

1 **Microplastic ingestion by a herring *Opisthonema sp.*, in the Pacific coast of**
2 **Costa Rica**

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18 **Abstract:** Despite there is a growing interest in studying the presence and effects of
19 microplastics (MP) in fishes and other aquatic species, knowledge is still limited in tropical
20 areas. In this study, we examined the presence of MP in the gastrointestinal content of 30 filter
21 feeders of thread herring, *Opisthonema* complex (Clupeiformes: Clupeidae) from the Central
22 Pacific coast of Costa Rica. We detected the presence of MP in 100% of the individuals with
23 an average of 36.7 pieces per fish, of which 79.5% were fibers and 20.5% particles. To our
24 knowledge, this is the first study in Costa Rica that demonstrates the presence of MP in
25 planktivorous fishes. The effects of microplastics ingestion by *O. libertate* and its transit
26 through aquatic food webs should be studied in greater detail, with greater number of sampling
27 points at different times of the year. However, our work confirms that contamination by
28 microplastics is having direct effects on the marine life of Costa Rica.

29

30 **Key words:** Plastic pollution, *Opisthonema*, marine pollution, Tropical Eastern Pacific

31 **Capsule:** This is the first multidisciplinary study in Costa Rica demonstrating the presence and
32 nature of microplastics in the digestive tract of planktivorous fish.

33 **Introduction**

34 Pollution by plastic in the oceans was reported since the second half of the 20th century
35 (Carpenter and Smith, 1972; Shiber, 1979), but the scientific investigation of its implications
36 in marine life has been addressed until recently (Law, 2017). In Costa Rica, few efforts have
37 been made to determine the presence of microplastics at different marine trophic levels, even
38 though it is estimated that more than 500 metric tons of solid waste is discarded per day.

39 Microplastics are defined as particles with a size less than 5 mm that can enter into the
40 environment by a primary form (cosmetic fragments and clothing fibers) or secondary,
41 generated from the decomposition of larger plastic objects by means of ultraviolet (UV) photo-
42 degradation, wave action and physical abrasion (Martin et al., 2017). The chemical
43 composition of these particles depends on different monomers that are used for their
44 production, among which polyethylene, polypropylene, and polystyrene are the most abundant
45 (Güven et al., 2017).

46 As the size of these particles overlaps with the size of zooplankton organisms,
47 planktivorous fish can ingest microplastics directly (Law and Thompson, 2014) or through
48 feeding zooplankton that has previously ingested microplastics (indirectly) (Cole et al., 2011).
49 In this line, (Setälä et al., 2014) evidenced the ingestion of 10 µm polystyrene microspheres in
50 all the planktonic organisms studied, including shrimp, copepods, cladocerans, rotifers, and
51 polychaete larvae. It also demonstrated the transfer of plastic microparticles through planktonic
52 guilds from lower trophic levels (mesozooplankton) to a higher level (macrozooplankton).

53 Among the organisms that could potentially ingest microplastics are filter feeders of
54 the Clupeidae family, whose feeding depends on planktonic life stages (Lozano, 1979). Recent
55 studies have found the presence of microplastics in the digestive tract of clupeid fish (Ory et
56 al., 2018; Tanaka and Takada, 2016). These fish represent a good study model since their short
57 life, and their tendency to group in homogeneous schools provide an updated image of the
58 amount and type of microplastic present in the marine landscape (at any given moment).

59 Due to the enormous increase registered over the last century, both in the production of
60 plastics and in their presence in the oceans worldwide (Ivar do Sul and Costa, 2014), it is
61 necessary to know their scope in marine ecosystems in order to determine its impact and take

62 mitigation measures of possible damages. The objective of this investigation was to determine
63 the incidence of microplastics in the gastrointestinal content of *Opisthonema* sp., a
64 planktivorous fish in the Pacific Coast of Costa Rica and a model for biomonitoring
65 microplastic pollution in the marine ecosystem.

66 **Materials and Methods**

67 *Study site*

68 Samples from commercial purse-seine catches, collected by the Costa Rican semi-
69 industrial sardine fleet, were obtained in October 2018 in the province of Puntarenas, Central
70 Pacific of Costa Rica (09°49.'242N, 084°42.087 W) (**Fig. 1**). Three species of the *Opisthonema*
71 complex (*O. libertate*, *O. bulleri*, *O. medirastre*) sustain the sardine fishery of Costa Rica,
72 although the Pacific thread herring, *O. libertate* tends to be most abundant in their mixed
73 schools during the sampling period (Vega-Corrales, 2010). Non-eviscerated whole specimens
74 were transported fresh from the landing dock to the Center for Research in Marine Sciences
75 and Limnology (CIMAR) of the University of Costa Rica (UCR).

76 *Analysis of samples*

77 Thirty individuals of *Opisthonema* complex (*O. libertate*, *O. bulleri*, *O. medirastre*)
78 underwent a series of biometric analyses in CIMAR laboratories. We measured the standard
79 length (SL), fork length (FL) and total length (TL) of each fish. Subsequently, their total and
80 eviscerated weights were determined on a Mettler PJ360 DeltaRange® granatary scale. Then,
81 each fish was opened with dissection scissors tracing a straight line in the belly from the anus
82 to the preopercular area, extracting the organs associated with the gastrointestinal tract. The
83 gastrointestinal tract was isolated from the mesentery and other structures in order to measure
84 and weigh it. Each specimen was sexed by examining the gonads.

85 *Processing of the intestinal tract*

86 A longitudinal section was made through the whole tract to obtain the gastric content,
87 then it was deposited in a filter paper. The tracts were washed with filtered distilled water and
88 filtered to obtain all the organic matter and the microplastics. The material was dissolved in
89 KOH 10% (previously pre-filtered though 0.2 µm) for the degradation of the organic matter.
90 This mixture was left standing for a minimum period of 48 hr in separate glass bottles.

91 After the incubation, each sample was vacuum-filtered using a Watman cellulose filter
92 paper. The filters were dried under a 100 Watts incandescent lamp and subsequently analyzed
93 under Motic® brand DM-143 stereoscopes. The material obtained were separated into fibers
94 and particles. As a negative control, a portion of the KOH was incubated and later filtered to
95 analyze it under the stereoscope and determine if any MP comes from our protocol.

96 *Fiber and particle analysis*

97 Scanning electron microscopy (SEM): The particles isolated from the gastrointestinal tract
98 of *O. libertate* were mounted on carbon tape and sputtered with gold using a Denton Vacuum
99 Desk V sputter system at 20 mA for 300 s. Images were taken using a JSM-6390LV (JEOL,
100 Tokyo, Japan) SEM, with an accelerating voltage of 20 kV, under high vacuum. Energy-
101 dispersive X-rays (EDX) were measured with liquid nitrogen cooled Inca X-sight Si detector
102 (Oxford Instruments). EDX data was analyzed with Inca Suite version 4.08.

103 Fourier-transform infrared spectroscopy with attenuated total reflection (FTIR-ATR): The
104 spectra were collected in the range 4000-500 cm^{-1} , using a Nicolet 6700 Thermo Scientific
105 spectrophotometer with a diamond ATR crystal.

106 Differential scanning calorimetry (DSC). The analysis of *ca.* 1 mg of particles was performed
107 in a Q200 TA Instruments calorimeter, under nitrogen atmosphere in the temperature range -
108 80 – 200 °C, at a rate of 20 °C min^{-1} .

109 *Statistical analysis*

110 Data processing and statistical analysis were performed in R (R Core Team, 2018).
111 Visualizations were generated with the program ggplot (Wang et al., 2017) (Wickham, 2016).
112 The statistical differences in the number of microplastic particles were estimated using the non-
113 parametric Kruskal-Wallis test.

114

115 **Results**

116 The most notorious result of this work is that microplastics were found in 100% of the
117 individuals. We counted a total of 1100 pieces in the 30 individuals analyzed, of which 20.5%
118 were classified as particles and 79.5% were fibers. The average number of pieces per fish was
119 36 (range 32 to 42) where the average number of fibers was 29 (range 25 to 34), and the average
120 number of particles was 5 (range 6 to 10). These results represent the first stage of our work;
121 therefore, sample number should be increased in subsequent studies.

122 From the 30 specimens of *Opistonema* sp., five were females and 25 males. The
123 measurements of the main characteristics of the fishes (including standard, fork, and total
124 length; length of the gastrointestinal tract; average weight with and without evisceration;
125 average weight of the full and empty tract) grouped by sex are shown in **Fig. 2**. We also
126 observed a trend of higher total number of MPs in females than in males, but results were not
127 statistically significant (**Fig. 3**). Additional studies with more samples from each sex are
128 required to determine if females have higher ingestion rates of microplastics than males. We
129 did not find any other apparent association between the variables measured and the ingestion
130 of microplastics. This could be relevant since it shows that this species can be used as a standard
131 model for the biomonitoring of microplastic in the sea.

132 The main shapes and sizes of the particles and fibers found in the digestive tract of the
133 fish were photographed using a stereoscope (**Fig. 4**). In addition, SEM was used to determine
134 which of the structures found corresponded to MP. The analysis showed three types of
135 structures to be screened (**Fig. 5**): fibers (tagged as A), incrustations (attached to fibers, tagged
136 as B) and a couple of standalone particles (tagged as C). EDX spectroscopy was run on selected
137 structures to determine the chemical composition. When no incrustations were observed around
138 the fibers, only C and O were detected, which agrees with organic substances. Incrustations
139 typically added Ca, K and Si to the composition. Similar crusts have been reported on other
140 MP studies (Wang et al., 2017). A partially mineralized microorganism is tagged with an
141 asterisk symbol. Other SEM images showing mineraloid incrustations in the fibers are available
142 in supplementary information. The standalone particles tagged as (C) in **Fig. 5** were inorganic,
143 with no Carbon detected (see EDX results for rod-like structure as an example -the other
144 standalone structure tagged as (C) was an iron-rich aluminosilicate). This is relevant since, in
145 many cases, the classification of microplastics in fibers and particles by optical microscopy
146 could generate an overestimation of MP if their chemical nature is not verified by other
147 methods.

148 FTIR-ATR and DSC were used to determine the types of polymers contained in the
149 samples. The FTIR-ATR spectrum (**Fig. 6**) shows typical signals for thermoplastic polyolefins,
150 such as polyethylene or polypropylene: the CH₂ asymmetric stretching (2917 cm⁻¹) and CH₂
151 symmetric stretching (2839 cm⁻¹), the CH₃ symmetric deformation (1376 cm⁻¹) and the CH₂
152 bending deformation (1456 cm⁻¹) (Gulmine et al., 2002). Moreover, there are some weak
153 signals around 3300 cm⁻¹ and 1700 cm⁻¹, suggesting a small grade of oxidation of the polymer.

154 The DSC curve (**Figure 6.B**) shows two main signals, an exothermic signal at 120 °C
155 and an endothermal signal at 162 °C, which represent the crystallization temperature (T_c) and
156 the melting point (T_M) of the sample, respectively. As many thermoplastics, polypropylene has
157 a wide range of melting and crystallization temperatures, depending on molecular weight and
158 presence of functional groups. (Majewsky et al., 2016) reported the endothermic peaks (T_M)
159 by DSC for typical polymers in order to identify them in MP samples. The reported T_M for
160 polyethylene was 101 °C, 164 °C for polypropylene, and 250 °C for polyethylene terephthalate
161 (PET).

162 The FTIR-ATR and DSC results of the batch analyzed suggest the presence of
163 polypropylene in our MP samples, which is expected since it is widely used in packaging,
164 labeling, containers, and others. It is important to mention that these results do not exclude the
165 possible presence of other less concentrated polymers in the sample. In addition, the proportion
166 of the sample analyzed was very low compared to the total number of pieces that were found.

167

168 **Discussion**

169

170 *The utility of Opistonema as a model for biomonitoring*

171

172 To our knowledge, this is the first study in Costa Rica demonstrating the presence of
173 microplastics in the digestive tract of planktivorous fish of the Clupeidae family. Although
174 there are no other published studies in the country to compare with, this is the first study in the
175 region showing 100% of the samples containing microplastics in their intestinal tracts and also
176 accounting for the highest number of microplastic pieces per fish (Espinoza and Bertrand,
177 2008; Ory et al., 2018, 2017; Tanaka and Takada, 2016). Despite the low number of samples
178 analyzed, this study sets a precedent and calls for the continuous monitoring of the effects of
179 microplastic contamination on the coasts of the region.

180 The three measures of length showed little variation among the fish, and we found no
181 correlation between average weight and length with the number of microplastics. The

182 homogeneous biometric characteristics of the mixed schools of *Opisthonema* allows us to
183 suggest that this species complex could be used as a model biomonitoring studies of MP
184 pollution. In addition, the feeding type of this species makes it more prone to the intake of
185 microplastic, since, by suction, they cannot discriminate the presence of microplastic (Moore
186 et al., 2002, 2001). This type of feeding could explain the high number of pieces found per fish
187 as well as the small differences between individuals. In this regard, (McNeish et al., 2018)
188 concluded that filter fish tend to have more particles and plastic fibers than others with a
189 different type of feeding, due to the trophic transfer of the plastic elements consumed by the
190 prey.

191 As we found fibers of different colors, it is possible that there is no discrimination for
192 this characteristic (Tanaka and Takada, 2016). In contrast (Ory et al., 2017) reported that in
193 *Decapterus muroadsi* (Carangidae) the microplastic capture could be due to a confusion
194 between the color of the particle and the color of its prey. The fact of having found so many
195 pieces in short-lived fish might serve as a base for estimating the number of pieces that are
196 floating in the photic zone, which is where this species usually inhabit. To have a complete
197 picture, it will be necessary to analyze water samples from the nearby areas as well (Güven et
198 al., 2017).

199

200 *Comparison between the Pacific coast of different countries*

201

202 Different studies performed on the Pacific coast of other countries have shown the
203 presence of MP in the different levels of the trophic chain (Law and Thompson, 2014). For
204 example, research conducted in the Pacific coast has shown the existence of MP in filter fish
205 from Japan (77%), Chile (Easter Island) (80%) and in California, United States (35%).
206 However, in countries such as Peru, Colombia, and Panama, no microplastics were detected
207 ((Boerger et al., 2010; Espinoza and Bertrand, 2008; Ory et al., 2018, 2017; Tanaka and
208 Takada, 2016) (**Table I**). Since *O. libertate* was also used in Colombia for biomonitoring MP
209 contamination, we propose this species as a model for comparison between countries in the
210 Pacific coast of the region. An explanation for the differences between countries in the region
211 can be related to the effects of the marine currents, that transport the microplastics, and that
212 convergence near to Central America and North America (Law, 2017). However, we consider
213 that the proximity to urban areas with a high degree of pollution is the most important agent.

214

215

216 *Implications in marine life*

217 Floating plastics can be transported to greater depths by increasing the density induced
218 by biofouling and can be ingested by migratory species. Many of these processes have been
219 demonstrated in laboratory and field experiments, but their rates at a global scale remain
220 unknown (Law, 2017). The latter could also present a risk to other organisms of different
221 trophic levels, such as crustaceans or birds that feed on other fish. About other implications of
222 the MP for the marine species, it has been proven that MP releases toxic substances that include
223 residual monomers, plasticizers, coloring agents, among other additives, that can be ingested
224 and produce bioaccumulation (Worm et al., 2017). In laboratory studies, it was found that MP
225 particles ingested by fish of the Clupeidae family (*Alosa fallax*) passed from the digestive
226 system to the circulatory system and later to other organs (Neves et al., 2015). Likewise, in
227 other experiments, it has been shown that the exposure of reef fish to water sources that had
228 previously been exposed to polypropylene bags raises the levels of nonylphenol in the fish,
229 which led to their short and long-term death (Worm et al., 2017). Another consequence of the
230 presence of MP in the digestive tract of fish could include choking, histological damage and
231 alteration of the microbiome (Batel et al., 2018; Jin et al., 2018; Karami et al., 2017).

232 Plastic debris can also harbor pathogens that are often associated with disease
233 outbreaks, e.g., in coral reefs, since microbial communities can colonize microplastics. An
234 example of this is the bacteria of the genus *Vibrio* (Zettler et al., 2013), an opportunistic
235 pathogenic bacterium known to cause coral diseases worldwide (Lamb et al., 2018).

236

237 **Conclusions**

238 This study represents an emerging research field for Costa Rica as a response of the
239 efforts made in the region to characterize the presence of microplastics in marine life, and its
240 possible ecological implications. Although this is a small study, our results help to integrate
241 existing information on MP contamination in marine life. The validation of *Opistonema* sp. as
242 a model species in the MP biomonitoring will require more studies. It is also necessary to
243 replicate this type of studies systematically in different points of the Pacific coast and at
244 different times of the year, to better understand the effect of local and regional currents on the
245 dynamics of microplastic masses. In the Caribbean region, it is also necessary to carry out this
246 type of research to compare the state of the two Costa Rican coasts.

247 It is important to develop strategies that attempt to identify the possible physiological
248 effects of MP at different levels of marine life. In the case of our model species, it would be

249 necessary to investigate if there is any involvement at the histological level, at the metabolic
250 level or even in the microbiome (dysbiosis) due to the high presence of MP.

251

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253

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261

262 **Declarations of interest**

263 None

264

265

266 **References**

267

268 Batel, A., Borchert, F., Reinwald, H., Erdinger, L., Braunbeck, T., 2018. Microplastic
269 accumulation patterns and transfer of benzo[a]pyrene to adult zebrafish (*Danio rerio*)
270 gills and zebrafish embryos. *Environmental Pollution*, 235, 918–930.
271 <https://doi.org/10.1016/j.envpol.2018.01.028>

272 Boerger, C.M., Lattin, G.L., Moore, S.L., Moore, C.J., 2010. Plastic ingestion by planktivorous
273 fishes in the North Pacific Central Gyre. *Marine Pollution Bulletin* 60, 2275–2278.
274 <https://doi.org/10.1016/j.marpolbul.2010.08.007>

275 Carpenter, E.J., Smith, K.L., 1972. Plastics on the Sargasso sea surface. *Science* 175, 1240–
276 1241.

277 Cole, M., Lindeque, P., Halsband, C., Galloway, T.S., 2011. Microplastics as contaminants in
278 the marine environment: A review. *Marine Pollution Bulletin* 62, 2588–2597.
279 <https://doi.org/10.1016/j.marpolbul.2011.09.025>

280 Espinoza, P., Bertrand, A., 2008. Revisiting Peruvian anchovy (*Engraulis ringens*)
281 trophodynamics provides a new vision of the Humboldt Current system. *Progress in*
282 *Oceanography* 79, 215–227. <https://doi.org/10.1016/j.pocean.2008.10.022>

283 Gulmine, J.V., Janissek, P.R., Heise, H.M., Akcelrud, L., 2002. Polyethylene characterization
284 by FTIR. *Polymer Testing* 21, 557–563. [https://doi.org/10.1016/S0142-](https://doi.org/10.1016/S0142-9418(01)00124-6)
285 [9418\(01\)00124-6](https://doi.org/10.1016/S0142-9418(01)00124-6)

286 Güven, O., Gökdağ, K., Jovanović, B., Kıdeys, A.E., 2017. Microplastic litter composition of
287 the Turkish territorial waters of the Mediterranean Sea, and its occurrence in the
288 gastrointestinal tract of fish. *Environmental Pollution*, 223, 286–294.
289 <https://doi.org/10.1016/j.envpol.2017.01.025>

290 Ivar do Sul, J.A., Costa, M.F., 2014. The present and future of microplastic pollution in the
291 marine environment. *Environmental Pollution*, 185, 352–364.
292 <https://doi.org/10.1016/j.envpol.2013.10.036>

293 Jin, Y., Xia, J., Pan, Z., Yang, J., Wang, W., Fu, Z., 2018. Polystyrene microplastics induce
294 microbiota dysbiosis and inflammation in the gut of adult zebrafish. *Environmental*
295 *Pollution*, 235, 322–329. <https://doi.org/10.1016/j.envpol.2017.12.088>

296 Karami, A., Golieskardi, A., Ho, Y.B., Larat, V., Salamatinia, B., 2017. Microplastics in
297 eviscerated flesh and excised organs of dried fish. *Sci Rep* 7, 5473.
298 <https://doi.org/10.1038/s41598-017-05828-6>

- 299 Lamb, J.B., Willis, B.L., Fiorenza, E.A., Couch, C.S., Howard, R., Rader, D.N., True, J.D.,
300 Kelly, L.A., Ahmad, A., Jompa, J., Harvell, C.D., 2018. Plastic waste associated with
301 disease on coral reefs. *Science* 359, 460–462. <https://doi.org/10.1126/science.aar3320>
- 302 Law, K.L., 2017. Plastics in the Marine Environment. *Annu. Rev. Mar. Sci.* 9, 205–229.
303 <https://doi.org/10.1146/annurev-marine-010816-060409>
- 304 Law, K.L., Thompson, R.C., 2014. Microplastics in the seas. *Science* 345, 144–145.
305 <https://doi.org/10.1126/science.1254065>
- 306 Lozano Cabo, F. 1979. *Ictiología del Mar Menor (Murcia)*. Secretariado de Publicaciones,
307 Universidad de Murcia, Murcia.
- 308 Majewsky, M., Bitter, H., Eiche, E., Horn, H., 2016. Determination of microplastic
309 polyethylene (PE) and polypropylene (PP) in environmental samples using thermal
310 analysis (TGA-DSC). *Science of The Total Environment* 568, 507–511.
311 <https://doi.org/10.1016/j.scitotenv.2016.06.017>
- 312 Martin, J., Lusher, A., Thompson, R.C., Morley, A., 2017. The Deposition and Accumulation
313 of Microplastics in Marine Sediments and Bottom Water from the Irish Continental
314 Shelf. *Sci Rep* 7, 10772. <https://doi.org/10.1038/s41598-017-11079-2>
- 315 McNeish, R.E., Kim, L.H., Barrett, H.A., Mason, S.A., Kelly, J.J., Hoellein, T.J., 2018.
316 Microplastic in riverine fish is connected to species traits. *Sci Rep* 8, 11639.
317 <https://doi.org/10.1038/s41598-018-29980-9>
- 318 Moore, C.J., Moore, S.L., Leecaster, M.K., Weisberg, S.B., 2001. A comparison of plastic and
319 plankton in the North Pacific central gyre. *Mar. Pollut. Bull.* 42, 1297–1300.
- 320 Moore, C.J., Moore, S.L., Weisberg, S.B., Lattin, G.L., Zellers, A.F., 2002. A comparison of
321 neustonic plastic and zooplankton abundance in southern California’s coastal waters.
322 *Mar. Pollut. Bull.* 44, 1035–1038.
- 323 Neves, D., Sobral, P., Ferreira, J.L., Pereira, T., 2015. Ingestion of microplastics by commercial
324 fish off the Portuguese coast. *Marine Pollution Bulletin* 101, 119–126.
325 <https://doi.org/10.1016/j.marpolbul.2015.11.008>
- 326 Ory, N., Chagnon, C., Felix, F., Fernández, C., Ferreira, J.L., Gallardo, C., Garcés Ordóñez,
327 O., Henostroza, A., Laaz, E., Mizraji, R., Mojica, H., Murillo Haro, V., Ossa Medina,
328 L., Preciado, M., Sobral, P., Urbina, M.A., Thiel, M., 2018. Low prevalence of
329 microplastic contamination in planktivorous fish species from the southeast Pacific
330 Ocean. *Marine Pollution Bulletin* 127, 211–216.
331 <https://doi.org/10.1016/j.marpolbul.2017.12.016>

- 332 Ory, N.C., Sobral, P., Ferreira, J.L., Thiel, M., 2017. Amberstripe scad *Decapterus muroadsi*
333 (Carangidae) fish ingest blue microplastics resembling their copepod prey along the
334 coast of Rapa Nui (Easter Island) in the South Pacific subtropical gyre. *Science of The*
335 *Total Environment* 586, 430–437. <https://doi.org/10.1016/j.scitotenv.2017.01.175>
- 336 R Core Team. 2018. R: A language and environment for statistical computing. R Foundation
337 for Statistical Computing, Vienna, Austria.
- 338 Setälä, O., Fleming-Lehtinen, V., Lehtiniemi, M., 2014. Ingestion and transfer of microplastics
339 in the planktonic food web. *Environmental Pollution* 185, 77–83.
340 <https://doi.org/10.1016/j.envpol.2013.10.013>
- 341 Shiber, J.G., 1979. Plastic pellets on the coast of Lebanon. *Marine Pollution Bulletin* 10, 28–
342 30. [https://doi.org/10.1016/0025-326X\(79\)90321-7](https://doi.org/10.1016/0025-326X(79)90321-7)
- 343 Tanaka, K., Takada, H., 2016. Microplastic fragments and microbeads in digestive tracts of
344 planktivorous fish from urban coastal waters. *Sci Rep* 6, 34351.
345 <https://doi.org/10.1038/srep34351>
- 346 Vega-Corrales, L.A. 2010. Evaluación poblacional del stock explotable del complejo
347 *Opisthonema* (Pisces:Clupeidae) en el Golfo de Nicoya, Costa Rica. *Rev. Mar. Cost.*
348 2:83-94.
- 349 Wang, Z.-M., Wagner, J., Ghosal, S., Bedi, G., Wall, S., 2017. SEM/EDS and optical
350 microscopy analyses of microplastics in ocean trawl and fish guts. *Science of The Total*
351 *Environment* 603–604, 616–626. <https://doi.org/10.1016/j.scitotenv.2017.06.047>
- 352 Wickham, H., 2016. *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York.
- 353 Worm, B., Lotze, H.K., Jubinville, I., Wilcox, C., Jambeck, J., 2017. Plastic as a Persistent
354 Marine Pollutant. *Annu. Rev. Environ. Resour.* 42, 1–26.
355 <https://doi.org/10.1146/annurev-environ-102016-060700>
- 356 Zettler, E.R., Mincer, T.J., Amaral-Zettler, L.A., 2013. Life in the “Plastisphere”: Microbial
357 Communities on Plastic Marine Debris. *Environ. Sci. Technol.* 47, 7137–7146.
358 <https://doi.org/10.1021/es401288x>
- 359
- 360

361 **Figure legends**

362

363 **Figure 1.** The geographic location of the sampling site (in red) in the central Pacific coast of
364 Costa Rica.

365 **Figure 2.** Measurements of the main characteristics of the samples of *O. libertate* grouped by
366 sex.

367 **Figure 3.** Number of particles of microplastics identified in the gastrointestinal tract of *O.*
368 *libertate* and grouped by sex.

369 **Figure 4.** Micrographs of microplastics found in the digestive tract of *O. libertate*. The images
370 A, B, and C correspond to particles, and the images D, E, and F correspond to fibers.

371 **Figure 5.** SEM images of contents in the digestive tract of *O. libertate* and their % weight of
372 each element by Energy Dispersive X-ray Spectroscopy (EDX). The low magnification view
373 on the left shows different types of solids that can be categorized as (A) fibers, (B)
374 incrustations, and (C) standalone particles. Fiber in A showed a composition by weight % of
375 Carbon (60 ± 2) and Oxygen (40 ± 2). In B, EDX showed a variable composition; Aluminum
376 (4 ± 1), Calcium (10 ± 2), Carbon (35 ± 11), Potassium (4 ± 1), Oxygen (38 ± 7) and Silicon (9 ± 2).
377 Particle in C, showed the following composition: Calcium (57 ± 4), Magnesium (5 ± 1) and
378 Oxygen (38 ± 5).

379 **Figure 6. A)** -ATR spectrum for contents in the digestive tract of *O. libertate*. Range:500-4000
380 cm^{-1} . **B)** DSC graph: heat flow (mW) versus temperature ($^{\circ}\text{C}$) for *ca* 1 mg of contents in the
381 digestive tract of *O. libertate* under nitrogen atmosphere. Heating/cooling rate: $20^{\circ}\text{C min}^{-1}$.

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383 **Table 1.** Comparison of the percentage of fish that had microplastic and their respective
384 average in their digestive tract, according to the country (year) and species of study.

Country	Specie	Number of fishes	Percentage of fish with MP (%)	Average of MP per fish (\pm s.d.)	Total of MP
Costa Rica (2018)	<i>Opisthonema libertate</i>	30	100	36.7(\pm 0.86)	1101
Easter Island (2017)	<i>Decapterus muroadsi</i>	20	80	2.5(\pm 0.4)	48
Japan (2015)	<i>Engraulis japonicas</i>	64	77	2.3(\pm 2.5)	150
Colombia (2016)	<i>Opisthonema libertate</i>	27	0	0(\pm 0)	0
Panama (2018)	<i>Cetengraulis mysticetus</i>	10	0	0(\pm 0)	0
Peru (2018)	<i>Engraulis ringens</i>	40	0	0(\pm 0)	0

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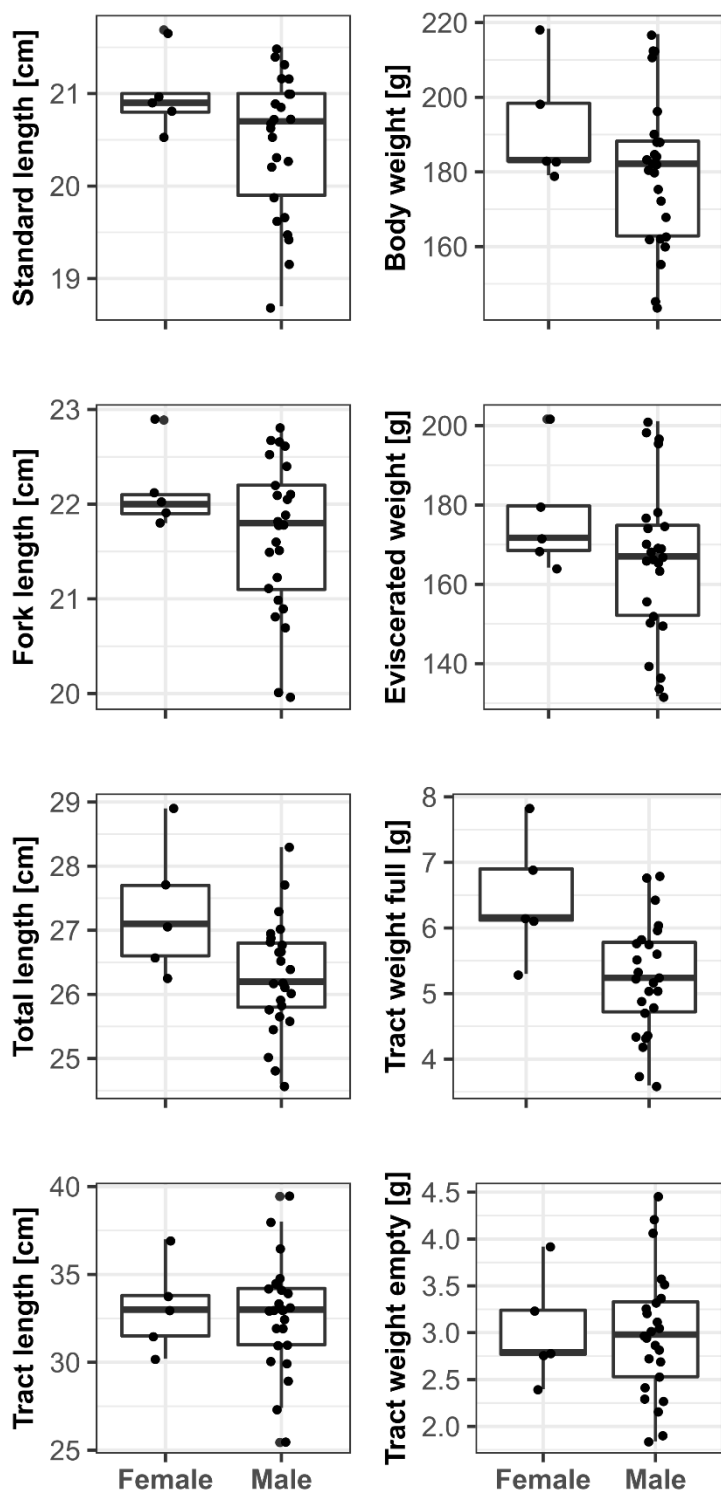


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Figure 1



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Figure 2

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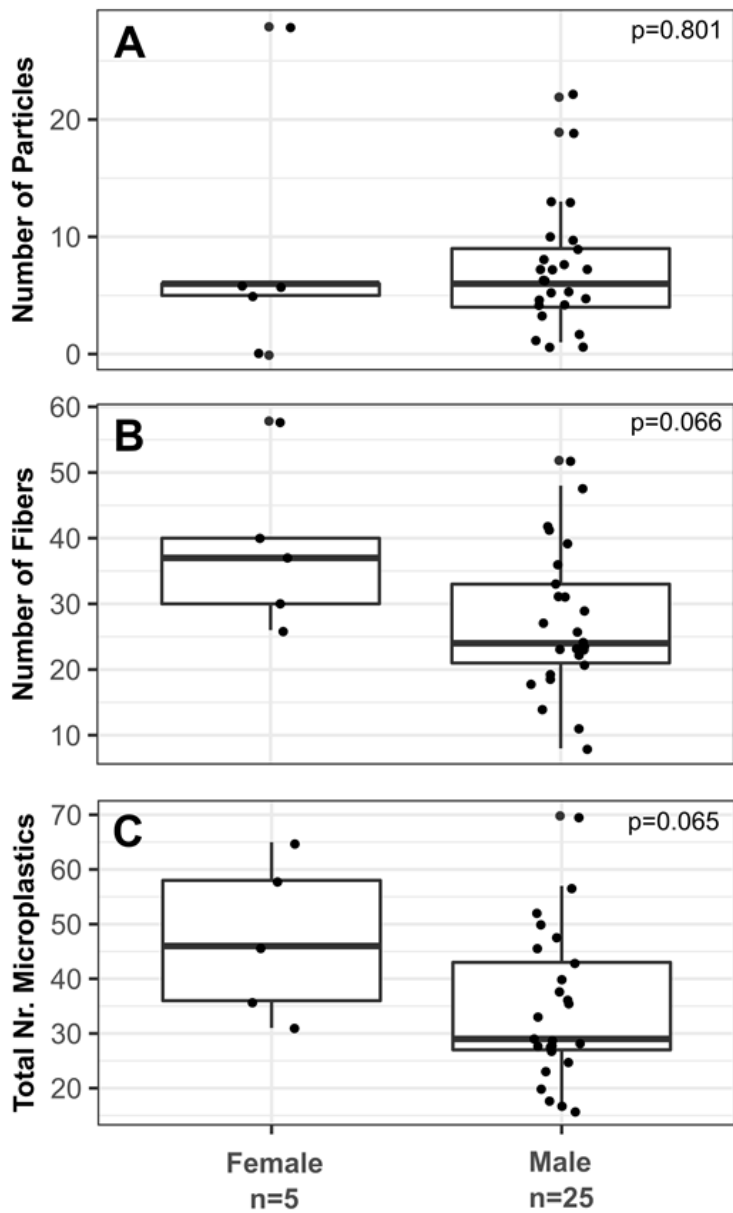
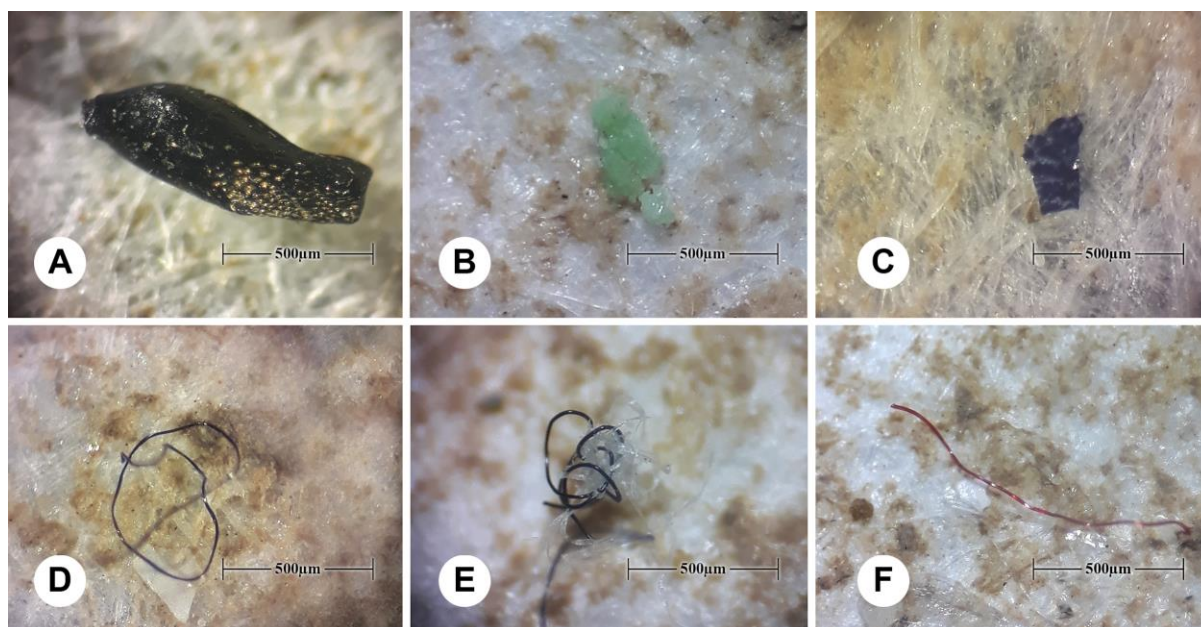


Figure 3



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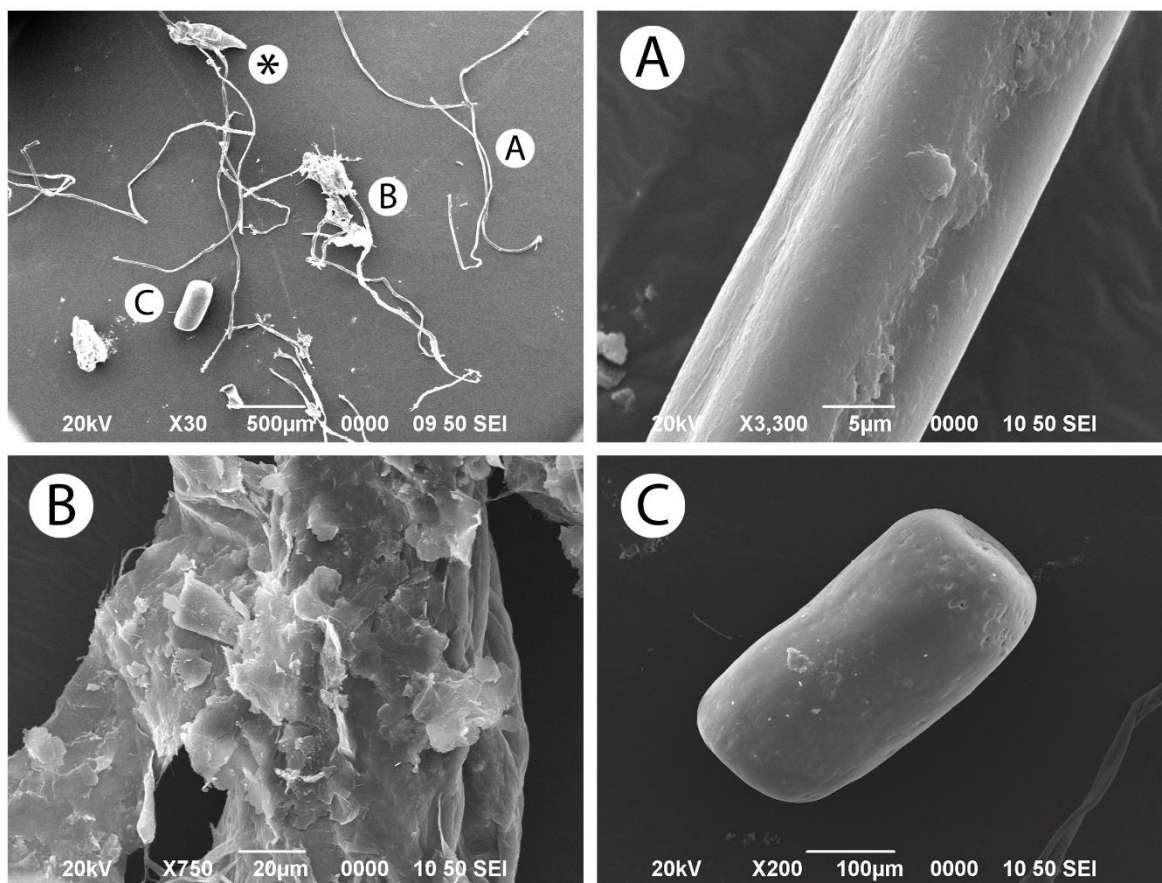
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Figure 4

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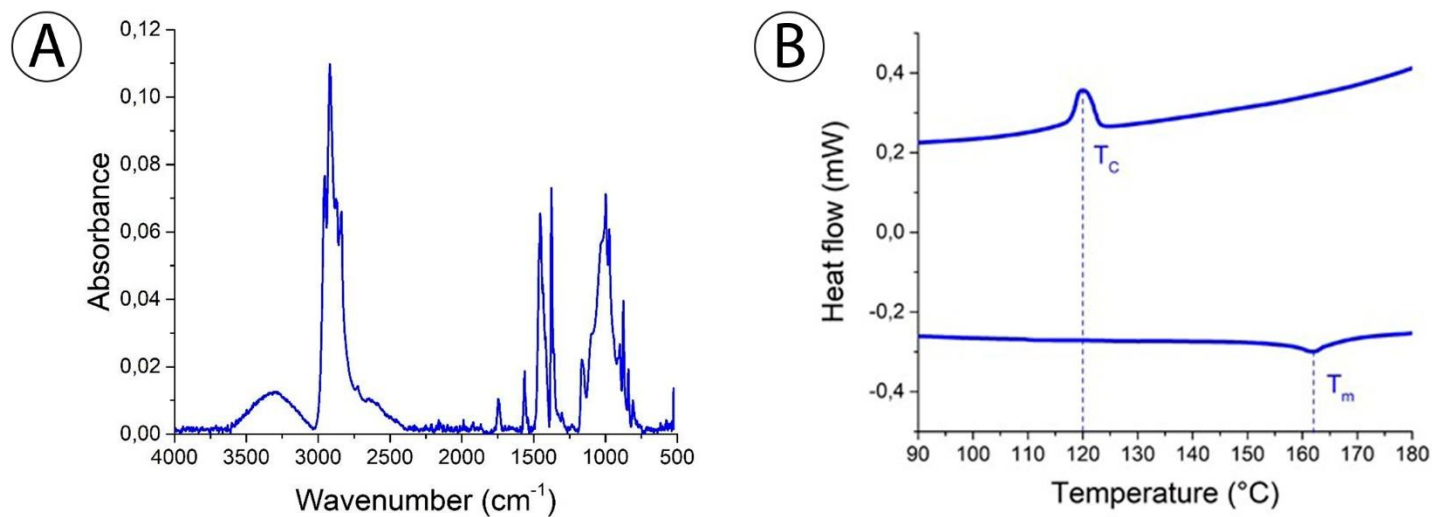


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Figure 5

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Figure 6

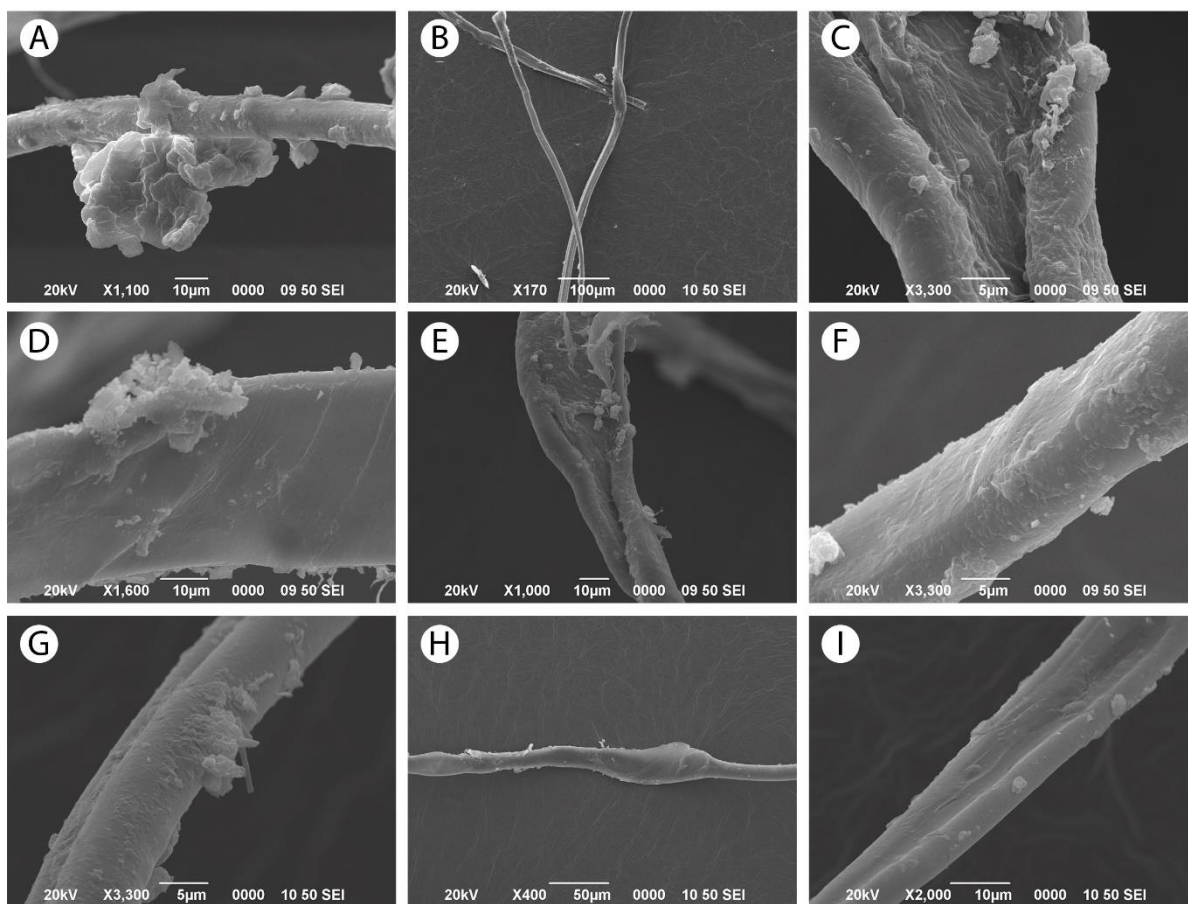
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Supplementary information

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449 **Supplementary Figure 1**

450 SEM images of fibers found in the digestive tract of *O. libertate*. The presence of incrustations
451 in the fibers was evident at ~3000X. It was possible to confirm the mineraloid nature of the
452 incrustations based on the composition obtained by Energy Dispersive X-ray Spectroscopy
453 (EDX). Incrustations are more evident in A, C, D, G.

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