

1 Variation in neotropical river otter (*Lontra longicaudis*) diet: Effects of an invasive prey
2 species

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4 Effects of invasive prey on Neotropical river otter diet

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6 Diego Juarez Sanchez^{1*}, John G. Blake^{1¶}, Eric C. Hellgren^{1¶}

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8 ¹Department of Wildlife Ecology and Conservation, University of Florida, Gainesville,
9 Florida United States of America.

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11 * Corresponding author

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13 E-mail: adjuarez@ufl.edu

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15 ¶ These authors contributed equally to this work.

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45 **Abstract**

46 Due to human activities, some species have expanded their distribution into areas
47 that were historically difficult or impossible to reach by natural dispersal. Such species
48 may become invasive if they successfully establish reproductive populations. Predation is
49 one of the main barriers that exotic species may face in newly colonized areas. We
50 evaluated the effect of an invasive prey (armored catfish: *Pterygoplichtys* sp.) on the
51 dietary niche breadth and trophic level of a native predator (Neotropical river otter:
52 *Lontra longicaudis*) in northern Guatemala. We examined otter scats from three rivers:
53 two where the invasive armored catfish occurred and one without the invasive fish.
54 Samples were collected two and seven years after the first report of the catfish in the
55 area. We performed gross scat analysis and stable isotope analyses of nitrogen and carbon
56 of fecal matter. Where the invasive armored catfish occurred, it was the main prey item
57 for *L. longicaudis*. Particularly in the river outside of protected areas seven years after the
58 first report of the catfish, where it accounted for 49% of the otter diet. Concordance was
59 found between the two techniques to estimate dietary niche breadth and trophic level.
60 The dietary niche breath of otters was narrower seven years after the invasion in
61 comparison to two years after the invasion in both invaded rivers, but, the extent of the
62 reduction was less inside the protected area. Finally, the trophic level of otters also
63 showed a reduction related to the occurrence of the armored catfish on their diet.

64 **Resumen**

65 Como producto de las actividades humanas algunas especies han expandido su
66 distribución hacia áreas que históricamente eran difícil o imposible de alcanzar mediante
67 de dispersión natural. Estas especies pueden convertirse en invasoras si establecen
68 exitosamente poblaciones reproductivas. La depredación es una de las principales
69 barreras que las especies exóticas deben afrontar en las áreas recientemente colonizadas.
70 Evaluamos los efectos de una especie invasora (el pez diablo: *Pterygoplichtys* sp.) sobre la
71 amplitud de nicho alimenticio y el nivel trófico de un depredador nativo (la nutria de río
72 Neo-tropical: *Lontra longicaudis*) en el norte de Guatemala. Examinamos las excretas de
73 nutrias provenientes de tres ríos: dos donde el pez diablo se encuentra presente y uno
74 donde este invasor aún está ausente. Las muestras fueron colectadas dos y siete años
75 después del primer reporte de del pez diablo en le área. Realizamos un análisis
76 macroscópico de las excretas y análisis de isotopos estables de nitrógeno y carbono de la
77 materia fecal. Donde el pez diablo invasor estaba presente, fue el principal ítem
78 alimenticio de *L. longicaudis*. Particularmente en el río ubicado fuera de áreas protegidas
79 siete años después del primer reporte del pez diablo, donde este consistió en el 49% de la
80 dieta de la nutria. Encontramos concordancia entre las dos técnicas para estimar la
81 amplitud de nicho dietario y nivel trófico. La amplitud de nicho dietario de las nutrias fue
82 más angosto siete años después de la invasión en comparación con dos años luego de la
83 invasión en ambos ríos invadidos, pero la magnitud de la reducción fue inferior dentro del

84 área protegida. Finalmente, observamos una reducción en el nivel trófico de las nutrias
85 relacionada con la ocurrencia del pez diablo en su dieta.

86 Introduction

87 Predators may change their diet after an exotic prey species becomes established
88 and abundant in the predator's range [1–4]. Inclusion of such a species in a predator's diet
89 can lead to a shift in the predator's dietary niche, which may become wider or narrower,
90 depending on the intensity of use of the new resource and changes in the use of
91 alternative native prey [5]. Furthermore, the type of prey that a predator eats defines its
92 trophic level (e.g., primary consumer, secondary consumer). Both niche breadth and
93 trophic levels can be evaluated using gross scat analysis and stable isotopes analyses.

94
95 An important group of invasive species in freshwater communities are the armored
96 catfishes of the South American family Loricariidae, a diverse group of fishes with 928
97 valid species and eight subfamilies, including the genus *Pterygoplichthys*, commonly
98 known as the suckermouth armored catfish (hereafter ACF; [6]). These catfish are very
99 popular in the aquarium trade, easily domesticated, exhibit parental care [7], possess
100 physiological tolerance to adverse conditions [8–12], have wide distribution ranges [13],
101 and possess high reproductive and growth rates [14,15]. They feed on detritus, an
102 abundant resource, especially in human-modified areas, and therefore have a low
103 fractional trophic level (FTL) [13]. These traits contribute to their invasiveness, as they
104 fulfill the six life-history variables associated with species that successfully establish
105 invasive populations [16]. The presence of ACF as an invasive species has been
106 documented for at least 21 countries in five continents [17]. In 2005, an established
107 population of *Pterygoplichthys pardalis* was found in Laguna Frontera at the mouth of the
108 Usumacinta River, Tabasco, Mexico [18]. Two years later, *P. pardalis* was reported in
109 Guatemala in the headwaters of the San Pedro River, a tributary of the Usumacinta River
110 (Juarez-Sanchez and J. F. Moreira, in prep.). The species identification, however, has not
111 been confirmed because *P. pardalis* can be misidentified and confused with other species
112 of *Pterygoplichthys* given that identification is based on ventral color patterns and
113 hybridization with *P. disjunctivus* has been reported elsewhere [19–21].

114
115 The ACF has been reported to have positive effects by generating nutrient
116 hotspots, making nutrients available for producers in nutrient-depleted areas [22].
117 However, the amount of nutrients released by the ACF does not compensate for its
118 grazing pressure [23]. Other negative impacts of ACF have been documented in places
119 where they have established invasive populations. These impacts include asphyxiating
120 native predators in Puerto Rico [24]; preying on native fish eggs and first-feeding fry in
121 Thailand [25]; competing for forage with native species, reducing biofilm from the
122 substrate, and changing the proportions of dissolved nutrients in the Philippines and
123 Mexico [23,26,27]; harassing manatees [28–30]; and possibly promoting erosion with their
124 nesting burrows in Florida and Mexico [7,31]. These impacts could occur anywhere ACF
125 establish an invasive population. Invasive ACF are preyed upon by native piscivorous

126 predators such as common snook (*Centropomus undecimalis*) and the Neotropical
 127 cormorant (*Phalacrocorax brasilianus*) [32,33], although their effects on these and other
 128 native predators have not been evaluated.

129

130 Otters (Lutrinae) are mid-sized carnivores that are top predators in freshwater
 131 wetlands and riverine systems because of their high energetic demand and trophic
 132 position [34,35]. The Neotropical river otter (*Lontra longicaudis*; hereafter NRO) is a semi-
 133 aquatic mustelid that preys primarily on benthic slow-moving fish [36], but also feeds on
 134 crustaceans, mollusks, reptiles, and mammals (Table 1). This species is distributed from
 135 northern Mexico to northern Argentina, coexisting with different community assemblages
 136 of prey species, and adapting its foraging behavior according to the local community [37].
 137 Where ACF are native, they coexist with the NRO and constitute one of the most
 138 important prey items in its diet [37–40]. However, the role of ACF as a prey item for NRO
 139 in areas where ACF has been introduced is unknown and may be reshaping the foraging
 140 ecology of the NRO in those areas.

141

142 **Table 1 Food items reported as present in diets of Neotropical river otters across their geographic range.**

Locality	Primary item	Other items	Citation
Oaxaca, México.	crustaceans (53.0%)	fish (33.1%), insects (9.8%) and amphibians (4.0%)	[41]
México state, México.	fish (92.4%)	invertebrates (3.5%), amphibians (2.9%) and plant matter (1.8%)	[42]
Alto Cauca, Colombia.	fish (76.7%)	insects (12.7%), reptiles (0.7%), and others (9.9%)	[39]
Salta, Argentina.	fish (53%)	insects (24%), crustaceans (16%), amphibians (7%), and reptiles, mammals and mollusks (<0.1%)	[37]
Rio de Janeiro, Brazil.	fish (86%)	crustaceans (71%), amphibians (10%), mammals (3%), birds (0.6%), reptiles (0.2%) and others (0.7%)	[43]
Rio Grande do Sul, Brazil.	Fish (Loricariidae 41.1%, Cichlidae 21%, Pimelodidae 12.6%, Characidae 6.5 %)	other fish (12.5 %), Megaloptera (3.6%), mammals (1.2%), insects (0.4%), Decapoda (0.1%), birds (0.3%), snakes (0.3%) and plant matter (0.4%)	[38]
Rio Grande do Sul, Brazil.	fish (82.6 %)	crustaceans (20.6%), birds (4.5%), mammals and snakes (3.7%), Coleoptera (1.2%), amphibians (0.8%) and mollusks (0.4%)	[44]
Rio Grande do Sul, Brazil	fish	mammals, amphibians, birds, snakes, insects, crustaceans mollusks and eggs.	[45]

143 Percent values are frequency of occurrence and do not add to 100%.

144 The main objective of this study was to determine if invasive armored catfish
145 affected the diet of Neotropical river otters. Given that NRO feed on ACF in areas where
146 native populations overlap [38,46,47], we hypothesized that NRO will change their diet to
147 include ACF in rivers where invasive populations of ACF occur. We predicted that where
148 ACF are present, they will become the main prey of NRO and reduce the niche breadth of
149 NRO. If ACF become the main prey of the NRO, we also predicted a lower trophic level for
150 the NRO in areas where ACF are present due to the low trophic level of the ACF.

151 **Materials and methods**

152 **Study area**

153 The study area is located at northern Guatemala in the district of Petén (between
154 15.50° and 17.50° N and -88.50° and -91.25° W) and includes the Usumacinta and Mopan
155 rivers (Fig. 1). Precipitation ranges from 1,200 to 4,000 mm/year on a gradient decreasing
156 northward (INSIVUMEH, 2016). Major habitat types in the study area consist of
157 subtropical moist forest in the north, subtropical very moist forest in the south, and
158 tropical very moist forest in the southeast [48]. The entire study area consists of lowland
159 forest, with elevations ranging from 0 to 1000 masl.

160
161 In northern Guatemala, rivers flow into the Gulf of Mexico or into the Caribbean
162 Sea watersheds (Fig. 1). Thus, bodies of fresh water are isolated by large expanses of land
163 in the headwaters, and large distances between river mouths along the coast. The Mopan
164 River flows northwards from southern Petén and then east in central Belize into the
165 Caribbean Sea. The Usumacinta River runs northwest into the Gulf of Mexico. Samples
166 were collected from the Mopan River and two tributaries of the Usumacinta River: The
167 San Pedro River and the Pasion River. In Guatemala, the San Pedro River flows along the
168 southern border of Laguna del Tigre National Park whereas the Pasion and the Mopan
169 rivers mainly run through private lands that are under different land uses.

170
171 **Fig 1. Study area** for collection of Neotropical river otter scats in northern Guatemala. Grey circles represent
172 samples collected in 2009-2010; black solid circles represent samples collected in 2015; black hollow circle
173 represents the area where samples were collected in 2016. The dashed area represents the Usumacinta
174 basin divided in sub-basins, where the armored catfish has been reported (ACF). The striped area represents
175 the Caribbean runoff where no ACF has been reported. Grey areas represent protected areas.

176
177 The Usumacinta basin has at least 61 fish species distributed in 25 families. The
178 two main families in Usumacinta basin are Cichlidae with 18 species and Poeciliidae with
179 10 species [6,13,49,50]. To our knowledge, no peer-reviewed document has been
180 published that describes fishes of the Mopan River within the borders of Guatemala. Thus,
181 information about the fish assemblage in this river is based on information from the
182 estuarine area in Belize. Therefore, the number of fish species that we are considering as
183 present and potential prey for otters in the river headwaters within Guatemalan territory
184 may be inflated. In Mopan River, there are at least 103 fish species distributed in 32

185 families, including the invasive tilapia (*Oreochromis aureus*). The main families are
186 Cichlidae with 14 species and Poeciliidae with 16 species [6,13,51]. Exotic tilapia is
187 widespread across all Guatemala due to multiple introductions, both accidental and
188 deliberate from aquaculture or governmental fisheries restocking. The Asian grass carp
189 (*Ctenopharyngodon idella*) and the ACF have been found in the Usumacinta basin, but the
190 origins of these invasions are not clear.

191 **Scat Collection**

192 Samples were collected during three periods: May 2009 – April 2010, May – July
193 2015, and June 2016. The search for otter scats was conducted from a small boat moving
194 at slow speeds (< 5 km/h) close to the shoreline, with scats and latrines typically found on
195 protruding structures (e.g. rocks or fallen trees). This search was conducted along both
196 shorelines of the river in opposite directions. All scats were collected, placed in paper
197 and/or plastic bags with silica gel, and stored in a dry environment. Otter scats were
198 identified by their appearance, as no other species present in the study area have similar
199 scats (located on protruding sites along the river shore, low fecal matter and high content
200 of fish or crab remains) [52]. If a scat was found but its identification was doubtful, it was
201 collected and included in the analysis only if otter hair from grooming was found on it.
202 Each scat was assigned a unique code and the geographic coordinates of its location were
203 recorded using a handheld GPS unit (GARMIN © Astro 320, Garmin Ltd. Kansas City, USA).
204

205 We sampled the Usumacinta basin during 2009-2010 using continuous searches
206 along the rivers, including 38.5 km of the San Pedro River (starting from Paso Caballos
207 village and moving west) and 89.1 km of the Pasion River (starting from Sayaxche town
208 and heading west). We sampled the Usumacinta and Mopan basins in 2015 by organizing
209 the search for scats into segments of 10 km, with segments separated by at least 10 km. In
210 the Usumacinta basin, we sampled along 40, 50, and 30 km of the San Pedro, La Pasion,
211 and Usumacinta rivers, respectively. Surveys began in Paso Caballos for the San Pedro, in
212 Sayaxche for the Pasion, and in Betel town for the Usumacinta River. The Mopan River
213 was sampled along 10 km in 2015 near La Polvora military base. In June 2016, local
214 fisherman collected samples for in the Mopan River near La Polvora military base, no
215 exact georeference was collected per sample (Fig. 1).

216 **Scat Sample Preparation and Analysis**

217 Samples of fecal matter were collected from each scat, homogenized using a
218 porcelain mortar and pestle, stored in glass vials and sent to the Light Stable Isotope Mass
219 Spectrometry Laboratory in the Department of Geological Sciences at the University of
220 Florida for stable isotope analysis (SIA) of $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$. Samples were analyzed using a
221 Thermo Electron DeltaV Advantage isotope ratio mass spectrometer coupled with a
222 ConFlo II interface linked to a Carlo Erba NA 1500 CNHS Elemental Analyzer. All carbon
223 isotopic results are expressed in standard delta notation relative to VPDB. All nitrogen
224 isotopic results are expressed in standard delta notation relative to AIR. Hard remains (i.e.,

225 scales, skeleton pieces) were separated and identified to the lowest possible taxonomic
226 level. A list of potential prey species for otters was made, consisting of all the fish species
227 reported in the study area that have a reported maximum total length ≥ 100 mm (S1
228 Table). Size selection was based on the assumption that otters prefer to feed on fish
229 within the 100-150 mm size range [53]. Prey remains that could be identified were fish
230 scales, otoliths or vertebra; crustacean shells; and mammal hairs. A scale guide was
231 constructed for 68 of the 80 scaled fish species that are found in the sampled river basins
232 and that were consider potential prey of the NRO [54]. Scales were obtained from
233 museum specimens at the Florida Museum of Natural History (FLMNH) and El Colegio de
234 la Frontera Sur in México (ECOSUR). Scales from these fish species were cleaned with
235 water and alcohol, placed on glass slides with nail polish, and sealed with a coverslip to
236 make semi-permanent slides. For 10 catfish species that do not have scales, the
237 identification was based on fin spines, using reference material from the
238 zooarchaeological collection at FLMNH. Hairs found in the scats were identified using a
239 hair-identification guide [55] and reference material from the mammal collection of the
240 Museo de Historia Natural (MUSNAT) at the Universidad de San Carlos de Guatemala
241 (USAC). Otter hair (product of grooming) was saved and pressed between glass slides and
242 coverslips for future analysis.

243 **Data Analyses**

244 For data analyses, the sampling units were the rivers (San Pedro, La Pasion,
245 Mopan) with year as factor (2009-2010 and 2015-2016). The year effect represents 2 and
246 7 years after the advent of the ACF invasion. Comparisons were made over time (i.e.,
247 same river, different year) only using data from Pasion and San Pedro rivers where the
248 ACF are present; we additionally looked at differences across river basins in the same
249 sampling years (i.e., different river, same year), combining 2015-2016 records as one year
250 and including the Mopan River where ACF do not occur.

251
252 The importance of each prey species can be biased by abundant and conspicuous
253 hard remains that are identifiable for some species, even if those species are consumed in
254 low numbers, due to differential digestibility of prey items. This overestimation of some
255 species can then lead to an underestimate of overall diet diversity. On the other hand,
256 when using SIA of predators, one can measure diet diversity breadth and comparative
257 trophic levels but with no taxonomic information about the prey. For this reason, we used
258 both techniques, expecting to find concordance between them.

259 **Gross scat analysis (GSA)**

260
261
262 Accumulation curves were constructed using program EstimateS (© Colwell 2013,
263 Connecticut, USA) where the expected number of prey species found in a given number of
264 scats is obtained by

$$265 \tau(h) = S_{obs} - \sum \alpha_{jh} S_j$$

267
$$\alpha_{jh} = (H-h)!(H-j)!/(H-h-j)!H!$$

268

269 were $\tau(h)$ is the estimated number of species for h number of scats; S_{obs} is the
270 number of species actually observed; S_j is the number of prey species found in j scats; α_{jh}
271 is a combinatorial coefficient; H is the total number of scats; h is the number of possible
272 combination of scats that add up to j scats; and j is the number of scats per moment or
273 segment of the curve [56].

274

275 The importance of different food items, including the ACF, in the NRO diet was
276 assessed through GSA, using the percentage of occurrence. Percentage of occurrence was
277 estimated for a prey item by dividing the number of scats with item i by the total number
278 of reported items. To compare the NRO niche breadth between basins, with different prey
279 communities, Levin's index was used:

280

281
$$B = 1 / \sum p_i^2$$

282

283 where p is the proportion of food items from category i [57]. The Levin's niche-
284 breadth index can be standardized using:

285

286
$$B_a = B - 1 / n - 1$$

287

288 where B_a is the standardized Levin's niche-breadth index, B is Levin's niche-
289 breadth index, and n is the number of recorded species. Levin's index ranges from 1 to n
290 and from 0 to 1 in its standardized version. In both cases, its minimum value is reached
291 when all reported prey belongs to only one species (specialist predator) and is at its
292 maximum when all the species are consumed in the same proportion (generalist
293 predator). It has been suggested that values of $B_a > 0.6$ represent a generalist and values
294 of $B_a < 0.4$ a specialist [58,59]. To estimate confident intervals the samples (scats) were
295 randomly selected with replacement (bootstrap), then the index was re-estimated with
296 the resulting set of samples. This procedure was repeated 1000 times and the confidence
297 intervals calculated.

298

299 The NRO's fractional trophic level, which represents the trophic distance of a
300 consumer species from producers, in each basin was estimated using Pauly and
301 Palomares's (2005) formula:

302

303
$$FTL_i = 1 + \sum_j (FTL_j DC_{ij})$$

304

305 where FTL_i is the fractional trophic level of the consumer, +1 is a constant
306 increment for the FTL of a consumer, FTL_j is the fractional trophic level of the prey j , and
307 DC_{ij} is the proportion of contribution of prey j to the diet of consumer i . Prey FTL_j values
308 were obtained from FishBase database[13]. The DC_{ij} was based on proportion of
309 occurrence values by river-year combination in the otter scats. To estimate confidence
310 intervals a bootstrap procedure was developed as explained above.

311

312 **Stable isotope analysis (SIA)**

313

314 Stable isotope analysis (SIA) measures the proportion of heavy to light stable
315 isotopes in a sample [63,64]; its values are expressed in delta notation (δ) or per mil (‰)
316 and estimated with this equation:

317

$$318 \quad \delta = ((R. \text{ sample}/R. \text{ standard}) - 1) \times 1000$$

319

320 where R = heavy isotope / light isotope obtained with a mass spectrometer.

321

322 Isotopic values of a predator are higher than those of its prey due to a process
323 called fractionation, wherein the molecules with the lighter isotopes, given their lighter
324 overall weight, react faster and can be metabolized and excreted faster than the heavier
325 ones. This process results in the predator being enriched with a higher proportion of
326 heavier isotopes than its prey [61,64]. The mean value of this fractionation across taxa is
327 3.4‰ (1 SD = 1‰) for $\delta^{15}\text{N}$ and 0.4‰ (1 SD = 1.3‰) for $\delta^{13}\text{C}$ [61]. These values are the
328 expected increment of the isotopic value when molecules are assimilated from prey tissue
329 to predator tissue (from lower to higher trophic levels;[65]).

330

331 Isotopic values of different tissues, such as bone, blood, hair or muscle, have been
332 used to evaluate the diets of a wide range of species [66–73]. Normally, tissue samples are
333 obtained from dead or captured specimens but these invasive techniques are sometimes
334 difficult or impossible to use, especially for secretive, rare or endangered species.
335 However, controlled experiments have shown that SIA based on feces is sensitive to
336 changes in the diet over periods of 3 hours for insectivorous bats [74] and, thus, represent
337 the isotopic values of the latest meals of the individual that produced the scat[75]. In
338 carnivores and omnivores, SIA based on scats can be used to estimate the main type of
339 prey and nutrient flow, using $\delta^{15}\text{N}$ to infer the range of trophic positions or FTLs at which a
340 predator eats, and $\delta^{13}\text{C}$ to determine the type of producers that supported the specific
341 trophic chain [61,76–78]. Further, the variance of isotopic values of a population may
342 represent the niche width (or breadth) of a consumer [79].

343

344 Taking $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values from individual scats as samples from each river, we
345 evaluated the data for normality using histograms, qq-plots and a Shapiro-Wilk normality
346 test; all values followed a normal distribution. To evaluate differences between variances
347 in $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ as a niche breadth metric, a Levene's homoscedasticity test was used. To
348 test for differences in mean $\delta^{15}\text{N}$ values between rivers and years we use a two-factor
349 ANOVA after a log transformation of the data to correct for lack of homoscedasticity; a
350 post-hoc paired *t*-test with Bonferroni adjusted *p*-values was used to evaluate where the
351 differences occurred. All the statistical procedures except for the species accumulation
352 curves were performed using the program R [80].

353

354 Results

355 Field collection of scats yielded 286 samples identified as coming from the NRO.
 356 After eliminating scats that had some type of contamination (e.g., wood, mud, or termite
 357 nest), 177 samples of fecal matter were sent for isotopic analysis (Table 2). We identified
 358 35 scaled fish species, including three nonnative fish species (*Oreochromis aureus*,
 359 *Ctenopharyngodon idella* and *Pterygoplichtys* sp.) from otter scats. In addition, remains of
 360 unidentified insects, one reptile, one unidentified mammal, and unidentified crabs and
 361 crayfish were recovered from the scats (Table 3).

362
 363

Table 2. Scats of Neotropical river otters collected in northern Guatemala.

River	No. of scats collected (year)	No. of scats without contamination (year)
Usumacinta	1 (2015)	0
	1 Total	0 Total
San Pedro	36 (2009)	20 (2009)
	117 (2015)	55 (2015)
	153 Total	75 Total
La Pasion	52 (2010)	36 (2010)
	40 (2015)	34 (2015)
	92 Total	70 Total
Mopan	1 (2015)	1 (2015)
	39 (2016)	31 (2016)
	40 Total	32 Total

364 Only scats without contamination were used for fecal matter isotope analyses

365
 366
 367
 368
 369

Table 3. Number of records (No), and percentage of the total of prey species (%) found in otter scats collected from the Mopan, Pasion and San Pedro (San Pe.) rivers, northern Guatemala, during 2009-2010 (09-10) and 2015-2016 (15-16).

	Mopan 15-16		Pasion 09-10		Pasion 15-16		San Pe. 09-10		San Pe. 15-16	
	No	%	No	%	No	%	No	%	No	%
Belonidae										
<i>Strongylura hubbsi</i>	0	0	0	0	0	0	3	2.2	0	0
<i>Strongylura marina</i>	1	1.1	0	0	0	0	0	0	0	0
Carangidae										
<i>Caranx latus</i>	1	1.1	0	0	0	0	0	0	0	0
Centropomidae										
<i>Centropomus ensiferus</i>	1	1.1	0	0	0	0	0	0	0	0
Characidae										
<i>Astianax fasciatus</i>	0	0	0	0	0	0	9	6.5	1	0.3
Cichlidae										
<i>Chuco intermedius</i>	6	6.6	0	0	0	0	0	0	1	0.3

<i>Cincolichthys bocourti</i>	1	1.1	0	0	1	1.4	0	0	1	0.3
<i>Cincolichthys pearsei</i>	0	0	0	0	0	0	0	0	1	0.3
<i>Cribroheros robertsoni</i>	1	1.1	13	9.1	4	5.5	22	15.8	12	4.1
<i>Kihmchithys ufermammi</i>	0	0	0	0	0	0	3	2.2	1	0.3
<i>Maskaheros argenteus</i>	0	0	1	0.7	0	0	0	0	0	0
<i>Mayaheros urophtalmus</i>	2	2.2	5	3.5	5	6.8	5	3.6	17	5.9
<i>Oreochromis aureus</i>	8	8.8	5	3.5	6	8.2	4	2.9	21	7.3
<i>Parachromis friedrichsthalii</i>	2	2.2	9	6.3	5	6.8	1	0.7	4	1.4
<i>Petenia splendida</i>	0	0	1	0.7	0	0	0	0	3	1
<i>Rheoheros lentiginosus</i>	0	0	1	0.7	1	1.4	0	0	0	0
<i>Rocio octofasciata</i>	0	0	2	1.4	1	1.4	0	0	3	1
<i>Thorichthys affinis</i>	0	0	2	1.4	0	0	0	0	1	0.3
<i>Thorichthys aureus</i>	6	6.6	0	0	0	0	0	0	0	0
<i>Thorichthys meeki</i>	5	5.5	11	7.7	2	2.7	10	7.2	25	8.6
<i>Thorichthys pasionis</i>	0	0	10	7.0	1	1.4	10	7.2	8	2.8
<i>Trichromis salvini</i>	0	0	0		0	0	0	0	2	0.7
<i>Vieja bifasciata</i>	0	0	3	2.1	6	8.2	1	0.7	21	7.3
<i>Vieja melanurus</i>	0	0	3	2.1	1	1.4	0	0	8	2.8
Cyprinidae										
<i>Ctenopharyngodon idella</i>	0	0	7	4.9	0	0	0	0	5	1.7
Eleotridae										
<i>Dormitator maculatus</i>	1	1.1	0	0	0	0	0	0	0	0
Gerreidae										
<i>Eugerres mexicanus</i>	0	0	0	0	0	0	0	0	4	1.4
Hemiramphidae										
<i>Hyporhamphus mexicanus</i>	0	0	0	0	0	0	9	6.5	2	0.7
Lepisosteidae										
<i>Aractosteus tropicus</i>	0	0	0	0	0	0	1	0.7	0	0
Loricariidae										
<i>Pterygoplichthys</i> sp	0	0	14	9.9	36	49.3	23	16.5	75	25.9
Megalopidae										
<i>Megalops atlanticus</i>	0	0	0	0	0	0	0	0	1	0.3
Mugilidae										
<i>Mugil cephalus</i>	0	0	1	0.7	0	0	0	0	0	0
Poeciliidae										
<i>Belonesox belizanus</i>	0	0	5	3.5	0	0	1	0.7	12	4.1
<i>Poecilia mexicana</i>	1	1.1	3	2.1	1	1.4	17	12.2	24	8.3
<i>Poecilia petenensis</i>	0	0	11	7.7	1	1.4	17	12.2	26	9
Ariidae, Heptapteridae, Ictaluridae										
Catfish	8	8.8	3	2.1	1	1.4	3	2.2	4	1.4
Crab	32	35.2	30	21.1	0	0	0	0	3	1

Crayfish	0	0	0	0	0	0	0	0	1	0.3
Insect	3	3.3	0	0	0	0	0	0	0	0
Reptile	3	3.3	0	0	1	1.37	0	0	0	0
Unknown	8	8.8	2	1.4	0	0	0	0	2	0.7
Unknown mammal	1	1.1	0	0	0	0	0	0	0	0
Totals	44	100	109	100	72	100	138	100	283	100

370

371 The precision (one standard deviation of standards) of the $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ reads was
372 0.097 and 0.080 respectively, n=34.

373

374 Niche breadth

375

376 Pterygoplichtys sp. was the main identifiable prey item in all samples from the
377 Usumacinta basin. Occurrence of ACF in scat samples was highest (49%) in samples
378 collected from Pasion River 7 years after the first report of the catfish, an increase from
379 9.9% in 2010 (Table 3). ACF occurrence also increased in the San Pedro River, but less than
380 in the Pasion River. *O. aureus* was an important item (percentage of occurrence > 5%) for
381 otters in the Pasion and San Pedro rivers in 2015 and the Mopan River in 2016 (Table 3).

382

383 Based on species accumulation curves, the expected number of prey species was
384 marginally lower in 2015 than in 2010 in Pasion River samples (Fig 2A); no difference was
385 seen for San Pedro River samples (Fig 2B). When all three rivers were compared based on
386 data from 2015-2016, otters from the San Pedro River were expected to have more prey
387 species, those from the Pasion River fewer species, and those from the Mopan River were
388 expected to have a middle number of prey species. Confidence intervals around expected
389 numbers were wide and overlapped, especially between curves from the Mopan River and
390 the other two rivers (Fig 2C). Further, the assumption that all samples used to construct
391 the accumulation curves were independent may have been violated because some of the
392 scats were collected from the same latrine.

393

394 **Fig 2.** Species accumulation curves for prey species found in scats of Neotropical river otter in the **(A)** Pasion
395 River, Guatemala 2010 (Pa10) and 2015 (Pa15); **(B)** San Pedro River, Guatemala, in 2009 (Sp09) and 2015
396 (Sp15); and **(C)** Mopan River 2016 (Mo16), Pasion River 2015 (Pa15) and San Pedro River 2015 (Sp15).

397

398 Niche breadth (Levin's index, B_a) of the Neotropical river otter was lower 7 years
399 after the ACF invasion when compared to 2 years after the invasion in the San Pedro River
400 ($B_a = 0.53$ in 2009 vs 0.29 in 2015). A similar situation was found in Pasion River ($B_a = 0.47$
401 in 2010 vs 0.18 in 2015). NRO niche breadth varied among the three rivers in 2015, with
402 similar values in San Pedro River and Mopan River and lower values in Pasion River ($B_a =$
403 0.29, 0.28 and 0.18 respectively; Table 4).

404

405 **Table 4** Neotropical river otter niche breadth (Levin's index, B_a) in the study area.

River	year	Ba	2.5% quantile	97.5% quantile
Mopan	2016	0.29	0.36	0.23
San Pedro	2009	0.53	0.6	0.5
San Pedro	2015	0.29	0.33	0.25
Pasion	2010	0.47	0.59	0.39
Pasion	2015	0.18	0.25	0.11

406 Quantiles estimated using 1,000 bootstrap randomizations.

407

408 Isotope values ranged from 5.89 to 16.39 $\delta^{15}\text{N}$ and -38.31 to -20.61 $\delta^{13}\text{C}$ (Fig 3) and
 409 did not depart from a normal distribution so no transformations were needed. Variance of
 410 $\delta^{15}\text{N}$ signatures from fecal samples differed among groups (Levene's test for
 411 homoscedasticity; $W = 2.54$, $p = 0.042$; Fig 4A). Based on pairwise comparisons, variance
 412 of $\delta^{15}\text{N}$ signatures from the Pasion River did not differ significantly between years ($\sigma^2 =$
 413 2.45 in 2010 and $\sigma^2 = 1.80$ in 2015; $W = 0.78$, $p = 0.37$; Fig 4A). In contrast, variance of $\delta^{15}\text{N}$
 414 differed significantly between years in samples from San Pedro River ($\sigma^2 = 4.83$ in 2009
 415 and $\sigma^2 = 1.73$ in 2015; $W = 6.68$, $p < 0.01$; Fig 4A). The $\delta^{13}\text{C}$ variances also differed among
 416 groups ($W = 3.23$, $p < 0.01$; Fig 4B), with pairwise contrasts indicating that $\delta^{13}\text{C}$ variances
 417 increased across years for Pasion River ($\sigma^2 = 3.65$ in 2010 and $\sigma^2 = 6.49$ in 2015; $W = 3.83$,
 418 $p = 0.05$; Fig 4B) and San Pedro River ($\sigma^2 = 2.04$ in 2009 and $\sigma^2 = 7.09$ in 2015; $W = 6.75$, p
 419 $= 0.01$; Fig 4B).

420

421 **Fig 3 Isotopic values of $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ from Neotropical river otter scats collected from the study area.**

422 Error bars are one sd. Mo15= samples from Mopan River 2015 (n=1); Pa10, Pa15 = samples from Pasion
 423 River 2010 and 2015 (n = 36 in 2010 and 34 in 2015); Sp09, Sp15 = samples from San Pedro River 2009 and
 424 2015 (n = 20 in 2010 and 55 in 2015).

425

426 **Fig 4 Isotopes Variance from the mean in fecal samples from Neotropical river otters for (A) $\delta^{15}\text{N}$ and (B)**

427 **$\delta^{13}\text{C}$.** The mean is set to 0 to help visualize the magnitude of the variances. Pasion River n = 36 in 2010 and
 428 34 in 2015; San Pedro River n = 20 in 2010 and 55 in 2015.

429

430 Trophic level

431

432 Calculations of FTL values excluded information from *Maskaheros argenteus*
 433 (found in one sample from the Pasion River), insects, reptiles and unknown species
 434 because no data on the FTL of those prey items were available. Similarly, crabs and
 435 crayfish were excluded because they consume items from different trophic levels and
 436 their specific diets are not known for the study area, although crabs were important prey
 437 items in Mopan 2016 and Pasion 2010. The highest FTL values for NRO came from the
 438 Mopan River in 2016, Pasion River in 2010, and San Pedro River in 2009 with lower values
 439 from the Pasion and San Pedro rivers in 2015 (Table 5).

440

441 **Table 5 Neotropical river otter fractional trophic level (FTL) in the study area.**

River	year	FTL	2.5% quantile	97.5% quantile
Mopan	2016	3.98	4.11	3.83
San Pedro	2009	3.71	3.79	3.64

San Pedro	2015	3.47	3.53	3.41
Pasion	2010	3.78	3.9	3.68
Pasion	2015	3.48	3.61	3.37

442 Quantiles estimation using 1,000 bootstrap randomizations.

443

444 Values of $\delta^{15}\text{N}$ from NRO samples were highest in the Mopan River in 2015 (based
445 on only one specimen), followed by mean values from the Pasion River in 2010 and the
446 San Pedro River in 2009 (Fig 5). Lowest values came from the Pasion and San Pedro rivers
447 in 2015 (Fig 5). Values of $\delta^{15}\text{N}$ from sites in the Usumacinta basin differed across years
448 (ANOVA, $F_{1,141} = 67.98$; $p < 0.001$) and across rivers (ANOVA, $F_{1,141} = 15.53$; $p < 0.001$) with
449 no interaction between the two factors (ANOVA, $F_{1,141} = 2.76$; $p = 0.10$). Higher values
450 were found from scats collected during the early sampling years in the Pasion and San
451 Pedro rivers, two years after the first report of the ACF (post-hoc pairwise t -test with
452 Bonferroni adjusted p -values: Pasion 2010 vs. 2015 $t = 5.37$, $df = 68$, $p < 0.001$; San Pedro
453 2009 vs. 2015, $t = 5.31$, $df = 24.122$, $p < 0.001$). Mean values of NRO $\delta^{15}\text{N}$ did not differ
454 between the Pasion and San Pedro rivers from same sampling years (Pasion vs. San Pedro
455 2010-2009 $t = -0.40$, $df = 54$, $p = 1.0$; Pasion vs. San Pedro 2015, $t = 2.42$, $df = 87$ $p =$
456 0.23).

457

458 **Fig. 5. Boxplots for $\delta^{15}\text{N}$ in fecal samples from Neotropical river otters in Guatemala.** Mo16 = samples from
459 Mopan River 2016 ($n = 1$); Pa10, Pa15 = samples from Pasion River 2010 and 2015 ($n = 36$ in 2010 and $n = 34$
460 in 2015); Sp09, Sp15 = samples from San Pedro River 2009 and 2015 ($n = 20$ in 2009 and $n = 55$ in 2015).

461

462 Discussion

463 Concordance between the gross scat analysis and stable isotope analysis values
464 strongly supports the idea that an increase in consumption of the armored catfish
465 reduced the dietary niche breadth of the neotropical river otter and trophic level at which
466 the otter feeds in northern Guatemala. As predicted, ACF became the main prey species
467 for the NRO in invaded rivers and, consequently, NRO $\delta^{15}\text{N}$ variances and mean values
468 decreased over time in both invaded rivers (with a weaker decline in Pasion River). The
469 same pattern was observed in the standardized niche breadth index (B_a). Further, the
470 wider niche breadth (B_a values) in the San Pedro River may be related to its higher
471 environmental integrity (located adjacent to a national park) that could help sustain the
472 richness of native NRO prey or reduce the invasiveness of the ACF. This conclusion is
473 supported by the species accumulation curves. Invasive species are predicted to have
474 better chances of establishment in native assemblages that are depleted or disrupted and
475 more likely to have long-term success in systems highly altered by human activity [81].
476 The increase in $\delta^{13}\text{C}$ variation over time suggests that the NRO diet has included a prey
477 that consumes different producer types or a prey that consumes producers in a different
478 proportion, likely because of the ability of ACF to exploit a different range/proportion of
479 plant resources than natives from the same trophic guild [82]. Furthermore, the decrease
480 in FTL across rivers (Mopan River showing similar values to San Pedro River and higher

481 than Pasion River) combined with lower mean values of $\delta^{15}\text{N}$ provide evidence of a
482 reduction in the NRO trophic level associated with ACF presence.

483 The range of prey types exploited by NRO changed after the invasion of ACF, with
484 the lowest dietary niche breadth found in Pasion River seven years after the invasion. The
485 dietary NRO niche breadth changed from that of a mild generalist to that of a specialist
486 ($0.6 > B_a > 0.4$ to $B_a < 0.4$, Levin's standardized index) in Pasion and San Pedro rivers, even
487 though the number of prey species consumed by NRO was highest in the San Pedro River
488 in 2015. This result is concordant with the idea that a specialist can use a wide range of
489 resources but still concentrate on a subset of those resources [83]. It also supports the
490 statement that NRO prey mostly on slow-moving and territorial prey species [36]; the
491 main prey species for NRO in this study included Loricariidae, Cichlidae, large Poeciliidae,
492 and crabs (Table 3).

493
494 Results based on GSA and $\delta^{15}\text{N}$ variances were similar for both indexes, with
495 narrower niche breadth 7 years after initiation of the ACF invasion compared to 2 years
496 after the invasion. The narrower dietary niche breath found in the Pasion River in all
497 situations and with both indexes in relation to the San Pedro River supports the idea that
498 the Pasion River prey community was already depleted before the arrival of the ACF, and
499 that the Laguna del Tigre National Park provided some type of protection to the San Pedro
500 River. A similar result was seen in a Bahamas mangrove system for grey snapper (*Lutjanus*
501 *griseus*) with a reduced niche breadth (based on SIA) found in disturbed areas [5].
502 Therefore, it is possible that the higher values of NRO niche breadth in San Pedro River in
503 relation to Pasion River are related to differences in the resilience of the two rivers due to
504 differences in habitat conservation. Disturbances may facilitate the ACF or depress
505 populations of native fish. For example, in the Guadalquivir marshes of southwestern
506 Spain, the Eurasian otter (*Lutra lutra*) included high levels of an invasive species (75%;
507 North American red swamp crayfish, *Procambarus clarkii*) in its diet within 10 years of the
508 invasion. In the same area, various waterbirds similarly consumed this invasive species at
509 higher rates in disturbed locations than in natural marshes [1].

510
511 In contrast to results from $\delta^{15}\text{N}$, variances of $\delta^{13}\text{C}$ in fecal samples were greater
512 seven years after the ACF invasion compared to two years after the first sighting. Values of
513 $\delta^{13}\text{C}$ represent the plant source of a food chain and a wider variance may indicate that
514 primary consumers exploit a greater range of producers. Loricariidae may exploit a diverse
515 variety of basal sources or a portion that the natives does not exploit, which may help
516 explain the increase in the variance of $\delta^{13}\text{C}$ in NRO scats, given the increased presence of
517 ACF in the NRO diet [84].

518
519 Native predators may act to reduce invasive species numbers [85,86], and such
520 predation could be one of the main biological drivers by which streams resist the invasion
521 of exotic species [87]. Further, predators from different taxa often adapt to and benefit
522 from the consumption of invasive species [3,4,88]. In this context, NRO may act as a buffer
523 to hold ACF populations at low levels and minimize their potential negative effects on the
524 system. The question that arises from this situation, as in other systems where an invasive

525 species becomes the main prey of a native predator [1], is whether the consumption of
526 ACF by NRO and other native predators can facilitate the predators [89]. Greater predator
527 populations might increase depredation on native prey that are threatened by
528 overexploitation or habitat loss [90]. This effect is a valid concern in our study area, where
529 cichlids, a group of fish that is highly appreciated by the local artisanal fisheries [91] were
530 exploited as a group without much change when the consumption of ACF increased (Table
531 3). Also, concern for the increase of negative interactions between native predator and
532 humans becomes relevant when wild predators establish dense populations in or near
533 human-dominated areas [92,93].

534

535 Both GSA and $\delta^{15}\text{N}$ values indicated a reduction in the trophic level at which otters
536 feed in rivers where ACF are present in northern Guatemala. Based on GSA, there were
537 reductions in the FTL of NRO of approximately 0.33 FTL in the Pasion River and 0.2 FTL in
538 the San Pedro River. These reductions may not represent much ecological difference. GSA
539 may, however, under-estimate the consumption of some species and over-estimate the
540 consumption of others either because of differences in digestibility of prey or because we
541 measured presence of prey remains rather than consumed biomass, regardless of the
542 amount of remains (not all remains were identifiable; e.g., spines). In contrast to GSA, SIA
543 may give a more accurate result. Differences in mean $\delta^{15}\text{N}$ were as great as 1.88‰ for
544 Pasion River and 2.78‰ for San Pedro River. If we use the widely accepted 3.4‰
545 enrichment ($\Delta^{15}\text{N}$) per FTL, these differences in mean $\delta^{15}\text{N}$ may represent changes of 0.5
546 to 0.8 FTLs in the Pasion and San Pedro rivers, respectively. The 3.4‰ $\Delta^{15}\text{N}$ value has,
547 however, been criticized. Models and empirical data have shown that this enrichment
548 factor can underestimate FTL of marine predators [94]. In any case, the observed mean
549 $\delta^{15}\text{N}$ values for NRO in both the Pasion and San Pedro rivers apparently represents a
550 decrease in trophic level.

551

552 A reduction in the trophic level at which otters feed can have diverse effects on the
553 riverine ecosystem. These effects may be difficult to anticipate and can compete with or
554 interact with each other. It could mean predator release for other prey species that would
555 benefit from reduced predation pressure [95,96]. On the other hand, consumption of the
556 invasive species may benefit the predator, eventually leading to higher predator densities
557 that could increase pressure on other native species. A model evaluating this situation
558 suggests that predation on native prey by a native predator whose numbers have been
559 enhanced by consumption of an invasive species can be more harmful than direct
560 competition between native and invasive species [97]. Empirical data using SIA for golden
561 eagles (*Aquila chrysaetos*) suggests that these eagles colonized the California Channel
562 Islands after the introduction of feral pigs (*Sus scrofa*) [90]. Nonetheless, eagles still
563 preyed on endemic meso-carnivores, including a fox (*Urocyon littoralis*) and skunk
564 (*Spilogale gracilis amphiala*), pushing the fox towards extinction [90].

565

566 Another potential effect that needs to be evaluated is the reduction of trophic
567 levels in the system by moving energy directly from primary consumers to top predators.
568 This results can occur by eliminating food-web links in the mid-trophic levels through

569 competition or predation facilitated by a numerical response of predators in
570 response to the high abundance of the invasive [1]. A similar situation was found in
571 the United Kingdom, where researchers compared the fish assemblage in a pond
572 with a low-trophic-level invasive cyprinid (*Pseudorasbora parva*) composing > 99%
573 of fish present to that in another pond without the cyprinid. They reported a
574 reduction in the $\delta^{15}\text{N}$ values of piscivorous fish and a mean reduction in the $\delta^{15}\text{N}$ of
575 the complete fish community [98]. Further studies are needed to investigate the
576 effect of different types of land management, as well as factors that indicate the
577 ecological integrity of communities, on the ability of communities to resist or
578 facilitate the invasion of exotic species and their interactions with native predators.
579

580 Acknowledgments

581 We wish to acknowledge the institutions and all the people who helped us through those
582 institutions to make the fieldwork and laboratory work possible: Eli.S.A., WCS-Guatemala,
583 BALAM, CONAP-Peten, Propeten, Defensores de la Naturaleza Peten, FUNDAECO, CECON,
584 the Univeridad de San Carlos de Guatemala Escuela de Biología, the University of Florida
585 Departement of Geological Sciences Light Stable Isotopes Mass Spectrometry laboratory,
586 Roan Balas, Byron Castellanos, Yobany Tut, Julio Madrid, Rony Garcia, Rafael Cevallos,
587 Mercedes Barrios, Rosalito Barrios, Manuel Lepe, Werner Paz, Silja Ramirez, Leonel Zisse,
588 Marta Pujol and Jason Curtis. The people who helped in the data gathering: Alejandro
589 Chen, Alejandro Mérida, Alfredo Choc, Belarmino García, Carlos Cifuentes, Dastin Ramírez,
590 Elder Godoy, Elmer Monzón, Ervin Flores, Fredy Tot, Jeovany Nolberto Tut Pacheco, René
591 de Jesús Mauricio Te, Ricardo Coc Caal, Samuel Yatz, Juan Rodas but specially to Andrea
592 Paiz and Francisco Cordova who were present during most of the sampling occasions in
593 2015-2016 and Yasmín Quintana in 2010. Also, to the people whose comments, reviews,
594 and advice helped to improve this document: Christina Romagosa, Bill Pine, Harry Jones,
595 Laura Gelin, Jose Soto, Claudio Moraga, Audrey Wilson, Yasmín Quintana, Flavia Montalvo,
596 Farah Carrasco, and Jason Curtis.

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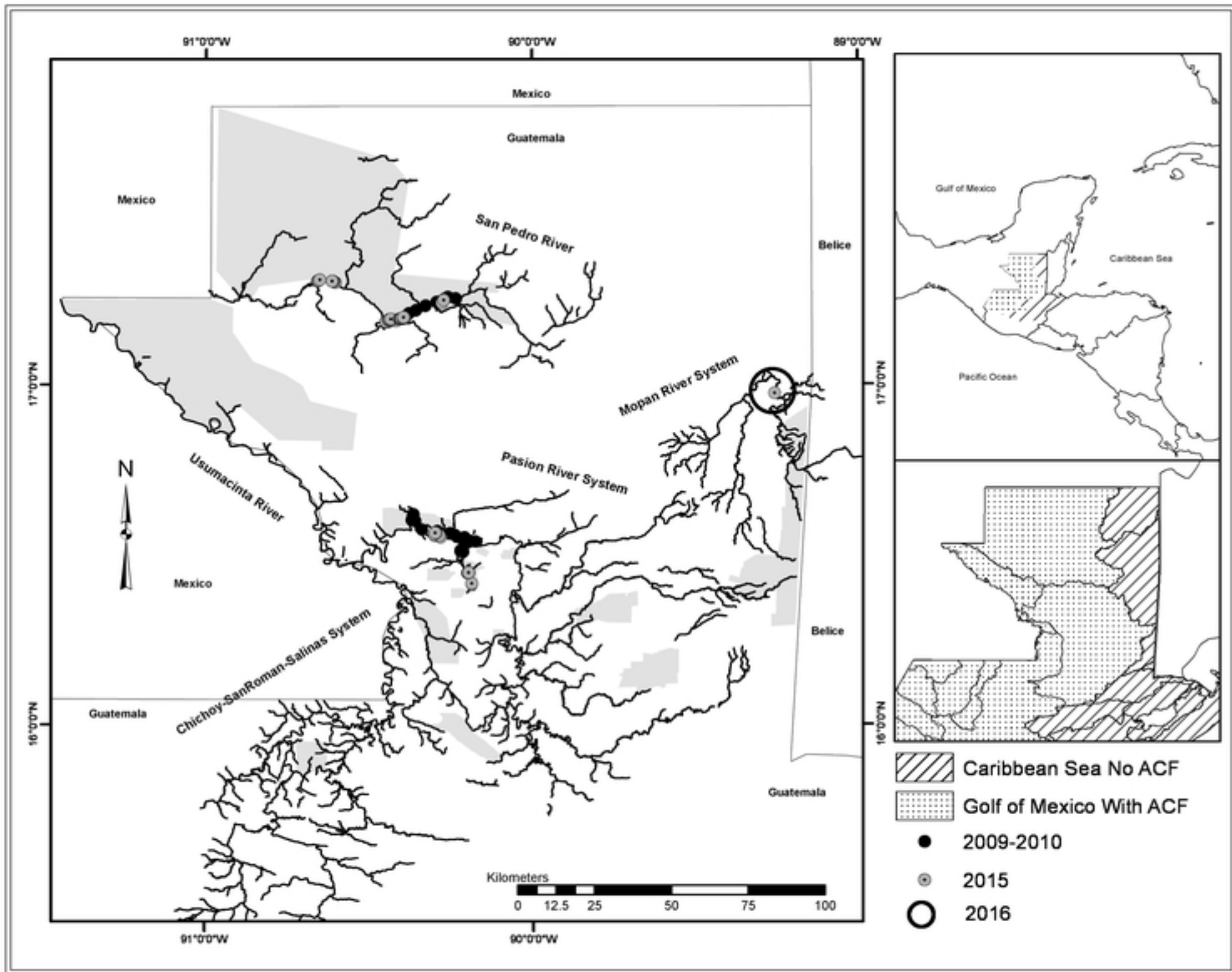
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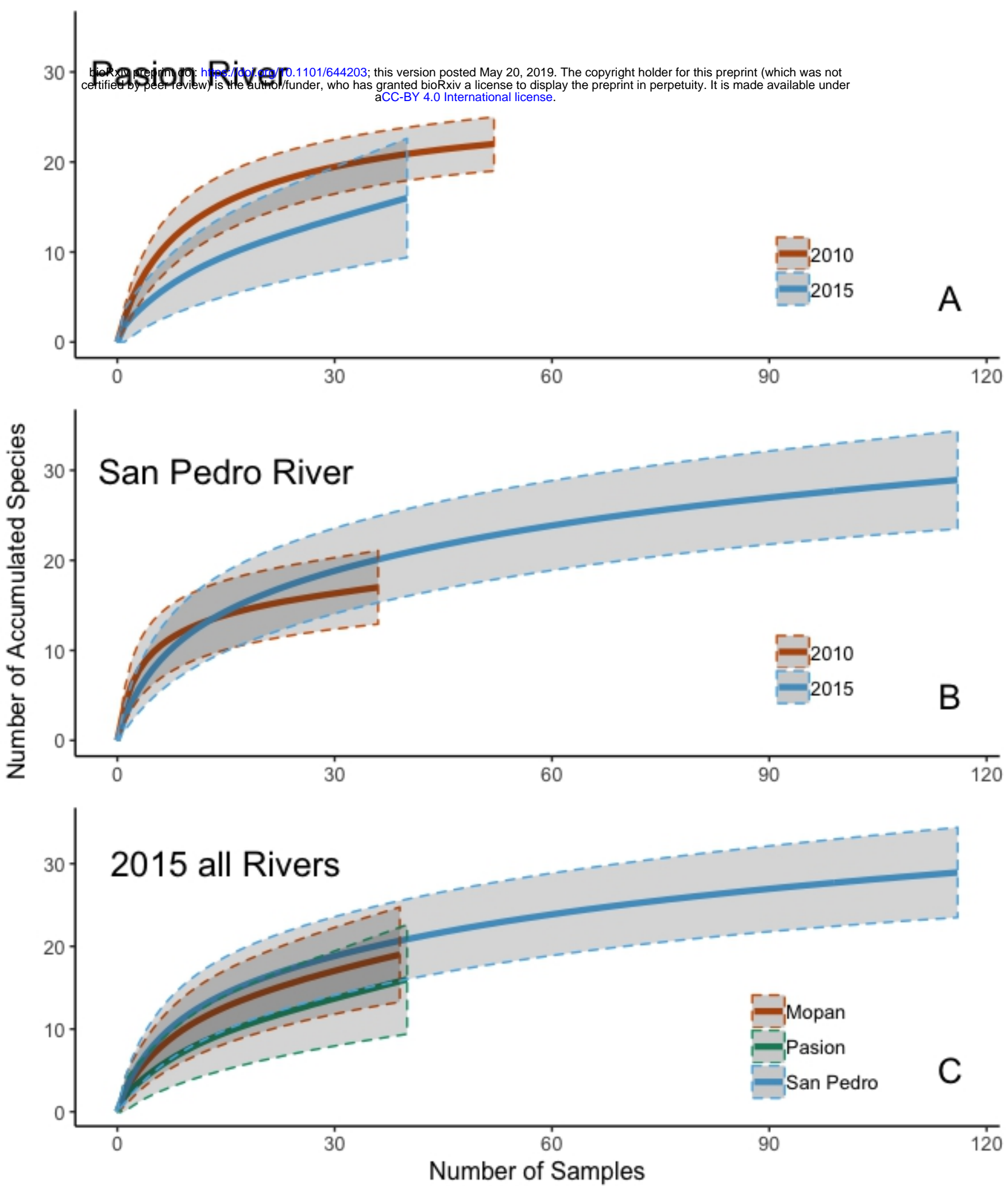
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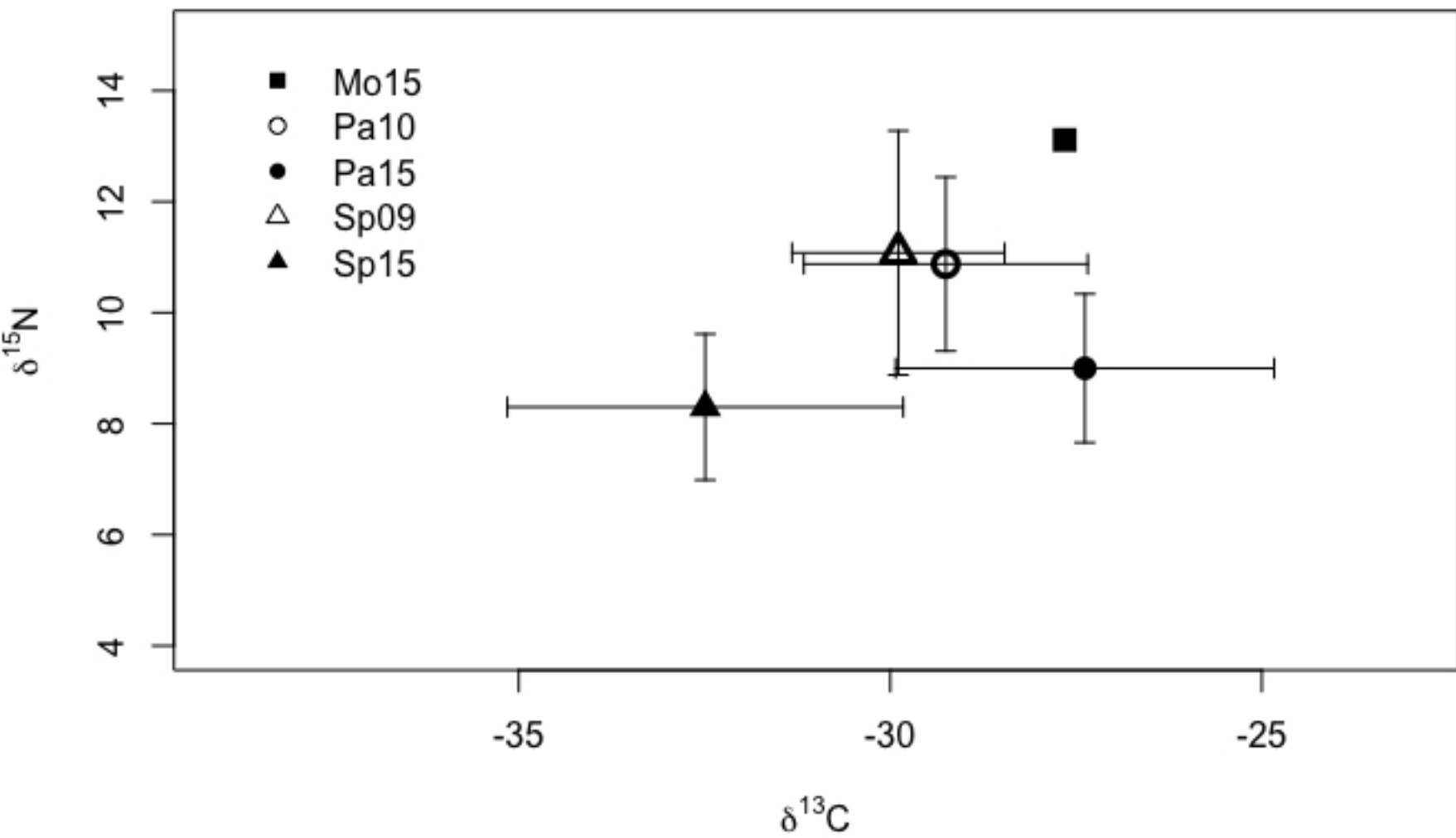
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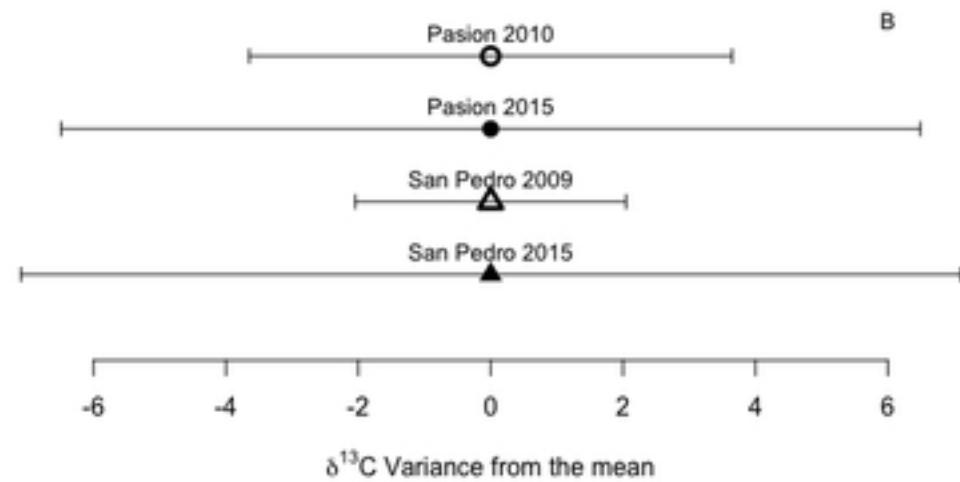
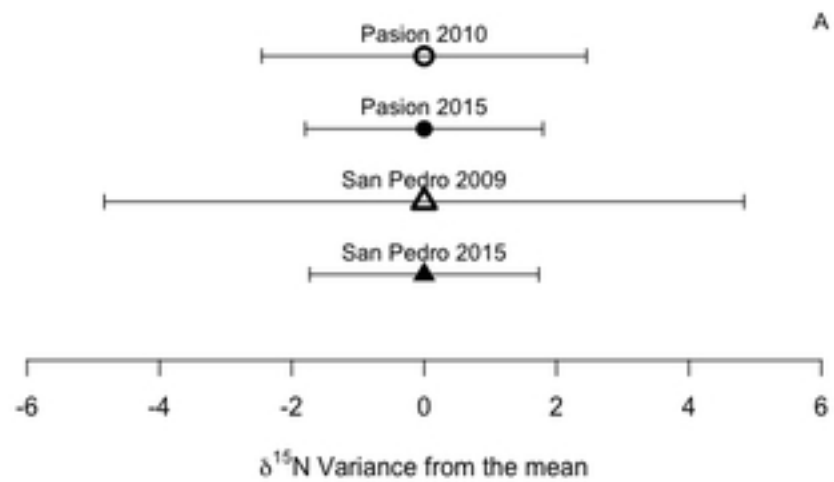
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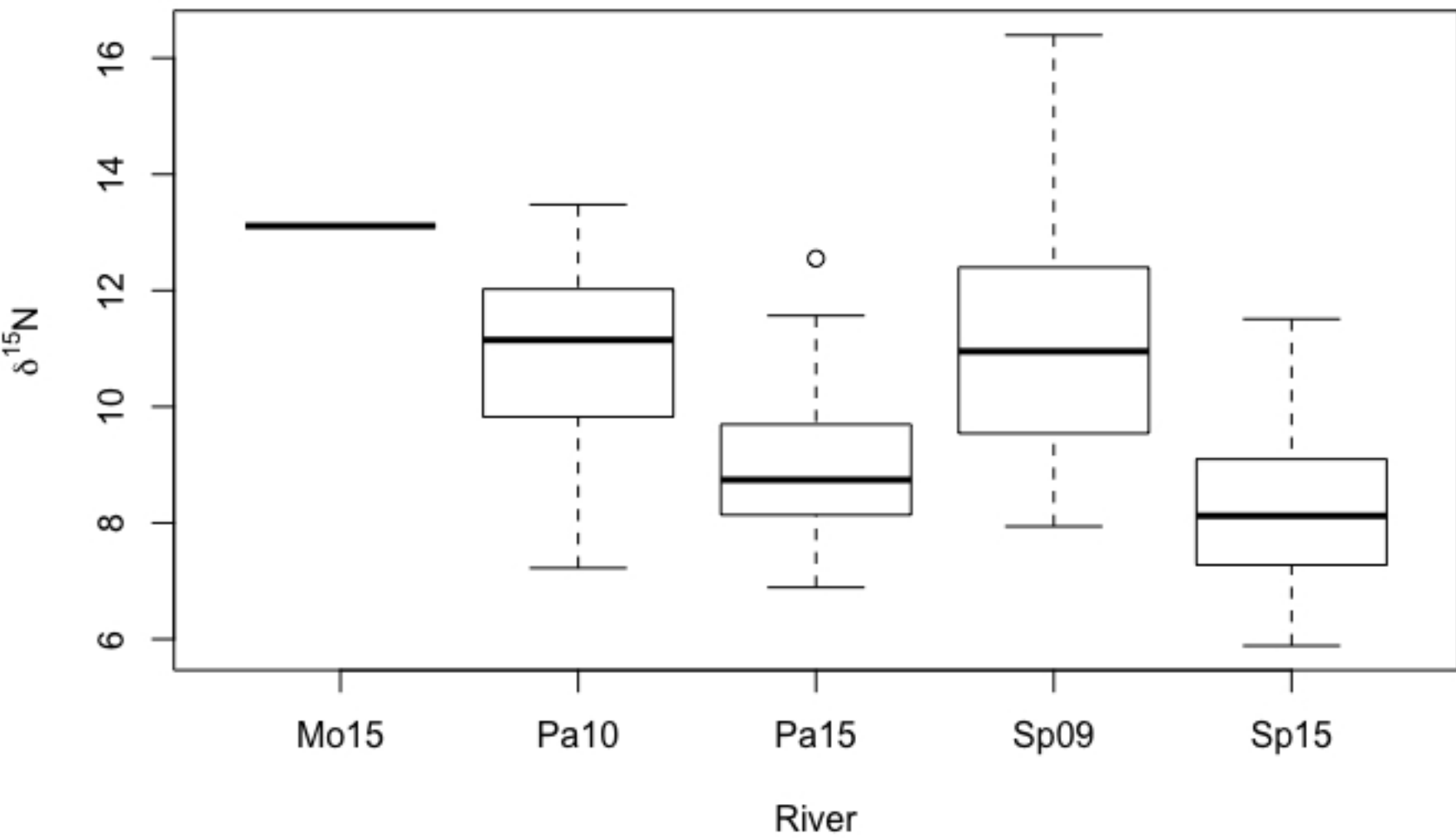
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