5638a	AUACCAAACUCUCUCACUUU	<i>RBOHC</i>
ath-miR398a-3p	UGUGUUCUCAGGUCACCCCUU	<i>NRX1</i>
ath-miR3434-3p	UCAGAGUAUCAGCCAUGUGA	<i>TCTP</i>

157

158 The expression levels of the SOD gene in chloroplasts and mitochondria were monitored
159 and we found the *SOD2* gene in the chloroplast was positively correlated with superoxide
160 dismutase activity. The *SOD1* gene in mitochondria was negatively correlated with the activity
161 of superoxide dismutase. It is concluded that the superoxide dismutase produced by *Z.*
162 *bungeanum* under drought stress comes mainly from the chloroplast.

163 In addition, some expression patterns of pathway genes activated by drought stress were
164 also monitored. *P5CS* is a key enzyme in the proline synthesis process. *ABI* is a key inhibitor of
165 the ABA signaling pathway and participates in the closure of stomata and *TCTP* is involved in
166 ABA and calcium ion-mediated stomatal closure. In addition, the relative expression levels of
167 *MAPK1* and *PDI52* were also up-regulated (Figure 4). At the same time, the relative expression
168 of *JAR1*, a gene related to jasmonic acid synthesis, was also up-regulated under the induction of
169 drought stress. However, other genes were inhibited, such as *RBOHC*, *NRX1*, *EPFL9*.



170

171 **Figure 4.** miRNAs and their target genes.

172

173 DISCUSSION

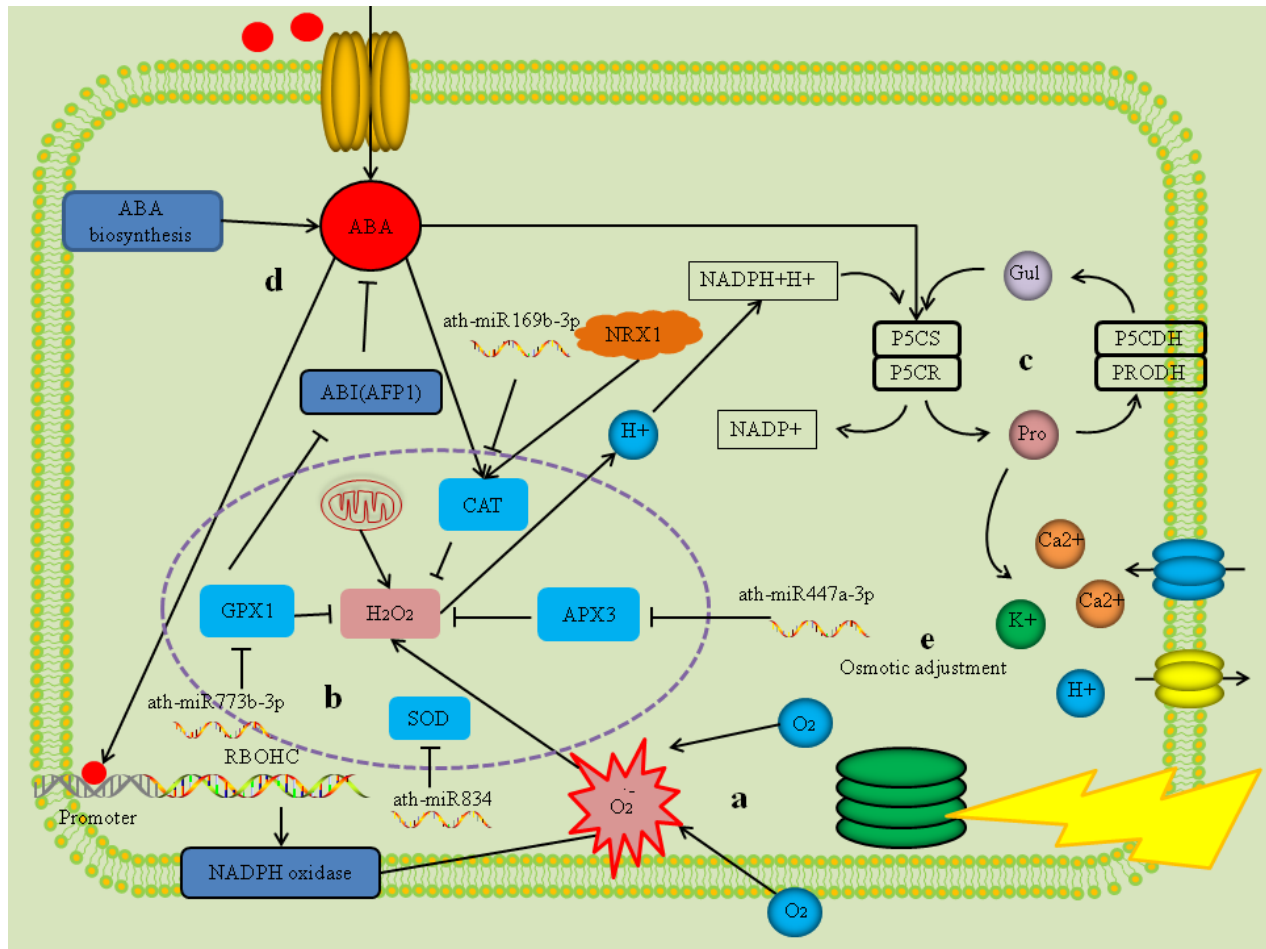
174 Response of antioxidant enzymes under drought stress

175 The results show that most of antioxidant enzymes trend upward under continuing drought stress,
 176 while SOD shows a downward trend. The trend of the *SOD2* gene and SOD in the chloroplasts
 177 are consistent, which indicates the chloroplast antioxidant system was severely damaged during
 178 the drought. In addition, it can be explained that in the antioxidant system, several other

179 antioxidant enzymes (such as POD, CAT, APX) exert major antioxidant effects. Proline and
180 MDA increased gradually during the drought stress and may also be involved in signal
181 transduction and protection. Studies have shown that *Barbula fallax* and *Zanthoxylum*
182 *bungeanum* have similar patterns of antioxidant enzyme activity, where SOD showed a
183 downward trend in the early stage of drought stress, while POD and CAT activities were positive
184 in response to drought stress and increased in the early stage of drought stress (Zhang et al.,
185 2017). In many species, the activities of antioxidant enzymes such as SOD, POD, CAT, and
186 APX generally rise under drought. Chickpea accumulates proline and increases the activity of
187 SOD, APX, GPX and CAT under drought stress (Dalvi et al., 2017). The activities of SOD, POD
188 and CAT in alfalfa increase significantly under drought stress (Tina et al., 2017). This also
189 occurs in pea (Mittler. & Zilinskas., 1994), rice (Sharma & Dubey, 2005), Kentucky bluegrass
190 (Bian & Jiang, 2009) and sesame (FAZELI. et al., 2007).

191 **Antioxidant signaling pathway regulatory factors interaction**

192 Many signal pathways are activated in plants under drought stress, including signal transduction,
193 gene interaction, physiological changes and so on. Under drought stress, the accumulation of
194 ROS in plants seriously affects the normal growth and development of plants. Antioxidant
195 system can effectively prevent the damage caused by ROS produced by plants, and is an
196 indispensable self-protection system for plants. It plays a very complex process in the antioxidant
197 system, involving the synthesis of antioxidant enzymes, transcription and modification of
198 functional genes, transport of ions, and so on. According to the experimental results and previous
199 research results, the factors involved in the regulation of antioxidant system were analyzed and a
200 signal regulation model of antioxidant system was constructed.



201

202 **Figure 5.** Antioxidant signaling pathway genes interaction model. (a) Oxygen produced by
 203 photosynthesis and oxygen in vitro are the main sources of ROS. (b) SOD, POD, CAT, APX,
 204 GPX convert superoxide anion to hydrogen peroxide and eventually decompose into oxygen and
 205 water. (c) The antioxidative enzyme decomposes H^+ produced by hydrogen peroxide as a
 206 substrate for the synthesis of proline from glutamate. (d) ABA can bind to the *RBOHC* promoter
 207 to promote the synthesis of NADPH oxidase. (e) Proline can regulate the concentration of ions in
 208 cells.

209 Under drought, mitochondrial respiration can produce ROS. Oxygen produced by
 210 chloroplast photosynthesis and external oxygen are also sources of ROS in plants (Figure 5a).

211 Reactive oxygen species can destroy cell membranes and interfere with normal growth of plants,
212 while the accumulation of ROS is toxic to plants. The production of ROS activates the plant's
213 antioxidant system. SOD, POD, CAT, APX, GPX and other enzymes can convert superoxide
214 anion to H₂O₂ and eventually decompose it into non-toxic H₂O and O₂ (Wang et al., 2018, Duan
215 et al., 2009) (Figure 5b).

216 In this process, miRNAs are involved in the regulation of the synthesis of antioxidant
217 enzymes, directly inhibiting the transcription or degradation of the corresponding mRNA,
218 resulting in a decrease in the amount of antioxidant enzyme synthesis. However, by analyzing
219 the expression levels of miRNAs, the expression levels of miRNAs associated with antioxidant
220 systems are mostly declining. The above analysis can show that in order to protect itself from
221 ROS damage under drought stress, plants have largely relieved the inhibitory effect of miRNAs
222 on antioxidant system. During the scavenging of H₂O₂ by antioxidant enzymes, it is susceptible
223 to oxidative stress, resulting in reduced clearance. NRX1 is able to reduce oxidized antioxidant
224 enzymes and has a stable antioxidant system (Kneeshaw. et al., 2017). However, in the gene
225 expression level study, the expression level of *NRX1* was down-regulated. It is concluded that
226 drought interfered with the reduction of NRX1 against oxidase. The antioxidant system
227 decomposes ROS to produce H⁺, which provides a substrate for glutamate synthesis in the
228 proline pathway (Figure 5c).

229 Proline is an important osmo-regulatory substance, and the main way to regulate the
230 osmotic potential is to regulate the concentration of ions in the cell. Proline produced under
231 drought stress protects cells from damage by controlling the concentration of ions, and becomes
232 an important regulator of plant self-protection (Tieleman et al., 2001, Woolfson et al., 1991).

233 *P5CS* is an important synthetic gene in the proline synthesis pathway, and *ath-miR397b* can
234 inhibit the expression level of *P5CS*. Under drought stress, the relative expression level of *ath-*
235 *miR397b* was decreased, which inhibited the inhibition of *P5CS* and promoted the synthesis of
236 proline. The accumulation of ROS can activate the Ca^{2+} channel on the cell membrane(Figure
237 5e), causing a large amount of Ca^{2+} to enter the cell, and so can increase the Ca^{2+} concentration
238 of the guard cells and change their osmotic potential (Singh et al., 2017). At the same time, high
239 concentration of Ca^{2+} can suppress the input of K^+ and the output of H^+ . In addition, *SLAC1*
240 transports anions out (Vahisalu et al., 2008), resulting in an increase in the concentration of
241 cations in the membrane. The combination of ABA and TCTP can induce stomatal closure, and
242 CPK can phosphorylate CAT as well as promote stomatal closure (Zou et al., 2010).

243 At the same time, plants can also synthesize ABA under drought stress, and ABA can
244 promote the synthesis of proline from glutamate, which eventually leads to a large accumulation
245 of proline (Figure 5d) (Strizhov. et al., 1997). Proline is an important osmotic adjustment
246 substance in plants, so the above reaction is beneficial, allowing plants to cope better with
247 drought. ABA binds to the promoter of *RBOHC* and promotes the production of respiratory burst
248 oxidase (NADPH oxidase) (Zhao. et al., 2001). NADPH oxidase (NOX) also activates Ca^{2+}
249 channels on the cell membrane (Kurusu et al., 2015). In addition, it has been shown to play an
250 important protective role in plant drought stress, preventing leaves from being destroyed by ROS
251 (Duan et al., 2009, Miller. et al., 2009). Under drought stress, the ABA signaling pathway and
252 antioxidant system of plants are activated, and there is close interaction between them (Qi. et al.,
253 2015, He et al., 2018). On the one hand, ABA can promote the synthesis of CAT and improve
254 the efficiency of the antioxidant system. While, on the other hand, *GPXI* can inhibit *ABI*, thereby
255 relieving the inhibition of ABA by plants (Lim et al., 2017).

256 **ACKNOWLEDGMENTS**

257 The authors would like to thank Yao Ma for his participation in the manuscript
258 discussion. This study was financially supported by the National Key Research and
259 Development Program Project Funding (2018YFD1000605).

260 **REFERENCES**

- 261 Bates LS, Waldren RP, Teare ID, 1973. Rapid determination of free proline for water-stress
262 studies. *Plant Soil* **39**, 205–7.
- 263 Bian S, Jiang Y, 2009. Reactive oxygen species, antioxidant enzyme activities and gene
264 expression patterns in leaves and roots of Kentucky bluegrass in response to drought stress and
265 recovery. *Scientia Horticulturae* **120**, 264-70.
- 266 Chance. B, Maehly. AC, 1955. Assay of catalases and peroxidases. *Methods in Enzymology* **2**,
267 764–75.
- 268 Cornic. G, 2000. Drought stress inhibits photosynthesis by decreasing stomatal aperture--not by
269 affecting ATP synthesis. *Trends in Plant Science* **5**, 187-8.
- 270 Dalvi US, Naik RM, Lokhande PK, 2017. Antioxidant defense system in chickpea against
271 drought stress at pre- and post- flowering stages. *Indian Journal of Plant Physiology* **23**, 16-23.
- 272 Duan ZQ, Bai L, Zhao ZG, *et al.*, 2009. Drought-stimulated activity of plasma membrane
273 nicotinamide adenine dinucleotide phosphate oxidase and its catalytic properties in rice. *J Integr*
274 *Plant Biol* **51**, 1104-15.
- 275 Fazeli. F, Ghorbanli. M, Niknam. V, 2007. Effect of drought on biomass, protein content, lipid
276 peroxidation and antioxidant enzymes in two sesame cultivars. *Biologia Plantarum* **51**, 98-103.
- 277 Fei X, Shi Q, Yang T, Fei Z, Wei A, 2018. Expression Stabilities of Ten Candidate Reference

278 Genes for RT-qPCR in *Zanthoxylum bungeanum* Maxim. *Molecules* **23**.

279 Fu. J, Huang. B, 2001. Involvement of antioxidants and lipid peroxidation in the adaptation of
280 two cool-season grasses to localized drought stress. *Environmental and Experimental Botany* **45**,
281 105–14.

282 Garcí'a-Mata. C, Lamattina. L, 2001. Nitric Oxide Induces Stomatal Closure and Enhances the
283 Adaptive Plant Responses against Drought Stress. *Plant Physiology* **126**, 1196-204.

284 Gill SS, Tuteja N, 2010. Reactive oxygen species and antioxidant machinery in abiotic stress
285 tolerance in crop plants. *Plant Physiol Biochem* **48**, 909-30.

286 He F, Wang HL, Li HG, *et al.*, 2018. PeCHYR1, a ubiquitin E3 ligase from *Populus euphratica*,
287 enhances drought tolerance via ABA-induced stomatal closure by ROS production in *Populus*.
288 *Plant Biotechnol J* **16**, 1514-28.

289 Kneeshaw. S, Keyani. R, Delorme-Hinoux. V, *et al.*, 2017. Nucleoredoxin guards against
290 oxidative stress by protecting antioxidant enzymes. *PNAS* **114**, 8414–9.

291 Kurusu T, Kuchitsu K, Tada Y, 2015. Plant signaling networks involving Ca(2+) and Rboh/Nox-
292 mediated ROS production under salinity stress. *Front Plant Sci* **6**, 427.

293 Li J, Li Y, Yin Z, *et al.*, 2017. OsASR5 enhances drought tolerance through a stomatal closure
294 pathway associated with ABA and H₂O₂ signalling in rice. *Plant Biotechnol J* **15**, 183-96.

295 Lim IK, Choi JA, Kim EY, *et al.*, 2017. TIS21(/BTG2) inhibits doxorubicin-induced stress fiber-
296 vimentin networks via Nox4-ROS-ABI2-DRF-linked signal cascade. *Cell Signal* **30**, 179-90.

297 Lima CS, Ferreira-Silva SL, Carvalho FEL, *et al.*, 2018. Antioxidant protection and PSII
298 regulation mitigate photo-oxidative stress induced by drought followed by high light in cashew
299 plants. *Environmental and Experimental Botany* **149**, 59-69.

300 Martin-Stpaul N, Delzon S, Cochard H, 2017. Plant resistance to drought depends on timely

301 stomatal closure. *Ecol Lett* **20**, 1437-47.

302 Miller. G, Schlauch. K, Tam. R, *et al.*, 2009. The Plant NADPH Oxidase RBOHD Mediates
303 Rapid Systemic Signaling in Response to Diverse Stimuli. *Science Signaling* **2**, ra45.

304 Mittler. R, Zilinskas. BA, 1994. Regulation of pea cytosolic ascorbate peroxidase and other
305 antioxidant enzymes during the progression of drought stress and following recovery from
306 drought. *The Plant Journal* **5**, 397-405.

307 N. GC, K. RS, 1977. Superoxide dismutases: I . Occurrence in higher plants. *Plant Physiology*
308 **59**, 309-14.

309 Panchuk, Ii, Volkov RA, Schoffl F, 2002. Heat stress- and heat shock transcription factor-
310 dependent expression and activity of ascorbate peroxidase in Arabidopsis. *Plant Physiol* **129**,
311 838-53.

312 Qi. W, Zhang. L, Feng. W, Xu. H, Wang. L, Jiao. Z, 2015. ROS and ABA Signaling Are Involved
313 in the Growth Stimulation Induced by Low-Dose Gamma Irradiation in Arabidopsis Seedling.
314 *Applied Biochemistry and Biotechnology* **175**, 1490–506.

315 Rong R, Cui MY, Zhang QL, *et al.*, 2016. Anesthetic constituents of *Zanthoxylum bungeanum*
316 Maxim.: A pharmacokinetic study. *J Sep Sci* **39**, 2728-35.

317 Sharma P, Dubey RS, 2005. Drought Induces Oxidative Stress and Enhances the Activities of
318 Antioxidant Enzymes in Growing Rice Seedlings. *Plant Growth Regulation* **46**, 209-21.

319 Singh R, Parihar P, Singh S, Mishra RK, Singh VP, Prasad SM, 2017. Reactive oxygen species
320 signaling and stomatal movement: Current updates and future perspectives. *Redox Biol* **11**, 213-8.

321 Strizhov. N, Braha´M. EA, Sz. LSOK, *et al.*, 1997. Differential expression of two P5CS genes
322 controlling proline accumulation during salt-stress requires ABA and is regulated by ABA1,

- 323 ABI1 and AXR2 in Arabidopsis. *The Plant Journal* **12**, 557–69.
- 324 Tieleman D, Shrivastava I, Ulmschneider M, Sansom M, 2001. Proline-induced hinges in
325 transmembrane helices: Possible roles in ion channel gating. *Proteins-structure Function &*
326 *Bioinformatics* **44**, 63.
- 327 Tina RR, Shan XR, Wang Y, *et al.*, 2017. Response of antioxidant system to drought stress and
328 re-watering in Alfalfa during branching. *IOP Conference Series: Earth and Environmental*
329 *Science* **94**, 012129.
- 330 Vahisalu T, Kollist H, Wang YF, *et al.*, 2008. SLAC1 is required for plant guard cell S-type anion
331 channel function in stomatal signalling. *Nature* **452**, 487-91.
- 332 Wang Y, Branicky R, Noe A, Hekimi S, 2018. Superoxide dismutases: Dual roles in controlling
333 ROS damage and regulating ROS signaling. *J Cell Biol* **217**, 1915-28.
- 334 Woolfson DN, Mortishire-Smith RJ, Williams DH, 1991. Conserved positioning of proline
335 residues in membrane-spanning helices of ion-channel proteins. *Biochem Biophys Res Commun*
336 **175**, 733-7.
- 337 Yang LC, Li R, Tan J, Jiang ZT, 2013. Polyphenolics composition of the leaves of *Zanthoxylum*
338 *bungeanum* Maxim. grown in Hebei, China, and their radical scavenging activities. *J Agric Food*
339 *Chem* **61**, 1772-8.
- 340 Zhang WJ, Guo SS, You CX, *et al.*, 2016a. Chemical Composition of Essential Oils from
341 *Zanthoxylum bungeanum* Maxim. and Their Bioactivities against *Lasioderma serricorne*. *J Oleo*
342 *Sci* **65**, 871-9.
- 343 Zhang X, Li K, Xing R, *et al.*, 2018. miRNA and mRNA Expression Profiles Reveal Insight into
344 Chitosan-Mediated Regulation of Plant Growth. *J Agric Food Chem* **66**, 3810-22.
- 345 Zhang X, Zhao Y, Wang S, 2017. Responses of antioxidant defense system of epilithic mosses to

346 drought stress in karst rock desertified areas. *Acta Geochimica* **36**, 205-12.

347 Zhang Y, Luo Z, Wang D, He F, Li D, 2014. Phytochemical profiles and antioxidant and
348 antimicrobial activities of the leaves of *Zanthoxylum bungeanum*. *ScientificWorldJournal* **2014**,
349 181072.

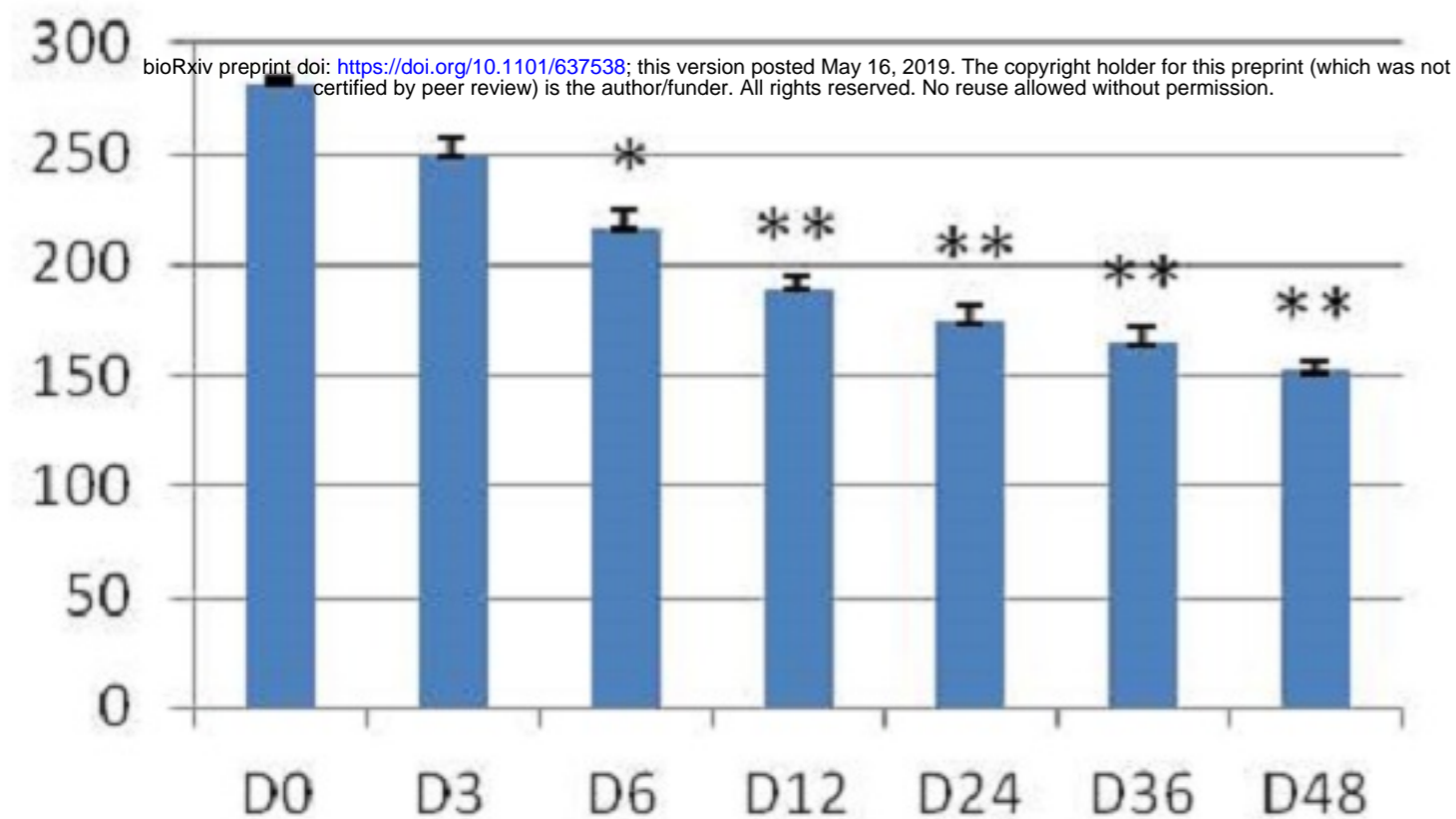
350 Zhang Z, Liu J, Shen P, *et al.*, 2016b. *Zanthoxylum bungeanum* pericarp extract prevents dextran
351 sulfate sodium-induced experimental colitis in mice via the regulation of TLR4 and TLR4-
352 related signaling pathways. *Int Immunopharmacol* **41**, 127-35.

353 Zhao. Z, Chen. G, Zhang. C, 2001. Interaction between reactive oxygen species and nitric oxide
354 in drought-induced abscisic acid synthesis in root tips of wheat seedlings. *Australian Journal of*
355 *Plant Physiology* **28**, 1055–61.

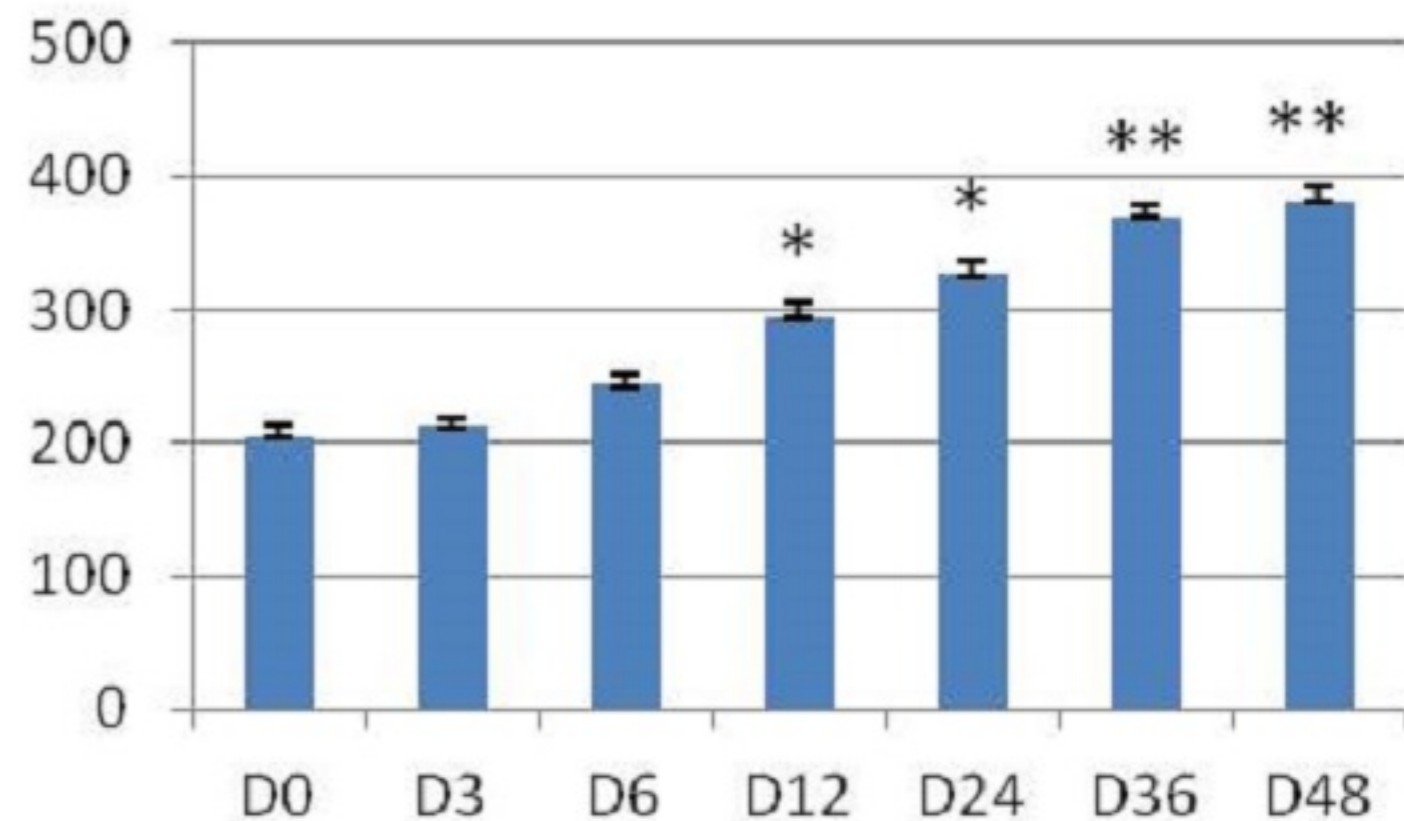
356 Zou JJ, Wei FJ, Wang C, *et al.*, 2010. Arabidopsis calcium-dependent protein kinase CPK10
357 functions in abscisic acid- and Ca²⁺-mediated stomatal regulation in response to drought stress.
358 *Plant Physiol* **154**, 1232-43.

359

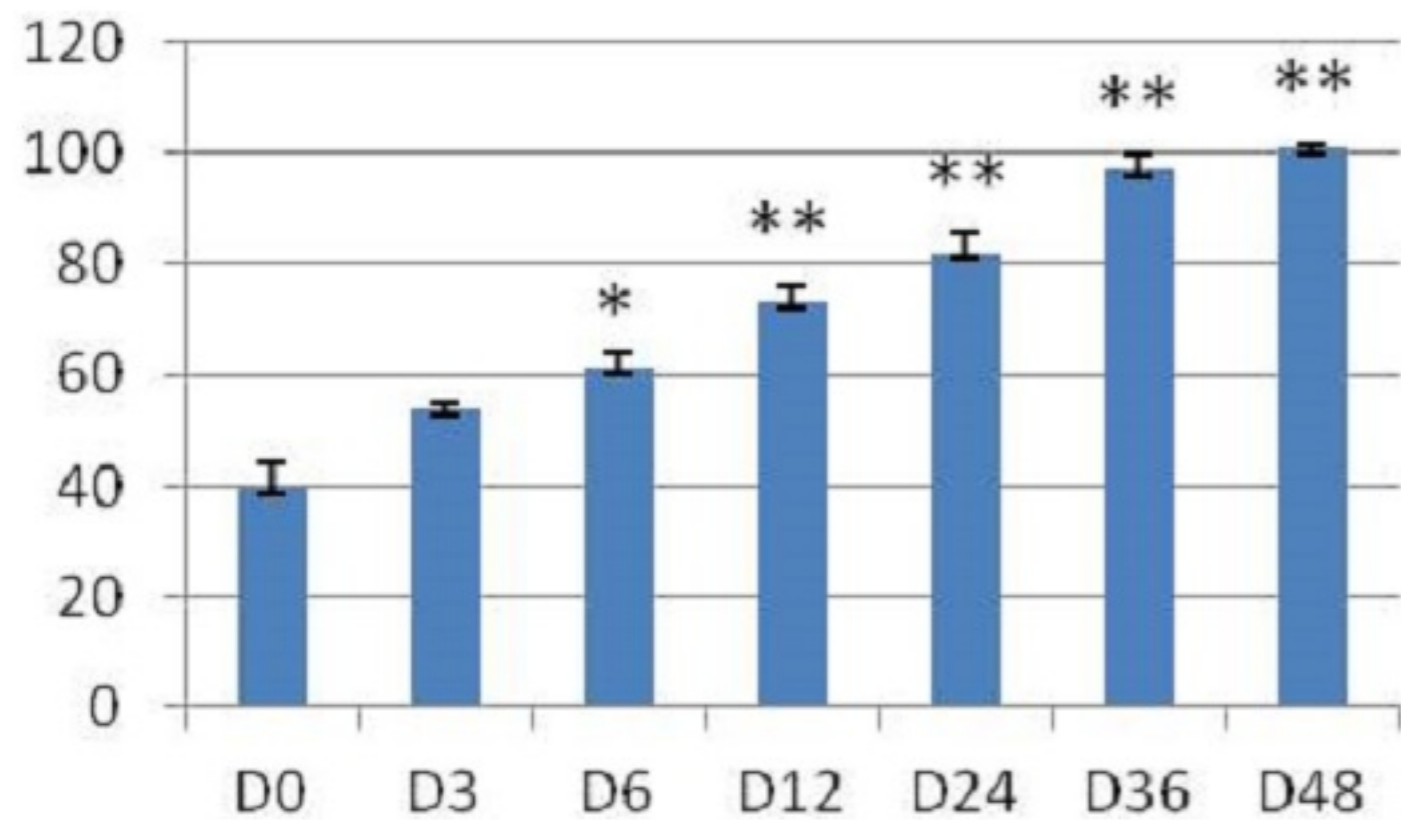
SOD (U/ml)



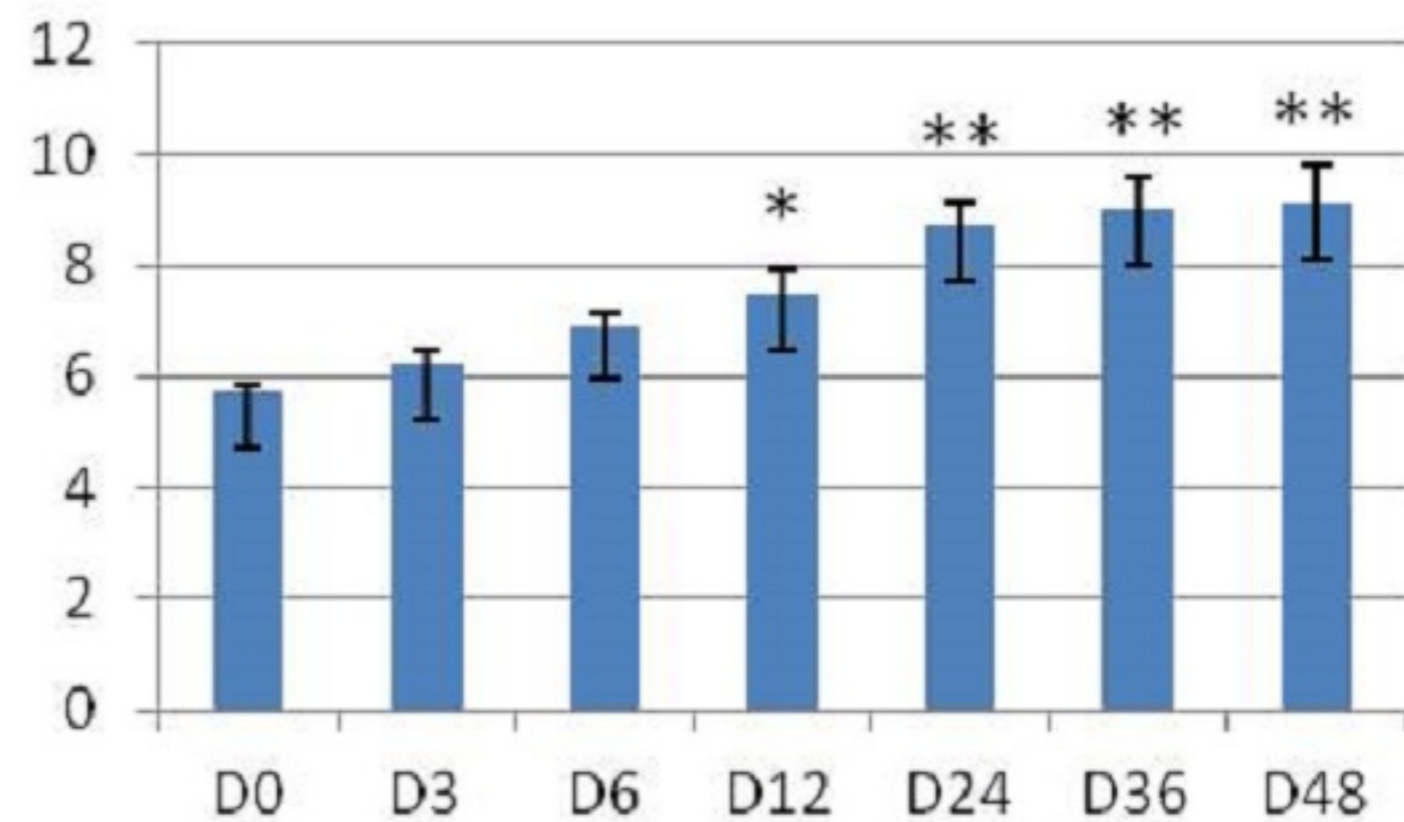
POD (U/L)



CAT (U/ml)

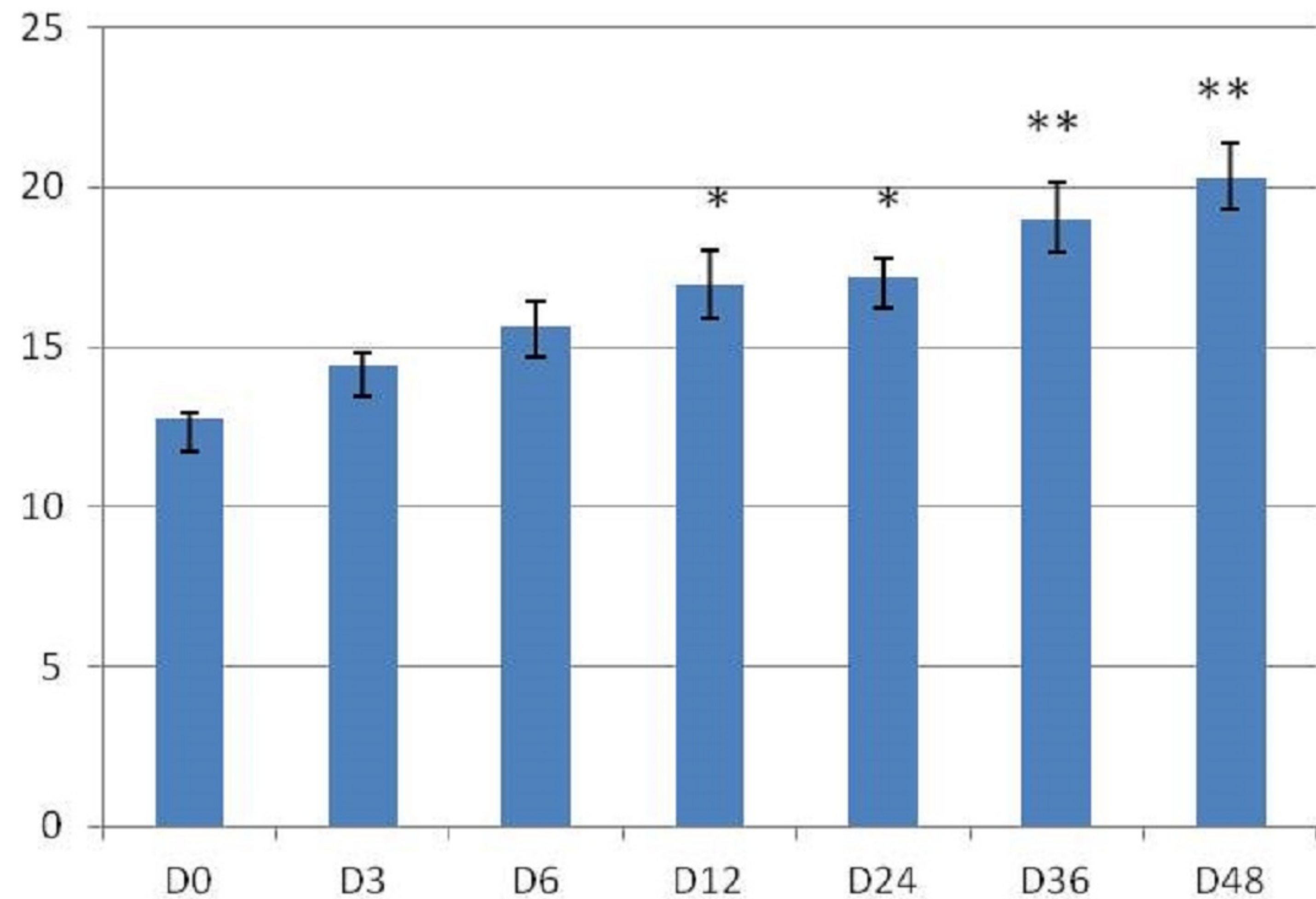


APX (U/ml)



Proline ($\mu\text{g/ml}$)

bioRxiv preprint doi: <https://doi.org/10.1101/637538>; this version posted May 16, 2019. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.



MDA (nmol/ml)

