

1 **Lactation curve model with explicit representation of perturbations as a phenotyping**
2 **tool for dairy livestock precision farming.**

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16 All relevant data are within the paper and its Supporting Information files. The R code for the
17 Perturbed Lactation Model is available under request for academic purposes.

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24 **Abstract**

25 **Background**

26 Understanding the effects of environment on livestock provides valuable information on how
27 farm animals express their production potential, and on their welfare. Ruminants are often
28 confronted with perturbations that affect their performance. Evaluating the effect of these
29 perturbations on animal performance could provide metrics to quantify how animals cope
30 with their environment and therefore better manage them. In dairy systems, milk production
31 records can be used to evaluate this effect because (1) they are easily accessible, (2) the
32 overall dynamics throughout the lactation process have been widely described, and (3)
33 perturbations often occur and cause milk loss. In this study, a lactation curve model with
34 explicit representation of perturbations was developed.

35 **Methods**

36 The perturbed lactation model is composed of two components. The first one describes a
37 theoretical unperturbed lactation curve (unperturbed lactation model), and the second
38 describes deviations from the unperturbed lactation model. The model was fitted on 319
39 complete lactation data from 181 individual dairy goats allowing the characterization of
40 individual perturbations in terms of their starting date, intensity, and shape.

41 **Results**

42 The fitting procedure detected a total of 2354 perturbations with an average of 7.40
43 perturbations per lactation. Loss of production due to perturbations varied between 2 % and
44 19 %. Results show that it is not the number of perturbations is not the major factor
45 explaining the loss in milk yield over the lactation, suggesting that there are different types of
46 animal response to disturbing factors.

47 **Conclusions**

48 By incorporating explicit representation of perturbations, the model allowed the
49 characterization of potential milk production, deviations induced by perturbations, and
50 thereby comparison between animals. These indicators are likely to be useful to move from
51 raw data to decision solutions in dairy production.

52 INTRODUCTION

53 In the context of precision livestock farming, simple interpretive tools are required to convert
54 raw time series datasets, now routinely recorded in animals, into useful information for on-
55 farm decision-making. Such tools are not only expected to provide farmers with good
56 information on performance level of individual animals, but also to detect pathological,
57 nutritional or environmental problems affecting production traits at individual or herd scales.
58 In dairy systems, it is well known that milk yield can be affected by problems such as udder
59 health problems [1], lameness [2], heat stress [3] or nutritional challenges [4]. Such problems
60 induce perturbations in the course of the lactation process and result in a serrated shape
61 pattern in the lactation curve. These perturbations can be seen as deviations of the lactation
62 curve from its typical profile. This typical profile reflects that lactation is a physiological
63 process common to mammal females and as a result, its expression through time follows a
64 general pattern [5]. It can be described in 3 phases. The first phase starts after parturition with
65 the initial milk yield increasing to a maximum or peak yield. The second phase is a plateau-
66 like period in which milk yield is maintained for a more or less long time. The third phase is
67 the decrease from the peak yield. This last phase can be divided into two parts according to
68 the speed of decrease, the first one corresponding to an approximately constant declining rate
69 of milk production after the peak yield and the second corresponding to an acceleration of the
70 decline as pregnancy progresses before the start of the dry period when lactation stops [6–8].
71 Modelling the lactation curve is a long standing issue [9] and numerous authors have
72 proposed mathematical models allowing the characterization of milk yield dynamics, *i.e.*, the
73 transformation of a series of temporal data into a vector of estimated parameters via a fitting
74 procedure. The most famous and most used model is the one of Wood published in 1967 [10].
75 The overall objective of lactation models is to reduce the variability or noise in data by
76 extracting a profile and therefore being able to characterize an average animal milk

77 production or to compare the production of different animals. This strategy of using lactation
78 models as phenotyping tools has been very useful in the past years (for instance, test-day
79 models for genetic selection) and in a context of scarce raw data. An important limitation of
80 these modelling approaches is that short-term perturbations are removed during fitting
81 procedure in order to extract an unperturbed phenotype, corresponding to a typical lactation
82 curve. However, characterizing perturbations can be highly relevant for better understanding
83 the resilience of milk production and for making management decisions. Evaluating the effect
84 of perturbations on animal performance could provide metrics to quantify how animals cope
85 with their environment, and therefore better manage them. Taking into account this type of
86 information can provide a proxy to estimate the frequency and severity of disorders such as
87 clinical mastitis [11]. Studying perturbations in lactation curves also makes it possible to
88 compare animals facing the same stress and detect the ones with the greatest adaptive
89 capacities. Finally, the on-farm early detection of perturbations in milk yield can provide
90 farmers with an alert system on udder health. Recently, Huybrechts et al. [12] tested and
91 developed the synergistic control concept for early detection of anomalies in dairy cows based
92 on detection of shifts in milk yield per hour. Of the 49 mastitis cases, 31 cases were detected
93 using this methodology at the same time or earlier than they were detected by the farmer.

94 The need for incorporating perturbations into lactation curve models is also driven by the
95 development of precision livestock farming. Now, we have more frequent and reliable data
96 and we can move from the logic of reducing variability around average profiles to the logic of
97 extracting variability to provide information as such. High throughput data has led to the
98 development and use of statistical methods to understand perturbations (*e.g.* Codrea et al.
99 [13]). However, such smoothing methods are limited by their lack of an a priori representation
100 of the typical "unperturbed" lactation curve. As such, the quantification of perturbations in
101 such models is underestimated, especially for perturbations of long duration. A final

102 limitation of these purely statistical methods is that the model coefficients in themselves do
103 not have direct biological meaning. There is thus a lack of tools for phenotyping milk
104 production with a systemic representation of perturbations.

105 We developed a Perturbed Lactation Model (PLM) that incorporates an explicit representation
106 of perturbations and that converts individual raw time-series data into biological meaningful
107 parameters. The fitting procedure of PLM allows the detection and the characterization of
108 perturbations in milk time-series. The objective of the present paper is (1) to introduce the
109 PLM model and the explicit representation of perturbations, (2) to describe the use of PLM to
110 detect and characterize perturbations in milk yield time series with an example in dairy goats,
111 and (3) to illustrate the role of PLM as a phenotyping tool by analyzing the variability in
112 perturbed lactation curves on the basis of the fitting results obtained on the dairy goat dataset.

113 MATERIALS AND METHODS

114 The Perturbed Lactation Model (PLM) is composed of a lactation model, denoted Y^* ,
115 describing the theoretical unperturbed dynamics of milk yield along the lactation, and a
116 perturbation model, denoted π , describing deviations from the lactation model.

117 The dynamics of daily milk yield ($Y(t)$, in kg) during the lactation is thus given by:

$$118 \quad Y(t) = Y^*(t) \cdot \pi(t)$$

119 where t is the time after parturition in days.

120 Unperturbed lactation model

121 Among the numerous mathematical models developed to study lactation curves, the
122 incomplete Gamma function proposed by Wood [10] has been widely used in different
123 mammals (*e.g.* rabbits [14], sheep [15]). This model gives a general expression for the
124 dynamics of milk yield along the lactation. In this article, we have selected this model as an
125 example to define the unperturbed lactation curve. Because the structure of PLM is generic,
126 any other lactation model can be used.

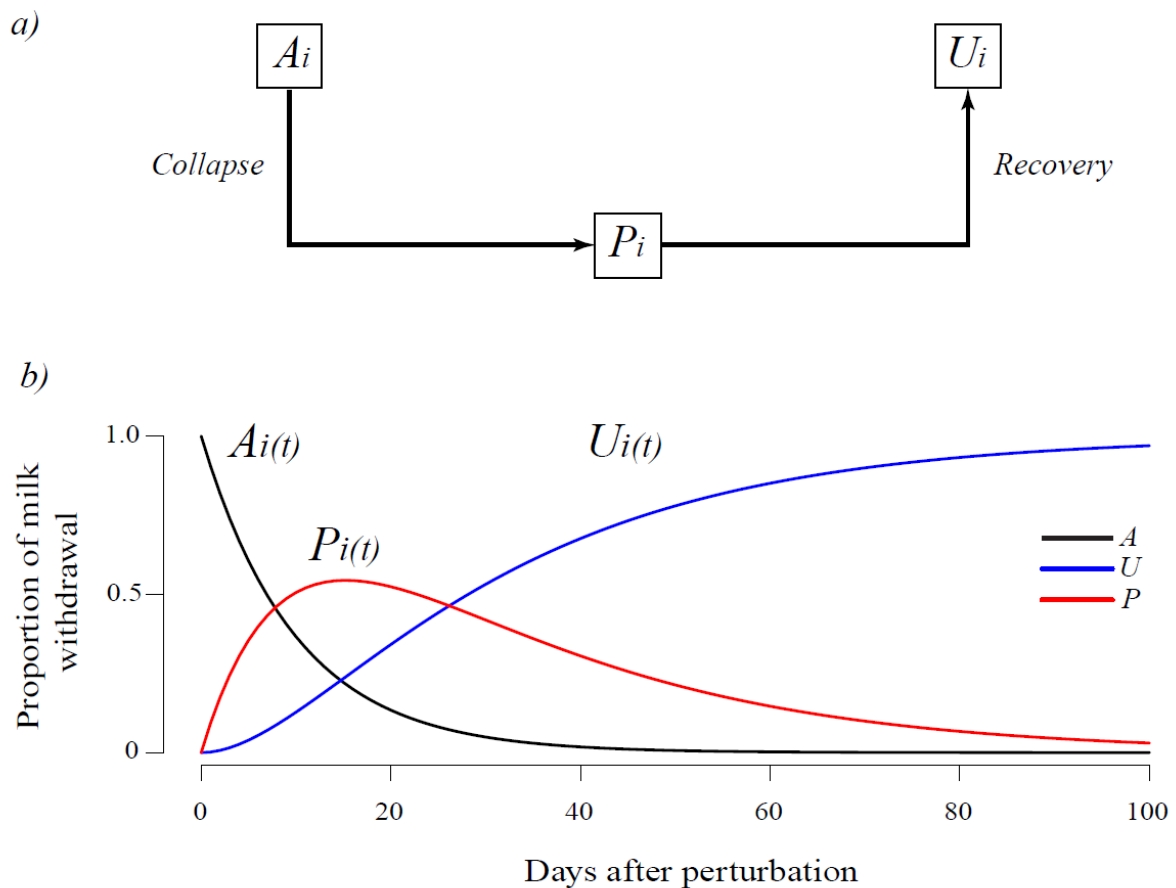
127 The Wood model is given by:

$$128 \quad Y^*(t) = a \cdot t^b \cdot e^{-c \cdot t}$$

129 where $Y^*(t)$ is the unperturbed daily milk yield in kg, t is the time in days after parturition
130 and a , b , c are positive parameters that determine the shape of the lactation curve (a scales the
131 general level of the curve, b controls the type and magnitude of the curvature of the function,
132 and c regulates the rate of decrease in milk yield after the lactation peak). Values of these
133 parameters can be used to calculate some essential features of the lactation curve such as the
134 time of peak yield (b/c , in days), the lactation persistency, *i.e.*, the extent to which peak yield
135 is maintained ($-(b + 1) \cdot \ln(c)$ in $\text{kg} \cdot \text{d}^{-1}$), or the peak yield ($a \cdot (b/c)^b \cdot e^{-b}$ in kg)[16].

136 Perturbation model

137 The perturbation model is based on the idea that each single perturbation I affecting lactation
 138 dynamics can be described as a transient proportional decrease in milk yield, through a
 139 sequence of collapse and recovery. Each perturbation can thus be modelled by way of a 3-
 140 compartment model (Figure 1) representing the dynamics of the proportion of milk withdrawn
 141 from the theoretical unperturbed yield.



142

143 Figure 1. Conceptual model of a single perturbation. A: proportion affected by the
 144 perturbation, P: proportion effectively affected by the perturbation, U: proportion unaffected
 145 by the perturbation. a) Model diagram and b) Solution dynamics.

146 The three compartments of the model are: A_i , the maximal proportion potentially affected by
 147 the i^{th} perturbation, U_i , the proportion unaffected by the i^{th} perturbation, and P_i , the proportion
 148 effectively affected by the i^{th} perturbation. Given the structure of the compartmental model,
 149 forming a path from A_i to U_i through P_i , and given that the model is defined such as $A_i + P_i +$
 150 $U_i = 1$, the dynamics of P_i represents the proportional deviation in milk yield.

151 The perturbation model for a single perturbation i is defined by the following simple
 152 differential system:

$$153 \quad \text{if } t \geq t_{P_i}: \begin{cases} \frac{dA_i}{dt} = -k_{1,i} \cdot A_i \\ \frac{dP_i}{dt} = +k_{1,i} \cdot A_i - k_{2,i} \cdot P_i \\ \frac{dU_i}{dt} = +k_{2,i} \cdot P_i \end{cases} \text{ otherwise: } \begin{cases} \frac{dA_i}{dt} = 0 \\ \frac{dP_i}{dt} = 0 \\ \frac{dU_i}{dt} = 0 \end{cases}$$

154

155 with the following initial conditions at parturition time ($t = 0$):

$$156 \quad \begin{cases} A_i(0) = k_{0,i} \\ P_i(0) = 0 \\ U_i(0) = 1 - k_{0,i} \end{cases}$$

157 and where t_{P_i} is the time of start of the i^{th} perturbation, $k_{0,i}$ is the parameter of intensity of the
 158 i^{th} perturbation ($k_{0,i} \in]0; 1]$), $k_{1,i}$ is the parameter of collapse speed of the i^{th} perturbation and
 159 $k_{2,i}$ is the parameter of recovery speed of the i^{th} perturbation.

160 Assuming that $k_{1,i} \neq k_{2,i}$, the algebraic solution of this differential system is given by:

$$161 \quad \begin{cases} A_i(t) = k_{0,i} \cdot e^{-k_{1,i} \cdot \Delta_i(t)} \\ P_i(t) = \frac{k_{0,i} \cdot k_{1,i}}{k_{1,i} - k_{2,i}} \cdot (e^{-k_{2,i} \cdot \Delta_i(t)} - e^{-k_{1,i} \cdot \Delta_i(t)}) \\ U_i(t) = 1 - \frac{k_{0,i}}{k_{1,i} - k_{2,i}} \cdot (k_{1,i} \cdot e^{-k_{2,i} \cdot \Delta_i(t)} - k_{2,i} \cdot e^{-k_{1,i} \cdot \Delta_i(t)}) \end{cases}$$

162 Where $\Delta_i(t)$ is the elapsed time since the beginning of the i^{th} perturbation and is given by:

$$163 \quad \Delta_i(t) = \begin{cases} 0 & \text{if } t < t_{P_i} \\ t - t_{P_i} & \text{if } t \geq t_{P_i} \end{cases}$$

164 Finally, the perturbation model, including n individual perturbations affecting the lactation
 165 curve is given by:

$$166 \quad \pi(t) = \prod_{i=1}^n (1 - P_i(t))$$

167

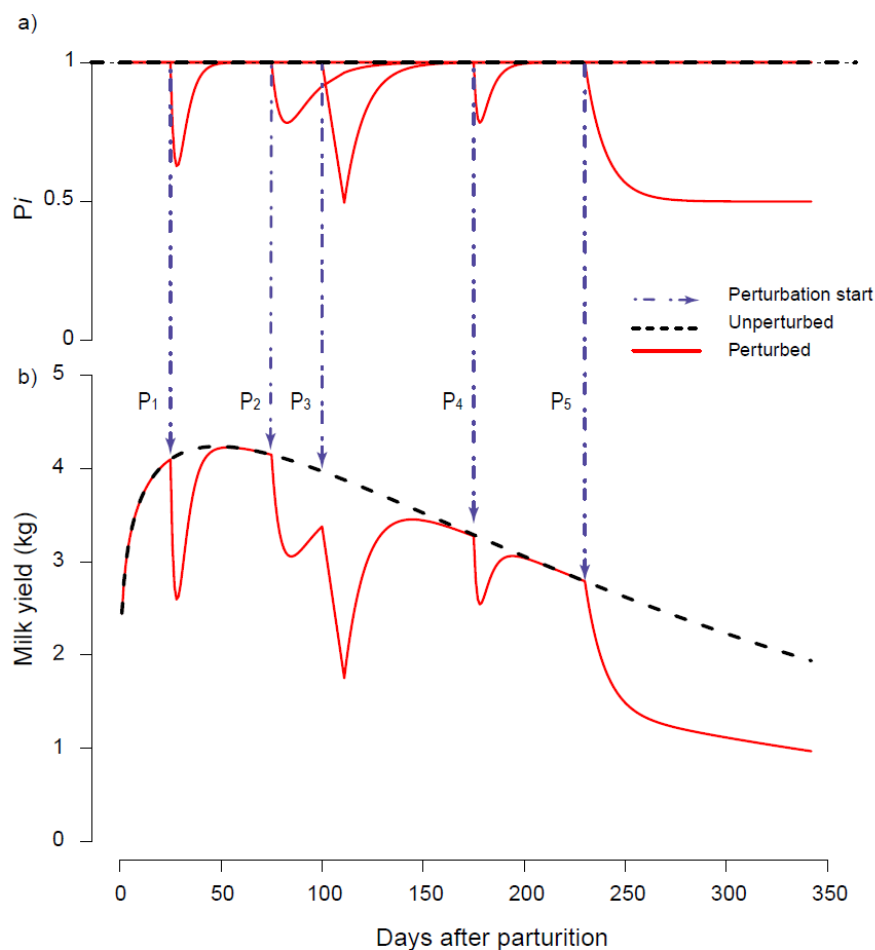
168 Model Formalism

169 The detailed algebraic formula of PLM with n individual perturbations is given by:

$$170 \quad Y(t) = a \cdot t^b \cdot e^{-c \cdot t} \cdot \prod_{i=1}^n \left(1 - \frac{k_{0,i} \cdot k_{1,i}}{k_{1,i} - k_{2,i}} \cdot (e^{-k_{2,i} \cdot \Delta_i(t)} - e^{-k_{1,i} \cdot \Delta_i(t)}) \right)$$

171 The model includes the three parameters of the Wood model (a , b , and c) to define the
172 unperturbed lactation curve, one parameter to define the number of perturbations affecting the
173 lactation curve (n), and four parameters per individual perturbation i (t_{P_i} , $k_{0,i}$, $k_{1,i}$, and $k_{2,i}$)
174 so that the total number of parameters to define PLM is equal to $4 + 4 \cdot n$.

175 A simulation of PLM with five perturbations over 300 days of lactation is shown in Figure 2
176 as an illustration of the model behavior.



177

178 Figure 2. Example of a simulation of the Perturbed Lactation Model (PLM) including five
179 perturbations with a) individual perturbations dynamics expressed as the proportion of
180 unperturbed lactation curve (P_i) and b) unperturbed and perturbed milk yield dynamics.

181 Perturbations are considered individually, so that a perturbation can occur within another one
182 (see P_3 in Figure 2 $att_{P_3} = 100$). Given that individual perturbations are proportional
183 deviations multiplied between them, when a perturbation is added during another
184 perturbation, the new perturbation is a proportion of the already perturbed curve. Moreover,
185 perturbations can be used to simulate the effect of pregnancy (see P_5 in Figure 2 $att_{P_5} = 225$)
186 with the recovery parameter $k_{2,i}$ set to zero.

187 **Fitting procedure**

188 PLM is aimed at detecting perturbations in milk yield time-series data and thus provide
189 estimates of (1) a theoretical unperturbed lactation curve and (2) the number, timing and
190 shape of the perturbations leading to the observed perturbed lactation curve. A dedicated
191 algorithm was developed in R (R Core Development Team, 2018) with the aim of fitting PLM
192 on lactation data and deriving parameter estimates a , b , and c to characterize the unperturbed
193 lactation curve, n to define the number of perturbation and parameter estimates
194 $(t_{P_i}, k_{0,i}, k_{1,i}, \text{ and } k_{2,i})$ for each i^{th} detected perturbation. Preliminary tests have shown that
195 repeated fittings using different starting values can lead to the detection of perturbations
196 differing in total number and detection order. This raised the question of the theoretical
197 identifiability of the model parameters (for further details on identifiability see [17]) and of
198 the use of a stop criterion to estimate n . The structure of the model does not allow a classical
199 identifiability analysis to be performed if n is unknown. However, by using the software
200 DAISY (Differential Algebra for Identifiability of Systems[18]), we could assess that for one
201 perturbation the PLM parameters are locally identifiable. To facilitate the identification of the
202 model parameters, we adopted a fitting strategy in two steps: first, performing numerous
203 repeated fittings to estimate the most frequent number of perturbations. In the second step, we
204 fixed as known the number of perturbations detected in step 1 and proceed to estimate the

205 remaining parameters of the model. This strategy ultimately makes it possible to estimate an
206 optimal number of perturbations and facilitates the estimation of the model parameters.

207 In the following section, PLM_n stands for PLM with n perturbations, k_{W_n} stands for the
208 triplet of parameters (a, b, c) of Wood's model estimated with n perturbations (n ranging from
209 0 to n_{max}) and $k_{P_{i,n}}$ stands for the quadruplet $(t_{P_i}, k_{0,i}, k_{1,i}, k_{2,i})$ of the i^{th} perturbation (n
210 ranging from 1 to n_{max}).

211 The `nls.multstart` package [19] performing non-linear least squares regression with the
212 Levenberg-Marquardt algorithm and with multiple starting values was used for each single fit.

213 Two different sampling schemes of starting parameters were used: random sampling of
214 starting parameters from a uniform distribution within the starting parameter bounds or
215 selection of combinations of starting parameters at equally spaced intervals across each of the
216 starting parameter bounds. These two fitting methods are hereafter referred to as 'shotgun
217 search' and 'gridstart search' respectively. Starting parameter bounds are defined as follows:

218 a : [0; 100]; b : [0; 1]; c : [0; 1]; t_{P_i} : [t_0 ; t_3] (where t_0 and t_3 are the times of first and last
219 records of the dataset); $k_{0,i}$: [0; 1]; $k_{1,i}$: [0; 10]; $k_{2,i}$: [0; 10]. For the 'shotgun search', the
220 number of random combinations of starting parameters was set to 100 000. For the 'gridstart
221 search', the number of combinations of starting parameters (*i.e.*, the size of the grid), was set
222 to five for parameters $a, b, c, k_{0,i}, k_{1,i}, k_{2,i}$ and to 10 for the parameter t_{P_i} . Consequently, for
223 the fit of one perturbation (*i.e.*, estimating $3 + 4 = 7$ parameters) the number of tested
224 combinations of starting parameters was $7^6 \times 10 = 1176490$. For both search methods, the best
225 model was selected on the basis of the lowest Akaike Information Criterion (AIC) score [20].

226 The whole fitting procedure includes repetitions of a fitting sequence that proceeds by
227 successive addition of perturbations. This fitting sequence is defined in such a way that the
228 estimate of the parameters of each new perturbation is obtained while the parameters of the
229 previously added perturbations are kept fixed. Therefore, the fitting of PLM_i provides

230 parameters estimates for the new added i^{th} perturbation and for a new version of Wood
231 model's parameters k_{W_i} (*i.e.*, each time a new perturbation is added, a new version of the
232 unperturbed lactation is refined). For a given lactation dataset composed of daily milk yield
233 records, the preliminary fitting of PLM₀ (*i.e.*, the original Wood's model without any
234 perturbation) was first performed to estimate k_{W_0} . Then, the fitting sequence starts by the
235 fitting of PLM₁ (*i.e.*, PLM with 1 perturbation) thus providing estimates k_{W_1} and $k_{P_{1,1}}$. Then,
236 the fitting of PLM₂ consists in estimating k_{W_2} and $k_{P_{2,2}}$ with $k_{P_{1,2}}$ fixed equal to $k_{P_{1,1}}$. Then,
237 the fitting of PLM₃ consists in estimating k_{W_3} and $k_{P_{3,3}}$ with $k_{P_{1,3}}$ and $k_{P_{2,3}}$ fixed equal to
238 $k_{P_{1,2}}$ and $k_{P_{2,2}}$, respectively. The procedure is applied stepwise until the maximum number of
239 perturbation n_{max} is reached. This maximum number is an *a priori* user defined value to fix a
240 stop criterion. Preliminary tests have shown that setting $n_{max} = 15$ was sufficient. The end of
241 the fitting sequence consists in reordering the n_{max} detected perturbations in decreasing order
242 according to the time of perturbation t_{P_i} (the original obtained order of perturbations is based
243 on the opportunities found by the fitting procedure to improve the goodness of fit for each
244 added perturbation).

245 Finally, the whole fitting procedure is carried out following the 3 following steps:

246 Step1: Repeat 100 times the fitting sequence with the 'shotgun search' and $n_{max} = 15$.

247 Step2: Compare the fitting results of the 100 repetitions obtained in step1 and identify
248 perturbations systematically detected at $t_{P_i} \pm 3$ days. This was performed by counting, for the
249 15 perturbations over the 100 fitting results, the number of occurrences of the rounded
250 value $t_{P_i}^* = round(t_{P_i}/7) \cdot 7$. This step provides the optimal number of perturbations
251 denoted N with an estimate of t_{P_i} for each perturbation (calculated as the median of the
252 t_{P_i} with the same rounded value $t_{P_i}^*$).

253 Step3: Perform the fitting sequence with the ‘gridstart search’, with $n_{max} = N$ and with
254 starting parameters bounds for each t_{P_i} reset to $[t_{P_i} - 10 ; t_{P_i} + 10]$. This last fit provides the
255 final estimates k_{W_N} and $(k_{P_{1,N}}, \dots, \text{and } k_{P_{N,N}})$ characterizing respectively the best fit for the
256 unperturbed model and the N detected perturbations. The Root Mean Square Error was
257 calculated to indicate the goodness-of-fit of PLM_N. Additionally, the percentage of loss ‘ L ’
258 was calculated using the formula $L = 1 - S_0/S_N$ where S_0 and S_N are the total milk yield
259 over $[t_0; t_3]$ calculated with Wood’s model without perturbation using parameters a , b and c
260 from PLM_N and PLM_N (i.e., Wood’s model with N perturbations).

261 To provide complementary information on lactation time-series and refine PLM outputs
262 analysis, the model of Grossman et al. [21] was also fit to lactation data as described in Martin
263 and Sauvant [22]. This fitting cuts the lactation period into three stages corresponding to
264 early, middle and late stages (respectively intervals $[t_0; t_1]$: increasing phase, $[t_1; t_2]$: plateau-
265 like phase and $[t_2; t_3]$: decreasing phase). This triphasic model, based on a smoothing logistic
266 transition between intersecting straight lines, specifies the cut points of the three stages
267 (instead of a priori number of days in milk). This fit was performed using the ‘gridstart
268 search’ with $[t_0; t_3]$ as starting parameters bounds for the interval terminals t_1 and t_2 .

269 **Dairy goat dataset**

270 In this study we used data from 319 lactations (126 primiparous and 193 multiparous; parity
271 ranging from 1 to 7) including 80773 milk records from the dairy goat herd of the INRA-
272 AgroParisTech Systemic Modelling Applied to Ruminants research unit (Paris, France)
273 between 2015 and 2018. Data concerned 181 goats (94 Alpine and 87 Saanen) born between
274 2009 and 2017. Records are shown in supplementary Figure 1 by breed and parity. All
275 lactations considered had at least one record in the first 5 days of lactation and a last record
276 between 150 and 350 days of lactation (no extended lactation included).

277

278 **Statistical analysis**

279 All statistical analyses were performed using R (R Core Development Team, 2018).
280 Fixed effects of breed (Saanen *vs.* Alpine) and parity (1 *vs.* 2 and more) were tested on root
281 mean square error (RMSE), on parameters a , b , and c , on estimated peak milk yield, peak time
282 and total milk yield over $[t_0; t_3]$ for Wood and PLM models, and on the estimated number of
283 perturbation and percentage loss for PLM model, with a mixed analysis of variance model
284 with goat as a random factor. Fixed effect of lactation stage (early *vs.* middle *vs.* late) was
285 tested on RMSE and on PLM parameters t_p, k_0, k_1, k_2 with a mixed analysis of variance
286 model with parity as a random factor. Pearson linear correlations were calculated for PLM
287 parameters: intra-class of breed and parity for a, b, c, N , and L and intra-class of stage of
288 lactation for t_p, k_0, k_1 , and k_2 .

289 **RESULTS**

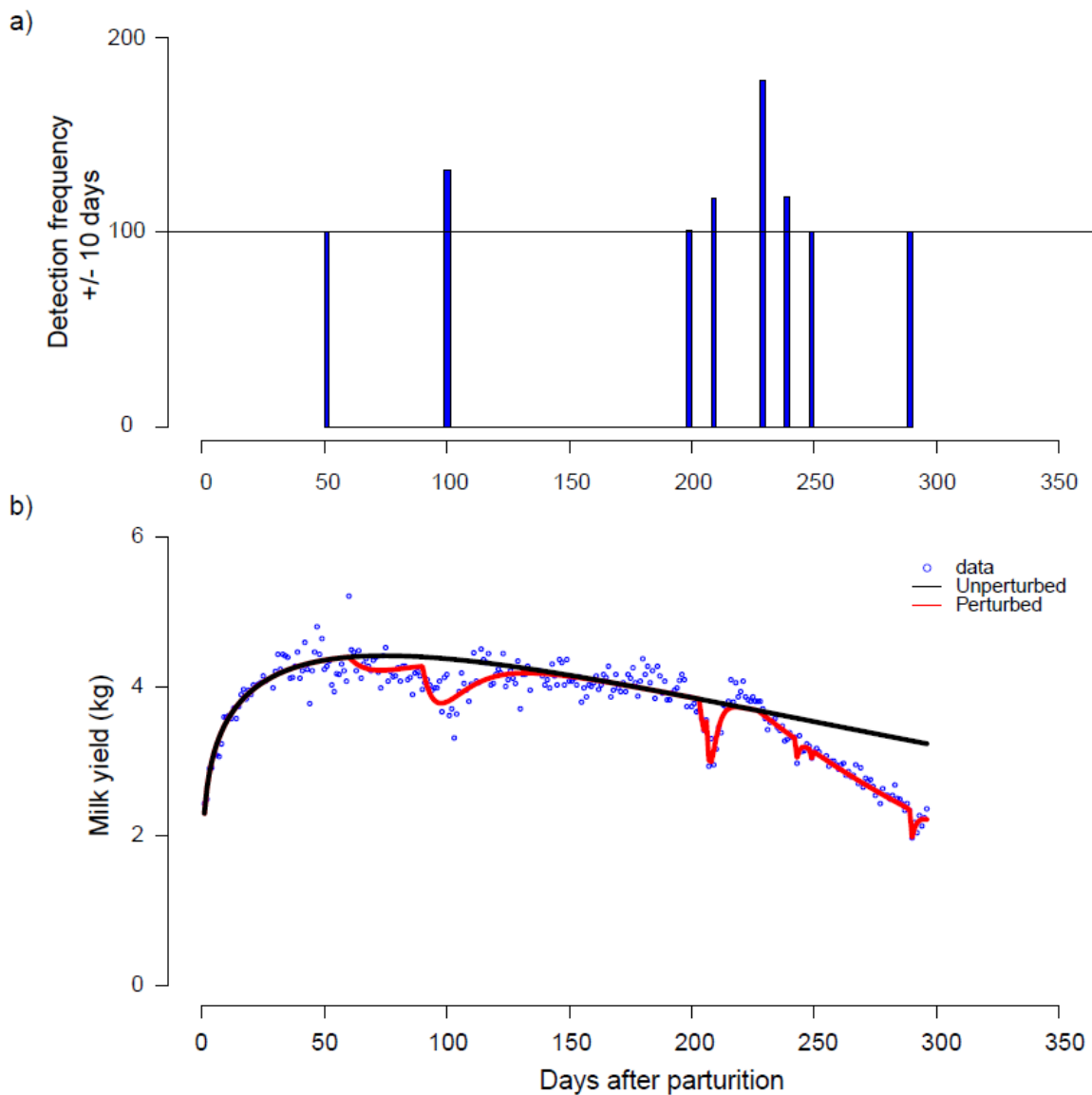
290 Lactation duration ranged from $t_0 = 1.21 \pm 0.64$ to $t_3 = 270.30 \pm 40.77$ days in milk. Early,
291 middle and late lactation stages determined with Grossman's model were [1.21, 34.45],
292 [34.45, 171.05] and [171.05, 270.30], respectively.

293 **Fitting**

294 The fitting procedure converged for the 319 lactations and detected a total of 2354
295 perturbations with an average of 7.40 perturbations per lactation. Figure 3 shows the fitting of
296 PLM on one lactation dataset. The fitting results on individual lactations exhibiting the
297 minimum and maximum values for respectively the RMSE (0.11 kg and 0.41 kg) are provided
298 in supplementary Figure 2. The number of perturbations varied between 4 and 11, the
299 percentage loss between 2 % and 19 %, the total unperturbed milk yield was between 393.56
300 kg and 1557 kg and the record interval length $t_0 - t_3$ was between ([1, 5] to [165, 358] in
301 days). During the first fitting steps, the Wood's parameters were stabilized on average after
302 the detection of the first 4 perturbations (supplementary Figure 3). This indicates the
303 robustness of the unperturbed curve.

304 Descriptive statistics of the results obtained from the fitting procedure of PLM_n are given in
305 Table 1 by breed and parity and are compared to the results obtained with PLM_0 ,
306 corresponding to an adjustment of the Wood model without any perturbation. The value for
307 the parameter a greatly increased between the Wood model and PLM_n . The values for
308 parameters b and c decreased between the Wood model and PLM_n . As a consequence, values
309 for peak milk and peak time increased between the Wood model and PLM_n . Both models did
310 not give a similar level of variance of error according to breed or parity level. Regarding the
311 quality of fitting, the RMSE values showed a fairly significant decline between the Wood
312 model and PLM_n (0.17 ± 0.08 kg). Considering explicit perturbations in the fitting of the

313 Wood model with PLM compare to fitting directly the Wood function to data led to a
314 decrease in RMSE, reflecting an improvement in the quality of the adjustment procedure.



315

316 Figure3. Example of the perturbed lactation model fitting procedure result on a lactation
317 dataset. a) frequency of detection of a single perturbation within ± 10 days; b: unperturbed
318 and perturbed lactation models plotted against data.

319 Table 1. Results of the fitting procedure.

model	All				SAA (143)				ALP (176)				P-value	
	1 (126*)		2 + (193*)		1 (59*)		2 + (84*)		1 (67*)		2 + (109*)		Breed	Parity
Wood ¹	Mean	sd	Mean	sd	Mean	sd	Mean	sd	Mean	sd	Mean	sd		
<i>a</i>	1.884	0.626	2.385	0.793	1.840	0.551	2.443	0.844	1.922	0.686	2.339	0.761	NS	***
<i>b</i>	0.217	0.111	0.242	0.114	0.215	0.109	0.226	0.107	0.218	0.114	0.254	0.118	NS	NS
<i>c</i>	0.004	0.002	0.004	0.002	0.003	0.002	0.004	0.002	0.003	0.001	0.005	0.002	***	***
RMSE ³ (kg/d)	0.308	0.082	0.441	0.136	0.319	0.871	0.461	0.153	0.297	0.076	0.425	0.120	*	***
peak milk ⁴ (kg)	3.543	0.550	4.723	0.715	3.558	0.585	4.686	0.704	3.528	0.521	4.751	0.727	NS	***
peak time ⁵ (d)	63.850	32.180	56.809	22.007	74.280	39.884	60.232	24.758	54.658	19.504	54.170	19.334	*	*
total milk (kg)	719.601	149.136	972.838	204.343	731.910	150.040	986.850	223.174	708.762	148.614	962.039	188.906	NS	***
PLM²														
<i>a</i>	2.159	0.599	2.771	0.690	2.137	0.488	2.890	0.712	2.178	0.684	2.679	0.661	NS	***
<i>b</i>	0.167	0.077	0.185	0.078	0.160	0.065	0.162	0.066	0.174	0.086	0.203	0.081	***	NS
<i>c</i>	0.003	0.001	0.003	0.002	0.002	0.001	0.003	0.001	0.003	0.001	0.004	0.001	***	***
RMSE ³ (kg/d)	0.184	0.040	0.245	0.051	0.193	0.050	0.246	0.042	0.176	0.026	0.244	0.057	NS	***
peak milk ⁴ (kg)	3.573	0.472	4.812	0.709	3.559	0.442	4.751	0.680	3.586	0.500	4.857	0.725	*	***
peak time ⁵ (d)	63.505	25.649	69.459	37.333	77.73	45.065	67.810	32.259	57.795	24.106	60.564	33.258	***	NS
S _N ⁶ (kg)	712.252	147.601	962.423	201.667	723.989	148.533	976.645	220.736	701.917	147.123	951.362	185.470	NS	***
S ₀ ⁷ (kg)	766.280	164.168	1053.915	232.294	780.748	165.600	1069.684	255.555	753.540	163.072	1041.654	212.872	NS	***
N	7.587	1.304	7.380	1.471	7.525	1.278	7.440	1.508	7.642	1.333	7.333	1.447	NS	NS
L (%)	6.016	2.383	7.427	3.502	6.186	2.751	7.512	3.655	5.865	2.014	7.361	3.394	NS	***

320 Signification codes: 0.001: '***', 0.01: '**', 0.05: '*', NS : not significant.

321 ♦ Number of lactation curves

322 ¹ Wood model (1967): *a*, *b*, and *c*: estimated Wood parameters, ²Perturbated Lactation Model based on Wood, ³RMSE: root mean square error of model fit, ⁴peak
 323 milk= $a \cdot (\frac{b}{c})^b \cdot e^{-b}$, ⁵peak time = $\frac{a}{b}$, ⁶total milk based on the PLM perturbed lactation curve: $S_N = \sum_{t_0}^{t_1} y(t)$, ⁷total milk based on the PLM unperturbed lactation curve: $S_0 =$

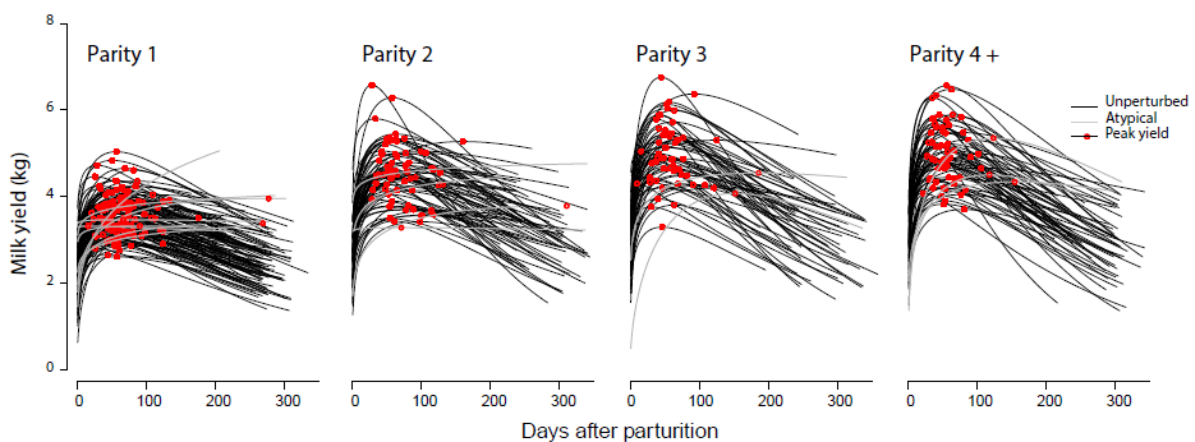
324 $\sum_{t_0}^{t_1} y^*(t)$, N: number of perturbation detected, L :rate of loss milk yield

325

326 **Unperturbed lactation curve**

327 Descriptive statistics of the parameters a , b and c for the unperturbed lactation curves are
328 presented in Table 1 for the overall dataset, breed and parity. The parameter a , which drives
329 the general scaling of the curve, was not significantly different for the two breeds ($2.52 \pm$
330 0.71). Consequently, no significant breed effect was found for the peak milk or for the total
331 unperturbed milk production. The same statistical effects were found with the Wood
332 adjustment without perturbation. The parameter a was significantly affected by the parity of
333 the lactation, with first lactations having a lower value for parameter a than the two and more
334 parities. Consequently, there was a significant parity effect on the peak milk and on the total
335 milk production. The parameter b , which drives the curvature of the lactation curve, was
336 significantly affected by breed. Alpine goats exhibited higher values of b compared to Saanen
337 goats (Alpine: 0.19 ± 0.08 ; Saanen: 0.16 ± 0.06). Parity also had a significant effect on the
338 parameter b , with first lactations having a lower value for parameter b than two and more
339 lactations. Regarding the parameter c , which drives the rate of decrease of milk production
340 after the peak, both parity and breed effects were highly significant. Alpine goats exhibited a
341 higher value for the parameter c than the Saanen goats (Alpine: 0.003 ± 0.001 ; Saanen: 0.002
342 ± 0.001). For this parameter, first lactations had a lower value than two and more lactations
343 (Primiparous: 0.002 ± 0.001 ; Multiparous: 0.003 ± 0.001). The peak time of the unperturbed
344 curve, resulting from both b and c parameters, was significantly affected by breed, with
345 Saanen goats exhibiting a peak 14 days later in lactation than the Alpine goats. The statistical
346 effects found for PLM_n parameters were consistent with the effects found for the Wood model
347 (PLM_0), except for the peak time. Regarding peak time, the Wood model peak time was
348 slightly affected by both breed and parity, while for the PLM_n peak time, breed had a very
349 significant effect and parity was not significant.

350 Individual unperturbed lactation curves obtained with PLM_n for increasing parities are shown
351 in Figure 4. Some of these individual adjusted curves were considered as atypical, in the sense
352 they were not similar to conventional definition of lactation curves. An individual lactation
353 was considered “atypical” if the persistence estimated by PLM, *i.e.* the value of parameter *c*,
354 was an outlier, defined as a value either 3 times above the inter-quartile range (IQR) (above
355 the third quartile of the distribution for the *c* parameter) or 3 times below the IQR (below the
356 first quartile of the distribution for the *c* parameter). A total of 18 curves were classified as
357 atypical. Generally, these atypical curves come from the same goat in different parities or for
358 primiparous that have not started the second parity. The peaks milk of the unperturbed
359 lactation curve were increased by 27.47 % between the first parity and the second parity, by
360 9.46 % between the second parity and the third parity and by -0.29 % between the third parity
361 and the fourth parity (Figure 4). The total milk production for the unperturbed curve was
362 increased by 32.55 % between the first parity and the second parity, 5.20 % between the
363 second parity and the third parity and by 1.01 % between the third parity and the fourth parity.
364 These results are consistent with Arnal et al. [23].

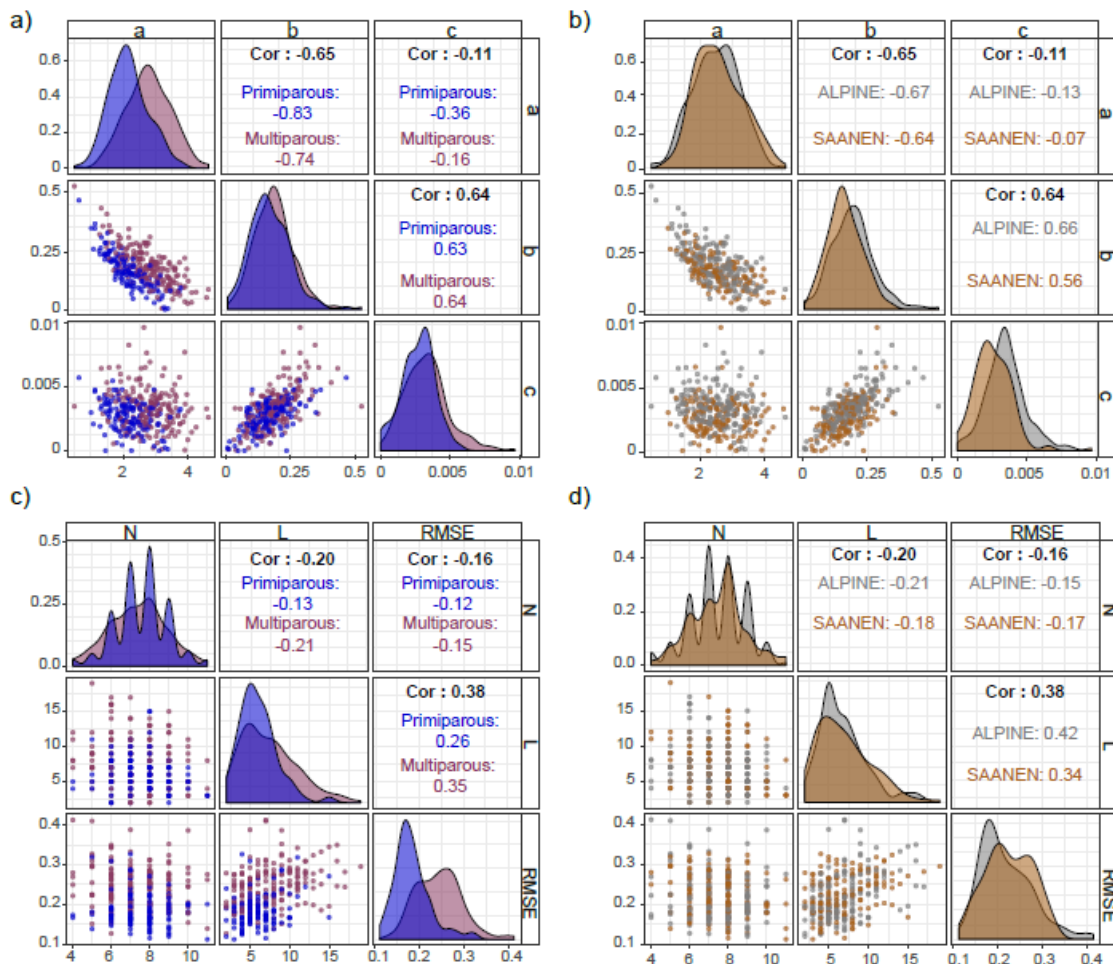


365

366 Figure 4: Individual unperturbed PLM-based lactation curves for increasing parity number (fit
367 on 319 lactation data; atypical curves correspond to outlying estimates of the parameter *c*
368 governing milk persistency).

369

370 The Pearson linear correlation matrix by breed and parity between the PLM-based
 371 unperturbed parameters is shown in Figure 5 (panels a and b). A strong negative correlation
 372 was found between a and b (-0.65), indicating that high values of a (scaling of the lactation
 373 curve), were associated with low values of b (shaping the curve). A positive correlation was
 374 found between the parameters c and b (0.64) indicating a positive association between the
 375 shape of the curve and the rate of decrease of lactation. Finally, a low negative correlation
 376 between c and a (-0.11) was found. These results are consistent with the well-known features
 377 of lactation curves: higher milk at peak yield being associated with higher speed of decline
 378 after peak [24].



379

380 Figure 5. Pearson linear correlation matrix on the PLM-based parameters estimates: panels (a)
 381 and (b): the a , b , c parameters defining the unperturbed curve (a: by parity and b: by breed).
 382 (c) and (d) : the number of perturbations N , milk loss and RMSE (c: by parity and d: by
 383 breed).

384 **Number of perturbations and milk loss**

385 The effects of parity and breed on the total number of perturbations were not significant (7.59
386 for the primiparous 7.38 for the multiparous and 7.45 for the Alpine and 7.47 for the Saanen).
387 By contrast, the rate of milk yield loss was significantly affected by the parity. A Pearson
388 linear correlation matrix by breed and parity between PLM-based estimates of the number of
389 perturbations (N), percentage loss of milk yield (L) and goodness of fit RMSE was also
390 carried out (Figure 5, panels c and d). A positive correlation was noted between RMSE and
391 milk loss (0.38). However, weak negative correlations between the number of detected
392 perturbations and RMSE (-0.16), and the number of perturbations and the milk loss (-0.20)
393 were also noted. Distributions of N , L and RMSE showed an even larger difference according
394 to the parity than to the breeds. These results show that it is not the number of perturbations
395 that contribute the most to the loss in milk yield over the lactation.

396 **Perturbation timing and shape**

397 Table 2 gives descriptive statistics on the parameters of PLM characterizing the 2354
398 perturbations detected during the fitting procedure: time t_p , intensity k_0 , collapse speed k_1 and
399 recovery speed k_2 according to the lactation stage determined with Grossman's model. Most of
400 the perturbations were detected during the late stage of lactation ($n = 1063$). The number of
401 perturbations tended to decrease in middle stage ($n = 1054$) and for early stage ($n = 237$). The
402 parameter k_0 increased from early, middle and late lactation stage. These results suggest that
403 throughout the lactation process, perturbations become more intense. The parameter k_1
404 decreased from early to late stages of lactation. This suggests that perturbations tended to be
405 sharper at the beginning of lactation, with a high speed of collapse and recovery, while they
406 tended to be more smooth as lactation progressed. Several factors (*e.g.*, breed, parity,
407 seasonality and season of kidding) can affect characteristics of the lactation curve. The
408 differences found in this study between primiparous and multiparous goats are consistent with

409 previous studies [23,24] with primiparous goats being less productive, with a lower peak yield
 410 and a greater persistency. Despite the lack of a significant effect of parity, our results are
 411 consistent with previous studies [24] where primiparous goats had a peak later than
 412 multiparous (see Table 1). The strong breed effect we observed on peak time is consistent
 413 with previous studies [24] with Saanen goats having a peak yield later than Alpine goats.

414 Table 2. Descriptive statistics of perturbation parameters for the 2354 perturbations detected
 415 by the perturbed lactation model in the dairy goat lactation dataset.

Perturbations	Stage of lactation (2354)					
	Early (237)		Middle(1054)		Late (1063)	
	Mean	sd	Mean	sd	Mean	sd
tp : time	33.767	34.000	107.183	62.996	202.182	59.584
k0 : intensity	0.450	0.331	0.506	0.349	0.672	0.359
k1 : collapse	4.013	4.170	3.407	3.870	2.760	3.694
k2 : recovery	1.128	1.961	1.181	1.794	0.954	1.714

416
 417 The PLM parameter k_0 , which drives the intensity of the perturbation, varied considerably
 418 between 0.001 and 1 (set as a boundary). The parameter k_1 , which drives the collapse speed of
 419 the perturbation varied between 0 and 10. The parameter k_2 , which drives the speed of
 420 recovery, varied between 0 and 10. A gradient according to the stage lactation was noted for
 421 these parameters with a gradual increase in k_0 and a gradual decrease in k_1 and k_2 according
 422 to early, middle and late lactation stages. In the late stage, 30.20 % of the perturbations were
 423 detected with a parameter k_2 equal to 0, which implied a perturbation without any recovery
 424 period. Among these perturbations, 85.39 % had a k_0 value equal to 1. On the other hand, in
 425 the early and middle stages, the perturbations detected with an k_2 equal to 0 were 1.70 % and
 426 7.07 %, respectively.

427

428

429 **Discussion**

430 **1) Combining two types of models**

431 In this study, we described the PLM model, a tool for extracting simultaneously perturbed and
432 unperturbed lactation curves from daily milk time-series. The key original feature of PLM is
433 to combine an explicit representation of perturbations with a mathematical representation of
434 the lactation curve.

435 Regarding the mathematical representation of the lactation curve, the structure of PLM is
436 generic and any equation can be used to describe the general pattern of milk production
437 throughout lactation (see appendix including Figure 4 showing illustrating results with other
438 lactation models). The Wood model was chosen in this study as it is one of the most well-
439 known and commonly used mathematical model of lactation curve. Behind the choice of
440 considering a general pattern of lactation that is distorted by perturbations, the biological
441 assumption is that the dairy female has a theoretical production potential (the unperturbed
442 curve) corresponding to the expression of its genetics. This genetic potential may be not fully
443 expressed in the farm environment because of perturbations (the perturbed curve).

444 Regarding the representation of perturbations, we chose an explicit formalism with a
445 compartmental structure. Developing models that are able to capture perturbations in lactation
446 curve is a longstanding issue in animal sciences. Historically, perturbations in milk production
447 data were considered as impairing the quality of fitting of the mathematical equation of the
448 lactation curve. Therefore, authors have developed approaches to take into account external
449 factors that alter the lactation curve. Wood [25] himself was the first to modify his model in
450 order to consider external factors affecting the shape of the lactation curve with a depressed
451 production during the winter months (18-8 % in January) and an increased production in
452 spring (14-7 % in May) regardless of the stage of lactation. With the same idea of altering the
453 general model of the lactation curve to increase the goodness of fit, models were developed to

454 be more representative of the variability in the lactation curve. For example, Dhanoa [26]
455 showed that by considering the time required to achieve maximum milk yield in the Wood
456 model, the correlations between non-linear parameters were reduced. After this, Dhanoa and
457 Le Du [27] introduced the autocorrelation notion between milk yield in a given stage of
458 lactation and yield in the preceding stage. Another example is provided by Goodall and
459 Sprevak [28] that, based on the Wood model, developed a stochastic model for milk yield to
460 improve the fit of the lactation curve. The relationship between the maximum milk yield and
461 Wood's parameter a is linear, but all three parameters affect maximum milk yield. Thus, any
462 model attempting to explicitly represent alterations in milk yield and under-achievement
463 relative to a theoretical potential should not be conceptually applied to parameter a but to the
464 whole function.

465 With the development of on-farm data acquisition, allowing more frequent milk production
466 measurements, and the development of more sophisticated statistical methods, modelling
467 approaches have moved toward an explicit consideration of perturbations, instead of just
468 eliminating them to improve the overall fitting of the lactation curve. Codrea et al. [13]
469 studied the effect of nutritional challenge on the lactation curve using differential smoothing
470 procedures for quantifying biological perturbations in animal performance. Results of this
471 experience highlighted the decline in milk yield during the challenge period for each cow, and
472 showed the presence of other deviations with unknown causes or unrelated to the "off-feed"
473 experiment. Friggens et al. [4] used a clustering procedure linked to a piecewise mixed model
474 to characterize different responses between lactation stages and types of response for the
475 nutritional challenges. Other studies have highlighted the large differences in milk production
476 in goats that are subject to the same dietary and environmental conditions [29]. There are few
477 other approaches to describe the shape of the lactation curves from animals faced to health
478 problems. Lescourret and Coulon [30] had shown the huge variability of milk production

479 response to mastitis in both form and intensity. Adriaens et al. [1] developed a novel
480 methodology to predict quarter milk yield during clinical mastitis. The main shortcoming of
481 these approaches is the lack of an explicit representation of perturbations which are only
482 captured through statistical objects. To overcome this limit, models have been developed with
483 a more explicit representation of perturbations. In the work of Revilla et al. [31] on growing
484 piglets, a classical Gompertz equation, used to capture the unperturbed growth curve, is
485 combined to an equation of the perturbation, used to capture the perturbation in body weight
486 change induced by the weaning event. Sadoul et al. [32] used a model based on a spring and a
487 damper to capture perturbations in physiological responses to challenges. This formalism
488 allows to characterize perturbations with stiffness and resistance to the change of the system.
489 The same concept has been applied to dry matter intake data [33]. These recent developments
490 exhibit limits for capturing perturbations in the lactation curve. They were not extended to
491 make it possible to capture multiple perturbations that may be imbricated. PLM overcomes
492 these limitations as it allows the capture of multiple perturbations with contrasted features:
493 from a sharp and short drop (for instance due to a diarrhoea episode) to a long and slow
494 decrease (for instance due the gestation status). PLM also allows to determine the time at
495 which the perturbations occur. This last point is of great interest to add value to on-farm data
496 where challenge imposed to animals are not controlled and arise from the farm environment.
497 Like all lactation models, the good functioning of PLM depends on several factors. The most
498 important factor is the quality of datasets. If there is an inconsistency in the data, PLM loses
499 its relevance.

500 By combining a general model of lactation curve with an explicit model of perturbations,
501 PLM provides two key outputs: first, the unperturbed curve of the female which reflects its
502 production potential in a non-perturbed environment and second the perturbed curve which
503 reflects the production permitted by the farm environment. The PLM parameters ($k_{0,i}$,

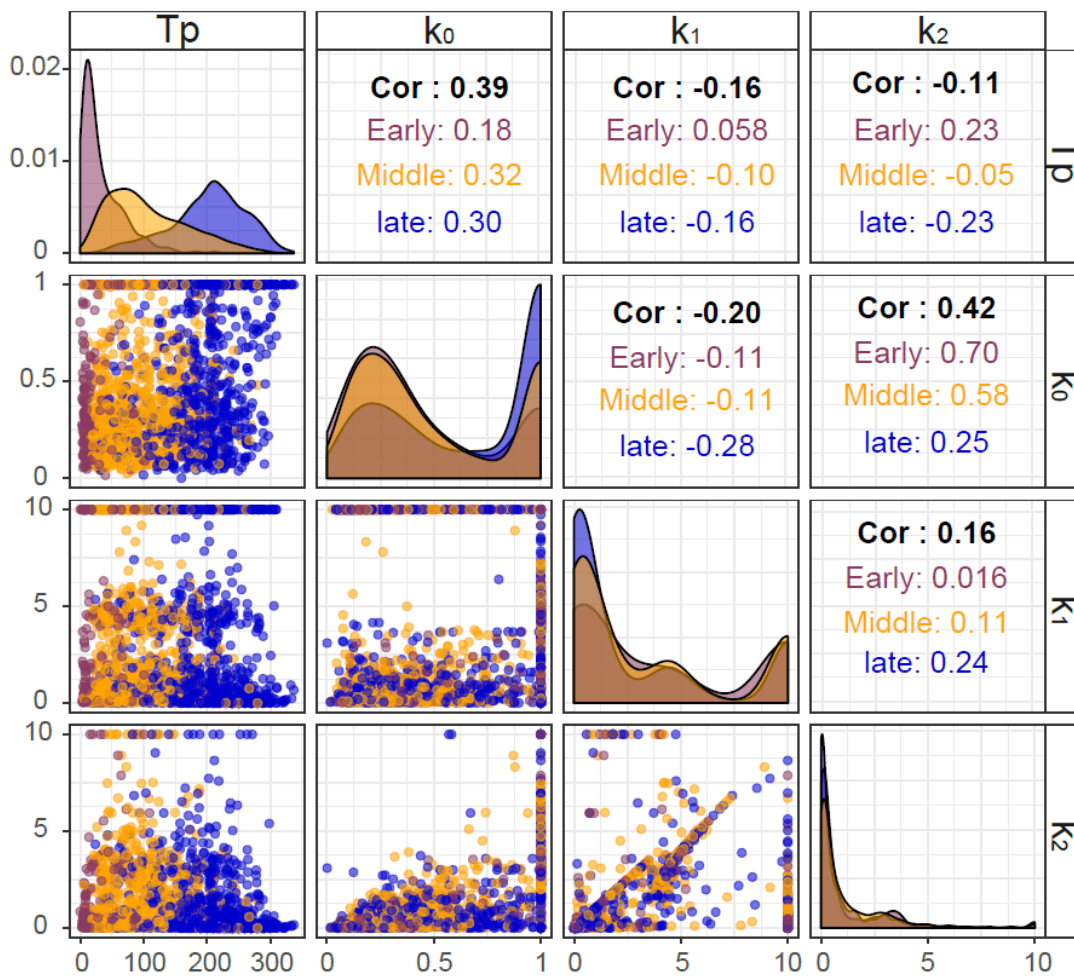
504 $k_{1,i}$ and $k_{2,i}$) provides the most useful information on characteristics of the perturbed lactation
505 curve including scale and shape for each perturbation. Indeed, by providing a perturbed curve,
506 we give an estimate of the number of perturbations and for each perturbation an estimate of
507 intensity ($k_{0,i}$, the collapse $k_{1,i}$ and recovery $k_{2,i}$ speed with a good capture of the time of the
508 perturbation. This not only allows PLM to be flexible in capturing different types of
509 perturbations (*e.g.* gestation, drying), but also to produce metrics to compare the effect of
510 these perturbations on milk yield. In such cases, and by introducing the information
511 concerning these perturbations as an explicit component in the Wood model, we force the
512 model to take into account these perturbations to build the unperturbed curve.
513 With the development of on-farm technology measurements, an interesting perspective for
514 PLM is to be used on other biological time-series data (*e.g.* body weight, dry matter intake,
515 hormones).

516 **2) Fitting algorithm**

517 Beyond the original concepts behind PLM, a key methodological development has been the
518 fitting algorithm. The number of parameters to be determined is substantial, between on the
519 one hand the Wood parameters of the unperturbed curve, and on the other the PLM
520 parameters (time of perturbation and 3 parameters for each perturbation). To overcome the
521 difficulty of estimating a high number of parameters, a 2-step algorithm was implemented.
522 The first step of the procedure is to determine Wood parameters and the time when the
523 perturbation starts. The second step of the procedure is to determine PLM parameters.
524 Another difficulty in PLM development has been the choice of a maximum number of
525 perturbations. After several attempts, this 2-step algorithm was selected for three main
526 reasons. The first one was related to the visual quality of the fitting results itself. Indeed, the
527 obtained fitted curve is always very close to what the human hand would have drawn after
528 simply looking at the raw data and wondering what could be the curve without perturbations.

529 This proximity to what the human eye could have inferred was considered decisive, although
530 subjective. The second reason was related to the issue of finding the number of perturbations.
531 The procedure allows an automated determination of an optimal number of perturbations,
532 without *a priori* or use of an arbitrarily chosen stopping criterion. Preliminary results have
533 shown that allowing a maximal number of 15 perturbations to be detected in the first step of
534 the algorithm was enough for the considered dataset. The third reason pertained to the model
535 identifiability issue [17]. Since the fitting is based on a huge number of repeated fittings from
536 which the systematically detected times of perturbations are retained, the 2-step fitting
537 algorithm facilitates the practical identifiability of the model parameters. Indeed, the overall
538 fitting algorithm was applied several times to the same dataset. That the parameter estimates
539 were the same between the different runs strengthen the convergence properties of the
540 algorithm.

541 Fitting results (see Figure 6) have shown that, in some cases, parameter estimates
542 characterizing an individual perturbation reached their initial upper boundaries (1 for
543 parameter $k_{0,i}$ and 10 for parameters $k_{1,i}$ and $k_{2,i}$). This situation concerns perturbations with
544 a narrow and deep peak-shape. By construction, as a percentage, the value of the parameter
545 k_0 is not supposed to exceed 1. For the parameters k_1 and k_2 , a value of 10 already represents
546 a very abrupt collapse or recovery, respectively. These results are therefore considered
547 relevant. However, a next step may be to test the model on a larger dataset to assess the need
548 to broaden these boundaries. Furthermore, another working step will consist in developing an
549 application where the settings of the PLM algorithm can be user-defined (for instance, the
550 maximal number of detectable perturbations or the size of the search grid in step one,
551 boundaries of parameters, etc)



552

553 Figure 6: Pearson linear correlation matrix on the PLM parameters by stage of lactation: (t_p :
 554 perturbations times detected; k_0 : intensity, k_1 : collapse and k_2 : recovery of perturbation.)

555 3) Phenotyping tool

556 PLM has been developed to improve our ability to phenotype animals by extracting biological
 557 meaningful information from raw data. The unperturbed curve fitted by PLM makes it
 558 possible to compare animals based on their potential of production. With this information,
 559 animals can be ranked based on the production level they would have achieved in a non-
 560 perturbed environment, instead of being ranked based on the measured production level. This
 561 ranking may be of interest for the farmer's breeding strategy, avoiding to cull animals that
 562 have faced a challenge and decreased their production while still having high genetic merit.

563 The perturbed curve and the characteristics of each perturbation (time, intensity, collapse and
 564 recovery) open the perspective of working on perturbations as such and using this information

565 for breeding and management. As a phenotyping tool, PLM can be useful for genetic
566 selection. Studying characteristics of perturbations throughout many lactations of a large
567 number of individuals and linking them to genetic or genomic information opens perspectives
568 to evaluate their heritability and their potential genetic basis. PLM can also be a valuable tool
569 for on-farm management. Linking perturbations with other information on the animals (such
570 as lactation stage, parity, gestation stage...) can help to detect sensitive periods where
571 perturbations are more likely to occur. By cross-checking information on perturbations from
572 all animals with information on the farm environment (for instance temperature, diet quality),
573 it would be possible to detect synchronous occurrences of perturbations and link them to farm
574 environment. With this better understanding of environmental effects on animal production,
575 preventive measures at farm scale could be undertaken.

576 Understanding the effects of the environment on farm animals and how they cope with
577 perturbations is crucial to gain insights on resilience and robustness. These complex dynamic
578 properties are highly desirable to face the changes occurring in the livestock sector [34].
579 While the conceptual framework to work on resilience and robustness is now well defined in
580 animal sciences, we still need operational metrics [35]. Such metrics have been proposed for a
581 single perturbation (*e.g.*, Revilla et al., [31]; Sadoul et al. [32]). To our knowledge, existing
582 metrics for the lactation curve, as proposed by Elgersma et al. [36], are based on a variance
583 approach applied to the whole curve. Fluctuations in milk yield are summarized with a single
584 statistical measure. Complementary to this type of approach, PLM can decompose the whole
585 curve and characterize each perturbation, with metrics that are consistent with the concept of
586 resilience (intensity, collapse, recovery). It offers a way of quantifying the consequences of
587 external factors and exploring hypotheses about the biological types of response. By giving a
588 biological meaning to these parameters, we reconcile a phenotyping tool with the opportunity
589 of an explanatory approach.

590 **Conclusion**

591 By combining a general description of the lactation curve with an explicit representation of
592 perturbations, the PLM model allows the characterization of the potential milk production,
593 reflecting animal genetics, and the deviations induced by the environment, reflecting how
594 animals cope with real farm conditions. The translation of raw time series data into
595 quantitative indicators makes it possible to compare animals and bring insights on their
596 resilience to external factors. In that sense, PLM is a valuable phenotyping tool and it
597 contributes to provide decision solutions for dairy production that are grounded in a
598 biologically meaningful framework. Further modelling studies should strive for integrating
599 high throughput data analysis with such biological framework.

600

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609

610 **References**

- 611 1. Adriaens I, Huybrechts T, Aernouts B, Geerinckx K, Piepers S, De Ketelaere B, et al.
612 Method for short-term prediction of milk yield at the quarter level to improve udder
613 health monitoring. *Journal of Dairy Science*. 2018;101: 10327–10336.
614 doi:10.3168/jds.2018-14696
- 615 2. Huxley JN. Impact of lameness and claw lesions in cows on health and production.
616 *Livestock Science*. 2013;156: 64–70. doi:10.1016/j.livsci.2013.06.012

- 617 3. West JW. Effects of Heat-Stress on Production in Dairy Cattle. *Journal of Dairy Science*.
618 2003;86: 2131–2144. doi:10.3168/jds.S0022-0302(03)73803-X
- 619 4. Friggens NC, Duvaux-Ponter C, Etienne MP, Mary-Huard T, Schmidely P.
620 Characterizing individual differences in animal responses to a nutritional challenge:
621 Toward improved robustness measures. *Journal of Dairy Science*. 2016;99: 2704–2718.
622 doi:10.3168/jds.2015-10162
- 623 5. Pond CM. The significance of lactation in the evolution of mammals. *Evolution*. 1977;31:
624 177–199. doi:10.1111/j.1558-5646.1977.tb00995.x
- 625 6. Coulon JB, Pérochon L, Lescourret F. Modelling the effect of the stage of pregnancy on
626 dairy cows' milk yield. *Animal Science*. 1995;60: 401–408.
627 doi:10.1017/S1357729800013278
- 628 7. Dijkstra J, France J, Dhanoa MS, Maas JA, Hanigan MD, Rook AJ, et al. A Model to
629 Describe Growth Patterns of the Mammary Gland During Pregnancy and Lactation.
630 *Journal of Dairy Science*. 1997;80: 2340–2354. doi:10.3168/jds.S0022-0302(97)76185-X
- 631 8. Grossman M, Koops WJ. Modeling Extended Lactation Curves of Dairy Cattle: A
632 Biological Basis for the Multiphasic Approach1. *Journal of dairy science*. 2003;86: 988–
633 998. doi:10.3168/jds.S0022-0302(03)73682-0
- 634 9. Delage J, Leroy AM, Poly J. Une étude sur les courbes de lactation. *Annales de*
635 *zootechnie*. 1953. pp. 225–267.
- 636 10. Wood P. Algebraic model of the lactation curve in cattle. *Nature*. 1967;216: 164.
- 637 11. Erb HN, Smith RD, Oltenacu PA, Guard CL, Hillman RB, Powers PA, et al. Path Model
638 of Reproductive Disorders and Performance, Milk Fever, Mastitis, Milk Yield, and
639 Culling in Holstein Cows. *Journal of Dairy Science*. 1985;68: 3337–3349.
640 doi:10.3168/jds.S0022-0302(85)81244-3
- 641 12. Huybrechts T, Mertens K, De Baerdemaeker J, De Ketelaere B, Saeys W. Early warnings
642 from automatic milk yield monitoring with online synergistic control. *Journal of Dairy*
643 *Science*. 2014;97: 3371–3381. doi:10.3168/jds.2013-6913
- 644 13. Codrea MC, Højsgaard S, Friggens NC. Differential smoothing of time-series
645 measurements to identify disturbances in performance and quantify animal response
646 characteristics: An example using milk yield profiles in dairy cows1. *Journal of Animal*
647 *Science*. 2011;89: 3089–3098. doi:10.2527/jas.2010-3753
- 648 14. Casado C, Piquer O, Cervera C, Pascual JJ. Modelling the lactation curve of rabbit does:
649 Towards a model including fit suitability and biological interpretation. *Livestock Science*.
650 2006;99: 39–49. doi:10.1016/j.livprodsci.2005.05.019
- 651 15. Ruiz R, Oregui LM, Herrero M. Comparison of Models for Describing the Lactation
652 Curve of Latxa Sheep and an Analysis of Factors Affecting Milk Yield. *Journal of Dairy*
653 *Science*. 2000;83: 2709–2719. doi:10.3168/jds.S0022-0302(00)75165-4
- 654 16. France J, Thornley JH. *Mathematical models in agriculture*. Butterworths. 1984.

- 655 17. Muñoz-Tamayo R, Puillet L, Daniel JB, Sauvant D, Martin O, Taghipoor M, et al.
656 Review: To be or not to be an identifiable model. Is this a relevant question in animal
657 science modelling? *animal*. 2018;12: 701–712. doi:10.1017/S1751731117002774
- 658 18. Bellu G, Saccomani MP, Audoly S, D'Angiò L. DAISY: A new software tool to test
659 global identifiability of biological and physiological systems. *Computer Methods and*
660 *Programs in Biomedicine*. 2007;88: 52–61. doi:10.1016/j.cmpb.2007.07.002
- 661 19. Padfield D, Matheson G. nls. multstart: robust non-linear regression using AIC scores. R
662 package version. 2018;1.
- 663 20. Akaike H. A new look at the statistical model identification, *IEEE T. Automat. Contr.*, 19,
664 716–723. 1974;
- 665 21. Grossman M, Hartz SM, Koops WJ. Persistency of Lactation Yield: A Novel Approach.
666 *Journal of Dairy Science*. 1999;82: 2192–2197. doi:10.3168/jds.S0022-0302(99)75464-0
- 667 22. Martin O, Sauvant D. Metaanalysis of Input/Output Kinetics in Lactating Dairy Cows.
668 *Journal of Dairy Science*. 2002;85: 3363–3381. doi:10.3168/jds.S0022-0302(02)74424-X
- 669 23. Arnal M, Robert-Granié C, Larroque H. Diversity of dairy goat lactation curves in France.
670 *Journal of Dairy Science*. 2018;101: 11040–11051. doi:10.3168/jds.2018-14980
- 671 24. Gipson T, Grossman M. Lactation curves in dairy goats: a review. *Small Ruminant*
672 *Research*. 1990;3: 383–396. doi:10.1016/0921-4488(90)90019-3
- 673 25. Wood PDP. Factors affecting the shape of the lactation curve in cattle. *Animal*
674 *Production*. 1969;11: 307–316. doi:10.1017/S0003356100026945
- 675 26. Dhanoa MS. A note on an alternative form of the lactation model of Wood. *Animal*
676 *Production*. 1981;32: 349–351. doi:10.1017/S0003356100027276
- 677 27. Dhanoa MS, Du YLPL. A partial adjustment model to describe the lactation curve of a
678 dairy cow. *Animal Production*. 1982;34: 243–247. doi:10.1017/S0003356100010175
- 679 28. Goodall EA, Sprevak D. A Bayesian estimation of the lactation curve of a dairy cow.
680 *Animal Production*. 1985;40: 189–193. doi:10.1017/S0003356100025290
- 681 29. Gaddour A, Najari S, Ouni M. Dairy performances of the goat genetic groups in the
682 southern Tunisian. *Agricultural Journal*. 2007;2: 248–253. doi:aj.2007.248.253
- 683 30. Lescourret F, Coulon JB. Modeling the Impact of Mastitis on Milk Production by Dairy
684 Cows. *Journal of Dairy Science*. 1994;77: 2289–2301. doi:10.3168/jds.S0022-
685 0302(94)77172-1
- 686 31. Revilla M, Friggens NC, Broudicou LP, Lemonnier G, Blanc F, Ravon L, et al. Towards
687 the quantitative characterisation of piglets' robustness to weaning: a modelling approach.
688 *animal*. 2019; 1–11. doi:10.1017/S1751731119000843
- 689 32. Sadoul B, Martin O, Prunet P, Friggens NC. On the Use of a Simple Physical System
690 Analogy to Study Robustness Features in Animal Sciences. Pant AB, editor. *PLOS ONE*.
691 2015;10: e0137333. doi:10.1371/journal.pone.0137333

- 692 33. Nguyen Ba H, Van Milgen J, Taghipoor M. A procedure to quantify the feed intake
693 response of growing pigs to perturbations. *Animal*. 2019;
- 694 34. Dumont B, González-García E, Thomas M, Fortun-Lamothe L, Ducrot C, Dourmad JY, et
695 al. Forty research issues for the redesign of animal production systems in the 21st century.
696 *Animal*. 2014;8: 1382–1393. doi:10.1017/S1751731114001281
- 697 35. Friggens NC, Blanc F, Berry DP, Puillet L. Review: Deciphering animal robustness. A
698 synthesis to facilitate its use in livestock breeding and management. *animal*. 2017;11:
699 2237–2251. doi:10.1017/S175173111700088X
- 700 36. Elgersma GG, de Jong G, van der Linde R, Mulder HA. Fluctuations in milk yield are
701 heritable and can be used as a resilience indicator to breed healthy cows. *Journal of Dairy*
702 *Science*. 2018;101: 1240–1250. doi:10.3168/jds.2017-13270
- 703