

Bayesian multivariate reanalysis of large genetic studies identifies many new associations

Michael C. Turchin^{1,3} and Matthew Stephens^{1,2,†}

¹Department of Human Genetics, The University of Chicago

²Department of Statistics, The University of Chicago

³Current Address: Center for Computational Molecular Biology, Department of Ecology And Evolutionary Biology, Brown University

[†]To whom correspondence should be addressed:

mstephens@uchicago.edu

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Abstract

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Genome-wide association studies (GWAS) have now been conducted for hundreds of phenotypes of relevance to human health. Many such GWAS involve multiple closely-related phenotypes collected on the same samples. However, the vast majority of these GWAS have been analyzed using simple univariate analyses, which consider one phenotype at a time. This is despite the fact that, at least in simulation experiments, multivariate analyses have been shown to be more powerful at detecting associations. Here, we conduct multivariate association analyses on 13 different publicly-available GWAS datasets that involve multiple closely-related phenotypes. These data

11 include large studies of anthropometric traits (GIANT), plasma lipid traits
12 (GlobalLipids), and red blood cell traits (HaemgenRBC). Our analyses iden-
13 tify many new associations (433 in total across the 13 studies), many of which
14 replicate when follow-up samples are available. Overall, our results demon-
15 strate that multivariate analyses can help make more effective use of data
16 from both existing and future GWAS.

17 **1 Author Summary**

18 Genome-wide association studies (GWAS) have become a common and powerful
19 tool for identifying significant correlations between markers of genetic variation
20 and physical traits of interest. Often these studies are conducted by comparing
21 genetic variation against single traits one at a time ('univariate'); however, it has
22 previously been shown that it is possible to increase your power to detect significant
23 associations by comparing genetic variation against multiple traits simultaneously
24 ('multivariate'). Despite this apparent increase in power though, researchers still
25 rarely conduct multivariate GWAS, even when studies have multiple traits readily
26 available. Here, we reanalyze 13 previously published GWAS using a multivariate
27 method and find >400 additional associations. Our method makes use of univariate
28 GWAS summary statistics and is available as a software package, thus making it
29 accessible to other researchers interested in conducting the same analyses. We also

30 show, using studies that have multiple releases, that our new associations have high
31 rates of replication. Overall, we argue multivariate approaches in GWAS should
32 no longer be overlooked and how, often, there is low-hanging fruit in the form of
33 new associations by running these methods on data already collected.

34 **2 Introduction**

35 Genome wide association studies (GWAS) have been widely used to identify genetic
36 factors – particularly single nucleotide polymorphisms (SNPs) and copy number
37 variations (CNVs) – associated with human disease risk and other phenotypes
38 of interest ([Price et al., 2015](#); [Visscher et al., 2017](#)). Indeed, at time of writing
39 over 24,000 such associations have been identified as ‘genome-wide significant’
40 ([MacArthur et al., 2017](#)).

41 The vast majority of these many genetic association analyses consider only
42 one phenotype at a time (“univariate association analysis”). This is despite the
43 fact that measurements on multiple phenotypes are often available, and joint as-
44 sociation analysis of multiple phenotypes (“multivariate association analysis”) can
45 substantially increase power ([Jiang and Zeng, 1995](#); [Zhu and Zhang, 2009](#); [Shriner,](#)
46 [2012](#); [Yang and Wang, 2012](#); [Galesloot et al., 2014](#)). There are likely multiple rea-
47 sons for the preponderance of univariate analyses. One possible reason is that
48 initial association analyses are usually performed under tight time constraints,

49 and at a time when many other analysis issues (e.g. quality control, population
50 stratification) are competing for attention. In these conditions it makes sense to
51 focus on the simplest possible approach that will quickly yield new associations,
52 without overly worrying about loss of efficiency. In addition analysts may be legit-
53 imately concerned that deviation from the most widely adopted analysis pipeline
54 may invite unwanted additional reviewer attention.

55 Nonetheless, we believe that multivariate association analysis has an important
56 role to play in making the most of costly and time-consuming GWAS studies. One
57 way forward is to conduct multivariate analyses of previously-published GWAS,
58 checking for additional associations that may have been missed by the initial uni-
59 variate association analyses. This is greatly facilitated by the fact that many
60 GWAS now make summary data from single-SNP tests freely available ([Willer
61 et al., 2013](#); [Wood et al., 2014](#); [Locke et al., 2015](#); [Shungin et al., 2015](#); [Astle et al.,
62 2016](#)), and that simple multivariate analysis can be conducted using such summary
63 data ([Stephens, 2013](#); [Pickrell et al., 2016](#); [Hormozdiari et al., 2016](#)).

64 Here we demonstrate the potential benefits of reanalyzing published GWAS
65 using multivariate methods. Specifically we apply multivariate methods from
66 [Stephens 2013](#) to reanalyze 13 different GWAS whose initial publications reported
67 only univariate results. In most cases our multivariate analyses find many new
68 associations. For example, in GIANT 2014/5 we find over 150 new associations.

69 In studies with multiple data releases, we find that new multivariate associations
70 found in initial releases typically replicate in subsequent releases, supporting that
71 many of the new associations are likely real. We also demonstrate that the multi-
72 variate approach is not equivalent to simply relaxing the univariate GWAS signif-
73 icance threshold. Finally, we point out some limitations of the specific framework
74 we used here, and suggest some alternative strategies that may help address those
75 limitations in future multivariate GWAS analyses.

76 **3 Results**

77 **Multivariate association analyses**

78 To facilitate multivariate association analyses using the methods from [Stephens](#)
79 [2013](#), we implemented them in an R package `bmass` (Bayesian multivariate anal-
80 ysis of summary statistics). The software requires as input univariate GWAS
81 summary statistics, for the same set of SNPs, on d related phenotypes. Then, for
82 each SNP, it attempts to categorize each phenotype as belonging to one of three
83 categories: **Unassociated**, **Directly Associated**, or **Indirectly Associated** with the
84 SNP. The difference between **D** and **I** is that an indirect association disappears
85 after controlling for associations with other phenotypes (see Online Methods and
86 Supplementary Figure 1).

87 For d phenotypes, there are 3^d possible assignments of phenotypes to these 3
88 categories, and each assignment corresponds to a different “model” γ . For example,
89 one model corresponds to the “null” that all phenotypes are **Unassociated**; another
90 model corresponds to the model that all phenotypes are **Directly** associated; an-
91 other model corresponds to just the first phenotype being **Directly** associated, etc.
92 The goal of the association analysis is to determine which of these models is con-
93 sistent with the data and, in particular, to assess overall evidence against the null
94 model.

95 The support in the data for model γ , relative to the null model, is summarized
96 by a Bayes Factor (BF_γ). Large values of BF_γ indicate strong evidence for model
97 γ compared against the null. One advantage of Bayes Factors over p -values is that
98 the Bayes Factors from different models can be easily compared and combined. For
99 example, the overall evidence against the null is given by the (weighted) average
100 of these BFs:

$$\text{BF}_{\text{av}} := \sum_{\gamma} w_{\gamma} \text{BF}_{\gamma} \quad (1)$$

101 where the weights w_{γ} are chosen to reflect the relative plausibility of each model
102 γ . In `bmass` we implemented the Empirical Bayes approach from [Stephens 2013](#)
103 that learns appropriate weights from the data (see Online Methods).

104 Comparisons with published univariate analyses

105 To provide a benchmark against which to compare our multivariate analysis re-
106 sults, we compiled a list of “previous univariate associations”: SNPs that were
107 both reported as significant in the original publication and exceeded the original
108 publication’s definition for genome-wide significance in at least one phenotype in
109 the publicly-available (univariate) summary data analyzed here. This does not
110 include all SNPs reported in every original publication because in some studies
111 SNPs became genome-wide significant only after adding additional samples not
112 included in the publicly available summary data.

113 We used these previous univariate associations to determine a significance
114 threshold for our multivariate associations. Specifically, we declared a multivariate
115 association as significant if its BF_{av} exceeds that of any previous univariate associa-
116 tion’s BF_{av} in the same study (Stephens, 2013). The rationale is that the evidence
117 for these multivariate associations exceeds the evidence for previously-reported
118 genome-wide significant associations, which are generally regarded as likely to be
119 (mostly) real associations.

120 Finally, we defined a list of “new multivariate associations”, which are SNPs
121 that are significant in our multivariate analysis but are not a “previous univariate
122 association”. To avoid double-counting of signals due to linkage disequilibrium
123 (LD), we pruned the list of new multivariate associations so that they are all at

124 least 0.5Mb apart. For additional details, see Online Methods.

125 **Many new loci identified in reanalyzing 13 publicly available** 126 **GWAS studies**

127 We applied `bmass` to 13 publicly available GWAS studies, representing 10 dif-
128 ferent collections of phenotypes (Table 1). Phenotypic collections include blood
129 lipid traits (GlobalLipids: (Teslovich et al., 2010; Willer et al., 2013)), body mor-
130 phological traits (GIANT: (Lango Allen et al., 2010; Speliotes et al., 2010; Heid
131 et al., 2010; Wood et al., 2014; Locke et al., 2015; Shungin et al., 2015)), red blood
132 cell traits (HaemgenRBC: (van der Harst et al., 2012; Astle et al., 2016)), blood
133 pressure traits (International Consortium for Blood Pressure Genome-Wide Asso-
134 ciation et al., 2011; Wain et al., 2011), bone density traits (Zheng et al., 2015), and
135 kidney function traits (Kottgen et al., 2010; Boger et al., 2011). For three of these
136 phenotypic collections (GlobalLipids, GIANT, and HaemgenRBC), two different
137 releases were available from the source consortiums. We conducted basic QC as
138 described in Online Methods.

139 Our multivariate analyses identify, in total, hundreds of new associations. The
140 numbers of previous univariate associations and new multivariate associations are
141 summarized in Figure 1 (see also Supplementary Table 2). For example, we iden-
142 tify 162 new multivariate associations in GIANT2014/5, 65 in GlobalLipids2013,

Dataset	Release	<i>N</i>	Phenotypes
GlobalLipids	2010	95454	LDL, HDL, TC, TG ^a
	2013	188577	LDL, HDL, TC, TG
GIANT	2010	77167	Height, BMI, WHRadjBMI ^b
	2014/5	224459	Height, BMI, WHRadjBMI
HaemgenRBC	2012	135367	RBC, PCV, MCV, MCH, MCHC, Hb ^c
	2016	173480	RBC, PCV, MCV, MCH, MCHC, Hb
ICBP	2011	69395	SBP, DBP, PP, MAP ^d
MAGIC	2010	46186	FstIns, FstGlu, HOMA_B, HOMA_IR ^e
GEFOS	2015	32965	FA, FN, LS ^f
GIS	2014	48972	Iron, Sat, TrnsFrn, Log10Frtn ^g
SSGAC	2016	343072	NEB_Pooled, AFB_Pooled ^h
CKDGen	2010/1	67093	Crea, Cys, CKD, UACR, MA ⁱ
ENIGMA2	2015	30717	ICV, Accumbens, Amygdala, Caudate, Hippocampus, Pallidum, Putamen, Thalamus ^j

Table 1: **Dataset Summary.** *N* is the maximum number of samples contributing to each study.

a - Low-Density Lipoproteins (LDL), High-Density Lipoproteins (HDL), Total Cholesterol (TC), Total Triglycerides (TG)

b - Body Mass Index (BMI), Waist-Hip Ratio adjusted for BMI (WHRadjBMI)

c - Red Blood Cell Count (RBC), Packed Cell Volume (PCV), Mean Cell Volume (MCV), Mean Cell Haemoglobin (MCH), Mean Cell Haemoglobin Concentration (MCHC), Haemoglobin (Hb)

d - Systolic Blood Pressure (SBP), Diastolic Blood Pressure (DBP), Pulse Pressure (PP), Mean Arterial Pressure (MAP)

e - Fasting Insulin (FstIns), Fasting Glucose (FstGlu), Homeostatic Model Assessment of Beta Cell Function (HOMA_B), Homeostatic Model Assessment of Insulin Resistance Function (HOMA_IR)

f - Forearm Bone Mineral Density (FA), Femoral Neck Bone Mineral Density (FN), Lumbar Spine Bone Mineral Density (LS)

g - Serum Iron (Iron), Serum Transferrin Saturation (Sat), Serum Transferrin (TrnsFrn), Log-Transformed Ferritin (Log10Frtn)

h - Number of Children Ever Born, Male & Female (NEB_Pooled), Age at First Birth, Male & Female (AFB_Pooled)

i - Serum Creatine (Crea), Serum Cystatin (Cys), Chronic Kidney Disease (CKD), Urinary Albumin-to-Creatine Ratio (UACR), Microalbuminuria (MA)

j - Intracranial Volume (ICV), specified subcortical brain structures refer to MRI-derived volume measurements for each one

143 and 60 in HaemgenRBC2016. These represent power increases from 10% to 45%
144 compared with previous univariate analyses.

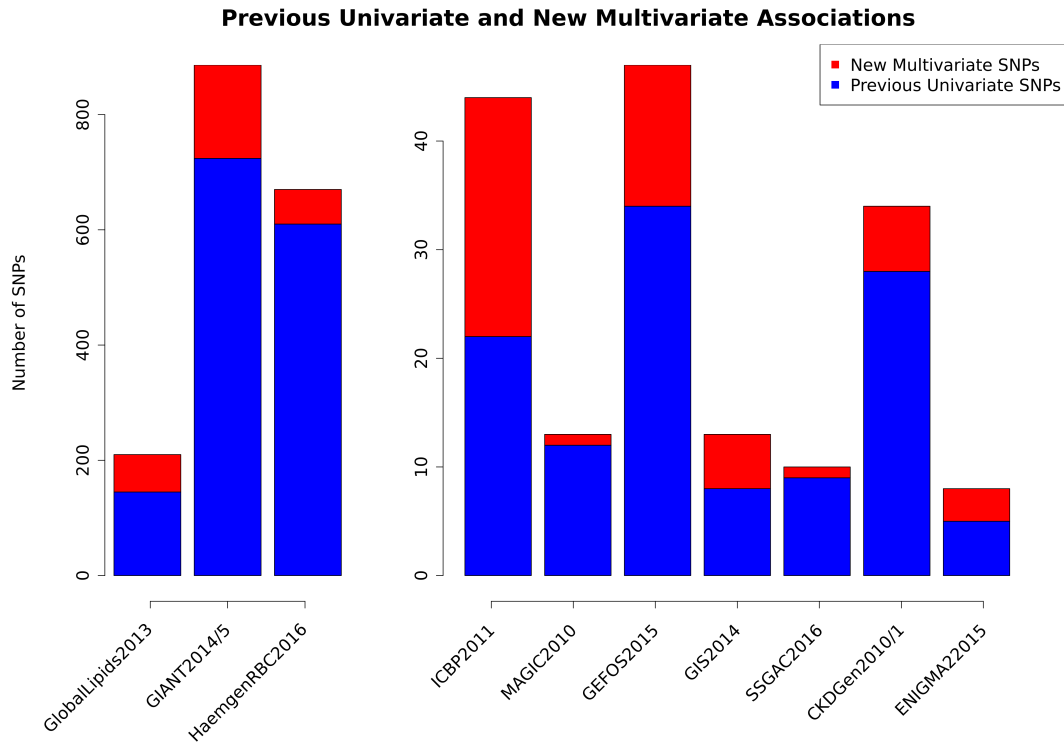


Figure 1: **Number of Independent Significant SNPs, By Study.** The barplot shows the number of independent SNPs that were significant in previous univariate analyses (blue) and the number of additional significant associations in our new multivariate analyses (red). For univariate analysis, significance levels were set by the original study. For multivariate analyses, we declared a SNP to be significant if its weighted average Bayes Factor (BF_{av}) exceeded that of the smallest BF_{av} among the previous univariate significant SNPs. We considered SNPs more than .5Mb apart to be independent. See Table 1 and Online Methods for phenotype details, Online Methods for further analysis details, and Supplementary Tables 2-4 for lists of significant SNPs from each dataset.

Dataset	Release	—SNP Associations—		BF_{av} Thresh	Overlap With Next Release
		Previous Univariate	New Multivariate		
GlobalLipids	2010	102	19	4.35	13/19
	2013	145	65	4.29	-
GIANT	2010	144	60	4.11	49/60
	2014/5	724	162	4.49	-
HaemgenRBC	2012	63	16	5.21	9/16
	2016	610	60	4.68	-
ICBP	2011	22	22	5.24	-
MAGIC	2010	12	1	6.90	-
GEFOS	2015	34	13	5.06	-
GIS	2014	8	5	7.04	-
SSGAC	2016	9	1	5.43	-
CKDGen	2010/1	28	6	4.10	-
ENIGMA2	2015	5	3	7.48	-

Table 2: **Summary of New Multivariate Associations Identified.** Previous Univariate: the number of previous genome-wide significant univariate associations based on the publicly available summary data. New Multivariate: the number of new genome-wide significant multivariate associations. BF_{av} Thresh: the Bayes Factor threshold used in declaring new multivariate associations to be significant. Overlap With Next Release: for GlobalLipids2010, GIANT2010, and HaemgenRBC2012, the last column shows the number of new multivariate associations that overlap with the univariate GWAS associations in the next release from the same consortium; overlap is defined as being within 50kb of the univariate GWAS variant.

145 **Replication of multivariate associations across releases**

146 To demonstrate that many of these new multivariate associations are likely to
147 be real we take advantage of three datasets that each have two releases sepa-
148 rated by several years (GlobalLipids, GIANT, and HaemgenRBC). In each case
149 we performed multivariate association analysis of the earlier release and checked
150 how the new multivariate associations fared in univariate analyses of the later
151 release (Figure 2). Since later releases include the samples from earlier releases,
152 to assess “replication” we focus on whether the association in the new release is
153 more significant than the original release – that is, whether the signal in the new
154 (non-overlapping) samples provides additional evidence *over and above* the original
155 signal. By this measure the results show high replication rates for the new mul-
156 tivariate associations: in total, 84 of 94 new associations have smaller minimum
157 univariate p -values in the later release (at exactly the same SNP), and indeed the
158 majority of these reach univariate GWAS significance in the later release.

159 **Multivariate analysis is different from multiple univariate** 160 **analyses**

161 Because multivariate analysis takes account of *joint* patterns across phenotypes, its
162 ranking of significance of SNPs can change compared with that from the univariate
163 p -values alone. That is, multivariate analysis is not simply equivalent to multiple

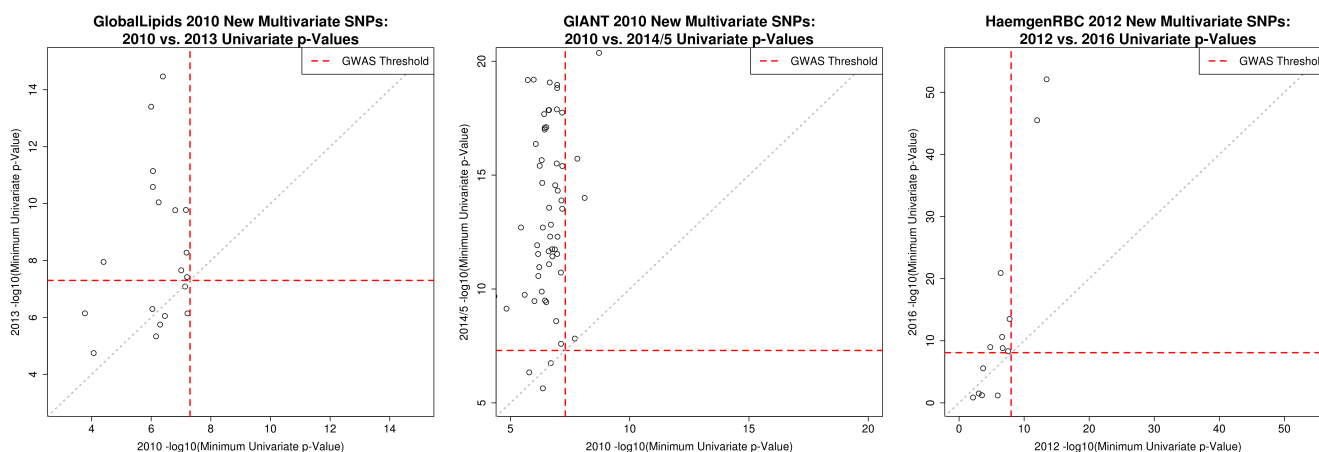


Figure 2: Replication of New Multivariate Associations. The figure shows results based on earlier and later releases from studies with multiple releases (GlobalLipids, GIANT, and HaemgenRBC). Each point represents a new multivariate association identified in our multivariate analysis of the earlier release. The x - and y -axes show the minimum (across phenotypes) of the $-\log_{10}$ univariate p -values from the earlier release (x -axis) vs. the later release (y -axis). Dashed red lines represent the univariate significance GWAS thresholds used for each study's releases. Across all three studies, 84 out of 94 new multivariate associations from the earlier releases have smaller minimum univariate p -values in the later release, and 68 out of 84 new multivariate associations that did not reach GWAS significance in the earlier release do so in the later release (see Supplementary Table 5 for a per-dataset breakdown).

164 univariate analyses. To illustrate this we examined, in three well-powered stud-
165 ies, the associations that would be declared significant if the univariate significance
166 threshold were relaxed, and assessed which of them would also be significant in our
167 multivariate analysis (i.e. we assess whether, if we go deeper into the univariate re-
168 sults, we find the same SNPs as appear in our multivariate results). The results are
169 shown in Figure 3. Although there is, understandably, substantial overlap between
170 the significant SNPs, any non-trivial relaxation of the univariate threshold includes
171 many SNPs that are not multivariate significant in our analysis; for example, at

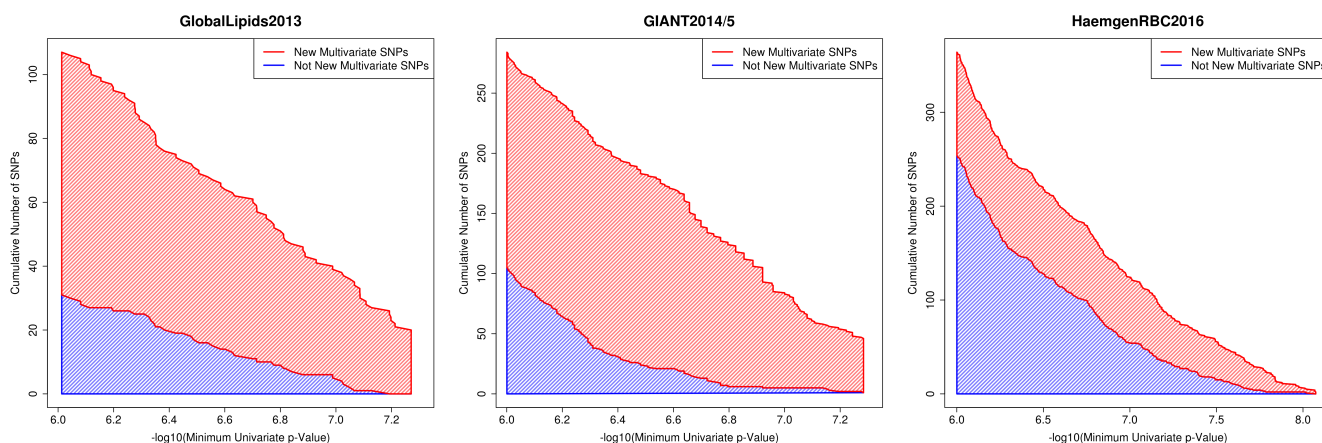


Figure 3: Comparison of New Multivariate Hits vs. Relaxing Univariate p -Value Threshold. For each data set the graph shows how many associations become significant as the univariate p -value threshold is relaxed (moving from right to left on the x -axis), and how many of these are declared as new multivariate hits in our analysis. In both cases results are pruned to avoid counting associations of SNPs in strong LD; see Online Methods for details. The appearance of appreciable blue areas indicates that the multivariate analysis is reordering the significance of SNPs compared with performing multiple univariate analyses.

172 a univariate threshold of 5×10^{-7} only 66% of the univariate significant SNPs are
173 also multivariate significant across these three studies. This demonstrates that,
174 indeed, our multivariate approach reorders significance of SNPs compared with
175 multiple univariate analyses.

176 **Reanalysis also identifies new univariate associations**

177 During our multivariate reanalyses we noticed many SNPs that appeared to be
178 genome-wide univariate significant but were – somewhat mysteriously – not re-
179 ported as such by the original studies (i.e. SNPs whose univariate p -values crossed
180 the significance threshold, as defined by the given study, in at least one trait).

181 Supplementary Table 1 reports 79 such associations.

182 There may be many reasons why such variants went unreported, but one rea-
183 son may be physical proximity to a variant with a stronger signal. Indeed, more
184 than half of the variants described above are within 1Mb of a previously-reported
185 univariate GWAS association. Refraining from reporting multiple near-by associa-
186 tions seems a reasonable – if conservative – strategy to avoid reporting redundant
187 associations due to LD. Further, even when redundant associations due to LD can
188 be ruled out (e.g. by directly examining LD rather than by simply using physi-
189 cal distance), it might be assumed that multiple nearby associated variants may
190 all act through the same biological mechanism and therefore provide redundant
191 biological insights. However, we found that multi-phenotype patterns of associa-
192 tion can differ between nearby SNPs, suggesting that they act through different
193 mechanisms.

194 To highlight just one example, consider rs7515577 – which is an original uni-
195 variate association in GlobalLipids2010 – and rs12038699 – which is a new multi-
196 variate association in GlobalLipids2013. We note that rs12038699 actually reached
197 univariate genome-wide significance in the GlobalLipids2013 dataset, but was not
198 reported (Supplementary Table 6). This is possibly because the latter SNP is rel-
199 atively close, in genomic terms, to the former SNP (549kb). However, these SNPs
200 are not in strong LD ($r^2 = .08$), and so these associations almost certainly repre-

201 sent non-redundant associations. This is further supported by the effect sizes in
202 each phenotype, which clearly reveal very different multivariate patterns of effect
203 sizes among phenotypes (Supplementary Figure 2 & Supplementary Table 6). In-
204 deed the very different multivariate patterns of effect size suggest that not only are
205 these associations non-redundant but likely involve different biological mechanisms
206 as well.

207 These results suggest that, moving forward, it may pay to be more careful
208 in designing filters designed to avoid reporting redundant associations, and that
209 multi-phenotype analyses may have a helpful role to play here.

210 **Limitations**

211 One goal of the multivariate approach introduced in [Stephens 2013](#) was to increase
212 interpretability of multivariate analyses; in particular, the goal was to not only
213 *test* for associations but also to help *explain* the associations by partitioning the
214 phenotypes into “Unassociated”, “Directly Associated”, and “Indirectly Associated”
215 categories. In principle one might hope to use these classifications to gain insights
216 into the relationships among phenotypes and also perhaps to identify different
217 “types” of multivariate association - effectively clustering associations into different
218 groups. However, in practice we find that these discrete classifications are often not
219 as helpful as one might hope. One reason is the difficulty of reliably distinguishing

220 between direct and indirect effects (Stephens, 2013). Another reason is widespread
221 associations with multiple phenotypes. Indeed, we find that, consistently across
222 data sets, the most common multivariate models involve associations – either direct
223 or indirect – with many phenotypes (Supplementary Table 7) and many SNPs
224 are classified as being associated with many phenotypes (Figure 4A). Further,
225 SNPs are very rarely confidently classified as “Unassociated” with any phenotype
226 (Figure 4B). This last observation can be explained by the fact that it is essentially
227 impossible to distinguish ‘unassociated’ from ‘weakly associated’. Nonetheless
228 when all SNPs show similar classifications it is difficult to get insights into different
229 patterns of multivariate association.

230 Moving forward, rather than relying on the discrete classifications of “Unas-
231 sociated”, “Directly Associated”, and “Indirectly Associated” to identify different
232 patterns of multivariate association, we believe it will be more fruitful to use
233 multivariate methods that take a more quantitative approach, such as identifying
234 different patterns of *effect size* (including direction of effect) among phenotypes
235 (Urbut et al., 2017). Focusing on effect sizes has the potential to be much more
236 informative than discrete classification, which can hide effect size differences. For
237 example, when multiple SNPs are classified as associated with all phenotypes,
238 they can still show very different patterns of estimated effect sizes/direction (see
239 Supplementary Figure 3).

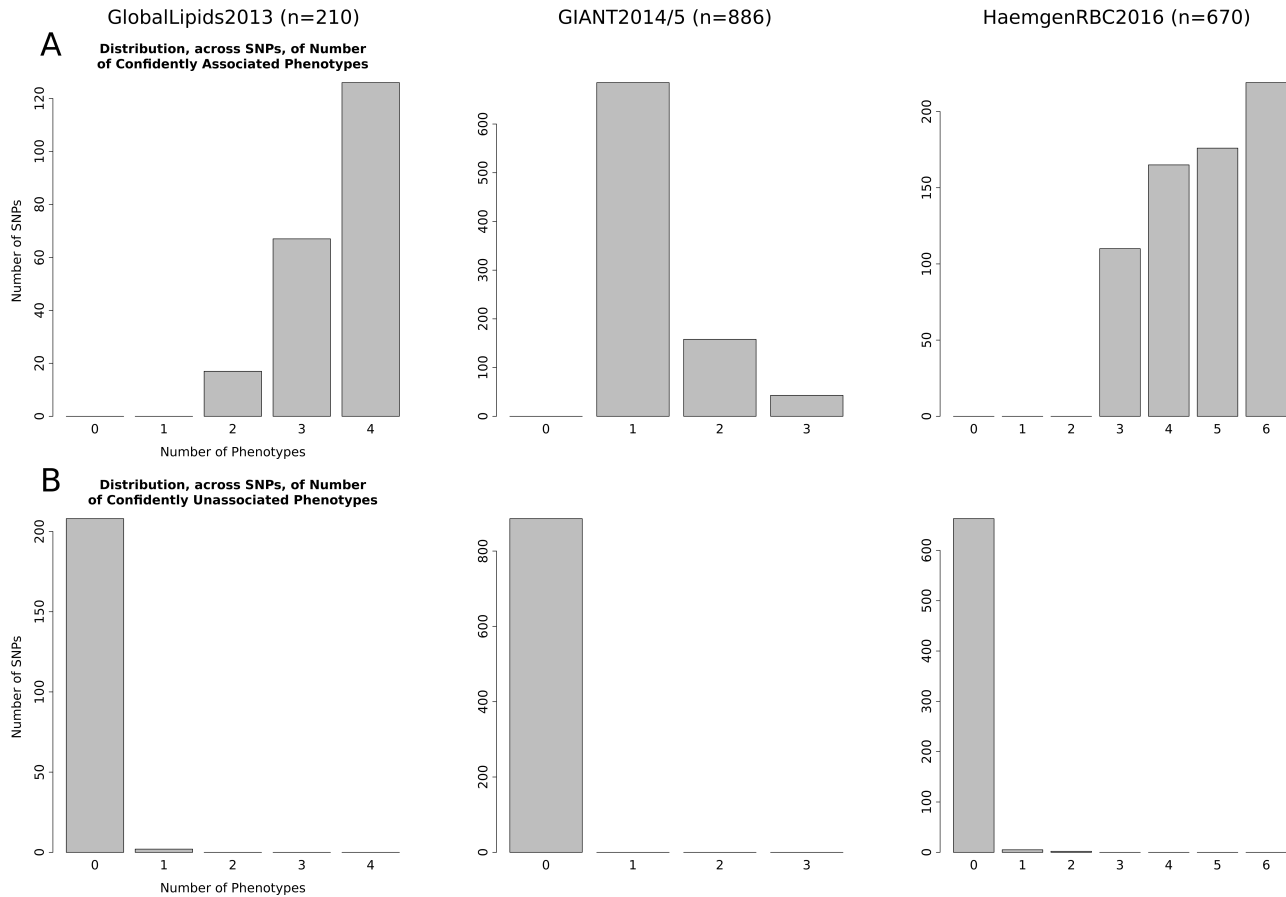


Figure 4: Distribution, Across Significant SNPs, of Number of Phenotypes That Are Confidently Associated (A) or Confidently Unassociated (B). Results are shown for three well-powered datasets: GlobalLipids2013, GIANT2014/5, and HaemgenRBC2016. Here “confident” means with probability > 0.95 , so a SNP is considered “confidently associated” with a phenotype if the sum of its probabilities in the “Directly Associated” and “Indirectly Associated” categories exceeds 0.95 (A), and is considered confidently unassociated with the phenotype if this probability is less than 0.05 (B). The set of significant SNPs includes both previous univariate associations and new multivariate associations.

240 Another limitation of our multivariate methods is that they can lead to (what
241 appear to be) false positive associations when applied to test SNPs with very low
242 minor allele frequencies. Specifically we saw examples where very low-frequency
243 SNPs (e.g. $MAF < .001$) showed strong signals of multivariate association despite
244 showing very little signal in any univariate test. Although such results are not
245 impossible, we believe that most of these cases were likely false positives, and we
246 applied a MAF cut-off (of 0.01 or 0.005) to avoid these issues. Consequently we
247 recommend caution in interpreting results of multivariate analyses at very low-
248 frequency SNPs, and more generally we recommend that multivariate results be
249 compared against univariate results to check they make sense – highly significant
250 multivariate associations that do not also show at least a moderate level of uni-
251 variate association should be treated with caution.

252 4 Discussion

253 We reanalyzed 13 publicly available GWAS datasets using a Bayesian multivari-
254 ate approach and identified many new genetic associations. Turning genetic as-
255 sociations into biological discoveries remains, of course, a challenging problem.
256 Nonetheless, our results suggest that the increased power of multivariate associa-
257 tion analysis that has been reported in many simulation studies ([Stephens, 2013](#);
258 [Galesloot et al., 2014](#); [Porter and O'Reilly, 2017](#)) also translates to discovery of

259 many new associations in practice.

260 Our results exploit the public availability of summary data from several large
261 GWAS. Despite progress toward easier availability of individual-level data for large
262 studies (Sudlow et al., 2015), in many cases summary data remain much easier
263 to obtain and work with; there are big practical advantages as well to modular
264 pipelines that first compute summary data and then use these as inputs to sub-
265 sequent (more sophisticated) analyses. For example, the multivariate analyses we
266 present here are simplified by assuming that the summary data were computed
267 while adequately adjusting for population stratification. And our results illustrate
268 the potential for reanalysis of summary data to yield novel inferences. In this
269 regard we also emphasize the importance of consortia releasing carefully-chosen
270 summaries. For example, Z -scores are much more helpful than p -values because
271 they preserve information on the direction of the effect. Even better would be
272 both the effect size and standard error that created the Z -score. More generally,
273 although not necessarily essential for our analyses here, it is always helpful to in-
274 clude additional key meta-data (e.g. the reference allele, or effect allele, the minor
275 allele frequency, and sample size).

276 The specific multivariate methods used here were derived under the assumption
277 that the summary data from each phenotype has been obtained from the same
278 sampled individuals (which is true, at least approximately, for studies analyzed

279 here). However, multivariate analysis of summary data is also possible even when
280 data were obtained from different samples for each phenotype. The main difference
281 between these settings is that, for data from overlapping samples, the “noise” is
282 correlated as well as the signal: i.e. the summary data are correlated under the null
283 due to sample overlap, and correlated under the alternative due to both sample
284 overlap and any shared genetic effects. In contrast, for data from non-overlapping
285 samples the noise is uncorrelated (whereas the signal may remain correlated if
286 genetic factors are shared). Our methods use data at (empirically) null SNPs
287 to estimate the noise correlation, and so their overall assessment of associations
288 should be relatively robust to whether samples for different phenotypes overlap
289 (however, our definitions of **D** (direct) vs **I** (indirect) associations requires the
290 same samples to be measured across phenotypes.)

291 Moving forward, we expect multivariate association analyses to play an in-
292 creasingly important role in detecting and understanding genetic associations and
293 relationships among phenotypes. Large studies are now collecting, and making
294 available, rich human genetic and phenotypic information on many complex phe-
295 notypes, most notably the UKBioBank ([Sudlow et al., 2015](#)). In addition, there
296 are increasingly large studies linking genetic variation and molecular phenotypes
297 such as gene expression (e.g. the GTEx project ([GTEx Consortium, 2013](#))), as
298 well as epigenetic modifications and transcript degradation ([Gaffney, 2013](#); [Pai](#)

299 [et al., 2015](#); [Birney et al., 2016](#); [Stricker et al., 2017](#)). Analysis of multiple molec-
300 ular traits can help yield insights into causal connections among traits ([Li et al.,](#)
301 [2016](#)), and joint analysis of molecular traits with complex phenotypes may also
302 shed light on functional mechanisms (as in “co-localization” analyses ([Hormozdiari](#)
303 [et al., 2016](#); [Li and Kellis, 2016](#); [Zhu et al., 2016](#); [Wen et al., 2017](#))). Even simply
304 moving from single phenotype to pairwise analysis can shed considerable light on
305 sharing of genetic effects and possible causal connections ([Pickrell et al., 2016](#); [Shi](#)
306 [et al., 2017](#)).

307 These increasingly-complex new data also bring new analytic and computa-
308 tional challenges. Here we have restricted our analyses to relatively small sets
309 of closely-related traits, and indeed the specific multivariate framework we used
310 here – which performs an exhaustive search over all possible multivariate models
311 – is fully tractable for only moderate numbers of traits (up to about 10). Scal-
312 ing methods up to dealing with larger number of traits may well be helpful for
313 some settings, and recent multivariate analysis methods can deal with dozens of
314 outcomes ([Dahl et al., 2016](#); [Urbut et al., 2017](#)). In addition, developing multivari-
315 ate methods to perform *fine-mapping* of genetic associations simultaneously across
316 multiple phenotypes ([Lewin et al., 2016](#)) seems an important and challenging area
317 for future work.

318 5 URLs

319 bmass R package: <https://github.com/mturchin20/bmass>

320 6 Acknowledgments

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326 GM118652.

327 7 Author Contributions

328 MS conceived the original statistical framework. MS and MCT conceived the study
329 design. MCT performed the data collection, processing, and analyses. MCT wrote
330 the R package bmass. MS supervised the project. MCT and MS wrote the paper.

331 8 Materials and Methods

332 8.1 GWAS Datasets

333 Below are specific details regarding retrieval and data-processing for each dataset
334 analyzed. Where applicable, these details include the sample size (N), minor allele
335 frequency (MAF), and p -value thresholds that were applied (based on the thresh-
336 olds used in the original publications). For each dataset variants were dropped if
337 they satisfied at least one of the following criteria: did not contain information for
338 every phenotype; had missing MAF; were fixed (MAF of 0); had effect size exactly
339 0 (i.e. direction of effect would be indeterminable); or did not contain the same
340 reference and alternative alleles across each phenotype. For a handful of studies,
341 external databases were used to retrieve chromosome, basepair information, and
342 MAF based on rsID#; in these studies SNPs for which this information could not
343 be retrieved were also dropped.

344 **GlobalLipids2010 (Teslovich et al., 2010):** Original merged, processed,
345 and GWAS-hit annotated summary data from Stephens 2013 (Stephens, 2013) for
346 HDL, LDL, TG, and TC was downloaded from [https://github.com/stephens999/](https://github.com/stephens999/multivariate)
347 [multivariate](#) (*dtlesssignif.annot.txt* and *RSS0.txt*).

348 **GlobalLipids2013 (Willer et al., 2013):** Summary data for HDL, LDL,
349 TG, and TC was downloaded from <http://csg.sph.umich.edu/abecasis/public/>

350 [lipids2013/](#). We used a minimum N threshold of 50,000, a MAF threshold of
351 1%, and a univariate significant GWAS p -value threshold of 5×10^{-8} . All vari-
352 ants were oriented to the HDL minor allele. The final merged and QC'd datafile
353 contained 2,004,701 SNPs. rsID#'s of published GWAS SNPs were retrieved for
354 all four phenotypes from [https://www.nature.com/ng/journal/v45/n11/full/
355 ng.2797.html](https://www.nature.com/ng/journal/v45/n11/full/ng.2797.html) via Supplementary Tables 2 and 3.

356 **GIANT2010 (Lango Allen et al., 2010; Speliotes et al., 2010; Heid
357 et al., 2010)**: Summary data for Height, BMI, and WHRadjBMI were down-
358 loaded from [https://www.broadinstitute.org/collaboration/giant/index.
359 php/GIANT_consortium_data_files](https://www.broadinstitute.org/collaboration/giant/index.php/GIANT_consortium_data_files). We used a minimum N threshold of 50,000,
360 a MAF threshold of 1%, and a univariate significant GWAS p -value threshold of
361 5×10^{-8} . Chromosome and basepair position per variant were retrieved from
362 dbSNP130 (Sherry et al., 2001). All variants were oriented to the Height minor
363 allele. The final merged and QC'ed datafile contained 2,363,881 SNPs. rsID#'s
364 of published GWAS SNPs were retrieved for Height from [https://www.nature.
365 com/nature/journal/v467/n7317/full/nature09410.html](https://www.nature.com/nature/journal/v467/n7317/full/nature09410.html) via Supplementary
366 Table 1, for BMI from [https://www.nature.com/ng/journal/v42/n11/full/
367 ng.686.html](https://www.nature.com/ng/journal/v42/n11/full/ng.686.html) via Table 1, and for WHRadjBMI from [https://www.nature.com/
368 ng/journal/v42/n11/full/ng.685.html](https://www.nature.com/ng/journal/v42/n11/full/ng.685.html) via Table 1.

369 **GIANT2014/5 (Wood et al., 2014; Locke et al., 2015; Shungin et al.,**

370 **2015**): Summary data for Height, BMI, and WHRadjBMI were downloaded from
371 [https://www.broadinstitute.org/collaboration/giant/index.php/GIANT_consortium_](https://www.broadinstitute.org/collaboration/giant/index.php/GIANT_consortium_data_files)
372 [data_files](#). We used a minimum N threshold of 50,000, a MAF threshold of 1%,
373 and a univariate significant GWAS p -value threshold of 5×10^{-8} . Chromosome
374 and basepair position per variant were retrieved from dbSNP130 (Sherry et al.,
375 2001). All variants were oriented to the Height minor allele. The final merged and
376 QC'ed datafile contained 2,340,715 SNPs. rsID#'s of published GWAS SNPs were
377 retrieved for Height from [https://www.nature.com/ng/journal/v46/n11/full/](https://www.nature.com/ng/journal/v46/n11/full/ng.3097.html)
378 [ng.3097.html](#) via Supplementary Table 1, for BMI from [https://www.nature.](https://www.nature.com/nature/journal/v518/n7538/full/nature14177.html)
379 [com/nature/journal/v518/n7538/full/nature14177.html](#) via Supplementary
380 Tables 1 and 2, and for WHRadjBMI from [https://www.nature.com/nature/](https://www.nature.com/nature/journal/v518/n7538/full/nature14132.html)
381 [journal/v518/n7538/full/nature14132.html](#) via Supplementary Table 4.

382 **HaemgenRBC2012 (van der Harst et al., 2012)**: Summary data for RBC,
383 PCV, MCV, MCH, MCHC, and Hb were downloaded from the European Genome-
384 Phenome Archive via accession number EGAS00000000132 ([https://www.ebi.](https://www.ebi.ac.uk/ega/studies/EGAS00000000132)
385 [ac.uk/ega/studies/EGAS00000000132](#)). We used a minimum N threshold of
386 10,000, a MAF threshold of 1%, and a univariate significant GWAS p -value thresh-
387 old of 1×10^{-8} . Chromosome, basepair position, and MAF per variant were
388 retrieved from HapMap release 22 (International HapMap, 2003). All variants
389 were oriented to the RBC minor allele. The final merged and QC'ed datafile

390 contained 2,327,567 SNPs. rsID#'s of published GWAS SNPs were retrieved
391 for all six phenotypes from [https://www.nature.com/nature/journal/v492/
392 n7429/full/nature11677.html](https://www.nature.com/nature/journal/v492/n7429/full/nature11677.html) via Table 1.

393 **HaemgenRBC2016 (Astle et al., 2016)**: Summary data for RBC, PCV,
394 MCV, MCH, MCHC, and Hb were shared via personal communication with the
395 authors. We used a MAF threshold of 1% and a univariate significant GWAS p -
396 value threshold of 8.319×10^{-9} . Since sample size was not provided per variant, the
397 following overall study sample sizes were used as proxies per phenotype: 172,952 for
398 RBC, 172,433 for PCV, 173,039 for MCV, 172,332 for MCH, for 172,925 MCHC,
399 and 172,851 for Hb. All variants were oriented to the RBC minor allele. Only SNPs
400 were analyzed. The final merged and QC'ed datafile contained 8,649,095 SNPs.
401 We then used these summary data to create a list of (non-redundant) "Previous
402 univariate associations". This was done separately for each phenotype by collecting
403 all SNPs that exceeded the univariate significant GWAS p -value threshold and
404 greedily pruning the SNPs: i.e. we went down the list, removing SNPs that were
405 less significant than another SNP within 500kb. The pruned lists of previous
406 univariate associations for each phenotype were then combined to produce the
407 final SNP list of "published GWAS results". Published CNVs that tagged regions
408 that were not identified by this 'final published SNP list' were also included to
409 avoid erroneously claiming downstream a region as a 'new unpublished result';

410 these CNVs however were only used to mask additional loci as being ‘nearby a
411 published univariate GWAS result’ and for nothing else in the `bmass` analysis
412 pipeline.

413 **ICBP2011 (International Consortium for Blood Pressure Genome-**
414 **Wide Association et al., 2011; Wain et al., 2011)**: Summary data for
415 SBP, DBP, PP, and MAP were downloaded from dbGaP via accession number
416 phs000585.v1.p1 ([https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.](https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000585.v1.p1)
417 [cgi?study_id=phs000585.v1.p1](https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000585.v1.p1)). We used a minimum N threshold of 10,000, a
418 MAF threshold of 1%, and a univariate significant GWAS p -value threshold of $5 \times$
419 10^{-8} . Chromosome and basepair position per variant were retrieved from HapMap
420 release 21 ([International HapMap, 2003](#)). All variants were oriented to the SBP mi-
421 nor allele. The final merged and QC’ed datafile contained 2,387,851 SNPs. rsID#’s
422 of published GWAS SNPs were retrieved for SBP and DBP from [https://www.](https://www.nature.com/nature/journal/v478/n7367/full/nature10405.html)
423 [nature.com/nature/journal/v478/n7367/full/nature10405.html](https://www.nature.com/nature/journal/v478/n7367/full/nature10405.html) via Supple-
424 mentary Table 5, and for PP and MAP from [https://www.nature.com/ng/journal/](https://www.nature.com/ng/journal/v43/n10/full/ng.922.html)
425 [v43/n10/full/ng.922.html](https://www.nature.com/ng/journal/v43/n10/full/ng.922.html) via Table 1 and Supplementary Table 2F. Addition-
426 ally, we gratefully acknowledge the International Consortium for Blood Pressure
427 Genome-Wide Association Studies (Nature. 2011 Sep 11;478(7367):103-9, Nat
428 Genet. 2011 Sep 11;43(10):1005-11) for generating and sharing these data.

429 **MAGIC2010 (Dupuis et al., 2010)**: Summary data for FstIns, FstGlu,

430 HOMA_B, and HOMA_IR were downloaded from <https://www.magicinvestigators.org/downloads/>. We used a MAF threshold of 1% and a univariate significant
431 GWAS p -value threshold of 5×10^{-8} . Since sample size was not provided per
432 variant, the overall study sample size of 46,186 was used as a proxy. Chromo-
433 some and basepair position per variant were retrieved from HapMap release 22
434 ([International HapMap, 2003](#)). All variants were oriented to the FstIns minor
435 allele. The final merged and QC'ed datafile contained 2,333,328 SNPs. rsID#'s
436 of published GWAS SNPs were retrieved for all four phenotypes from <https://www.nature.com/ng/journal/v42/n2/full/ng.520.html>
437 via Table 1.

439 **GEFOS2015 ([Zheng et al., 2015](#))**: Summary data for FA, FN, and LS were
440 downloaded from <http://www.gefos.org/?q=content/data-release-2015>. We
441 used a MAF threshold of .5% and a univariate significant GWAS p -value threshold
442 of 1.2×10^{-8} . Since sample size was not provided per variant, the overall study sam-
443 ple size of 32,965 was used as a proxy. All variants were oriented to the FA minor
444 allele. The final merged and QC'ed datafile contained 8,938,035 SNPs. rsID#'s of
445 published GWAS SNPs were retrieved for all four phenotypes from <https://www.nature.com/nature/journal/v526/n7571/full/nature14878.html>
446 via Supple-
447 mentary Table 13.

448 **GIS2014 ([Benyamin et al., 2014](#))**: Summary data for Iron, Sat, TrnsFrn,
449 and Log10Frtn were shared via personal communication with the authors. We

450 used a MAF threshold of 1% and a univariate significant GWAS p -value threshold
451 of 5×10^{-8} . Since sample size was not provided per variant, the overall study
452 sample size of 48,972 was used as a proxy. All variants were oriented to the Iron
453 minor allele. The final merged and QC'ed datafile contained 1,985,313 SNPs.
454 rsID#'s of published GWAS SNPs were retrieved for all four phenotypes from
455 <https://www.nature.com/articles/ncomms5926/> via Table 1.

456 **SSGAC2016 (Barban et al., 2016)**: Summary data for NEB_Pooled and
457 AFB_Pooled were downloaded from <https://www.thessgac.org/data>. We used
458 a MAF threshold of 1% and a univariate significant GWAS p -value threshold of
459 5×10^{-8} . Since sample size was not provided per variant, the following overall
460 study sample sizes were used as proxies per phenotype: 251,151 for NEB_Pooled
461 and 343,072 for AFB_Pooled. All variants were oriented to the NEB_Pooled
462 minor allele. The final merged and QC'ed datafile contained 2,395,561 SNPs.
463 rsID#'s of published GWAS SNPs were retrieved for all four phenotypes from
464 <https://www.nature.com/ng/journal/v48/n12/full/ng.3698.html> via Table
465 1.

466 **CKDGen2010/1 (Kottgen et al., 2010; Boger et al., 2011)**: Summary
467 data for Crea, Cys, CKD, UACR, and MA were downloaded from [https://www.](https://www.nhlbi.nih.gov/research/intramural/researchers/pi/fox-caroline/datasets)
468 [nhlbi.nih.gov/research/intramural/researchers/pi/fox-caroline/datasets](https://www.nhlbi.nih.gov/research/intramural/researchers/pi/fox-caroline/datasets).
469 We used a MAF threshold of 1% and a univariate significant GWAS p -value thresh-

470 old of 5×10^{-8} . Since sample size was not provided per variant, the following
471 overall study sample sizes were used as proxies per phenotype: 67,093 for Crea,
472 20,957 for Cys, 62,237 for CKD, 31,580 for UACR, and 30,482 for MA. All vari-
473 ants were oriented to the Crea minor allele. The final merged and QC'ed datafile
474 contained 2,333,498 SNPs. rsID#'s of published GWAS SNPs were retrieved for
475 Crea, Cys, and CKD from [https://www.nature.com/ng/journal/v42/n5/full/
476 ng.568.html](https://www.nature.com/ng/journal/v42/n5/full/ng.568.html) via Table 2.

477 **ENIGMA22015 (Hibar et al., 2015)**: Summary data for ICV, Accum-
478 bens, Amygdala, Caudate, Hippocampus, Pallidum, Putamen, and Thalamus were
479 downloaded from <http://enigma.ini.usc.edu/research/download-enigma-gwas-results/>.
480 We used a minimum N threshold of 10,000, a MAF threshold of 1% and a uni-
481 variate significant GWAS p -value threshold of 5×10^{-8} . All variants were oriented
482 to the ICV minor allele. The final merged and QC'ed datafile contained 6,271,117
483 SNPs. rsID#'s of published GWAS SNPs were retrieved for all 8 phenotypes from
484 <https://www.nature.com/nature/journal/v520/n7546/full/nature14101.html>
485 via Table 1.

486 8.2 bmass

487 **bmass** implements in an R package the statistical methods described in [Stephens](#)
488 [2013](#), which should be consulted for full details. In particular, the sections “Com-

489 putation” and “Detailed Methods (Global Lipids Analysis)” in [Stephens 2013](#) de-
490 scribe how multivariate analyses are applied to GWAS summary data, and `bmss`
491 implements the data analysis pipeline described in the “Detailed Methods (Global
492 Lipids Analysis)” section. The `bmss` R package also includes two vignettes to help
493 users begin processing GWAS summary data and implementing these methods.

494 **8.3 Additional Details for Figure 3**

495 For each dataset we made a list of “marginally-significant” SNPs, with p -values
496 smaller than 1×10^{-6} but not genome-wide significant at the relevant datasets’
497 GWAS threshold. We then greedily pruned these lists of marginally-significant
498 SNPs: that is we repeatedly went through the lists removing SNPs that were less
499 significant than another SNP within 500kb. We then removed any SNPs that were
500 within 500kb of a new multivariate association, and merged the resulting list with
501 the list of new multivariate associations, and sorted this merged list of SNPs by
502 their minimum univariate p -value.

503 This results in a non-redundant list of marginally-significant SNPs – some of
504 which are new multivariate associations and some of which are not – sorted by
505 their smallest univariate p -value. The plot shows how the number of SNPs of each
506 type varies as the p -value threshold is relaxed from the GWAS threshold to 10^{-6}
507 (the HaemgenRBC2016 results show only the top 500 SNPs due to the abundance

508 of SNPs between 8.31×10^{-9} and 1×10^{-6}).

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960 9 Figure Legends

961 **Figure 1: Number of Independent Significant SNPs, By Study.** The barplot
962 shows the number of independent SNPs that were significant in previous univariate
963 analyses (blue) and the number of additional significant associations in our new
964 multivariate analyses (red). For univariate analysis, significance levels were set by
965 the original study. For multivariate analyses, we declared a SNP to be significant
966 if its weighted average Bayes Factor (BF_{av}) exceeded that of the smallest BF_{av}
967 among the previous univariate significant SNPs. We considered SNPs more than
968 .5Mb apart to be independent. See Table 1 and Online Methods for phenotype
969 details, Online Methods for further analysis details, and Supplementary Tables 2-4
970 for lists of significant SNPs from each dataset.

971 **Figure 2: Replication of New Multivariate Associations.** The figure
972 shows results based on earlier and later releases from studies with multiple re-
973 leases (GlobalLipids, GIANT, and HaemgenRBC). Each point represents a new
974 multivariate association identified in our multivariate analysis of the earlier release.
975 The x - and y -axes show the minimum (across phenotypes) of the $-\log_{10}$ univariate
976 p -values from the earlier release (x -axis) vs. the later release (y -axis). Dashed red
977 lines represent the univariate significance GWAS thresholds used for each study's
978 releases. Across all three studies, 84 out of 94 new multivariate associations from
979 the earlier releases have smaller minimum univariate p -values in the later release,

980 and 68 out of 84 new multivariate associations that did not reach GWAS signifi-
981 cance in the earlier release do so in the later release (see Supplementary Table 5
982 for a per-dataset breakdown).

983 **Figure 3: Comparison of New Multivariate Hits vs. Relaxing Univariate**
984 **ate p -Value Threshold.** For each data set the graph shows how many associ-
985 ations become significant as the univariate p -value threshold is relaxed (moving
986 from right to left on the x -axis), and how many of these are declared as new mul-
987 tivariate hits in our analysis. In both cases results are pruned to avoid counting
988 associations of SNPs in strong LD; see Online Methods for details. The appearance
989 of appreciable blue areas indicates that the multivariate analysis is reordering the
990 significance of SNPs compared with performing multiple univariate analyses.

991 **Figure 4: Distribution, Across Significant SNPs, of Number of Phe-**
992 **notypes That Are Confidently Associated (A) or Confidently Unassoci-**
993 **ated (B).** Results are shown for three well-powered datasets: GlobalLipids2013,
994 GIANT2014/5, and HaemgenRBC2016. Here “confident” means with probability
995 > 0.95 , so a SNP is considered “confidently associated” with a phenotype if the
996 sum of its probabilities in the “Directly Associated” and “Indirectly Associated”
997 categories exceeds 0.95 (A), and is considered confidently unassociated with the
998 phenotype if this probability is less than 0.05 (B). The set of significant SNPs

999 includes both previous univariate associations and new multivariate associations.

1000 10 Supporting Information Legends

1001 Supplementary Figure 1: **Graphical Model of Multivariate Categories**. Shown
1002 here is a Directed Acyclic Graphical (DAG) model of our multivariate categories
1003 in the context of our vector of phenotypes \mathbf{Y} (e.g. $\mathbf{Y} = \{\mathbf{Y}_U, \mathbf{Y}_D, \mathbf{Y}_I\}$) and their
1004 connections with the variant of interest \mathbf{g} . The relationships described in-text
1005 can be seen here. \mathbf{Y}_U , our unassociated phenotypes, have no connection with \mathbf{g} .
1006 \mathbf{Y}_D , our directly associated phenotypes, have a direct connection with \mathbf{g} . And
1007 \mathbf{Y}_I , our indirectly associated phenotypes, have a connection with \mathbf{g} only by going
1008 through \mathbf{Y}_D first. Note that if \mathbf{Y}_D were not observed, \mathbf{Y}_I would appear as a direct
1009 connection.

1010 Supplementary Figure 2: **Refining Association Signals – GlobalLipids2013**
1011 **rs7515577 & rs12038699**. Shown are the $-\log_{10}$ univariate p -values from the
1012 GlobalLipids2013 analysis for both the previous univariate association rs7515577
1013 (“Previous Univariate SNP”) and the new multivariate association rs12038699
1014 (“New Multivariate SNP”) across all four phenotypes analyzed. rs7515577 is repre-
1015 sented as a triangle and rs12038699 is represented as a square. Also shown are the
1016 $-\log_{10}$ univariate p -values of SNPs within 1Mb of the midpoint between rs7515577

1017 and rs12038699. Color-coding of the SNPs represent the degree of linkage disequi-
1018 librium between variants and the new association rs12038699 based on the GBR
1019 cohort of 1000Genomes ([Genomes Project et al., 2015](#)); for color coding details,
1020 see legend.

1021 **Supplementary Figure 3: Effect Size Heterogeneity Among SNPs With**
1022 **Identical Multivariate Model Assignments.** Shown are the phenotype effect
1023 sizes (points), and ± 2 standard errors (bars), for four significantly associated SNPs
1024 from HaemgenRBC2016. All four SNPs were classified as being “associated” with
1025 all six phenotypes (i.e. marginal posterior probability of association $\geq 95\%$ for
1026 each phenotype). However, they clearly show different patterns of effect sizes.
1027 Therefore focusing simply on binary calls of “associated” vs “unassociated” can
1028 hide different patterns of multivariate association.

1029 **Supplementary Table 1: Summary of Associations in Each Dataset.**

1030 ^a Number of new multivariate associations discovered by our analysis. Note that
1031 we required a multivariate association to be at least 500kb from a previous re-
1032 ported association to be considered “new”.

1033 ^b Univariate GWAS significance p -value threshold used by the original study pub-
1034 lication.

1035 ^c These are new multivariate SNPs that were not reported by the original study

1036 despite having a univariate association (in the public summary data) that was
1037 genome-wide significant by the original study's univariate significance threshold.

1038 ^d A “previous association” means an association reported by the original GWAS;
1039 “near” means within 1Mb (but these are all more than 500kb away from a previous
1040 association since our classification of new multivariate SNPs requires this).

1041 Supplementary Tables 2a-m: **Lists of New *bmass* Multivariate Associa-**
1042 **tions, per Dataset.** Attached Excel sheets list new *bmass* associations for each
1043 dataset analyzed.

1044 Supplementary Tables 3a-m: **Lists of Retrieved Univariate Associations**
1045 **From Original Publications, per Dataset.** Attached Excel sheets list the
1046 rsID#'s of the univariate significant SNPs that were retrieved from the original
1047 publication(s) associated with each dataset (see Online Methods for details).

1048 Supplementary Tables 4a-m: **Results for Previous Univariate Associa-**
1049 **tions, per Dataset.** Attached Excel sheets give *bmass* results for previous uni-
1050 variate associations, per dataset. Note that these results may not include all SNPs
1051 from Tables 3a-m, because some SNPs were dropped during QC and other SNPs
1052 were dropped because they did not reach univariate significance in the publicly
1053 available summary data (see Online Methods for details).

1054 **Supplementary Table 5: Replication of New Multivariate Associations.**
1055 Shown are example metrics of how well our new multivariate associations replicate
1056 in datasets that allow such an evaluation. Specifically, for three of the studies
1057 used (GlobalLipids, GIANT, and HaemgenRBC), there are multiple dataset re-
1058 leases. To examine how well our new multivariate **bmss** associations replicate,
1059 we compared the results from the first releases (“1st”) with the univariate GWAS
1060 associations of the second releases (“2nd”). In essence, each of these approaches aim
1061 to increase power – one by using a multivariate approach (**bmss**) and the other by
1062 increasing sample size (the 2nd releases) – thus allowing us to compare the results
1063 against one another. Univariate *p*-Value Threshold: univariate GWAS significance
1064 *p*-value thresholds used by the original publication(s) for both the earlier (1st) and
1065 later (2nd) releases. New Multivariate SNPs in 1st: number of new multivariate
1066 associations from the earlier release. Lower Univariate *p*-Value in 2nd: number of
1067 new multivariate associations from the earlier release that also have lower univari-
1068 ate *p*-values in the later release. Below 2nd Univariate Threshold: number of new
1069 multivariate associations from the earlier release that also cross the later release’s
1070 univariate GWAS significance threshold.

1071 **Supplementary Table 6: *p*-Values for rs7515577 & rs12038699 in 2010**
1072 **and 2013 GlobalLipids Releases** – In the 2010 release rs7515577 has a univari-
1073 ate *p*-value that crosses the 5×10^{-8} threshold (TC), whereas rs12038699 does not.

1074 Since rs12038699 is near to rs7515577 it may get masked for future analyses; how-
1075 ever in the 2013 data rs12038699 not only has a lower minimum univariate p -value,
1076 but also has a different multivariate p -value pattern as compared to rs7515577.
1077 Both these signals suggest that rs12038699 should be viewed as a separate GWAS
1078 hit for GlobalLipids2013.

1079 **Supplementary Table 7: Top Multivariate Model Examples per SNP.**

1080 List of multivariate models that most frequently have the highest posterior prob-
1081 abilities per SNP. Top 5 models are shown from across both the previous uni-
1082 variate associations analyzed and the new multivariate associations discovered in
1083 the GlobalLipids2013, GIANT2014/5, and HaemgenRBC2016 datasets. Pheno-
1084 type ordering is shown in the header, where 0, 1, and 2 refer to the multivariate
1085 categories of **Unassociated**, **Directly Associated**, and **Indirectly Associated**. n is
1086 the number of SNPs that show the specified model as having the largest posterior
1087 probability, with Mean Posterior displaying the average posterior probability of
1088 the given model across the n SNPs, and Original Prior showing the prior estab-
1089 lished for the given model from training on all the previous univariate associations
1090 from that dataset.

1091 11 Tables

1092 Table 1: **Dataset Summary.** N is the maximum number of samples contributing
1093 to each study.

1094 a - Low-Density Lipoproteins (LDL), High-Density Lipoproteins (HDL), Total
1095 Cholesterol (TC), Total Triglycerides (TG)

1096 b - Body Mass Index (BMI), Waist-Hip Ratio adjusted for BMI (WHRadjBMI)

1097 c - Red Blood Cell Count (RBC), Packed Cell Volume (PCV), Mean Cell Volume
1098 (MCV), Mean Cell Haemoglobin (MCH), Mean Cell Haemoglobin Concentration
1099 (MCHC), Haemoglobin (Hb)

1100 d - Systolic Blood Pressure (SBP), Diastolic Blood Pressure (DBP), Pulse Pres-
1101 sure (PP), Mean Arterial Pressure (MAP)

1102 e - Fasting Insulin (FstIns), Fasting Glucose (FstGlu), Homeostatic Model As-
1103 sessment of Beta Cell Function (HOMA_B), Homeostatic Model Assessment of
1104 Insulin Resistance Function (HOMA_IR)

1105 f - Forearm Bone Mineral Density (FA), Femoral Neck Bone Mineral Density (FN),
1106 Lumbar Spine Bone Mineral Density (LS)

1107 g - Serum Iron (Iron), Serum Transferrin Saturation (Sat), Serum Transferrin
1108 (TrnsFrn), Log-Transformed Ferritin (Log10Frtn)

1109 h - Number of Children Ever Born, Male & Female (NEB_Pooled), Age at First
1110 Birth, Male & Female (AFB_Pooled)

1111 i - Serum Creatine (Crea), Serum Cystatin (Cys), Chronic Kidney Disease (CKD),
1112 Urinary Albumin-to-Creatine Ratio (UACR), Microalbuminuria (MA)
1113 j - Intracranial Volume (ICV), specified subcortical brain structures refer to MRI-
1114 derived volume measurements for each one

1115 **Table 2: Summary of New Multivariate Associations Identified.** Pre-
1116 vious Univariate: the number of previous genome-wide significant univariate as-
1117 sociations based on the publicly available summary data. New Multivariate: the
1118 number of new genome-wide significant multivariate associations. BF_{av} Thresh:
1119 the Bayes Factor threshold used in declaring new multivariate associations to be
1120 significant. Overlap With Next Release: for GlobalLipids2010, GIANT2010, and
1121 HaemgenRBC2012, the last column shows the number of new multivariate asso-
1122 ciations that overlap with the univariate GWAS associations in the next release
1123 from the same consortium; overlap is defined as being within 50kb of the univariate
1124 GWAS variant.