Discovery of compounds inhibiting the ADP-ribosyltransferase activity of pertussis toxin

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RUNNING TITLE

Inhibitory compounds for pertussis toxin

ABSTRACT

Bordetella pertussis causes the highly contagious respiratory disease pertussis, also known as

whooping cough. The resurgence of pertussis has been witnessed even in highly vaccinated

populations, and macrolide-resistant strains have been isolated. One attractive target for drug

development is the pertussis toxin – an important type IV secretion system-dependent virulence

factor of *B. pertussis*. The AB_5-topology pertussis toxin is composed of a pentameric PtxS2-S5

(1:1:2:1) complex mediating toxin binding to cell surface receptors, and one ADP-

ribosyltransferase subunit PtxS1. Once internalized into the host cell, PtxS1 ADP-ribosylates α-

subunits of heterotrimeric Gαi-superfamily members, thereby disrupting G-protein-coupled

receptor (GPCR) signaling. Here, we describe protocols to purify mg-levels of truncated but

highly active recombinant *B. pertussis* PtxS1 from *E. coli* and an *in vitro* high throughput-

compatible assay to quantify NAD^+ consumption during PtxS1-catalyzed Gαi ADP-ribosylation.

The *in vitro* NAD^+ consumption assay was used to screen compounds inhibiting the PtxS1

activity. Two inhibitory compounds (NSC 29193 and NSC 228155) with low micromolar IC_{50}-

values were identified that also were potent in an independent *in vitro* assay monitoring

conjugation of ADP-ribose to Gαi. Docking and molecular dynamics simulations identified

plausible binding poses of NSC 228155 and in particular NSC 29193, most likely owing to the

rigidity of the latter ligand, at the NAD^+-binding pocket of PtxS1. NSC 228155 inhibited the

pertussis AB_5 holotoxin-catalyzed ADP-ribosylation of Gαi in living human cells in low

micromolar concentration. NSC 228155 and NSC 29193 might prove useful as lead compounds

in targeted drug development in pertussis.

INTRODUCTION

Pertussis, i.e. whooping cough, is a globally distributed acute respiratory disease (1). Pertussis affects all age groups. However, young children are the most affected age group where pertussis may lead to death despite hospital intensive care and use of antibiotics. Worldwide estimates of pertussis cases and deaths in children younger than 5 years in 2014 were 24.1 million and 160.700, respectively (2). Despite efficient global vaccine campaign pertussis remains endemic and recent data, e.g. from USA (3), indicate that the number of pertussis cases is increasing. Sizeable outbreaks have also been reported (1), but the reasons for the resurgence of pertussis are incompletely understood. On the one hand, improved diagnostics and surveillance methods as well as increased awareness of pertussis by health care professionals might contribute (1). On the other hand, molecular changes in the pathogenic *B. pertussis* lineages and waning immunity especially after receipt of acellular pertussis vaccines have been extensively debated (1). These data highlight the need to improve the vaccine formulations and vaccination campaigns, but also to develop alternative means to treat pertussis.

Pertussis toxin is included in the acellular pertussis vaccine formulations in a detoxified form (1), and has provided good protection even as a single component (4). At the same time, clinical isolates lacking pertussis toxin, i.e. vaccine escape mutants, have turned out to be extremely rare (5). Indeed, pertussis toxin is considered as a major virulence factor of *B. pertussis* (6, 7). When administrated systemically to experimental animals, e.g. to mice (8), pertussis toxin recapitulated the leukocytosis (increase in number of circulating white blood cells) seen in young children with pertussis (1). Rats experimentally infected with *B. pertussis* developed paroxysmal coughing lasting several days, but an isogenic pertussis toxin-deficient strain did not cause such pathology (9, 10). Seven-day-old neonatal mice infected with a pertussis toxin-deficient strain of *B. pertussis* fully survived a challenge, which caused 100% mortality with the parental strain (11).

Therefore, targeting of pertussis toxin might prove beneficial in the treatment of pertussis,

especially in young children who still lack the vaccine-induced protection against pertussis.

Pertussis toxin is composed of five non-covalently bound subunits (PtxS1-S5), which are

arranged in an AB_5-topology (12, 13). The B_5-assembly is formed by the PtxS2-S5 subunits

(PtxS2, PtxS3, 2 copies of PtxS4, PtxS5) (12, 13). Pertussis toxin is secreted from the bacteria via

Sec-pathway and Ptl type IV secretion system (14). The B_5-assembly mediates binding of the

secreted AB_5 holotoxin on the surface of various different cell types in a carbohydrate-dependent

manner (13). Subsequent cell entry is followed by dissociation of the B_5-assembly and the PtxS1-

subunit (15), which belongs to the family of ADP-ribosyltransferases (16). PtxS1 ADP-

ribosylates a single C-terminal cysteine residue in α-subunits of most heterotrimeric Gαi-

superfamily members such as Gαi, Gαo, and Gαt (17-19). The C-terminus of the heterotrimeric

G-protein α-subunits makes functionally important contacts with the plasma membrane-localized

G-protein coupled receptors (GPCRs) (20). Therefore, the bulky PtxS1-catalyzed ADP-ribose

modification decouples the G-protein αi-subunit from the GPCRs and inhibits signal propagation

initiated upon agonist stimulation of GPCRs.

Seven-day-old neonatal mice infected with a *B. pertussis* strain expressing a catalytically

inactive mutant of PtxS1 fully survived a challenge, which causes 100% mortality with the

parental strain of *B. pertussis* (11). Purified AB_5 holotoxin containing the same PtxS1 mutant is

incapable of inducing leukocytosis in mice in doses that are 100-fold more than the median lethal

dose of wild-type AB_5 holotoxin (21). It appears, therefore, that the enzymatic activity of PtxS1

is important for the pathological effects of pertussis toxin. In this study, we set out to identify

small molecular weight compounds inhibiting the Gαi-specific ADP-ribosyltransferase activity of

pertussis toxin, and thereby to interfere with the cellular effects of pertussis toxin.

RESULTS

Purification of recombinant PtxS1 from *E. coli* – We did not succeed in purifying the full length N- or C-terminally HIS-tagged recombinant PtxS1 from *E. coli* due to weak solubility and proteolytic instability (data not shown). Therefore, we engineered a double deletion to *ptxA* resulting into expression of an N-terminally HIS-tagged version of *B. pertussis* Tohama I strain PtxS1 (UniProt_P04977), referred hereafter to as rPtxS1, which lacks the N-terminal secretion signal sequence as well as part of the C-terminus (Fig. 1, Fig. 2A). The C-terminal truncation exposes the ADP-ribosyltransferase (ART) catalytic site of PtxS1 including the NAD^+-binding pocket (Fig. 1)(12, 13). Structural data of the pertussis AB_5 holotoxin demonstrate that the C-terminus of PtxS1 masks the active site, and indicate that di-sulfide bond reduction and significant movement of the C-terminus would be required for catalysis (12, 13).

rPtxS1, as purified with metal affinity and size exclusion chromatography, is proteolytically stable and migrates between the 25 and 20 kDa protein markers in SDS-PAGE (Fig. 2B), in accordance with a theoretical size of 23.4 kDa. In order to study the oligomerization state of rPtxS1 in solution, size exclusion chromatography coupled to static light scattering (SEC-MALS) was used. Purified rPtxS1 appeared as a single peak (Fig. 2C), with an estimate of 23.04 ± 0.2 kDa in a triplicate run indicating that rPtxS1 is monomeric in solution. rPtxS1 is well folded as evidenced by a sigmoidal curve with a melting temperature (Tm) of 51°C in a differential scanning fluorimetry (DSF)-assay (Fig. 2D).

Analysis of the ART activity of rPtxS1 – ART activity of rPtxS1 towards Gαi was first studied with a HEK293T cell membrane preparation having an endogenous level of Gαi in an NAD^+-biotin Western blot-assay. rPtxS1 ADP-ribosylated one major protein from the complex

membrane proteome (Fig. 2E). This protein migrated between the 55 and 35 kDa protein markers

in SDS-PAGE (Fig. 2E), in accordance with a theoretical size of endogenous Gαi, e.g. 40.4 kDa

for isoform 1 (UniProt_P63096). Next, ART activity of rPtxS1 towards Gαi was studied with

recombinant N-terminally HIS-tagged Gαi (isoform 1, UniProt_P63096) purified from *E. coli*,

hereafter referred to as rGαi, in an NAD^+ Western blot-assay. rPtxS1 ADP-ribosylated rGαi (Fig.

2F), indicating that Gαi may efficiently serve as a substrate without the G-protein βγ-complex or

other cellular constituents. SEC-analysis of rPtxS1-rGαi complex solution topology shows the

presence of only single monomeric rPtxS1 and rGαi proteins (Fig. S1). rPtxS1-rGαi complex

formation during catalysis therefore appears to be weak, i.e. the rGαi modification is based on a

transient kiss-and-run interaction. Interestingly, rPtxS1 modifies itself (Fig. 2E), and it already

had become auto-ADP-ribosylated in the *E. coli* expression host (Fig. 2F). This activity was

drastically diminished in a Q127D/E129D double mutant of rPtxS1, hereafter referred to as

rPtxS1-mutant (Fig. S2A). The rPtxS1-mutant was also incapable of ADP-ribosylating rGαi (Fig.

S2B). This double mutant of rPtxS1 was analyzed, because structurally identical mutations in a

recently discovered pertussis-like toxin from *E. coli* caused catalytic inactivation, although NAD^+

was still capable of binding to the protein (22)(Fig. S2C). Q127 and E129 of rPtxS1 are also

conserved with several other bacterial ART-toxins where these residues position NAD^+, in

particular the nicotinamide end (Fig. S2C), and promote the transfer of ADP-ribose to a substrate

amino acid residue (16). To assess the substrate amino acid specificity of the extensively

truncated rPtxS1 (see Fig. 1 and Fig. 2A), we analyzed the C351A mutant of rGαi

(UniProt_P63096), hereafter referred to as rGαi-mutant, in an NAD^+-biotin Western blot-assay.

rPtxS1 ADP-ribosylated rGαi, but it was incapable of ADP-ribosylating the rGαi-mutant (Fig.

S3). Based on a DSF-assay, the Tm of the rGαi-mutant was 44.33 ± 0.23 °C as compared to Tm

of the rGαi-wt (Tm of 43.83 ± 0.23 °C), indicating that the C351A mutation does not affect the

protein folding. Therefore, rPtxS1 is not only catalytically active towards rGαi, but it also retains

the Gαi substrate amino acid specificity evidenced with the pertussis AB$_5$ holotoxin (17-19).

Multiwell-based assay set-up for screening of rPtxS1 inhibitors – We analyzed the suitability

of a fluorometric method to screen for rPtxS1 inhibitors previously developed for poly-ADP-

ribose synthesizing enzymes (23) and later extended to mono-ADP-ribose synthesizing enzymes

(24). Briefly, the end point assay is based on conversion of NAD$^+$ to a stable fluorophore with

emission maximum at 444 nm (23). The decrease in fluorescence in comparison to the non-

enzyme control is a measure of NAD$^+$-consuming enzymatic activity (23). Incubation of rPtxS1

with rGαi for 40 min resulted in a strong decrease of fluorescence, almost comparable to the

fluorescence signal obtained with the buffer control where no NAD$^+$ was added to the reaction

(Fig. 3A). No fluorescence decrease was detected with the rPtxS1-mutant under identical

conditions (Fig. 3A). Assays with different concentrations of rPtxS1 were stopped at various time

points to measure NAD$^+$ consumption in the presence of a constant amount of rGαi substrate.

These data demonstrate that an increase in the rPtxS1 concentration increases NAD$^+$

consumption that also progresses over time (Fig. 3B). These results indicate that the detected

reduction in fluorescence is not an effect of fluorescence quenching but indeed NAD$^+$-consuming

activity of rPtxS1. rPtxS1 could also consume NAD$^+$ in the absence of rGαi, although much

slower than with the rGαi substrate (Fig. 3A). This NAD$^+$ glycohydrolase activity, i.e. enzyme-

catalyzed reaction between NAD$^+$ and water to yield ADP-ribose and nicotinamide, was not

detected with the rPtxS1-mutant (Fig. 3A). The effect of DMSO on the NAD$^+$-consuming activity

of rPtxS1 was also studied because small molecules in chemical libraries are usually dissolved in

DMSO typically resulting in assay concentrations of 0.1% – 1% DMSO. DMSO did not have a

significant effect on the NAD^+-consuming activity of rPtxS1 in the presence of rGαi substrate up

to 0.2% DMSO, while 1% DMSO already significantly ($p = 10^{-4}$) reduced activity (Fig. 3C). We

chose to perform screenings in the presence of 0.1% DMSO. To test reproducibility of the

fluorometric assay, maximum (NAD^+ as incubated in plain buffer) and minimum (NAD^+ as

incubated in rPtxS1- and rGαi-containing buffer) signals were measured from five independent

runs to test plate-to-plate and day-to-day variability. The average Z' value for the assay was 0.68,

indicating that the fluorometric assay is suitable for high throughput screening. The statistical

parameters of the assay are summarized in Table S1.

Screening for rPtxS1 inhibitors – We screened a chemical library of 1,695 compounds obtained

from the National Cancer Institute (NCI) Developmental Therapeutics program repository

(https://dtp.cancer.gov/). A total of nine compound hits were identified, which inhibited the

NAD^+-consumption activity of rPtxS1 in the presence of rGαi more than 50% (screening data not

shown). These nine compounds were analyzed in an independent NAD^+-biotin Western blot-

assay, which measures the ADP-ribosylation activity of rPtxS1, i.e. rPtxS1-catalyzed conjugation

of ADP-ribose-biotin onto rGαi. Five out of the nine primary compound hits showed strong

inhibition of the rPtxS1 ADP-ribosylation activity (Fig. 4A). NSC 119875 (Cisplatin, DNA

alkylating agent used in cancer treatments) and NSC 44750 affected rGαi and/or rPtxS1 protein

integrity in the NAD^+-biotin Western-blot based assay (Fig. 4A), even if the assay was performed

with a 10-fold lower compound concentration (data not shown). NSC 119875 and NSC 44750

were therefore excluded from further studies. To evaluate the potency of the remaining three

compounds for rPtxS1 inhibition, dose response studies were performed using the fluorometric

NAD^+-consumption assay in the presence of rGαi. Two of the compounds had an IC_{50}-value of <

10 μM with NSC 228155 and NSC 29193 having an IC_{50}-value of 3.0 and 6.8 μM, respectively

(Fig. 4B). NSC 149286 was less potent than the other two compounds with an IC_{50}-value of 20

μM (Fig. 4B), and was excluded from further analyses. We did not detect NSC 228155- or NSC

29193-induced transition of the rPtxS1 sigmoidal melting curves towards higher temperatures in

a DSF-assay (Fig. S4). This indicates that either the compound binding is not sufficient to alter

the thermal stability of rPtxS1 fold, or that the compound affinity to rPtxS1 is low. Importantly,

the transition of the sigmoidal melting curve towards lower temperatures was also not evident,

indicating that the compounds do not destabilize rPtxS1. The two hit compounds were not

identified as pan-assay interference compounds or aggregators (zinc15.docking.org). In summary,

compound library screening resulted into identification of two compounds (NSC 228155 and

NSC 29193), which inhibited the rPtxS1-catalyzed ADP-ribosylation of rGαi.

Putative binding poses of NSC 29193 and NSC 228155 to PtxS1 − In order to evaluate

possible binding modes of the compounds to PtxS1, we performed docking and molecular

dynamic simulations (MDS). Docking of NSC 228155 produced multiple binding poses but some

features were shared among the top ranked poses, including the placement of the aromatic

pyridine ring of NSC 228155 in an enclosed region where the nicotinamide ring of NAD^+ would

bind to PtxS1, with the double benzoxadiazole ring positioned near the center of the NAD^+-

binding pocket. The benzoxadiazole ring is joined to the pyridine ring via a central sulfur atom,

and the rotatable bonds permit the adoption of varying conformations. In pose 1 (Fig. 5A), with

the lowest free energy of binding (ΔG_{bind} of -51.2 kcal/mol; docking score of -4.08 kcal/mol), the

pyridine ring oxygen of the ligand forms a hydrogen bond with the main-chain nitrogen atom of

Tyr10, anchoring the ligand to an aromatic area of the binding pocket, and with π-π stacking

interactions observed for the pyridine ring with nearby Tyr59 and for the benzoxadiazole ring

with Tyr63. In pose 2 (Fig. 5B), having the best docking score (-4.20 kcal/mol; ΔG_{bind} of -43.3

kcal/mol), the pyridine ring is rotated 90° relative to pose 1, losing the backbone hydrogen bond

with Tyr10, but making π-π interactions with both Tyr63 and Tyr10, and ionic interactions with

Arg9 and Arg67. In MDS, pose 1 showed stability and retains its position with respect to the

original docked position, only differing on average by an RMSD of 0.9 Å (Fig. S5A), and the

RMSD for the ligand orientations differ on average by 2.3 Å when the coordinates of PtxS1 are

superposed over the backbone atoms. In contrast, pose 2 was very unstable at the initial binding

position and occupied different locations of the binding pocket with an average 22 Å RMSD for

the ligand based on superposing the PtxS1 coordinates from the simulation (Fig. S5B).

In contrast to NSC 228155, NSC 29193 is rigid and almost exclusively resulted in two docked

poses. Pose 1 of NSC 29193 is positioned near the binding site for the nicotinamide ring of

NAD$^+$ in the PtxS1, hydrogen bonding with main-chain nitrogen and oxygen atoms of Tyr10

(Fig. 5C). Additionally, NSC 29193 forms π-cation interactions with Arg9, π-π stacking with

Tyr63 and hydrogen bonding with the side-chain hydroxyl group of Ser52. NSC 29193, pose 2, is

bound near the location where the adenine ring of NAD$^+$ would be bound in the pertussis toxin,

stacking against the aromatic ring of Trp26 and hydrogen bonding with the main-chain oxygen

atom of Thr24 (Fig. 5D) as well as forming ionic interactions with Arg9 and Arg13. During

MDS, NSC 29193 pose 1 was quite stable, with an average RMSD of 0.1 and 1.0 Å with respect

to the ligand and receptor backbone atoms (Fig. S5C), and interactions between the ligand and

main-chain atoms of Tyr10 were conserved for over 90% of the simulation time. The π-π

interactions with Tyr63 and π-cation interactions with Arg9 were also retained for the majority of

the simulation time. In contrast, the binding mode of pose 2 was disrupted at around 20 ns of the

simulation (Fig. S5D), mainly due to the reorientation of the indole ring of Trp26, which appears

to be one of the key interaction partners of NSC 29193. Consequently, in pose 2 the ligand moves

from the initial binding position and binds to several areas of the binding pocket. This is reflected

by the average 12 Å RMSD of the ligand with respect to the superposed coordinates of the toxin

from the simulation (Fig. S5D). Taken together, docking and MDS resulted in identification of

plausible binding poses of NSC 228155 and in particular for NSC 29193, most likely owing to

the rigidity of the latter ligand, at the NAD^+-binding pocket of PtxS1.

Inhibitory action of NSC 228155 and NSC 29193 in a living human cell-based assay – First,

to find the optimal toxin dosage, we titrated the pertussis AB_5 holotoxin and detected the ADP-

ribosylation of endogenous Gαi by Western blot-assay using a polyclonal mono-ADP-ribose-

recognizing antibody. Incubation of cells for 2 h with the holotoxin resulted, in a concentration

dependent-manner, mono-ADP-ribosylation of a single protein migrating between the 35 and 55

kDa protein markers in SDS-PAGE (Fig. 6A), in accordance with a theoretical size of

endogenous Gαi, e.g. 40.4 kDa for isoform 1 (UniProt_P63096). Next, we pre-incubated the cells

with the compounds NSC 228155 or NSC 29193 prior to addition of the holotoxin and

subsequent co-incubation. We did not detect any inhibitory action for the compound NSC 29193

despite multiple analyzed conditions (data not shown), e.g. i) 30 min pre-incubation with

inhibitor (0.1, 1, 10 and 50 µM) + 2 h with holotoxin (10 ng/mL) or ii) 1, 2, 3, 4 and 5 h pre-

incubation with 50 µM inhibitor + 2 h with holotoxin (10 ng/mL). However, compound NSC

228155 inhibited the Gαi-specific ADP-ribosylation activity of holotoxin in a concentration-

dependent manner (Fig. 6B). Pre-incubation of the cells for 30 min with 5 µM of NSC 228155

prior to holotoxin addition and subsequent co-incubation resulted in a near complete inhibition of

mono-ADP-ribosylation of Gαi (Fig. 6B). Importantly, NSC 228155 inhibited the holotoxin

activity in concentrations that were not deleterious for the cell viability. First of all, we did not

detect major visual alterations in HEK293T monolayers upon 5 µM NSC 228155 incubation

during the 2.5 h *in vivo* ADP-ribosylation assay period (data not shown). Secondly, cell viability,

as analyzed by the MTT-assay at 3.5 h after addition of NSC 21855, was significantly affected

only after 20 µM or higher concentration was used (Fig. S6A). IC_{50} of the NSC 228155

cytotoxicity in HEK293T was 15.33 µM (Fig. S6A). Thirdly, 5 µM NSC 228155 did not

significantly increase the DNA-damage induced PARP1 (also called ARTD1) poly-ADP-

ribosylation or cell death-associated PARP1/ARTD1 proteolytic processing from the

approximately 120 kDa full length form into approximately 90 kDa form (Fig. S6B). Taken

together, compound NSC 228155 inhibits the AB_5 holotoxin-catalyzed ADP-ribosylation of Gαi

in living HEK293T cells in concentrations that are nor deleterious for the cell viability.

DISCUSSION

We report the identification of the first small molecular weight compounds inhibiting the ADP-

ribosyltransferase activity of pertussis toxin *in vitro* and in living cells. Compound library

screening was made possible due to our ability to purify mg-levels of a truncated but highly

active recombinant PtxS1 (rPtxS1) under native conditions from *E. coli*. Moreover, rPtxS1

retained the high amino acid specificity of pertussis AB_5 holotoxin toward the single C-terminal

cysteine in Gαi (17-19). Used truncation positions in rPtxS1 are functionally justified. PtxS1

contains an N-terminal signal sequence ending at Ala34 (25, 26) that gets cleaved upon the Sec-

mediated secretion of PtxS1 from the cytoplasm into the periplasm where the AB_5 holotoxin is

assembled prior to Ptl type IV secretion system-mediated export. Asp35 is classically numbered

as the first amino acid of mature PtxS1 (see Fig. 2). The truncated part of the PtxS1 C-terminus

(see Fig. 1, 2A) masks the NAD^+-binding pocket in the AB_5 holotoxin involving a stabilizing

intramolecular di-sulfide bond between Cys41 and Cys201 (12, 13). Our truncation approach

bypasses the need for disulfide bond reduction (27) and extensive conformational movement of

the C-terminus (12, 13) to activate PtxS1, which inside the host cell has also been proposed to

involve proteolysis (28).

The inhibitory compounds NSC 29193 and NSC 228155 with low micromolar IC_{50}-values

were identified in the *in vitro* NAD^+ consumption assay from a chemical library of 1,695

compounds. These compounds were also potent in an independent *in vitro* assay where we

monitored the amount of rGαi-conjugated ADP-ribose-biotin upon rPtxS1 catalysis by

streptavidin-HRP Western blotting (see Fig. 4A). It is noteworthy that other compounds were

also positive in the *in vitro* NAD^+ consumption assay, but apparently via deleterious effects for

protein stability as evidenced by the independent *in vitro* ADP-ribose conjugation assay, i.e.

compounds NSC 44750 and NSC 119875 (cisplatin) (see Fig. 4A). We did not detect NSC

228155- or NSC 29193-induced transition of the rPtxS1 sigmoidal melting curves towards higher

temperatures in a DSF-assay (see Fig. S4). This indicates that either the compound binding is not

sufficient to alter the thermal stability of rPtxS1 fold, or the affinity of the compounds to rPtxS1

is low. With respect to NSC 228155, we also attempted to utilize its inherent fluorescence (29) in

a fluorescence polarization assay, but we did not detect significant changes in fluorescence

polarization to make reliable conclusions on the affinity of NSC 228155-rPtxS1 interaction (data

not shown). Our docking and molecular dynamic simulations resulted in variability among poses

for both ligands. More consistent, energetically favorable binding poses were observed for NSC

29193. This is in accordance with the compound structures (see Fig. 4C). NSC 29193 (purine-

2,8-dithiol) is a rather rigid purine analogue that mimics the structure of the adenine base of

NAD^+. NSC 228155, on the other hand, contains two major molecular structures, connected by a

rotatable linker, one ring compound mimicking the adenine base of NAD^+ and the other ring

compound mimicking the nicotinamide of NAD^+. In summary, compound library screening

resulted into the identification of two compounds, NSC 29193 and NSC 228155, which inhibited

the rPtxS1-catalyzed ADP-ribosylation of rGαi *in vitro*.

NSC 228155, but not NSC 29193, was a potent inhibitor of pertussis holotoxin-mediated

ADP-ribosylation of Gαi in living HEK293T cells. We tried multiple different experimental set-

ups with NSC 29193 (see Results section, last chapter), including concentrations as high as 50

µM that still did not cause visible alterations for HEK293T cell monolayers. In respect of

previous drug screening approaches there appears to be little if any published information on

NSC 29193. The cell permeability of NSC 29193 is not known, but might be weak and thus

explain our negative inhibitory data. In sharp contrast, we detected a near complete inhibition of

the ADP-ribosylation with 5 µM NSC 228155 (see Fig. 6). In part, this could relate to the fact

that NSC 228155 easily permeates cells. It has been shown in MDA-MB-468 breast cancer cells

that 5 min incubation with 100 µM NSC 228155 caused rapid movement of this inherently

fluorescent molecule across cell membranes and dispersal to both the cytoplasm and nucleus

(29). However, we witnessed significant toxicity of NSC 228155 for HEK293T cells with 20 µM

or higher concentrations after a 3.5-h incubation (cytotoxicity IC_{50} – 15.33 µM). This cytotoxicity

might be caused by the proposed NSC 228155-mediated production of reactive oxygen species

(ROS) inside the cell (29). In this respect, it is noteworthy that, upon incubation of cells with 5

µM of NSC 228155 for 2.5 h, we did not detect auto-ADP-ribosylation of PARP1/ARTD1 (see

Fig. S6), which is induced in cells upon ROS-induced DNA damage (30). Moreover, we did not

detect caspase-mediated proteolytic PARP1/ARTD1 cleavage (see Fig. S6), which is a robust

readout for the onset of programmed cell death. However, future lead development needs to

address the apparent cell toxicity effects of NCS 228155. Taken together, NSC 228155 is a cell

permeable and potent compound to inhibit pertussis holotoxin-mediated ADP-ribosylation of Gαi

in living cells in low micromolar concentration.

Resurgence of pertussis has been witnessed even in highly vaccinated populations (1-3), and

there are no specific therapeutics to treat whooping cough. Macrolide antibiotics, when

administrated at a very early stage, show therapeutic effects in certain patient subgroups such as

in infants <3 months of age (31). However, macrolide resistant *B. pertussis* strains have been

reported (32, 33). Pertussis toxin is a major virulence factor of *B. pertussis*, and appears to

contribute to the development of whooping cough (6-11). Targeting of pertussis toxin might

prove beneficial in the treatment of whooping cough, especially in young children who still lack

the vaccine-induced protection against pertussis. In this respect, several drug modes of action

could be envisioned, including to inhibit – i) secretion, ii) host cell recognition, iii) endocytosis,

iv) intracellular trafficking, and v) enzymatic activity (i.e. ADP-ribosylation) inside the host cell.

Humanized monoclonal antibodies have been developed that block pertussis toxin cell surface

receptor interaction or the subsequent internalization and retrograde trafficking (34). These

humanized antibodies prevented the characteristic signs of pertussis in mouse and baboon models

(35). Our current study shows that small molecular weight compounds inhibiting the ADP-

ribosyltransferase activity of pertussis toxin might also have therapeutic potential. Similar

findings have been published on other ADP-ribosylating bacterial toxins, such as ExoA and ExoS

of *Pseudomonas aeruginosa* and cholix-toxin of *Vibrio cholerae* (36, 37). We conclude that NSC

29193 and NSC 228155 are useful templates for future lead development to specifically inhibit

pertussis toxin activity, the immediate challenge being to enhance target specificity and to

increase affinity.

MATERIALS AND METHODS

Expression plasmids – *i) rPtxS1-wt* Synthetic DNA fragment (Eurofins Genomics) encoding for amino acids D35-I221 of *Bordetella pertussis* strain Tohama I (UniProt_P04977) was cloned with NdeI and BamHI into pET15b (Novagen) allowing expression of an N-terminally HIS-tagged rPtxS1 (MGSSHHHHHHSSGLVPRGSHM-D35-PtxS1-I221). The D35-I221 truncation positions of PtxS1 are based on (38). Of note, Asp35 is classically numbered as the first amino acid of PtxS1. *ii) rPtxS1-Q161D/E163D* pET15b-rPtxS1 plasmid was linearized with PCR using 5'-phosphorylated oligonucleotide primers (Eurofins Genomics) prAPV-351 (<u>GAT</u>AGCGA<u>T</u>TATCTGGCACACCGGCGCATTCCG, mutagenic nucleotides underlined) and prAPV-352 (GTAGGTGGCCAGCGCGCCGGCGAGGATACG). The PCR product was gel-isolated, re-ligated and transformed to acquire the mutant plasmid. *iii) rGαi-wt* Synthetic DNA fragment (GenScript) encoding for *E. coli* codon-optimized full-length human Gαi (isoform 1, UniProt_P63096-1) was cloned into pNIC-Bsa4 (Structural Genomics Consortium) using the ligation independent cloning method allowing expression of an N-terminally HIS-tagged rGαi (MHHHHHHSSGVDLGTENLYFQS-Gαi). *iii) rGαi-Cys351Ala* rGαi-encoding plasmid was used as a template in PCR to amplify Cys351Ala-mutant-encoding mutant allele using oligonucleotide primers (Eurofins Genomics) prAPV-418 (tacttccaatccATGGGTTGCACCCTGAGCGCGGAA, LIC-cloning overhangs in lowercase) and prAPV-419 (tatccacctttactgTCAGAACAGGCC<u>CGC</u>ATCCTTCAGGTTGTTCTTG, mutagenic nucleotides underlined, LIC-cloning overhangs in lowercase). The PCR product was cloned into pNIC-Bsa4 (Structural Genomics Consortium) using the ligation independent cloning method allowing expression of an N-terminally HIS-tagged rGαi-Cys351Ala mutant similar to rGαi-wt. All expression plasmids were verified by sequencing.

Protein expression and purification – Expression plasmids were transformed into BL21(DE3)

(Novagen) and selected overnight at 37°C on Luria-Bertani (LB) agar with appropriate

antibiotics. Next morning, the bacterial lawn from the LB-plates was transferred into Terrific

broth autoinduction medium (Formedium, AIMTB0205) supplemented with 0.8% (w/v) of

glycerol with appropriate antibiotics. Cultures were grown at 37°C with 250 rpm until optical

density at 600 nm reached 1 (typically 3-5 h), and temperature was reduced to 18°C. Bacteria

were collected after 24 h by centrifugation and were either frozen to -80°C as pellets, in lysis

buffer or directly used for purification. Pefabloc protease inhibitor (Roche, 11585916001) was

added to 0.1 mM in the thawed biomass in lysis buffer [100 mM Hepes (pH 7.5), 500 mM NaCl,

10% (w/v) glycerol, 0.5 mM Tris(2-carboxyethyl)phosphine hydrochloride (TCEP), 10 mM

imidazole]. Samples were sonicated and clarified by centrifugation. Supernatant was loaded to 5

mL HisTrap HP column (GE Healthcare). With rPtxS1 proteins, column was washed with 10

column volumes of wash buffer I and II. Wash buffer I and II have identical compositions to lysis

buffer with the exception of imidazole concentration of 25 and 50 mM, respectively. rPtxS1

proteins were eluted with elution buffer [100mM Hepes (pH 7.5), 500 mM NaCl, 10% (w/v)

glycerol, 0.5 mM TCEP, 500 mM imidazole], concentrated using a 10 kDa cut-off concentrator

(Thermo Scientific) and subjected to size exclusion chromatography on Superdex75 16/600

Hiload Superdex column (GE Healthcare) using SEC buffer [100 mM Hepes (pH 7.5), 500 mM

NaCl, 10% (w/v) glycerol, 0.5 mM TCEP]. With Gαi proteins, HisTrap HP column was washed

with 15 column volumes of wash buffer [20mM Hepes (pH 7.5), 500 mM NaCl, 10% (w/v)

glycerol, 0.5 mM TCEP, 25 mM imidazole] and eluted with elution buffer [20 mM Hepes (pH

7.5), 500 mM NaCl, 10% (w/v) glycerol, 0.5 mM TCEP, 500 mM imidazole]. Fractions were

concentrated using a 10 kDa cut-off concentrator (Thermo Scientific) and further purified by size

exclusion chromatography on Superdex75 16/600 Hiload Superdex column (GE Healthcare) with

SEC buffer [30 mM Hepes (pH 7.5), 350 mM NaCl, 1 mM MgCl$_2$, 0.5 mM TCEP]. Protein

fractions were pooled, concentrated using a 10 kDa cut-off concentrator (Thermo Scientific),

flash frozen and stored in -80°C.

Static light scattering – All experiments were conducted with SEC-MALS buffer [100 mM

Hepes (pH 7.5), 500 mM NaCl, 10% (w/v) glycerol, 0.5 mM TCEP] with a flow rate of 0.150 mL

per minute. Buffer was filtered with 0.1 μm filter to remove small particles. Typically 100 μg of

protein samples were injected into Superdex 200 10/300 increase column (GE Healthcare) by a

Shimadzu autosampler coupled HPLC machine. Light scattering data were recorded using a

multiangle light scattering detector (miniDAWN TREOS, Wyatt technology).   Data were

analyzed using Astra software (Wyatt technology). For complex formation studies, 1:1 molar

ratio of rPtxS1 and rGαi (100 μg in total) was incubated on ice for 2 h and then injected into the

column.

Differential scanning fluorimetry (DSF) – rPtxS1 at a concentration of 0.25 mg/mL was used

in PBS. Protein was incubated with 5 × SYPRO orange (Thermo Scientific) for 10 minutes. The

samples were heated from 20-90°C with 1°C increments (1 min/1°C). The experiment was run on

2500 Real-Time PCR systems (Applied Biosystems). The resulting data was analyzed with

Boltzmann sigmoidal equation using GraphPad (GraphPad software, Inc.). Thermal stability of

rGαi, rGαi-mutant and rPtxS1 alone as well as in the presence of NSC 228155 or NSC 29193,

was analyzed by DSF using a CFX96 Real-Time PCR detection system (Bio-Rad). rGαi, rGαi-

mutant and rPtxS1 at a concentration of 0.2 mg/mL was used in 20 mM Hepes (pH 7.5), 500 mM

NaCl, 10% (w/v) glycerol, 0.5 mM TCEP. The inhibitor concentrations used in the assay ranged

from 50 µM to 1 mM. Samples were incubated with 5 × SYPRO Orange (Thermo Scientific) for

5 min. The samples were heated from 20-90°C with 0.5°C increments (1 min/1°C). Tm-values

were determined by using the CFX96 Real-Time PCR detection system (Bio-Rad) software.

In vitro* ADP-ribosylation assays – *i) enzyme-excess condition Reactions (typically in 120 µL)

contained 10 µM rPtxS1 proteins, 10 µM biotinylated NAD$^+$ (Trevigen, 4670-500-01) or 10 µM

NAD$^+$ (Sigma, N3014) and either 4 µM rGαi proteins or membrane fraction of HEK293T cells

(30 µg of total protein) as the substrate in 100 mM Hepes (pH 7.5), 500 mM NaCl and 10% (w/v)

glycerol. The membrane fraction was prepared essentially as described in (39). 80% confluent 10

cm cell culture plate of HEK293T cells was placed on ice and washed twice with PBS. Cells

were collected by scraping into 1 mL of hypotonic lysis buffer [20 mM Hepes (pH 7.5), 2.5 mM

MgCl$_2$, 1 mM DTT supplemented with Pierce Protease and Phosphatase Inhibitor Mini Tablets

(40 µL/mL of stock solution – one tablet / 2 mL H$_2$O, Thermo Scientific, 88668) and 25 U/mL of

benzonase (Merck-Millipore, 70664-3)]. The cells were incubated at 4°C for 1 h in rotation to

allow them to swell and partially lyse. The partial lysates were drawn 20 times through 27-gauge

needles and centrifuged with low speed (600 × g, 4°C, 10 min) to pellet the nuclei and insoluble

cell debris. The low-speed supernatant was subjected to high-speed centrifugation (16100 × g,

4°C, 30 min) to pellet the membranes. The membranes were resolubilized into 50 µL of 50 mM

Hepes (pH 7.5), 200 mM NaCl, 1 mM EDTA and 10 mM DTT, 0.3% (w/v) SDS and 2% Triton

X-100 supplemented with protease and phosphatase inhibitors, in concentration as described

above. Protein concentration was measured with Bradford assay. The ADP-ribosylation reactions

were carried out at room temperature for 3 h with shaking at 300 rpm. Reactions were stopped by

addition of Laemmli loading dye to 1 × and heating for 10 minutes at 95°C. The samples were

run on SDS-PAGE and transferred to nitrocellulose membranes, followed by blocking with 1%

(w/v) casein blocking buffer (Bio-Rad, 161-0782). Membranes were incubated with streptavidin

conjugated to horse radish peroxidase (1:5000) (GE Healthcare, RPN1231VS) in 1% (w/v) casein

blocking buffer (Bio-Rad, 161-0782) for 3 h at 4°C in rotation and washed thrice with Tris-

buffered saline [10 mM Tris-HCl (pH 7.5), 150 mM NaCl] containing 0.05% Tween 20 (TBST)

for ten minutes each time. Alternatively, after blocking with 4% (w/v) BSA in TBST, membranes

were probed in TBST containing 2% (w/v) BSA (24 - 48 h at 4°C in rotation) for HIS-tagged

rPtxS1 and rGαi proteins with mouse monoclonal anti-HIS (1:1000) (R&D Systems, MAB050),

for Gαi with mouse monoclonal anti-Gαi (1:500) (Santa Cruz Biotechnology, sc-136478) or

mono-ADP-ribose with a rabbit polyclonal anti-mono-ADP-ribose (1:1000) antibody (Hottiger-

laboratory). Primary antibody membranes were washed thrice with TBST containing 2% (w/v)

BSA for ten minutes each time. Primary antibody membranes were incubated with mouse IgG

kappa binding protein conjugated to horseradish peroxidase (1:2500) (sc-516102, Santa Cruz

Biotechnology) or goat anti-rabbit IgG conjugated to horseradish peroxidase (1:2500) (sc-2004,

Santa Cruz Biotechnology) for 3 h at 4°C in rotation and washed thrice with TBST for ten

minutes each time. All membranes were subsequently developed with WesternBright ECL

(Advansta) and imaged on ImageQuant LAS 4000 (GE Healthcare). *ii) substrate-excess*

condition Reactions (typically in 50 μL) contained 50 nM PtxS1 proteins and 500 nM Gαi in 50

mM sodium phosphate (pH 7.0) and 1 μM biotinylated NAD^+ (Trevigen, 4670-500-01). The

reactions were carried at room temperature for 40 minutes. For validating hits from chemical

screening, 10 μM of the indicated compound and 0.1% DMSO (control reaction) were used.

Reactions were stopped by addition of Laemmli loading dye to 1 × and heating for 3 minutes at

90°C. The samples were run on SDS-PAGE and transferred to nitrocellulose membranes,

followed by blocking with 1% (w/v) casein blocking buffer (Bio-Rad, 161-0782). Membranes

were incubated with streptavidin conjugated to horse radish peroxidase (1:7000) (PerkinElmer,

NEL750001EA) in 1% (w/v) casein blocking buffer (Bio-Rad, 161-0782) for 3 h at 4°C in

rotation and washed thrice with TBST for ten minutes each time. Alternatively, for anti-His

blotting, Penta His HRP conjugate (Qiagen, 34460) was diluted 1:5000 in 1% (w/v) casein

blocking buffer (Bio-Rad, 161-0782). Blots were incubated at room temperature for 2 h, followed

by 3 washes with TBST each for 5 minutes. Membranes were subsequently developed with

WesternBright ECL (Advansta) and imaged on ChemiDoc XRS+ (Bio-Rad).

In vitro* NAD$^+$ consumption assay – *i) basic reaction set-up Reactions were carried out in a U-

shaped 96-well black plate (Greiner BioOne, 650209). Typically, reactions were conducted with

50 mM sodium phosphate (pH 7.0) at 25°C with shaking at 300 rpm with a reaction volume of 50

μL. The reactions were stopped by adding 20 μL of 20% acetophenone (diluted with ethanol) and

20 μL of 2M KOH and incubated at room temperature for 10 minutes. 90 μL of formic acid was

added and further incubated for 20 minutes. The plates were read using Tecan infinity M1000 pro

with excitation and emission wavelengths set at 372 and 444 nm, respectively. Maximum signal

was defined as the NAD$^+$ buffer control and the minimum signal was the rPtxS1-catalyzed

reaction in the presence of rGαi. The raw fluorescence values were always subtracted from blank

containing buffer. The assay was optimized to reach 60% NAD$^+$ consumption. The optimized

conditions for the assay are 125 nM rPtxS1, 500 nM NAD$^+$ and 1 μM rGαi. The reaction has

incubation time of 40 minutes at 25°C with shaking at 300 rpm. The DMSO concentration used

was 0.1%. DMSO tolerance test was done with optimized conditions (0.1-5%). For rGαi

substrate-independent NAD$^+$ consumption activity of rPtxS1, identical conditions were used

except that rGαi was excluded with longer incubation time of 60 minutes. Statistical analyses on

typically 8 parallel values was conducted using two-tailed Student's t-test two sample equal

variance (homoscedastic). ***ii) validation of the NAD$^+$ consumption assay*** In order to establish

repeatability of values for maximum and minimum signals between plates, wells and days, five

control plates were tested. Experimental conditions and protein batches used for assay validation

were the same. Three plates were made on one day while second and third days had one plate

each. Each plate had 40 wells for maximum and minimum signals individually with buffer

blanks. The CV% were calculated separately for minimum and maximum signals using (Standard

deviation/average) *100. Assay parameters such as signal-to-noise (S/N), (S/B) signal-to-

background, screening window coefficient (Z') were calculated as described previously (23, 24).

iii) small molecule library screening Each plate had buffer blank, control reaction (minimum

signal, with rPtxS1-wt) and maximum signal (NAD$^+$ and rGαi). Both maximum and minimum

signals contain 0.1% DMSO. To correct for inherent fluorescence of compounds separate

controls with NAD$^+$, Gαi and compounds were prepared. Compounds were tested at a

concentration of 10 μM. Compounds displaying inhibition more than 50% were considered as

hits. Compound libraries for chemical screening were obtained from NCI repository. ***iv) IC$_{50}$***

measurements Assay incubation time was adjusted such that the substrate conversion did not

exceed 30% in order to minimize the effect of reduction in substrate concentration while

maintaining a robust signal. Compounds were tested from a concentration range of (100-0.01

μM) in half-log dilution series. Each plate had buffer blank, positive control where PtxS1-wt is

added which corresponds to 100% activity and negative control (without enzyme) where the

activity was 0%. Control values were included as two half-log units below and above the

inhibitor concentration series for reactions with 0% and 100% activity. IC$_{50}$ values were obtained

by fitting data to log (inhibitor) vs response - variable slope using GraphPad (GraphPad software,

Inc.) Chemical structures were drawn using MarvinSketch 5.11.3 (chemAxon) or ChemDraw

18.0 (Perkin-Elmer).

Structure preparation and molecular docking – The 2.7 Å resolution crystal structure of

pertussis toxin was accessed from the Protein Data Bank (PDB) (PDB_1BCP (15)). Herein, we

considered only the catalytic PtxS1 subunit (1BCP, chain A). Residues 222-269 of PtxS1 were

also removed from the coordinate file (see Fig. 1 and 2). The truncated structure was then

prepared for docking using the protein preparation wizard (40) in Maestro (Schrödinger Release

2019-1: Maestro, Schrödinger, LLC, New York, NY, 2019). Hydrogen atoms were added,

protonation states of ionizable groups were determined and the structure was energy minimized.

A grid outlining the binding pocket was specified based on the superimposition of the PtxS1

structure with the S1 subunit of the pertussis-like toxin structure from *E. coli* with bound NAD$^+$

(PDB_4Z9D; (22)). The two toxins are 30% identical in primary amino acid sequence. The

ligands NSC 228155 (7-nitro-4-(1-oxidopyridin-1-ium-2-yl)sulfanyl-2,1,3-benzoxadiazole) and

NSC 29193 (purine-2,8-dithiol) were prepared for docking using the LigPrep program in Maestro

(Schrödinger Release 2019-1: LigPrep, Schrödinger, LLC, New York, NY, 2019). Possible

ionization states of the ligands at pH 7.0 ± 2.0 were determined and, in the case of NSC 29193,

four tautomeric states were also produced. The ligands were docked to the pertussis S1 structure

using the glide XP and SP methods (41), keeping the protein structure rigid but allowing ligand

flexibility, producing up to 20 poses for each ligand and tautomer. Since the resulting XP docking

poses exhibited few interactions, the SP docking results were pursued further. The poses were

ranked according to the glide docking score and free energy of binding calculations made with

the Prime MM-GBSA module (Schrödinger Release 2019-1: Prime, Schrödinger, LLC, New

York, NY, 2019).

Molecular dynamics simulation – Molecular dynamics simulation (MDS) of the selected

binding poses of NSC 228155 and NSC 29193 bound to the S1 structure was used to study the

dynamics of the protein-ligand complexes using the Desmond program (42) in Maestro. The

complexes were solvated using a TIP3P water model (43) in an octahedral box, with a 10 Å

distance between solute surface atoms and an edge of the box. The systems were neutralized by

adding Na^+ counterions. Additional Na^+/Cl^- ions were added to bring the systems to a 150 mM

salt concentration. The simulations were carried out using the OPLS3e force field (44) at constant

temperature (300 K) and constant pressure (1 atm), which were respectively regulated using the

Nose-Hoover chain thermostat (45) and Martyna-Tobias-Klein barostat (46). Short-range and

long-range interactions were computed with a 9 Å distance cutoff. The RESPA integrator (47)

was employed with a 6.0 fs time step for long-range non-bonded interactions and a 2.0 fs time

step for bonded and short range non-bonded interactions. The systems were relaxed with the

default equilibration protocol in Desmond, followed by a 100 ns production simulation. Energies

were saved every 1.2 ps, whereas coordinates were recorded every 100 ps. The resulting

trajectories were analyzed in terms of ligand stability (as root mean squared deviation, RMSD)

and lifetime of protein-ligand interactions using the Simulation Interactions Diagram application

in Desmond.

In vivo **ADP-ribosylation assay** – HEK293T cells grown in DMEM + 10% FBS were seeded in

3 mL volumes in 6-well plates (500 000 cells/well) in the late afternoon. The next morning fresh

media containing NSC 228155 at concentrations of 0.1, 1 and 5 µM was exchanged to the cells

(0.05% DMSO in all reactions, including control reactions). Cells were incubated for 30 minutes

at 37°C under normal cell culturing conditions, after which 10 ng/mL of pertussis AB_5 holotoxin

(List Biological Laboratories Inc., 179A) was added to the cells and incubation was continued for

2 h. Cells were then transferred on ice and washed twice with 1 × PBS and collected into 70 µL

of lysis buffer [50 mM Tris-HCl (pH 7.5), 400 mM NaCl, 0.1% sodium deoxycholate, 1% NP-40,

75 µM tannic acid (PARG inhibitor), 40 µM PJ34 (PARP inhibitor) supplemented with Pierce

Protease and Phosphatase Inhibitor Mini Tablets (40 µL/mL of stock solution – one tablet / 2 mL

H_2O, Thermo Scientific, 88668). Samples were kept on ice for 30 minutes and centrifuged for 15

minutes, 4°C, 16100 × g. Protein concentration was measured from the supernatants with

Bradford protein assay. The samples were scaled for protein content to allow loading of 30 µg of

total protein per lane in the SDS-PAGE. Laemmli loading dye was added to 1 × and the samples

were boiled for 10 minutes at 95°C. The samples were run on SDS-PAGE and transferred to

nitrocellulose membranes, followed by blocking with 4% (w/v) BSA in TBST. Membranes were

probed in TBST containing 2% (w/v) BSA for mono-ADP-ribose with custom in-house rabbit

polyclonal anti-mono-ADP-ribose (1:1000) (Hottiger-laboratory), for GAPDH with mouse

monoclonal anti-GAPDH (1:1000) (Abcam, ab9484), for Gαi with mouse monoclonal anti-Gαi

(1:500) (Santa Cruz Biotechnology, sc-136478), for poly-ADP-ribose with rabbit polyclonal anti-

PAR (1:1000) (Enzo Life Sciences, ALX-210-890A) and for PARP1 with mouse monoclonal

anti-PARP1 (1:300) (Santa Cruz Biotechnology, sc-8007). Membranes were washed thrice with

TBST containing 2% (w/v) BSA for ten minutes each time. Membranes were incubated in TBST

containing 2% (w/v) BSA with mouse IgG kappa binding protein conjugated to horseradish

peroxidase (1:5000) (sc-516102, Santa Cruz Biotechnology) or goat anti-rabbit IgG conjugated to

horseradish peroxidase (1:5000) (sc-2004, Santa Cruz Biotechnology) for 3 h at 4°C on a rotary

and washed thrice with TBST for ten minutes each time. Membranes were subsequently

developed with WesternBright ECL (Advansta) and imaged on ImageQuant LAS 4000 (GE

Healthcare).

MTT cell viability assay – HEK293T cells grown in DMEM + 10% FBS were seeded in 100 μL

volumes in 96-well plates (20 000 cells/well) in the late afternoon. The next morning fresh media

containing varying concentrations (0, 0.1, 0.5, 1, 2.5, 5, 10, 20, 40, 60 μM) of NSC 228155 was

added in triplicate. In all the wells final concentration of DMSO was 0.6%. Cells were incubated

for 2.5 h at 37°C under normal cell culturing conditions. Cell viability was investigated using the

CellTiter 96 Non-Radioactive Cell Proliferation Assay (MTT) (Promega, G4002) according to

the manufacturer's instructions. Incubation time with the MTT dye solution was 1 h at 37°C

under normal cell culturing conditions. NSC 228155 cytotoxicity IC_{50} value was calculated using

GraphPad (GraphPad software, Inc.) Data are displayed as means and the standard error of the

mean. Data were normalized to cells treated with 0.6% DMSO (negative controls, 100%

viability) and cells killed with 200 μM H_2O_2 (positive control, 0% viability) and expressed as

percentage of these controls. Normalized response is compared to common log of the inhibitor

concentration (μM) and the IC_{50} calculated using a variable slope. Statistical analyses were

conducted using two-tailed Student's t-test two sample equal variance (homoscedastic).

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     FIGURE LEGENDS

Figure 1. PtxS1 of pertussis toxin. A) Surface representation of the crystal structure of PtxS1

(PDB_1BCP, chain A). The truncated C-terminal fragment and the truncation-uncovered ART

active site are shown in yellow and dark grey, respectively. **B)** Possible binding mode of NAD^+

to PtxS1 derived by superimposing the PtxS1 crystal structure (PDB_1BCP, chain A) to the

crystal structure of active site mutant of pertussis-like toxin from *E. coli* with bound NAD^+

(PDB_4Z9D). Color-coding of atoms in NAD^+: yellow, carbon; blue, nitrogen; red, oxygen;

magenta, phosphorus.

Figure 2. **Catalytic activity of rPtxS1. A)** Construct design. rPtxS1 lacks the N-terminal

secretion signal (S) as well as part of the C-terminus (C-term). N-terminus of the mature PtxS1

starting with Asp35 is classically numbered as the first amino acid of PtxS1. **B)** PageBlue-stained

SDS-PAGE gel of SEC-purified rPtxS1. **C)** Representative SEC-MALS result of SEC-purified

rPtxS1. **D)** Representative DSF result of SEC-purified rPtxS1. **E)** *In vitro* ADP-ribosylation assay

(enzyme-excess condition) for rPtxS1 with NAD^+-biotin and HEK293T membrane fraction with

endogenous level of Gαi. Protein-conjugated ADP-ribose-biotin is detected with streptavidin-

HRP. **F)** *In vitro* ADP-ribosylation assay (enzyme-excess condition) for rPtxS1 with NAD^+ and

recombinant N-terminally HIS-tagged Gαi. Protein-conjugated ADP-ribose is detected with a

rabbit polyclonal antibody specific for mono-ADP-ribose (MAR). Same samples were analyzed

in three (Fig. 2E) and two (Fig. 2F) parallel membranes, respectively.

Figure 3. Multiwell fluorometric NAD$^+$ quantitation assay. Decrease in fluorescence over time is a measure of NAD$^+$-consuming enzymatic activity. **A)** rPtxS1 and rPtxS1-mutant were incubated for 40 min in the presence or for 60 min in the absence of rGαi substrate. **B)** Time and concentration dependency of NAD$^+$-consuming enzymatic activity of rPtxS1 in the presence of rGαi substrate. **C)** Effect of DMSO on NAD$^+$-consuming enzymatic activity of rPtxS1 in the presence of rGαi substrate. Statistics based on two-tailed Student's t-test two sample equal variance (homoscedastic).

Figure 4. Evaluation of primary compound hits from rPtxS1 inhibitor screen. A) *In vitro* ADP-ribosylation assay (substrate-excess condition) for N-terminally HIS-tagged rPtxS1 with NAD$^+$-biotin and N-terminally HIS-tagged rGαi. Protein-conjugated biotin-ADP-ribose was detected with streptavidin-HRP. The reactions contained 200-fold molar excess of inhibitors over rPtxS1. Compounds marked with asterisks affect rPtxS1 and/or rGαi protein integrity, and were therefore excluded from further studies. Same samples were analyzed in two parallel membranes. **B)** IC$_{50}$-curves of the hit compounds selected based on Fig. 3A data. **C)** Chemical structures of NAD$^+$ and hit compounds NSC 228155 and NSC 29193 with < 10 μM IC$_{50}$-values.

Figure 5. Prediction of binding poses of NSC 228155 and NSC 29193 to PtxS1. Selected

binding poses 1 **A)** and 2 **B)** of NSC 228155 and binding poses 1 **C)** and 2 **D)** of NSC 29193 to

PtxS1 (PDB_1BCP, chain A), and residues involved in ligand interactions. Binding mode of

NAD$^+$ to PtxS1 in the corresponding area of the NAD$^+$-binding pocket is shown in each panel on

the right (see also Fig. 1). Color-coding of atoms in NAD$^+$: yellow, carbon; blue, nitrogen; red,

oxygen; magenta, phosphorus. Hydrogen bond interactions are shown in dotted lines.

Figure 6. Evaluation of NSC 228155 in a living human cell-based assay. NSC 228155 inhibits

pertussis holotoxin-mediated mono-ADP-ribosylation of Gαi in living HEK293T cells. **A)** Effect

of pertussis holotoxin concentration on mono-ADP-ribosylation of Gαi in 2 h incubation with

living HEK293T cells. **B)** Effect of NSC 228155 concentration on mono-ADP-ribosylation of

Gαi in living HEK293T cells. Inhibitors were added 30 min before starting the 2 h holotoxin

incubation. Protein-conjugated ADP-ribose is detected with a rabbit polyclonal antibody specific

for mono-ADP-ribose (MAR). [#]Western blot of a parallel membrane with the same Fig. 6B

samples, otherwise Fig. 6A and Fig. 6B blots probed, stripped and re-probed in the order of 1)

anti-MAR, 2) anti-GAPDH and 3) anti-Gαi.

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