Ecological scaffolding and the evolution of individuality: the transition from cells to

multicellular life

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ABSTRACT

Evolutionary transitions in individuality are central to the emergence of biological complexity. Recent

experiments provide glimpses of processes underpinning the transition from single cells to multicellular

life and draw attention to the critical role of ecology. Here we emphasise this ecological dimension and

argue that its current absence from theoretical frameworks hampers development of general explanatory

solutions. Using mechanistic mathematical models, we show how a minimal ecological structure

comprised of patchily distributed resources and between patch dispersal can scaffold Darwinian-like

properties on collectives of cells. This scaffolding causes cells to participate directly in the process of

evolution by natural selection as if they were members of multicellular collectives, with collectives

participating in a death-birth process arising from the interplay between the timing of dispersal events

and the rate of resource utilisation by cells. When this timescale is sufficiently long and new collectives

are founded by single cells, collectives experience conditions that favour evolution of a reproductive

division of labour. Together our simple model makes explicit key events in the major evolutionary

transition to multicellularity. It also makes predictions concerning the life history of certain pathogens

and serves as an ecological recipe for experimental realisation of evolutionary transitions.

INTRODUCTION

Evolutionary transitions in individuality (ETIs) are central to the emergence of biological complexity[1-3]. Each ETI involved the formation of collective-level entities from the interaction of particles[4,5]. For example, chromosomes evolved from the joining of once independently replicating genes. Sexually reproducing types evolved from asexual organisms. Multicellular life evolved from independently replicating cells .

Central to each of these transitions was the emergence of properties at the newly formed level that allowed individuals — at this level — to participate directly in the process of evolution by natural selection[5-9]. This required newly formed collectives to be discrete and vary one to another, to reproduce and to leave offspring that resemble parental types[10]. These essential and intertwined Darwinian properties of variation, differential reproduction and heredity are such fundamental features of living systems that it is easy to overlook the fact that individuality is a derived state and in need of evolutionary explanation[3,7-9,11-13].

With focus on multicellular life, it is evident that reproduction, in even simple multicellular forms, is a complex process[9,11,12,14]. It is therefore tempting to invoke selection as its cause. But this is problematic because the earliest collectives lacked capacity for collective-level reproduction and thus to invoke selection at the collective level as the cause of collective-level reproduction is to invoke the trait requiring explanation as the cause of its own evolution. Clearly such an explanation is unsatisfactory.

One way to avoid this dilemma is to recognise opportunities for co-option of pre-existing cellular traits. For example, in the colonial volvocine algae, group formation evolved by co-option and expansion of cell cycle regulation evident in the unicellular ancestor *Chlamydomonas*[15]. In experimentally evolved snowflake yeast, collective-level reproduction emerged via co-option of apoptotic capacity already apparent in single cell precursors[16].

We do not wish to downplay the importance of co-option, but there is conceivable value in asking whether Darwinian properties might emerge in the absence of co-option. Such a take-nothing-for-granted line of inquiry presents a challenge as it requires conceiving possibilities for the emergence of properties essential for collectives to participate in the process of evolution by natural selection from starting states that lack any manifestation of collective-level Darwinian properties. In essence it begs explanations for how Darwinian properties might emerge from non-Darwinian entities and therefore by non-Darwinian means. Solutions stand to inform not only how multicellular states arise from single cells, but how Darwinian properties might emerge during each of the major evolutionary transitions, including that from non-living matter.

A solution that we advance draws heavily on ecology, the significance of which we suggest has been overlooked — even though the importance of population structure has been emphasised by literature on the levels of selection[17]. It recognises that Darwinian properties can be "scaffolded" by the environment — that these properties can be exogenously imposed in such a way as to cause lower level entities (e.g., cells) to become unwitting participants in a selective process that occurs over a longer timescale than the timescale over which cell-level selection occurs, and as part of a larger entity (a collective). In time, such exogenously imposed — Darwinian-like — properties stand to become endogenous features of evolving systems (for development of general views on scaffolding processes see[18]).

Ecological scaffolding underpinned a recent (and on-going) experimental exploration on the evolution of multicellularity[19,20]. Discrete lineages established from the bacterium *Pseudomonas fluorescens* were propagated under conditions that required, for long-term persistence, repeated completion a two-phase life cycle involving soma and germ-like states. In the experiment, variation was discretised using glass microcosms, but the design is synonymous with an environment such as a pond in which reeds extend from the water[19,21]. Each reed allows establishment of a single microbial mat (the soma-like phase), with the spacing of reeds ensuring variation at the level of mats. Mats that collapse, for example, through physical disturbance, are extinguished, allowing the possibility that an extant mat might, via production of a dispersing (germ-like) phase, increase its representation among the population of mats. Thus the possibility of a selective process unfolds at the level of mats. After ten lifecycle generations, the fitness of derived mats significantly improved, with the most successful lineage having even evolved a simple genetic switch that ensured reliable developmental change between soma and germ-line phases[19]. Not only does this study demonstrate that scaffolding works, but it also showed that externally imposed Darwinian properties can begin the shift toward endogenisation. Or to quote Van Valen: "*evolution is the control of development by ecology*"[22].

Our goal here is to move from the experimental *Pseudomonas* study to a more general formulation. Our focus is the transition from single cells to multicellular life, but argue that scaffolding has broad implications. Our goal is to show that a minimal set of ecological conditions (and ensuing evolutionary responses) are sufficient to effect evolutionary transitions in individuality. We develop our thesis using mechanistic models.

RESULTS

Scaffolding Darwinian properties

Before moving to mathematical models, we describe the simplest conceivable example of a population structure that confers Darwinian-like properties on collectives of particles (cells). Consider an environment in which resources are distributed across patches. A single cell founds a patch. Available resources allow exponential growth of the founding type, however, because resources within patches are finite, they are rapidly depleted causing the population to decline (equations describing the birth / death process and relationship with resources are shown in Box 1 and the Supplementary Information file). Long-term persistence of cells requires dispersal to a new patch. Dispersal occurs at a fixed regular time interval via, for example, some external factor such as wind, water splash or tidal flow.

Cell fate within the environment of patches depends on performance over two timescales. The first timescale is defined by the doubling time of cells. The second is defined by the timing of dispersal events. To make apparent the impact of the second timescale on the ensuing evolutionary dynamics, consider a second variant cell. This type grows faster than the former (cells consume resources more rapidly), which means that in a patch founded by both types, faster growing cells rapidly exclude slower growing cells. In the following we therefore limit the number of colonisers to a single founding cell type, thus limiting within patch competition.

Consider a single slow (depicted in Figure 1 as green) and fast (blue) cell that colonise separate patches (Figure 1). Cells of both types grow and divide, but different growth rates mean that blue cells deplete resources more rapidly than green cells. If dispersal occurs early (Figure 1A) when cells are in exponential growth, then the number of extant blue cells exceeds the number of green cells and thus future recursions of patches are dominated by blue cells (Figure 1B). If, however, dispersal occurs at a later time point, for example, once resources are depleted and population size is in decline, as in Figure 1C, then future patch recursions are dominated by green types despite the fact that within a patch, green types lose in competition with blue types (Figure 1D). Under this scenario parallels are evident with models of virulence evolution in pathogens, where patches equate with hosts, cells are pathogens and dispersal is synonymous with transmission[23-26]. Virulent pathogens that kill their host before transmission face extinction.

Thus far, our focus has been the consequences of this population structure on the long-term fate of cells with different growth rates, but it is possible to switch perspective: there is a coupled evolutionary dynamic occurring at the level of patches (Figures 1C and 1D). Patches manifest Darwinian-like properties of variation (spatially distributed resources ensure that variation is discretised and that

patches vary one to another), differential reproduction (successful patches give rise to patches via

dispersal) and heredity (offspring patches resemble parental patches because new patches are founded by

single cells) that are also features of the founding cells. These properties are externally imposed

(scaffolded) on patches by virtue of the structure of the environment.

Note that we refer to the properties of patches as "Darwinian-*like*". Indeed, it makes no sense to think

of patches as multicellular organisms (they are not) — if the ecological scaffold was to be removed

(patchily distributed resources and a means of dispersal) — the Darwinian-like properties of the patches

would instantly disappear. Yet, under the scenario outlined, cell fate is determined by selective

conditions operating over the second (longer) timescale, just as if the cells themselves were members of

multicellular collectives. Such a scaffolded framework of patch-level selection, based on nothing other

than patchily distributed resources and a means of dispersal between patches, establishes conditions

sufficient for the evolution of traits that are adaptive at the level of patches. We elaborate the

mechanistic bases using models developed in the following section.

Evolution in nested Darwinian populations

To explore the evolutionary dynamics of the above ecological model we allow mutation to affect the

growth rate of individual cells (the within-patch model is described in Box 1). With such a model it

becomes possible to determine the effect of the timing of dispersal — the second timescale — on the

dynamics of within- and between-patch competition. Mathematical details are provided in the

Supplementary Information file, but see also Box 1.

The full evolutionary model consists of M patches that are each founded by a single cell of a single

phenotype (growth rate β). Cells within patches replicate and consume resources with mutation giving

rise to types that vary in growth rate. Once resources are depleted the population size within patches

declines. After a fixed time interval, T, which defines the second timescale, dispersal takes place.

Dispersal is effected by randomly selecting M patches (with replacement) in proportion to the number

of cells within each patch, and then randomly selecting a cell, within chosen patches, in proportion to

numbers within the patch. In effect, the procedure is equivalent to pooling all viable cells from all

patches at the time of dispersal and picking M cells at random. The dispersal regime thus rewards

patches containing the greatest number of cells.

The bottleneck wrought at the moment of dispersal means that types founding new patches are freed

from competition with faster growing types. Figure 2 shows the number of cells within a patch for a

single realisation with initial (arbitrarily chosen) growth rate $\beta = 1.8$. The bottleneck imposes a strong

homogeneity on the composition of the patch as the original population has to grow significantly before

mutants start to arise. The peak number of cells within the patch is reached at time $T = 16.5$, thus for

cells with this initial growth rate, setting a dispersal time of $T = 10$ is fast (i.e., still within the

exponentially growing phase), and $T = 30$ is slow (cells have significantly declined since their peak

numbers).

From the patch perspective, the bottleneck reduces within patch variation and ensures high fidelity of

transmission of patch phenotype (the size of the patch at the time of dispersal). Cells chosen for

dispersal are individually transferred to new patches thus marking the founding of a new generation of

patches. The mechanistic nature of the model allows the average growth rate of cells within a

generation, number of cells in patches at the time of dispersal, and genealogy to be determined.

Figure 3 shows the time resolved dynamics of 50 independent realisations of the full evolutionary

model, where patches experience 200 recursions, under slow ($T = 30$) and fast ($T = 10$) dispersal

regimes. In these simulations the maximum cell growth is set to rate of $\beta = 2$, which in a real system

would arise from chemical and physical constraints to the rate of cell replication. The state of

populations at the time of dispersal are shown in Figures 4A and 4B. Single realisations of the model

under slow and fast dispersal regimes are also shown in Supplementary Movie Files 1 and 2.

Under both fast and slow dispersal regimes patch fitness (the number of cells within patches at time of

dispersal) increases rapidly before reaching a plateau (Figure 3B and 3D). This is consistent with

predictions arising from the logic of Darwinism: imposition (by ecological scaffolding) of Darwinian-

like properties on patches ensures patches participate in a selection process akin to evolution by natural

selection, one that could be the starting point for patches to be units in their own right, provided they

eventually acquire features classically associated with evolutionary individuals[5,27]. The plateau arises

because under the slow dispersal regime growth rate evolves to maximise the number of particles

available at the time of dispersal. Under the fast dispersal regime, the plateau is a consequence of

reaching the maximum limit imposed by the growth rate. As this maximum rate is arbitrarily set,

allowing it to increase would result in the evolution of patches of larger final size.

The cause of enhanced evolutionary success of patches resides in properties of individual cells. Under

both fast and slow dispersal regimes, selection favours patches that harbour the greatest number of cells

at the time of dispersal. Under both regimes fast growing cells outcompete slow growing types within

patches, however, under the slow dispersal regime, selection rewards patches containing slow growing

mutants and selects against patches dominated by fast growing cells. The opposite is true of patches

evolving under the fast dispersal regime.

Under the slow dispersal regime this results in the seemingly counter intuitive finding that patch fitness

increases at the expense of cell fitness (Figures 3A and 3B). Yet within our model, this is readily

explained: fitness of a cell is measured over the short timescale while patch fitness is measured over the

long timescale[28,29]. This captures precisely — and explains mechanistically — the notion of "fitness

decoupling" thought to occur during the earliest stages of the evolution of multicellular life, but which

has often been difficult to intuit[3,30].

Under the fast dispersal regime, fast growing cells are favoured both within patches and over the second

timescale. From the perspective of the evolution of multicellular life, the selection regime imposes the

same directionality at both timescales leading to the view that fitness at both timescales (levels) are

"coupled".

It is interesting to note the difference in speed of the selective response under the two dispersal regimes

and also the magnitude of difference in patch population size at equilibrium. The slower response

under the slow dispersal regime is a consequence of the time taken for slow growing mutants to invade

from rare in the face of within-patch competition for fast growth. The maximum population size under

the fast dispersal regime (for these parameters) is a consequence of the imposed maximum limit on the

growth rate. If faster rates were allowed, then larger population sizes would evolve (up to a limit where

rate maximises patch size at the dispersal time).

Figures 5A and 5B show the evolutionary fate (genealogy) of 10 independent lineages under the slow

and fast dispersal regimes, respectively. Mapped on the phylogenies are changes in cell growth rate and

patch size at time of dispersal. That a genealogical representation is possible derives from both the

mechanistic nature of the model and the fact that patches are founded from single cell types. Movie

versions of the Figures 5A and 5B are shown in Supplementary Movie Files 3 and 4.

It is important to emphasise that the parameters and timescales chosen in the above simulations are

arbitrary. For any initial growth rate > 1, it is always possible to choose fast and slow dispersal times

relative to the time of the peak patch population that will result in selection over the timescale of

dispersal feeding back to affect the growth rate of cells. In contrast, if the dispersal time is equal to the

initial peak time, then no evolutionary change in cell growth rate will be observed.

Evolution of patch traits

The above model shows how a second timescale, defined by dispersal events necessary for establishment

of new patches, affects the evolution of cell growth rate, and how changes in cell growth rate affect the

evolutionary dynamics of patches. From the patch perspective, derived patches are more fit than

ancestral patches, but this is not a consequence of traits adaptive at the patch level. Under both slow

and fast dispersal regimes, selection favours cells whose growth rate maximises the number of cells

available for dispersal. Changes in cell growth rate thus fully explain the evolutionary dynamics of

patches. This cell-level perspective further emphasises the previous comment that patches are not to be

confused with even the most basic manifestations of multicellular life forms.

Nonetheless, our prediction is that an ecological scaffold that couples short and long-term timescale

dynamics establishes conditions conducive to the evolution of traits adaptive at the level of patches. By

this we mean traits that would be difficult to explain from the view point of cells. This prediction

becomes intuitive upon switching perspectives: from a cell-level to a patch-level perspective. Although

patches are endowed with Darwinian-like properties, there is scope for patches to evolve genuine

Darwinian properties — in a ratchet-like manner[31] — so that patches participate in the process of

evolution by natural selection and thus become bearers of adaptations at the patch level.

What might such patch-level adaptations entail and what might constitute their mechanistic (cell-level)

basis? A fundamental requirement given the need for patches to pass through single bottlenecks at each

recursion, is evolution of a stochastic epigenetic switch (a simple developmental programme), such as

observed previously in numerous experiments[32,33] including those arising from experimental

explorations of the evolution of multicellular life[19].

To investigate this possibility we extend the basic model to include two types of cell. The first type,

which we denote G, is essentially the same as in our first model, with the exception that at each

reproduction event there is some probability, q, that instead of giving rise to another G cell, a different

cell type, denoted S, is produced instead. The S cells also consume resources, but unlike G cells, S cells

cannot replicate or be dispersed. The production of S cells is thus costly: they deplete resources and

reduce the number of cells available for dispersal. Full mathematical details of the model are given in

the Supplementary Information file, but see also Box 2. The phenotype of G cells is quantified by their

growth rate, β, and the probability of production of S cells, q. All other parameters are fixed.

In this formulation, S cells are a rough approximation for soma. Like soma, S cells are an evolutionary

dead end. In this switch to considering S cells as proxy for soma, it follows that G cells approximate

germ cells: like germ cells, these dispersing cells found the next collective generation.

To connect with our previous results, simulations of the model were first performed with dispersal

depending solely on the number of G cells within the patch at the time of dispersal, thus patches that

optimise the number of G cells maximise the number of descendent patches. As to be expected given

the cost of maintaining S cells, in repeated simulations of the model in which mutation affects both

growth rate and the probability of production of *S* cells, the rate of *S* cell production under both slow

and fast dispersal regimes declines to zero (Figure 6A-C). The equilibrium fitness of both cells and

patches tend to the same values as in the previous model.

To determine whether the ecological scaffold established by patchily distributed resources and dispersal

between patches establishes conditions favouring evolutionary emergence of a division of labour, the

model was re-run, but with *S* cells now endowed with ability to aid dispersal of *G* types. Mathematically

this was achieved by defining the probability that a cell within a patch is chosen for dispersal be a

function of both the number of *G* and *S* cells in the patch (see Box 2 for details).

As shown in Figures 6D-F (and especially 6E) *S* cells are favoured under the slow dispersal regime (the

probability of *S* cell production rapidly evolves away from zero and plateaus at 0.08). Under this

scenario the equilibrium cell growth rate is higher than when dispersal depends solely on the number of

G cells (contrast this with the solid lines in Figures 6A and D). This is because increased production of

S cells slows the rate of production of *G* cells allowing the population to peak at a comparatively higher

growth rate. Mean patch fitness depends on the contribution that *S* cells make toward dispersal of *G*

types.

Under the fast regime *S* cells are not favoured and the growth rate of *G* simply increases to its

maximum limit. However, if the maximum allowable growth rate is increased beyond the limit of

$\beta = 2$, production of *S* cells under the fast dispersal regime can be favoured. The key point is that

under the slow dispersal regime, production of *S* is always favoured. When dispersal time is fast,

production of *S* cells is favoured only if cell growth rate can increase to the point at which peak

population size is reduced (through early and rapid depletion of resources). In real systems it is likely

that cells would already be close to their maximum growth rate and thus further increases would

depend on rare beneficial mutations. In contrast, decreases in growth rate are readily achieved via

deleterious mutations.

DISCUSSION

The major evolutionary transitions in individuality pose some of the most intriguing and complex problems in biology. Numerous perspectives have been offered, ranging from theoretical multi-level selection frameworks[3,34-39], to views that give prominence to explanations for the evolution of cooperation[40-42]; from perspectives that emphasise the importance of specific mechanisms[4,43-45] through those, like us, that emphasise the pivotal importance of the origins of group-level Darwinian properties[5,7,9,11,13,46,47].

Encompassed within these diverse views are central concepts that are often ambiguous. This is particularly true of scenarios in which ETIs are described in terms of "shifts in levels of selection", or more specifically, shifts between multi-level selection (MLS) frameworks MLS1 (where individual cells are the focus of attention) and MLS2 (where groups are replete with Darwinian properties). A thorough analysis lead Okasha[3] to conclude the existence of a "grey area" between early and later stages of ETIs where both a MLS1 and MLS2 perspective can be taken. Closely allied is the notion of "fitness decoupling"[30] — a sense that as selection shifts from a lower to a higher level the fitness of the higher level decouples from that of the lower — and the related idea of "de-Darwinisation" of lower level components[5]. While to the initiated all these terms convey meaning, they remain metaphorical and descriptive: discussion of issues surrounding ETIs needs to become mechanistic. The challenge is to know where to start.

Our mechanistic approach places emphasis on simplicity, causality and gives prominence to ecological factors. The ability of natural selection to act on collectives of cells depends on emergence of some manifestation of heritable variance in fitness at the collective level. In our take-nothing-for-granted approach the possibility that this arises from co-option of pre-existing cell-level traits was recognised, but put aside. While resulting in a high bar, it gives emphasis to the fact that reproduction, heredity and variation are derived traits and their existence should not be presumed[7,11,14]. It has also made transparent a genuine dilemma, namely, the need to explain how Darwinian properties emerge from non-Darwinian entities and thus by non-Darwinian means. If Darwinian properties do not pre-exist, or cannot arise by co-option of pre-existing lower-level traits, then their earliest manifestation necessarily lies in some exogenous factor(s). The solution we advocate involves recognising the continuity between organisms and their environments; the idea that Darwinian-like properties can be scaffolded by the environment in much the same way that reproduction in viruses is scaffolded by the host cell[5], or that development can be scaffolded by overlap of parts between parents and offspring[48].

The mechanistic models outlined here show that certain ecological structures can scaffold Darwinian-

like properties on collectives, causing the constituent cells to experience selective conditions as if they

were members of nascent multicellular organisms — even to the point where traits emerge that are

defining features of multicellular life. The circumstances are minimal: nothing more than patchily

distributed resources and a means of dispersal between patches. The existence of patches ensures that

variation among collectives is discreet, while establishment of future recurrences of patches via single

founding cells not only reinforces discreteness but is akin to reproduction. At the same time passage

through a single cell bottleneck establishes high fidelity between parent and offspring patches.

The second timescale is of critical importance in that it underpins a death-birth process at the level of

patches[49,50]. Without this feature there would be no, or minimal, evolutionary impact of the second

timescale on the fate of cells. Patches fail or succeed based on properties of the cells. The fact that slow

growing cells are favoured when dispersal time is long is a direct consequence of the feedback between

the patch-level birth-death process and the evolutionary dynamics of cells. Although within-patch

selection favours fast growing cells, patches dominated by fast growing cells contain few viable cells for

dispersal. Long-term success of cells thus comes from alignment of cell and patch fitness. The model

thus explicates the concept of fitness decoupling. If the growth rate of cells is fast relative to the longer

timescale, such that there is suboptimal patch occupancy at the time of dispersal, then patch-level

selection will drive the evolution of reduced cell growth rate, leading to enhanced patch fitness. As

noted above there are numerous parallels with certain models of disease evolution.

An at first unexpected, albeit important, subtlety surrounding the second timescale arises from its

frequency of occurrence relative to the initial growth rate of cells. Beginning from a position where the

growth rate of cells leads to suboptimal patch occupancy at the time of dispersal, as in Figure 3, a

dispersal time that coincides with the exponential growth phase of cells drives an increase in cell growth

rate (Figure 3D), while also marginally increasing patch fitness. More significant though is the fact that

the fast dispersal regime is not conducive to the evolution of a reproductive division of labour. Under

the fast dispersal regime growth rate of cells is the sole factor governing patch success: any reduction in

total yield of G cells due to production of S cells is not offset by contributions that S cells make to

dispersal.

For the evolutionary emergence of a reproductive division of labour the second timescale needs to occur

when cells are not in exponential growth phase. When evolutionary success depends on being the

fastest, limited opportunities exist for exploration of phenotypic e novelty[51-53]. This stems from the

simple fact that manifestations of phenotypic novelty typically come at some cost to growth rate. In

other words, opportunities for evolution to explore phenotypic space require the possibility that slow

growing types not be excluded by selection. Selection comes to reward persistence and not simply types

that grow fastest.

There are additional aspects of the longer timescale that are illuminating. It is usual when discussing the

transition from cells to multicellular life to consider the cell the "lower level" and the group the "higher

level". Accordingly, the transition from cells to multicellular life is often referred to as a levels of

selection problem[3,5,34,37]. The same is true for any of the major ETIs. It is clear from our patch model

that the evolution of multicellular life is better articulated as a problem to be solved by understanding

conditions leading to the emergence of a second timescale over which a birth-death process operates on

discretely packaged variation. This shift from levels to timescales does much to clarify the kinds of

conditions necessary to effect ETIs[22,28,29].

The occurrence of ecological conditions in nature that generate birth-death dynamics at a timescale

longer than the doubling time of particles are likely rare, perhaps explaining why transitions to

multicellular life are rare[13]. Elsewhere we have articulated spatially structured environments afforded by

reeds in ponds about which surface-colonising microbial mats form, allowing cells to access oxygen that

is otherwise limiting. Periodic collapse of mats marks death events that allow the possibility that new

mats arise by dispersal from extant types[7,21]. Precisely this scenario underpinned design of on-going

lineage selection experiments exploring the evolution of multicellular life[19,20].

Once attention focusses on dispersal events, along with recognition that such events may effect

collective reproduction, then attention turns to emergence of simple primordial lifecycles. Those

involving more than a single phase, for example those involving soma- and germ-like phases, establish

by virtue of the life cycle, a second timescale over which selection acts[19]. A significant aspect of such life

cycles is the fact that birth-death events depend on the efficacy of developmental processes that become

the focus of selection. Indeed, the evolution of lifecycles is intimately connected to the transition of

multicellular life. Even in our simple conceptual model outlined above, the moment that S cells have a

selected advantage, then a rudimentary life cycle manifests, with selection able to set the developmental

programme via effects on q, the rate at which S cells are produced. Arguably this marks an early step in

the process of endogenisation: the process by which externally imposed Darwinian-like properties

become integral features of the new entity[3,14]. Another possibility would be for S cells to provision new

environments with resources as does the cotyledon in a plant seed, thus freeing evolving collectives from

dependence on patchily distributed resources. In the *Pseudomonas* experiment, as generations of

selection have passed, dependence of the evolving lineages on the scaffold has lessened. This is

especially noticeable via evolution of a simple developmental programme controlling the switch

between soma-like and germ-like phases[19].

Thus far we have been silent on cooperation and the causes of cooperation that typically feature so

prominently in discussions on the evolution of multicellular life[41,54]. There is no doubt that cooperation

is a basic feature of multicellular organisms, but the scaffolding perspective does not presuppose

cooperation among cells as a necessary first step. Nonetheless behaviours that might reasonably be

labelled as cooperation stand to evolve given appropriate ecological scaffolds. For example, the slow

growing cells favoured when dispersal time is slow (Fig 3A, 4A and 5A) could be labelled cooperating

types because they show restraint in the face of plentiful resources and the fast growing mutants arising

within patches might be termed "cheats", but there is no need to use such labels. Indeed, applying

these labels brings focus to the individual cell, and detracts from the ecological context and importance

of timescales that is critical to scaffolding, and to where causal processes lie.

One behaviour that is readily labelled as an extreme form of cooperation — suicidal altruism — is

evident in the *S* cells. Such behaviours can be seen from the perspective of single cells with the

temptation to invoke inclusive fitness[55], but to do so, would be to ignore the importance of ecology[56]

and population structure[17]. It is the meta population structure determined by ecological circumstances

that ensures patches are founded by single cells and this both limits within patch conflict, while also

being necessary for the emergence and maintenance of a reproductive division of labour. In our model

within-patch close kinship is thus a consequence of environmental structure.

The models developed in this paper are the simplest possible constructs that illustrate the process of

ecological scaffolding. In simplifying to its most basic formulation a strict ecology has been assumed.

For example, a single cell is only ever dispersed into a new patch, there is no migration between patches,

and dispersal occurs for all patches at the same moment on a fixed, deterministic, timescale. On-going

work explores the effects of relaxing these assumptions and in a forthcoming paper we show that the

overall effects of scaffolding are robust, up to a point, to increases in bottleneck size and to

asynchronous dispersal.

The concept of ecological scaffolding has a number of applications and implications. In concluding we

mention briefly three areas. The first concerns the emergence of the first self-replicating chemistries at

the moment life emerges from non-living material. Recognition that Darwinian-like properties might

emerge from the interplay between chemistry and environment opens the door to conceptual and

experimental scenarios whereby chemistries that lack capacity for autonomous replication might begin

to transition, through a process of templated production of bioactive compounds, toward a replicative

process. Alkaline thermal vents appear to offer as much: these highly compartmentalised structures sit at the interface between hydrogen-rich fluids arising from the flow of heated water across serpentine, and acidic ocean waters with the possibility that carbon-dioxide is reduced to biologically active species[57,58]. Ensuing "growth" of products within the porous compartments sets in place the possibility of a replicative process. Incorporation of such ecological structures in future experimental designs may provide Darwinian ingredients that are typically absent from explorations of the chemical origins of life.

The second area of relevance is infectious disease biology. To a pathogen, the eukaryotic host offers a discrete patch of resource. Pathogens that rely on transmission for long-term persistence experience selection over two timescales. Our model leads to the prediction that pathogens that passage through restrictive bottlenecks, such as HIV, are likely to have evolved more complex life histories than currently appreciated, involving, for example, a division of labour. This seems to be true of *Salmonella typhimurium* that switches stochastically between virulent and avirulent cell types, that invade the lumen, or colonise the gut, respectively. Cells that colonise the lumen, trigger the inflammatory response, which benefits the faster growing avirulent cells in the gut. However, unlike cells in the gut, lumen colonising cells are killed by the intestinal innate immune defences and are thus, like soma, an evolutionary dead-end[59]. A division of labour is also hinted at in the case of HIV and other chronic RNA viruses in humans that appear to escape the deleterious effects of short-sighted within-host evolution over prolonged time intervals. Growing evidence suggests that this is attributable to establishment of germ-line-like lineages[60].

The third example concerns application of ecological scaffolding for in vitro engineering of evolutionary transitions and particularly for top-down engineering of microbial communities in which communities eventually become a single symbiotic entity. In the laboratory environment, and aided particularly by advances in micro / millifluidic technologies, it is a relatively trivial matter to confine populations and / or communities to thousands of discretised droplets that can then be subject to a death-birth process[21]. In a forthcoming paper we detail this process and its outcome for the evolution of interactions that build integrated communities.

ACKNOWLEDGEMENTS

We are grateful to Silvia De Monte, Guilhem Doulcier and members of our respective teams for stimulating discussion. AJB acknowledges an ARC DECRA fellowship (DE160100690) and support from both the ARC Centre of Excellence for Mathematical and Statistical Frontiers (CoE ACEMS), and the Australian Government NHMRC Centre for Research Excellence in Policy Relevant Infectious diseases Simulation and Mathematical Modelling (CRE PRISM²). PB acknowledges a Macquarie University Research Fellowship and a Large Grant from the John Templeton Foundation (Grant ID 60811). PBR acknowledges generous financial support from MPG core funding and previously from the Marsden Fund Council from New Zealand Government funding, administered by the Royal Society of New Zealand.

SUPPLEMENTARY MATERIAL

Supplementary Information

Supplementary Movie File 1

Supplementary Movie File 2

Supplementary Movie File 3

Supplementary Movie File 4

BOX 1. Mathematical model

We construct the simplest possible model to demonstrate the dynamics of two nested Darwinian populations. We assume a fixed population of M patches each provisioned with a fixed amount of resource that is consumed by the cells in order to divide. The dynamics within each patch are independent. Within each patch, cells undergo a birth/ death process for a fixed length of time, T, after which a dispersal event occurs leading to a new generation of M patches (with replenished resources).

We first describe the basic birth / death dynamics within a patch (with no mutations), we then describe how this is modified by allowing for mutation. Cells have a mean life time that, without loss of generality, is set to 1 and individual cells reproduce at a rate β in proportion to the density of resource in the patch. Under these dynamics, the density of cells within the patch follows modified Lotka–Volterra equations, where there is no replenishment of the resource, i.e.,

$$\frac{dy}{dt} = -\beta x y,$$
$$\frac{dx}{dt} = (\beta y - 1)x, \tag{1}$$

where $y(t)$ and $x(t)$ are the proportions of resource and cells within the patch respectively. The scaling for these proportions is taken to be the initial amount of resource within the patch, N 61. Initially, $y(0) \approx 1$, so cells grow exponentially with the resource being depleted at the same rate. At some point, the resource becomes significantly depleted leading to a reduction in growth rate. The population peaks once the rate of growth matches the rate that cells die. After this point the population of cells begins to decline (the trajectories shown in Figure 1 are examples of these dynamics). The amount of growth and hence the peak population size is limited by the initial amount of resource in the patch, so high growth rates lead to large populations that peak early, but which then decrease quickly.

We now outline how mutations and hence different competing cell types are included in the model. We assume that mutations only affect the growth rate of cells and, for simplicity, possible growth rates are discretised (with step size μ) so that cells can be grouped according to their growth rate. We define the growth rate of the i'th type as β_i. At each reproduction event there is a probability, p, of creating a child cell with a different growth rate, and hence with probability $1 - p$ a cell of the same type is produced. If a mutant is produced there is a probability ν that it has a growth rate larger than the parent, and therefore $1 - \nu$ it is smaller than the parent cell. As before, each cell type has the same mean life span which is set to 1. Unless otherwise indicated these parameters are set as $p = 0.01$, $\mu = 0.05$ and $\nu = 0.5$.

Simulating such a model is more difficult than just solving a system of ordinary differential equations

(ODEs) as the introduction of new types through mutations is a stochastic process. Simulation of a full

stochastic model of the process defined above is possible, but undesirable as it is computationally

expensive. Instead we approximate the dynamics with a piecewise deterministic process, where the times

of the introduction of new types via mutations are modelled stochastically, but the growth dynamics

between these times are modelled deterministically[62]. Hence between the introduction of new types,

the proportions of each type already in the patch evolves via the set of $m + 1$ ODEs,

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$$\frac{dy}{dt} = -y \sum_{i=1}^{m} \beta_i x_i,$$
$$\frac{dx_i}{dt} = (\beta_i y - 1)x_i, \quad i = 1,\ldots,m,$$
(2)

Where m is the number of types currently in the patch. This is equivalent to Lotka–Volterra dynamics

with competition, but where resources are not replenished.

The use of the piecewise deterministic models, where times of new mutants arising are stochastic but

the birth-approach is a pragmatic comprise between computational efficiency and realism, but ignores

other stochastic effects, such as the time for the cells to grow to an appreciable number before

exponential growth is fully underway. Full stochastic models can account for this and display a

distribution of patch sizes with a much larger variance. Forthcoming work shows that the evolutionary

dynamics are similar.

The particular dynamics of the system allow further approximation to improve the computational

efficiency of the model. The bottleneck imposes a strong homogeneity to the patch, which means that

new mutants cannot reach appreciable frequencies within a single patch generation. This means it is

possible to limit the number of mutants in the system, typically to just the first two types, which means

that it is not necessary to allow mutant types to create further mutants. An example trajectory is shown

in Figure 2 with full details provided in the Supplementary Information.

Simulation of the full model over multiple generations proceeds as follows. Each of the M patches are

seeded with a single cell (at the same time). For each patch the model described above is simulated up

to the dispersal time T. For the first model, the dispersal dynamics only depend on the proportion of

cells within each patch. A new generation of patches is founded by randomly selecting a patch in

proportion to the number of cells within it and then randomly selecting a cell, within the chosen patch,

again in proportion to its number within the patch. This procedure is equivalent to simply picking

particles randomly from the whole population of patches. Hence the larger the number of a given type

within the population, the more likely it is to be dispersed. This two-step procedure is simulated for a

given number of generations and quantities, such as the average growth rate within a generation, can be

calculated. Because the model is mechanistic, it is possible to track the genealogies of both the cells and

patches, as shown in Figure 5.

Box 2: Mathematical model with two cell types

We take a similar approach to simulating this version of the model, employing a piecewise-deterministic approximation where the times of the introduction of new types due to mutations are stochastic and the birth / death dynamics are deterministic. Hence between mutations the dynamics evolve as

$$\frac{dy}{dt} = -y \sum_{i=1}^{m} \beta_i x_i - dyz,$$

$$\frac{dx_i}{dt} = \beta_i (1 - q_i) y x_i - x_i, \quad i = 1, \dots, m(t),$$

$$\frac{dz}{dt} = y \sum_{i=1}^{m} q_i \beta_i x_i - z,$$

where $z(t)$ is the proportion of S cells in the patch. This model introduces two new parameters: q_i, which is the per event probability of producing an S cell and d, which is the rate at which S cells consume the resource. The parameter d remains fixed, but q_i is subject to mutation. As the phenotype space is now two-dimensional the scheme for generating mutants is different to the first model, but the number of new mutants remains limited to the first two. We assume that only G cells can be dispersed, hence the bottleneck enforced by the dispersal mechanism means a patch is always seeded from a single G type cell. More details of the model and mutational process are given in the Supplementary Information.

For dispersal we assume the system to be composed of $k = 1, \dots, K$ patches. Then let $x_i^{(k)}$ be the proportion of type G_i and $y^{(k)}$ be the proportion of type S in patch k. Each patch is founded by a single G cell with phenotype $(\beta, q)^{(k)}$. These cells reproduce, mutate, and create S cells until the dispersal time, T, at which point a sample is taken from the resulting populations to create the next generation. This occurs in two steps:

1. Randomly select a patch k, in proportion to its weight, w_k, which is function of the proportion of its constituents, $x_i^{(k)}$ $i = 1,2,3$ and $y^{(k)}$.

2. From the patch selected in step 1, randomly select a G cell from the total patch population.

We consider two ways to assign a weight to patches for the first step. Most simply, if we take $w_k = \sum_{i=1}^{m} x_k$, then the dispersal process is as in the first model, i.e., the probability of choosing a patch is proportional only to the number of G cells, so patches with more G cells at the time of dispersal are more likely to be sampled from. The other function we consider is

$$w_k = (1 + 200y^{(k)}) \sum_{i=1}^{m} x_i^{(k)}.$$

This can be interpreted as the S cells aiding dispersal from the patch, for example by attracting the dispersal agent. The constant multiplying $y^{(k)}$ term represents the strength of this effect.

REFERENCES

Buss, L. W. *The Evolution of Individuality*. (Princeton University Press, 1987).
Maynard Smith, J. & Szathmáry, E. *The Major Transitions in Evolution*. (Freeman, 1995).
Okasha, S. Evolution and the Levels of Selection. (Oxford University Press, 2006).
Michod, R. E. Darwinian Dynamics: Evolutionary Transitions in Fitness and Individuality. (Princeton University Press, 1999).
Godfrey-Smith, P. Darwinian Populations and Natural Selection. (Oxford University Press, 2009).
Hull, D. L. Individuality and selection. *Ann. Rev. Ecol. Syst.* **11**, 311-332, (1980).
Rainey, P. B. & Kerr, B. Cheats as first propagules: a new hypothesis for the evolution of individuality during the transition from single cells to multicellularity. *Bioessays* **32**, 872-880, (2010).
Libby, E. & Rainey, P. B. A conceptual framework for the evolutionary origins of multicellularity. *Phys. Biol.* **10**, 035001, (2013).
De Monte, S. & Rainey, P. B. Nascent multicellular life and the emergence of individuality. *J. Biosci.* **39**, 237-248, (2014).
Lewontin, R. C. The units of selection. *Ann. Rev. Ecol. Syst.* **1**, 1-18, (1970).
Griesemer, J. The units of evolutionary transition. *Selection* **1**, 67-80, (2000).
Bourrat, P. From survivors to replicators: Evolution by natural selection revisited. *Biol. Phil.* **29**, 517-538, (2014).
Rainey, P. B. & De Monte, S. Resolving conflicts during the evolutionary transition to multicellular life. *Annu. Rev. Ecol. Evol. Syst.* **45**, 599-620, (2014).
Okasha, S. The strategy of endogenization in evolutionary biology. *Synthese*, doi.org/10.1007/s11229-11018-11832-11226, (2018).
Hanschen, E. R. *et al.* The Gonium pectorale genome demonstrates co-option of cell cycle regulation during the evolution of multicellularity. *Nat. Commun.* **7**, 11370, (2016).
Ratcliff, W. C., Denison, R. F., Borrello, M. & Travisano, M. Experimental evolution of multicellularity. *Proc. Natl. Acad. Sci. USA* **109**, 1595–1600, (2012).
Wade, M. J. Adaptation in Metapopulations: How Interactions Change Evolution. (University of Chicago Press, 2016).
Caporael, L. R., Griesemer, J. R. & Wimsatt, W. C. in *Vienna Series in Theoretical Biology* (eds G. B. Müller, G. P. Wagner, & W. Callebaut) 426 (The MIT Press, Cambridge, MA, 2014).
Hammerschmidt, K., Rose, C., Kerr, B. & Rainey, P. B. Life cycles, fitness decoupling and the evolution of multicellularity. *Nature* **515**, 75-79, (2014).
Rose, C. J., Hammerschmidt, K. & Rainey, P. B. Meta-population structure and the evolutionary transition to multicellularity. *bioRxiv*, doi.org/10.1101/407163, (2019).
Rainey, P. B., Remigi, P., Farr, A. D. & Lind, P. A. Darwin was right: where now for experimental evolution? *Curr Opin Genet Dev* **47**, 102-109, (2017).
Van Valen, L. Energy and evolution. *Evolutionary Theory* **1**, 179-229, (1976).
Ewald, P. W. *Evolution of Infectious Disease*. (Oxford University Press, 1994).
Levin, B. R. & Bull, J. J. Short-sighted evolution and the virulence of pathogenic microorganisms. *Trends Microbiol.* **2**, 76-81, (1994).
Frank, S. A. Natural selection. III. Selection versus transmission and the levels of selection. *J Evol Biol* **25**, 227-243, (2012).
Lythgoe, K. A., Pellis, L. & Fraser, C. Is HIV short-sighted? insights from a multistrain nested model. *Evolution* **67**, 2769-2782, (2013).
Clarke, E. The problem of biological individuality. *Biol. Theor.* **5**, 312-325, (2010).

| 597 | 28 | Bourrat, P. Levels of selection are artefacts of different temporal fitness measures. *Ratio* **28**, 40-|
| 598 | | 50, (2015). |

28 Bourrat, P. Levels of selection are artefacts of different temporal fitness measures. *Ratio* **28**, 40-
50, (2015).

29 Bourrat, P. Levels, time and fitness in evolutionary transitions in individuality. *Philos Theor Biol*
7, e601, (2015).

30 Michod, R. E. & Roze, D. in Mathematical and Computational Biology: Computational
Morphogenesis, Hierarchical Complexity, and Digital Evolution   (ed C. L. Nehaniv) 47-92
(American Mathematical Society, 1999).

31 Libby, E. & Ratcliff, W. C. Ratcheting the evolution of multicellularity. *Science* **346**, 426-427,
(2014).

32 Gallie, J. *et al.* Bistability in a metabolic network underpins the de novo evolution of colony
switching in *Pseudomonas fluorescens*. *PLoS Biol.* **13**, e1002109, (2015).

33 Remigi, P. *et al.* Ribosome provisioning activates a bistable switch coupled to fast exit from
stationary phase. *Mol Biol Evol*, 10.1093/molbev/msz1041, (2019).

34 Calcott, B. & Sterelny, K. in *The Vienna Series in Theoretical Biology*   (eds G. B. Müller, G. P.
Wagner, & W. Callebaut) 319 (The MIT Press, Cambridge MA, 2011).

35 Godfrey-Smith, P. & Kerr, B. Gestalt-switching and the evolutionary transitions. *Brit J Philos Sci*
64, 205-222, (2013).

36 Clarke, E. Origins of evolutionary transitions. *J. Biosci.* **39**, 1-14, (2014).

37 Shelton, D. E. & Michod, R. E. Group selection and group adaptation during a major
evolutionary transition: insights from the evolution of multicellularity in the volvocine algae. *Biol.*
Theor. **9**, 452-469, (2014).

38 Clarke, E. A levels-of-selection approach to evolutionary individuality. *Biol Philos* **31**, 893-911,
(2016).

39 Bourrat, P. Evolutionary transitions in individuality: A formal analysis. *Synthese*, in press, (2019).

40 Queller, D. C. & Strassmann, J. E. Beyond society: the evolution of organismality. *Phil. Trans. R.*
Soc. Lond. B **364**, 3143-3155, (2009).

41 Bourke, A. F. *Principles of Social Evolution*. 267 (Oxford University Press, 2011).

42 West, S. A., Fisher, R. M., Gardner, A. & Kiers, E. T. Major evolutionary transitions in
individuality. *Proc. Natl. Acad. Sci. USA* **112**, 10112-10119, (2015).

43 Boraas, M. E., Seale, D. B. & Boxhorn, J. E. Phagotrophy by a flagellate selects for colonial prey:
A possible origin of multicellularity. *Evol Ecol* **12**, 153-164, (1998).

44 van Gestel, J. & Tarnita, C. E. On the origin of biological construction, with a focus on
multicellularity. *Proc. Natl. Acad. Sci. USA* **114**, 11018-11026, (2017).

45 Herron, M. D. *et al.* De novo origins of multicellularity in response to predation. *Sci Rep* **9**,
2328, (2019).

46 Rainey, P. B. Unity from conflict. *Nature* **446**, 616, (2007).

47 Bourrat, P. Evolutionary transitions in heritability and individuality. *Theor Biosci*, in press,
(2019).

48 Griesemer, J. in *Towards a Theory of Development*   eds A. Minelli & T. Pradeu) 183-202
(Oxford University Press, 2014).

49 Simon, B., Fletcher, J. A. & Doebeli, M. Towards a general theory of group selection. *Evolution*
67, 1561-1572, (2012).

50 Doebeli, M., Ispolatov, Y. & Simon, B. Towards a mechanistic foundation of evolutionary
theory. *eLife* **6**, (2017).

51 Muller, G. B. & Wagner, G. P. Novelty in Evolution - Restructuring the Concept. *Ann. Rev.*
Ecol. Syst. **22**, 229-256, (1991).

52 Pfeiffer, T., Schuster, S. & Bonhoeffer, S. Cooperation and competition in the evolution of ATP-
producing pathways. *Science* **292**, 504-507, (2001).

53 Pfeiffer, T. & Bonhoeffer, S. An evolutionary scenario for the transition to undifferentiated
multicellularity. *Proc. Natl. Acad. Sci. USA* **100**, 1095-1098, (2003).

54 Wilson, D. S. & Sober, E. Reviving the superorganism. *J. Theor. Biol.* **136**, 337-356, (1989).

55 Hamilton, W. D. The genetical evolution of social behavior, I & II. *J. Theor. Biol.* **7**, 1–52,
(1964).

56 Wilson, E. O. & Holldobler, B. Eusociality: origin and consequences. *Proc. Natl. Acad. Sci. USA*
102, 13367-13371, (2005).

57 Martin, W. & Russell, M. J. On the origin of biochemistry at an alkaline hydrothermal vent.
Phil. Trans. R. Soc. Lond. B **362**, 1887-1925, (2007).

58 Lane, N. The Vital Question: Why is Life the Way it is? , 352 (Profile Books Ltd, 2015).

59 Diard, M. *et al.* Stabilization of cooperative virulence by the expression of an avirulent
phenotype. *Nature* **494**, 353-356, (2013).

60 Lythgoe, K. A., Gardner, A., Pybus, O. G. & Grove, J. Short-Sighted Virus Evolution and a
Germline Hypothesis for Chronic Viral Infections. *Trends Microbiol.* **25**, 336-348, (2017).

61 Black, A. J. & McKane, A. J. Stochastic formulation of ecological models and their applications.
Trends Ecol. Evol. **27**, 337-345, (2012).

62 Davis, M. H. A. Piecewise-deterministic Markov processes: A general class of non-diffusion
stochastic models. *J. R. Statist. Soc. B* **46**, 353-388, (1984).

Figure 1. Scaffolding Darwinian properties. Patchily distributed resources provide opportunity for two cell types (blue and green) to replicate (blue cells grow faster than green according to equation 1 (Box). Single cells of each type colonise discrete patches at time t = 0 and consume resources. Difference in growth rate means the relationship between cell density and time differs for blue versus green populations. Should a dispersal event occur during exponential growth (dashed line) then more blue cells will be dispersed relative to green (A) and thus the blue population will be more successful over the long term (B). Conversely, should dispersal occur at a later stage and after resources are depleted (C), then the population of green cells will out compete green over the long term (D).

Figure 2. Single realisation of the within patch model with mutation. The number of cells is plotted as a function of time starting from a single cell with growth rate β = 1.8 and the amount of resource $N=10^6$. The darker shaded regions show the numbers of mutant cells. Darker and lighter colours correspond to faster and slower growing cells, respectively. The circles below the main plot are representations of cell numbers (used in the video and Figures 3 and 4) at times 7, 10, 16.5, 22, 26 and 30. In this representation, the *area* of each region is proportional to the number of cells of each type within the patch (with the same colour scheme). The peak number of cells within the patch is reached at time ~16.5, thus for the initial growth rate of β = 1.8, setting a dispersal time of T=10 is considered fast, and T=30 slow (shown by the dashed lines).

Figure 3. Effect of dispersal timescale on properties of cells and patches. Each grey line is from an independent realisation of the stochastic model. The solid black lines are averages over 50 realisations. A and B show the evolution of the average growth rate over all patches in the generation with slow dispersal (T=30). C and D are the same but with fast dispersal (T=10). Both regimes start with a homogenous population of cells with β = 1.8 and in both cases the average patch size increases, but for slow dispersal this is achieved by cells decreasing their average growth rate. Figure 4 shows the 64 patches at the moment of dispersal for two single realisations (shown in red above) after a number of generations.

Figure 4. Evolution of patch size under slow and fast regimes. The dynamic of patch-size evolution and corresponding effects on cell growth rate under slow (A) and fast (B) dispersal regimes. Movies of simulations are shown in Supplementary Movie Files 1 and 2. Colour corresponds to cell growth rate and patch size is proportional to the number of cells within patches at the moment immediately prior to dispersal. Generation number is indicated above each panel and corresponds to the realisations highlighted in red in Figure 3.

Figure 5. Genealogy of patches under slow (A) and fast (B) dispersal regimes. The simulations to produce these have only 10 patches and modified mutational parameters compared with those in Figures 2 and 3. This is to allow a clearer visualisation of the process, which otherwise requires many more generations to see change. Movie versions of these are included in the Supplementary Movie Files 3 and 4. As in the previous figures, the cell numbers in each patch are proportional to the area of the circles and the growth rates are indicated by the colours, as shown by the colour bar in Figure 4. The mutational parameters are larger for these simulations (μ = 0.05, p = 0.05) so evolution occurs on a quicker timescale as compared with the results shown in Figures 3 and 4.

Figure 6. Simulations of the two-type model. A, B and C show the simulations the the model where the probability of dispersal only depends on the number of G cells in the patch at T. A shows the mean growth rate (averaged over generations), B is the mean probability of a reproduction event creating a B cell, and C shows the mean patch fitness. Each grey line is a single stochastic realisation and the solid and dashed lined are averages over 100 independent realisations for slow (T=30) and fast dispersal (T=10) respectively. Panels D, E, and F, show the similar simulations, but where the probability of dispersal from a patch is a function of both the number of G and S cells (see Box 2). In all scenarios, the maximum cell growth rate is limited to 2. The difference in the total number of generations between these simulations (100) and previous results (200) are due to different mutational schemes between the models.