Title:

Highly diverse mesophotic reef fish communities in Raja Ampat, West Papua

Dominic A. Andradi-Brown[1,2]*, Angela J. E. Beer[3,4], Luigi Colin[1], Hastuti[3,5], Catherine E. I.

Head[1,6], Nur Ismu Hidayat[3], Steven J Lindfield[7], Catherine R. Mitchell[1,2], Defy N. Pada[3],

Nikola M. Piesinger[1], Purwanto[8], Gabby N. Ahmadia[2]

[1]Department of Zoology, University of Oxford, Oxford, OX1 3PS, UK

[2]Ocean Conservation, World Wildlife Fund–US, 1250 24th St. NW, Washington, D.C., 20037,

USA

[3]Conservation International Indonesia, Sorong, Papua Barat, 98414, Indonesia

[4]School of Environment and Sustainability, Royal Roads University, Victoria, British

Columbia, V9B 5Y2, Canada

[5] Marine Science, Hasanuddin University, Makassar, Sulawesi Selatan, 90245, Indonesia

[6]Institute of Zoology, Zoological Society of London, Regent's Park, London, NW1 4RY, UK

[7]Coral Reef Research Foundation, PO Box 1765 Koror, PW, 96940, Palau

[8]Universitas Papua, Manokwari, Papua Barat, 98314, Indonesia

*Corresponding author: dominic.andradi-brown@wwfus.org

Abstract

Mesophotic coral ecosystems (MCEs; reefs 30-150 m depth) are poorly studied, with existing research heavily geographically biased away from the most species-rich reef regions. Yet, MCEs are of high interest because of their unique species and potential to act as refuges from the impacts of fishing. Using baited remote underwater video systems, we surveyed reef fish communities from depths of 2 to 85 m throughout the Raja Ampat archipelago in West Papua, Indonesia – an area considered the heart of the Coral Triangle where coral reef biodiversity is greatest. Here we show—similar to shallow reefs—Raja Ampat MCEs are exceptionally diverse, with 152 fish species recorded at depths greater than 40 m. We found that fish communities were highly depth driven, with declines in fish abundance at increased depth. In contrast to previous studies elsewhere in the world, we found that the proportion of planktivores declined across the shallow reef to MCE depth gradient. While greater human population density correlated with lower reef fish biomass, we did not observe differential impacts based on depth, and so found no evidence that MCEs provide a depth refuge from fishing. Our results expand and contrast some previously established MCE-depth patterns in fish communities, highlighting the need for future MCE studies within the Coral Triangle region.

Introduction

Mesophotic coral ecosystems (MCEs)—reefs from approximately 30-150 m depth—are of increased conservation and research interest because of their unique species assemblages (Rocha et al. 2018; Pinheiro et al. 2019) and potential to act as refuges from shallow reef impacts (Hinderstein et al. 2010; Loya et al. 2016). MCEs are found in tropical and sub-tropical waters globally, and contain a rich array of biodiversity including corals, fishes, sponges, and algae (Kahng et al. 2010; Baker et al. 2016). MCEs have been poorly documented compared to shallower coral reefs because of sampling limitations at greater depths. With advances in survey technology there has been a greater understanding of MCEs, especially in regions such as the Caribbean, Red Sea, Australia, and Hawaii (Turner et al. 2017; Laverick et al. 2018). The Coral Triangle (comprising: Indonesia, Malaysia, Papua New Guinea, the Philippines, Timor Leste, and the Solomon Islands) hosts the greatest shallow reef biodiversity globally (Veron et al. 2009). Yet—despite this diversity—there is comparatively little research effort in the Coral Triangle (Fisher et al. 2011) especially for MCEs (Turner et al. 2017; Laverick et al. 2018). Coral Triangle MCE surveys have been mostly limited to several sites in the Philippines, where highly diverse MCEs have been identified (Cabaitan et al. 2019; Pinheiro et al. 2019).

Based on ecological community composition, MCEs can be divided into upper-MCEs (approximately 30-60 m depth) and lower-MCEs (approximately 60-150 m depth) (Loya et al. 2016). Coral reef fish species richness, abundance, and biomass has broadly been reported to decline across the depth gradient from shallow reefs to MCEs (Kahng et al. 2010; 2014). While reef fish communities are variable between geographical locations, some patterns have begun to emerge. For example, habitat complexity is an important predictor of fish abundance and biomass on MCEs (Boland and Parrish 2005; Brokovich et al. 2008; Bryan et al. 2013). Herbivorous fish—though present in upper-MCEs—often decline in abundance with increased depth and can be near-absent from lower-MCEs (Thresher and Colin 1986; Brokovich et al. 2008; Rocha et al. 2018). While planktivores, in contrast, often increase as a proportion of the fish community on deeper reefs (Thresher and Colin 1986; Brokovich et al. 2008; Rocha et al. 2018).

MCEs may act as fish refuges because shallow reefs maybe more exposed to habitat damage and overfishing, or MCEs may be located at greater distances from shore and so be

subject to fewer human impacts (Bridge et al. 2013). Greater biomass of fisheries target

species has been observed on MCEs compared to adjacent shallow reefs in many locations

(Bejarano et al. 2014; Lindfield et al. 2016; Andradi-Brown et al. 2017). This supports the

idea that some MCEs may be exposed to lower fisheries pressures than adjacent shallow

reefs. Many small-scale fisheries gears are highly depth specific, with differential impacts

across small depth gradients (Ashworth and Ormond 2005; Tyler et al. 2009; Goetze et al.

2011). Small-scale fisheries gears that reach MCEs require additional cost and effort

compared to those used in the shallows (Wood et al. 2006; Lindfield et al. 2014). Yet, in

some regions MCEs may support important fisheries – for example, in Papua New Guinea

mesophotic species comprise large a proposition of some subsistence fish catches

(Longenecker et al. 2017; 2019). Testing potential MCE refuge effects is challenging,

however, as fish abundance, biomass, and community composition transitions with depth

(Thresher and Colin 1986; Brokovich et al. 2008; Rocha et al. 2018; Pinheiro et al. 2019) and

with benthic composition and structural complexity (Boland and Parrish 2005; Gratwicke

and Speight 2005; Bridge et al. 2012). Despite some potential protection by virtue of their

depth, MCEs are still vulnerable to many human impacts including some fisheries,

sedimentation, and global stressors such as coral bleaching and increased tropical storm

frequency and intensity (Andradi-Brown et al. 2016a; Rocha et al. 2018). With few—though

increasing—MCE studies in the Coral Triangle, human impacts on Coral Triangle MCEs

remain poorly understood.

Raja Ampat, located in West Papua, Indonesia, and at the center of the Coral Triangle

contains extensive reefs, as well as mangrove forests and seagrass beds (Mangubhai et al.

2012). Raja Ampat reefs contain the greatest richness of scleractinian corals (Veron et al.

2009) and fishes (Allen and Erdmann 2009) recorded in a single region – with over 550

scleractinian coral species and over 1,400 fish species (Mangubhai et al. 2012). Given Raja

Ampat's unique diversity, there have been several focused efforts to build regional species

lists. For example, Allen and Erdmann (2009) combined detailed field surveys with other

records to produce a fish species list for the region. Scleractinian corals have been recorded

to 160 m depth within Raja Ampat, though reefs beyond the depths of conventional scuba

diving remain poorly characterized (Mangubhai et al. 2012). Unlike many reefs globally, the

shallow reefs of Raja Ampat have not faced widespread mortality from bleaching events and

shallow reef scleractinian coral cover remained stable between 2010-2016 at approximately

30% (Ahmadia et al. 2017).

Local people living within Raja Ampat have high marine resource dependence, with

approximately two-thirds reliant on marine fish for over half their dietary protein needs,

and approximately a quarter reliant on marine capture fisheries as their primary occupation

(Ahmadia et al. 2017). Gleaning, hand-lines, and spear guns are the dominant fishing gears

used by local communities, though commercial trawlers fish in the region (Mangubhai et al.

2012; Ahmadia et al. 2017). Destructive fishing gears such as dynamite and cyanide, while

present, are generally less prevalent in Raja Ampat than elsewhere in Indonesia (Mangubhai

et al. 2012). Since the 1990s, several non-governmental organizations (NGOs) have worked

in Raja Ampat in partnership with local communities and the district and provincial

government to establish a marine protected area (MPA) network. These MPAs are zoned

based on periodic harvest closure areas, no-take areas, and large sustainable fisheries areas

(Grantham et al. 2013), with the intention of allowing local communities to control their

marine resources and reduce the number of commercial vessels from elsewhere in

Indonesia fishing within Raja Ampat (Mangubhai et al. 2012; Ahmadia et al. 2017). In

addition, the entirety of Raja Ampat regency was declared a shark and ray sanctuary in

2012, banning all shark and ray fisheries within the region[1]. Despite fisheries impacts, Raja

Ampat reefs retain high fish biomass including top predators compared to many reefs

elsewhere in the world (Ahmadia et al. 2017). These conservation initiatives—while not

specifically targeting MCEs—provide benefits to many MCEs. For example, the Raja Ampat

MPA network extends protection beyond shallow coral reefs over many MCE and deep-sea

areas. While the shark and ray sanctuary is declared based on the boundary of Raja Ampat

Regency—representing a political boundary rather than an ecological boundary—and so

encompassing all reefs regardless of depth within Raja Ampat.

In this study, we investigate fish communities on MCEs within Raja Ampat and compare

them to adjacent shallow reefs. We specifically identify: (i) whether the high fish species

richness that is known to occur on the shallow reefs of Raja Ampat extends into the

mesophotic zone; (ii) which fish species are indicative of Raja Ampat MCEs; (iii) patterns in

fish species richness, abundance, and trophic groups across the depth gradient; and (iv) we

[1]Peraturan Daerah No. 9 Tahun 2012.

130   consider how human impacts have differentially affected fish biomass on MCEs compared

131   to shallow reefs. Our study represents one of the largest MCE fish community sampling

132   efforts undertaken to date within the Coral Triangle region, and the first within Raja Ampat.

133   Material and methods

134   Study sites

Surveys were conducted in January–February 2014 by UPTD Raja Ampat and

Conservation International Indonesia staff from the KM Imbekwan research vessel. Two

MPAs within the Raja Ampat MPA network were sampled in this study: Suaka Alam Perairan

(SAP; Marine Sanctuary) Kepulauan Waigeo Sebelah Barat and Kawasan Konservasi Perairan

Daerah (KKPD; Regional Marine Protected Area) Selat Dampier (Fig. 1). SAP Kepulauan

Waigeo Sebelah Barat is 271,630 ha in area and was declared in 2009[2], while KKPD Selat

Dampier is 336,200 ha and was declared in 2008[3]. At the time of the surveys, provisional

zonation plans were being developed and implemented for both MPAs and there was

limited MPA enforcement. Sites for SAP Kepulauan Waigeo Sebelah Barat were within the

provisionally designated no-take MPA zones and in areas to the south around Kawe Island

(henceforth grouped with the MPA sites as Waigeo). Sites in KKPD Selat Dampier were

within the provisionally designated no-take and sustainable fisheries zones (henceforth

Dampier) and covering both Batanta (on the south side of the Dampier Strait) and Kri (on

the north side of the Dampier Strait). As these MPAs were new, and zonation

implementation and MPA patrolling and enforcement were still under development, it is

likely that any differences between sites inside the MPA and outside, or between different

MPA zones reflect pre-MPA differences in reef condition. In this study we therefore do not

compare sites inside and outside the MPAs (Waigeo), or between no-take and sustainable

fishing zones (Dampier).

Surveys were conducted using baited remote underwater video (BRUV) during

daylight hours (07:00-16:00). At each site, surveys were conducted within three depth

zones: (i) shallow (2-12 m), (ii) intermediate/mid (15-30 m), and (iii) deep/MCE (40-85 m). A

minimum of two BRUV drops were conducted per depth zone at each site - see Electronic

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Supplementary Material (ESM) Table 1. BRUV replicates within a single site and depth

combination were deployed a minimum of 500 m apart. Sites were located a minimum of

1.5 km apart, with 14 sites surveyed within Waigeo, and 10 sites surveyed in Dampier. In

total across all sites and depths 160 BRUV drops were completed (see ESM Table 2 for GPS

locations). The BRUV consisted of a single GoPro Hero 3 camera, mounted in a frame 0.6 m

above the benthos, and recording forward at the BRUV bait arm and seascape. The BRUV

bait arm was 1.5 m long and was baited with 0.8-1 kg of crushed skipjack tuna. More details

on the survey design and methods are available in Beer (2015). BRUVs were left for 60 min

on the benthos before being recovered.

Video analysis and data processing

Video footage from each BRUV drop was analyzed for all fish species using

EventMeasure (SeaGIS, Melbourne, Australia). The first 50 min of footage was analyzed

from when the BRUV reached the seabed and stabilized, recording the maximum number

(MaxN) for each species. MaxN is defined as the largest number of individuals of a species

visible in a single video frame and therefore is a relative measure of fish abundance. MaxN

is a widely used metric for static video fish analysis, as it avoids problems of counting

individual fish multiple times as they leave and then re-enter the camera field of view

(Watson et al. 2010). MaxN has been shown to provide reliable fish community data when

compared to other conventional reef survey techniques (e.g. Watson et al. 2010; Andradi-

Brown et al. 2016c). Fish identification followed Allen and Erdmann (2012), and all fish were

identified to species level or the greatest taxonomic resolution possible (e.g. genus or

family) if full species identification was not possible.

We categorized each fish species based on trophic group and commercial value. All

fish data were filtered to remove records of manta and devil rays (Mobulidae), and eagle

rays (Myliobatidae), as these are large-bodied transient species that are unlikely to be

accurately recorded in our BRUV sampling. Whitetip, blacktip, and grey reef sharks

(Carcharhinidae), nurse sharks (Ginglymostomatidae), bamboo sharks (Hemiscylliidae),

wedgefishes (Rhinidae), and whiptail rays (Dasyatidae) were retained in our analysis

alongside all other reef fishes. Trophic groups were allocated following the Fishbase 'Food

Items' table, using the Food I-III hierarchical classification of food items eaten by a species,

based on diet composition of >20% of recorded items accessed through rfishbase (Boettiger
et al. 2012; Froese and Pauly 2015). Trophic groups were: (i) Herbivore, (ii) Herbivore and
Planktivore, (iii) Planktivore, (iv) Planktivore and Mobile Invertebrate Feeder, (v) Omnivore,
(vi) Corallivore, (vii) Mobile Invertebrate Feeder, (viii) Piscivore and Mobile Invertebrate
Feeder, and (ix) Piscivore. High commercial value fish species were identified based on the
Fishbase commercial value listing, with species listed at 'high' or 'very high' commercial
value included, and sharks of the families Carcharhinidae, Ginglymostomatidae,
Hemiscylliidae, and Rhinidae also included.

We explored how broad human and environmental impacts affect reef fish biomass
across the depth gradient and region. As our BRUV was single camera, and so only provided
MaxN abundance values for each species, we assigned length estimates to each fish to allow
estimated total biomass per BRUV to be calculated. Fish lengths were assigned in the
following way: Both Waigeo and Dampier have an active shallow reef monitoring program
which records visual fish length estimates along transects at 10 m depth (Ahmadia et al.
2017). We used these locally sourced fish length records from 2014 visual fish surveys and
assigned the median fish species length recorded to each species within each region.
Median length was used as it provides a good estimate for missing fish lengths when
conducting fish biomass calculations (Wilson et al. 2018). For species that were not
recorded in the 2014 shallow reef visual surveys, we instead used the 'Common Length' fish
size estimates for each species from Fishbase (Froese and Pauly 2015). We sourced length-
weight conversion values following the method outlined in Andradi-Brown et al. (2016b)
from Fishbase, enabling us to estimate the biomass of each species at MaxN. The biomass of
each species MaxN per BRUV drop was then summed to give an estimate of relative total
fish biomass per BRUV that could be compared between depths and across the region. We
sourced three additional human/environmental variables from the Marine Socio-
Environmental Covariates database (Yeager et al. 2017) for each BRUV drop. These variables
were: (i) mean annual wave exposure (kW/m), (ii) 2015 human population within 10 km,
and (iii) linear distance to major regional market – in this case Sorong (Fig. 1) on the West
Papuan mainland (km). A 10 km human population buffer was used as studies in the Coral
Triangle have suggested that fishers using reef gears typically fish at distances <10 km, and
fishers generally fish closer to their villages when possible (Soede et al. 2001; Teh et al.
2012).

Fish communities are highly correlated with benthic habitat cover and complexity on

both shallow reefs (Gratwicke and Speight 2005) and MCEs (Brokovich et al. 2008; Bryan et

al. 2013). To account for varying habitat complexity at each BRUV drop, we visually assessed

from the video the Habitat Assessment Score (HAS) following Gratwicke and Speight (2005).

HAS is based on six benthic categories: (i) rugosity of substratum, (ii) variety of benthic

organism growth forms, (iii) benthic habitat architecture height, (iv) refuge size categories

present in the reef, (v) live cover of benthic organisms, and (vi) % hard substratum present

(Gratwicke and Speight 2005). Each benthic category is given a value between 1-5, with 5

representing the greatest habitat complexity/structure/cover. All raw data is available from

DOI: 10.6084/m9.figshare.8097788. [DOI to be activated upon article publication]

Data analysis

We analyzed fish communities based on both abundance and biomass. To visualize

changes in benthic habitat and fish communities across the depth gradient we used a

Principal Component Analysis (PCA) based on the six benthic categories (each scored

between 1-5) and fourth root transformed fish abundance (MaxN). Fourth root

transformation was used to down weight the influence of highly abundant species. PCAs

based on the correlation matrix were fitted using the 'prcomp' function in R (R Core Team

2013). Plots were visualized with 95% confidence ellipses using the ggbiplot package (Vu

2011).

To identify differences in the fish community based on region (Waigeo or Dampier),

site, benthic habitat, and depth we used permutational multivariate analysis of variance

(PERMANOVA) based on a fourth root transformed Bray-Curtis dissimilarity matrix of fish

abundance where each BRUV drop represented a row in the matrix. PERMANOVA was run

for 99999 permutations using the 'adonis' function from vegan (Oksanen et al. 2013). As we

had unbalanced data (i.e. differing number of replicates per site and depth combination),

and were testing multiple factors, the interaction sums of squares model terms are not

independent (Anderson et al. 2008). We therefore used Type 1—sequential—sums of

squares, where each term is fitted after taking account of previous fitted terms. While this

means that the order in which terms are fitted can affect our results, this is considered

appropriate for nested experimental designs where model terms exhibit a natural

hierarchical ordering (Anderson et al. 2008). We fitted PERMANOVAs in the form of Region

+ Site + Benthic PC1 + Depth. Region represents whether the sites were in the Waigeo or

Dampier. Benthic PC1 represents the first principal component axis of a PCA of the benthic

data. This was used to combine the six HAS benthic categories into a single explanatory

variable, as the six categories exhibited high collinearity.

To identify fish species indicative of different depths, we used Dufrêne and Legendre

indicator species analysis (Dufrêne and Legendre 1997) on the fish abundance data across

the three depth zones. Dufrêne and Legendre indicator species analysis identifies the

association between fish species and a particular depth zone. Dufrêne and Legendre

indicator species analysis was conducted and permutational p-values were generated using

the 'indval' function from the labdsv package (Roberts 2013).

To identify how fish community trophic structure changed across the depth gradient

we summed fish abundance per trophic group per BRUV (sum of MaxN for each individual

species within the trophic group) for each depth zone at each site, and then conducted

permutational analysis of variance (permutational-ANOVA) to test for differences with

depth. Depth was coded as 1 (shallow), 2 (intermediate), 3 (MCE) and treated as a

continuous variable to incorporate the gradient of depths. For trophic groups with

significant effects of depth on abundance, we identified potential fish species that could be

driving depth effects by: Firstly, we constructed a PCA of the untransformed fish abundance

data for species belonging to that trophic group. We then tested each species within the

trophic group for a correlation between the species abundance and the first and second

trophic group principal component axes. Species with a Pearson's correlation coefficient

>|0.4| were then tested for depth effects using permutational-ANOVA.

To identify how total and commercially valuable fish biomass across the depth

gradient was affected by human and environmental conditions we used linear mixed-effect

models. These models used fourth root transformed total fish biomass per BRUV as the

response variable and included Site and Region as random factors. Models initially had the

form: fish biomass$^{(1/4)}$ ~ Depth * Benthic PC1 * Wave exposure * Human population *

Distance to market + (1|Site) + (1|Region). Where '*' means the separate fixed effect and

interaction between the variables. Models were fitted using 'lmer' set to optimize the log-

likelihood criterion in the package lme4 (Bates et al. 2015). We first fit this full model, but

then simplified by using likelihood ratio tests to remove non-significant interactions.

Reported p values were generated by likelihood ratio tests removing the

variable/interaction of interest (Bolker et al. 2009). We used partial effects plots to visualize

the effects of human population density and distance to market on fish biomass based on

the remef package (Hohenstein and Kliegl 2019).

Results

Benthic habitat complexity

Habitat complexity declined as depth increased, from a mean total HAS per BRUV of

26 ± 3 (mean ± SE) and 27 ± 3 on shallow reefs in Dampier and Waigeo respectively, to 11 ±

2 and 13 ± 2 on MCEs (Fig. 2A). All six HAS categories (rugosity, variety of growth forms,

height, refuge size category, live cover, and hard substratum) strongly correlate with the

first principal component axis (explaining 80% of benthic variation observed), which shows

clear depth groupings (ESM Fig. 1A).

Fish species richness and abundance

Overall, we recorded 521 fish species across the two regions and all depth zones,

with 328, 307, and 152 fish species recorded on shallow, intermediate, and mesophotic

reefs respectively. Our surveys identified 31 species not previously recorded by the Allen

and Erdmann (2009) fish species list for Raja Ampat (ESM Table 3). Of these, 13 were

potential new records for the region, and 18 had previously been recorded from the

adjacent West Papuan reefs of Cenderawasih Bay, Fakfak, or Kaimana. We found 99 fish

species at greater depths than their maximum depth listed on Fishbase, of which 76 were

extensions onto MCEs in the 40-85 m depth range, and 23 were extensions on shallow or

intermediate depth reefs (<40 m maximum depth; ESM Table 4). Species with depth

extensions included commercially important families such as emperor (Lethrinidae – nine

species with depth extensions) and snapper (Lutjanidae – one species). Some important

fisheries species were also exclusively recorded on MCEs (ESM Table 5) – for example four

emperor species (*Gymnocranius elongatus*, *Lethrinus laticaudis*, *Lethrinus lentjan*, and

Lethrinus rubrioperculatus) and one snapper (*Lutjanus argentimaculatus*). We recorded

International Union for Conservation of Nature (IUCN) red listed species on MCEs in Raja

Ampat, such as the endangered humphead wrasse (*Cheilinus undulatus*) and the vulnerable

green bumphead parrotfish (*Bolbometopon muricatum*). Humphead wrasse were found

across all depths at a similar density (ESM Table 5), but bumphead parrotfish were only

recorded on intermediate depth reefs and MCEs – where they were observed to 41 m

maximum depth (ESM Table 4; ESM Table 5).

Fish species richness declined as depth increased, from a mean number of fish

species per BRUV drop of 49 ± 10 and 57 ± 8 on shallow reefs to 22 ± 6 and 19 ± 2 on MCEs

in Dampier and Waigeo respectively (Fig. 2B). Fish abundance also declined with increasing

depth, however, the magnitude of difference varied between locations (Fig. 2C). For

Dampier, fish abundance only slightly declined between shallow (156 ± 41 ind./BRUV) and

intermediate reefs (144 ± 30 ind./BRUV) but was much lower on MCEs (73 ± 27 ind./BRUV).

For Waigeo, fish abundance declined steadily between shallow reefs and MCEs from 194 ±

26 to 53 ± 7 ind./BRUV.

Differences in fish communities were identified between the two regions, individual

sites, benthic habitat structure, and depth (Table 1). Depth was a major driver of fish

communities (ESM Fig. 1B), and indicator species analysis identified 74 species associated

with the shallows (ESM Table 6), 24 fish species associated with intermediate depths, and

13 species associated with MCEs (Table 2). Of particular note on MCEs were the presence of

three Nemipteridae species (*Pentapodus nagasakiensis*, *Pentapodus paradiseus*, and

Scolopsis vosmeri), which were commonly recorded on MCEs. Purple sand tilefish

(*Hoplolatilus purpureus*) were also indicative of MCEs, which given the lower MCE HAS score

and that this species associates with sand/rubble areas is unsurprising. Overall, 99 fish

species significantly changed in abundance across the depth gradient, with 82 of these fish

species at lower abundance on MCEs than shallow reefs, while 17 fish species were at

greater abundance on MCEs than shallow reefs (ESM Table 5).

Fish trophic structure

Mobile invertebrate feeders and joint piscivore and mobile invertebrate feeders

comprised the largest component of MCE fish communities, together accounting for 65%

(Dampier) and 55% (Waigeo) of the community by abundance (Fig. 3A). The majority (83%)

of indicator species associated with MCEs were also mobile invertebrate feeders (both

exclusively or combined with piscivory) (Table 2). This MCE community contrasts with the

shallows, where herbivorous and planktivorous fishes made up 53% (Dampier) and 57%

(Waigeo) of the community abundance (Fig 3A). Shallow reef indicator species were

comprised by 14% herbivores, 11% corallivores, and only 19% mobile invertebrate feeders

and/or piscivores (ESM Table 6). Removing commercially valuable fish species altered the

reef fish community trophic structure considerably (Fig. 3B) by disproportionately removing

mobile invertebrate feeders and piscivores. This led to a shallow reef community dominated

by herbivores and planktivores, but a more even trophic group distribution on MCEs (Fig

3B).

We tested for differences in overall abundance of each trophic group across the

depth gradient, finding that abundance of herbivores, planktivores, omnivores, and

corallivores declined at increased depth (Table 3). Herbivores had the greatest abundance

decline—with a percentage change of -82% between shallow reefs and MCEs—followed by

planktivores at -79% (Table 3). Piscivores and mobile invertebrate feeders showed the

opposite pattern – with a 50% increase from shallow reefs to MCEs (Table 3). To identify the

species responsible for trophic group abundance change across the depth gradient, we

tested each species within the trophic group for correlations with the first two axes of a PCA

of the trophic group (Table 4). Two herbivores were found to decline in abundance with

depth—the surgeonfish, *Acanthurus maculiceps,* and the damselfish, *Dischistodus*

melanotus—and both of these species were absent from the MCE BRUV drops. For

planktivores, two species—the fusilier, *Caesio lunaris,* and the triggerfish, *Odonus niger*—

correlated with the planktivore PCA and changed abundance across the depth gradient.

Caesio lunaris declined in abundance at increased depths, while *Odonus niger* increased in

abundance from 4.93 ± 1.89 ind./BRUV in the shallows, to 7.29 ± 2.15 ind./BRUV at

intermediate depths, but then declined to 1.33 ± 0.59 ind./BRUV on MCEs. Omnivores

showed mixed patterns, with five species declining in abundance at increased depths, while

three species had greatest abundance at intermediate depths (Table 4). Only one corallivore

correlating with the corallivore PCA changed in abundance—the wrasse, *Diproctacanthus*

xanthurus—that declined with increased depth (Table 4). Two piscivore and mobile

invertebrate feeding species increased in abundance on MCEs, the emperor, *Lethrinus*

microdon, and the threadfin bream, *Pentapodus emeryii* (Table 4).

Human impacts on reef fish biomass

Total fish biomass per BRUV declined as the human population within 10 km

increased (Table 5; Fig. 4A). Though there was high variation, our partial effects plot

indicated approximately 8 kg/BRUV decline in total fish biomass per 100-person population

increase within 10 km distance of the reef site after controlling for other factors (Fig. 4A).

We found the interaction between human population and depth to be non-significant,

suggesting that human population density was not having differential effects on fish

biomass dependent on depth, so we removed the interaction from the model. Total fish

biomass declined with increased distance from the main fish markets in Sorong (Table 5; Fig

4B) – i.e. more remote sites had lower fish biomass than sites closer to the market. Again,

while much variation was present in this relationship, our partial effects plot indicated a

decline of approximately 15 kg/BRUV per 25 km increase in distance away from Sorong (Fig.

4B). We found the interaction between depth and distance to market was non-significant,

suggesting distance to market effects do not vary based on depth, and so removed the

interaction to simplify the model. To identify whether these relationships between human

population and distance to market could be confounded by many non-fisheries target

species, we also ran the model for only commercially valuable fish biomass (Table 5). Using

commercially valuable fish biomass, however, did not detect any effects of human

population or distance to Sorong. As expected from our earlier fish abundance analysis

(Table 1), we found fish biomass declined as depth increased for both total fish biomass and

commercially valuable fish biomass (Table 5). We also identified a significant interaction

between depth and benthic habitat structure for total and commercially valuable fish

biomass—suggesting more complex reefs have greater fish biomass, but the effect of

complexity varies based on depth (Table 5).

Discussion

This study represents one of the largest in situ assessment of coral reef fish

community structure across the shallow-mesophotic depth gradient within the Coral

Triangle region, based on 160 BRUV drops conducted across the 2-85 m depth gradient. Our

results show that fish community structure in Raja Ampat is strongly driven by both depth

and benthic habitat complexity, and that diverse mesophotic fish communities exist within

Raja Ampat. We identified commercially valuable and IUCN red listed fish species on MCEs

and found 99 fish species at depths greater than their Fishbase recorded depth range. We

found evidence of depleted reef fish biomass in areas with higher human population, but

that this effect of human population was not mediated by depth. Overall, we recorded 521

fish species across all depth zones, with 152 fish species recorded on MCEs, placing Raja

Ampat MCEs amongst the most fish species rich MCEs identified globally.

Changes in benthic habitat complexity with depth

Benthic habitat structure declined with depth, in a similar pattern at both Dampier and

Waigeo (Fig. 2A). Unsurprisingly, all six benthic catagories recorded in HAS (rugosity, variety

of growth forms, height, refuge size category, live cover, and hard substratum) were highly

correlated, with many of the deepest BRUV drops associated with low relief reefs or even

recording flat sandy benthos with soft corals. Our results contrast with the only previous

shallow-MCE benthic habitat study within Indonesia, which was conducted within the

Wakatobi Marine National Park (South East Sulawesi), where reef hard substrate cover was

lowest in the shallows and increased down to 80 m depth (Bell et al. 2019). Of the previous

MCE studies in the Coral Triangle, changes in benthic communities and complexity in Raja

Ampat appear most similar to observations from Apo Island in the Philippines, where the

proportion of sand/rubble increased exponentially with depth from 10-80 m depth

(Abesamis et al. 2018). Our HAS results suggested a consistent decline in benthic habitat

structure with increased depth, with intermediate depth reefs having intermediate habitat

structure when compared to shallow reefs and MCEs. Studies in the Philippines have found

more variable benthic community composition across the depth gradient, including that

shallow benthic communities may not be a good predictor of adjacent MCE benthic

communities (Dumalagan et al. 2019). Within the Coral Triangle—and for MCEs more

broadly—MCEs are highly variable and influenced by local conditions, for example river

outflows can increase MCE sedimentation (Cabaitan et al. 2019) or storms can damage reefs

(Abesamis et al. 2018). We used BRUVs to survey benthic communities, unlike most

previous MCE benthic studies from Indonesia (Bell et al. 2019) and the Philippines (Cabaitan

et al. 2019; Dumalagan et al. 2019; Quimpo et al. 2019), which used diver transects or

remote-operated vehicles. BRUVs have to rest on the seabed, preventing our surveys from

incorporating walls or steep slopes. Therefore, the areas we surveyed are likely to have

differing benthic complexity to previous studies in the region, as reef slope is an important

predictor of MCE benthic communities (Lesser et al. 2018).

Fish species richness and indicator species

The number of fish species recorded on Raja Ampat MCEs—152 species—were amongst

the highest recorded to date on MCEs anywhere in the world. For example, in Hawaii 162

fish species were recorded on MCEs by diver surveys spanning 161 sites over 2010-2016

(Fukunaga et al. 2017). In the Philippines a combination of BRUV and technical diving

recorded 277 fish species on MCEs (Pinheiro et al. 2019) – though only 92 MCE fish species

were recorded by BRUV (Abesamis et al. 2018). These results from the Philippines, however,

were based on 29 BRUV drops and so are likely under-sampling fish species richness in

comparison to our 160 BRUV drops in Raja Ampat. In Kimbe Bay, Papua New Guinea, 61

species were recorded at 30 m depth by video transects (MacDonald et al. 2016). While

focused fish collection surveys detect greater fish species richness than BRUVs on MCEs

(Pinheiro et al. 2019), work on Caribbean MCEs suggests that BRUVs detect greater fish

species richness than standardized video transects (Andradi-Brown et al. 2016c). Given the

wide variation in methods used, it is challenging to directly compare fish species richness

between geographical locations. With increased survey effort—including other sampling

methods such as targeted fish collection by technical diving—more fish species are likely to

be recorded on Raja Ampat MCEs. Therefore, our results should be considered an

underestimate of the true fish species richness on Raja Ampat MCEs. Our results, however,

confirm that the exceptional fish species richness present on shallow coral reefs within Raja

Ampat (Allen and Erdmann 2009) also extends into the mesophotic zone.

Our surveys indicate Raja Ampat MCEs have approximately 40% of the species richness

of adjacent shallow reefs. Declines in mean fish species richness at increased depths are

recognized across shallow reef and MCE depth gradients at many sites globally (Kahng et al.

2010; 2014). For example, declines have been recorded in the Caribbean (Bejarano et al.

2014; Pinheiro et al. 2016; Andradi-Brown et al. 2016b), Red Sea and Indian Ocean

(Brokovich et al. 2008; Andradi-Brown et al. 2019), and Pacific (Thresher and Colin 1986;

Lindfield 2015; Pyle et al. 2016; Asher et al. 2017). In the Coral Triangle, previous

comparisons of whole community fish species richness have mostly been limited to smaller

depth gradients. In Kimbe Bay, Papua New Guinea, fish species richness declined 15% from

5 to 30 m depth (MacDonald et al. 2016). In the Philippines, fish species richness was lower

on MCEs (30-35 m) than shallow reefs (8-20 m), though declines with depth are highly

variable – ranging from relatively modest declines to species richness more than halving

(Quimpo et al. 2018; 2019).

We identified 12 fish indicator species of MCEs in Raja Ampat, of which ten were

piscivores and/or mobile invertebrate feeding species (Table 2). Reef fish species

composition across depth gradients are known to be driven by factors such as habitat

availability (MacDonald et al. 2018), water temperature (Simon et al. 2016; Gress et al.

2017), fish morphological traits (Bridge et al. 2016), and broader trophic groups (Bejarano et

al. 2014; Andradi-Brown et al. 2016b). Many of the MCE indicator species we identified have

also been identified as deep species by studies in the Philippines, such as *Hoplolatilus*

purpureus and *Parupeneus heptacanthus* (Abesamis et al. 2018). Species identified as

characteristic of Raja Ampat MCEs included two species of *Pentapodus* – *Pentapodus*

nagasakiensis and *Pentapodus paradiseus*. We also found five *Pentapodus* species

correlated with our piscivore and mobile invertebrate feeder PCA (Table 4), and so

contributed to the trend of greater piscivore and mobile invertebrate feeder abundance on

MCEs. Four *Pentapodus* species increased in abundance on MCEs, though one species

declined (ESM Table 5). Several *Pentapodus species* have elongate trailing caudal fin

filaments (Allen and Erdmann 2012), which has been associated with reducing

hydrodynamic disturbance when swimming (Bridge et al. 2016). This is believed to aid

predator avoidance on deeper reefs where water turbulence is less (Bridge et al. 2016).

Pentapodus species are also known to favor sandy areas immediately adjacent to reefs

(Gomelyuk 2009), which are similar to MCE habitats we recorded in Raja Ampat. As several

of our analyses have identified MCE *Pentapodus*, this genus could be used as an easy to

identify indicator group for MCE fish communities in Raja Ampat.

Whilst there was broad agreement between our indicator species and other studies

from the Coral Triangle, several Raja Ampat indicator species were identified at different

depths to previous studies. For example, *Sufflamen fraenatum* was found to be a depth

generalist between 10-80 m in the Philippines (Abesamis et al. 2018) and *Pentapodus*

paradiseus was also found to be common at 30-35 m depth in the Philippines (Quimpo et al.

2018), but our indicator species analysis clustered both as MCE species in Raja Ampat. In

even greater contrast, work in the Philippines identified two of our MCE indicator species,

Lethrinus lentjan and *Pomacentrus nagasakiensis*, as shallow water species (Abesamis et al.

2018). These differences could reflect real differences between the Philippines survey sites

and Raja Ampat or they could be the result of methodological differences between surveys

that affect detection of species. Surveys using standardized methods across the region are

required to directly identify differences in the fish community, but our results imply that

depth ranges should not be generalized for species across the Coral Triangle.

Our intermediate depth fish indicator species more broadly aligns with previous

studies. For example, *Forcipiger flavissimus* and *Pomacentrus nigromanus*, which we

identified as intermediate depth species, are both common at intermediate depths in Papua

New Guinea (MacDonald et al. 2016). Studies from Papua New Guinea have highlighted the

absence of Acanthuridae from MCEs (Pyle 2000; Longenecker et al. 2019), but their

presence at intermediate depths and the transition zone with upper-MCEs (MacDonald et

al. 2016; Longenecker et al. 2019). Comparing with our results, we find that *Acanthurus*

thompsoni was identified as an intermediate depth indicator species in Raja Ampat. This

species is common in coral reef communities at intermediate depths in both Papua New

Guinea (MacDonald et al. 2016) and the Philippines (Quimpo et al. 2018).

Fish abundance and community structure across the depth gradient

Raja Ampat fish abundance declined with increased depth in both Dampier and

Waigeo (Fig. 2). Globally, differences in fish abundance across shallow reef to MCE depth

gradients are highly variable. In many locations abundance declines at increased depths, for

example in Honduras (Andradi-Brown et al. 2016b), Curaçao (Pinheiro et al. 2016), Red Sea

(Brokovich et al. 2008; 2010), the Chagos Archipelago (Andradi-Brown et al. 2019), and

Hawaii (Asher et al. 2017). Whereas, abundance and biomass increases with depth in some

locations, for example Bermuda (Pinheiro et al. 2016) and the Mariana Islands (Lindfield et

al. 2016), before reducing on lower MCEs (>70 m depth). In the Coral Triangle, fish

abundance reflects this variation, with declines at increased depth in Papua New Guinea

and the Philippines (MacDonald et al. 2016; Quimpo et al. 2018; 2019) – but some locations

where fish abundance increases from the surface to a peak at 30 m depth before declining

(Abesamis et al. 2018; Quimpo et al. 2018). As we only recorded three broad depth groups,

it is not possible to identify fine scale changes in fish abundance in Raja Ampat. However,

differences in abundance with depth between Waigeo and Dampier could be driven by

benthic complexity. Previous BRUV studies in Hawaii have found that benthic structure

complexity alters fish patterns across the depth gradient, with unconsolidated substrates

having similar overall densities of fish across the depth gradient, while hard bottom habitats

show declines (Asher et al. 2017). Given that we detected a significant interaction between

depth and habitat complexity when predicting fish biomass (Table 5), it is likely that changes

in abundance are also affected by differences in benthic habitats.

We found that MCE fish communities shifted towards increased mobile invertebrate

feeders and piscivores at deeper depths, and away from omnivores, corallivores, and

planktivores which were more common in the shallows (Fig. 3A). Increases in piscivore

abundance on MCEs compared to shallow reefs is consistent with other studies, as are

declines in omnivores and corallivores (Abesamis et al. 2018; Quimpo et al. 2019). However,

reductions planktivores as a proportion of the fish community at increased depths is

unusual. Previously, studies—including some based on BRUV surveys—from the western

Atlantic (Bejarano et al. 2014; Pinheiro et al. 2016; Andradi-Brown et al. 2016b), the

Marshall Islands (Thresher and Colin 1986), the Red Sea (Brokovich et al. 2008; 2010), and

the Philippines (Abesamis et al. 2018; Quimpo et al. 2018; 2019) have characterized MCEs

fish communities as planktivore dominated. The decline in planktivores as a proportion of

the fish community at increased depths in Raja Ampat could in part be explained by

commercially valuable species. Many locations where planktivores are dominant on MCEs

have experienced high fishing pressure – removing commercially valuable fish species. Raja

Ampat has been exposed to limited fishing pressure in the past and so retains a high

abundance of commercially valuable fish species. Filtering our data to remove commercially

valuable fish species (Fig. 3B) shifted the MCE fish community towards a greater proportion

of planktivores.

Human impacts on MCE reef fishes

Increased human population density in areas where local communities have high

marine resource dependence is well-known to be associated with lower reef fish biomass

(e.g. Bellwood et al. 2012; Williams et al. 2015). Our results show that locations closer to

higher population densities also have lower fish biomass (Table 5; Fig. 4A). MCEs are

particularly likely to act as refuges from fisheries when spear-fishing is a dominant fishing

method (Lindfield et al. 2014; Andradi-Brown et al. 2017). Community surveys in Dampier

region show that people typically use simple hand gears for subsistence fishing, including

spear-fishing and single hand lines, but it was not reported which was the most dominant

form (Claborn et al. 2017). If MCEs were acting as fisheries refuges, we would have expected

to identify a significant interaction between human population density and depth, as spear-

fishers would be unable to reach the full depth range of MCEs. The lack of interaction in our

results suggests that hand lines are likely to be more dominant in the fishery. Caution is

required, however, in interpreting our fish biomass data across depth gradients as we were

unable to record individual fish lengths. We allocated average fish lengths to individual fish

species, using region-specific length data from shallow reef surveys in Waigeo and Dampier

(Ahmadia et al. 2017) supplemented by Fishbase (Froese and Pauly 2015) for species

without regional data available. Studies have shown fish length distributions change across

shallow reef to MCE depth gradients, with larger individuals often present on deeper reefs

(Andradi-Brown et al. 2016b; Gress et al. 2018). Our total fish biomass estimates for each

BRUV does not account for biomass differences caused by changes in fish body size with

depth for each individual species. Therefore, these values should be considered estimates.

Further research is needed to identify whether changes in body size with depth occurs for

species in Raja Ampat, and the effect this has on biomass patterns with depth.

Remote reefs often have greater fish biomass (Graham and McClanahan 2013;

Pinheiro et al. 2016), largely attributed to increased accessibility cost and effort required for

fishers (Maire et al. 2016).We found reefs further away from Sorong— the major urban

center and fish market within the Raja Ampat region (Ahmadia et al. 2017)—had lower fish

biomass. At a global scale, fish biomass is often unchanged within 0-14 km of markets, but

then increases exponentially away from the market (Cinner et al. 2013), though some

studies have found no effect (e.g. Campbell et al. 2018). While linear distance strongly

correlates with market access at a global scale, it can be locally highly variable (Maire et al.

2016). For example, travel times for reefs 105-115 km from the nearest market can vary

from 2-13 hours (Maire et al. 2016). Reefs in this study varied from 70-183 km from Sorong,

with Dampier sites 70-112 km and Waigeo sites 148-183 km. However, while this static view

of market centers may be useful at the global scale, it misses local nuance. In the period up

to 2014, Waigeo reefs were visited by large freezer-equipped vessels that purchased fish

from villages, effectively creating mobile market centers. In addition, prior to the 2012 shark

fishing ban many shark fishers from Indonesian provinces such as Maluku travelled

eastwards to fish within Raja Ampat regency (Jaiteh et al. 2016). Therefore, reefs further

from Sorong—such as Waigeo—may actually have been more accessible to this historic

fishing effort.

Dampier MPA is subject to high and increasing reef tourism (Spalding et al. 2017;

Papilaya et al. 2019). During the period 2012-2016 many residents in Dampier shifted

primary occupation from wild capture fisheries to wage labor (believed to be tourism

related), which was accompanied by a decline in the number of households fishing regularly

(Claborn et al. 2017). Despite this decline in fisheries, Dampier residents continue to fish at

the highest rate within Raja Ampat (Ahmadia et al. 2017), and demand for fish for tourist

consumption is likely to be growing. Tourism brings a range of sustainability and

environmental management challenges for the region – including managing coastal

development and pollution. Elsewhere in the world, MCEs are known to be damaged by

fisheries overexploitation, sedimentation, and pollution (Andradi-Brown et al. 2016a; Rocha

et al. 2018). It is therefore important for tourism development in the region to be

sustainable managed to ensure that benefits are received by local communities and it does

not lead to environmental degradation (Atmodjo et al. 2017; Papilaya et al. 2019).

Our results provide the first overview of fish communities on MCEs in Raja Ampat

and represent one of the most extensive MCE fish surveys conducted in the Coral Triangle

to date. These results suggest that MCEs in Raja Ampat are likely amongst the most fish

species rich globally and retain substantial abundance of higher fish trophic levels. Our

findings help increase global understanding of MCE fish ecology and provide the first insight

of how human interactions can shape MCEs in the center of coral reef biodiversity.

Conflict of interest

623   On behalf of all authors, the corresponding author states that there is no conflict of interest.

Acknowledgments

We wish to thank: Balai Kawasan Konservasi Perairan Nasional (BKKPN) Kupang of the Ministry of Marine Affairs and Fisheries (MMAF) through the Satker KKPN Raja Ampat; the Walton Family Foundation who provide funding for the field surveys; Captain Wempy Boari and Captain Andreas of MV Imbekwan and their crew; Conservation International Indonesia staff who assisted with surveys (including: Abraham Sianipar, Ronald Mambrasar, Abdy Hasan); UPTD Raja Ampat staff who assisted with surveys (including: Aser Burdam, Risyart Mirino, Elvis Mambraku, Yance Mayor, Naftali Manggara, Imanuel Mofu); students from the International Master of Science in Marine Biodiversity and Conservation (EMBC+) and the International Master in Marine Biological Resources (IMBRSea) and the University of Oxford who assisted with BRUV video analysis. We also thank the following who assisted with this study: Edy Setyawan, Florencia Cerutti, Matthew Fox, Meity Mongdong, Timore Kristiani, Alberth Nebore, Permenas Mambrasar, Alex David Rogers, and Mark Erdmann. DAAB acknowledges a Fisheries Society of the British Isles (FSBI) PhD studentship. AJEB acknowledges a NSERC IPS grant. CEIH is supported by the Bertarelli Foundation as part of the Bertarelli Programme in Marine Science. We thank the reviewers and the editor for comments that substantially improved this manuscript.

References

Abesamis RA, Langlois T, Birt M, Thillainath E, Bucol AA, Arceo HO, Russ GR (2018) Benthic habitat and fish assemblage structure from shallow to mesophotic depths in a storm-impacted marine protected area. Coral Reefs 37:81–97

Ahmadia GN, Awaludinnoer A, Glew L, Pakiding F, Harris J, Hidayat N, Ihsan E, Mascia MB, Matualage D, Mohebalian P, Prada D, Purwanto P (2017) 2016 State of the Bird's Head Seascape MPA Network Report. 1–35

Allen GR, Erdmann MV (2009) Reef fishes of the Bird's Head Peninsula, West Papua, Indonesia. Check List 5:587–628

Allen GR, Erdmann MV (2012) Reef Fishes of the East Indies. Conservation International Foundation.

Anderson MJ, Gorley RN, Clarke KR (2008) PERMANOVA+ for PRIMER: Guide to Software and Statistical Methods. PRIMER-E, Plymouth, U.K.

Andradi-Brown D, Laverick J, Bejarano I, Bridge T, Colin P, Eyal G, Jones R, Kahng SE, Reed J,
Smith TB, Spalding HL, Weil E, Wood E (2016a) Threats to mesophotic coral ecosystems
and management options. In: Baker E.K., Puglise K.A., Harris P.T. (eds) Mesophotic Coral
Ecosystems - A lifeboat for coral reefs? The United Nations Environment Programme
and GRID-Arendal, Nairobi and Arendal, pp 67–82

Andradi-Brown DA, Dinesen Z, Head CEI, Tickler DM, Rowlands G, Rogers AD (2019) The
Chagos Archipelago. In: Loya Y., Puglise K.A., Bridge T.C.L. (eds) Mesophotic Coral
Ecosystems. Springer, pp 215–229

Andradi-Brown DA, Gress E, Wright G, Exton DA, Rogers AD (2016b) Reef Fish Community
Biomass and Trophic Structure Changes across Shallow to Upper-Mesophotic Reefs in
the Mesoamerican Barrier Reef, Caribbean. PLoS ONE 11:e0156641

Andradi-Brown DA, Grey R, Hendrix A, Hitchner D, Hunt CL, Gress E, Madej K, Parry RL,
Régnier-McKellar C, Jones OP, Arteaga M, Izaguirre AP, Rogers AD, Exton DA (2017)
Depth-dependent effects of culling—do mesophotic lionfish populations undermine
current management? Royal Society Open Science 4:170027

Andradi-Brown DA, Macaya-Solis C, Exton DA, Gress E, Wright G, Rogers AD (2016c)
Assessing Caribbean Shallow and Mesophotic Reef Fish Communities Using Baited-
Remote Underwater Video (BRUV) and Diver-Operated Video (DOV) Survey Techniques.
PLoS ONE 11:e0168235

Asher J, Williams ID, Harvey ES (2017) Mesophotic Depth Gradients Impact Reef Fish
Assemblage Composition and Functional Group Partitioning in the Main Hawaiian
Islands. Front. Mar. Sci. 4:98

Ashworth JS, Ormond RFG (2005) Effects of fishing pressure and trophic group on
abundance and spillover across boundaries of a no-take zone. Biol Conserv 121:333–344

Atmodjo E, Lamers M, Mol A (2017) Financing marine conservation tourism: Governing
entrance fees in Raja Ampat, Indonesia. Marine Policy 78:181–188

Baker EK, Puglise KA, Harris PT (2016) Mesophotic Coral Ecosystems - A lifeboat for coral
reefs? The United Nations Environment Programme and GRID-Arendal, Nairobi and
Arendal

Bates D, Mächler M, Bolker B, Walker S (2015) Fitting Linear Mixed-Effects Models Using
lme4. Journal of Statistical Software 67

Beer AJE (2015) Diversity and abundance of sharks in no-take and fished sites in the marine
protected area network of Raja Ampat, West Papua, Indonesia, using baited remote
underwater video (BRUVs). Royal Roads University. Victoria, Canada.

Bejarano I, Appeldoorn RS, Nemeth M (2014) Fishes associated with mesophotic coral
ecosystems in La Parguera, Puerto Rico. Coral Reefs 33:313–328

Bell JJ, Jompa J, Haris A, Werorilangi S, Shaffer M, Mortimer C (2019) Domination of
mesophotic ecosystems in the Wakatobi Marine National Park (Indonesia) by sponges,
soft corals and other non-hard coral species. Journal of the Marine Biological
Association of the United Kingdom 99:771-775

Bellwood DR, Hoey AS, Hughes TP (2012) Human activity selectively impacts the ecosystem
roles of parrotfishes on coral reefs. Proceedings of the Royal Society of London B:
Biological Sciences 279:1621–1629

Boettiger C, Lang DT, Wainwright PC (2012) rfishbase: exploring, manipulating and
visualizing FishBase data from R. Journal of Fish Biology 81:2030–2039

Boland RC, Parrish FA (2005) Description of Fish Assemblages in the Black Coral Beds off
Lahaina, Maui, Hawai'i. Pacific Science 59:411–420

Bolker BM, Brooks ME, Clark CJ, Geange SW, Poulsen JR, Stevens MHH, White J-SS (2009)
Generalized linear mixed models: a practical guide for ecology and evolution. Trends in
Ecology & Evolution 24:127–135

Bridge T, Scott A, Steinberg D (2012) Abundance and diversity of anemonefishes and their
host sea anemones at two mesophotic sites on the Great Barrier Reef, Australia. Coral
Reefs 31:1057–1062

Bridge TCL, Hughes TP, Guinotte JM, Bongaerts P (2013) Call to protect all coral reefs.
Nature Climate Change 3:528–530

Bridge TCL, Luiz OJ, Coleman RR, Kane CN, Kosaki RK (2016) Ecological and morphological
traits predict depth-generalist fishes on coral reefs. Proceedings of the Royal Society of
London B: Biological Sciences 283:20152332

Brokovich E, Ayalon I, Einbinder S, Segev N, Shaked Y, Genin A, Kark S, Kiflawi M (2010)
Grazing pressure on coral reefs decreases across a wide depth gradient in the Gulf of
Aqaba, Red Sea. Marine Ecology Progress Series 399:69–80

Brokovich E, Einbinder S, Shashar N, Kiflawi M, Kark S (2008) Descending to the twilight-
zone: changes in coral reef fish assemblages along a depth gradient down to 65 m.
Marine Ecology Progress Series 371:253–262

Bryan DR, Kilfoyle K, Gilmore RG Jr, Spieler RE (2013) Characterization of the mesophotic
reef fish community in south Florida, USA. Journal of Applied Ichthyology 29:108–117

Cabaitan PC, Quimpo TJR, Dumalagan EE, Munar J, Calleja MAC, Olavides RDD, Go K, Albelda
R, Cabactulan D, Tinacba EJC, Doctor MAA, Villanoy CL, Siringan FP (2019) The
Philippines. Mesophotic Coral Ecosystems. Springer, Cham, pp 265–284

Campbell SJ, Edgar GJ, Stuart-Smith RD, Soler G, Bates AE (2018) Fishing-gear restrictions
and biomass gains for coral reef fishes in marine protected areas. Conservation Biology
32:401–410

Cinner JE, Graham NAJ, Huchery C, MacNeil MA (2013) Global Effects of Local Human
Population Density and Distance to Markets on the Condition of Coral Reef Fisheries.
Conservation Biology 27:453–458

Claborn K, Glew L, Matualage D, Mohebalian P, Pakiding F, Ahmadia GN, Mascia MB (2017)
Status and trends in human well-being in Selat Dampier MPA. World Wildlife Fund,
University of Papua, and Conservation International. Washington, D.C., United States,
Manokwari, Indonesia, and Arlington, V.A., United States.

Dufrêne M, Legendre P (1997) Species assemblages and indicator species: the need for a
flexible asymmetrical approach. Ecological Monographs 67:345–366

Dumalagan EE Jr, Cabaitan PC, Bridge TCL, Go KT, Quimpo TJR, Olavides RDD, Munar JC,
Villanoy CL, Siringan FP (2019) Spatial variability in benthic assemblage composition in
shallow and upper mesophotic coral ecosystems in the Philippines. Marine
Environmental Research 150:104772

Fisher R, Ben T Radford, Knowlton N, Brainard RE, Michaelis FB, Caley MJ (2011) Global
mismatch between research effort and conservation needs of tropical coral reefs.
Conservation Letters 4:64–72

Froese R, Pauly D (2015) FishBase. www.fishbase.org

Fukunaga A, Kosaki RK, Wagner D (2017) Changes in mesophotic reef fish assemblages along
depth and geographical gradients in the Northwestern Hawaiian Islands. Coral Reefs
36:785–790

Goetze JS, Langlois TJ, Egli DP, Harvey ES (2011) Evidence of artisanal fishing impacts and
depth refuge in assemblages of Fijian reef fish. Coral Reefs 30:507–517

Gomelyuk VE (2009) Fish assemblages composition and structure in three shallow habitats
in north Australian tropical bay, Garig Gunak Barlu National Park, Northern Territory,
Australia. Journal of the Marine Biological Association of the United Kingdom 89:449–
460

Graham NAJ, McClanahan TR (2013) The Last Call for Marine Wilderness? BioScience
63:397–402

Grantham HS, Agostini VN, Wilson J, Mangubhai S, Hidayat N, Muljadi A, Muhajir, Rotinsulu
C, Mongdong M, Beck MW, Possingham HP (2013) A comparison of zoning analyses to
inform the planning of a marine protected area network in Raja Ampat, Indonesia.
Marine Policy 38:184–194

Gratwicke B, Speight MR (2005) The relationship between fish species richness, abundance
and habitat complexity in a range of shallow tropical marine habitats. Journal of Fish
Biology 66:650–667

Gress E, Andradi-Brown DA, Woodall L, Schofield PJ, Stanley K, Rogers AD (2017) Lionfish
(*Pterois* spp.) invade the upper-bathyal zone in the western Atlantic. PeerJ 5:e3683

Gress E, Arroyo-Gerez MJ, Wright G, Andradi-Brown DA (2018) Assessing mesophotic coral
ecosystems inside and outside a Caribbean marine protected area. Royal Society Open
Science 5:180835

Hinderstein LM, Marr JCA, Martinez FA, Dowgiallo MJ, Puglise KA, Pyle RL, Zawada DG,
Appeldoorn R (2010) Theme section on "Mesophotic Coral Ecosystems:
Characterization, Ecology, and Management." Coral Reefs 29:247–251

Hohenstein S, Kliegl R (2019) remef: Remove Partial Effects. R package version 1.0.6.9000
https://github.com/hohenstein/remef/:

Jaiteh VF, Lindfield SJ, Mangubhai S, Warren C, Fitzpatrick B, Loneragan NR (2016) Higher
Abundance of Marine Predators and Changes in Fishers' Behavior Following Spatial
Protection within the World's Biggest Shark Fishery. Front. Mar. Sci. 3:511

Kahng SE, Copus JM, Wagner D (2014) Recent advances in the ecology of mesophotic coral
ecosystems (MCEs). Current Opinion in Environmental Sustainability 7:72–81

Kahng SE, Garcia-Sais JR, Spalding HL, Brokovich E, Wagner D, Weil E, Hinderstein L, Toonen
RJ (2010) Community ecology of mesophotic coral reef ecosystems. Coral Reefs 29:255–
275

Laverick JH, Piango S, Andradi-Brown DA, Exton DA, Bongaerts P, Bridge TCL, Lesser MP, Pyle
RL, Slattery M, Wagner D, Rogers AD (2018) To what extent do mesophotic coral
ecosystems and shallow reefs share species of conservation interest? A systematic
review. Environmental Evidence 7:15

Lesser MP, Slattery M, Mobley CD (2018) Biodiversity and Functional Ecology of Mesophotic
Coral Reefs. Annu. Rev. Ecol. Evol. Syst. 49:49–71

Lindfield S (2015) Depth refuge and the impacts of fishing on coral reef fish communities.
University of Western Australia

Lindfield SJ, Harvey ES, Halford AR, McIlwain JL (2016) Mesophotic depths as refuge areas
for fishery-targeted species on coral reefs. Coral Reefs 35:125–137

Lindfield SJ, McIlwain JL, Harvey ES (2014) Depth Refuge and the Impacts of SCUBA
Spearfishing on Coral Reef Fishes. PLoS ONE 9:e92628

Longenecker K, Langston R, Bolick H, Crane M, Donaldson T, Franklin E, Kelokelo M, Kondio
U, Potuku T (2017) Rapid reproductive analysis and length–weight relations for five
species of coral-reef fishes (Actinopterygii) from Papua New Guinea: Nemipterus
isacanthus, Parupeneus barberinus, Kyphosus cinerascens, Ctenochaetus striatus
(Perciformes), and Balistapus undulatus (Tetraodontiformes). Acta Ichthyologica et
Piscatoria 47:107–124

Longenecker K, Roberts TE, Colin PL (2019) Papua New Guinea. Mesophotic Coral
Ecosystems. Springer, Cham, pp 321–336

Loya Y, Eyal G, Treibitz T, Lesser MP, Appeldoorn R (2016) Theme section on mesophotic
coral ecosystems: advances in knowledge and future perspectives. Coral Reefs 35:1–9

MacDonald C, Bridge TCL, Jones GP (2016) Depth, bay position and habitat structure as
determinants of coral reef fish distributions: Are deep reefs a potential refuge? Marine
Ecology Progress Series 561:217–231

MacDonald C, Tauati MI, Jones GP (2018) Depth patterns in microhabitat versatility and
selectivity in coral reef damselfishes. Marine Biology 165:138

Maire E, Cinner J, Velez L, Huchery C, Mora C, D'Agata S, Vigliola L, Wantiez L, Kulbicki M,
Mouillot D (2016) How accessible are coral reefs to people? A global assessment based
on travel time. Ecol Letters 19:351–360

Mangubhai S, Erdmann MV, Wilson JR, Huffard CL, Ballamu F, Hidayat NI, Hitipeuw C,
Lazuardi ME, Muhajir, Pada D, Purba G, Rotinsulu C, Rumetna L, Sumolang K, Wen W
(2012) Papuan Bird's Head Seascape: Emerging threats and challenges in the global
center of marine biodiversity. Marine Pollution Bulletin 64:2279–2295

Oksanen J, Kindt R, Legendre P, OHara B, Simpson GL, Solymos P, Stevens MHH, Wagner H
(2013) vegan: Community Ecology Package. www.cran.r-project.org

Papilaya RL, Boli P, Nikijuluw VPH (2019) Carrying Capacity of Diving Tourism in Dampier
Strait Marine Conservation Area – District of Raja Ampat. IOP Conf. Ser.: Earth Environ.
Sci. 246:012060

Pinheiro HT, Goodbody-Gringley G, Jessup ME, Shepherd B, Chequer AD, Rocha LA (2016)
Upper and lower mesophotic coral reef fish communities evaluated by underwater
visual censuses in two Caribbean locations. Coral Reefs 35:139–151

Pinheiro HT, Shepherd B, Castillo C, Abesamis RA, Copus JM, Pyle RL, Greene BD, Coleman
RR, Whitton RK, Thillainath E, Bucol AA, Birt M, Catania D, Bell MV, Rocha LA (2019)
Deep reef fishes in the world's epicenter of marine biodiversity. Coral Reefs 38:985–995

Pyle RL (2000) Assessing Undiscovered Fish Biodiversity on Deep Coral Reefs Using
Advanced Self-Contained Diving Technology. Marine Technology Society Journal 34:82–
91

Pyle RL, Boland R, Bolick H, Bowen BW, Bradley CJ, Kane C, Kosaki RK, Langston R,
Longenecker K, Montgomery A, Parrish FA, Popp BN, Rooney J, Smith CM, Wagner D,
Spalding HL (2016) A comprehensive investigation of mesophotic coral ecosystems in
the Hawaiian Archipelago. PeerJ 4:e2475

Quimpo TJR, Cabaitan PC, Go KTB, Dumalagan EE, Villanoy CL, Siringan FP (2019) Similarity in
benthic habitat and fish assemblages in the upper mesophotic and shallow water reefs
in the West Philippine Sea. Journal of the Marine Biological Association of the United
Kingdom 99:1507–1517

Quimpo TJR, Cabaitan PC, Olavides RDD, Dumalagan EE Jr, Munar J, Siringan FP (2018)
837   Spatial variability in reef-fish assemblages in shallow and upper mesophotic coral
838   ecosystems in the Philippines. Journal of Fish Biology 37:81

839   R Core Team (2013) R: A language and environment for statistical computing. www.R-
840   project.org

841   Roberts DW (2013) labdsv: Ordination and Multivariate Analysis for Ecology. R package
842   version 1.0.6.9000 http://CRAN.R-project.org/package=labdsv:

843   Rocha LA, Pinheiro HT, Shepherd B, Papastamatiou YP, Luiz OJ, Pyle RL, Bongaerts P (2018)
844   Mesophotic coral ecosystems are threatened and ecologically distinct from shallow
845   water reefs. Science 361:281–284

846   Simon T, Pinheiro HT, Moura RL, Filho AC, Rocha LA, Martins AS, Mazzei E, Filho RBF, Filho
847   GMA, Joyeux JC (2016) Mesophotic fishes of the Abrolhos Shelf, the largest reef
848   ecosystem in the South Atlantic. Journal of Fish Biology 89:990–1001

849   Soede CP, Van Densen WLT, Hiddink JG, Kuyl S, Machiels MAM (2001) Can fishermen
850   allocate their fishing effort in space and time on the basis of their catch rates? An
851   example from Spermonde Archipelago, SW Sulawesi, Indonesia. Fisheries Management
852   and Ecology 8:15–36

853   Spalding M, Burke L, Wood SA, Ashpole J, Hutchison J, zu Ermgassen P (2017) Mapping the
854   global value and distribution of coral reef tourism. Marine Policy 82:104–113

855   Teh LCL, Teh LSL, Meitner MJ (2012) Preferred Resource Spaces and Fisher Flexibility:
856   Implications for Spatial Management of Small-Scale Fisheries. Hum Ecol 40:213–226

857   Thresher RE, Colin PL (1986) Trophic structure, diversity and abundance of fishes of the
858   deep reef (30-300m) at Enewetak, Marshall Islands. Bulletin of Marine Science 38:253–
859   272

860   Turner JA, Babcock RC, Hovey R, Kendrick GA (2017) Deep thinking: a systematic review of
861   mesophotic coral ecosystems. ICES Journal of Marine Science

862   Tyler E, Speight M, Henderson P, Manica A (2009) Evidence for a depth refuge effect in
863   artisanal coral reef fisheries. Biol Conserv 142:652–667

864   Veron JEN, Devantier LM, Turak E, Green AL, Kininmonth S, Stafford-Smith M, Peterson N
865   (2009) Delineating the Coral Triangle. Galaxea, Journal of Coral Reef Studies 11:91–100

866   Vu VQ (2011) ggbiplot: A ggplot2 based biplot. R package version 0.55

867   Watson DL, Harvey ES, Fitzpatrick BM, Langlois TJ, Shedrawi G (2010) Assessing reef fish
868   assemblage structure: how do different stereo-video techniques compare? Marine
869   Biology 157:1237–1250

Williams ID, Baum JK, Heenan A, Hanson KM, Nadon MO, Brainard RE (2015) Human,
Oceanographic and Habitat Drivers of Central and Western Pacific Coral Reef Fish
Assemblages. PLoS ONE 10:e0120516

Wilson SK, Graham NAJ, Holmes TH, MacNeil MA, Ryan NM (2018) Visual versus video
methods for estimating reef fish biomass. Ecological Indicators 85:146–152

Wood E, Suliansa MS, Mustapa I (2006) Fishing practices, status of coral reef resources and
tactics for reversing unsustainable use on the Semporna Island reefs (Sabah, Malaysia).
Proceedings of 10th International Coral Reef Symposium 1211–1217

Yeager LA, Marchand P, Gill DA, Baum JK, McPherson JM (2017) Marine Socio-
Environmental Covariates: queryable global layers of environmental and anthropogenic
variables for marine ecosystem studies. Ecology 98:1976–1976

Figures

884
885 Figure 1. Study sites and marine protected areas within Raja Ampat.
886

887
Figure 2. Overall benthic and fish community differences with depth. (A) Habitat assessment
score (HAS), (B) fish species richness, and (C) fish abundance. (A) shows the mean total HAS
per BRUV drop, with values per BRUV potentially ranging between 6-30 based on scoring a
value of 1-5 for each of: rugosity, variety of growth forms, height, refuge size category, live
cover, and hard substratum. Abundance represents the mean number of individual fish per
BRUV (ind./BRUV), based on summing the MaxN for each species per BRUV. Error bars show
1 standard error above and below the mean. Asterix (*) signified a significant (p <0.05)
difference with depth.
896

897
898 Figure 3. Community composition of each trophic group by depth. (A) abundance of all fish,
899 (B) abundance of all fish excluding high/very high value fish and sharks.
900

901
Figure 4. Partial effects plots for (A) human population and (B) distance to market based on
total fish biomass per BRUV. Partial effect plots adjust fish biomass for all model fixed
effects (Depth, Benthic PC1, Wave exposure, the Depth:Benthic PC1 interaction, and either
human population or distance to market) and random effects (Area, Site) with the exception
of the plotted fixed effect (human population or distance to market). Adjusted biomass,
therefore, allows the relationship between the fixed effect of interest and fish biomass to be
examined after controlling for all other fixed and random parts of the model. See Table 5 for
full model.
910

911 ## Tables

912 Table 1. PERMANOVA testing effects of region, site, benthic principal component 1 (PC1),
913 and depth on fish community structure. Fish community structure is calculated as fourth-
914 root transformed abundance (MaxN) per BRUV drop using Bray-Curtis dissimilarities.
915 Degrees of freedom (DF), sum of squares (SS), and mean square (MS) are shown.

	DF	SS	MS	Pseudo-F	p(perm)
Region	1	0.92	0.92	2.52	<0.001
Site	22	10.51	0.48	1.31	<0.001
Benthic PC1	1	4.16	4.16	11.40	<0.001
Depth	1	1.38	1.38	3.78	<0.001
Residuals	134	48.92	0.37		
Total	159	65.89			

Table 2. Dufrêne and Legendre indicator species analysis showing fish species indicative of
intermediate and MCE depth zones. Analysis is based on fish abundance with results
showing the Dufrêne and Legendre indicator value (measuring the association between the
fish species and the depth group) and the associated permutational p-value (based on
99999 permutations). Only fish species with a p<0.05 association for the depth band are
shown. See ESM Table 6 for an expanded version of this table including fish species
indicative of the shallow depth zone.

Depth/Family	Species	Indicator Value	p(perm)	Trophic Group
Intermediate				
Acanthuridae	*Acanthurus thompsoni*	0.22	0.008	Planktivore
	Naso thynnoides	0.32	<0.001	Omnivore
Balistidae	*Odonus niger*	0.33	0.032	Planktivore
	Sufflamen bursa	0.39	0.001	Mobile Invert.
Caesionidae	*Pterocaesio digramma*	0.17	0.028	Planktivore
Carcharhinidae	*Triaenodon obesus*	0.18	0.042	Piscivore & Mobile Invert.
Chaetodontidae	*Coradion chrysozonus*	0.22	0.033	Omnivore
	Forcipiger flavissimus	0.17	0.030	Mobile Invert.
	Forcipiger longirostris	0.19	0.015	Mobile Invert.
Labridae	*Bodianus diana*	0.19	0.009	Mobile Invert.
	Bodianus dictynna	0.22	0.010	Mobile Invert.
	Choerodon zosterophorus	0.22	0.012	Mobile Invert.
	Halichoeres chrysus	0.39	<0.001	Mobile Invert.
	Halichoeres prosopeion	0.33	0.002	Mobile Invert.
Lethrinidae	*Monotaxis grandoculis*	0.22	0.020	Mobile Invert.
Lutjanidae	*Macolor macularis*	0.29	0.004	Piscivore & Mobile Invert.
Monacanthidae	*Paraluteres prionurus*	0.17	0.031	Omnivore
Pomacanthidae	*Centropyge bicolor*	0.3	0.002	Mobile Invert.
	Genicanthus lamarck	0.29	0.001	Planktivore
Pomacentridae	*Pomacentrus amboinensis*	0.36	<0.001	Herbivore & Planktivore
	Pomacentrus nigromanus	0.27	0.007	Planktivore
Serranidae	*Plectropomus oligacanthus*	0.19	0.016	Piscivore & Mobile Invert.
	Variola albimarginata	0.19	0.034	Piscivore
Tetraodontidae	*Canthigaster valentini*	0.17	0.031	Omnivore

MCE				
Balistidae	*Abalistes stellatus*	0.41	<0.001	Piscivore & Mobile Invert.
	Sufflamen fraenatum	0.23	0.023	Mobile Invert.
Carangidae	*Carangoides ferdau*	0.35	0.011	Piscivore & Mobile Invert.
Labridae	*Choerodon jordani*	0.25	0.003	Mobile Invert.
Lethrinidae	*Lethrinus amboinensis*	0.25	0.017	Piscivore & Mobile Invert.
	Lethrinus lentjan	0.17	0.031	Piscivore & Mobile Invert.
Malacanthidae	*Hoplolatilus purpureus*	0.25	0.002	Planktivore
Mullidae	*Parupeneus heptacanthus*	0.20	0.027	Piscivore & Mobile Invert.
Nemipteridae	*Pentapodus nagasakiensis*	0.36	0.004	Mobile Invert.
	Pentapodus paradiseus	0.39	<0.001	Piscivore & Mobile Invert.
	Scolopsis vosmeri	0.25	0.003	Mobile Invert.
Pomacentridae	*Pomacentrus nagasakiensis*	0.17	0.031	Herbivore & Planktivore

Table 3. Fish trophic group species richness and comparison of abundance across the depth
gradient. Richness represents the number of unique species recorded for a trophic grouping
within a depth zone. Abundance represents the mean total abundance per BRUV drop
within a depth zone based on summing species level MaxN for all species within a tropic
group per BRUV drop. Changes in abundance across the depth gradient were tested with a
permutational-ANOVA of the form Trophic Group Abundance ~ Region + Depth. As
permutational-ANOVA uses sequential sums of squares, the biomass depth effects
represent the effect of depth on trophic group biomass after controlling for the effect of
region. Depth was treated as continuous coded as 1 (shallow), 2 (intermediate), 3 (MCE) to
incorporate the gradient of depths. Significant p-values at the $p<0.05$ level are shown in
bold.

Trophic Group	Shallow		Intermediate		MCE		Abundance depth effect	
	Richness	Abundance (ind./BRUV)	Richness	Abundance (ind./BRUV)	Richness	Abundance (ind./BRUV)	Pseudo-F	p(perm)
Herbivore	50	12.92 ± 1.91	38	7.26 ± 0.84	9	2.35 ± 0.52	8.94	**0.001**
Herbivore & Planktivore	36	17.61 ± 3.65	35	14.56 ± 2.89	10	9.39 ± 4.11	1.25	0.301
Planktivore	56	46.06 ± 7.19	59	46.35 ± 9.38	26	8.62 ± 1.9	8.78	**<0.001**
Planktivore & Mobile Invertebrate	11	2.88 ± 0.62	14	4.77 ± 1.56	7	2.5 ± 0.97	1.02	0.394
Omnivore	43	10.67 ± 3.14	29	7.05 ± 1.24	18	3.41 ± 0.63	2.66	**0.030**
Corallivore	22	6.14 ± 0.53	20	2.98 ± 0.39	7	2.42 ± 0.38	22.77	**<0.001**
Piscivore & Mobile Invertebrate	95	11.83 ± 0.83	95	16.21 ± 1.65	66	17.72 ± 3.04	2.66	**0.071**
Piscivore	16	4.52 ± 1.45	18	3.95 ± 1.05	10	1.75 ± 0.3	1.93	0.134

Table 4. Fish species potentially driving changes in herbivore, planktivore, omnivore,
corallivore, and piscivore and mobile invertebrate feeder abundance across the depth
gradient. Abundance represents the mean abundance per BRUV drop (MaxN) within a depth
zone. Changes in abundance across the depth gradient were tested with a permutational-
ANOVA of the form MaxN ~ Region + Depth. Species mean MaxN for each site and depth
combination was used in this analysis. As permutational-ANOVA uses sequential sums of
squares, the depth effect represent the effect of depth on species abundance after
controlling for the effect of region. Depth was treated as continuous coded as 1 (shallow), 2
(intermediate), 3 (MCE) to incorporate the gradient of depths. Significant p-values at the
p<0.05 level are shown in bold.

Species	Biomass (g/BRUV)			Biomass depth effect	
	Shallow	Intermediate	MCE	Pseudo-F	p(perm)
Herbivore					
Acanthurus blochii	0.92 ± 0.64	0.69 ± 0.22	0 ± 0	1.50	0.161
Acanthurus maculiceps	2.93 ± 1.45	0.36 ± 0.21	0 ± 0	3.54	**0.003**
Acanthurus nigricans	0.17 ± 0.12	0.06 ± 0.06	0 ± 0	1.20	0.289
Centropyge tibicen	0.14 ± 0.09	0.07 ± 0.05	0 ± 0	1.35	0.304
Centropyge vrolikii	0.06 ± 0.04	0 ± 0	0 ± 0	2.04	0.330
Chlorurus microrhinos	0.11 ± 0.07	0 ± 0	0 ± 0	2.79	0.064
Dischistodus melanotus	0.35 ± 0.23	0.03 ± 0.03	0 ± 0	2.14	**0.015**
Dischistodus pseudochrysopoecilus	0.39 ± 0.3	0.11 ± 0.09	0 ± 0	1.24	0.325
Naso vlamingii	0.36 ± 0.28	0.06 ± 0.04	0.08 ± 0.08	1.00	0.422
Scarus niger	0.03 ± 0.03	0 ± 0	0 ± 0	1.00	1.000
Scarus psittacus	0.11 ± 0.07	0.06 ± 0.06	0 ± 0	1.21	0.371
Toxotes jaculatrix	0.08 ± 0.06	0 ± 0	0 ± 0	1.83	0.327
Planktivore					
Caesio cuning	3.33 ± 1.65	6.15 ± 3.14	0 ± 0	2.25	0.076
Caesio lunaris	7.03 ± 2.79	2.03 ± 1.35	0.31 ± 0.25	3.76	**0.020**
Caesio teres	3.5 ± 2.05	2.58 ± 2.36	0.03 ± 0.03	1.02	0.415
Crenimugil crenilabis	1.22 ± 1.22	0 ± 0	0 ± 0	1.00	1.000
Neopomacentrus azysron	0.11 ± 0.09	0 ± 0	0 ± 0	1.62	0.329
Odonus niger	4.93 ± 1.82	7.29 ± 2.15	1.33 ± 0.59	3.52	**0.030**
Pterocaesio marri	2.58 ± 1.89	3.79 ± 2.32	2.33 ± 1.99	0.14	0.904
Omnivore					
Acanthurus triostegus	0.22 ± 0.2	0 ± 0	0 ± 0	1.29	0.326
Canthigaster amboinensis	0 ± 0	0.06 ± 0.06	0 ± 0	1.00	1.000
Canthigaster valentini	0 ± 0	0.17 ± 0.09	0 ± 0	3.21	**0.008**
Chaetodon citrinellus	0.24 ± 0.1	0 ± 0	0 ± 0	5.29	**0.005**
Chaetodon ephippium	0.36 ± 0.14	0.14 ± 0.08	0 ± 0	3.62	**0.019**
Chaetodon semeion	0.18 ± 0.07	0 ± 0	0 ± 0	6.85	**0.001**
Chaetodon ulietensis	0.14 ± 0.1	0.14 ± 0.08	0.03 ± 0.03	0.77	0.482
Chlorurus bowersi	0.03 ± 0.03	0 ± 0	0 ± 0	1.00	1.000
Kyphosus bigibbus	0.61 ± 0.61	0 ± 0	0 ± 0	1.00	1.000
Kyphosus cinerascens	1.03 ± 0.97	0 ± 0	0 ± 0	1.12	0.106
Kyphosus vaigiensis	1.72 ± 1.59	0 ± 0	0 ± 0	1.18	0.324
Naso thynnoides	0.08 ± 0.08	3.56 ± 1.54	0.03 ± 0.03	5.10	**0.001**

Neoglyphidodon melas	0.36 ± 0.14	0.06 ± 0.06	0 ± 0	4.97	**0.007**
Paraluteres prionurus	0 ± 0	0.19 ± 0.12	0 ± 0	2.67	**0.030**
Pomacanthus navarchus	0.06 ± 0.04	0.03 ± 0.03	0 ± 0	1.03	0.515
Corallivore					
Chaetodon kleinii	0.99 ± 0.26	0.78 ± 0.17	0.61 ± 0.2	0.78	0.480
Chaetodon oxycephalus	0.17 ± 0.09	0.06 ± 0.04	0.06 ± 0.06	0.91	0.455
Diproctacanthus xanthurus	1.04 ± 0.35	0.25 ± 0.15	0.08 ± 0.06	5.31	**0.004**
Piscivore & Mobile Invertebrate					
Grammatorcynus bilineatus	0 ± 0	0.08 ± 0.05	0.11 ± 0.09	1.04	0.425
Gymnocranius grandoculis	0.03 ± 0.03	0.19 ± 0.12	0.25 ± 0.12	1.33	0.270
Lethrinus microdon	0 ± 0	0.25 ± 0.07	0.17 ± 0.08	3.78	**0.023**
Lethrinus nebulosus	0 ± 0	0.18 ± 0.08	0.28 ± 0.17	1.63	0.172
Lethrinus semicinctus	0.12 ± 0.06	1.11 ± 0.63	0.97 ± 0.31	1.71	0.153
Lutjanus argentimaculatus	0 ± 0	0 ± 0	0.03 ± 0.03	1.00	1.000
Parupeneus cyclostomus	0.17 ± 0.06	0.51 ± 0.16	0.35 ± 0.14	1.83	0.168
Pentapodus aureofasciatus	0.08 ± 0.05	1.47 ± 0.47	1.29 ± 0.72	2.27	0.089
Pentapodus caninus	0.03 ± 0.03	1.54 ± 0.92	3.72 ± 1.86	2.38	0.071
Pentapodus emeryii	0.25 ± 0.15	0.68 ± 0.16	1.04 ± 0.31	3.30	**0.037**
Pentapodus numberii	0 ± 0	0.14 ± 0.07	0.46 ± 0.43	0.87	0.611
Pentapodus porosus	0.06 ± 0.04	0.86 ± 0.37	0.31 ± 0.31	2.15	0.095

Table 5. Linear mixed-effect model of the effects of depth, benthic cover, wave exposure,
human population within 10 km, and the distance to major market on fourth root
transformed fish biomass. Results are shown for total fish biomass, and for high/very high
value species and sharks. Biomass (g) was fourth root transformed prior to analysis. Sharks
included in commercially valuable biomass were all individuals of the families:
Carcharhinidae, Ginglymostomatidae, Hemiscylliidae, and Rhinidae. Benthic PC1 indicates
the first principal components axis from a PCA of each component of the benthic HAS per
BRUV drop.

	Estimate	SE	t-value	$\chi 2$	p
Total fish biomass					
Intercept	21.354	2.823	7.57		
Depth	-2.884	0.865	-3.33	10.74	**0.001**
Benthic PC1	0.549	0.285	1.92	3.65	0.056
Wave exposure	-0.719	0.580	-1.24	1.47	0.225
Human population	-0.003	0.001	-2.07	4.19	**0.041**
Distance to market	-0.020	0.009	-2.28	4.91	**0.027**
Depth:Benthic PC1	-0.407	0.132	-3.08	9.22	**0.002**
Commercially valuable fish biomass					
Intercept	19.867	3.175	6.26		
Depth	-3.235	0.974	-3.32	10.63	**0.001**
Benthic PC1	0.551	0.227	2.43	5.74	**0.017**
Wave exposure	-0.584	0.652	-0.90	0.79	0.374
Human population	-0.003	0.001	-1.89	3.48	0.062
Distance to market	-0.017	0.010	-1.69	2.81	0.094
Depth:Benthic PC1	-0.372	0.105	-3.54	11.99	**0.001**

 Electronic Supplementary Material

958
959 ESM Figure 1. PCA of benthic and fish community structure differences based on depth and
960 region. (A) Benthic communities based on mean HAS per depth at each site. (B) Fish
961 communities weighed on abundance. 95% confidence ellipses are shown. Variable labels on
962 (A) are: rugosity (R), variety of growth forms (GF), height (H), refuge size category (RS), live
963 cover (LC), and hard substratum (HS).
964

965 ESM Table 1. Number of BRUV drops per site and depth for both regions. A minimum of two
966 BRUV drops were conducted per depth zone at each site with the exception of MCEs at
967 WNTZ site 7 where recorded footage was only analyzable from one BRUV.

Region/Site	Number of replicate BRUV drops per depth		
	Shallow	Intermediate	MCE
Dampier (DNTZ)			
1	3	2	3
2	3	2	2
3	3	2	3
4	2	2	2
5	2	2	2
Dampier (DUZ)			
1	3	3	2
2	3	2	2
3	3	2	2
4	2	2	3
5	2	2	3
Waigeo (WNTZ)			
1	2	2	3
2	2	3	2
3	2	2	2
4	2	2	2
5	2	4	2
6	2	2	2
7	2	2	1
Waigeo (WUZ)			
1	2	2	2
2	2	2	2
3	2	2	2
4	2	2	2
5	2	2	2
6	2	4	2
7	2	2	2

ESM Table 2. GPS coordinates for each BRUV drop. 'Region' indicates both the region and
fisheries type as follows: DNTZ – Dampier no-take zone, DUZ – Dampier use zone
(sustainable fisheries zone within the MPA), WNTZ – Waigeo no-take zone, and WUZ –
Waigeo use area (area open to fisheries outside the MPA). Depth represents depth zone as
follows: S – Shallow, M – Mid (intermediate), and D – Deep (MCE). 'Site number' refers to
the site number as used in the manuscript. The 'Raw data site number' and 'Replicate
numbers' represent the original Conservation International survey site and replicate
numbers, which are used in the raw data. Not all sites/replicates surveyed were analyzed in
this study, hence the numbers are not continuous. All GPS points are given in WGS84 format
decimal degrees.

Region	Site number	Raw data site number	Depth	Replicate number	Longitude	Latitude
DNTZ	1	1	D	1	130.530697	-0.794603
DNTZ	1	1	D	2	130.526488	-0.7905658
DNTZ	1	1	D	3	130.523987	-0.783556
DNTZ	1	1	M	1	130.524713	-0.795062
DNTZ	1	1	M	3	130.52132	-0.7972476
DNTZ	1	1	S	1	130.523	-0.794309
DNTZ	1	1	S	2	130.520595	-0.798318
DNTZ	1	1	S	3	130.532855	-0.7987367
DNTZ	2	2	D	1	130.508027	-0.7989828
DNTZ	2	2	D	2	130.513062	-0.7867608
DNTZ	2	2	M	3	130.509315	-0.7927859
DNTZ	2	2	M	4	130.515608	-0.7952866
DNTZ	2	2	S	1	130.507272	-0.793176
DNTZ	2	2	S	2	130.497068	-0.794196
DNTZ	2	2	S	3	130.511291	-0.8056208
DNTZ	3	3	D	1	130.452206	-0.843469
DNTZ	3	3	D	2	130.454281	-0.851093
DNTZ	3	3	D	3	130.448862	-0.855004
DNTZ	3	3	M	1	130.455182	-0.847037
DNTZ	3	3	M	2	130.449541	-0.839482
DNTZ	3	3	S	1	130.449029	-0.83451
DNTZ	3	3	S	2	130.453843	-0.855849
DNTZ	3	3	S	3	130.452008	-0.859498
DNTZ	4	4	D	2	130.434489	-0.863857
DNTZ	4	4	D	3	130.440475	-0.8744669
DNTZ	4	4	M	1	130.441935	-0.8590817
DNTZ	4	4	M	2	130.449238	-0.8638667
DNTZ	4	4	S	1	130.439157	-0.857953
DNTZ	4	4	S	2	130.434941	-0.860401
DNTZ	5	8	D	1	130.353733	-0.49002
DNTZ	5	8	D	2	130.34567	-0.488285
DNTZ	5	8	M	1	130.357852	-0.487746
DNTZ	5	8	M	3	130.338402	-0.4898128

DNTZ	5	8	S	1	130.362512	-0.486074
DNTZ	5	8	S	3	130.358208	-0.483863
DUZ	1	1	D	2	130.635423	-0.788386
DUZ	1	1	D	3	130.641301	-0.788727
DUZ	1	1	M	1	130.626402	-0.793261
DUZ	1	1	M	2	130.625086	-0.796811
DUZ	1	1	M	4	130.631527	-0.791067
DUZ	1	1	S	1	130.626423	-0.794241
DUZ	1	1	S	2	130.630811	-0.791413
DUZ	1	1	S	3	130.635497	-0.790103
DUZ	2	2	D	1	130.544367	-0.82834
DUZ	2	2	D	3	130.527688	-0.827546
DUZ	2	2	M	1	130.541011	-0.824919
DUZ	2	2	M	2	130.536438	-0.826965
DUZ	2	2	S	1	130.540425	-0.824592
DUZ	2	2	S	2	130.536746	-0.826036
DUZ	2	2	S	3	130.537214	-0.830708
DUZ	3	4	D	1	130.570988	-0.788506
DUZ	3	4	D	2	130.575959	-0.787782
DUZ	3	4	M	1	130.575876	-0.795152
DUZ	3	4	M	3	130.563995	-0.797067
DUZ	3	4	S	1	130.570746	-0.792993
DUZ	3	4	S	2	130.579564	-0.797969
DUZ	3	4	S	3	130.561255	-0.802375
DUZ	4	7	D	1	130.637406	-0.569417
DUZ	4	7	D	2	130.632069	-0.570607
DUZ	4	7	D	3	130.626858	-0.571275
DUZ	4	7	M	2	130.623917	-0.572667
DUZ	4	7	M	3	130.638624	-0.569266
DUZ	4	7	S	2	130.634321	-0.5711356
DUZ	4	7	S	3	130.61781	-0.573148
DUZ	5	9	D	1	130.598434	-0.598411
DUZ	5	9	D	2	130.603696	-0.594229
DUZ	5	9	D	3	130.607279	-0.589442
DUZ	5	9	M	1	130.576687	-0.609784
DUZ	5	9	M	3	130.594597	-0.599805
DUZ	5	9	S	1	130.566139	-0.61247
DUZ	5	9	S	2	130.571207	-0.610765
WNTZ	1	1	D	1	130.069786	0.134305
WNTZ	1	1	D	2	130.06853	0.15673002
WNTZ	1	1	D	3	130.070163	0.14955629
WNTZ	1	1	M	2	130.061787	0.15101625
WNTZ	1	1	M	3	130.065415	0.15681
WNTZ	1	1	S	2	130.060455	0.153615
WNTZ	1	1	S	3	130.065935	0.15262331
WNTZ	2	2	D	2	130.120479	0.137607

WNTZ	2	2	D	3	130.13899	0.133177
WNTZ	2	2	M	2	130.130658	0.134857
WNTZ	2	2	M	3	130.136195	0.13559832
WNTZ	2	2	M	4	130.147438	0.133255
WNTZ	2	2	S	2	130.128722	0.134322
WNTZ	2	2	S	3	130.142698	0.13507001
WNTZ	3	3	D	1	130.223638	0.09694
WNTZ	3	3	D	3	130.210911	0.105589
WNTZ	3	3	M	2	130.216657	0.1027
WNTZ	3	3	M	3	130.212705	0.10462
WNTZ	3	3	S	1	130.219081	0.099818
WNTZ	3	3	S	2	130.228548	0.095535
WNTZ	4	5	D	1	130.125093	0.12996401
WNTZ	4	5	D	3	130.129542	0.120686
WNTZ	4	5	M	2	130.132819	0.12261601
WNTZ	4	5	M	3	130.138764	0.12245701
WNTZ	4	5	S	1	130.123098	0.125012
WNTZ	4	5	S	2	130.133877	0.120297
WNTZ	5	6	D	1	130.076229	0.129769
WNTZ	5	6	D	2	130.067475	0.13330202
WNTZ	5	6	M	1	130.062805	0.13976103
WNTZ	5	6	M	2	130.080999	0.147264
WNTZ	5	6	M	3	130.079717	0.13855998
WNTZ	5	6	M	4	130.078602	0.13337703
WNTZ	5	6	S	1	130.072124	0.13152597
WNTZ	5	6	S	3	130.07027	0.13389001
WNTZ	6	7	D	1	129.998199	0.16885721
WNTZ	6	7	D	2	130.004007	0.163066
WNTZ	6	7	M	1	130.002036	0.17793842
WNTZ	6	7	M	2	130.002848	0.17372911
WNTZ	6	7	S	1	130.006936	0.17944406
WNTZ	6	7	S	2	130.010678	0.178129
WNTZ	7	9	D	3	130.030903	0.17054901
WNTZ	7	9	M	2	130.040895	0.17433504
WNTZ	7	9	M	3	130.030178	0.165611
WNTZ	7	9	S	1	130.02632	0.178
WNTZ	7	9	S	2	130.039838	0.167497
WUZ	1	2	D	1	130.171925	0.004977
WUZ	1	2	D	2	130.177582	0.006271
WUZ	1	2	M	1	130.168195	-0.003426
WUZ	1	2	M	2	130.166462	0.001249
WUZ	1	2	S	2	130.176871	0.00416229
WUZ	1	2	S	3	130.1771	-0.000226
WUZ	2	3	D	2	130.143002	0.004364
WUZ	2	3	D	3	130.118273	0.001968
WUZ	2	3	M	2	130.12406	0.000755

WUZ	2	3	M	3	130.137578	0.00481004
WUZ	2	3	S	1	130.121792	-0.0110666
WUZ	2	3	S	2	130.124705	-0.0037417
WUZ	3	4	D	2	130.123823	-0.026527
WUZ	3	4	D	3	130.128838	-0.025605
WUZ	3	4	M	1	130.146495	-0.018221
WUZ	3	4	M	3	130.141314	-0.021141
WUZ	3	4	S	1	130.134777	-0.025737
WUZ	3	4	S	3	130.126522	-0.036198
WUZ	4	5	D	1	130.159497	-0.056352
WUZ	4	5	D	2	130.160028	-0.05167
WUZ	4	5	M	1	130.165126	-0.061347
WUZ	4	5	M	3	130.157356	-0.045629
WUZ	4	5	S	2	130.154767	-0.0556245
WUZ	4	5	S	3	130.15199	-0.0405267
WUZ	5	6	D	1	130.174079	-0.0831957
WUZ	5	6	D	2	130.188744	-0.091517
WUZ	5	6	M	1	130.178132	-0.085585
WUZ	5	6	M	3	130.184552	-0.102447
WUZ	5	6	S	1	130.191393	-0.098528
WUZ	5	6	S	3	130.179898	-0.094061
WUZ	6	8	D	1	130.10519	-0.0707138
WUZ	6	8	D	3	130.11609	-0.09311
WUZ	6	8	M	1	130.109248	-0.068807
WUZ	6	8	M	2	130.113974	-0.089298
WUZ	6	8	M	3	130.114045	-0.084448
WUZ	6	8	M	4	130.114045	-0.084448
WUZ	6	8	S	1	130.111878	-0.0725609
WUZ	6	8	S	2	130.097381	-0.06316
WUZ	7	9	D	1	130.067343	-0.067851
WUZ	7	9	D	2	130.043359	-0.073457
WUZ	7	9	M	3	130.048681	-0.074707
WUZ	7	9	M	4	130.027712	-0.073842
WUZ	7	9	S	2	130.067135	-0.072522
WUZ	7	9	S	3	130.038239	-0.074227

ESM Table 3. Potential new species records for Raja Ampat. Table contains species we
recorded in Raja Ampat absent from Allen and Erdmann (2009) Raja Ampat records. Allen
and Erdmann combined field surveys within the Bird's Head Seascape (BHS) including Raja
Ampat with literature searching to compile the most comprehensive fish species list for the
region. Where Allen and Erdmann (2009) identified a species record from the wider BHS
region—but not Raja Ampat—this is noted with the location. Other BHS locations include
Cenderawasih Bay (CB) and Fakfak/Kaimana (FK). Species in this table represent absences
from Allen and Erdmann (2008), and we believe may represent new records for Raja Ampat.

Species	Species Authority	Notes
Bodianus dictynna	Gomon, 2006	Previous records of *Bodianus diana* (Lacepède, 1801) from the region are likely to be *Bodianus dictynna*.
Carangoides oblongus	Cuvier, 1833	
Carcharhinus obscurus	Lesueur, 1818	
Cheilodipterus parazonatus	Gon, 1993	Previously recorded in CB
Chelmon muelleri	Klunzinger, 1879	Previously recorded in FK
Choerodon monostigma	Ogilby, 1910	
Chromis fumea	Tanaka, 1917	
Chrysiptera flavipinnis	Allen & Robertson, 1974	
Chrysiptera glauca	Cuvier, 1830	Previously recorded in CB
Chrysiptera parasema	Fowler, 1918	
Cirrhilabrus beauperryi	Allen, Drew & Barber, 2008	
Cirrhilabrus cyanopleura	Bleeker, 1851	
Coradion melanopus	Cuvier, 1831	Previously recorded in CB, FK
Gymnocranius elongatus	Senta, 1973	Previously recorded in FK
Hoplolatilus chlupatyi	Klausewitz, McCosker, Randall & Zetzsche, 1978	Previously recorded in FK
Lactoria cornuta	Linnaeus, 1758	Previously recorded in CB
Lagocephalus sceleratus	Gmelin, 1789	
Lethrinus microdon	Valenciennes, 1830	Previously recorded in FK
Lethrinus rubrioperculatus	Sato, 1978	
Lutjanus maxweberi	Popta, 1921	Previously recorded in CB
Myripristis botche	Cuvier, 1829	Previously recorded in CB, FK
Naso caesius	Randall & Bell, 1992	
Parupeneus chrysopleuron	Temminck & Schlegel, 1843	Previously recorded in FK
Pentapodus caninus	Cuvier, 1830	Previously recorded in CB
Pentapodus nagasakiensis	Tanaka, 1915	

Pentapodus porosus	Valenciennes, 1830	Previously recorded in FK
Pseudojuloides splendens	Victor, 2017	Previously recorded in FK as *Pseudojuloides cerasinus*
Rhabdamia spilota	Allen & Kuiter, 1994	Previously recorded in CB, FK
Scomberoides tol	Cuvier, 1832	Previously recorded in CB, FK
Siganus doliatus	Guérin-Méneville, 1829-38	Previously recorded in FK
Valenciennea wardii	Playfair, 1867	Previously recorded in FK

989 ESM Table 4. Depth extensions for species. Max fishbase depth represents the maximum
990 depth recorded for the species as stated in fishbase (Froese & Pauly, 2019). Max. Raja
991 Ampat depth represents the maximum BRUV drop depth that the species was recorded on.

FAMILY	Species	Max. Fishbase Depth	Max. Raja Ampat Depth
ACANTHURIDAE	*Ctenochaetus striatus*	35	42
	Naso caeruleacauda	45	50
	Naso thynnoides	40	45
APOGONIDAE	*Ostorhinchus hoevenii*	30	40
BALISTIDAE	*Odonus niger*	40	66
	Rhinecanthus verrucosus	20	47
	Sufflamen chrysopterum	30	52
BLENNIIDAE	*Aspidontus taeniatus*	25	61
	Meiacanthus crinitus	20	51
	Meiacanthus grammistes	20	59
CAESIONIDAE	*Pterocaesio marri*	30	50
	Pterocaesio randalli	30	40
CHAETODONTIDAE	*Chaetodon oxycephalus*	40	43
	Chaetodon ulietensis	30	42
	Chaetodon vagabundus	30	44
	Coradion altivelis	30	60
CIRRHITIDAE	*Cirrhitichthys aprinus*	40	47
EPHIPPIDAE	*Platax batavianus*	40	64
	Platax orbicularis	30	59
GERREIDAE	*Gerres erythrourus*	40	49
HAEMULIDAE	*Plectorhinchus lineatus*	35	59
	Plectorhinchus polytaenia	40	41
LABRIDAE	*Anampses neoguinaicus*	30	52
	Choerodon jordani	30	66
	Choerodon zosterophorus	40	49
	Coris dorsomacula	40	50
	Diproctacanthus xanthurus	25	38
	Halichoeres bicolor	20	42
	Halichoeres chloropterus	10	27
	Halichoeres melanochir	25	55
	Halichoeres nigrescens	10	20
	Halichoeres prosopeion	40	64
	Halichoeres scapularis	20	50
	Iniistius twistii	20	65
	Labroides bicolor	40	60
	Labroides dimidiatus	40	66
	Oxycheilinus celebicus	40	47
	Stethojulis bandanensis	30	35
	Thalassoma amblycephalum	15	59
LETHRINIDAE	*Gnathodentex aureolineatus*	30	52

	Lethrinus atkinsoni	30	60
	Lethrinus amboinensis	30	61
	Lethrinus erythropterus	25	55
	Lethrinus harak	20	65
	Lethrinus laticaudis	35	60
	Lethrinus obsoletus	30	52
	Lethrinus ornatus	30	47
	Lethrinus semicinctus	35	66
LUTJANIDAE	*Lutjanus biguttatus*	36	49
MICRODESMIDAE	*Gunnellichthys pleurotaenia*	15	44
MONACANTHIDAE	*Amanses scopas*	18	55
	Paramonacanthus japonicus	46	61
	Pseudomonacanthus macrurus	10	18
MONODACTYLIDAE	*Monodactylus argenteus*	12	27
	Upeneus tragula	42	60
NEMIPTERIDAE	*Pentapodus aureofasciatus*	35	55
	Pentapodus emeryii	35	61
	Pentapodus trivittatus	30	43
	Scolopsis bilineata	25	47
	Scolopsis ciliata	25	66
	Scolopsis margaritifera	25	50
	Scolopsis temporalis	35	51
	Scolopsis trilineata	20	61
	Scolopsis vosmeri	25	64
PHOLIDICHTHYIDAE	*Pholidichthys leucotaenia*	30	50
PINGUIPEDIDAE	*Parapercis hexophtalma*	25	28
	Parapercis lineopunctata	35	51
	Parapercis snyderi	40	51
	Parapercis tetracantha	25	47
POMACANTHIDAE	*Centropyge bicolor*	25	43
	Chaetodontoplus dimidiatus	35	60
	Chaetodontoplus mesoleucus	20	40
POMACENTRIDAE	*Amblyglyphidodon batunai*	12	30
	Amblypomacentrus clarus	25	46
	Cheiloprion labiatus	3	35
	Chromis margaritifer	20	52
	Chrysiptera bleekeri	35	52
	Chrysiptera cyanea	10	28
	Dischistodus fasciatus	8	27
	Dischistodus melanotus	12	35
	Dischistodus pseudochrysopoecilus	5	27
	Neoglyphidodon crossi	12	18
	Neoglyphidodon oxyodon	4	21
	Neopomacentrus filamentosus	12	27
	Pomacentrus adelus	8	15

	Pomacentrus amboinensis	40	59
	Pomacentrus chrysurus	5	27
	Pomacentrus lepidogenys	12	28
	Pomacentrus littoralis	5	28
	Pomacentrus nagasakiensis	35	47
	Pomacentrus simsiang	10	35
	Pomacentrus tripunctatus	3	35
	Stegastes albifasciatus	4	15
PSEUDOCHROMIDAE	Pseudochromis perspicillatus	27	51
SCARIDAE	Bolbometopon muricatum	40	41
	Scarus oviceps	20	40
	Scarus tricolor	25	43
TETRAODONTIDAE	Arothron immaculatus	30	49
	Arothron mappa	30	45

993 ESM Table 5. Change in fish species abundance with depth across all sites. Abundance represents the mean MaxN per BRUV drop within a
994 depth zone. 'Mid' represents intermediate depths. Changes in abundance across the depth gradient were tested with a permutational-ANOVA
995 (9999 permutations) of the form Abundance ~ Depth. Pseudo-F and p(perm) values represent F and p values associated with the depth effect.
996 Depth was treated as continuous coded as 1 (shallow), 2 (intermediate), 3 (MCE) to incorporate the gradient of depths. Abundance data with
997 significant depth effects (p<0.05) are shown in bold.
998

Family	Species	Shallow	Mid	Deep	Pseudo-F	p(perm)
Acanthuridae	*Acanthurus bariene*	0.03 ± 0.03	0.02 ± 0.02	0 ± 0	0.97	0.441
Acanthuridae	*Acanthurus blochii*	0.8 ± 0.64	0.56 ± 0.22	0 ± 0	2.15	0.150
Acanthuridae	*Acanthurus fowleri*	0.33 ± 0.1	0.56 ± 0.33	0.02 ± 0.02	1.17	0.367
Acanthuridae	*Acanthurus leucocheilus*	**0.15 ± 0.07**	**0.13 ± 0.06**	**0 ± 0**	**4.66**	**0.039**
Acanthuridae	*Acanthurus lineatus*	**0.87 ± 0.31**	**0 ± 0**	**0 ± 0**	**11.63**	**<0.001**
Acanthuridae	*Acanthurus maculiceps*	**2.31 ± 1.07**	**0.24 ± 0.12**	**0 ± 0**	**6.84**	**<0.001**
Acanthuridae	*Acanthurus mata*	0.29 ± 0.27	0.44 ± 0.37	0.06 ± 0.06	0.37	0.614
Acanthuridae	*Acanthurus nigricans*	0.13 ± 0.09	0.03 ± 0.03	0 ± 0	2.59	0.140
Acanthuridae	*Acanthurus nigricauda*	**0.75 ± 0.39**	**0.04 ± 0.03**	**0 ± 0**	**5.35**	**0.011**
Acanthuridae	*Acanthurus nigrofuscus*	0.16 ± 0.09	0.13 ± 0.07	0 ± 0	2.86	0.095
Acanthuridae	*Acanthurus nubilus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Acanthuridae	*Acanthurus olivaceus*	0.25 ± 0.16	0.15 ± 0.06	0.04 ± 0.04	2.18	0.178
Acanthuridae	*Acanthurus pyroferus*	0.16 ± 0.08	0.23 ± 0.07	0 ± 0	3.20	0.081
Acanthuridae	*Acanthurus thompsoni*	0.04 ± 0.04	0.34 ± 0.15	0 ± 0	0.10	0.785
Acanthuridae	*Acanthurus triostegus*	0.17 ± 0.15	0 ± 0	0 ± 0	1.95	0.223
Acanthuridae	*Acanthurus xanthopterus*	**0.13 ± 0.06**	**0.01 ± 0.01**	**0 ± 0**	**7.75**	**0.004**
Acanthuridae	*Ctenochaetus binotatus*	0 ± 0	0.04 ± 0.03	0 ± 0	0.00	1.000
Acanthuridae	*Ctenochaetus cyanocheilus*	0.08 ± 0.05	0.08 ± 0.06	0 ± 0	1.59	0.256
Acanthuridae	*Ctenochaetus striatus*	**1.14 ± 0.25**	**0.52 ± 0.21**	**0.02 ± 0.02**	**18.14**	**<0.001**
Acanthuridae	*Ctenochaetus tominiensis*	0 ± 0	0.08 ± 0.06	0 ± 0	0.00	1.000
Acanthuridae	*Naso annulatus*	0.3 ± 0.17	0.21 ± 0.15	0.06 ± 0.05	1.57	0.236
Acanthuridae	*Naso brachycentron*	0.35 ± 0.2	0.3 ± 0.27	0 ± 0	1.68	0.215
Acanthuridae	*Naso brevirostris*	**0.63 ± 0.24**	**0.02 ± 0.02**	**0 ± 0**	**9.88**	**<0.001**
Acanthuridae	*Naso caeruleacauda*	0.35 ± 0.27	0.08 ± 0.08	0.1 ± 0.1	0.98	0.395
Acanthuridae	*Naso caesius*	0.46 ± 0.19	0.39 ± 0.22	0.02 ± 0.02	3.40	0.076
Acanthuridae	*Naso hexacanthus*	0.51 ± 0.25	0.43 ± 0.16	0.13 ± 0.11	2.18	0.150
Acanthuridae	*Naso lituratus*	**0.46 ± 0.12**	**0.17 ± 0.06**	**0 ± 0**	**17.59**	**<0.001**
Acanthuridae	*Naso lopezi*	0.03 ± 0.02	0 ± 0	0 ± 0	3.01	0.214
Acanthuridae	*Naso mcdadei*	0.03 ± 0.03	0 ± 0	0 ± 0	1.51	0.680
Acanthuridae	*Naso minor*	0.65 ± 0.62	0.14 ± 0.11	0.2 ± 0.19	0.69	0.544

Acanthuridae	*Naso thynnoides*	0.04 ± 0.04	2.11 ± 0.84	0.02 ± 0.02	0.00	0.982
Acanthuridae	*Naso tonganus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Acanthuridae	*Naso unicornis*	**0.2 ± 0.1**	**0.03 ± 0.02**	**0 ± 0**	**5.42**	**0.010**
Acanthuridae	*Naso vlamingii*	0.27 ± 0.21	0.06 ± 0.05	0.06 ± 0.06	1.33	0.354
Acanthuridae	*Paracanthurus hepatus*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.675
Acanthuridae	*Zebrasoma scopas*	**0.47 ± 0.08**	**0.13 ± 0.08**	**0.04 ± 0.04**	**18.43**	**<0.001**
Acanthuridae	*Zebrasoma velifer*	0.06 ± 0.05	0.08 ± 0.08	0 ± 0	0.65	0.592
Antennariidae	*Antennatus rosaceus*	0 ± 0	0 ± 0	0.02 ± 0.02	1.51	0.667
Apogonidae	*Cheilodipterus nigrotaeniatus*	0.02 ± 0.02	0.01 ± 0.01	0 ± 0	1.22	0.436
Apogonidae	*Cheilodipterus parazonatus*	0.08 ± 0.05	0.04 ± 0.03	0.13 ± 0.07	0.58	0.490
Apogonidae	*Ostorhinchus cavitensis*	0.1 ± 0.1	0 ± 0	0 ± 0	1.51	0.669
Apogonidae	*Ostorhinchus hoevenii*	0 ± 0	0 ± 0	0.04 ± 0.04	1.51	0.663
Apogonidae	*Pristiapogon exostigma*	0.27 ± 0.27	0 ± 0	0 ± 0	1.51	0.665
Apogonidae	*Rhabdamia spilota*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Apogonidae	*Taeniamia fucata*	0 ± 0	0.08 ± 0.08	0 ± 0	0.00	1.000
Atherinidae	*Atherinomorus lacunosus*	0.19 ± 0.19	0 ± 0	0 ± 0	1.51	0.666
Aulostomidae	*Aulostomus chinensis*	0.1 ± 0.05	0.04 ± 0.04	0 ± 0	3.72	0.086
Balistidae	*Abalistes stellatus*	**0.01 ± 0.01**	**0.08 ± 0.05**	**0.45 ± 0.13**	**14.93**	**<0.001**
Balistidae	*Balistapus undulatus*	**2.24 ± 0.37**	**0.81 ± 0.16**	**0.28 ± 0.07**	**33.84**	**<0.001**
Balistidae	*Balistoides conspicillum*	0.04 ± 0.03	0.08 ± 0.05	0.08 ± 0.04	0.55	0.590
Balistidae	*Balistoides viridescens*	0.03 ± 0.02	0.08 ± 0.04	0.03 ± 0.02	0.00	1.000
Balistidae	*Canthidermis maculata*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Balistidae	*Melichthys niger*	0 ± 0	1.01 ± 1.01	0 ± 0	0.00	1.000
Balistidae	*Melichthys vidua*	**0.31 ± 0.1**	**0.2 ± 0.07**	**0.04 ± 0.03**	**6.70**	**0.011**
Balistidae	*Odonus niger*	3.65 ± 1.3	7.51 ± 2.39	1 ± 0.43	1.27	0.279
Balistidae	*Pseudobalistes flavimarginatus*	0.04 ± 0.03	0.08 ± 0.04	0.06 ± 0.05	0.15	0.851
Balistidae	*Pseudobalistes fuscus*	0.21 ± 0.21	0.48 ± 0.46	0.07 ± 0.04	0.11	0.883
Balistidae	*Rhinecanthus rectangulus*	0.01 ± 0.01	0.08 ± 0.05	0 ± 0	0.11	0.834
Balistidae	*Rhinecanthus verrucosus*	0.08 ± 0.07	0.01 ± 0.01	0.02 ± 0.02	1.23	0.357
Balistidae	*Sufflamen bursa*	0.04 ± 0.03	0.41 ± 0.08	0.25 ± 0.09	3.71	0.058
Balistidae	*Sufflamen chrysopterum*	0.27 ± 0.1	0.28 ± 0.09	0.12 ± 0.09	1.34	0.258
Balistidae	*Sufflamen fraenatum*	**0 ± 0**	**0.14 ± 0.06**	**0.22 ± 0.07**	**9.04**	**0.003**
Balistidae	*Xanthichthys auromarginatus*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.661
Belonidae	*Tylosurus crocodilus*	0.04 ± 0.03	0 ± 0	0 ± 0	3.13	0.218
Blenniidae	*Aspidontus taeniatus*	0.06 ± 0.04	0.02 ± 0.02	0.01 ± 0.01	1.05	0.385
Blenniidae	*Ecsenius midas*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Blenniidae	*Meiacanthus atrodorsalis*	0 ± 0	0.03 ± 0.02	0 ± 0	0.00	1.000
Blenniidae	*Meiacanthus crinitus*	0 ± 0	0.02 ± 0.02	0.08 ± 0.08	1.43	0.440
Blenniidae	*Meiacanthus grammistes*	0.01 ± 0.01	0.12 ± 0.06	0.03 ± 0.02	0.14	0.727

Blenniidae	*Plagiotremus rhinorhynchos*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Caesionidae	*Caesio caerulaurea*	2.83 ± 2.15	0.08 ± 0.06	0 ± 0	2.60	0.061
Caesionidae	*Caesio cuning*	3.89 ± 2.28	5.27 ± 2.4	0 ± 0	2.04	0.174
Caesionidae	*Caesio lunaris*	**4.85 ± 1.92**	**1.52 ± 1.01**	**0.17 ± 0.13**	**7.03**	**0.008**
Caesionidae	*Caesio teres*	2.63 ± 1.51	1.94 ± 1.77	0.01 ± 0.01	1.91	0.186
Caesionidae	*Gymnocaesio gymnoptera*	0.1 ± 0.1	0.04 ± 0.04	0 ± 0	1.29	0.439
Caesionidae	*Pterocaesio digramma*	0.14 ± 0.14	0.68 ± 0.39	0 ± 0	0.16	0.713
Caesionidae	*Pterocaesio marri*	2.36 ± 1.79	3.09 ± 1.82	1.75 ± 1.49	0.06	0.805
Caesionidae	*Pterocaesio pisang*	0.02 ± 0.02	0.97 ± 0.69	0 ± 0	0.00	0.989
Caesionidae	*Pterocaesio randalli*	0 ± 0	0 ± 0	0.04 ± 0.04	1.51	0.665
Caesionidae	*Pterocaesio tile*	**3.11 ± 1.36**	**0.31 ± 0.31**	**0.04 ± 0.04**	**7.19**	**0.004**
Carangidae	*Alectis ciliaris*	0.05 ± 0.03	0 ± 0	0.01 ± 0.01	1.33	0.364
Carangidae	*Alepes kleinii*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Carangidae	*Atule mate*	0.01 ± 0.01	0.08 ± 0.05	0 ± 0	0.11	0.840
Carangidae	*Carangoides bajad*	0.33 ± 0.17	0.42 ± 0.28	0.41 ± 0.35	0.05	0.841
Carangidae	*Carangoides ferdau*	**0.17 ± 0.08**	**0.17 ± 0.07**	**0.81 ± 0.3**	**5.98**	**0.008**
Carangidae	*Carangoides fulvoguttatus*	0.08 ± 0.06	0.04 ± 0.04	0.04 ± 0.03	0.30	0.660
Carangidae	*Carangoides gymnostethus*	0.06 ± 0.03	0.04 ± 0.03	0 ± 0	2.94	0.158
Carangidae	*Carangoides oblongus*	0.05 ± 0.03	0.06 ± 0.05	0.14 ± 0.1	0.90	0.409
Carangidae	*Carangoides plagiotaenia*	0.06 ± 0.04	0.21 ± 0.12	0.02 ± 0.02	0.10	0.794
Carangidae	*Caranx ignobilis*	0.38 ± 0.29	0.1 ± 0.07	0.04 ± 0.04	1.84	0.221
Carangidae	*Caranx melampygus*	1.15 ± 0.4	1.25 ± 0.48	0.29 ± 0.08	2.77	0.101
Carangidae	*Caranx papuensis*	0.1 ± 0.06	0.02 ± 0.02	0.02 ± 0.02	1.92	0.253
Carangidae	*Caranx sexfasciatus*	0.06 ± 0.05	0 ± 0	2.6 ± 2.58	1.46	0.592
Carangidae	*Elagatis bipinnulata*	0 ± 0	0 ± 0	0.06 ± 0.05	2.80	0.212
Carangidae	*Gnathanodon speciosus*	0.03 ± 0.02	0.25 ± 0.23	0.03 ± 0.03	0.00	0.979
Carangidae	*Scomberoides commersonnianus*	0 ± 0	0.13 ± 0.09	0 ± 0	0.00	1.000
Carangidae	*Scomberoides lysan*	**0.12 ± 0.06**	**0 ± 0**	**0 ± 0**	**5.62**	**0.024**
Carangidae	*Scomberoides tol*	0.4 ± 0.4	0 ± 0	0 ± 0	1.51	0.665
Carangidae	*Selar boops*	0 ± 0	0 ± 0	0.23 ± 0.23	1.51	0.668
Carangidae	*Trachinotus blochii*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.663
Carcharhinidae	*Carcharhinus albimarginatus*	0 ± 0	0 ± 0	0.01 ± 0.01	1.51	0.666
Carcharhinidae	*Carcharhinus amblyrhynchos*	0.02 ± 0.02	0.02 ± 0.02	0.02 ± 0.02	0.00	1.000
Carcharhinidae	*Carcharhinus melanopterus*	**0.5 ± 0.1**	**0.28 ± 0.06**	**0.18 ± 0.07**	**8.95**	**0.004**
Carcharhinidae	*Carcharhinus obscurus*	0 ± 0	0.03 ± 0.02	0 ± 0	0.00	1.000
Carcharhinidae	*Triaenodon obesus*	0 ± 0	0.11 ± 0.04	0.04 ± 0.03	0.91	0.385
Chaetodontidae	*Chaetodon auriga*	0.06 ± 0.05	0.08 ± 0.04	0.03 ± 0.03	0.44	0.560
Chaetodontidae	*Chaetodon baronessa*	**0.35 ± 0.1**	**0.01 ± 0.01**	**0 ± 0**	**16.78**	**<0.001**
Chaetodontidae	*Chaetodon bennetti*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000

Chaetodontidae	*Chaetodon citrinellus*	**0.18 ± 0.08**	**0 ± 0**	**0 ± 0**	**7.33**	**0.005**
Chaetodontidae	*Chaetodon ephippium*	**0.26 ± 0.1**	**0.1 ± 0.06**	**0 ± 0**	**7.38**	**0.004**
Chaetodontidae	*Chaetodon kleinii*	1.03 ± 0.25	0.8 ± 0.17	0.51 ± 0.18	3.29	0.074
Chaetodontidae	*Chaetodon lineolatus*	**0.28 ± 0.09**	**0.07 ± 0.05**	**0 ± 0**	**11.08**	**0.001**
Chaetodontidae	*Chaetodon lunula*	**0.17 ± 0.06**	**0.05 ± 0.03**	**0 ± 0**	**9.42**	**0.003**
Chaetodontidae	*Chaetodon lunulatus*	**0.48 ± 0.11**	**0.02 ± 0.02**	**0 ± 0**	**25.64**	**<0.001**
Chaetodontidae	*Chaetodon melannotus*	0.03 ± 0.03	0.04 ± 0.03	0 ± 0	0.72	0.508
Chaetodontidae	*Chaetodon meyeri*	0.04 ± 0.04	0.04 ± 0.04	0 ± 0	0.76	0.669
Chaetodontidae	*Chaetodon ocellicaudus*	0.01 ± 0.01	0.09 ± 0.08	0 ± 0	0.04	0.811
Chaetodontidae	*Chaetodon octofasciatus*	0.09 ± 0.05	0.03 ± 0.02	0 ± 0	3.82	0.060
Chaetodontidae	*Chaetodon ornatissimus*	0.04 ± 0.04	0.08 ± 0.04	0 ± 0	0.84	0.426
Chaetodontidae	*Chaetodon oxycephalus*	0.11 ± 0.06	0.04 ± 0.03	0.04 ± 0.04	1.13	0.321
Chaetodontidae	*Chaetodon punctatofasciatus*	0.03 ± 0.03	0 ± 0	0 ± 0	1.51	0.664
Chaetodontidae	*Chaetodon rafflesii*	**0.5 ± 0.13**	**0 ± 0**	**0 ± 0**	**19.48**	**<0.001**
Chaetodontidae	*Chaetodon selene*	0 ± 0	0 ± 0	0.04 ± 0.04	1.51	0.665
Chaetodontidae	*Chaetodon semeion*	**0.16 ± 0.07**	**0 ± 0**	**0 ± 0**	**7.52**	**0.002**
Chaetodontidae	*Chaetodon speculum*	**0.22 ± 0.08**	**0.04 ± 0.03**	**0 ± 0**	**10.47**	**0.001**
Chaetodontidae	*Chaetodon trifascialis*	**0.32 ± 0.09**	**0 ± 0**	**0 ± 0**	**18.31**	**<0.001**
Chaetodontidae	*Chaetodon ulietensis*	0.1 ± 0.07	0.1 ± 0.06	0.01 ± 0.01	1.37	0.285
Chaetodontidae	*Chaetodon unimaculatus*	0.04 ± 0.04	0.02 ± 0.02	0 ± 0	1.22	0.434
Chaetodontidae	*Chaetodon vagabundus*	**0.63 ± 0.14**	**0.18 ± 0.09**	**0.1 ± 0.07**	**11.96**	**0.001**
Chaetodontidae	*Chelmon muelleri*	0 ± 0	0 ± 0	0.02 ± 0.02	1.51	0.672
Chaetodontidae	*Chelmon rostratus*	0.04 ± 0.04	0 ± 0	0 ± 0	1.51	0.660
Chaetodontidae	*Coradion altivelis*	0.01 ± 0.01	0.11 ± 0.05	0.08 ± 0.05	1.29	0.293
Chaetodontidae	*Coradion chrysozonus*	0.03 ± 0.02	0.27 ± 0.11	0.1 ± 0.06	0.49	0.538
Chaetodontidae	*Coradion melanopus*	0.02 ± 0.02	0 ± 0	0.02 ± 0.02	0.00	1.000
Chaetodontidae	*Forcipiger flavissimus*	0 ± 0	0.07 ± 0.04	0 ± 0	0.00	1.000
Chaetodontidae	*Forcipiger longirostris*	0.02 ± 0.02	0.17 ± 0.07	0 ± 0	0.10	0.800
Chaetodontidae	*Hemitaurichthys polylepis*	0 ± 0	0.17 ± 0.17	0 ± 0	0.00	1.000
Chaetodontidae	*Heniochus acuminatus*	0 ± 0	0.06 ± 0.06	0.02 ± 0.02	0.15	0.894
Chaetodontidae	*Heniochus chrysostomus*	0.03 ± 0.02	0.03 ± 0.03	0 ± 0	1.33	0.362
Chaetodontidae	*Heniochus monoceros*	0 ± 0	0.06 ± 0.05	0 ± 0	0.00	1.000
Chaetodontidae	*Heniochus singularius*	0.12 ± 0.06	0.04 ± 0.04	0 ± 0	3.92	0.061
Chaetodontidae	*Heniochus varius*	**0.23 ± 0.09**	**0.09 ± 0.06**	**0 ± 0**	**7.47**	**0.009**
Chaetodontidae	*Parachaetodon ocellatus*	0 ± 0	0.06 ± 0.04	0 ± 0	0.00	1.000
Chanidae	*Chanos chanos*	0.1 ± 0.1	0 ± 0	0 ± 0	1.51	0.662
Cirrhitidae	*Cirrhitichthys aprinus*	0 ± 0	0.02 ± 0.02	0.04 ± 0.04	1.22	0.448
Cirrhitidae	*Cirrhitichthys falco*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Cirrhitidae	*Paracirrhites forsteri*	**0.1 ± 0.04**	**0 ± 0**	**0 ± 0**	**8.82**	**0.006**

Dasyatididae	*Pastinachus sephen*	0 ± 0	0 ± 0	0.03 ± 0.02	3.01	0.218
Dasyatididae	*Taeniura lymma*	0 ± 0	0.03 ± 0.02	0 ± 0	0.00	1.000
Diodontidae	*Diodon hystrix*	0.03 ± 0.02	0.02 ± 0.02	0 ± 0	1.77	0.290
Echeneidae	*Echeneis naucrates*	0.11 ± 0.07	0.09 ± 0.04	0.02 ± 0.02	2.00	0.172
Ephippidae	*Platax batavianus*	0 ± 0	0.02 ± 0.02	0.01 ± 0.01	0.46	0.669
Ephippidae	*Platax boersii*	0.03 ± 0.03	0.24 ± 0.12	0.04 ± 0.03	0.02	0.915
Ephippidae	*Platax orbicularis*	0.11 ± 0.11	0.09 ± 0.07	0.43 ± 0.42	0.81	0.542
Ephippidae	*Platax pinnatus*	0.01 ± 0.01	0 ± 0	0 ± 0	1.51	0.663
Fistulariidae	*Fistularia commersonii*	0.1 ± 0.05	0.08 ± 0.04	0.06 ± 0.03	0.58	0.485
Gerreidae	*Gerres erythrourus*	0 ± 0	0 ± 0	0.02 ± 0.02	1.51	0.670
Ginglymostomatidae	*Nebrius ferrugineus*	0.04 ± 0.03	0.02 ± 0.02	0 ± 0	2.09	0.293
Gobiidae	*Amblyeleotris steinitzi*	0 ± 0	0.04 ± 0.04	0 ± 0	0.00	1.000
Gobiidae	*Valenciennea strigata*	0.02 ± 0.02	0.02 ± 0.02	0 ± 0	0.76	0.669
Gobiidae	*Valenciennea wardii*	0 ± 0	0 ± 0	0.02 ± 0.02	1.51	0.660
Haemulidae	*Diagramma pictum*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.659
Haemulidae	*Plectorhinchus albovittatus*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.665
Haemulidae	*Plectorhinchus gibbosus*	0 ± 0	0.04 ± 0.03	0 ± 0	0.00	1.000
Haemulidae	*Plectorhinchus lessonii*	0.04 ± 0.03	0 ± 0	0 ± 0	3.13	0.222
Haemulidae	*Plectorhinchus lineatus*	0.04 ± 0.04	0.08 ± 0.08	0.01 ± 0.01	0.13	0.892
Haemulidae	*Plectorhinchus polytaenia*	0.02 ± 0.02	0.09 ± 0.06	0.02 ± 0.02	0.00	1.000
Haemulidae	*Plectorhinchus schotaf*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Hemiscylliidae	*Chiloscyllium punctatum*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Holocentridae	*Myripristis berndti*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Holocentridae	*Myripristis botche*	0.07 ± 0.05	0.04 ± 0.04	0 ± 0	1.77	0.297
Holocentridae	*Sargocentron caudimaculatum*	0.06 ± 0.05	0.02 ± 0.02	0 ± 0	2.35	0.214
Holocentridae	*Sargocentron violaceum*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.661
Kyphosidae	*Kyphosus bigibbus*	0.46 ± 0.46	0 ± 0	0 ± 0	1.51	0.668
Kyphosidae	*Kyphosus cinerascens*	0.76 ± 0.73	0 ± 0	0 ± 0	1.66	0.070
Kyphosidae	*Kyphosus vaigiensis*	1.26 ± 1.19	0 ± 0	0 ± 0	1.69	0.219
Labridae	*Anampses caeruleopunctatus*	0.02 ± 0.02	0.06 ± 0.03	0 ± 0	0.39	0.761
Labridae	*Anampses geographicus*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.671
Labridae	*Anampses meleagrides*	0.09 ± 0.07	0.08 ± 0.07	0 ± 0	1.41	0.288
Labridae	*Anampses neoguinaicus*	0.06 ± 0.03	0.02 ± 0.02	0.04 ± 0.04	0.09	0.835
Labridae	*Bodianus axillaris*	0.06 ± 0.03	0.02 ± 0.02	0 ± 0	3.66	0.117
Labridae	*Bodianus bilunulatus*	0.02 ± 0.02	0.01 ± 0.01	0 ± 0	1.22	0.439
Labridae	*Bodianus bimaculatus*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Labridae	*Bodianus diana*	0.01 ± 0.01	0.13 ± 0.05	0 ± 0	0.08	0.844
Labridae	*Bodianus dictynna*	0.02 ± 0.02	0.16 ± 0.06	0 ± 0	0.14	0.765
Labridae	*Bodianus mesothorax*	0.08 ± 0.05	0.09 ± 0.04	0 ± 0	2.77	0.107

Labridae	*Cheilinus chlorourus*	0.02 ± 0.02	0.02 ± 0.02	0 ± 0	0.76	0.671
Labridae	*Cheilinus fasciatus*	0.15 ± 0.05	0.21 ± 0.08	0 ± 0	3.88	0.058
Labridae	*Cheilinus trilobatus*	0.05 ± 0.03	0.06 ± 0.03	0 ± 0	2.05	0.190
Labridae	*Cheilinus undulatus*	0.1 ± 0.07	0.04 ± 0.03	0.06 ± 0.03	0.40	0.672
Labridae	*Cheilio inermis*	0.04 ± 0.03	0.09 ± 0.04	0.02 ± 0.02	0.23	0.697
Labridae	*Choerodon anchorago*	**0.17 ± 0.05**	**0.01 ± 0.01**	**0 ± 0**	**12.61**	**0.001**
Labridae	*Choerodon jordani*	**0 ± 0**	**0 ± 0**	**0.24 ± 0.15**	**4.07**	**0.002**
Labridae	*Choerodon monostigma*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Labridae	*Choerodon schoenleinii*	0.03 ± 0.02	0 ± 0	0 ± 0	3.01	0.216
Labridae	*Choerodon zamboangae*	0 ± 0	0.04 ± 0.04	0.11 ± 0.07	2.87	0.117
Labridae	*Choerodon zosterophorus*	0 ± 0	0.31 ± 0.13	0.04 ± 0.03	0.13	0.762
Labridae	*Cirrhilabrus beauperryi*	0 ± 0	0.33 ± 0.31	0 ± 0	0.00	1.000
Labridae	*Cirrhilabrus condei*	0 ± 0	0 ± 0	0.29 ± 0.29	1.51	0.665
Labridae	*Cirrhilabrus cyanopleura*	0.1 ± 0.1	0.29 ± 0.29	0 ± 0	0.17	0.895
Labridae	*Cirrhilabrus exquisitus*	0 ± 0	0.1 ± 0.1	0 ± 0	0.00	1.000
Labridae	*Cirrhilabrus flavidorsalis*	0.96 ± 0.84	0 ± 0	0 ± 0	1.97	0.220
Labridae	*Cirrhilabrus lubbocki*	0.42 ± 0.42	0 ± 0	0 ± 0	1.51	0.666
Labridae	*Cirrhilabrus tonozukai*	0.15 ± 0.15	0 ± 0	0 ± 0	1.51	0.671
Labridae	*Coris batuensis*	0.01 ± 0.01	0.07 ± 0.03	0 ± 0	0.21	0.751
Labridae	*Coris dorsomacula*	0 ± 0	0 ± 0	0.02 ± 0.02	1.51	0.662
Labridae	*Coris gaimard*	0.02 ± 0.02	0.04 ± 0.03	0.01 ± 0.01	0.05	0.864
Labridae	*Coris pictoides*	0.68 ± 0.36	1.18 ± 0.64	0.69 ± 0.29	0.00	0.997
Labridae	*Diproctacanthus xanthurus*	**0.92 ± 0.36**	**0.15 ± 0.08**	**0.05 ± 0.03**	**8.44**	**<0.001**
Labridae	*Epibulus brevis*	0.06 ± 0.05	0.02 ± 0.02	0 ± 0	2.35	0.226
Labridae	*Epibulus insidiator*	0.01 ± 0.01	0 ± 0	0 ± 0	1.51	0.665
Labridae	*Gomphosus varius*	0.2 ± 0.09	0.13 ± 0.09	0 ± 0	4.00	0.050
Labridae	*Halichoeres bicolor*	0.52 ± 0.38	0.1 ± 0.07	0.04 ± 0.03	2.32	0.094
Labridae	*Halichoeres biocellatus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Labridae	*Halichoeres chloropterus*	0.12 ± 0.05	0.04 ± 0.04	0.01 ± 0.01	3.50	0.075
Labridae	*Halichoeres chrysus*	0.02 ± 0.02	0.74 ± 0.29	0.03 ± 0.03	0.00	0.991
Labridae	*Halichoeres hortulanus*	**0.23 ± 0.06**	**0.09 ± 0.07**	**0 ± 0**	**9.89**	**0.001**
Labridae	*Halichoeres margaritaceus*	0.06 ± 0.06	0 ± 0	0 ± 0	1.51	0.667
Labridae	*Halichoeres marginatus*	0.04 ± 0.03	0 ± 0	0 ± 0	3.13	0.219
Labridae	*Halichoeres melanochir*	0.01 ± 0.01	0.02 ± 0.02	0.02 ± 0.02	0.07	0.885
Labridae	*Halichoeres melanurus*	**0.13 ± 0.05**	**0 ± 0**	**0 ± 0**	**10.56**	**0.002**
Labridae	*Halichoeres melasmapomus*	0.01 ± 0.01	0 ± 0	0 ± 0	1.51	0.669
Labridae	*Halichoeres miniatus*	0.04 ± 0.04	0 ± 0	0 ± 0	1.51	0.661
Labridae	*Halichoeres nigrescens*	0.01 ± 0.01	0.02 ± 0.02	0 ± 0	0.46	0.660
Labridae	*Halichoeres papilionaceus*	0.03 ± 0.02	0 ± 0	0 ± 0	3.01	0.223

Labridae	*Halichoeres podostigma*	**0.12 ± 0.07**	**0 ± 0**	**0 ± 0**	**4.53**	**0.021**
Labridae	*Halichoeres prosopeion*	0.06 ± 0.03	0.38 ± 0.12	0.03 ± 0.02	0.06	0.845
Labridae	*Halichoeres richmondi*	0.06 ± 0.05	0 ± 0	0 ± 0	2.80	0.222
Labridae	*Halichoeres scapularis*	0.16 ± 0.11	0 ± 0	0.08 ± 0.07	0.56	0.526
Labridae	*Halichoeres solorensis*	**0.76 ± 0.68**	**0.03 ± 0.02**	**0 ± 0**	**1.91**	**0.037**
Labridae	*Halichoeres trimaculatus*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.663
Labridae	*Hemigymnus fasciatus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Labridae	*Hemigymnus melapterus*	**0.46 ± 0.15**	**0.11 ± 0.05**	**0 ± 0**	**12.43**	**<0.001**
Labridae	*Hologymnosus annulatus*	**0.32 ± 0.16**	**0.07 ± 0.05**	**0 ± 0**	**5.77**	**0.003**
Labridae	*Hologymnosus doliatus*	0.02 ± 0.02	0.06 ± 0.05	0 ± 0	0.25	0.821
Labridae	*Iniistius aneitensis*	0.02 ± 0.02	0 ± 0	0.17 ± 0.09	3.79	0.075
Labridae	*Iniistius twistii*	0 ± 0	0 ± 0	0.02 ± 0.02	1.51	0.657
Labridae	*Labrichthys unilineatus*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.660
Labridae	*Labroides bicolor*	0.08 ± 0.04	0.03 ± 0.02	0.02 ± 0.02	2.07	0.171
Labridae	*Labroides dimidiatus*	**1 ± 0.26**	**0.71 ± 0.16**	**0.25 ± 0.09**	**8.57**	**0.002**
Labridae	*Labroides pectoralis*	0.06 ± 0.05	0.01 ± 0.01	0 ± 0	2.59	0.144
Labridae	*Labropsis manabei*	0.04 ± 0.04	0 ± 0	0.02 ± 0.02	0.30	0.895
Labridae	*Leptojulis cyanopleura*	0.18 ± 0.12	0 ± 0	0 ± 0	3.67	0.067
Labridae	*Macropharyngodon negrosensis*	0.02 ± 0.02	0.15 ± 0.11	0 ± 0	0.05	0.911
Labridae	*Novaculichthys taeniourus*	0 ± 0	0.06 ± 0.04	0 ± 0	0.00	1.000
Labridae	*Oxycheilinus arenatus*	0.03 ± 0.02	0 ± 0	0 ± 0	3.01	0.211
Labridae	*Oxycheilinus celebicus*	0.04 ± 0.04	0.15 ± 0.13	0.02 ± 0.02	0.04	0.940
Labridae	*Oxycheilinus digramma*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.673
Labridae	*Oxycheilinus orientalis*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Labridae	*Oxycheilinus unifasciatus*	0.06 ± 0.03	0.17 ± 0.11	0.01 ± 0.01	0.27	0.705
Labridae	*Paracheilinus filamentosus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Labridae	*Paracheilinus walton*	0.04 ± 0.04	0 ± 0	0 ± 0	1.51	0.671
Labridae	*Pseudocoris heteroptera*	1.03 ± 0.83	0.17 ± 0.08	0.06 ± 0.04	2.07	0.075
Labridae	*Pseudocoris yamashiroi*	0.65 ± 0.48	0.31 ± 0.27	0 ± 0	2.11	0.193
Labridae	*Pseudodax moluccanus*	0.02 ± 0.02	0.04 ± 0.03	0 ± 0	0.51	0.733
Labridae	*Pseudojuloides splendens*	0.02 ± 0.02	0 ± 0	0.01 ± 0.01	0.12	0.891
Labridae	*Stethojulis bandanensis*	0 ± 0	0.22 ± 0.15	0.06 ± 0.06	0.21	0.740
Labridae	*Stethojulis interrupta*	0.06 ± 0.06	0 ± 0	0 ± 0	1.51	0.659
Labridae	*Stethojulis strigiventer*	0.02 ± 0.02	0.26 ± 0.23	0 ± 0	0.01	0.885
Labridae	*Thalassoma amblycephalum*	0.76 ± 0.34	0.22 ± 0.17	0.35 ± 0.31	1.04	0.328
Labridae	*Thalassoma hardwicke*	**1.94 ± 0.36**	**0.01 ± 0.01**	**0 ± 0**	**37.24**	**<0.001**
Labridae	*Thalassoma jansenii*	0.08 ± 0.05	0.04 ± 0.04	0 ± 0	2.55	0.186
Labridae	*Thalassoma lunare*	**2.83 ± 0.6**	**0.32 ± 0.12**	**0 ± 0**	**29.70**	**<0.001**
Lethrinidae	*Gnathodentex aureolineatus*	0.15 ± 0.15	0 ± 0	0.1 ± 0.07	0.12	0.823

Lethrinidae	*Gymnocranius elongatus*	0 ± 0	0 ± 0	0.02 ± 0.02	1.51	0.670
Lethrinidae	*Gymnocranius grandoculis*	0.02 ± 0.02	0.13 ± 0.09	0.17 ± 0.09	2.31	0.138
Lethrinidae	*Lethrinus amboinensis*	**0.01 ± 0.01**	**0.16 ± 0.08**	**0.53 ± 0.2**	**8.54**	**0.003**
Lethrinidae	*Lethrinus atkinsoni*	0.01 ± 0.01	0.05 ± 0.03	0.14 ± 0.11	1.90	0.196
Lethrinidae	*Lethrinus erythracanthus*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Lethrinidae	*Lethrinus erythropterus*	0.02 ± 0.02	0 ± 0	0.02 ± 0.02	0.00	1.000
Lethrinidae	*Lethrinus harak*	0.42 ± 0.14	0.11 ± 0.07	0.47 ± 0.35	0.02	0.913
Lethrinidae	*Lethrinus laticaudis*	0 ± 0	0 ± 0	0.08 ± 0.08	1.51	0.667
Lethrinidae	*Lethrinus lentjan*	**0 ± 0**	**0 ± 0**	**0.15 ± 0.08**	**5.37**	**0.020**
Lethrinidae	*Lethrinus microdon*	0 ± 0	0.18 ± 0.05	0.17 ± 0.09	3.68	0.058
Lethrinidae	*Lethrinus nebulosus*	0 ± 0	0.14 ± 0.06	0.16 ± 0.09	3.47	0.060
Lethrinidae	*Lethrinus obsoletus*	**0.02 ± 0.02**	**0.28 ± 0.12**	**0.35 ± 0.14**	**4.47**	**0.036**
Lethrinidae	*Lethrinus olivaceus*	0.06 ± 0.03	0.17 ± 0.08	0.1 ± 0.05	0.18	0.708
Lethrinidae	*Lethrinus ornatus*	0 ± 0	0.03 ± 0.02	0.17 ± 0.15	1.91	0.162
Lethrinidae	*Lethrinus ravus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Lethrinidae	*Lethrinus rubrioperculatus*	0 ± 0	0 ± 0	0.11 ± 0.11	1.51	0.657
Lethrinidae	*Lethrinus semicinctus*	0.12 ± 0.07	0.77 ± 0.46	0.88 ± 0.3	2.87	0.091
Lethrinidae	*Lethrinus variegatus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Lethrinidae	*Monotaxis grandoculis*	0.04 ± 0.03	0.22 ± 0.08	0.05 ± 0.03	0.01	0.953
Lethrinidae	*Monotaxis heterodon*	0.06 ± 0.05	0.09 ± 0.08	0 ± 0	0.65	0.510
Lutjanidae	*Aphareus furca*	0 ± 0	0.02 ± 0.02	0.1 ± 0.1	1.46	0.444
Lutjanidae	*Aprion virescens*	0.03 ± 0.02	0.18 ± 0.06	0.13 ± 0.05	1.96	0.173
Lutjanidae	*Lutjanus argentimaculatus*	0 ± 0	0 ± 0	0.02 ± 0.02	1.51	0.668
Lutjanidae	*Lutjanus biguttatus*	0.43 ± 0.43	0.04 ± 0.03	0.04 ± 0.04	1.22	0.551
Lutjanidae	*Lutjanus bohar*	**0.72 ± 0.18**	**0.27 ± 0.09**	**0.06 ± 0.03**	**15.58**	**<0.001**
Lutjanidae	*Lutjanus carponotatus*	**0.18 ± 0.09**	**0.01 ± 0.01**	**0 ± 0**	**5.97**	**0.002**
Lutjanidae	*Lutjanus decussatus*	**0.34 ± 0.11**	**0.08 ± 0.04**	**0 ± 0**	**13.00**	**<0.001**
Lutjanidae	*Lutjanus ehrenbergii*	0.08 ± 0.05	0 ± 0	0 ± 0	4.29	0.066
Lutjanidae	*Lutjanus fulviflamma*	0.03 ± 0.02	0 ± 0	0 ± 0	3.01	0.220
Lutjanidae	*Lutjanus fulvus*	**0.2 ± 0.08**	**0 ± 0**	**0 ± 0**	**8.30**	**0.001**
Lutjanidae	*Lutjanus gibbus*	0.27 ± 0.27	0.41 ± 0.25	0 ± 0	0.82	0.409
Lutjanidae	*Lutjanus lemniscatus*	0.03 ± 0.02	0.01 ± 0.01	0.02 ± 0.02	0.24	0.733
Lutjanidae	*Lutjanus lunulatus*	0.06 ± 0.06	0 ± 0	0 ± 0	1.51	0.669
Lutjanidae	*Lutjanus lutjanus*	0.03 ± 0.03	0.01 ± 0.01	0 ± 0	1.33	0.446
Lutjanidae	*Lutjanus maxweberi*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.668
Lutjanidae	*Lutjanus monostigma*	0.08 ± 0.04	0.11 ± 0.05	0 ± 0	2.12	0.160
Lutjanidae	*Lutjanus rivulatus*	**0.15 ± 0.07**	**0.04 ± 0.03**	**0 ± 0**	**6.24**	**0.007**
Lutjanidae	*Lutjanus russellii*	0.15 ± 0.06	0 ± 0	0.04 ± 0.04	3.15	0.089
Lutjanidae	*Lutjanus semicinctus*	**0.88 ± 0.16**	**0.2 ± 0.06**	**0 ± 0**	**37.61**	**<0.001**

Lutjanidae	*Lutjanus vitta*	0.03 ± 0.02	0.27 ± 0.19	0.23 ± 0.1	1.19	0.317
Lutjanidae	*Macolor macularis*	0.23 ± 0.13	0.5 ± 0.19	0 ± 0	1.38	0.265
Lutjanidae	*Macolor niger*	0.13 ± 0.06	0.06 ± 0.05	0 ± 0	4.00	0.069
Lutjanidae	*Pinjalo lewisi*	0 ± 0	1 ± 1	0 ± 0	0.00	1.000
Malacanthidae	*Hoplolatilus chlupatyi*	0 ± 0	0.02 ± 0.02	0.01 ± 0.01	0.46	0.672
Malacanthidae	*Hoplolatilus cuniculus*	0 ± 0	0 ± 0	0.06 ± 0.04	2.47	0.218
Malacanthidae	*Hoplolatilus purpureus*	**0 ± 0**	**0 ± 0**	**0.38 ± 0.15**	**9.15**	**0.002**
Malacanthidae	*Malacanthus brevirostris*	**0 ± 0**	**0.01 ± 0.01**	**0.07 ± 0.04**	**3.45**	**0.046**
Malacanthidae	*Malacanthus latovittatus*	0.04 ± 0.04	0.01 ± 0.01	0.03 ± 0.02	0.03	0.964
Microdesmidae	*Gunnellichthys monostigma*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Microdesmidae	*Gunnellichthys pleurotaenia*	0.06 ± 0.03	0 ± 0	0.02 ± 0.02	1.27	0.357
Monacanthidae	*Aluterus scriptus*	0.04 ± 0.03	0 ± 0	0.08 ± 0.05	0.79	0.534
Monacanthidae	*Amanses scopas*	0.06 ± 0.03	0.05 ± 0.03	0.02 ± 0.02	1.15	0.312
Monacanthidae	*Cantherhines dumerilii*	0.15 ± 0.09	0.04 ± 0.03	0.05 ± 0.03	1.52	0.270
Monacanthidae	*Cantherhines fronticinctus*	0.04 ± 0.03	0 ± 0	0 ± 0	3.13	0.212
Monacanthidae	*Cantherhines pardalis*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Monacanthidae	*Paraluteres prionurus*	0 ± 0	0.13 ± 0.09	0 ± 0	0.00	1.000
Monacanthidae	*Paramonacanthus japonicus*	0 ± 0	0 ± 0	0.06 ± 0.04	2.47	0.218
Monacanthidae	*Pervagor janthinosoma*	0.04 ± 0.04	0 ± 0	0 ± 0	1.51	0.674
Monacanthidae	*Pseudomonacanthus macrurus*	0 ± 0	0 ± 0	0.02 ± 0.02	1.51	0.662
Monodactylidae	*Monodactylus argenteus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Mugilidae	*Crenimugil crenilabis*	0.61 ± 0.61	0 ± 0	0 ± 0	1.51	0.661
Mullidae	*Mulloidichthys flavolineatus*	0.04 ± 0.04	0.02 ± 0.02	0.07 ± 0.05	0.25	0.655
Mullidae	*Mulloidichthys vanicolensis*	0 ± 0	0 ± 0	0.02 ± 0.02	1.51	0.672
Mullidae	*Parupeneus barberinoides*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Mullidae	*Parupeneus barberinus*	0.53 ± 0.11	0.69 ± 0.17	0.26 ± 0.1	2.10	0.161
Mullidae	*Parupeneus chrysopleuron*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Mullidae	*Parupeneus crassilabris*	**0.33 ± 0.08**	**0.06 ± 0.05**	**0 ± 0**	**18.08**	**<0.001**
Mullidae	*Parupeneus cyclostomus*	0.13 ± 0.05	0.47 ± 0.15	0.29 ± 0.12	0.90	0.355
Mullidae	*Parupeneus heptacanthus*	**0 ± 0**	**0.1 ± 0.07**	**0.38 ± 0.19**	**5.31**	**0.013**
Mullidae	*Parupeneus indicus*	0.09 ± 0.05	0.08 ± 0.06	0.17 ± 0.1	0.55	0.501
Mullidae	*Parupeneus multifasciatus*	0.99 ± 0.17	1.14 ± 0.19	0.55 ± 0.2	2.74	0.104
Mullidae	*Parupeneus pleurostigma*	0 ± 0	0.07 ± 0.06	0 ± 0	0.00	1.000
Mullidae	*Upeneus tragula*	0.03 ± 0.02	0.03 ± 0.02	0.17 ± 0.15	1.42	0.347
Muraenidae	*Echidna nebulosa*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.665
Muraenidae	*Gymnothorax chilospilus*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.662
Muraenidae	*Gymnothorax enigmaticus*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.673
Muraenidae	*Gymnothorax flavimarginatus*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.664
Muraenidae	*Gymnothorax javanicus*	**0.13 ± 0.05**	**0.05 ± 0.03**	**0 ± 0**	**6.71**	**0.010**

Muraenidae	*Gymnothorax undulatus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Muraenidae	*Gymnothorax zonipectis*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Nemipteridae	*Pentapodus aureofasciatus*	0.06 ± 0.03	1.43 ± 0.42	1.39 ± 0.69	4.02	0.050
Nemipteridae	*Pentapodus caninus*	**0.01 ± 0.01**	**1.33 ± 0.73**	**2.55 ± 1.14**	**5.32**	**0.022**
Nemipteridae	*Pentapodus emeryii*	**0.17 ± 0.11**	**0.62 ± 0.14**	**0.97 ± 0.3**	**7.74**	**0.005**
Nemipteridae	*Pentapodus nagasakiensis*	**0 ± 0**	**0.4 ± 0.19**	**2.5 ± 1.28**	**5.59**	**0.002**
Nemipteridae	*Pentapodus numberii*	0 ± 0	0.09 ± 0.05	0.66 ± 0.65	1.58	0.266
Nemipteridae	*Pentapodus paradiseus*	**0.04 ± 0.04**	**0.08 ± 0.08**	**1.62 ± 0.76**	**6.30**	**<0.001**
Nemipteridae	*Pentapodus porosus*	0.03 ± 0.02	0.57 ± 0.25	0.46 ± 0.46	0.99	0.367
Nemipteridae	*Pentapodus trivittatus*	**0.82 ± 0.22**	**0.16 ± 0.09**	**0.06 ± 0.05**	**14.29**	**<0.001**
Nemipteridae	*Scolopsis affinis*	0.56 ± 0.42	0.61 ± 0.25	1.07 ± 0.25	1.34	0.267
Nemipteridae	*Scolopsis bilineata*	**0.53 ± 0.1**	**0.32 ± 0.1**	**0.04 ± 0.04**	**17.04**	**<0.001**
Nemipteridae	*Scolopsis ciliata*	0.15 ± 0.08	0.12 ± 0.07	0.12 ± 0.07	0.11	0.765
Nemipteridae	*Scolopsis lineata*	0.33 ± 0.27	0 ± 0	0 ± 0	2.26	0.065
Nemipteridae	*Scolopsis margaritifera*	0.19 ± 0.07	0.57 ± 0.36	0.19 ± 0.12	0.00	1.000
Nemipteridae	*Scolopsis monogramma*	0.04 ± 0.04	0.44 ± 0.34	0.03 ± 0.02	0.00	0.997
Nemipteridae	*Scolopsis temporalis*	0.21 ± 0.09	0.68 ± 0.43	0.15 ± 0.08	0.02	0.885
Nemipteridae	*Scolopsis trilineata*	0.18 ± 0.11	0.02 ± 0.02	0.04 ± 0.04	2.16	0.162
Nemipteridae	*Scolopsis vosmeri*	**0 ± 0**	**0 ± 0**	**0.4 ± 0.2**	**6.28**	**0.002**
Nemipteridae	*Scolopsis xenochroa*	0.17 ± 0.07	0.1 ± 0.07	0.17 ± 0.11	0.00	1.000
Ophichthidae	*Myrichthys colubrinus*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.671
Ostraciidae	*Lactoria cornuta*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Ostraciidae	*Ostracion cubicus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Ostraciidae	*Ostracion meleagris*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.674
Pempheridae	*Parapriacanthus ransonneti*	0 ± 0	2.75 ± 2.75	0 ± 0	0.00	1.000
Pholidichthyidae	*Pholidichthys leucotaenia*	0 ± 0	0 ± 0	0.46 ± 0.46	1.51	0.662
Pinguipedidae	*Parapercis clathrata*	0.03 ± 0.02	0 ± 0	0.03 ± 0.03	0.05	0.963
Pinguipedidae	*Parapercis cylindrica*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Pinguipedidae	*Parapercis hexophtalma*	0.07 ± 0.03	0.02 ± 0.02	0.01 ± 0.01	2.70	0.130
Pinguipedidae	*Parapercis lineopunctata*	0.02 ± 0.02	0.04 ± 0.03	0.02 ± 0.02	0.00	1.000
Pinguipedidae	*Parapercis snyderi*	0 ± 0	0 ± 0	0.13 ± 0.11	2.12	0.210
Pinguipedidae	*Parapercis tetracantha*	0.03 ± 0.03	0.1 ± 0.06	0.09 ± 0.05	0.83	0.407
Pinguipedidae	*Parapercis xanthozona*	0 ± 0	0.07 ± 0.04	0.04 ± 0.04	0.86	0.423
Plotosidae	*Plotosus lineatus*	0 ± 0	9.5 ± 9.5	0 ± 0	0.00	1.000
Pomacanthidae	*Apolemichthys trimaculatus*	0.07 ± 0.04	0.08 ± 0.04	0.14 ± 0.06	1.06	0.329
Pomacanthidae	*Centropyge bicolor*	0.06 ± 0.04	0.34 ± 0.11	0.02 ± 0.02	0.17	0.722
Pomacanthidae	*Centropyge bispinosa*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Pomacanthidae	*Centropyge fisheri*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Pomacanthidae	*Centropyge nox*	0.07 ± 0.04	0 ± 0	0 ± 0	4.78	0.070

Pomacanthidae	*Centropyge tibicen*	0.08 ± 0.05	0.08 ± 0.07	0 ± 0	1.58	0.304
Pomacanthidae	*Centropyge vrolikii*	0.03 ± 0.02	0 ± 0	0 ± 0	3.01	0.210
Pomacanthidae	*Chaetodontoplus dimidiatus*	**0 ± 0**	**0.1 ± 0.05**	**0.27 ± 0.12**	**6.45**	**0.008**
Pomacanthidae	*Chaetodontoplus mesoleucus*	0.18 ± 0.08	0.25 ± 0.09	0.02 ± 0.02	2.65	0.119
Pomacanthidae	*Chaetodontoplus poliourus*	0.06 ± 0.05	0 ± 0	0 ± 0	2.80	0.213
Pomacanthidae	*Genicanthus lamarck*	0 ± 0	0.44 ± 0.16	0.06 ± 0.06	0.18	0.740
Pomacanthidae	*Genicanthus melanospilos*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Pomacanthidae	*Paracentropyge multifasciata*	0.02 ± 0.02	0.02 ± 0.02	0.01 ± 0.01	0.07	0.893
Pomacanthidae	*Pomacanthus imperator*	0.12 ± 0.06	0.18 ± 0.06	0.07 ± 0.03	0.43	0.552
Pomacanthidae	*Pomacanthus navarchus*	0.03 ± 0.02	0.01 ± 0.01	0 ± 0	2.58	0.143
Pomacanthidae	*Pomacanthus semicirculatus*	0.01 ± 0.01	0.04 ± 0.04	0 ± 0	0.15	0.891
Pomacanthidae	*Pomacanthus sexstriatus*	0.07 ± 0.05	0.19 ± 0.1	0.07 ± 0.05	0.00	1.000
Pomacanthidae	*Pomacanthus xanthometopon*	0.01 ± 0.01	0.02 ± 0.02	0 ± 0	0.46	0.665
Pomacanthidae	*Pygoplites diacanthus*	**0.18 ± 0.07**	**0.21 ± 0.07**	**0 ± 0**	**5.03**	**0.030**
Pomacentridae	*Abudefduf bengalensis*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.669
Pomacentridae	*Abudefduf sexfasciatus*	0.17 ± 0.17	0 ± 0	0 ± 0	1.51	0.662
Pomacentridae	*Abudefduf vaigiensis*	0.72 ± 0.71	0 ± 0	0 ± 0	1.57	0.223
Pomacentridae	*Acanthochromis polyacanthus*	0.4 ± 0.14	0.76 ± 0.75	0 ± 0	0.42	0.672
Pomacentridae	*Amblyglyphidodon aureus*	0.02 ± 0.02	0.11 ± 0.08	0 ± 0	0.09	0.892
Pomacentridae	*Amblyglyphidodon batunai*	0.1 ± 0.1	0.03 ± 0.02	0 ± 0	1.44	0.371
Pomacentridae	*Amblyglyphidodon curacao*	**0.37 ± 0.18**	**0.02 ± 0.02**	**0 ± 0**	**6.01**	**0.007**
Pomacentridae	*Amblyglyphidodon leucogaster*	0.33 ± 0.16	0.19 ± 0.15	0 ± 0	3.30	0.079
Pomacentridae	*Amblyglyphidodon ternatensis*	**0.35 ± 0.18**	**0 ± 0**	**0 ± 0**	**5.46**	**0.021**
Pomacentridae	*Amblypomacentrus breviceps*	0.04 ± 0.03	0 ± 0	0 ± 0	3.13	0.214
Pomacentridae	*Amblypomacentrus clarus*	1.17 ± 1.15	0.02 ± 0.02	0.02 ± 0.02	1.51	0.409
Pomacentridae	*Cheiloprion labiatus*	0 ± 0	0.04 ± 0.04	0 ± 0	0.00	1.000
Pomacentridae	*Chromis alpha*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Pomacentridae	*Chromis amboinensis*	**1.76 ± 1.38**	**0.08 ± 0.05**	**0 ± 0**	**2.47**	**0.010**
Pomacentridae	*Chromis analis*	0.44 ± 0.26	0 ± 0	0 ± 0	4.08	0.065
Pomacentridae	*Chromis atripectoralis*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.666
Pomacentridae	*Chromis caudalis*	0.1 ± 0.07	0.01 ± 0.01	0.19 ± 0.11	0.71	0.436
Pomacentridae	*Chromis cinerascens*	0.06 ± 0.06	0 ± 0	0 ± 0	1.51	0.664
Pomacentridae	*Chromis fumea*	0.44 ± 0.35	0.13 ± 0.06	0 ± 0	2.25	0.088
Pomacentridae	*Chromis margaritifer*	**0.87 ± 0.25**	**0.08 ± 0.05**	**0.01 ± 0.01**	**16.02**	**<0.001**
Pomacentridae	*Chromis retrofasciata*	0 ± 0	0.06 ± 0.06	0 ± 0	0.00	1.000
Pomacentridae	*Chromis scotochiloptera*	**0.41 ± 0.23**	**0.15 ± 0.1**	**0 ± 0**	**3.96**	**0.039**
Pomacentridae	*Chromis ternatensis*	**3.39 ± 1.87**	**0.05 ± 0.03**	**0 ± 0**	**4.90**	**<0.001**
Pomacentridae	*Chromis viridis*	0.75 ± 0.57	0 ± 0	0 ± 0	2.63	0.069
Pomacentridae	*Chromis weberi*	**0.89 ± 0.41**	**0.22 ± 0.17**	**0 ± 0**	**6.10**	**0.006**

Pomacentridae	*Chromis xanthochira*	0.1 ± 0.05	0.04 ± 0.04	0 ± 0	3.64	0.077
Pomacentridae	*Chromis xanthura*	**0.38 ± 0.2**	**0.06 ± 0.03**	**0 ± 0**	**5.38**	**0.012**
Pomacentridae	*Chrysiptera bleekeri*	0 ± 0	0.33 ± 0.27	0.1 ± 0.07	0.18	0.739
Pomacentridae	*Chrysiptera cyanea*	**1.41 ± 0.77**	**0.01 ± 0.01**	**0 ± 0**	**5.00**	**0.002**
Pomacentridae	*Chrysiptera flavipinnis*	0 ± 0	0.1 ± 0.1	0.06 ± 0.06	0.40	0.669
Pomacentridae	*Chrysiptera glauca*	0.06 ± 0.05	0 ± 0	0 ± 0	2.80	0.221
Pomacentridae	*Chrysiptera hemicyanea*	0.04 ± 0.04	0.13 ± 0.11	0 ± 0	0.20	0.817
Pomacentridae	*Chrysiptera parasema*	0.1 ± 0.09	0.07 ± 0.06	0.02 ± 0.02	0.91	0.390
Pomacentridae	*Chrysiptera pricei*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.674
Pomacentridae	*Chrysiptera rex*	0.14 ± 0.08	0.08 ± 0.08	0 ± 0	2.27	0.183
Pomacentridae	*Chrysiptera rollandi*	0.1 ± 0.07	0.19 ± 0.09	0 ± 0	1.15	0.318
Pomacentridae	*Chrysiptera springeri*	0.06 ± 0.06	0.02 ± 0.02	0 ± 0	1.37	0.436
Pomacentridae	*Chrysiptera talboti*	0.06 ± 0.03	0 ± 0	0 ± 0	4.72	0.067
Pomacentridae	*Dascyllus aruanus*	0.12 ± 0.1	0.04 ± 0.04	0 ± 0	1.85	0.289
Pomacentridae	*Dascyllus melanurus*	0.13 ± 0.11	0 ± 0	0 ± 0	2.12	0.216
Pomacentridae	*Dascyllus reticulatus*	**0.33 ± 0.15**	**0.04 ± 0.04**	**0 ± 0**	**6.47**	**0.010**
Pomacentridae	*Dascyllus trimaculatus*	0.29 ± 0.17	0.04 ± 0.04	0.04 ± 0.04	2.78	0.096
Pomacentridae	*Dischistodus chrysopoecilus*	0.04 ± 0.04	0 ± 0	0 ± 0	1.51	0.664
Pomacentridae	*Dischistodus fasciatus*	0.02 ± 0.02	0.06 ± 0.04	0 ± 0	0.28	0.743
Pomacentridae	*Dischistodus melanotus*	**0.31 ± 0.18**	**0.02 ± 0.02**	**0 ± 0**	**4.38**	**0.010**
Pomacentridae	*Dischistodus perspicillatus*	0.22 ± 0.14	0 ± 0	0 ± 0	3.51	0.073
Pomacentridae	*Dischistodus prosopotaenia*	0.08 ± 0.05	0 ± 0	0 ± 0	3.94	0.071
Pomacentridae	*Dischistodus pseudochrysopoecilus*	0.22 ± 0.16	0.08 ± 0.07	0 ± 0	2.55	0.139
Pomacentridae	*Hemiglyphidodon plagiometopon*	0.1 ± 0.05	0.13 ± 0.13	0 ± 0	0.89	0.465
Pomacentridae	*Lepidozygus tapeinosoma*	0 ± 0	0.04 ± 0.04	0 ± 0	0.00	1.000
Pomacentridae	*Neoglyphidodon crossi*	0.03 ± 0.02	0.02 ± 0.02	0 ± 0	1.46	0.293
Pomacentridae	*Neoglyphidodon melas*	**0.26 ± 0.1**	**0.04 ± 0.04**	**0 ± 0**	**8.96**	**0.002**
Pomacentridae	*Neoglyphidodon nigroris*	0.19 ± 0.09	0.23 ± 0.16	0 ± 0	1.64	0.225
Pomacentridae	*Neoglyphidodon oxyodon*	0.05 ± 0.03	0.1 ± 0.07	0 ± 0	0.53	0.539
Pomacentridae	*Neoglyphidodon thoracotaeniatus*	0 ± 0	0.07 ± 0.05	0 ± 0	0.00	1.000
Pomacentridae	*Neopomacentrus azysron*	0.06 ± 0.05	0 ± 0	0 ± 0	2.80	0.214
Pomacentridae	*Neopomacentrus cyanomos*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Pomacentridae	*Neopomacentrus filamentosus*	0.02 ± 0.02	0.1 ± 0.1	0 ± 0	0.06	0.895
Pomacentridae	*Plectroglyphidodon dickii*	**0.15 ± 0.07**	**0 ± 0**	**0 ± 0**	**6.32**	**0.006**
Pomacentridae	*Plectroglyphidodon lacrymatus*	0.02 ± 0.02	0.02 ± 0.02	0 ± 0	0.76	0.664
Pomacentridae	*Plectroglyphidodon leucozonus*	0.03 ± 0.03	0 ± 0	0 ± 0	1.51	0.671
Pomacentridae	*Pomacentrus adelus*	0 ± 0	0.08 ± 0.08	0 ± 0	0.00	1.000
Pomacentridae	*Pomacentrus amboinensis*	0.02 ± 0.02	0.89 ± 0.45	0.03 ± 0.03	0.00	0.992
Pomacentridae	*Pomacentrus auriventris*	1.96 ± 1.16	1.66 ± 0.89	0.25 ± 0.25	2.01	0.169

Pomacentridae	*Pomacentrus bankanensis*	**0.47 ± 0.16**	**0.04 ± 0.04**	**0 ± 0**	**10.99**	**<0.001**
Pomacentridae	*Pomacentrus brachialis*	**0.24 ± 0.1**	**0.08 ± 0.05**	**0 ± 0**	**6.42**	**0.014**
Pomacentridae	*Pomacentrus chrysurus*	**0.51 ± 0.24**	**0.06 ± 0.06**	**0 ± 0**	**6.46**	**0.009**
Pomacentridae	*Pomacentrus coelestis*	**0.49 ± 0.28**	**0 ± 0**	**0 ± 0**	**4.56**	**0.006**
Pomacentridae	*Pomacentrus cuneatus*	0.01 ± 0.01	0 ± 0	0 ± 0	1.51	0.668
Pomacentridae	*Pomacentrus lepidogenys*	0.08 ± 0.05	0.08 ± 0.08	0 ± 0	0.96	0.405
Pomacentridae	*Pomacentrus littoralis*	0.02 ± 0.02	0.04 ± 0.03	0 ± 0	0.51	0.743
Pomacentridae	*Pomacentrus moluccensis*	**1.94 ± 0.57**	**0 ± 0**	**0 ± 0**	**16.24**	**<0.001**
Pomacentridae	*Pomacentrus nagasakiensis*	**0 ± 0**	**0 ± 0**	**0.16 ± 0.09**	**4.93**	**0.024**
Pomacentridae	*Pomacentrus nigromanus*	0.15 ± 0.09	0.42 ± 0.14	0.02 ± 0.02	0.74	0.419
Pomacentridae	*Pomacentrus nigromarginatus*	0 ± 0	0.06 ± 0.05	0 ± 0	0.00	1.000
Pomacentridae	*Pomacentrus opisthostigma*	0.08 ± 0.05	0 ± 0	0 ± 0	4.29	0.071
Pomacentridae	*Pomacentrus pavo*	0.04 ± 0.03	0 ± 0	0 ± 0	3.13	0.221
Pomacentridae	*Pomacentrus philippinus*	0.06 ± 0.05	0 ± 0	0 ± 0	2.80	0.218
Pomacentridae	*Pomacentrus reidi*	0.2 ± 0.15	0.03 ± 0.02	0 ± 0	2.53	0.113
Pomacentridae	*Pomacentrus simsiang*	**0.14 ± 0.08**	**0.04 ± 0.03**	**0 ± 0**	**4.48**	**0.037**
Pomacentridae	*Pomacentrus taeniometopon*	0 ± 0	0 ± 0	0.04 ± 0.04	1.51	0.664
Pomacentridae	*Pomacentrus tripunctatus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Pomacentridae	*Pomacentrus vaiuli*	0.02 ± 0.02	0.02 ± 0.02	0 ± 0	0.76	0.672
Pomacentridae	*Stegastes albifasciatus*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Pomacentridae	*Stegastes fasciolatus*	0.01 ± 0.01	0 ± 0	0 ± 0	1.51	0.671
Priacanthidae	*Priacanthus hamrur*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Pseudochromidae	*Pictichromis caitlinae*	0 ± 0	0.04 ± 0.04	0 ± 0	0.00	1.000
Pseudochromidae	*Pseudochromis fuscus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Pseudochromidae	*Pseudochromis jace*	0 ± 0	0 ± 0	0.02 ± 0.02	1.51	0.668
Pseudochromidae	*Pseudochromis perspicillatus*	0 ± 0	0 ± 0	0.07 ± 0.05	3.01	0.213
Pseudochromidae	*Pseudochromis pylei*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Ptereleotridae	*Ptereleotris evides*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Ptereleotridae	*Ptereleotris hanae*	0.03 ± 0.03	0 ± 0	0 ± 0	1.51	0.669
Ptereleotridae	*Ptereleotris heteroptera*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Rachycentridae	*Rachycentron canadum*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Scaridae	*Bolbometopon muricatum*	0 ± 0	0.03 ± 0.02	0.04 ± 0.03	1.84	0.215
Scaridae	*Cetoscarus ocellatus*	0.02 ± 0.02	0.02 ± 0.02	0 ± 0	0.76	0.673
Scaridae	*Chlorurus bleekeri*	**0.06 ± 0.03**	**0 ± 0**	**0 ± 0**	**6.49**	**0.020**
Scaridae	*Chlorurus bowersi*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.660
Scaridae	*Chlorurus japanensis*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.668
Scaridae	*Chlorurus microrhinos*	0.08 ± 0.05	0 ± 0	0 ± 0	4.29	0.070
Scaridae	*Chlorurus sordidus*	**0.21 ± 0.08**	**0.02 ± 0.02**	**0.01 ± 0.01**	**8.43**	**0.002**
Scaridae	*Hipposcarus longiceps*	0.01 ± 0.01	0 ± 0	0 ± 0	1.51	0.667

Scaridae	*Scarus chameleon*	0.06 ± 0.06	0 ± 0	0 ± 0	1.51	0.669
Scaridae	*Scarus dimidiatus*	**0.13 ± 0.06**	**0 ± 0**	**0 ± 0**	**6.00**	**0.019**
Scaridae	*Scarus flavipectoralis*	0.08 ± 0.03	0.18 ± 0.08	0 ± 0	1.22	0.295
Scaridae	*Scarus forsteni*	0.06 ± 0.05	0 ± 0	0 ± 0	2.80	0.214
Scaridae	*Scarus ghobban*	0.02 ± 0.02	0.02 ± 0.02	0 ± 0	0.76	0.665
Scaridae	*Scarus niger*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.659
Scaridae	*Scarus oviceps*	0.13 ± 0.06	0.1 ± 0.05	0.04 ± 0.04	1.30	0.284
Scaridae	*Scarus prasiognathos*	0.03 ± 0.02	0.02 ± 0.02	0 ± 0	1.77	0.300
Scaridae	*Scarus psittacus*	0.07 ± 0.04	0.04 ± 0.04	0 ± 0	1.96	0.237
Scaridae	*Scarus quoyi*	0.06 ± 0.04	0 ± 0	0 ± 0	4.51	0.065
Scaridae	*Scarus rivulatus*	0.21 ± 0.21	0.02 ± 0.02	0 ± 0	1.50	0.443
Scaridae	*Scarus rubroviolaceus*	0 ± 0	0.04 ± 0.03	0 ± 0	0.00	1.000
Scaridae	*Scarus schlegeli*	0.02 ± 0.02	0 ± 0	0.02 ± 0.02	0.00	1.000
Scaridae	*Scarus spinus*	0.04 ± 0.04	0 ± 0	0 ± 0	1.51	0.666
Scaridae	*Scarus tricolor*	0.02 ± 0.02	0.13 ± 0.07	0.02 ± 0.02	0.00	1.000
Scombridae	*Grammatorcynus bilineatus*	0 ± 0	0.06 ± 0.03	0.08 ± 0.07	1.95	0.250
Scombridae	*Gymnosarda unicolor*	0 ± 0	0.1 ± 0.1	0.06 ± 0.03	0.39	0.659
Scombridae	*Scomberomorus commerson*	0 ± 0	0.08 ± 0.06	0.01 ± 0.01	0.08	0.892
Serranidae	*Aethaloperca rogaa*	**0.19 ± 0.06**	**0.08 ± 0.04**	**0 ± 0**	**9.96**	**0.001**
Serranidae	*Cephalopholis argus*	**0.33 ± 0.09**	**0.16 ± 0.06**	**0 ± 0**	**14.13**	**<0.001**
Serranidae	*Cephalopholis boenak*	**0.25 ± 0.07**	**0.04 ± 0.03**	**0.01 ± 0.01**	**12.67**	**0.001**
Serranidae	*Cephalopholis cyanostigma*	0.01 ± 0.01	0.06 ± 0.05	0 ± 0	0.12	0.817
Serranidae	*Cephalopholis microprion*	0.03 ± 0.02	0.05 ± 0.04	0.01 ± 0.01	0.25	0.677
Serranidae	*Cephalopholis miniata*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Serranidae	*Cephalopholis polleni*	0 ± 0	0.04 ± 0.04	0 ± 0	0.00	1.000
Serranidae	*Cephalopholis sexmaculata*	0 ± 0	0.03 ± 0.02	0.03 ± 0.02	1.62	0.238
Serranidae	*Cephalopholis sonnerati*	0.05 ± 0.03	0 ± 0	0 ± 0	3.07	0.216
Serranidae	*Cephalopholis spiloparaea*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Serranidae	*Cephalopholis urodeta*	0.13 ± 0.05	0.11 ± 0.06	0 ± 0	3.58	0.070
Serranidae	*Diploprion bifasciatum*	0.13 ± 0.06	0.21 ± 0.09	0.17 ± 0.08	0.09	0.778
Serranidae	*Epinephelus areolatus*	0.04 ± 0.04	0 ± 0	0.08 ± 0.07	0.43	0.658
Serranidae	*Epinephelus bontoides*	0.01 ± 0.01	0.01 ± 0.01	0 ± 0	0.97	0.447
Serranidae	*Epinephelus coioides*	0.04 ± 0.03	0.08 ± 0.06	0 ± 0	0.53	0.586
Serranidae	*Epinephelus fasciatus*	0 ± 0	0.04 ± 0.03	0 ± 0	0.00	1.000
Serranidae	*Epinephelus fuscoguttatus*	0.02 ± 0.02	0.04 ± 0.03	0 ± 0	0.51	0.732
Serranidae	*Epinephelus macrospilos*	0.02 ± 0.02	0 ± 0	0 ± 0	1.51	0.669
Serranidae	*Epinephelus malabaricus*	0.02 ± 0.02	0.02 ± 0.02	0 ± 0	0.76	0.662
Serranidae	*Epinephelus merra*	0.04 ± 0.03	0 ± 0	0 ± 0	2.80	0.222
Serranidae	*Epinephelus polyphekadion*	0 ± 0	0.02 ± 0.01	0 ± 0	0.00	1.000

Serranidae	*Epinephelus spilotoceps*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Serranidae	*Gracila albomarginata*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Serranidae	*Plectropomus areolatus*	0.02 ± 0.02	0.08 ± 0.05	0 ± 0	0.24	0.710
Serranidae	*Plectropomus laevis*	0.01 ± 0.01	0 ± 0	0 ± 0	1.51	0.660
Serranidae	*Plectropomus leopardus*	0.02 ± 0.02	0.02 ± 0.02	0 ± 0	0.76	0.669
Serranidae	*Plectropomus maculatus*	0.01 ± 0.01	0.04 ± 0.04	0 ± 0	0.15	0.891
Serranidae	*Plectropomus oligacanthus*	0.01 ± 0.01	0.14 ± 0.07	0 ± 0	0.05	0.863
Serranidae	*Pseudanthias dispar*	0.71 ± 0.71	0 ± 0	0 ± 0	1.51	0.658
Serranidae	*Pseudanthias huchtii*	0.02 ± 0.02	0.04 ± 0.04	0 ± 0	0.30	0.889
Serranidae	*Pseudanthias pleurotaenia*	0 ± 0	0.11 ± 0.11	0 ± 0	0.00	1.000
Serranidae	*Variola albimarginata*	0.02 ± 0.02	0.18 ± 0.08	0.04 ± 0.03	0.09	0.819
Serranidae	*Variola louti*	0.16 ± 0.06	0.16 ± 0.07	0.1 ± 0.07	0.50	0.505
Siganidae	*Siganus argenteus*	**0.44 ± 0.33**	**0.04 ± 0.04**	**0 ± 0**	**2.57**	**0.023**
Siganidae	*Siganus canaliculatus*	0.01 ± 0.01	0 ± 0	0 ± 0	1.51	0.666
Siganidae	*Siganus corallinus*	**0.16 ± 0.07**	**0.13 ± 0.06**	**0 ± 0**	**4.37**	**0.045**
Siganidae	*Siganus doliatus*	**0.15 ± 0.08**	**0 ± 0**	**0 ± 0**	**5.29**	**0.021**
Siganidae	*Siganus guttatus*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Siganidae	*Siganus puellus*	**0.26 ± 0.1**	**0.06 ± 0.04**	**0 ± 0**	**9.23**	**0.002**
Siganidae	*Siganus punctatissimus*	0.08 ± 0.05	0.12 ± 0.09	0 ± 0	0.92	0.411
Siganidae	*Siganus punctatus*	0.17 ± 0.08	0.13 ± 0.09	0 ± 0	2.92	0.138
Siganidae	*Siganus vulpinus*	**0.24 ± 0.07**	**0.06 ± 0.04**	**0 ± 0**	**12.96**	**0.001**
Sphyraenidae	*Sphyraena barracuda*	0 ± 0	0 ± 0	0.03 ± 0.03	1.51	0.660
Syngnathidae	*Choeroichthys brachysoma*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Syngnathidae	*Dunckerocampus pessuliferus*	0 ± 0	0.01 ± 0.01	0 ± 0	0.00	1.000
Synodontidae	*Synodus variegatus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Tetraodontidae	*Arothron caeruleopunctatus*	0.02 ± 0.02	0.05 ± 0.03	0.02 ± 0.02	0.00	1.000
Tetraodontidae	*Arothron hispidus*	0.04 ± 0.03	0 ± 0	0 ± 0	3.13	0.221
Tetraodontidae	*Arothron immaculatus*	0 ± 0	0 ± 0	0.01 ± 0.01	1.51	0.662
Tetraodontidae	*Arothron mappa*	0.02 ± 0.02	0 ± 0	0.04 ± 0.03	0.51	0.732
Tetraodontidae	*Arothron nigropunctatus*	0.02 ± 0.02	0.08 ± 0.05	0 ± 0	0.22	0.834
Tetraodontidae	*Arothron stellatus*	0 ± 0	0 ± 0	0.02 ± 0.02	1.51	0.662
Tetraodontidae	*Canthigaster amboinensis*	0 ± 0	0.03 ± 0.03	0 ± 0	0.00	1.000
Tetraodontidae	*Canthigaster valentini*	0 ± 0	0.11 ± 0.07	0 ± 0	0.00	1.000
Tetraodontidae	*Lagocephalus sceleratus*	0 ± 0	0.02 ± 0.02	0 ± 0	0.00	1.000
Toxotidae	*Toxotes jaculatrix*	0.05 ± 0.03	0 ± 0	0 ± 0	3.07	0.225
Zanclidae	*Zanclus cornutus*	**0.82 ± 0.16**	**0.77 ± 0.16**	**0.26 ± 0.12**	**6.91**	**0.011**

 ESM Table 6. Dufrêne and Legendre indicator species analysis showing fish species
1001 indicative of shallow, intermediate, and MCE depth zones. Analysis is based on fish
1002 abundance with results showing the Dufrêne and Legendre indicator value (measuring the
1003 association between the fish species and the depth group) and the associated
1004 permutational p-value (based on 99999 permutations). Only fish species with a p<0.05
1005 association for the depth band are shown.

Family / Depth group	Species	Indicator value	P(perm)	Trophic Group
Shallow				
Acanthuridae	*Acanthurus lineatus*	0.38	<0.001	Herbivore
Acanthuridae	*Acanthurus maculiceps*	0.53	<0.001	Herbivore
Acanthuridae	*Acanthurus nigricauda*	0.16	0.044	Omnivore
Acanthuridae	*Acanthurus xanthopterus*	0.19	0.009	Planktivore
Acanthuridae	*Ctenochaetus striatus*	0.54	<0.001	Planktivore
Acanthuridae	*Naso brevirostris*	0.32	0.001	Herbivore
Acanthuridae	*Naso lituratus*	0.33	0.002	Herbivore_Planktivore
Acanthuridae	*Naso unicornis*	0.18	0.044	Herbivore
Acanthuridae	*Zebrasoma scopas*	0.49	<0.001	Herbivore
Balistidae	*Balistapus undulatus*	0.59	<0.001	Mobile.Invert
Caesionidae	*Caesio lunaris*	0.28	0.017	Planktivore
Caesionidae	*Pterocaesio tile*	0.26	0.004	Planktivore
Carangidae	*Scomberoides lysan*	0.17	0.031	Piscivore_Mobile.Invert
Carcharhinidae	*Carcharhinus melanopterus*	0.37	0.011	Piscivore_Mobile.Invert
Chaetodontidae	*Chaetodon baronessa*	0.44	<0.001	Corallivore
Chaetodontidae	*Chaetodon citrinellus*	0.21	0.009	Omnivore
Chaetodontidae	*Chaetodon ephippium*	0.21	0.021	Omnivore
Chaetodontidae	*Chaetodon lineolatus*	0.26	0.004	Corallivore
Chaetodontidae	*Chaetodon lunula*	0.23	0.011	Mobile.Invert
Chaetodontidae	*Chaetodon lunulatus*	0.48	<0.001	Corallivore
Chaetodontidae	*Chaetodon rafflesii*	0.46	<0.001	Corallivore
Chaetodontidae	*Chaetodon semeion*	0.25	0.003	Omnivore

Chaetodontidae	*Chaetodon speculum*	0.25	0.005	Corallivore
Chaetodontidae	*Chaetodon trifascialis*	0.46	<0.001	Corallivore
Chaetodontidae	*Chaetodon vagabundus*	0.4	<0.001	Corallivore
Chaetodontidae	*Heniochus varius*	0.21	0.022	Mobile.Invert
Cirrhitidae	*Paracirrhites forsteri*	0.21	0.009	Piscivore
Labridae	*Choerodon anchorago*	0.31	0.001	Mobile.Invert
Labridae	*Diproctacanthus xanthurus*	0.44	<0.001	Corallivore
Labridae	*Halichoeres hortulanus*	0.3	0.002	Mobile.Invert
Labridae	*Halichoeres melanurus*	0.25	0.003	Mobile.Invert
Labridae	*Halichoeres podostigma*	0.17	0.031	Mobile.Invert
Labridae	*Hemigymnus melapterus*	0.37	<0.001	Mobile.Invert
Labridae	*Hologymnosus annulatus*	0.24	0.014	Piscivore
Labridae	*Labroides dimidiatus*	0.38	0.013	Piscivore_Mobile.Invert
Labridae	*Thalassoma hardwicke*	0.87	<0.001	Mobile.Invert
Labridae	*Thalassoma lunare*	0.75	<0.001	Mobile.Invert
Lutjanidae	*Lutjanus bohar*	0.4	0.002	Mobile.Invert
Lutjanidae	*Lutjanus carponotatus*	0.23	0.006	Piscivore_Mobile.Invert
Lutjanidae	*Lutjanus decussatus*	0.33	0.001	Piscivore_Mobile.Invert
Lutjanidae	*Lutjanus fulvus*	0.25	0.003	Piscivore_Mobile.Invert
Lutjanidae	*Lutjanus rivulatus*	0.2	0.033	Piscivore_Mobile.Invert
Lutjanidae	*Lutjanus russellii*	0.19	0.014	Piscivore_Mobile.Invert
Lutjanidae	*Lutjanus semicinctus*	0.61	<0.001	Piscivore_Mobile.Invert
Mullidae	*Parupeneus crassilabris*	0.42	<0.001	Mobile.Invert
Muraenidae	*Gymnothorax javanicus*	0.18	0.048	Piscivore_Mobile.Invert
Nemipteridae	*Pentapodus trivittatus*	0.36	0.001	Piscivore_Mobile.Invert
Nemipteridae	*Scolopsis bilineata*	0.45	<0.001	Mobile.Invert
Nemipteridae	*Scolopsis trilineata*	0.19	0.033	Piscivore_Mobile.Invert

Pomacentridae	*Amblyglyphidodon curacao*	0.2	0.015	Herbivore_Planktivore
Pomacentridae	*Amblyglyphidodon ternatensis*	0.17	0.031	Herbivore_Planktivore
Pomacentridae	*Chromis margaritifer*	0.45	<0.001	Herbivore_Planktivore
Pomacentridae	*Chromis ternatensis*	0.29	0.003	Planktivore
Pomacentridae	*Chromis weberi*	0.27	0.006	Planktivore
Pomacentridae	*Chrysiptera cyanea*	0.25	0.004	Herbivore_Planktivore
Pomacentridae	*Dascyllus reticulatus*	0.18	0.022	Planktivore
Pomacentridae	*Dischistodus melanotus*	0.2	0.023	Herbivore
Pomacentridae	*Neoglyphidodon melas*	0.25	0.003	Omnivore
Pomacentridae	*Plectroglyphidodon dickii*	0.21	0.009	Piscivore
Pomacentridae	*Pomacentrus bankanensis*	0.34	<0.001	Herbivore_Planktivore
Pomacentridae	*Pomacentrus chrysurus*	0.22	0.008	Omnivore
Pomacentridae	*Pomacentrus coelestis*	0.21	0.009	Mobile.Invert_Planktivore
Pomacentridae	*Pomacentrus moluccensis*	0.58	<0.001	Omnivore
Scaridae	*Chlorurus bleekeri*	0.17	0.030	Herbivore
Scaridae	*Chlorurus sordidus*	0.29	0.002	Herbivore
Scaridae	*Scarus dimidiatus*	0.17	0.031	Herbivore
Serranidae	*Aethaloperca rogaa*	0.24	0.016	Piscivore_Mobile.Invert
Serranidae	*Cephalopholis argus*	0.31	0.004	Piscivore
Serranidae	*Cephalopholis boenak*	0.31	0.001	Piscivore_Mobile.Invert
Siganidae	*Siganus argenteus*	0.19	0.035	Omnivore
Siganidae	*Siganus doliatus*	0.17	0.032	Herbivore
Siganidae	*Siganus puellus*	0.21	0.008	Omnivore
Siganidae	*Siganus vulpinus*	0.31	0.001	Herbivore_Planktivore
Zanclidae	*Zanclus cornutus*	0.37	0.023	Omnivore
Intermediate				
Acanthuridae	*Acanthurus thompsoni*	0.22	0.008	Planktivore
Acanthuridae	*Naso thynnoides*	0.32	<0.001	Omnivore
Balistidae	*Odonus niger*	0.33	0.032	Planktivore
Balistidae	*Sufflamen bursa*	0.39	0.001	Mobile.Invert

Caesionidae	*Pterocaesio digramma*	0.17	0.028	Planktivore
Carcharhinidae	*Triaenodon obesus*	0.18	0.042	Piscivore_Mobile.Invert
Chaetodontidae	*Coradion chrysozonus*	0.22	0.033	Omnivore
Chaetodontidae	*Forcipiger flavissimus*	0.17	0.030	Mobile.Invert
Chaetodontidae	*Forcipiger longirostris*	0.19	0.015	Mobile.Invert
Labridae	*Bodianus diana*	0.19	0.009	Mobile.Invert
Labridae	*Bodianus dictynna*	0.22	0.010	Mobile.Invert
Labridae	*Choerodon zosterophorus*	0.22	0.012	Mobile.Invert
Labridae	*Halichoeres chrysus*	0.39	<0.001	Mobile.Invert
Labridae	*Halichoeres prosopeion*	0.33	0.002	Mobile.Invert
Lethrinidae	*Monotaxis grandoculis*	0.22	0.020	Mobile.Invert
Lutjanidae	*Macolor macularis*	0.29	0.004	Piscivore_Mobile.Invert
Monacanthidae	*Paraluteres prionurus*	0.17	0.031	Omnivore
Pomacanthidae	*Centropyge bicolor*	0.3	0.002	Mobile.Invert
Pomacanthidae	*Genicanthus lamarck*	0.29	0.001	Planktivore
Pomacentridae	*Pomacentrus amboinensis*	0.36	<0.001	Herbivore_Planktivore
Pomacentridae	*Pomacentrus nigromanus*	0.27	0.007	Planktivore
Serranidae	*Plectropomus oligacanthus*	0.19	0.016	Piscivore_Mobile.Invert
Serranidae	*Variola albimarginata*	0.19	0.034	Piscivore
Tetraodontidae	*Canthigaster valentini*	0.17	0.031	Omnivore
MCE				
Balistidae	*Abalistes stellatus*	0.41	<0.001	Piscivore_Mobile.Invert
Balistidae	*Sufflamen fraenatum*	0.23	0.023	Mobile.Invert
Carangidae	*Carangoides ferdau*	0.35	0.011	Piscivore_Mobile.Invert
Labridae	*Choerodon jordani*	0.25	0.003	Mobile.Invert
Lethrinidae	*Lethrinus amboinensis*	0.25	0.017	Piscivore_Mobile.Invert
Lethrinidae	*Lethrinus lentjan*	0.17	0.031	Piscivore_Mobile.Invert

Malacanthidae	*Hoplolatilus purpureus*	0.25	0.002	Planktivore
Mullidae	*Parupeneus heptacanthus*	0.2	0.027	Piscivore_Mobile.Invert
Nemipteridae	*Pentapodus nagasakiensis*	0.36	0.004	Mobile.Invert
Nemipteridae	*Pentapodus paradiseus*	0.39	<0.001	Piscivore_Mobile.Invert
Nemipteridae	*Scolopsis vosmeri*	0.25	0.003	Mobile.Invert
Pomacentridae	*Pomacentrus nagasakiensis*	0.17	0.031	Herbivore_Planktivore

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