mPFC Spindle Cycles Organize Sparse Thalamic Activation and Recently Active CA1 cells During non-REM Sleep

Varela C[1,2], Wilson MA[1].

[1]Massachusetts Institute of Technology, Cambridge, MA.

[2]Florida Atlantic University, Boca Raton, FL.

Abstract

Sleep oscillations in neocortex and hippocampus are critical for the integration of new episodic memories into stable generalized representations in neocortex. However, the role of the thalamus in this process is poorly understood.

To determine the thalamic contribution to non-REM oscillations (sharp-wave ripples, SWRs; slow/delta; spindles), we recorded units and local field potentials (LFPs) simultaneously in the limbic thalamus, mPFC, and CA1 in rats. We report that the cycles of neocortical spindles provide a key temporal window that coordinates CA1 SWRs with sparse but consistent activation of thalamic units. Thalamic units were phase-locked to delta and spindles in mPFC, and fired at consistent lags with other thalamic units within spindles, while CA1 units that were active during spatial exploration were engaged in SWR-coupled spindles after behavior. The sparse thalamic firing could promote an incremental integration of recently acquired memory traces into neocortical schemas through the interleaved activation of thalamocortical cells.

Introduction

Two complementary learning systems acquire and process episodic memories in order to learn adaptive models of the world that generalize beyond specific experiences and continue to incorporate information through the lifetime of an animal[1–4]. The hippocampus can form memory traces rapidly; however, over time, interactions between the hippocampus and neocortex are thought to extract statistical regularities from the organism's daily experiences, and update neocortical models or schemas to increase their adaptive value[5,6]. During non-REM sleep, the interactions between the hippocampal and neocortical cell ensembles that underlie these processes are reflected in nested oscillations in the local field potentials (LFPs). The up-states of the slow oscillation (1–4 Hz) provide a processing window in which faster dynamics lead to synaptic plasticity in relevant cell populations. In the neocortex, some cells are preferentially active at the start or at the end of up-states[7,8], and in the hippocampal CA1 region, sharp-wave ripple oscillations (SWRs, 100–275 Hz) occur primarily during up-states[9]. Hippocampal SWRs are brief (up to ~100 ms) oscillations that correlate with the reactivation of spatial representations by hippocampal cell ensembles[10,11] which are thought to contribute the content in the consolidation of episodic memory[12–14]. Spindle oscillations (6–14 Hz) are coordinated with the slow oscillation and with SWRs[15–17]. Spindles are generated in the thalamus when the inhibitory cells of the thalamic reticular nucleus (TRN) evoke inhibitory post-synaptic potentials and rebound bursting in cells of the dorsal thalamus, which then entrain their postsynaptic targets in neocortex[18,19]. However, the role of spindles in gating and coordinating multi-region communication remains to be explored at the cellular and systems level.

The coupling between these non-REM oscillations appears to be causally linked to memory consolidation, as suggested by the disruption or enhancement of memory following closed-loop manipulations[20,21], and by the association of spindle dysfunction with perturbation of memory function in some neuropsychiatric disorders[22,23]. However, we are far from understanding the specific mechanisms and single-cell dynamics involved in the hippocampal-neocortical interaction. In particular, information regarding the thalamic involvement is limited. Evidence from anesthetized animals suggest a correlation of thalamic firing with both the neocortical slow oscillation[24,25] and hippocampal SWRs[26], while fMRI results in monkeys indicate an

overall decrease in thalamic activity during SWRs[27]. A recent study in naturally sleeping rats[28], found a decrease in activity in cells of the thalamic mediodorsal (MD) nucleus at the time of CA1 SWRs, particularly during SWRs uncoupled from spindles recorded in the skull EEG. This emerging evidence emphasizes the need to study the thalamus, but we have been lacking a multi-region approach (thalamus, neocortex, hippocampus) that would compare in the same preparation the correlations of cells in each region to the sleep oscillations, and that would investigate the dynamics of the thalamic contribution down to the finer timescales relevant for plasticity.

We sought to address this gap by characterizing the fine temporal correlations of thalamic cells with sleep oscillations through extracellular recordings of single units and LFPs in the medial prefrontal cortex (mPFC), hippocampal CA1 region, and nuclei of the midline thalamus in freely behaving rats. We targeted these brain regions because of their anatomical and functional connectivity and their role in memory processes. The limbic thalamus has widespread connections with neocortical regions, and projections from the midline are particularly dense to mPFC[29–31], a critical region for the recall of consolidated memories[32]. The reuniens nucleus in the midline sends collaterals to CA1 and mPFC, making it a compelling candidate to coordinate mPFC-CA1 activity[33,34]. Recording from these areas, we find evidence for a dual role of the thalamus in hippocampal-neocortical sleep interactions; first, an increase in thalamic firing at the down to up-state transition could influence the dynamics of the slow oscillation, and second, the sparse but consistent firing of thalamic cells within individual spindles suggests an interleaved activation of thalamocortical synapses that may promote a flexible integration of recently acquired memory traces into neocortical schemas during systems consolidation.

Results

We used tetrodes to record extracellularly (single units, LFPs) from the midline thalamus, and from areas involved in recent (CA1) and remote (medial
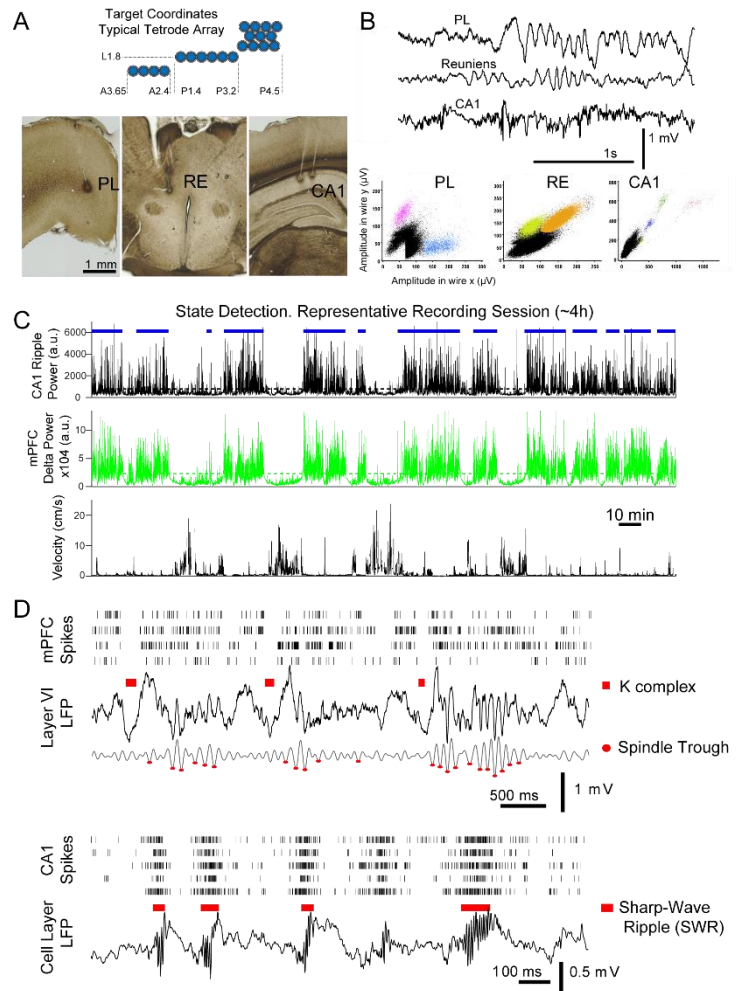
Figure 1. Identification of non-REM sleep oscillatory events in mPFC and CA1. *(A), Diagram of the cross-section of a typical multi-tetrode array and target coordinates. (B) Sample LFP traces and units simultaneously recorded in mPFC (prelimbic region, PL), reuniens (RE), and CA1. (C) Resting states (blue horizontal bars) were detected from the moving average of the squared filtered LFP, SWR-filtered from CA1 - top panel-, and delta-filtered from mPFC -middle panel-; selected periods used for analyses were confirmed by visual inspection and observation of the corresponding animal velocity -bottom panel-. (D) Multi-unit spike rasters (each row shows all spikes from one tetrode) and LFPs, illustrate the detection of sleep events used for analyses; top: detected K complexes (marked above raw LFP trace) and spindle troughs (marked on the 6–14 Hz filtered LFP) in mPFC; bottom: examples of CA1 sharp-wave ripple detection.*

prefrontal cortex, mPFC) memory recall[32,35]. During the 1–5 h recording sessions (in freely behaving rats), the animals remained in a quiet, square-shaped enclosure where they cycled through bouts of sleep and wakefulness; in 6 sessions from 3 rats, the

animals also explored a radial maze after the recording of sleep, and were brought back to the sleep area after behavior for comparison of sleep activity before and after spatial exploration. LFPs and 317 units were recorded from 7 rats; sessions from an additional animal in which no units were recorded were also included for LFP analyses for a total of n = 8 rats (Figure 1a displays a subset of tetrode locations in brain sections from one rat; Supplementary Figure 1, parts 1 and 2, document the trajectories in the brain for the electrodes and sessions used for analyses). We selected recording sessions in which there was at least one unit recorded in the thalamus and where reliable detection of sleep events could be performed (slow oscillation and spindle activity in mPFC, sharp-wave ripples, SWRs, in CA1; examples of simultaneously recorded LFP traces and units shown in Figure 1b). The majority of recordings in the thalamus were from units in the reuniens (RE), and ventromedial (VM) nuclei, and some from the mediodorsal (MD), paratenial (PT), rhomboid (RH) and centromedial (CM; detailed population numbers in Supplementary Table 1). All of these nuclei are bi-directionally connected to mPFC, and are functionally related to mPFC and other higher order cognitive regions, including CA1, which also receives direct input from reuniens. Here we will use the term limbic thalamus to refer to the thalamic nuclei that we recorded from. See the Methods section for target anatomic coordinates, procedures to identify tetrode location, and additional inclusion/exclusion criteria for the analyses reported below.

To study the modulation of unit spikes in the three regions (mPFC, limbic thalamus, CA1) with non-REM sleep oscillations, we first identified times with both high delta (1–4 Hz) power in mPFC and high SWR (100–275 Hz) power in CA1 (as described in Methods); an example of such non-REM sleep state detection for a representative session is shown in Figure 1c. Within this state, we detected reference features that reflect non-REM sleep events that are key organizers of memory consolidation: K-complexes (KCs, marked by red bars in Figure 1d, mPFC LFP), the troughs of spindle cycles in mPFC (red dots in 1d), and SWRs in CA1 (red bars in Figure 1d, CA1 LFP). These LFP events are markers of cell population dynamics, namely, KCs mark the down-states of the neocortical slow oscillation, while SWRs correspond to the high-frequency oscillations

that reflect memory-related reactivation of ensembles of CA1 place cells[11,13,14,36]. Spindles (6–14 Hz) are generated through oscillatory bursting in the thalamic reticular nucleus (TRN), which is thought to then engage cells in the dorsal thalamus and, through them, neocortical networks[18,37–39]. We considered that the time window of each spindle cycle has a similar duration to that of SWRs (up to ~ 100ms) and sought to test if individual spindle cycles represent a window for multi-region coordination; we thus detected individual spindle cycles from the mPFC 6–14 Hz filtered LFP. The detection of individual cycles was also motivated by the occasional observation of few spindle cycles in succession (e.g., only 3–4 cycles following the second detected KC in Figure 1d), and by the broader variability in the morphology of the LFP trace in the spindle band compared to the more clearly discrete and consistent signatures of KCs and SWRs (examples in Figure 1d). KCs, spindle cycles, and SWRs were detected from the mPFC and CA1 LFPs after applying an amplitude threshold to the squared, filtered trace (1–4 Hz for KCs; 6–14 Hz for spindles; 100–275 Hz for SWRs), at a level of three (spindles) and four (KCs, SWRs) standard deviations above the average LFP voltage across sleep periods in each recording session (see Methods for details). Although the spindle band overlaps with the theta band, we limited the detection of spindle cycles (and KCs and SWRs) to resting states; peri-spindle spectrograms confirmed that detected spindle cycles are functionally different from theta (Supplementary Figure 2). Peri-event histograms of the LFP and local spiking activity, triggered on the timestamps at the time of largest amplitude of the detected events (trough for KCs and spindles, peak for SWRs), confirmed that the LFP detection identified events that reflect the modulated activity of local cell populations (Supplementary Figure 3). Quantifications are reported as mean ± standard deviation (sd), unless otherwise indicated.

Figure 2a illustrates the temporal and phase relations between non-REM sleep events, with examples from the raw and filtered mPFC and CA1 LFPs. The overlaid filtered LFP traces (which have been scaled for display purposes), show that increases in SWR amplitude occur preferentially in the positive half-cycle of the delta-filtered LFP, and in the negative halves of the spindle-filtered trace (inset). The average distribution of delta and spindle phases (estimated with the Hilbert transform, see Methods)

at the time of SWRs is shown in Figure 2b for all sessions. Figure 2c shows the peri-event histograms for non-REM detected population events (SWRs, KCs, spindle cycles) across sessions and rats.
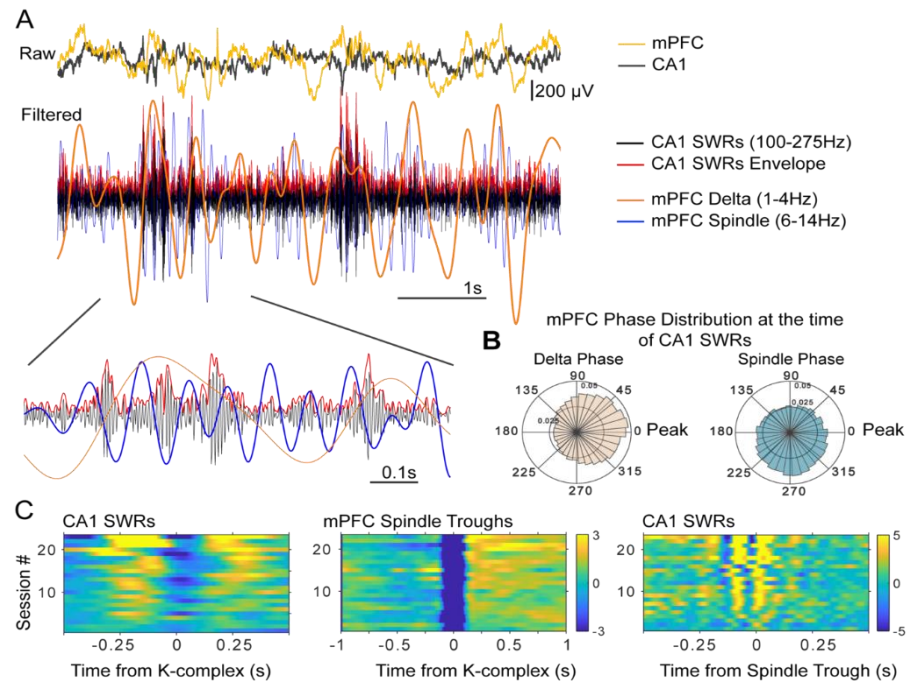
Confirming previous results[9,15–17], we found that KCs, spindles, and SWRs are temporally and phase-coupled; SWRs preferentially occurred following and preceding KCs, phase-locked to the peaks of the delta-filtered LFP, and to the troughs and ascending phase of spindle cycles (circular mean and standard deviation of delta phase at the time of SWRs = -0.04 ± 0.43 rad; spindle phase = -1.72 ± 0.93 rad; n = 23 sessions, 6 rats; Supplementary Figure 4 shows the delta and spindle phase distributions at the time of SWRs for all sessions included in this analysis).

hyperpolarization, which is a pre-requisite for T-channel activation, and they can enhance transmission in thalamocortical synapses and induce synaptic plasticity[42,43]. Therefore, analyzing burst
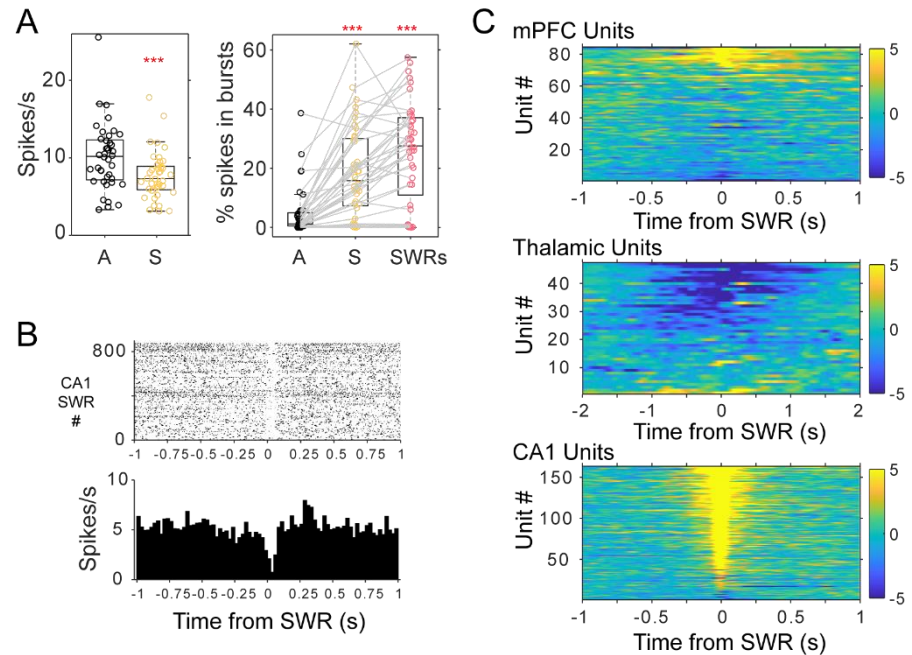

Figure 2. Time and phase coordination between non-REM sleep events. (A) Overlaid raw and filtered mPFC and CA1 LFPs display the time relations of the relevant oscillatory events (filtered traces have been scaled in amplitude for display purposes). (B) Average (across sessions and rats) distribution of delta and spindle phase at the time of SWRs, showing a preferential occurrence of SWRs near the peak of delta and the ascending phase of spindles. (C) Peri-event histograms of detected non-REM sleep events show that SWR probability increases in the ~250 ms windows near KCs (left panel), and are nested in the negative half-cycle of spindles (right), spindles are largest in amplitude following KCs (middle) (color scale bars indicate standard deviations with respect to a null, shuffled, distribution; scale bar in the middle panel applies also to the left panel).

Thalamic cells decrease their overall firing and increase bursting with CA1 SWRs

We next investigated the changes in the firing patterns of single units in the limbic thalamus with CA1 SWRs.

Most thalamic units presented a reduced average firing rate during rest compared to wakefulness. In 28 of 32 sessions the animals (n = 7) alternated between sleep and active wakefulness (in the other 4 sessions rats were only in wakefulness for less than 5 mins and were not included in this comparison), and the firing rate of thalamic units changed from an average of 10.10 ± 4.26 spikes/s during wakefulness to 7.73 ± 3.09 spikes/s during sleep (n = 40 units; p = 1.31 × 10^{-6}, Wilcoxon signed rank; Figure 3a). A decrease in firing rate may be due to the hyperpolarization of thalamic cells. Thalamic cells fire in two modes, burst and tonic, depending on the activation state of a transient conductance that relies on T-type calcium channels[40,41]. Bursts correlate with thalamic

firing can provide clues on the mechanism behind the decrease in firing rate in sleep. Bursting can be detected from extracellular recordings because of its characteristic combination of a relatively long inter-spike interval followed in rapid succession by a few spikes with brief inter-spike intervals[44], but the correlation between thalamic bursts and SWRs has not been explored.

We detected bursts in the thalamus as groups of action potentials with less than 5 ms of inter-spike interval preceded by at least 70 ms with no spikes, a criterion based on the dynamics of the T-channels underlying burst firing in the dorsal thalamus[41,44,45].

With this criterion, we found that 25 % of the thalamic cells had at least 5 % of their spikes in bursts during wakefulness, while 77.5 % did during sleep (average percentage of spikes in bursts during wakefulness 4.60 ± 7.97 %, and during sleep 18.65 ± 15.3 %; p = 1.8×10^{-7}; Wilcoxon signed rank; n = 40 units, 7 rats; Figure 3a). The increase in bursting was more pronounced around the time of SWRs occurrence in CA1 (25.52 ± 17.29 % of spikes in bursts in a 2 s window around SWRs; p = 1.8×10^{-7} Kruskall-Wallis; Figure 3a). Because the T-channels responsible for burst firing need a sustained hyperpolarization for about 50–100 ms to be primed for activation[41,46], the increase in bursting suggests a hyperpolarization of the majority of cells in the limbic thalamus during sleep, and particularly in correlation with hippocampal SWRs.

Indeed, peri-SWR triggered histograms showed that SWRs

Figure 3. Decrease of firing in thalamic units with SWRs. *(A) Left, Distributions of average firing rate for all units in the awake, A-black, compared to sleep periods, S-yellow. Right, distributions of the percent of spikes in bursts for the same units during wakefulness, sleep, and in a window of 2 s around the occurrence of CA1 SWRs. (B) Example of peri-SWR-triggered histogram for a unit recorded in the nucleus reuniens, showing a decrease in firing tightly locked to SWR occurrence (top plot is the raster of unit spikes for all detected SWRs). (C) Population results of peri-SWR triggered histogram of spikes for all mPFC (top), thalamic (middle), and CA1 (bottom) units; scale bars units are in standard deviations from the null, shuffled, distribution.*

occur in a background of decreased thalamic spiking. An example of a peri-SWR histogram for a unit from the RE nucleus is shown in Figure 3b, and the peri-SWR for the population of recorded cells in the three areas is shown in Figure 3c; 55.32 % of units in the thalamus showed a reduction in firing of more than 2 standard deviations (sd) from the null (average -4.23 ± 2.00 sd) in a 200 ms window around the time of SWRs occurrence. Instead, 90.18 % of CA1 units increased their firing by 19.85 ± 18.65 sd in a 200 ms window around SWRs (4.91 % of CA1 units showed an average decrease of 4.11 ± 1.59 sd), while the mPFC population included 30.95 % of units showing an increase (average 6.22 ± 4.32 sd) and 40.48 % a reduction in firing (average -3.22 ± 1.40 sd). Using a complementary approach, generalized linear models (GLMs) of CA1 activity (SWRs and multi-unit firing rate) with the firing rate in thalamic units as covariate, showed that the firing in CA1

varied inversely with the firing of most thalamic units (61.7 %, p < 0.001; only one unit in the MD nucleus had a significant positive correlation with SWRs rate). For the units that negatively correlated with SWRs, the models predict an average increase of 20.74 % in the rate of SWRs for every 1 spike/s drop in thalamic firing, suggesting anti-correlated activity in the limbic thalamus and CA1 (in contrast, as expected, the rate of SWRs was positively correlated with the spiking activity recorded in CA1; p value < 5×10^{-189} in all electrodes used for SWR detection). The modulation of thalamic units with SWRs was observed both in RE and in other nuclei. We observed some of the strongest negative correlations with SWRs in RE (unit in Figure 3b), and the size of the effect was marginally significant in RE compared to non-RE units (p = 0.05). However, there was no difference between the proportion of SWR-correlated units in RE compared to non-RE units (p = 0.159), suggesting that the decrease in thalamic firing with

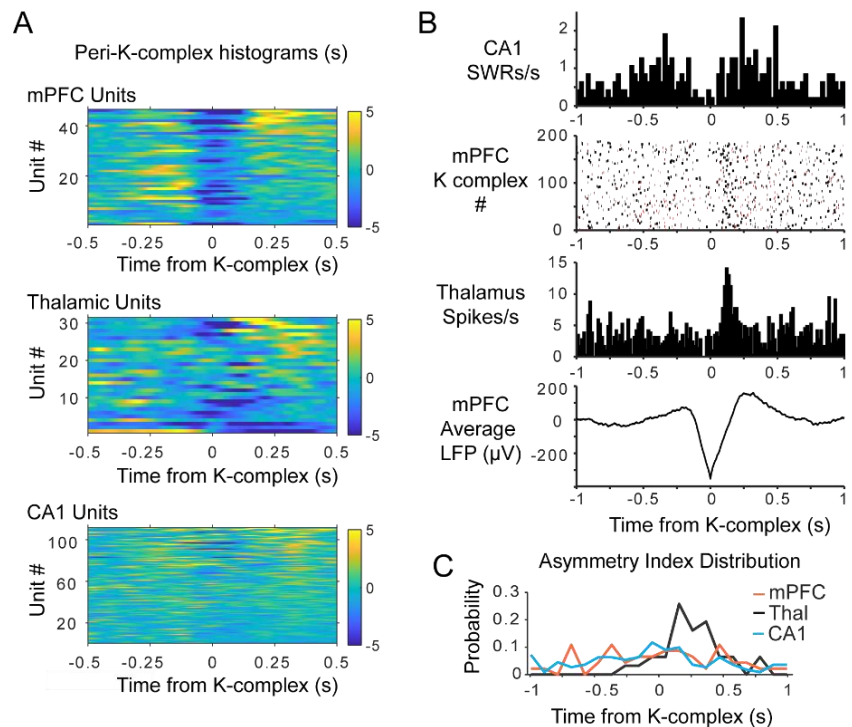SWRs relates to general excitability changes in the state of thalamocortical networks in the limbic thalamus.

Correlation of thalamic units with mPFC delta and spindle oscillations

Although we found a decrease in average thalamic firing and an increase in bursting that suggests a hyperpolarized thalamus during non-REM sleep, it is unclear how the remaining thalamic spikes relate to the mPFC delta (1–4 Hz) and spindle (6–14 Hz) oscillation bands, and if there are specific temporal windows with selective increase in thalamic firing. To investigate this, we used two approaches: we calculated firing rates with respect to local events detected in the mPFC LFP (KCs, spindle troughs); and we estimated the phase-preference of unit spikes to the delta and spindle-filtered mPFC LFP.

Thalamic rebound following the down-state of the mPFC slow oscillation

Figure 4a shows peri-KCs histograms for units in the three regions, as well as a representative example of the temporal relations between KCs, SWRs and thalamic activity in one recording session (Figure 4b). Units in the thalamus often fired asymmetrically with respect to KCs, with stronger firing in the few hundreds of miliseconds after the mPFC KC compared to before (average of 2.48 standard deviations at 274.76 ± 135.33 ms after KCs, compared to 1.28 standard deviations at 263.31 ± 145.31 ms before KCs; n = 31 thalamic units). The trough of thalamic firing occurred on average after the mPFC KCs and after the trough of mPFC unit spikes, and coinciding with the trough of SWRs in CA1, suggesting that the neocortex leads both the thalamus and CA1 in the slow oscillation (thalamic units 10.56 ± 130.75 ms after KCs, n = 31; mPFC units -7.70 ± 72.3 ms, before LFP KCs, n = 46; SWRs 10.33 ± 5.88 ms after KCs).

Previous evidence[8] suggests that the thalamus may have a specific role in the initiation of cortical up-


Figure 4. Rebound of thalamic activity following mPFC K-complexes (KCs). *(A) Population results for mPFC (top), thalamus (middle), and CA1 (bottom), showing the preferential firing of thalamic units following KCs, compared to the mPFC and CA1 populations. Scale bar units are standard deviations from the null, shuffled, distribution. (B) Example from one recording session with all the plots aligned to the mPFC KCs; top panel is the peri-KC histogram of SWR occurrence, middle plots show the peri-KC histogram and raster for a thalamic unit (red ticks on the raster indicate detected SWRs), bottom plot displays the average KC-triggered mPFC LFP for the session. (C) Distribution of the asymmetry index in the three cell populations highlights the asymmetric response in the thalamic population compared to the other two groups.*

states. To assess the specificity of thalamic firing to the down to up-state transition, we compared the firing of thalamic units before and after mPFC KCs (from the peri-KCs histograms in Figure 4a). For the population of thalamic cells, 38.71 % increased their activity above 2 sd (from the null distribution) following KCs, i.e., in the down to up-state transition, compared to 9.68 % of the cells with a significant peak preceding KCs, i.e., in the up to down-state transition (19.35 % units had a significant peak both before and after KCs). Instead, CA1 and mPFC activity was more similar preceding and following

KCs. SWR probability increased within the 250 ms around mPFC KCs (4.65 sd at 208.15 ± 90 ms before KCs, and 3.02 sd at 266.85 ± 100 ms after KCs; n = 23 sessions). This indicates a delta modulated increase in SWR probability in CA1, with the minimum rate of ripples following briefly after the trough of neocortical activity (10.33 ± 5.88 ms). The modulation of SWRs was paralleled by a modulation of firing in CA1 units (n = 111), which increased their
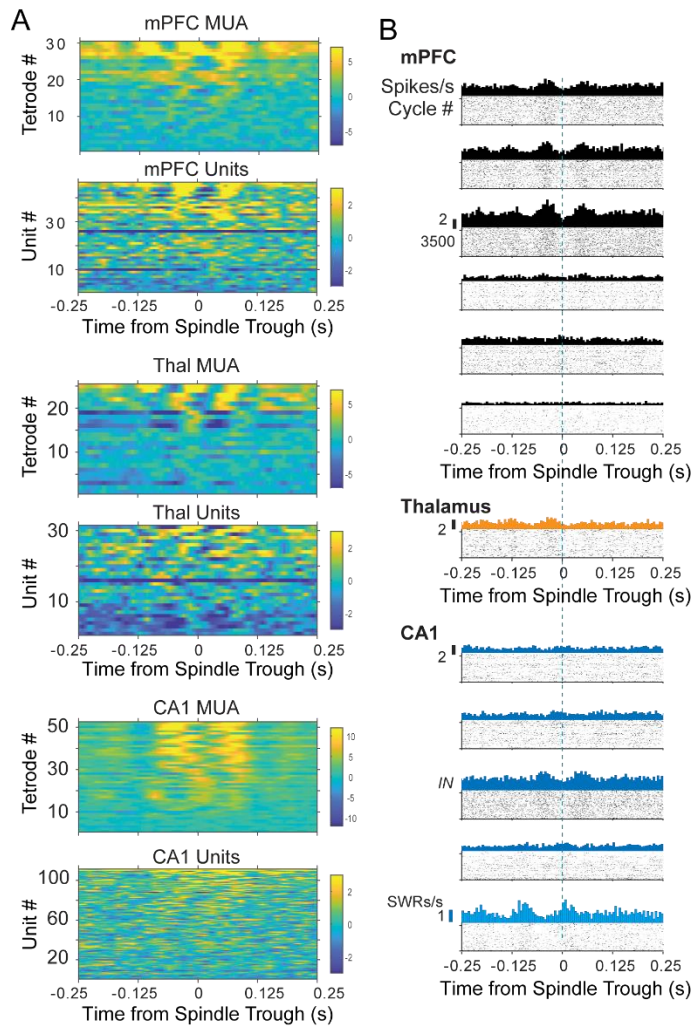

Figure 5. Modulation of firing with mPFC spindles. *(A) Spindle-trough triggered perievent histograms of multi-unit activity (MUA) and units in mPFC (top), thalamus (Thal, middle) and CA1 (bottom) in standard deviations (SD) with respect to the null, shuffled, distribution. (B) Sample session with SWRs and unit spike rasters and peri-spindle histograms; 6 units recorded in mPFC (black), one in thalamus (orange), 6 in CA1 (blue; IN = putative interneuron); the vertical dashed blue line marks time 0 across plots for reference.*

activity both before and after KCs (2.32 ± 1.49 sd at -233.7 ± 146 ms before KCs, and 2.35 ± 1.69 sd at 273 ± 140 ms after KCs); on average, CA1 units presented a trough of activity after KCs (19.53 ± 132 ms). Activity in mPFC (n = 46 units) was modulated in a similar way, with increases in unit firing preceding and following the down-state of the slow oscillation (2.33 standard deviations at -267.30 ± 110.40 ms, before KCs; 2.31 standard deviations at 295.20 ± 120 ms, after KCs).

In other words, while units from the three areas showed no difference in the peak of the histograms in the time window following KCs (p = 0.89, Kruskal-Wallis between regions), thalamic units had significantly lower firing before KCs compared to mPFC and CA1 units (p = 0.007, Kruskal-Wallis). This suggest that thalamic cells contribute spikes primarily as the new cycle of slow oscillation starts, while mPFC and CA1 units also fire at up to down transitions. We used an additional metric to quantify the differential firing of units in the three areas with respect to the time of mPFC KCs. For all the units, we calculated the difference between the maximum firing rate in the 0.5 s windows before and after KCs, normalized to the maximum firing in the two windows. This asymmetry index ranges between -1 and 1, with negative values indicating higher firing before KCs. The distribution of this index (Figure 4c) was biased to positive values for thalamic units, but was uniformly distributed for mPFC and CA1 units (average 0.24 ± 0.22 in thalamus; -0.004 ± 0.47 in mPFC; -0.06 ± 0.5 in CA1), consistent with a selective rebound of thalamic firing in the time window following mPFC KCs relative to thalamic activity preceding the down-state.

Sparse thalamic firing and consistent activation of cell pairs during spindles

We then analyzed the firing at the timescale of mPFC spindles (Figure 5), and found the strongest modulations in the CA1 multi-unit activity (MUA, all spikes recorded by a given electrode). Peri-spindle-trough histograms of MUA and single units are shown in Figure 5a (n = 30 tetrodes in mPFC; n = 25 tetrodes in thalamus; n = 52 tetrodes in CA1; n = 46 mPFC units; n = 31 thalamus units; n = 111 CA1 units). Figure 5b shows an example of the histograms for one session in which a unit in the mediodorsal nucleus of the thalamus was recorded at the same time as units in mPFC and CA1. Across the sessions, 84.62
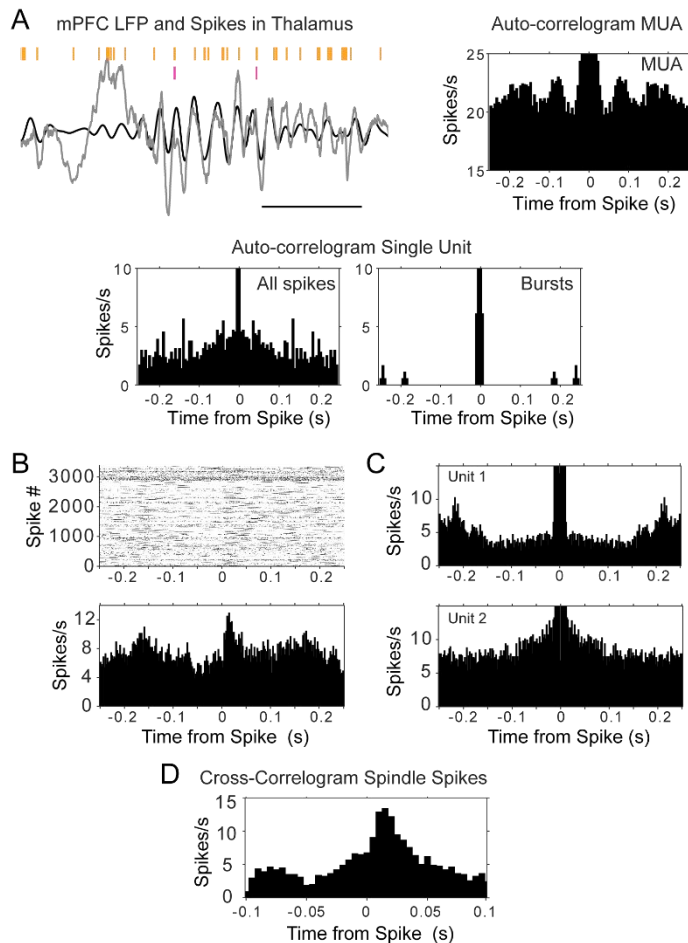
Figure 6. Auto- and cross-correlograms of thalamic spikes show lack of single unit rhythmicity but consistent activation of MUA and unit pairs. *(A) Top left, raw trace of mPFC LFP in gray, with overlaid filtered trace (6–14 Hz, black); spike raster in orange corresponds to all spikes (multi-unit activity, MUA) recorded by one thalamic tetrode, raster in magenta is from a single unit recorded by the same tetrode; scale bar 500 ms. Bottom, auto-correlograms of thalamic spikes during sleep for the single unit (all spikes were used for the plot on the left, burst spikes only for the plot on the right); top right, only the auto-correlogram for the MUA recorded by the same electrode displays rhythmic firing at spindle frequency. (B) Cross-correlogram between the sleep spikes of two thalamic units, showing increased modulation within about 50 ms, even when the auto-correlation of the spikes from each individual unit (C) does not show rhythmic firing. (D) Cross-correlogram of the same thalamic pair, calculated only with spikes within spindle cycles, suggests spindle-related co-activation.*

% of the CA1 tetrodes showed an increase in spike activity (at least 3 standard deviations from null distribution in the -100 to 100 ms window of the peri-

spindle-trough histogram), with 53.3 % and 36 % tetrodes showing a similar result in mPFC and thalamus. The increase was less clear and consistent in individual units, even when the local MUA had a clear modulation, which suggests a functional coordination at the cell population level with sparse engagement of individual cells. Spindle oscillations can vary in frequency[7] and individual units may not contribute spikes in every spindle cycle, flattening the average correlograms. To investigate the oscillatory activity of thalamic units directly, we looked for peaks in the auto-correlograms of the single unit spikes.

The auto-correlograms of thalamic units did not present peaks at the time intervals of spindle frequency, i.e., in the time window between 50-150 ms, suggesting that the cells did not fire rhythmically in that frequency. Figure 6a illustrates the sparsity of thalamic cell firing during mPFC spindles; the panel shows a trace of mPFC LFP with several cycles of spindle oscillation (6–14 Hz filtered trace overlaid in black); the MUA recorded by one tetrode in the thalamus is shown in orange and the spikes of a single unit in magenta. The auto-correlograms next to the raw data were calculated with the spikes fired during sleep, and show the lack of spindle modulation in the single unit (also when calculated with only burst spikes), in spite of the MUA modulation (which can also be seen in the spike raster). Similar results were obtained for all thalamic units, regardless of the nucleus, and including the units that showed the strongest rebound following KCs (Supplementary Figure 5). Because cells could be firing rhythmically only when they are bursting, we also calculated auto-correlograms with only burst spikes, as well as only for the spikes that occurred in the 0.5 s time window immediately following KCs (when spindle troughs are largest). Thus, these analyses were more restricted to conditions in which rhythmicity in the spindle band would be expected to be strongest, and still failed to reveal evidence of rhythmic firing in the thalamic cells that we recorded from (Supplementary Figure 5).

The lack of rhythmicity in individual thalamic units, together with the modulation of the MUA, raise the possibility that different groups of thalamic units contribute to activity in different spindle cycles. A prediction from this hypothesis is

that pairs of units should be more likely to be correlated in brief timescales than individual units with themselves. We tested this in sessions in which we recorded more than one isolated unit in the thalamus (n = 12 pairs, 3 rats), and found that half of the pairs showed a sleep cross-correlation peak above 3 standard deviations from the average cross-correlogram (average 2.86 ± 0.87 sd). An example is shown in Figure 6b, which displays the cross-correlation of two thalamic units during sleep; the plots show that the spikes of 'unit 1' follow reliably in a window of ~50 ms after those of 'unit 2' (reference), while neither of the units show a strong auto-correlation in that period (Figure 6c).

All the pairs that showed a peak in the sleep cross-correlograms were correlated in a time scale consistent with the duration of spindle cycles (Figure 6b; Supplementary Figure 6 for all pair correlations). We then identified individual spindle cycles by finding the peak of the voltage trace before and after detected spindle troughs. The cross-correlograms between pairs of units showed similar values when calculated only with the spikes that occurred within spindle cycles (Figure 6d; Supplementary Figure 6 for all pairs), providing further support to the hypothesis of a co-activation of sparsely synchronous thalamic cells within spindles (average peak cross-correlation with spindle spikes 2.42 ± 0.41 sd; p = 0.18, when comparing the peak correlations calculated with all sleep spikes and with only spindle spikes; Wilcoxon signed rank).

The coordinated activity of cell populations drives amplitude and phase dynamics in the local field potentials[47], and phase coordination has been suggested to underlie routing mechanisms
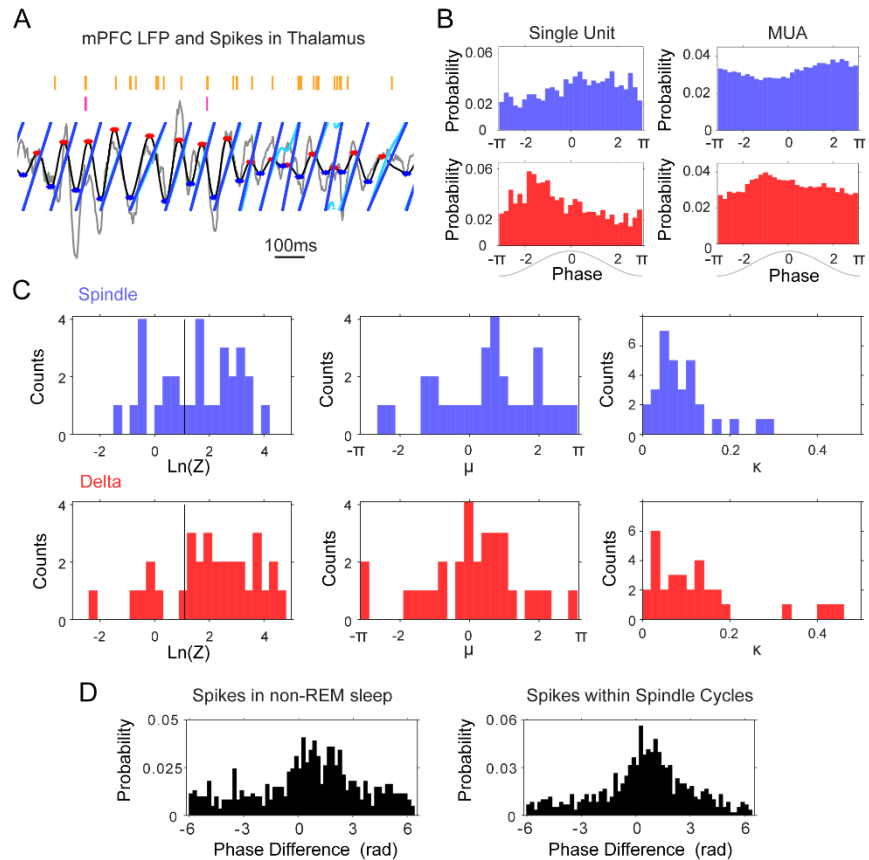
Figure 7. Phase-locking of thalamic units to mPFC delta and spindle frequencies. *(A) Instantaneous phase estimate for a filtered mPFC LFP trace using the Hilbert transform (cyan) or peak-trough interpolation (blue). (B) Spindle (blue) and delta (red) phase-distribution at the time of single unit and multi-unit spikes recorded during sleep in one tetrode in thalamus (sample rasters in Figure 7a); the distributions are non-uniform and suggest phase-locking of these spikes to the descending phase of the spindle oscillation and the ascending phase of delta. (C) Left, distribution of log-transformed Rayleigh's Z for all thalamic units, used to evaluate the significance of phase-locking to the two frequency bands; values to the right of the black vertical line are significant at the p = 0.05 level. Panels on the right side show the distribution of the μ (mean direction, middle panel) and κ (concentration, right panel) of the phase-distribution for all units, which suggest preferential firing at the peak of delta and descending phase of spindle oscillations. (D) Phase lag distributions for the thalamic pair in Figure 6; spindle phase-difference between the closest spikes from the two units was calculated for all spikes in non-REM sleep (Left) and for all spikes within spindle cycles (Right), and suggests that the two units are most likely to fire in sequence within about a quarter of a spindle cycle from each other.*

within and across brain regions[48–50]. Therefore, to confirm a functional correlation between thalamic spikes and mPFC non-REM oscillations, we assessed

the phase locking of thalamic units to the delta and spindle frequency bands of the mPFC LFP.

Phase-locking of thalamic units to delta and spindle oscillations

We studied the phase-locking of thalamic units to the delta and spindle frequencies by estimating the LFP phase using two complementary methods, the analytical signal and linear interpolation between consecutive troughs of the spindle and delta filtered LFPs (see Methods for details). Figure 7a shows the estimated phase obtained with both methods for the spindle trace in Figure 6a. Although the unit fires sparsely (raster above the trace), the spikes are more likely to occur on the descending phase of the spindle-filtered and ascending phase of delta-filtered LFP, shown by the phase histograms calculated for the whole sleep session (Figure 7b). We found similar preferential firing for the population of thalamic cells. The phase histograms were fitted for all units to a circular Gaussian (von Mises distribution), with mean and concentration parameters that indicate the direction and depth of the modulation or phase-locking. In order to prevent sampling bias due to non-uniform baseline phase distributions (e.g., due to non-sinusoidal LFP morphology), the unit phase distributions were also normalized to the baseline phase distributions during sleep[51].

We found that the majority of thalamic units (96.77 % of n = 31 units) showed preferential firing at the descending phases of the spindle-filtered mPFC LFP, as suggested by the positive values of the mean phase distribution (0.77 ± 1.15 rad; $p < 0.05$, Rayleigh test; Figure 7c). Thalamic units were phase-locked to the mPFC delta band, closer to the positive peak of the oscillation (96.77 %, $p < 0.05$, Rayleigh test; average 0.30 ± 1.04 rad). The amplitude of the thalamic rebound after KCs (reported above) had a tendency to correlate with a preference for earlier phases of delta; although the correlation was not significant ($p = 0.08$, $r = -0.32$), we did not observe such a trend when we regressed mean phase preference to the cell firing rate in a time window of the same length but before KCs ($p = 0.65$, $r = 0.08$).

Interestingly, we found that the phase lags between the spikes of unit pairs that had the largest temporal cross-correlations (Supplementary Figure 7) were more consistent than the phase lags between

uncorrelated pairs. In the sessions with thalamic pairs (n = 12), we calculated the smallest difference between the phase of the spikes during non-REM sleep, and between the phase of the spikes in spindle cycles. The phase difference would be expected to be distributed between -2π and 2π if units fire within one cycle of each other (positive or negative depending on their sequential order), and units that are functionally locked are expected to show a more consistent, less variable phase relation. Figure 7d shows the phase lag distribution for the pair described in Figure 6, calculated both from all non-REM spikes (left panel) and from the spikes that occurred within spindle cycles (right panel); the distributions suggest that most of the spikes occur within less than half a cycle of each other; the slight asymmetry with respect to zero suggests the consistent firing of one of the units before the other. We found similar results (Supplementary Figure 7) for the thalamic pairs that showed significant peaks in their cross-correlogram. The standard deviation of the distribution of phase lags was significantly lower for units with high spike cross-correlations during sleep (pair cross-correlograms > 3 sd) compared to the units that were not temporally correlated. The standard deviation of phase lag distributions in correlated units was 2.7 compared to 3.05 in uncorrelated units when calculated with all non-REM sleep spikes ($p = 0.026$, Mann-Whitney U-test), and was 2.46 compared to 2.72 for the spikes within spindles ($p = 0.009$, Mann-Whitney U-test). These results demonstrate the phase-locked firing of thalamic cells within the period of the spindle oscillation.

Overall, the phase-locking results are consistent with the peri-event histogram analyses in suggesting coordinated thalamic firing with the neocortical delta and spindle oscillations, and demonstrate the phase coordination of pairs of thalamic units within individual spindle cycles.

Spindle activation of thalamic units modulated with CA1 SWRs

Because SWRs were also locked to preferential phases of the spindle oscillation (Figure 2b and Supplementary Figure 4), we investigated the correlation between thalamus and SWRs within the timescale of spindle cycles. We calculated the ripple power within each spindle cycle, and classified cycles
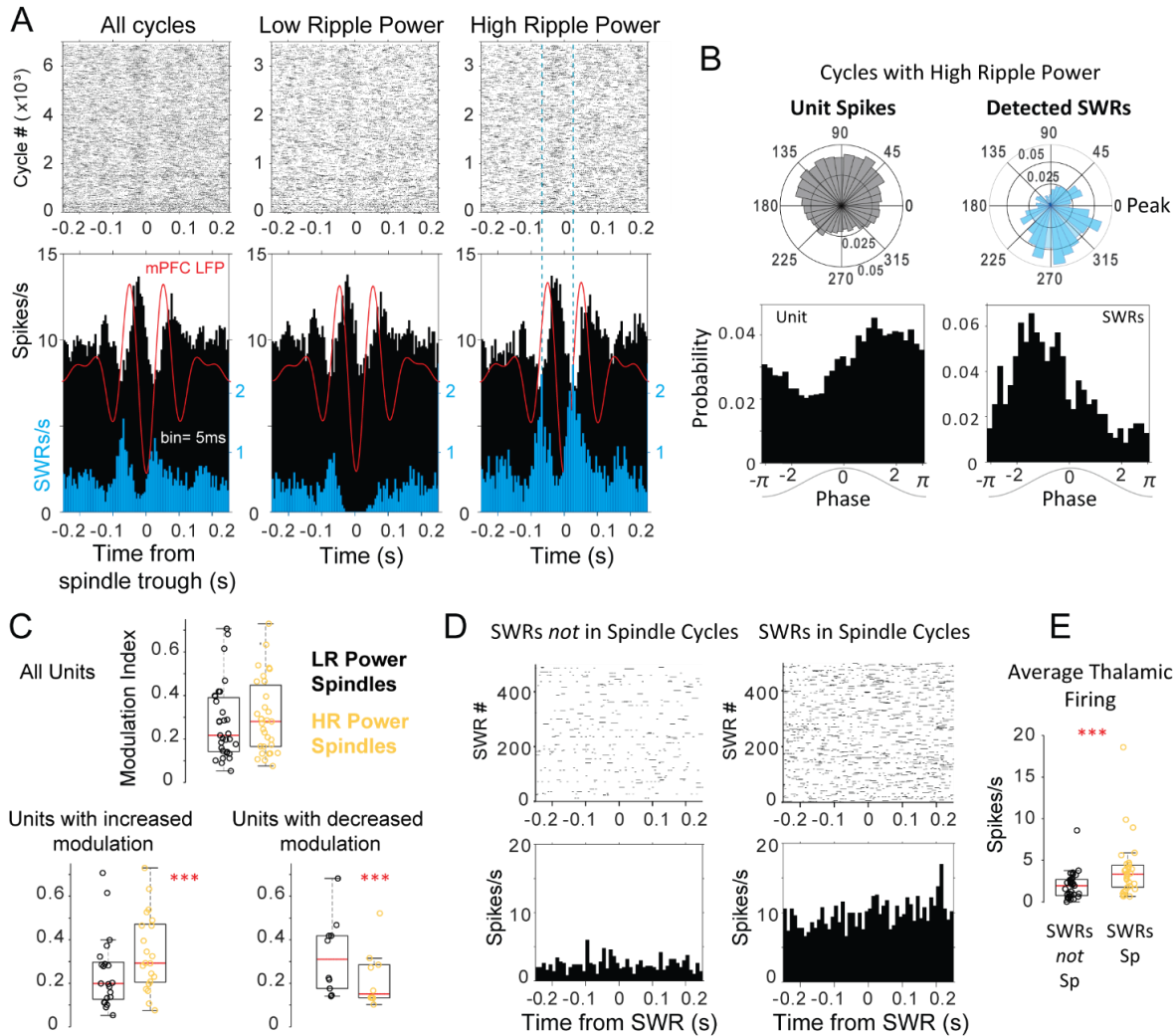
Figure 8. Modulation of spindle-related thalamic activity with SWRs. *(A) Spike rasters and peri-spindle-trough histograms of thalamic spikes from one unit and for SWRs recorded in the same session during sleep. Spindle cycles were classified into low and high ripple power cycles, according to the median power in the CA1 ripple band during the spindle cycle. In this unit there is an increased spindle modulation of firing in spindles with high ripple power (right plot), compared to those with low ripple power (middle); the spindle-triggered histogram of SWRs (blue) confirms the increased probability of SWR events in high-ripple power spindles. The average spindle-triggered mPFC LFP is overlaid in red. (B) Example of phase-locking offset between the unit spikes and CA1 SWRs; spikes from the thalamic unit in (A) occurred preferentially in the descending phase of spindles with high ripple power, in contrast to the SWR preference for ascending phases. (C) Population distribution of a modulation index shows that the majority of units (each circle one unit) increase in their modulation in cycles with high ripple power (HR); the distribution of modulation values is split below for units with increase or decrease in modulation with spindles. (D) Cross-correlation of SWRs nested (left) or not (right) in spindle cycles for the same session and unit as in (A); firing in the unit is higher with spindle-couple SWRs, although there is no strong temporal correlation with SWRs.(E) Average firing in peri-SWRs histograms for all thalamic units, shows an increase in firing with SWRs nested in spindles (SWRs-Sp, yellow) compared to those outside spindles.*

in two groups based on the median SWR power (detected spindle cycles and feature distributions for all sessions in Supplementary Figure 8). Cycles with ripple power above the median were slightly shorter

in duration (102.17 ± 2.01 ms compared to 103.22 ± 2.35 ms when ripple power was low; $p < 0.05$ in 65 % of the sessions although not significant for the mean duration across sessions, $p = 0.11$; Mann-

Whitney U-test) and larger in amplitude (229.21 ± 102.14 µV versus 219.47 ± 94.32 µV; $p < 0.05$ in 85 % of sessions, $p = 0.54$ across sessions; Mann-Whitney U-test).

Figure 8 displays the modulation of firing in one of the thalamic units with the strongest spindle modulation in our sample; the spindle-modulated spiking in this unit was enhanced in cycles with high SWR power in CA1. The peri-spindle-trough histogram of spikes is shown for all the spindle cycles in the session on the left panel (Figure 8a), and for the two functionally defined types of spindle cycles, with low and high SWR power, in the middle and right panels. The spindle-trough-triggered histogram of CA1 SWRs for the same session is shown in blue, and confirms that the spindle classification identifies cycles with low or high probability of SWRs in CA1.

The majority of thalamic units showed an increase in spindle cycle modulation in cycles with ripple power above the median (67.7 %, n = 31). A modulation index, calculated as the difference between peak to trough firing relative to the mean in a 200 ms window centered on the peri-spindle-trough histogram, was significantly larger in cells with an increase in modulation compared to those with a decrease in modulation in high-SWR-power spindles. The same metric was not different between the two groups of units during low-SWR-power spindles, suggesting an enhanced recruitment of some thalamic cells in high-SWR-power spindle cycles (the index was 0.34 ± 0.18 in cells with increased modulation, compared to 0.22 ± 0.13 in cells with decreased modulation in cycles with high ripple power; $p = 0.04$, Mann-Whitney). Cells with increased modulation in high-SWR-power spindles also showed a significantly higher concentration in the distribution of spindle phases at the time of their spikes, indicating that they are more tightly phase-locked to high-SWR-power spindles (concentration parameter 0.23 ± 0.14 compared to 0.13 ± 0.08 in cells with a decrease in modulation in cycles with high ripple power; $p = 0.03$, Mann-Whitney), without changes in the average preferred phase compared to the cells with decreased modulation (0.35 ± 1.64 rad for units with increased modulation, 0.19 ± 2.04 rad for units with decreased modulation; $p = 1$), or compared to the average preferred phase of the same units in low-ripple-power cycles (0.12 ± 1.67 rad; $p = 0.5$, Wilcoxon signed rank). In some of the units, like the example in

Figure 8, we observed a strong phase offset between SWRs and unit spikes (Figure 8b); however, this was not the case in all the units, and some had spindle phase preferences that overlapped with those of SWRs recorded in the same session.

The modulation of thalamic spikes with SWR power was stronger for unit pairs; we calculated the modulation index for the cross-correlation between thalamic pairs recorded in the same session and found that it increased by an average of 26.74 % in spindle cycles with high ripple power compared to low ripple power (average index value 2.11 ± 1.70 in low-ripple cycles, 2.51 ± 1.60 in high-ripple cycles; $p = 9.77 \times 10^{-4}$, Wilcoxon signed rank; n = 12 pairs), which points to an enhanced co-activation of thalamic cells within spindle cycles that engage hippocampal SWRs.

If thalamic modulation is enhanced with high-SWR-power spindles, it is possible that SWRs that occur within spindle cycles are selectively coordinated with thalamic units. At broader time scales (seconds), we had found an overall decrease in thalamic firing and increase in bursting with SWRs (Figure 3). However, it is possible that only spindle-nested SWRs are correlated with thalamic units. To test this possibility, we calculated the cross-correlations of thalamic units with SWRs that occurred within the time window of detected spindle cycles, and compared them to a sample of equal number of SWRs that did not occur within spindle cycles. We find that the average firing of thalamic cells is significantly higher near SWRs that occurred within spindle cycles (mean firing 3.86 ± 3.48 spikes/s with spindle-nested SWRs, compared to 1.94 ± 1.66; $p < 10^{-5}$, Wilcoxon signed rank). However, a precise temporal correlation between SWRs and thalamic spikes was unclear (Figure 8c), suggesting that although thalamic firing is higher with spindle-nested SWRs, there is no systematic time relation between SWRs and the firing of thalamic units. Instead, the spindle oscillation correlates with an enhancement of both thalamic and CA1 firing, and it is the phase of the spindle cycle that provides further organization.

Multi-region correlations after spatial exploration

To explore the possibility that awake behavior influences the multi-region coordination and thalamic contribution, we analyzed 6 sessions in which the rats (n = 3; 2 sessions from each rat) explored a radial maze (a 4-arm maze in one rat, 8-arm maze in the other two). The animals explored the maze freely for 30 mins, collecting pellet rewards at the end of each arm. In these sessions, we recorded an average of 52 mins of sleep before and 78 mins of sleep after spatial exploration to compare the non-REM sleep events and their correlation with single units before and after spatial experience. All of the sessions showed an increase in the amplitude (peak-to-trough) and rate (per second) of SWRs during sleep after the maze exploration, as well as an increase in the amplitude of KCs (p = 0.03, Wilcoxon signed rank), while spindles did not change significantly in number or amplitude (p > 0.5).

Although the number and amplitude of detected spindle cycles did not change, there was a significant increase in CA1 SWR power within spindle cycles (p = 0.03, Wilcoxon signed rank), consistent with the overall enhancement of SWR activity in CA1. To find out if there is selective activation of behavior-relevant CA1 cells during spindles, we classified CA1 units recorded in these sessions (n = 43) into those that were active (n = 28) or not (n = 15) when the animal was on the maze (based on visual inspection during spike sorting, i.e., unit clusters present both on the maze and during sleep or only in subsequent sleep; examples from one tetrode in Supplementary Figure 9a). These two groups of CA1 units did not differ in firing rate in the sleep post-behavior (calculated as the mean number of spikes over the total sleep time, or as the reciprocal of the average inter-spike interval; p > 0.1, Mann-Whitney U-test). However, units that were active during maze exploration were more likely to be active within spindle cycles than units not active on the maze (p = 0.0075, Mann-Whitney U-test for the average number of spikes in spindle cycles and p = 0.024 for the percent of spindle cycles with unit spikes), suggesting that cells that are engaged in spatial exploration are specifically correlated with thalamocortical oscillations. Furthermore, in the sleep after behavior, the peri-SWR histograms
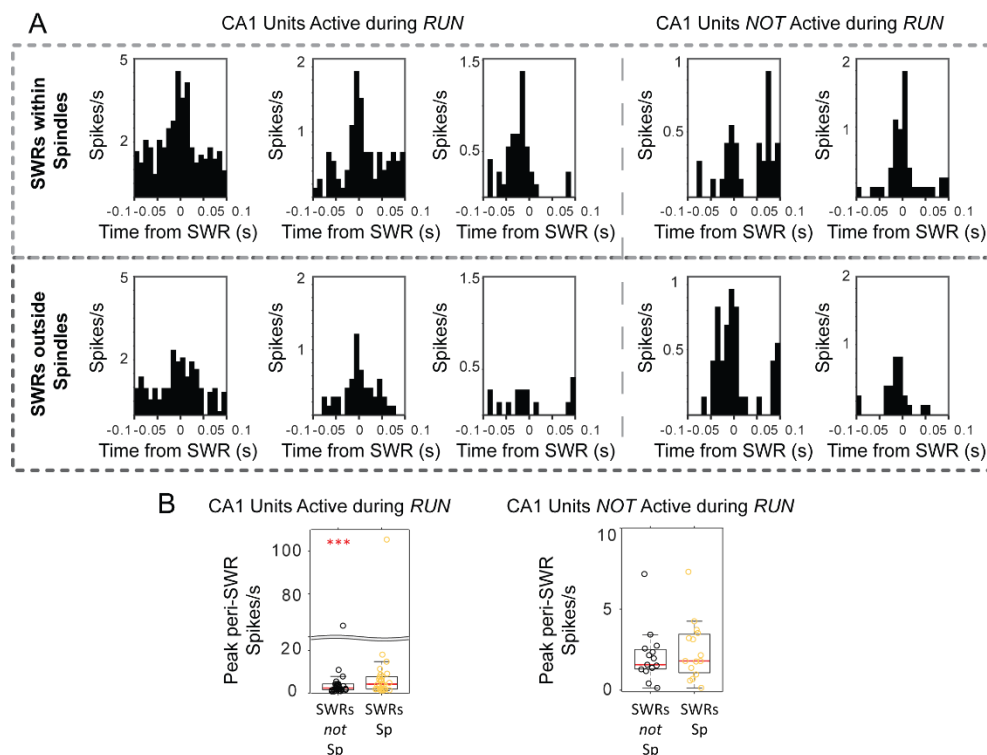

Figure 9. CA1 units active during 'run' show stronger correlations with SWRs nested within spindle cycles. (A) peri-SWR histograms of the spikes of 3 CA1 units active during exploratory behavior (left) and 2 CA1 units recorded by the same tetrode only active during the sleep after maze exploration (right); top plots, with SWRs that occurred nested in spindle cycles; bottom plots, with SWRs that were not nested in spindles (sorted clusters for all these units are shown in Supplementary Figure 7a). (B) Distribution of the peak firing in the peri-SWR histograms for all CA1 units that were active during run (left) or active in the sleep post-run (but not during behavior; right), separated by SWRs nested or not within spindle cycles (SWRs Sp vs. SWRs not Sp respectively).

of CA1 cells that had been active during run showed significantly higher peaks when calculated with SWRs that were nested in spindle cycles, compared to peri-SWR histograms calculated with a sample of an equal number of SWRs not coupled to spindles (p < 10^{-4}; Wilcoxon signed rank). Instead, peri-SWRs histograms of CA1 units that were not active during behavior were not different between spindle-coupled and spindle-uncoupled SWRs (p = 0.09; Wilcoxon signed rank). An example of peri-SWR histograms for units recorded by one tetrode in CA1 is shown in Figure 9. The panels illustrate the stronger correlations of SWRs nested in spindle cycles (top row in Figure 9a) for units that were active during behavior, compared with the correlations of the same units with SWRs that were not nested in spindle cycles (bottom row panels). The units that were not active during run had similar peri-SWRs regardless of the coupling with spindles (Figure 9b).

We then characterized post-exploration changes in thalamic activity, and did not find significant changes induced by maze exploration in this sample of thalamic units (n = 9). Figure 10a shows an example of a thalamic unit with a post-KC rebound, which had a similar profile before and after behavior. Thalamic units also had similar correlations with mPFC spindle cycles before and after behavior (Figure 10b), with no change in spindle phase distribution mean or concentration values (p > 0.3, Mann-Whitney U-test), although the modulation index estimated from peri-spindle-trough spike correlograms was on average higher after experience (0.21 ± 0.19 before exploration, 0.37 ± 0.31 after exploration; p = 0.02). This is in contrast to an increase in the spindle correlations with SWRs and with CA1 units that were active on the maze. Figure 10c-d, show examples of CA1 spindle-correlations from the same session in which the thalamic unit in Figure 10b was recorded. Although the peri-spindle-trough histograms for the thalamic unit are similar before and after exploration (or even show a slight decrease in modulation), the CA1 unit (which was active during run) and the SWRs show enhanced modulation with spindles post-exploration. On average, 60 % of the CA1 units fired more spikes in spindle cycles after exploration, and fired in a larger proportion of spindle cycles (distributions for all the CA1 units in Figure 10e).
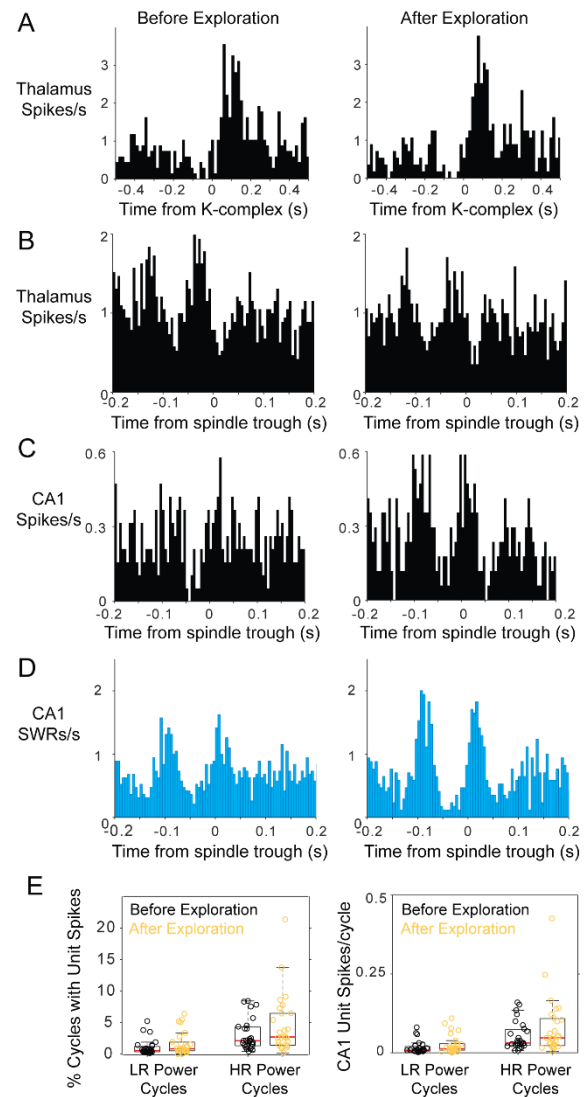
Figure 10. Modulation of thalamic and CA1 activity before and after free exploration on a radial-arm maze. Peri-event histograms display fairly stable modulation of spiking activity in the thalamus with K-complexes before and after spatial exploration; KC-triggered histograms of thalamic spikes are shown in (A) and peri-spindle histograms in (B); instead, a CA1 unit recorded in the same session increases its modulation with spindles (C) in the sleep following behavior. (D) Spindle-triggered SWR histograms show an increase in SWR modulation following behavior. (E) Distributions of the % of spindle cycles with CA1 unit spikes and the average number of CA1 spikes/spindle cycle before (black) and after exploration (yellow); each circle is one CA1 unit; most CA1 units increased their contribution to spindles post-behavior.

We tested the possibility of selective modulation of thalamic units with spindle cycles that engage CA1 units that were active during maze exploration (compared to spindle cycles with spikes from CA1 units that were not active during behavior). We calculated the correlation of thalamic spikes with spindle cycles with spikes from the CA1 units that had been active on the maze, and compared it with their correlation with spindle cycles with spikes from CA1 units that were not active during exploration. We found no significant differences between the peak or the average firing values of the cross-correlograms between the two types of spindles and thalamic units (p > 0.3, Wilcoxon signed rank), nor any differences between the mean number of thalamic spikes per spindle cycle or the percent of spindle cycles with spikes from thalamic units (p > 0.1).

Another possibility is that thalamic cells specifically change their modulation with spindles when CA1 units are reactivating representations of spatial trajectories that were experienced while the animal was on the maze. During exploratory behavior, CA1 cells with neighboring place fields become active as part of theta sequences, showing strong phase-locked activation in the theta band when the animal is running (Supplementary Figure 9b); these units then tend to be co-active in resting periods that follow behavior, often during SWRs[11,36]. We selected pairs of CA1 units that had a clear co-modulation during run (n = 19 CA1 pairs; examples in Supplementary Figure 9b), and calculated the correlation of thalamic units with spindles in which these pairs were active, compared to cycles in which they were not, and found no difference in thalamic activity (p > 0.9). Because the sample of thalamic units in these sessions was small (n = 9), we repeated these analyses with all the background spikes (MUA) recorded by thalamic tetrodes, finding no significant changes (p > 0.1).

In summary, these analyses suggest an increased correlation between recently active CA1 units and mPFC spindles but failed to demonstrate a selective modulation of thalamic units with mPFC spindles with behaviorally relevant CA1 unit activation. It is possible that studying a larger sample of thalamic units, or after behavioral tasks with stronger memory demands, will reveal task specific modulations between these three brain regions. Alternatively,

these results indicate that there is a selective modulation of task-relevant CA1 cells with spindles, while maintaining and overall similar background of at least some thalamic cell activity. This could selectively enhance signal to noise transmission to promote CA1-driven synaptic plasticity in mPFC networks during SWR reactivation, to facilitate the preferential consolidation of hippocampus-neocortical compared to thalamo-cortical ensemble activity.

Discussion

Here we report on the temporal correlations between cells in the limbic thalamus and the hallmark oscillations of non-REM sleep. We recorded single units and LFPs from functionally connected regions in the mPFC, CA1, and midline nuclei of the thalamus, including reuniens, mediodorsal and ventromedial nuclei. We found that thalamic cells in these nuclei often reduce their firing and are more bursty with CA1 SWRs during non-REM sleep. Although firing in the thalamus becomes sparse, spikes are correlated with other thalamic cells and with LFP population events in mPFC and CA1. Specifically, thalamic units were phase-locked to the delta and spindle frequency bands recorded in deep layers of mPFC, and some thalamic cells showed a strong rebound of activity following the down-state of the mPFC slow oscillation (1–4 Hz). In addition, individual thalamic cells rarely contributed spikes in consecutive cycles of the spindle oscillation, but showed consistent correlations with other thalamic cells within the period of spindle cycles. We also found that CA1 SWRs were strongly correlated with individual cycles of mPFC spindles, a correlation that was enhanced following spatial exploration; furthermore, CA1 units that were active during behavior became preferentially active during SWRs nested in spindle cycles, and coinciding with a relative increase in background thalamic firing. The evidence raises the hypothesis that sparse synchronicity in thalamic cell ensembles facilitates CA1-mPFC consolidation processes during non-REM sleep. The enhancement of ripple-spindle correlations after exploratory behavior and preferential engagement of recently active CA1 units in SWRs nested in spindles, further suggest that individual spindle cycles provide time windows in which the interleaved activation of thalamic cells can

promote incremental synaptic plasticity and facilitate the robust integration of recently acquired traces into neocortical networks.

Thalamic inhibition during hippocampal SWRs

The hypothesis of a multi-region interaction that stabilizes and integrates recently acquired episodic memory representations into neocortical schemas is supported by a stream of reports that point to the involvement of the thalamus[9,15–17,21,27,52,53]. However, the investigation of the role of thalamocortical cells in this processes has lagged behind; evidence from unit recordings in naturally sleeping animals is still fairly limited, and we have been lacking simultaneous recordings of LFPs and single units from these brain regions to study the dynamics of their correlations in the same animal. Current results point to a negative correlation between thalamic activity and CA1 SWRs, which is suggested by the decrease in fMRI bold signal in the thalamus of monkeys[27] and the reduction of firing in MD and RE thalamic cells with SWRs[26,28]. Our results extend these reports with the observation of an increase in thalamic bursting, which suggests that inhibition of the thalamus (as opposed to a decrease in excitatory drive) explains the negative correlations between hippocampal and thalamic activity. One possibility is that the GABAergic TRN inhibits the thalamus near the time of SWRs. It is also possible that changes in neuromodulatory tone during non-REM sleep could lead to the hyperpolarization of cells in the dorsal thalamus in coordination with a change in CA1 state that facilitates SWR occurrence. The finding that cells in all the thalamic nuclei we recorded from were coordinated with CA1 and mPFC suggests a non-specific process that may be under neuromodulatory and state control, without ruling out the involvement of the TRN. Because RE projects directly to CA1 and to the entorhinal cortex[54], a change in firing mode in RE cells that project to hippocampus could further influence CA1 state transitions and SWRs occurrence.

Thalamic cells with an increase in firing with SWRs were rare in our sample (~15 %), although we found a significant relative increase in cell firing when SWRs were locked to spindles. A recent study reported similar results although with different ratios, finding about 40 % of MD multi-unit activity increase
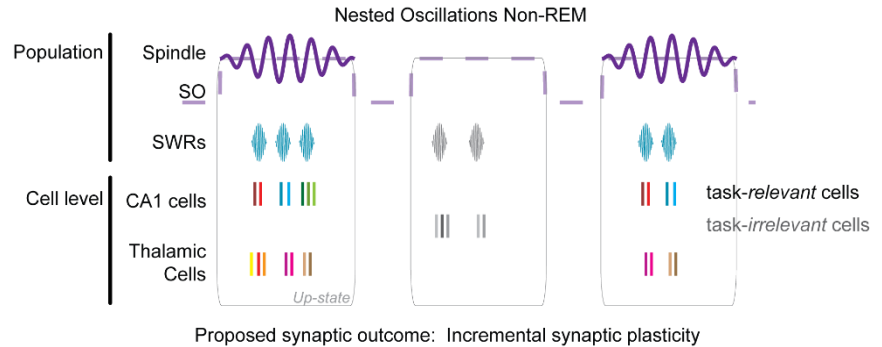
Figure 11. Model of spindle organization of thalamic and CA1 spike dynamics. *Task-relevant CA1 cells are preferentially engaged in SWRs locked to the spindle oscillation, coinciding with a relative increase in thalamic firing. The activation of different combinations of thalamic cells in subsequent spindle cycles could favor a broader range of synaptic associations at the neocortical level, while the sparsity of firing could ensure incremental synaptic changes and promote continuous learning. Each color in the spike rasters represents spikes from a different cell. SO = slow oscillation.*

with SWRs that were coupled to spindles[28]. In this study, spindles were detected from EEG skull screws using criteria that included at least 0.5 s of duration; spindle-coupled SWRs were then classified as those SWRs that occurred between the onset and offset of spindles. Our LFP recordings in mPFC show that time windows longer than 0.5 s are likely to encompass one or more KCs, which are correlated with SWRs and often followed by a thalamic rebound of firing. In rodents, spindles defined by a duration criterion that overlaps with the delta oscillation are likely to reflect KC-SWRs correlations (in addition to thalamo-SWRs correlations), and could increase the fraction of thalamic electrodes positively modulated with SWRs. The direct recording of LFP and KCs together with the analysis of shorter, within-spindle, timescales helps disambiguate the multi-oscillation correlations.

Computationally, higher order thalamic nuclei (which include those in the midline[55,56]) have been proposed to gate communication between neocortical areas. This 'gate' may be implemented through direct control of cortical synchronization and effective connectivity[57–59], and through the regulation of the signal-to-noise ratio in resting states[28]. The broad

decrease in thalamic activity at the time of potential memory reactivation in CA1, could facilitate effective information transfer between hippocampus and neocortex by reducing interference from the main input (thalamic) to neocortical areas. The signal-to-noise ratio may be particularly favorable to the transmission of novel traces between CA1-mPFC during spindles, when we find a selective enhancement of correlations with recently active CA1 cells. Although there is a selective relative increase in thalamic firing during spindles, the spindle oscillation may provide a protected functional window for select memory consolidation computations, for example those required to extract statistical regularities and organize synaptic plasticity. We find that there is an increase in thalamic bursting near SWRs, a firing mode that is more effective at activating post-synaptic cortical cells[42]. The function of thalamic bursting is not fully understood, but this thalamic response mode promotes better detectability of stimuli during wakefulness[60–62], and has been proposed to serve as a detection signal that can facilitate plasticity and learning in thalamocortical networks[63,64].

Functional windows for multi-region coordination in non-REM sleep

The up-states of the non-REM slow and delta oscillations are thought to provide discrete and relatively broad windows of hundreds of miliseconds in which hippocampal-neocortical interactions can occur[65,66]. Hippocampal SWRs happen mainly during up-states and nested within the cycles of spindles, and spindles are themselves largest in amplitude following the transition from the down-to-up state[17,67]. Thalamic cells often showed a rebound in firing after mPFC KCs (LFP markers of the slow oscillation trough), suggesting that SWRs that occur during the down-to-up state transition overlap in time with an increase in thalamic drive. The thalamic cells with post-KC rebound could influence CA1-mPFC interactions by influencing the dynamics within the slow oscillation up-state. Several reports have shown that subsets of neocortical and TRN cells are preferentially active at the start or at the end of neocortical slow oscillation up-states, although it is unclear what drives this differential activation[7,68–70]. Our data raise the possibility that cells in the dorsal thalamus (which can drive neocortical and TRN cells) could provide excitation early on in the slow oscillation up-state and influence the ensuing dynamics[8]. Cells in the thalamic nuclei that we recorded from project densely to superficial layers of the neocortex, where they can influence deeper layers through their synapses on the dendritic tufts of pyramidal cells or via layer I interneurons[31,71].

At finer time scales, individual SWRs events have durations that are strikingly similar to the period of the spindle oscillation (close to 100 ms), within the temporal range of spike-timing dependent synaptic plasticity[72,73]. Indeed, SWR- or spindle-like stimulation patterns induce LTP respectively in CA3-CA1[73] and in neocortical synapses[74]. These cell ensemble and circuit level oscillations and the nesting of SWRs in the spindle cycles can therefore induce the long-term synaptic remodeling behind memory consolidation. Both synaptic enhancement and weakening have been suggested to occur in non-REM sleep and contribute to consolidation[75–80]. Although our preparation does not address the synaptic level, our results demonstrate that CA1 cells that were active during exploratory behavior have enhanced correlations with SWRs nested in mPFC spindle cycles. A selective engagement of task-specific cells during SWR-spindles, in temporal windows that can promote synaptic plasticity, may explain the increase in memory following spindle-like stimulation[21], and suggests that spindle oscillations provide windows for synaptic reorganization of recently acquired memories. We observed an increase in spindle-CA1 correlations after behavioral experience even in the absence of significant changes of spindle-thalamic correlations. It is possible that behavioral tasks with stronger memory demands would reveal a modulation of thalamic cells in addition to the observed CA1 modulation. Nonetheless, the differential enhancement of CA1-mPFC relative to thalamo-mPFC correlations could contribute to differential synaptic plasticity changes in these two pathways[77,79]. Spindles could provide a thalamocortical mechanism for the selective modification of CA1-mPFC synapses following behavioral tagging.

Sparse firing of thalamic cells during spindles and its role in systems memory consolidation

Sparse firing of thalamocortical cells during spindles has been reported before[39,39,81–84], but it was

unclear how multiple cells in the dorsal thalamus would correlate with each other, and its relevance for the systems consolidation of memory has not been discussed. We find that pairs of thalamic cells are consistently phase-locked to the mPFC LFP within the period of spindle oscillations, suggesting that individual spindle cycles group and organize the sequential activation of thalamocortical ensembles. The repeated activation of thalamic cell ensembles through non-REM sleep could promote spike-timing dependent synaptic plasticity. Importantly, the sparse firing at the single cell level will allow for the interleaved excitation of different sets of thalamocortical synapses in subsequent spindle cycles, and in coordination with the reactivation of recently active CA1 place cells. The activation of varying combinations of thalamocortical synapses with CA1 SWRs that engage recently active cells (Figure 11) may promote a broad range of synaptic associations at the neocortical level, make the neocortical trace more robust to noise, and facilitate generalization. Critically, the sparse activation could prevent plasticity saturation[85] and contribute to gradual synaptic changes that would allow for extended computations[66] and facilitate continuous learning. One of the challenges in the development of artificial neural networks, has been to overcome the catastrophic interference that can occur during sequential training. While real-world settings require continuous learning, artificial network training algorithms are not yet optimized for these conditions. Sparsity and controlled weight changes have been suggested to facilitate robust representations and learning[86–89]. Future studies of thalamocortical sleep dynamics can provide clues on the biological implementation of continuous learning algorithms and help improve artificial networks and their applications.

Behaviorally, the systems consolidation of episodic memory leads to the stabilization of memories and to the formation of mental models or schemas that have higher adaptive value for the organism than the original details of the experience[2,3]. The oscillatory interactions between hippocampus and neocortex during sleep support the extraction of statistical consistencies and integration of hippocampal memory traces into gist-like neocortical schemas that are applicable beyond the specific conditions in which a memory was acquired[6,66]. Sparse thalamocortical population synchronicity during spindles may therefore promote incremental synaptic changes that optimize the interleaved extraction and incorporation of implicit information from recently acquired hippocampal memory traces into the neocortical representations.

It is worth noting that in certain states, such as anesthesia or epilepsy[90], cells in the thalamus may fire at the population rhythm, and sparse synchronicity in the thalamus might be a distinguishing feature of normal resting states[91]. The disruption of thalamocortical oscillations has been proposed as a pathophysiological model for several neuropsychiatric disorders[92,93]. Oscillatory stimulation (mimicking the abnormally increased delta rhythm in schizophrenia) of thalamic projections to CA1 disrupts working memory in rats[94], and spindle irregularities (reduced or abnormally enhanced spindles) have been observed in children with neurodevelopmental disorders[95], in schizophrenia[96], and in absence epilepsy[97]. A careful balance in the excitatory and inhibitory thalamic networks is required to generate the appropriate population level synchronicity and function. Studying spindle dynamics at the cellular and population level will thus be critical to understand the pathophysiology behind these disorders, and to design therapeutic interventions that preserve normal synchronicity in the thalamus as well as the multi-region coordination that underlies cognitive function.

Conclusion

Memory consolidation promotes the stabilization and incorporation of memory traces initially stored in the hippocampus into neocortical networks, contributing to the development and training of neocortical models that allow adaptive behavior. Recording from naturally sleeping rats, we present results that suggest that the individual cycles of the spindle oscillation provide a key processing time window that selectively engages recently active CA1 cell ensembles, as well as subsets of cells in thalamocortical networks. The rebound activation of thalamic cells following down-states could facilitate up-state dynamics. As the cycle of the slow oscillation progresses, the selective enhancement of spindle-CA1 activity relative to spindle-thalamic

activity following spatial experience could bias transmission in CA1-mPFC compared to thalamo-mPFC pathways. Lastly, the sparse and interleaved activation of groups of thalamic cells during spindles may provide a critical mechanism to facilitate the continuous integration of novel hippocampal memories into neocortical schemas for generalization and lifelong learning.

Competing interests: None.

Materials and Methods

We used 8 male Long-Evans rats (400–500 g) in the experiments reported here. Animals were housed individually, provided with food and water ad libitum, and monitored in a temperature-controlled room with a 12 h light/dark cycle (lights on/off at 7:00 am/7:00 pm). These experiments were approved by the Committee on Animal Care at the Massachusetts Institute of Technology and conform to US National Institutes of Health guidelines for the care and use of laboratory animals.

Electrode Arrays, Surgical Implantation and Electrophysiology Recordings

Multielectrode arrays (microdrives) with up to 21 nichrome wire tetrodes, 18 tetrodes for recording and 3 references, one per brain region (individual tetrode wires were 12.5 μm in diameter). The microdrives were prepared according to standard procedures in the laboratory, and the base was modified from previous designs (Kloosterman et al., 2009; Nguyen et al.,

2009) to target the three regions of interest. A diagram of a typical electrode configuration is shown in Figure 1a. The tetrodes spanned the following coordinates from Bregma (Paxinos and Watson atlas, 1986): Anterior 3.65 mm to 2.4 mm for mPFC, posterior 1.4 to 3.2 mm for the thalamus, and P3.1 to 4.4 mm for dorsal CA1. The arrays for mPFC and thalamus were positioned at 16 degrees from the vertical axis to prevent damage to the sagittal sinus. Sterile surgery procedures were performed for chronic microdrive implantation; anesthesia was induced by an intraperitoneal injection of a solution of ketamine (25 mg/kg), xylazine (3 mg/kg) and atropine (0.027 mg/kg), and maintained with 1-2 % inhaled isoflurane. Up to 8 bone screws were secured to the skull for support; three craniotomies were drilled over the target coordinates and the dura mater membrane was removed to allow for tetrode penetration. During surgery, the most anterior thalamic electrode was placed above P1.4 mm and lateral 1.8 mm, and the linear thalamic array was aligned parallel to the sagittal suture; this positioned the rest of the electrodes over their corresponding craniotomies. The implant was secured to the skull with dental cement after surrounding the exposed craniotomies and tetrodes with silicon grease to keep them from being fixed by the cement. Animals were preemptively injected with analgesics (Buprenorphine, 0.5–1 mg/kg, subcutaneous), and monitored by the experimenter and veterinarian staff for three days post-surgery.

The tetrodes were gradually advanced to the target depths over the course of 1.5–2 weeks before recordings started; the slow advancement of electrodes facilitated stable unit and local field potential (LFP) recordings over several hours. Once the target depth was reached, recordings were performed if units were present, and electrodes were lowered further in non-recording sessions to search for new units and allow for electrode drift before the next recording session. Recording sessions typically lasted at least 3 hours, during which the animal was in a quiet squared enclosure of about 50×50 cms, were they cycled through several bouts of sleep and wakefulness. Spikes were acquired at 32 kHz after the signal from any of the tetrode channels crossed a preset threshold. Individual units were isolated by manual clustering on peak spike waveform amplitudes across all channels using custom software (Xclust; M.A.W.). LFPs were sampled at 600 Hz after

anti-aliasing filtering and downsampling from up to 3.125 kHz. Two infrared LEDs (sampled at 30 Hz) attached to the headstages of the acquisition system (Neuralynx) were used to track the rat's position and estimate velocity.

Spatial exploration in the radial maze. In preparation for the 6 behavioral sessions, 3 of the implanted rats were food restricted for at least 5 days before behavior started. In each session, rats explored a radial maze (with 4 arms for one rat and 8 arms for the other two rats) that was surrounded by consistent visual cues on the walls of the room; rats retrieved one 45 mg sucrose pellet every time the animal reached the end of the arm (provided by the experimenter). The behavioral exploration (two sessions per animal) lasted 30 min, and we also recorded at least 30 mins of sleep pre- and post-behavior. All the rats had less than 7 days of experience exploring the maze at the time of these recording sessions. For one of the rats, the two behavioral sessions were days 1 and 2 of exploration (no previous maze experience before day 1), for rat 2, they were days 2 and 4, and for the last rat they were days 3 and 6. Therefore, all the recordings were performed during early experience in the mazes, when memory traces are expected to be dependent on the hippocampus for retrieval (Varela, 2016). We did not observe differences between the first and second session in each rat, and the data were pooled together.

Selection of Electrodes and Sessions for Analyses

Sessions were selected based on having at least one unit in two of the three regions of interest, clear sleep events (KCs, spindles, SWRs) in the neocortical and hippocampal LFPs, and clear histological confirmation of thalamic tetrode location. While CA1 electrode location can be confirmed via electrophysiological LFP features alone, we find that LFP signals alone do not reliably allow specific localization within the nuclei of the dorsal thalamus and histological confirmation is necessary. For arrays with independently moving tetrodes, this requires the identification of electrode lesions for all the tetrodes in the array to prevent ambiguity when matching signals to electrode position. Thus, for these experiments we only include data from rats in which we successfully confirmed the histological location of all implanted thalamic (and mPFC) tetrodes, and CA1 location was confirmed electrophysiologically. The location of electrodes used for analyses is reported in Supplementary figures 1 and 2. To prevent repeated counts of the same units across sessions, unit analyses include only units for which 1) the tetrode had been moved from the previous recording session at least 80 µm or 2) the amplitude projections used for spike sorting had different profiles compared to the preceding recorded session.

LFP State and Event Detection

For each rat, we selected one wire from one tetrode with clear sleep events (determined by visual inspection) for LFP analyses (one tetrode per brain region). To detect reference events for analyses, the three LFPs were filtered (with zero-phase distortion) in the delta (1-4 Hz), spindle (6-14 Hz), and ripple (100-275 Hz) bands using a band-pass finite impulse response filter designed with a blackman window and filter order 6 times the sampling rate/bandwidth ratio.

We detected reference events during states in which the power of both the delta, in the mPFC LFP, and ripple bands, in CA1, were above the mean for the session for at least two minutes; animal velocity and behavior during these time periods confirmed that the animal was immobile during detected offline states (figure 1). These electrophysiological and behavioral traits are defining features of non-REM sleep. The position information to calculate velocity was obtained from the LEDs attached to the headstages; gaps in the position information (due to occlusion of the LEDs) shorter than 300ms were linearly interpolated using nearby values; gaps longer than 300ms were not considered for further analysis. The space derivative was used to calculate velocity and then smoothed (Gaussian kernel, 250 ms standard deviation) and linearly interpolated to up-sample and match the 600 Hz of the LFP.

Non-REM periods detected with these combined criteria of high delta and SWR power, showed large transient deflections in the mPFC LFP, or K-complexes (KCs). To detect KCs, we first rectified the filtered (1–4 Hz) LFP (positive values were made equal to 0), the resulting trace was squared and KCs detected as local maxima, when the amplitude was above the mean plus 4 standard deviations (mean and

standard deviation calculated from the LFP in quiet states). We detected spindle troughs as local minima, 3 standard deviations below the mean of the spindle-band filtered LFP (6–14 Hz). The SWR detection algorithm detected times when the squared, filtered LFP (100–275 Hz) had an amplitude above the mean plus 3 standard deviations for at least 20 ms (mean and standard deviation calculated for the LFP when the animal was quiet). If two SWRs were closer than 20ms they were considered a single ripple event. The SWR timestamp was selected as the time with the largest absolute value in the ripple filtered LFP.

Correlation quantifications: Peri-event histograms and generalized linear model

We used peri-event histograms (PETHs) to quantify the correlations between sleep events (trough of KCs, spindles, peak of ripples) and the unit activity in each brain region. To compare across sessions and animals, we quantified the change (in units of standard deviation) of the event-triggered average with respect to a null distribution calculated through a permutation test. Specifically, we first calculated the average peri-KC, peri-spindle and peri-SWR histogram across detected events in the recording session (bin 15 ms, followed by smoothing with a Gaussian kernel with 15 ms standard deviation). All the event timestamps were then shifted by a number between 1.5–2.5 s (randomly chosen with uniform probability) to offset the event times with respect to the local spikes, while preserving the inter-event time intervals. The average PETH (15 ms bins, Gaussian smoothed) was calculated from the shifted event timestamps, and this procedure was repeated 100 times to generate a distribution of expected PETHs under the null hypothesis of no correlation between sleep events and local activity (Supplementary Figure 3).

To characterize the relation between thalamic firing and SWR rate more precisely, we implemented generalized linear models (GLM) of CA1 spikes and SWRs as a function of thalamic spikes. Multi-unit spikes from CA1 and the thalamus, and CA1 SWRs, were binned through the non-REM sleep periods in each session (bin 250 ms). A GLM was fit in matlab (equation 1) to estimate the log of counts of spikes or SWRs in CA1 (μ) as a linear function of the spikes in the thalamus (X), assuming a Poisson distribution.

From the fitted model, we calculated the estimated change in the response variable (firing in CA1) per unit change of the predictor (firing in thalamus) from the equivalent equation (2). The Wald test was used to test the significance of the thalamic parameter ($\beta 1$).

$$\log(\mu) = \beta 0 + \beta 1 X \quad (1) \qquad \mu = \exp(\beta 0 + \beta 1 X) \quad (2)$$

Phase-locking analyses

We used some of the phase analyses methods as in Siapas et al., 2005, and matlab´s circular statistics toolbox for quantification. We estimated the instantaneous phase using the Hilbert transform of the mPFC LFP filtered in the delta and spindle bands. Because asymmetry in the LFP shape can bias the phase distributions, the distribution of phases at the time of spikes or SWRs was normalized by the distribution of phases for the full delta or spindle-filtered LFP during non-REM sleep. After normalization, we used the Rayleigh test to determine if the angular phase at the spikes or LFP events timestamps were significantly different from a uniform distribution. A fit to the Von Misses distribution was then used to estimate the mean vector direction (mean preferred phase), and the degree of phase-locking from the concentration parameter *kappa*. The concentration parameter is a reciprocal measure of dispersion and increases as the density of values near the mean of the distribution increases. We also estimated phase based on linear interpolation between LFP peaks and troughs (arbitrarily assigned $-\pi$ and π values; equivalent to the 'extrema' method in Siapas et al., 2005); we did not find differences in the phase-locking based on either method and the results are reported for phase estimates based on the Hilbert transform.

References

1. Marr D. Simple memory: a theory for archicortex. Philos Trans R Soc Lond B Biol Sci. 1971 Jul 1;262(841):23–81. PMID: 4399412

2. McClelland JL, McNaughton BL, O'Reilly RC. Why there are complementary learning systems in the hippocampus and neocortex: insights from the successes and failures of connectionist models of learning and memory. Psychol Rev. 1995 Jul;102(3):419–457. PMID: 7624455

3. Kumaran D, Hassabis D, McClelland JL. What Learning Systems do Intelligent Agents Need? Complementary Learning Systems Theory Updated. Trends Cogn Sci. 2016 Jul;20(7):512–534. PMID: 27315762

4. Yu JY, Liu DF, Loback A, Grossrubatscher I, Frank LM. Specific hippocampal representations are linked to generalized cortical representations in memory. Nat Commun. 2018 07;9(1):2209. PMCID: PMC5992161

5. Morris RGM. Elements of a neurobiological theory of hippocampal function: the role of synaptic plasticity, synaptic tagging and schemas. Eur J Neurosci. 2006 Jun;23(11):2829–2846. PMID: 16819972

6. Moscovitch M, Cabeza R, Winocur G, Nadel L. Episodic Memory and Beyond: The Hippocampus and Neocortex in Transformation. Annu Rev Psychol. 2016;67:105–134. PMCID: PMC5060006

7. Gardner RJ, Hughes SW, Jones MW. Differential spike timing and phase dynamics of reticular thalamic and prefrontal cortical neuronal populations during sleep spindles. J Neurosci Off J Soc Neurosci. 2013 Nov 20;33(47):18469–18480. PMCID: PMC3834053

8. Neske GT. The Slow Oscillation in Cortical and Thalamic Networks: Mechanisms and Functions. Front Neural Circuits. 2015;9:88. PMCID: PMC4712264

9. Ji D, Wilson MA. Coordinated memory replay in the visual cortex and hippocampus during sleep. Nat Neurosci. 2007 Jan;10(1):100–107. PMID: 17173043

10. Wilson MA, McNaughton BL. Reactivation of hippocampal ensemble memories during sleep. Science. 1994 Jul 29;265(5172):676–679. PMID: 8036517

11. Davidson TJ, Kloosterman F, Wilson MA. Hippocampal replay of extended experience. Neuron. 2009 Aug 27;63(4):497–507. PMCID: PMC4364032

12. Buzsáki G. The hippocampo-neocortical dialogue. Cereb Cortex N Y N 1991. 1996 Apr;6(2):81–92. PMID: 8670641

13. Girardeau G, Benchenane K, Wiener SI, Buzsáki G, Zugaro MB. Selective suppression of hippocampal ripples impairs spatial memory. Nat Neurosci. 2009 Oct;12(10):1222–1223. PMID: 19749750

14. Ego-Stengel V, Wilson MA. Disruption of ripple-associated hippocampal activity during rest impairs spatial learning in the rat. Hippocampus. 2010 Jan;20(1):1–10. PMID: 19816984

15. Siapas AG, Wilson MA. Coordinated interactions between hippocampal ripples and cortical spindles during slow-

wave sleep. Neuron. 1998 Nov;21(5):1123–1128. PMID: 9856467

16. Sirota A, Csicsvari J, Buhl D, Buzsáki G. Communication between neocortex and hippocampus during sleep in rodents. Proc Natl Acad Sci U S A. 2003 Feb 18;100(4):2065–2069. PMID: 12576550

17. Peyrache A, Battaglia FP, Destexhe A. Inhibition recruitment in prefrontal cortex during sleep spindles and gating of hippocampal inputs. Proc Natl Acad Sci U S A. 2011 Oct 11;108(41):17207–17212. PMCID: PMC3193185

18. Steriade M, Domich L, Oakson G, Deschênes M. The deafferented reticular thalamic nucleus generates spindle rhythmicity. J Neurophysiol. 1987 Jan;57(1):260–273. PMID: 3559675

19. Contreras D, Destexhe A, Sejnowski TJ, Steriade M. Spatiotemporal patterns of spindle oscillations in cortex and thalamus. J Neurosci Off J Soc Neurosci. 1997 Feb 1;17(3):1179–1196. PMID: 8994070

20. Maingret N, Girardeau G, Todorova R, Goutierre M, Zugaro M. Hippocampo-cortical coupling mediates memory consolidation during sleep. Nat Neurosci. 2016 Jul;19(7):959–964. PMID: 27182818

21. Latchoumane C-FV, Ngo H-VV, Born J, Shin H-S. Thalamic Spindles Promote Memory Formation during Sleep through Triple Phase-Locking of Cortical, Thalamic, and Hippocampal Rhythms. Neuron. 2017 Jul 19;95(2):424-435.e6. PMID: 28689981

22. Purcell SM, Manoach DS, Demanuele C, Cade BE, Mariani S, Cox R, Panagiotaropoulou G, Saxena R, Pan JQ, Smoller JW, Redline S, Stickgold R. Characterizing sleep spindles in 11,630 individuals from the National Sleep Research Resource. Nat Commun. 2017 26;8:15930. PMCID: PMC5490197

23. Manoach DS, Stickgold R. Abnormal Sleep Spindles, Memory Consolidation, and Schizophrenia. Annu Rev Clin Psychol. 2019 May 7;15:451–479. PMID: 30786245

24. Slézia A, Hangya B, Ulbert I, Acsády L. Phase advancement and nucleus-specific timing of thalamocortical activity during slow cortical oscillation. J Neurosci Off J Soc Neurosci. 2011 Jan 12;31(2):607–617. PMCID: PMC3044867

25. Contreras D, Steriade M. Cellular basis of EEG slow rhythms: a study of dynamic corticothalamic relationships. J Neurosci Off J Soc Neurosci. 1995 Jan;15(1 Pt 2):604–622. PMID: 7823167

26. Lara-Vásquez A, Espinosa N, Durán E, Stockle M, Fuentealba P. Midline thalamic neurons are differentially engaged during hippocampus network oscillations. Sci Rep. 2016 Jul 14;6:29807. PMCID: PMC4944155

27. Logothetis NK, Eschenko O, Murayama Y, Augath M, Steudel T, Evrard HC, Besserve M, Oeltermann A. Hippocampal-cortical interaction during periods of subcortical silence. Nature. 2012 Nov 22;491(7425):547–553. PMID: 23172213

28. Yang M, Logothetis NK, Eschenko O. Occurrence of Hippocampal Ripples is Associated with Activity Suppression in the Mediodorsal Thalamic Nucleus. J Neurosci Off J Soc Neurosci. 2019 Jan 16;39(3):434–444. PMID: 30459228

29. Vertes RP, Hoover WB, Do Valle AC, Sherman A, Rodriguez JJ. Efferent projections of reuniens and rhomboid nuclei of the thalamus in the rat. J Comp Neurol. 2006 Dec 10;499(5):768–796. PMID: 17048232

30. Hoover WB, Vertes RP. Anatomical analysis of afferent projections to the medial prefrontal cortex in the rat. Brain Struct Funct. 2007 Sep;212(2):149–179. PMID: 17717690

31. Rubio-Garrido P, Pérez-de-Manzo F, Porrero C, Galazo MJ, Clascá F. Thalamic input to distal apical dendrites in neocortical layer 1 is massive and highly convergent. Cereb Cortex N Y N 1991. 2009 Oct;19(10):2380–2395. PMID: 19188274

32. Maviel T, Durkin TP, Menzaghi F, Bontempi B. Sites of neocortical reorganization critical for remote spatial memory. Science. 2004 Jul 2;305(5680):96–99. PMID: 15232109

33. Hoover WB, Vertes RP. Collateral projections from nucleus reuniens of thalamus to hippocampus and medial prefrontal cortex in the rat: a single and double retrograde fluorescent labeling study. Brain Struct Funct. 2012 Apr;217(2):191–209. PMID: 21918815

34. Varela C, Kumar S, Yang JY, Wilson MA. Anatomical substrates for direct interactions between hippocampus, medial prefrontal cortex, and the thalamic nucleus reuniens. Brain Struct Funct. 2014 May;219(3):911–929. PMID: 23571778

35. Varela C, Weiss S, Meyer R, Halassa M, Biedenkapp J, Wilson MA, Goosens KA, Bendor D. Tracking the Time-Dependent Role of the Hippocampus in Memory Recall Using DREADDs. PloS One. 2016;11(5):e0154374. PMCID: PMC4856306

36. Wilson MA, McNaughton BL. Reactivation of hippocampal ensemble memories during sleep. Science. 1994 Jul 29;265(5172):676–679. PMID: 8036517

37. Steriade M, Deschênes M, Domich L, Mulle C. Abolition of spindle oscillations in thalamic neurons disconnected from nucleus reticularis thalami. J Neurophysiol. 1985 Dec;54(6):1473–1497. PMID: 4087044

38. Steriade M, Nuñez A, Amzica F. Intracellular analysis of relations between the slow (< 1 Hz) neocortical oscillation and other sleep rhythms of the electroencephalogram. J Neurosci Off J Soc Neurosci. 1993 Aug;13(8):3266–3283. PMID: 8340807

39. Huguenard JR, McCormick DA. Thalamic synchrony and dynamic regulation of global forebrain oscillations. Trends Neurosci. 2007 Jul;30(7):350–356. PMID: 17544519

40. Perez-Reyes E, Cribbs LL, Daud A, Lacerda AE, Barclay J, Williamson MP, Fox M, Rees M, Lee JH. Molecular characterization of a neuronal low-voltage-activated T-type calcium channel. Nature. 1998 Feb 26;391(6670):896–900. PMID: 9495342

41. Gutierrez C, Cox CL, Rinzel J, Sherman SM. Dynamics of low-threshold spike activation in relay neurons of the cat lateral geniculate nucleus. J Neurosci Off J Soc Neurosci. 2001 Feb 1;21(3):1022–1032. PMID: 11157087

42. Swadlow HA, Gusev AG. The impact of "bursting" thalamic impulses at a neocortical synapse. Nat Neurosci. 2001 Apr;4(4):402–408. PMID: 11276231

43. Leresche N, Lambert RC. T-type calcium channels in synaptic plasticity. Channels Austin Tex. 2017 Mar 4;11(2):121–139. PMCID: PMC5398590

44. Lu SM, Guido W, Sherman SM. Effects of membrane voltage on receptive field properties of lateral geniculate neurons in the cat: contributions of the low-threshold Ca2+ conductance. J Neurophysiol. 1992 Dec;68(6):2185–2198. PMID: 1337104

45. Jahnsen H, Llinás R. Ionic basis for the electro-responsiveness and oscillatory properties of guinea-pig thalamic neurones in vitro. J Physiol. 1984 Apr;349:227–247. PMCID: PMC1199335

46. Jahnsen H, Llinás R. Electrophysiological properties of guinea-pig thalamic neurones: an in vitro study. J Physiol. 1984 Apr;349:205–226. PMID: 6737292

47. Buzsáki G, Anastassiou CA, Koch C. The origin of extracellular fields and currents--EEG, ECoG, LFP and spikes. Nat Rev Neurosci. 2012 May 18;13(6):407–420. PMCID: PMC4907333

48. Colgin LL. Oscillations and hippocampal-prefrontal synchrony. Curr Opin Neurobiol. 2011 Jun;21(3):467–474. PMCID: PMC3578407

49. Hasselmo ME, Stern CE. Theta rhythm and the encoding and retrieval of space and time. NeuroImage. 2014 Jan 15;85, Part 2:656–666.

50. Wilson MA, Varela C, Remondes M. Phase organization of network computations. Curr Opin Neurobiol. 2015 Apr;31:250–253. PMCID: PMC4375010

51. Siapas AG, Lubenov EV, Wilson MA. Prefrontal phase locking to hippocampal theta oscillations. Neuron. 2005 Apr 7;46(1):141–151. PMID: 15820700

52. Mölle M, Yeshenko O, Marshall L, Sara SJ, Born J. Hippocampal sharp wave-ripples linked to slow oscillations in rat slow-wave sleep. J Neurophysiol. 2006 Jul;96(1):62–70. PMID: 16611848

53. Staresina BP, Bergmann TO, Bonnefond M, van der Meij R, Jensen O, Deuker L, Elger CE, Axmacher N, Fell J. Hierarchical nesting of slow oscillations, spindles and ripples in the human hippocampus during sleep. Nat Neurosci. 2015 Nov;18(11):1679–1686. PMCID: PMC4625581

54. Vertes RP. Major diencephalic inputs to the hippocampus: supramammillary nucleus and nucleus reuniens. Circuitry and function. Prog Brain Res. 2015;219:121–144. PMCID: PMC4961211

55. Guillery RW, Sherman SM. Thalamic relay functions and their role in corticocortical communication: generalizations from the visual system. Neuron. 2002 Jan 17;33(2):163–175. PMID: 11804565

56. Varela C. Thalamic neuromodulation and its implications for executive networks. Front Neural Circuits. 2014;8:69. PMCID: PMC4068295

57. Saalmann YB, Kastner S. Gain control in the visual thalamus during perception and cognition. Curr Opin Neurobiol. 2009 Aug;19(4):408–414. PMCID: PMC3140205

58. Saalmann YB. Intralaminar and medial thalamic influence on cortical synchrony, information transmission and cognition. Front Syst Neurosci. 2014;8:83. PMID: 24847225

59. Jaramillo J, Mejias JF, Wang X-J. Engagement of Pulvino-cortical Feedforward and Feedback Pathways in Cognitive Computations. Neuron. 2019 Jan 16;101(2):321-336.e9. PMID: 30553546

60. Lesica NA, Stanley GB. Encoding of natural scene movies by tonic and burst spikes in the lateral geniculate nucleus. J Neurosci Off J Soc Neurosci. 2004 Nov 24;24(47):10731–10740. PMID: 15564591

61. Alitto HJ, Weyand TG, Usrey WM. Distinct properties of stimulus-evoked bursts in the lateral geniculate nucleus. J Neurosci Off J Soc Neurosci. 2005 Jan 12;25(2):514–523. PMID: 15647497

62. Ortuño T, Grieve KL, Cao R, Cudeiro J, Rivadulla C. Bursting thalamic responses in awake monkey contribute to visual detection and are modulated by corticofugal feedback. Front Behav Neurosci. 2014;8:198. PMCID: PMC4039069

63. Crick F. Function of the thalamic reticular complex: the searchlight hypothesis. Proc Natl Acad Sci U S A. 1984 Jul;81(14):4586–4590. PMCID: PMC345636

64. Varela C, Ahmad S. A dendritic mechanism for dynamic routing and control in the thalamus [Internet]. Cosyne Abstracts; 2019. Available from: http://cosyne.org/c/index.php?title=Cosyne_19

65. Destexhe A, Hughes SW, Rudolph M, Crunelli V. Are corticothalamic "up" states fragments of wakefulness? Trends Neurosci. 2007 Jul;30(7):334–342. PMCID: PMC3005711

66. Penagos H, Varela C, Wilson MA. Oscillations, neural computations and learning during wake and sleep. Curr Opin Neurobiol. 2017 Jun;44:193–201. PMID: 28570953

67. Gonzalez CE, Mak-McCully RA, Rosen BQ, Cash SS, Chauvel PY, Bastuji H, Rey M, Halgren E. Theta Bursts Precede, and Spindles Follow, Cortical and Thalamic Downstates in Human NREM Sleep. J Neurosci Off J Soc Neurosci. 2018 Nov 14;38(46):9989–10001. PMCID: PMC6234298

68. Puig MV, Ushimaru M, Kawaguchi Y. Two distinct activity patterns of fast-spiking interneurons during neocortical UP states. Proc Natl Acad Sci U S A. 2008 Jun 17;105(24):8428–8433. PMCID: PMC2448853

69. Fanselow EE, Connors BW. The roles of somatostatin-expressing (GIN) and fast-spiking inhibitory interneurons in UP-DOWN states of mouse neocortex. J Neurophysiol. 2010 Aug;104(2):596–606. PMCID: PMC2934925

70. Batterink LJ, Creery JD, Paller KA. Phase of Spontaneous Slow Oscillations during Sleep Influences Memory-Related Processing of Auditory Cues. J Neurosci Off J Soc Neurosci. 2016 Jan 27;36(4):1401–1409. PMCID: PMC4728733

71. Schuman B, Machold RP, Hashikawa Y, Fuzik J, Fishell GJ, Rudy B. Four Unique Interneuron Populations Reside in

Neocortical Layer 1. J Neurosci Off J Soc Neurosci. 2019 Jan 2;39(1):125–139. PMCID: PMC6325270

72. Buchanan KA, Mellor JR. The activity requirements for spike timing-dependent plasticity in the hippocampus. Front Synaptic Neurosci. 2010;2:11. PMCID: PMC3059701

73. Sadowski JHLP, Jones MW, Mellor JR. Sharp-Wave Ripples Orchestrate the Induction of Synaptic Plasticity during Reactivation of Place Cell Firing Patterns in the Hippocampus. Cell Rep. 2016 Mar 1;14(8):1916–1929. PMCID: PMC4785795

74. Rosanova M, Ulrich D. Pattern-specific associative long-term potentiation induced by a sleep spindle-related spike train. J Neurosci Off J Soc Neurosci. 2005 Oct 12;25(41):9398–9405. PMID: 16221848

75. Frey U, Morris RG. Synaptic tagging and long-term potentiation. Nature. 1997 Feb 6;385(6616):533–536. PMID: 9020359

76. Redondo RL, Morris RGM. Making memories last: the synaptic tagging and capture hypothesis. Nat Rev Neurosci. 2011 Jan;12(1):17–30. PMID: 21170072

77. Puentes-Mestril C, Roach J, Niethard N, Zochowski M, Aton SJ. How rhythms of the sleeping brain tune memory and synaptic plasticity. Sleep. 2019 Apr 26; PMID: 31100149

78. Seibt J, Frank MG. Primed to Sleep: The Dynamics of Synaptic Plasticity Across Brain States. Front Syst Neurosci. 2019;13:2. PMCID: PMC6367653

79. Tononi G, Cirelli C. Sleep and synaptic down-selection. Eur J Neurosci. 2019 Jan 5; PMID: 30614089

80. Langille JJ. Remembering to Forget: A Dual Role for Sleep Oscillations in Memory Consolidation and Forgetting. Front Cell Neurosci. 2019;13:71. PMCID: PMC6425990

81. Steriade M, Deschenes M. The thalamus as a neuronal oscillator. Brain Res. 1984 Nov;320(1):1–63. PMID: 6440659

82. von Krosigk M, Bal T, McCormick DA. Cellular mechanisms of a synchronized oscillation in the thalamus. Science. 1993 Jul 16;261(5119):361–364. PMID: 8392750

83. Crunelli V, Lőrincz ML, Connelly WM, David F, Hughes SW, Lambert RC, Leresche N, Errington AC. Dual function of thalamic low-vigilance state oscillations: rhythm-regulation and plasticity. Nat Rev Neurosci. 2018 Feb;19(2):107–118. PMCID: PMC6364803

84. Steriade M. Sleep oscillations and their blockage by activating systems. J Psychiatry Neurosci JPN. 1994 Nov;19(5):354–358. PMCID: PMC1188624

85. Watt AJ, Desai NS. Homeostatic Plasticity and STDP: Keeping a Neuron's Cool in a Fluctuating World. Front Synaptic Neurosci. 2010;2:5. PMCID: PMC3059670

86. Brunel N, Hakim V. Fast global oscillations in networks of integrate-and-fire neurons with low firing rates. Neural Comput. 1999 Oct 1;11(7):1621–1671. PMID: 10490941

87. Brunel N, Hakim V. Sparsely synchronized neuronal oscillations. Chaos Woodbury N. 2008 Mar;18(1):015113. PMID: 18377094

88. Kirkpatrick J, Pascanu R, Rabinowitz N, Veness J, Desjardins G, Rusu AA, Milan K, Quan J, Ramalho T, Grabska-Barwinska A, Hassabis D, Clopath C, Kumaran D, Hadsell R. Overcoming catastrophic forgetting in neural networks. Proc Natl Acad Sci U S A. 2017 28;114(13):3521–3526. PMCID: PMC5380101

89. Hawkins J, Ahmad S. Why Neurons Have Thousands of Synapses, a Theory of Sequence Memory in Neocortex. Front Neural Circuits. 2016;10:23. PMCID: PMC4811948

90. McCormick DA, Huguenard JR. A model of the electrophysiological properties of thalamocortical relay neurons. J Neurophysiol. 1992 Oct;68(4):1384–1400. PMID: 1331356

91. Akeju O, Hobbs LE, Gao L, Burns SM, Pavone KJ, Plummer GS, Walsh EC, Houle TT, Kim S-E, Bianchi MT, Ellenbogen JM, Brown EN. Dexmedetomidine promotes biomimetic non-rapid eye movement stage 3 sleep in humans: A pilot study. Clin Neurophysiol Off J Int Fed Clin Neurophysiol. 2018;129(1):69–78. PMCID: PMC5743618

92. Llinás RR, Ribary U, Jeanmonod D, Kronberg E, Mitra PP. Thalamocortical dysrhythmia: A neurological and neuropsychiatric syndrome characterized by magnetoencephalography. Proc Natl Acad Sci U S A. 1999 Dec 21;96(26):15222–15227. PMCID: PMC24801

93. Fogerson PM, Huguenard JR. Tapping the Brakes: Cellular and Synaptic Mechanisms that Regulate Thalamic Oscillations. Neuron. 2016 Nov 23;92(4):687–704. PMCID: PMC5131525

94. Duan AR, Varela C, Zhang Y, Shen Y, Xiong L, Wilson MA, Lisman J. Delta frequency optogenetic stimulation of the thalamic nucleus reuniens is sufficient to produce working memory deficits: relevance to schizophrenia. Biol Psychiatry. 2015 Jun 15;77(12):1098–1107. PMCID: PMC4444380

95. Gruber R, Wise MS. Sleep Spindle Characteristics in Children with Neurodevelopmental Disorders and Their Relation to Cognition. Neural Plast. 2016;2016:4724792. PMCID: PMC4958463

96. Wamsley EJ, Tucker MA, Shinn AK, Ono KE, McKinley SK, Ely AV, Goff DC, Stickgold R, Manoach DS. Reduced sleep spindles and spindle coherence in schizophrenia: mechanisms of impaired memory consolidation? Biol Psychiatry. 2012 Jan 15;71(2):154–161. PMID: 21967958

97. Lüttjohann A, Pape H-C. Regional specificity of cortico-thalamic coupling strength and directionality during waxing and waning of spike and wave discharges. Sci Rep. 2019 Feb 14;9(1):2100. PMCID: PMC6375974