

25 **Introduction**

26 Genome-wide association studies (GWAS) aim to identify single nucleotide
27 polymorphisms (SNPs) whose allelic variation is significantly tied to phenotypic variation. In
28 principle, the tie between the allelic variation and phenotypic variation can be measured based on
29 the variance among the phenotypic averages for all scores per each SNP (Kim, 2017; Kim, 2018a).
30 Greater variance indicates a stronger tie. Conventional GWAS practice has been largely conducted
31 using statistical methods such as the linear model and the linear mixed model (LMM). To date, the
32 use of the LMM has been widely encouraged because of the general perception that accounting
33 for a kinship matrix can reduce the noise between a phenotypic variable and an SNP variable, by
34 correcting the bias that genetic relationship among entities in a population introduces (Yu et al,
35 2006; Bradbury et al, 2007; Kang et al, 2008; Lipka et al, 2012; Hoffman, 2013; Kim et al, 2018b).
36 Recently, however, Kim (2019) demonstrated that the use of a kinship matrix actually makes the
37 LMM unreliable. In this regard, this study excluded the LMM.

38 Conventional GWAS practice based on the linear model often regresses each SNP variable
39 along with population-structure variables against a phenotypic variable, one by one across all SNPs.
40 Therein, the use of population-structure variables aims to obtain an adjusted phenotype calculated
41 by subtracting the estimated population-structure effect from the phenotype (Yu et al, 2006;
42 Bradbury et al, 2007; Kang et al, 2008; Lipka et al, 2012; Hoffman, 2013; Kim et al, 2018b). For
43 reliable GWAS practice, it is crucial to assure the adjusted phenotypes resulting across all SNPs
44 are consistent. Otherwise, every SNP variable will be regressed against a differently adjusted
45 phenotypic variable, which consequently confounds GWAS results. This study investigated
46 whether accounting for population structure in the linear model for GWAS assures the adjusted
47 phenotypes resulting across all SNPs to be consistent

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52 **Materials and Methods**

53 **Rice data set**

54 This study used a rice data set comprising SNP data, principle component analysis (PCA) data and
55 phenotypic data. The data set was originally used for GWAS by Zhao et al. (2011) and freely
56 available to public at <http://ricediversity.org/data/index.cfm>. Therefore, more information about the
57 data set can be found from the related paper. In the original data, 413 entities were genotyped with
58 36,901 SNPs. The number of SNPs was reduced to 12,983 by screening with a criterion of the
59 minor allele frequency (MAF) of 0.1. The phenotype chosen for this study was seed length.

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61 **Statistical model**

62 The two linear models were established as follows:

$$63 \quad y = \mu + \beta_1 x_{SNP} + \varepsilon \quad (1)$$

$$64 \quad y = \mu + \beta_1 x_{SNP} + \beta_2 x_{PCA1} + \beta_3 x_{PCA2} + \beta_4 x_{PCA3} + \beta_5 x_{PCA4} + \varepsilon \quad (2)$$

65 where y = the phenotypic observation; μ = the phenotypic mean; x_{SNP} = the SNP variable; x_{PCA1}
66 = the PCA1 variable; x_{PCA2} = the PCA2 variable; x_{PCA3} = the PCA3 variable; x_{PCA4} = the PCA4
67 variable; ε = the error term; β_1 = the coefficient for x_{SNP} ; β_2 = the coefficient for x_{PCA1} ; β_3 = the
68 coefficient for x_{PCA2} ; β_4 = the coefficient for x_{PCA3} ; β_5 = the coefficient for x_{PCA4} .

69 Equation 1 regresses the SNP variable against the phenotypic variable. Equation 2 regresses the
70 SNP variable along with the four PCA variables (x_{PCA1} , x_{PCA2} , x_{PCA3} , x_{PCA4}) against the
71 phenotypic variable. This means that Equation 2 regresses the SNP variable against the adjusted
72 phenotypic variable obtained by accounting for the four PCA variables. Equation 3 highlights the
73 adjusted phenotypic variable:

$$74 \quad y - \beta_2 x_{PCA1} - \beta_3 x_{PCA2} - \beta_4 x_{PCA3} - \beta_5 x_{PCA4} = \mu + \beta_1 x_{SNP} + \varepsilon \quad (3)$$

75 Equation 3 is compatible with Equation 2 and represents the adjusted phenotypic variable as $y -$
76 $\beta_2 x_{PCA1} - \beta_3 x_{PCA2} - \beta_4 x_{PCA3} - \beta_5 x_{PCA4}$.

77 **Manhattan plot**

78 The F test was implemented as a significance test, from which P values were obtained. The P
79 values transformed by $-\log_{10}$ were drawn in a Manhattan plot. It is important to note that the P
80 values resulting from the linear model for GWAS are prone to genomic inflation. Prior to
81 confirming the resultant Manhattan plot, therefore, it is necessary to calculate the genomic inflation
82 factor (λ_{GC}). The situation of $\lambda_{GC} > 1$ indicates the genomic inflation, which means that the
83 resultant P values are overly estimated compared with the χ^2 -distribution (van Iterson et al, 2017).
84 This study adjusted the genomic inflation using the genomic control. More information about the
85 genomic control can be found in previous studies (Devlin and Roeder, 1999; Yang et al, 2011; van
86 Iterson et al, 2017).

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88 **Integrity validation of accounting for population structure in GWAS**

89 Equation 3 (compatible with Equation 2) regresses each SNP variable against an adjusted
90 phenotypic variable. As GWAS handle numerous SNPs one by one at a time, it is important to
91 assure that the adjusted phenotypes resulting across all SNPs are consistent. Otherwise, each SNP
92 variable will be regressed against a differently adjusted phenotypic variable. The consistency
93 among the adjusted phenotypes resulting across all SNPs can be achieved, only if every coefficient
94 per each PCA variable is consistent across all SNPs. To check the consistency among the adjusted
95 phenotypes resulting across all SNPs, this study calculated Pearson coefficients between the
96 phenotype and every adjusted phenotype.

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98 **Data set and R code**

99 All computations were conducted using R (R Core Team, 2016). The data set and R scripts used
100 in this study are freely available at <https://github.com/bongsongkim/Population.Structure.GWAS>.

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103 Results

104 Validation of consistency across all adjusted phenotypes

105 Table 1 summarizes the coefficients per each PCA variable, resulting from applying all
106 SNPs to Equation 3. Figure 1 represents the estimated coefficients per each PCA variable, showing
107 large variation. Figure 2 represents the estimated Pearson correlation coefficients between the
108 phenotype and every adjusted phenotype, illustrating the adjusted phenotypes resulting across all
109 SNPs are not consistent. This means that each SNP variable is regressed against a differently
110 adjusted phenotypic variable.

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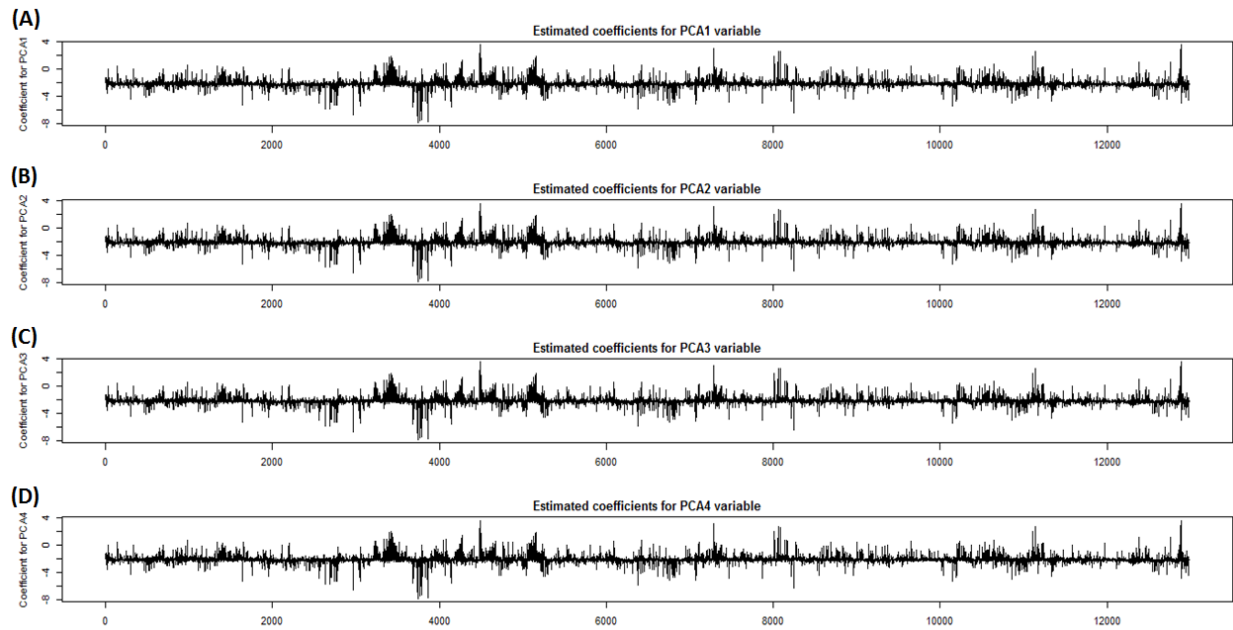
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113 **Table 1.** Summary of coefficients per each PCA variable in relation to Equation 3.

	Min.	1 st Qu.	Median	Mean	3 rd Qu.	Max
β_2	-7.833	-2.246	-2.162	-2.109	-2.024	3.629
β_3	-4.438	-1.097	-1.020	-1.007	-0.944	3.271
β_4	-13.610	-9.229	-9.194	-9.180	-9.160	0.659
β_5	-9.308	3.016	3.087	3.025	3.121	8.704

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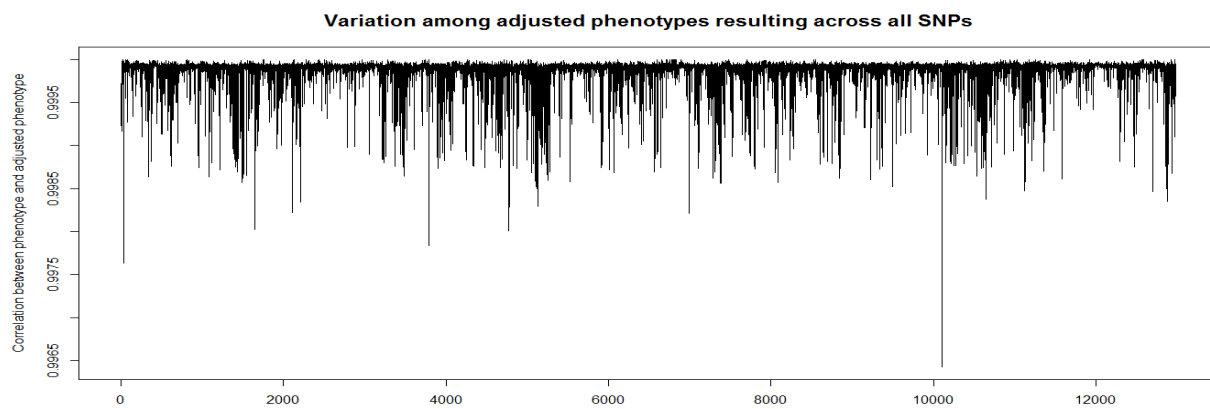
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117 **Figure 1.** (A) Estimated coefficients for the PCA1 variable, (B) estimated coefficients for the
118 PCA2 variable, (C) estimated coefficients for the PCA3 variable, (D) estimated coefficients for
119 the PCA4 variable.

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122 **Figure 2.** Pearson correlation coefficients between the phenotype and every adjusted phenotype.

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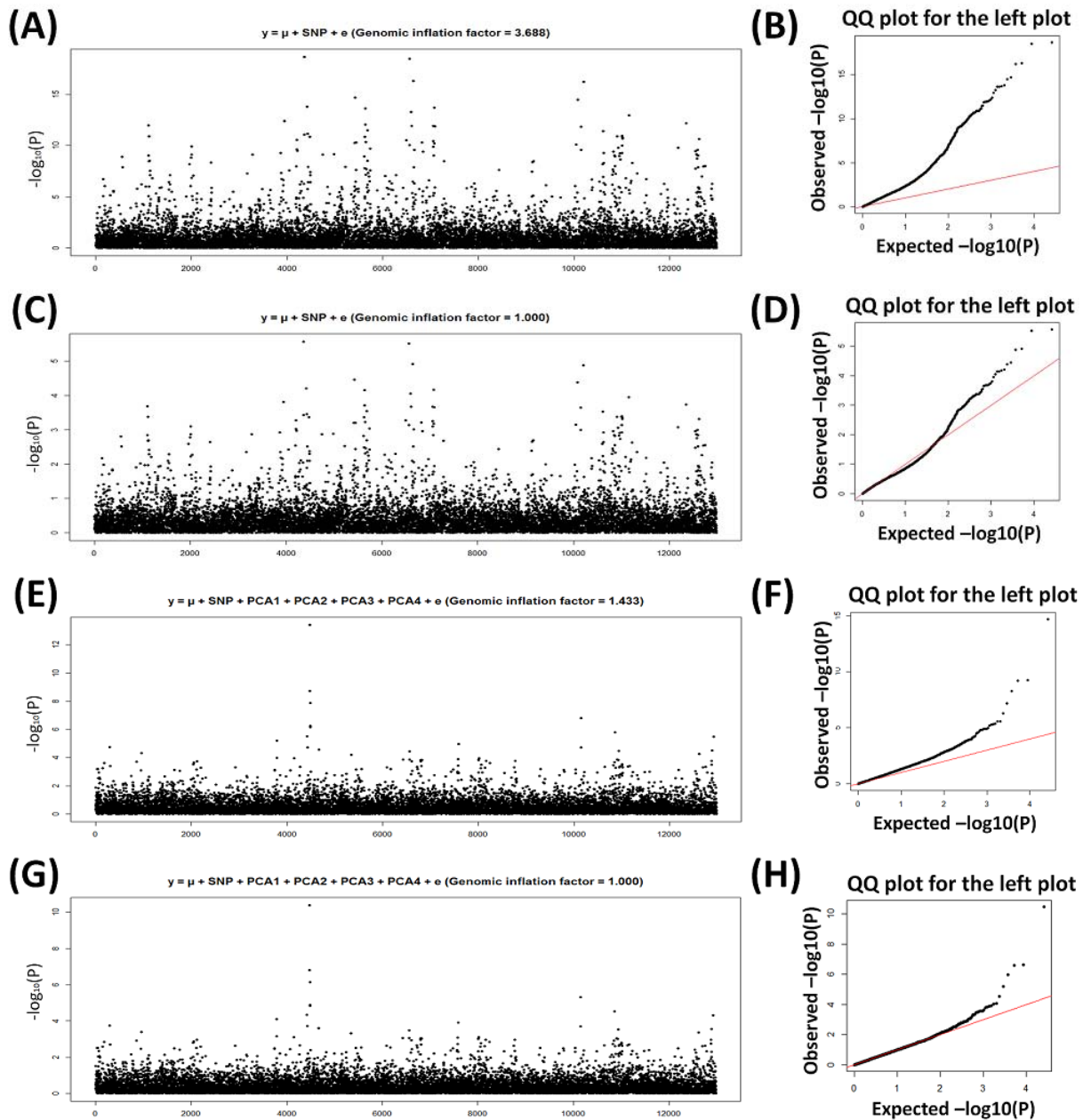
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125 **Impact of accounting for population structure in GWAS**

126 Figure 3 shows four Manhattan plots, for which the same SNP and phenotypic data were used.
127 Figure 3A represents the Manhattan plot in relation to Equation 1, in which the resultant λ_{GC} was
128 3.688. Figure 3C is the same as Figure 3A in shape. However, Figure 3C meets $\lambda_{GC} = 1$ by
129 implementing the genomic control with Figure 3A. Figure 3E represents the Manhattan plot in
130 relation to Equation 3, in which the resultant λ_{GC} was 1.433. Compared with Figure 3A, Figure 3E
131 has substantially lower λ_{GC} . This suggests that accounting for the four PCA variables was
132 impactful in diminishing the genomic inflation. Figure 3G was obtained by adjusting Figure 3E by
133 implementing the genomic control. This led to $\lambda_{GC} = 1$ in Figure 3G. It is apparent that Figure 3E
134 has clearer background than Figure 3A in relation to accounting for the four PCA variables. In this
135 regard, previous studies explained that accounting for population structure in the linear model for
136 GWAS eliminates the noise in SNP-phenotype associations, which results in clear background in
137 a resultant Manhattan plot (Yu et al, 2006; Kang et al, 2008; Korte and Farlow, 2013; Sul et al,
138 2018; Barton et al, 2019). However, Figure 4 illustrates that significant SNP-phenotype
139 associations are not consistent between Figures 3C and 3G. This means that the clear background
140 was not from eliminating the noise in SNP-phenotype associations, but from defining new SNP-
141 phenotype associations.

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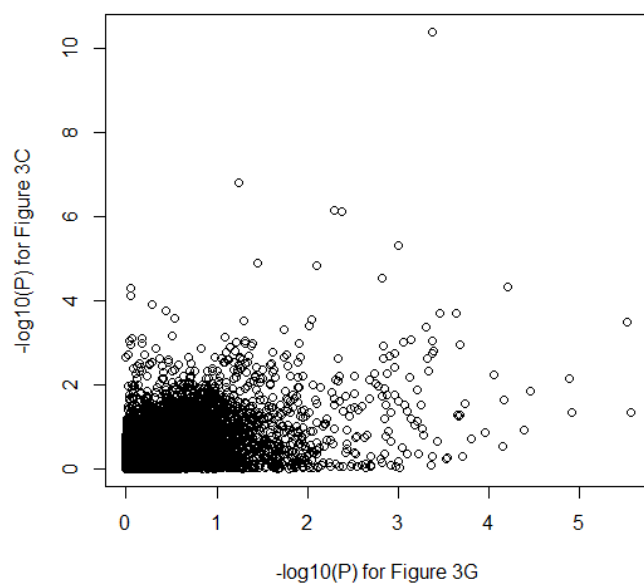
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145 **Figure 3.** (A) Manhattan plot obtained by not accounting for the four PCA variables ($\lambda_{GC} = 3.688$),
146 (C) Manhattan plot obtained by adjusting Figure 3A with implementing the genomic control (λ_{GC}
147 = 1.000), (E) Manhattan plot obtained by accounting for the four PCA variables ($\lambda_{GC} = 1.433$), (G)
148 Manhattan plot obtained by adjusting Figure 3C with implementing the genomic control ($\lambda_{GC} =$
149 1.000).

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152 **Figure 4.** Correlation plot between the $-\log_{10}(P)$ values obtained by not accounting for the four
153 PCA variables (Figure 3C) and the $-\log_{10}(P)$ values obtained by accounting for the four PCA
154 variables (Figure 3G).

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164 **Discussion**

165 It is generally perceived that accounting for population structure in GWAS improves the
166 quality of visual representation of a Manhattan plot by both suppressing genomic inflation and
167 reducing false-positive SNP-phenotype associations (Yu et al, 2006; Bradbury et al, 2007; Kang
168 et al, 2008; Lipka et al, 2012; Hoffman, 2013; Kim et al, 2018b). In fact, this study showed that
169 accounting for the four PCA variables was very effective in diminishing the genomic inflation.
170 Surprisingly, however, this study revealed that accounting for the four PCA variables breaks the
171 consistency among the adjusted phenotypes resulting across all SNPs. The loss of the consistency
172 consequently causes each SNP variable to be regressed against a differently adjusted variable,
173 making the GWAS process unreliable. The use of population-structure variables in the linear
174 model for GWAS implies two errors. First, the linear model is misused. Considering that the linear
175 model is suited for analyzing data in experimental blocks, the use of continuous variables rather
176 than factor variables necessarily causes an error. Second, the assumption for the relationship
177 between phenotype and population structure is unjustified. The linear model for GWAS generally
178 assumes that the population-structure variables additively contribute to the phenotypic variable.
179 However, how the population structure biologically influences the phenotype has yet been
180 unknown. Regardless of whether the additivity of the population-structure variables is true or false,
181 the current way of accounting for population structure is inappropriate in that population-structure
182 effects vary across all SNPs. The abovementioned errors consequently lead to the loss of the
183 consistency among the adjusted phenotypes resulting across all SNPs and cause each SNP variable
184 to be regressed against a differently adjusted phenotypic variable.

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186 **Conclusion**

187 The linear model assures to preserve the consistency among the adjusted phenotypes resulting
188 across all SNPs, only if factor variables such as years, locations, replications and treatments are
189 used. This study concluded that the conventional way of accounting for population structure makes
190 the GWAS process unreliable. This is because the population structure is represented as continuous
191 variables. If population structure can be represented as factor variables, accounting for the
192 population structure in the linear model for GWAS will be sound.

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