Non-additive effects of changing the cytochrome P450 ensemble: incorporation of CYP2E1 into human liver microsomes and its impact on CYP1A2.

Nadezhda Y. Davydova[1], Bikash Dangi[1], Marc A. Maldonado[1], Nikita E. Vavvlov[2], Victor G. Zgoda[2] and Dmitri R. Davydov[1]

[1] Department of Chemistry, Washington State University, Pullman, WA, 99164, and [2]Institute of Biomedical Chemistry, Moscow, 119121, Russia

Abstract

The aim of this study is to investigate the ability of ethanol-inducible CYP2E1 to interact with other cytochrome P450 species and affect the metabolism of their substrates. As a model system we used CYP2E1-enriched microsomes obtained by incorporation of purified CYP2E1 protein into HLM. Using the method based on homo-FRET in homo-oligomers of CYP2E1 labeled with BODIPY 577/618 maleimide we demonstrated that the interactions of CYP2E1 with microsomes result in dissociation of the protein homo-oligomers. The finding that this effect is much better pronounced in HLM as compared to the microsomes containing no P450 proteins indicates the formation of mixed oligomers of CYP2E1 with other P450 species that takes place in expense of dissociation of the homo-oligomers.

Incorporation of CYP2E1 into HLM results in a multifold increase in the rate of metabolism of CYP2E1-specific substrates *p*-Nitrophenol (pNP) and Chlorzaxozone (CLZ). The rate of their oxidation remains proportional to the amount of incorporated CYP2E1 up to the content of 0.3-0.4 nmol/mg protein (or about 50% CYP2E1 in the P450 pool). These results demonstrates that the incorporated CYP2E1 becomes a fully-functional member of the P450 ensemble and do not exhibit any detectable functional differences with the endogenous CYP2E1 in HLM.

Enrichment of HLM with CYP2E1 results in a pronounced alteration of the metabolism of 7-etoxy-4-cyanocoumarin (CEC), the substrate of CYP2C19 and CYP1A2, that suggests an important augmentation of the involvement of CYP1A2 in its metabolism. This effect goes together with a remarkable increase in the rate of dealkylation of CYP1A2-specific substrate 7-ethoxyresorufin by CYP2E1-enriched HLM. Furthermore, probing the interactions of CYP2E1 with model microsomes (Supersomes™) containing individual P450 enzymes we found that CYP2E1 efficiently interacts with CYP1A2, but lacks any ability to form complexes with CYP2C19. This finding goes inline with CYP2E1-induced redirection of the main route of CEC metabolism from CYP2C19 to CYP1A2.

Keywords: cytochrome P450; protein-protein interaction; oligomerization; membrane protein; alcohol-drug interactions, drug metabolism

Corresponding author: Dmitri R. Davydov, Department of Chemistry, Washington State University, MS 4630, Pullman, WA 99164-4630; Tel. +1 509 335-5983; E-mail: dmrdavyd@gmail.com

The functional properties of the drug-metabolizing system of human liver are largely determined by the properties of the ensemble of cytochromes P450, which is responsible for metabolism of over 75% of all marketed drugs and new drug candidates [1-3]. It is commonly recognized that the inter-individual, age-dependent and temporal variations in drug metabolism are largely dictated by the changes in the composition of the ensemble of multiple drug-metabolizing cytochrome P450 species. However. recent studies on a large selection of human liver microsomes (HLM) demonstrated the lack of a straight correlation between the composition of human cytochrome P450 ensemble and the profile of drug metabolism [4]. At the same time, there is an emerging recognition of the functional importance of physical interactions between multiple P450 species co-localized in the microsomal membrane [5-9]. Due to complex effects of these interactions any change in the content of a particular P450 enzyme may affect drug metabolism in a complex, hard-to-predict manner. Therefore, a systematic exploration of the network of P450-P450 interactions and identification of their functional consequences represents an issue of emerging importance.

Of particular practical significance are the functional effects of the changes in the composition of P450 ensemble induced by alcohol consumption. Although the multi-fold increase in the content of cytochrome P450 2E1 (CYP2E1) in liver observed in both alcoholics and moderate alcohol consumers is well documented [10-14], the role of CYP2E1 in the instances of alcohol-drug interactions is commonly considered insignificant due to a minor role of this enzyme in drug metabolism. However, the involvement of CYP2E1 in alcohol-drug interactions may stretch far beyond its immediate effects on the metabolism of its substrates.

There is a limited amount of information on possible functional effects of CYP2E1 on other human P450 species. In an early study with model microsomes Tan and co-authors demonstrated mutual inhibitory effects of CYP2E1 and CYP2A6, which was attributed to a competition for NADPH-cytochrome P450 reductase [15]. In the same study these authors also observed a potentially important effect of co-expression of CYP2E1 with CYP2A6 on H_2O_2 production by CYP2A6. While NADPH-dependent generation of H_2O_2 by CYP2A6 was drastically increased in the presence of coumarin, addition of this CYP2A6 substrate results in decreased H_2O_2 production by the system composed of CYP2A6, CYP2E1 and CPR [15]. In a latter study in a micellar reconstituted system Kelley and co-authors explored the reciprocal effects of rabbit CYP1A2 and CYP2E1 enzymes [16]. The authors demonstrate that, while the addition of CYP1A2 does not affect CYP2E1-dependent metabolism of N-nitrosodimethylamine, addition of CYP2E1 considerably intensifies CYP1A2-dependent dealkylation of 7-ethocyresorufin (ERR). Based on the observation that this

effect was most pronounced at sub-saturating concentrations of CPR the authors concluded that the stimulating effect of CYP2E1 on CYP1A2 may be exerted through the formation of the CYP1A2-CYP2E1 complexes where the interactions of CYP1A2 with CPR are promoted [16].

In our study of interactions between CYP2E1 and CYP3A enzymes in model microsomes we demonstrated the formation of the complexes of CYP2A1 with either CYP3A4 or CYP3A5, where the activity of CYP2E1 is boosted, while the functionality of the CYP3A enzymes remains unaffected [17]. In our recent study we also explored the interactions between CYP2E1 with CYP2D6 and observed the formation of their heteromeric complexes in microsomal membrane. We demonstrated that these interactions inhibit the activity of CYP2E1, while exerting a time-dependent activating effect on CYP2D6 in the presence of its specific substrates, 3-[2-(N,N-diethyl-N-methyl ammonium)ethyl]-7-methoxy-4-methylcoumarin (AMMC) and bufuralol [9]. Of particular importance might be the effect of CYP2E1 on the stoichiometry of futile cycling and substrate oxidation by CYP2D6 revealed in a decrease in the electron leakage from CYP2D6 through the peroxide-generating pathways [9].

All these results demonstrate complex functional relationships between CYP2E1 and other drug-metabolizing P450 species which deserves further attention in view of its potential involvement in alcohol-drug interactions. Of particular interest is possible crosstalk between CYP2E1 and CYP1A2 evidenced in the study of Kelley and co-authors [16]. The effect of ethanol-induced increase in CYP2E1 concentration in liver on CYP1A2 may be involved in pronounced interactions of alcohol with such drug substrates of CYP1A2 as tricyclic antidepressnts (amitryptiline, clomipramine, imipramine, etc.) [18-19]. Most of the drugs of this type are also metabolized by CYP2C19 and possible rerouting of drug metabolism between the two enzymes may also play a role in their interactions with alcohol.

The aim of the present study is to probe the interactions of CYP2E1 with cytochromes P450 present in HLM and explore the effect of these interactions on the functional properties of CYP1A2 and CYP2C19. In a search for a model system suitable for these studies we recently introduced a method based on incorporation of purified cytochromes P450 into microsomal membrane [9, 17, 20]. An important characteristic of this strategy is its adaptability for incorporating genetically or chemically modified proteins. This includes the incorporation of proteins bearing fluorescent probes and light-activating crosslinkers. Therefore, this strategy may be used for monitoring protein-protein interactions in HLM with various biophysical techniques, as well as for identifying the P450 interactions partners with the technique of molecular fishing. Results of the present study demonstrate high utility of this approach in the studies of interactions between multiple

cytochrome P450 species and their effect on drug metabolism. Using this strategy we demonstrated the formation of the complexes of CYP2E1 with other P450 species and demonstrated the effects of CYP2E1 on the activity of CYP1A2, the enzyme involved in metabolism of a wide variety of commonly used drugs known for their pharmacokinetic interactions with alcohol.

Materials and Methods

Materials. *p*-Nitrophenol, p-Nitroanizol, and DL-dithiothreitol (DTT) were the products of Sigma-Aldrich (St. Louis, MO). NADPH tetrasodium salt was from EMD Millipore (Billerica, MA). 7-ethoxyresorufin (ERR) was from AnaSpec (San Jose, CA, USA). 7-etoxy-4-cyanocoumarin (CEC), 7-hydroxy-4-cyanocoumarin (CHC) and BODIPY 577/618 maleimide were from Invitrogen/Molecular Probes Inc. (Eugene, CA).. Chlorzoxazone, 6-hydroxy chlorzoxazone and resorufin were the products of Cayman Chemical (Ann Arbor, MI). All other reagents were of ACS grade and used without additional purification.

Protein expression and purification. N-terminally truncated (Δ3-20) and C-terminally His-tagged CYP2E1 [21] was expressed in *E. coli* and purified following the procedure similar to that described for CYP2B6 [22] followed by ion-exchange chromatography on Macro-Prep CM Support resin (Bio-Rad Laboratories, Hercules, CA).

Microsomes from insect cells bearing recombinant human cytochromes P450 (SupersomesTM) were the products of BD Gentest, now a part of Corning Life Sciences (Tewksbury, MA). In the present study we used the preparations containing CYP2E1 (lot 44748), CYP1A2 (lot 69375), CYP2B6 (lot 31487), CYP2C8 (lot 8239002), CYP2C9 (lot 41274), CYP2C19 (lot 73445), CYP2D6 (lot 7114001) and CYP3A4 (lot 35933). All those preparation contained human NADPH-cytochrome P450 reductase (CPR) and cytochrome b5, except for SS2C8 and SS1A2, which had no cytochrome b_5 in their content. In the experiments on CYP2E1 interactions with microsomal membrane we also utilized Supersomes™ containing rat CPR (SS-RR, lot 5274006) or human CPR and cytochrome b5 (SS-B5, lot 41774) as control membranes bearing no cytochromes P450.

Preparations of human liver microsomes (HLM). In this study we used two different preparations of pulled human liver microsomes. The preparation refered here as HLM-1 was prepared by differential centrifugation from the sample of human liver S9 pool (lot number 3212595) obtained from BD Gentest, now a part of Corning Life Sciences (Tewksbury, MA). The HLM sample refered here as HLM-2 is an InVitroCYP™ M-class 50-donor mixed gender pooled HLM preparation (lot LFJ) obtained from BioIVT corporation (Baltimore, MD).

Both samples were characterized with measuring the total content of cytochromes P450 and the content of cytochrome b_5 with UV-Vis spectroscopic assays. We determined the content of CYP2E1, CYP1A2, CYP3A4 and NADPH-cytochrome P450 reductase in both samples with mass-spectrometry, as described below. The concentration of NADPH-cytochrome reductase (CPR) was also estimated on the basis of the rate of NADPH-dependent reduction of cytochrome *c*. Notably, the results of the two different methods of CPR quantification gave nearly identical results with both preparations of HLM.

Results of our analysis are summarized in Table 1. As seen from this table the two samples were significantly different in the composition of the P450 pool, as well as in the P450-to-phospholipids and P450-to-reductase molar ratios. HLM-2 preparation is characterized with higher P450 concentration (as judged from the phospholipids:P450 molar ratio, $R_{L/P}$) and has a higher molar excess of P450 over the CPR. At the same time HLM-1 has higher content of both CYP1A2 and CYP2E1, while it is characterized with considerably lower abundance of CYP3A4 (Table 1).

Incorporation of CYP2E1 into HLM was performed by incubation of undiluted suspensions of HLM (20-25 mg/ml protein, 10-13 mM phospholipid) in 125 mM K-Phosphate buffer containing 0.25M Sucrose with purified CYP2E1 for 16 - 20 hours at 4°C at continuous stirring. CYP2E1 was added in the amount ranging from 0.25 to 2 molar equivalents to the endogenous cytochrome P450 present in HLM. After incubation the suspension was diluted 4-8 times with 125 mM K-Phosphate buffer, pH 7.4 containing 0.25 M sucrose and centrifuged at 53,000 rpm (150,000 g) in an Optima TLX ultracentrifuge (Beckman Coulter Inc., Brea, CA, USA) with a TLA100.3 rotor for 90 min at 4 °C. The pellet was resuspended in the same buffer to the protein concentration of 15-20 mg/ml. The amount of incorporated cytochrome P450 was calculated from the difference between the heme protein added to the incubation media and the enzyme found in the supernatant. According to these assay the procedure resulted in incorporation of 96 -98% of the added protein.

Determinations of protein and phospholipid concentrations in microsomal suspensions were performed with the bicinchoninic acid procedure [23] and through determination of total phosphorus in a chloroform/methanol extract according to Bartlett [24] respectively.

The concentration of NADPH-cytochrome P450 reductase in microsomal and proteoliposomal membranes was determined based on the rate of NADPH-dependent reduction of cytochrome *c* at 25ºC [25] monitored by an increase in the difference in absorbencies at 550 and 541 nm and calculated using the extinction coefficient of 18 $mM^{-1}cm^{-1}$ [26]. Effective molar concentration of CPR was estimated using the turnover number of 3750 min^{-1}, which we determined with purified CPR quantified from its absorbance at 456 nm using the extinction coefficient of 21.4 mM^{-1} cm^{-1} [27].

The total concentration of cytochrome P450 in HLM was determined with a variant of "oxidized CO versus reduced CO difference spectrum" method [28]. In our assay we used Microsome Solubilization Buffer (MSB) composed of 100mM potassium phosphate pH 7.3, 10% Glycerol, 0.5% Sodium Cholate and 0.4% Igepal CO-630. Prior to addition of the microsomal suspension the buffer was saturated with CO through bubbling the gas for 40-60 sec. 100 µl of CO-saturated MSB were placed into a quartz microcuvette. After recording a baseline in 380-600 nm region the cell was supplemented with 5µl of HLM sample (10-20 mg/ml) and the spectrum was recorded. Reduction was achieved by addition of a small amount of dithionite dust. The process of reduction was followed through monitoring the changes in the difference of optical densities (DOD) at 450 and 490 nm. The spectrum of absorbance of the reduced sample was taken when the DOD stabilizes. The difference between the spectra taken after and before reduction was calculated and used to determine the concentration of cytochromes P450 using the extinction coefficient of 106 mM^{-1} cm^{-1} for DOD between the maximum of the peak around 450 nm (448-451 nm) and the plateau at 490 nm.

Determination of the content of cytochrome b5 in HLM was based on its NADH-dependent reduction and performed following the procedure described by Kennedy [29].

Mass-spectrometric analysis of the content of cytochromes P450 and NADPH-cytochrome P450 reductase in HLM was performed with a triple quadrupole mass spectrometer using the method of multiple reaction monitoring (MRM). For each protein, one standard peptide with three transitions was used. The peptides selected were arranged into one SRM assay. The information (m/z of precursors, m/z of transition ions, CE values, b, y ions transition ions and MS platform were used for the analysis) on SIS and the target peptides is provided in Table S1 found in Supplementary Material. Separation of peptides from digested HLM samples was carried out using the UPLC Agilent 1290 system including a pump and an autosampler. The sample was loaded into the analytical column Eclipse Plus SBC-18, 2.1 x 100 mm, 1.8 um, 100 A. Peptide elution was performed by applying a mixture of solvents A and B. Solvent A was HPLC grade water with 0.1% (v/v) formic acid, and solvent B was 80%(v/v) HPLC grade acetonitrile/water with 0.1% (v/v) formic acid. The separations were performed by applying a linear gradient from 3% to 32% solvent B over 50 min, then from 32% to 53% solvent B over 3 min at 300µl/min followed by a washing step (5 min at 90% solvent B) and an equilibration step (5 min at 3% solvent B). Ten microliters of each sample were applied on chromatographic column. The quantitative analysis was performed using Agilent 6495 Triple Quadrupole (Agilent, USA) equipped with a Jet Stream ionization source. The following parameters were used for the Agilent Jet Stream ionization source: the temperature of the drying

gas of 280°C, 18 psi pressure in the nebulizer, 14 L/min flow rate of the drying gas, and 3000V voltage on the capillary.

The standard samples of target peptides were obtained using the solid-phase peptide synthesis on the Overture (Protein Technologies, USA) according to the published method [30]. The isotope-labeled lysine (13C6,15N2) or arginine (13C6,15N4) was used for isotope-labeled peptide synthesis instead of the unlabeled lysine or arginine, correspondingly. Concentration of the synthesized peptides was quantified through acidic hydrolysis followed by analysis of derived amino acids with fluorimetric detection [31].

Activity measurements with fluorogenic substrates The rate of *O*-dealkylation of CEC and ERR were measured with a real-time continuous fluorometric assay. A suspension of microsomes was added to 250 µl of 0.1 M Na-HEPES buffer, pH 7.4,containing 60 mM KCl, to a final CPR concentration of 0.003 – 0.01 µM. The mixture was placed into a 5 x 5 mm quartz cell with continuous stirring and thermostated at 30 °C. An aliquot of a 15-20 mM stock solution of CEC or 0.4-2 mM stock solution of ERR in acetone was added to attain the desired concentration in the range of 0.5 - 250 µM (CEC) or 0.03 - 10 µM (ERR). The reaction was initiated by addition of 20mM solution of NADPH to the concentration of 200 µM. Increase in the concentration of products of dealkylation (CHC or resorufin) was monitored with a Cary Eclipse fluorometer (Agilent Technologies, Santa Clara, CA, USA) or a custom-modified PTI QM-1 fluorometer (Photon Technology International, New Brunswick, NJ) [9]. With both instruments the emission wavelength was set to 455 and 585 nm for the measurements with CEC and ERR, respectively. The excitation was performed with a monochromatic light centered at 405 or 560 nm (for CEC and ERR, respectively) with 5 nm bandwidth (Cary Eclipse) or with a light emitted by a 405 nm (CEC) or 532 nm (ERR) laser diode module (Thorlabs Inc, Newton, NJ. The rate of formation of the fluorescent products was estimated by determining the slope of the linear part of the kinetic curve recorded over a period of 3 – 5 min. Calibration of the assay was performed by measuring the intensity of fluorescence in a series of 5-10 samples of the same reaction mixture containing increasing concentrations of CHC or resorufin.

Measurements of the rate of hydroxylation of p-nitrophenol was preformed with absorbance spectroscopy [32]. To measure the rate of formation of *p*-nitrocatechol (pNC) by HLM we mixed 3-5µl of the microsomal suspension with reaction buffer (0.1 M Na-Hepes buffer containing 60mM KCl) and a desired amount of 10-20 mM stock solution of pNP in reaction buffer to the total volume of 80 µl. After a preincubation for 1 min in a water bath at 30 °C the reaction was started by addition of 2µl of 20 mM NADPH solution. The probes were incubated at 30 °C for 30 min in a

shaking water bath and the reaction was stopped by addition of 24 µl of 20% Trichloroacetic acid. The precipitated protein was removed by centrifugation at 15,000g for 2 min and the supernatant was supplemented with 16 µl of 5M NaOH. The spectra of absorbance of the samples in 450 – 700 nm region were recorded in a 100µl microcuvette (optical path 1 cm). As a baseline we used a spectrum of the sample treated as above and containing all ingredients except for pNP. To quantify the amount of the formed pNC and obtain the dependencies of the reaction rate on substrate concentration we subjected the series of spectra obtained with pNP concentrations increasing from 0 to 250 µM to the principal component analysis procedure. The spectra of the first two principal components were approximated with a combination of the standard spectra of absorbance of pNP and pNC supplemented with a first or second order polynomial, which was used to compensate for possible variation in turbidity. The maximum of absorbance of pNC was found at 514 nm and the value of the extinction coefficient of 10.3 mM^{-1} cm^{-1} at 546 nm [32] was used to normalize the spectral standard to 1µM concentration. The concentration factors found in this fitting were used to scale the first two vectors of loading factors and combine them into a dependence of the concentration of formed pNC on the concentration of pNP.

Activity measurements with chlorzoxazone were performed in 0.1 M Na-HEPES buffer, pH 7.4, containing 60mM KCl. The reaction was started by addition of NADPH to the final concentration of 0.5 mM and carried on for 10 min at 30 °C. Chlorzoxazone was added as a 25 mM solution in 60 mM KOH. Reaction was quenched with an addition of 25% of the incubation volume of 1 M solution of formic acid in acetonitrile containing a known concentration 3,5-dibromo-4-hydroxybenzoic acid as internal standard, followed by centrifugation at 9,300 g for 10 min.

The amount of formed 6-hydroxychlorzoxazone was quantified by liquid chromatography-mass spectrometry. An LC-20AD series high-performance liquid chromatography system (Shimadzu, Columbia, MD) fitted with a HTC PAL autosampler (LEAP Technologies, Carrboro, NC) was used to perform chromatography on a Kinetex® reverse-phase column (50 × 2.1 mm, 2.6 µm; Phenomenex, Torrance, CA). Chromatographic separation was performed using an isocratic method at the flow rate of 0.18 ml/min (mobile phase A) and 0.22 ml/min (mobile phase B) for 5 min. The quantification of the metabolite was conducted using an API 4000 Q-Trap tandem mass spectrometry system manufactured by Applied Biosystems/MDS Sciex (Foster City, CA) using turbospray ESI operating in negative ion mode. The mass spectrometer parameters were set at curtain gas, 20; collision gas, medium; ion spray voltage, -4500; ion source gas 1, 70; ion source gas 2, 50; temperature, 500, declustering potential, -550; entrance potential, -5; collision energy, -25, collision cell exist potential, 10. The analyte (6-hydroxychlorzoxazone) and the internal standard

(3,5-dibromo-4-hydroxybenzoic) were detected using multiple reaction monitoring (MRM) mode by monitoring the *m/z* transition from 184.0 to 120 and 294.8 to 251.0, respectively. Determination of the product amounts was achieved with a calibration curve ranging from 0 to 2000 nM concentration of 6-hydroxychlorzoxazone.

Normalization of the rates of substrate metabolism determined in HLM samples was performed based on the concentration of CPR determined form the rate of NADPH-dependent reduction of cytochrome *c* as described above. The rates of substrate metabolism by Supersomes™ were normalized on the concentration of cytochrome P450 (due to excess of CPR over cytochrome P450 in these preparations).

Monitoring the interactions of CYP2E1-BODIPY with microsomal membranes was performed with the use of a Cary Eclipse spectrofluorometer equipped with a Peltier 4-position cell holder. The excitation of donor phosphorescence was performed with monochromatic light centered at 505 nm with 20 nm bandwidth. Alternatively the measurements were done with the use a custom-modified PTI QM-1 fluorometer (Photon Technology International, New Brunswick, NJ) equipped with a thermostated cell holder and a refrigerated circulating bath. In this case the excitation at 505 nm was performed with an M505L3 LED module (Thorlabs Inc, Newton, NJ). The spectra in the 570 – 800 nm wavelength region were recorded repetitively with the time interval varying from 0.5 to 15 min during the course of monitoring (5 – 16 hours). The experiments were performed at continuous stirring at 4 °C in 100 mM Na-Hepes buffer containing 150 mM KCl and 250 mM sucrose.

Calculation of the surface density of cytochromes P450 in microsomal membrane was based on the molar ratio of microsomal phospholipids to cytochromes P450 ($R_{L/P}$). In these calculations we used the value of 0.95 nm² for the area of microsomal membrane corresponding to one molecule of phospholipid in a monolayer [33], similar to the approach used in our earlier reports [9, 17, 20].

Data analysis. Analysis of series of spectra obtained in fluorescence spectroscopy experiments was done by principal component analysis (PCA) [34], which was used as a method of global analysis that minimizes the contribution of incidental variation in the shape of the spectra and makes the results more robust to a minor displacement of the emission band (a blue shift of approx. 2 nm) that takes place upon the interactions of CYP2E1-BODIPY with the membrane. To quantify the changes in the fluorescence of BODIPY we used a linear least-squares fitting of the spectra of the first and second principal components by a combination of the standard spectra of emission of CYP2E1-BODIPY in solution and in a membrane-bound state.

The equation for the equilibrium of binary association (dimerization) used in the fitting of oligomerization isotherms (dependencies of FRET efficiency on the concentration of P450 in membranes) had the following form:

$$[X]=\tfrac{1}{2}\cdot K_D+[E]_0-\sqrt{\tfrac{1}{4}\cdot K_D^{\,2}+[E]_0\cdot K_D} \qquad \text{(Eq. 1)}$$

where $[E]_0$, $[X]$, and K_D are the total concentration of the associating compound (enzyme), the concentration of its dimers, and the apparent dissociation constant, respectively . In order to use this equation in fitting of the dependencies of the relative increase in fluorescence observed at enzyme concentration $[E]_0$ (R_E) equation (1) was complemented with the parameter R_{max} . This parameter corresponds to the value of R_E observed upon a thorough dissociation of completely oligomerized enzyme:

$$R_E = R_{max}\cdot([X]/[E]_0)$$

The parameter R_{max} is determined by the efficiency of FRET (E_{FRET}) according to the following relationship:

$$E_{FRET}= A_{max}/(1+A_{max})$$

Fitting of the titration isotherms to the above equations was performed with non-linear regression using a combination of Nelder-Mead and Marquardt algorithms as implemented in our SpectraLab software [34].

Results and Discussion

Incorporation of CYP2E1 into HLM and its effect on homo-oligomerization of the protein

In our previous studies we demonstrated that incubation of microsomal membranes, either the microsomes from insect cells (Supersomes™ or Baculosomes®) or HLM, results in efficient incorporation of purified cytochromes P450 into the membrane [9, 17, 20]. In order to further refine and better characterize this model system we developed a new technique for monitoring the process of CYP2E1 incorporation into the membrane. Our method utilizes fluorescence resonance energy transfer (homo-FRET) between fluorophores attached to neighboring CYP2E1 molecules in the oligomer. Short Stockes shift characteristic to fluorescent probes based on BODIPY moiety results in an efficient FRET between closely located fluorophores of the same kind (homo-FRET). This prompted us to use a BODIPY fluorophore in our study. For our experiments we chose BODIPY-577/618 maleimide due to a minimal spectral overlap of its emission band (λ_{max}=618 nm) with the absorbance bands of the P450 heme. With this florescent probe the effect of homo-FRET

between two BODIPY moieties (R_0=53 A) is expected to predominate over the effect of FRET from BODIPY to the P450 heme groups (R_0=40 A).

Labeling of purified CYP2E1 with BODIPY 577/618 maleimide taken in the 1:1 – 1:2 molar ratio to the protein causes neither conversion formation of the inactivated P420 state of the protein nor any detectable change in the P450 spin equilibrium. The FRET experiments reported here were performed with CYP2E1-BODIPY conjugates bearing two BODIPY fluorophores per one protein molecule.

The intensity of fluorescence of BODIPY 577/618 incorporated into CYP2E1 is heavily quenched in the oligomers in solution due to homo-FRET. Incorporation of the protein into the microsomal membrane is expected to cause an ample increase in fluorescence due to dissociation of the protein homo-oligomers and formation of complexes with other P450 species (where homo-FRET is not observed). Studying the dependence of the amplitude of this effect on the amount of the incorporated protein allows to determine the apparent dissociation constant of the homo-oligomers. Furthermore, comparison of the results obtained in HLM with those derived from the studies with model microsomes (Supersomes™) containing no P450 proteins or bearing a single recombinant P450 enzyme may be used to probe the interactions of CYP2E1 with other P450 species.

A series of spectra of fluorescence taken in the process of interactions of CYP2E1-BODIPY with HLM shown in Fig 1a demonstrate a multifold increase in the intensity of emission in full compliance with the expectations. Inset in Fig. 1a shows a series of kinetic curves of this increase recorded at various molar ratios of phospholipids of added HLM to CYP2E1-BODIPY. These time dependencies may be approximated with a three-exponential equation, where the characteristic times of the three phases amounts to 3+-0.5, 47+-20 and 600+-120 min. The maximal amplitude of the increase in fluorescence estimated from this approximation increases together with a decrease in the concentration of CYP2E1 in the membrane (or with an increase in the lipid-to-CYP2E1 ratio, $R_{L/P}$). This observation suggests that CYP2E1 incorporation into HLM results in dissociation of the homo-oligomers present in solution and the degree of this dissociation increases with decreasing the concentration of CYP2E1 in the membrane.

Similar effect was also observed during the incorporation of CYP2E1-BODIPY into the membrane of control Supersomes™ (SS-RR) containing recombinant CPR, but having no cytochromes P450 in their membranes (Fig 1B). In this case the dependence of the amplitude on $R_{L/P}$. was considerably more sharp than that observed with HLM. This difference reveals the effect of interactions of CYP2E1 with other P450 proteins present in HLM.

The dependencies of the amplitude of fluorescence increase on the surface density of CYP2E1-BODIPY in the membrane are shown in Fig. 2. Similar to the approach used in our previous studies [9, 17, 20] we determined the value of the apparent dissociation constant of the homo-oligomers (K_D) from the fitting of these dependencies to the equation for the equilibrium of binary association (dimerization) (Eq. 1), as described in Materials: and Methods.

The parameters of CYP2E1 interactions with HLM and SS-RR determined from the dependencies shown in Fig. 2 may be found in Table 2. This table also contains the parameters obtained in the studies of interactions of CYP2E1-BODIPY with CYP1A2 and CYP2C19 (see infra). As seen from this table the value of K_D for CYP2E1 homo-oligomers is much lower than that observed with the control microsomes. Therefore, the effect of CYP2E1 incorporation on its homo-oligomerization is much better pronounced in HLM as compared to the microsomes containing no P450 proteins. This observation evidences the formation of mixed complexes of CYP2E1 with other P450 species that takes place in the expense of dissociation of CYP2E1 homo-oligomers.

Effect of enriching HLM with CYP2E1 on the metabolism of CYP2E1-specific substrates.

In order to probe if the incorporation of CYP2E1 into HLM makes the exogenous enzyme a fully functional member of the P450 ensemble we probed the effect of enriching HLM with CYP2E1 on the metabolism of CYP2E1-specific substrates, *p*-Nitrophenol (pNP) and Chlorzoxazone (CLZ). In our studies with pNP we compared two different preparations of HLM, HLM-1 and HLM-2. The two preparations exhibited closely similar parameters of pNP metabolism and their dependencies on CYP2E1 concentration. With both preparations the dependencies of the reaction rate on pNP concentration revealed a noticeable homotropic cooperativity and obeyed the Hill equation with the Hill coefficient equal to 1.5±0.5 and 1.3±0.2 in HLM-1 and HLM-2 respectively.

As shown in Fig. 2, where we compare the substrate dependencies of the rate of oxidation of pNP and CLZ obtained with HLM with different concentrations of incorporated CYP2E1, enrichment of HLM with CYP2E1 results in a multifold increase in the rates of metabolism of both substrates. As illustrated in Fig. 3, the apparent k_{cat} derived from these titrations remains proportional to the amount of CYP2E1 in the membrane up to the content of 0.3-0.4 nmol/mg protein (or about 50% CYP2E1 in the P450 pool) and stabilizes (pNP) or decreases (CLZ) after that point. Other kinetic parameters of pNP and CLZ metabolism (K_M (CLZ) or S_{50} and the Hill coefficient (pNP)) do not show any statistically significant changes. These parameters are shown in Table 3. These results demonstrates that the incorporated CYP2E1 becomes a fully-functional member of the P450 ensemble and do not exhibit any detectable functional differences with the endogenous CYP2E1 in HLM.

Probing the ability of recombinant human P450 species to metabolize CEC and ERR.

In our further studies of the effect of added CYP2E1 on the metabolism of substrates of other P450 species we used fluorogenic substrates 7-etoxy-4-cyanocoumarin (CEC) and 7-ethoxyresorufin, which are known to be primarily metabolized in HLM by CYP1A2 and CYP2C19 [35-36]. In order to better characterize the ability of human cytochrome P450 species to metabolize these substrates we investigated their metabolism by recombinant human CYP1A2, CYP2B6, CYP2C8, CYP2C9, CYP2C19, CYP2D6, CYP2E1, and CYP3A4 using commercial preparations of Supersomes™ containing the respective enzymes. Results of these assays are shown in Table 4. The probed P450 species not found in this table did not exhibit any detectable activity with either substrate.

In good agreement with the previous reports the highest activity in dealkylation of ERR was exhibited by CYP1A2. Some activity with this substrate was also detected with CYP2C8 and CYP2C19. However, although affinity of these CYP2C enzymes to ERR was quite close to that observed with CYP1A2, the rates of reaction observed with them were considerably lower. Although CYP2E1 also reveals some activity in ERR dealkylation, the observed values of V_{max} and K_M (Table 4) leave out a possibility of any sizeable participation of CYP2E1 in ERR metabolism by HLM, at least at the substrate concentrations below 5 μM used in our studies with CYP2E1-enriched HLM.

Although the highest rate of metabolism of CEC was also exhibited by CYP1A2, the activity rendered by CYP2C19 is high enough to consider this enzyme as an important contributor to CEC metabolism, especially taking into account its relatively high abundance in HLM [4]. Another P450 enzyme capable of metabolizing CEC is CYP2B6. However, this enzyme is unlikely to make any important contribution to CEC metabolism in HLM due to low rate of turnover and low concentration of CYP2B6 in HLM.

Therefore, probing the impact of CYP2E1 incorporation on the parameters of metabolism of CEC and ERR by HLM is expected to provide information on the activities of CYP1A2 and CYP2C19 and discriminate between the CYP2E1 effects on the two enzymes.

Impact of CYP2E1 incorporation on the metabolism of CEC and ERR by HLM.

The dependencies of the rate of CEC dealkylation on substrate concentration obtained with intact and CYP2E1-enriched HLM samples HLM-1 and HLM-2 are shown in Fig. 5. The parameters obtained from the approximation of these dependencies with the Michaelis-Menten equation are summarized in Table 5. The rate of metabolism of CEC by HLM-1 was noticeably higher than that observed with HLM-2, consistent with higher concentration of CYP1A2 in HLM-1 as compared to HLM-2 (Table 1). As suggested by the titration traces shown in Fig. 5, CYP2E1 enrichment may

result in some increase in the rate of CEC metabolism in both preparations of HLM. However, this increase cannot be considered statistically significant (Table 5) due to a large bias of the results of individual measurements. More important is a statistically significant decrease in the value of K_M observed with both HLM preparations (Table 5). While in the intact supersomes this value is close to that characteristic to CYP2C19-dependent metabolism of CEC (Table 4), incorporation of CYP2E1 results in its decrease and makes it closer to that is expected for CYP1A2-catalyzed reaction (Table 5).

The change in the main route of CEC metabolism suggested by this initial analysis may be better revealed through fitting the titration curves to the equation of the sum of two Michaelis-Menten functions. In the case of HLM-1, however, this fitting was incapable to resolve two components with different K_M values and inefficient in improving the quality of approximation. However, in the case of HLM-2 the fitting of the datasets with a single Michaelis-Menten function results in systematic deviations suggesting a noticeable biphasisity of the titration curves (Fig. 5b). The use of the equation of the sum of two Michaelis-Menten functions for fitting the datasets obtained with HLM-2 (either with or without incorporated CYP2E1) suggests the values of Km for the two phases to be in the range of 2-4 and 30-70 μM.

Global fitting of the entire dataset obtained with a series of HLM-2 preparations containing various amounts of incorporated CYP2E1 gives the best results with the estimates of K_M of 2.5±0.9 and 58±13 μM for the high- and low-affinity components respectively. These estimates are in a reasonable agreement with the values of 2.3±0.1 and 24±5 μM 2.3 obtained with CYP1A2- and CYP219-containing Supersomes respectively (Table 4). As illustrated in Fig. 6a, the fraction of the high-affinity phase derived from the global fitting increases proportionally to the concentration of CYP2E1 in HLM up to the content of 0.2-0.3 nmol/mg protein and tends to stabilize after that point. This change is likely to be also associated with some increase in the rate of CEC metabolism, although the latter effect cannot be considered statistically significant (Fig. 5b).

The effect of CYP2E1 enrichment on the contribution of the high affinity component in the biphasic profile of CEC dealkylation suggests an important augmentation of the involvement of CYP1A2. In order to probe this possible activation further we investigated the effect of incorporation of CYP2E1 on the metabolism of ERR, the substrate almost exclusively metabolized by CYP1A2. As shown in Fig. 6 increase in the concentration of CYP2E1 causes a remarkable increase in the rate of ERR dealkylation up to the CYP2E1 content of 0.4 nmol/mg protein (or about 50% CYP2E1 in the P450 pool). The fact that this multifold increase in V_{max} is not associated with any substantial changes in the value of K_M (Fig. 6b, inset) rules out a possibility of direct

involvement of CYP2E1-dependent metabolism of CEC and substantiates our conclusion that incorporation of CYP2E1 results in an ample activation of CYP1A2 present in HLM.

Probing the association of CYP2E1 with CYP1A2 and CYP2C19 in Supersomes.

In order to probe if the observed activation of CYP1A2 may be attributed to a direct consequence of the formation of mixed oligomers of CYP1A2 and CYP2E1 we probed the association of these two proteins in the microsomal membrane using CYP1A2-containing Supersomes™ and our FRET-based technique that utilizes CYP2E1 labeled with BODIPY 577/618. The same approach was also used to probe possible interactions of CYP2E1 with CYP2C19, another enzyme metabolizing CEC. In order to have an appropriate control dataset we also monitored the interactions of CYP2E1 with Supersomes™ containing human CPR and cytochrome b_5 but bearing no P450 enzymes.

Similar to what was observed upon incorporation of CYP2E1-BODIPY into HLM (Fig. 1), addition of CYP2E1-BODIPY to the control Supersomes™ as well as to those containing CYP1A2 or CYP2C19 resulted in a stepwise increase in the intensity of fluorescence that obeyed three-exponential kinetics and took 18 – 20 hours for completion. The dependencies of the maximal amplitude of this increase on CYP2E1 concentration in the membrane (Fig. 7) demonstrate a dramatic difference between the CYP1A2-containing microsomes with the microsomes bearing no P450 protein or having CYP2C19 in their membranes. In the latter two cases a substantial degree of dissociation of the CYP2E1-BODIPY homo-oligomers was observed only at low surface densities of the incorporated enzyme, at $R_{L/P}$ ratio in the range of several thousands. In contrast, in the case of CYP1A2 containing microsomes 50% dissociation of homo-oligomers was observed at $R_{L/P}$ ratio around 400:1, similar to what was seen in HLM (Table 2). These results suggest that CYP2E1 efficiently forms mixed oligomers with CYP1A2 but lacks any ability to interact with CYP2C19, which stands in one-to-one correspondence with the observed effect of CYP2E1 on CYP1A2-specific activities in HLM.

Conclusion.

In this study we utilized the approach based on homo-FRET (fluorescent resonance energy transfer where donor and acceptor are represented by the fluorophores of one and the same structure) to monitor the incorporation of CYP2E1 into HLM and probe its interactions with other cytochrome P450 species in the microsomal membrane. Studying the effect of this incorporation on CYP2E1-specific activities of HLM we demonstrate that the incorporated enzyme becomes a fully-functional member of the P450 ensemble and do not exhibit any detectable functional differences

with the natively-present enzyme. Exploring the functional effect of increase in CYP2E1 concentrations we evidenced an important activation of CYP1A2 that results in rerouting of metabolism of CEC, the substrate jointly metabolized by CYP1A2 and CYP2C19, toward preferential dealkylation by CYP1A2. Furthermore, probing the interactions of CYP2E1 with model microsomes containing individual P450 enzymes we found that CYP2E1 efficiently interacts with CYP1A2, but lacks any ability to form complexes with CYP2C19. This observation stands in one-to-one correspondence with the observed effect of CYP2E1 on CYP1A2-specific activities in HLM.

Our results suggest that the ethanol-dependent induction CYP2E1 in human liver may have a pronounced effects on pharmacokinetics of drugs jointly metabolized by CYP1A2 and CYP2C enzymes. Further studies will probe the impact of CYP2E1 on the metabolism of such major drugs as tricyclic antidepressants (amitriptylline, imipramine), and anticonvulsants (phenytoin, carbamazepine), which fall into this category.

Acknowledgments: This research was supported by the National Institute On Alcohol Abuse And Alcoholism of NlH under Award Number R21AA024548. The authors are grateful to Jeffrey P. Jones (WSU) for research support, assistance in obtaining and analyzing LC-MS/MS data and continuous interest to this study. We also gratefully acknowledge the assistance of the "Human Proteome" Core Facility of the Institute of Biomedical Chemistry (IBMC, Moscow, Russia) in generating mass-spectrometry data used in characterization of HLM preparations .

Conflict of interest: The authors declare that they have no conflicts of interest with the contents of this article.

Author contributions: DRD and VGZ designed the study, DRD wrote the paper, DRD, ND, BD, NEV, and MAM conducted the experiments; DRD and VGZ analyzed the results.

References

1 Rendic, S. (2002) Summary of information on human CYP enzymes: Human P450 metabolism data. Drug Metabolism Reviews. **34**, 83-448

2 Guengerich, F. P. and Rendic, S. (2010) Update Information on Drug Metabolism Systems-2009, Part I. Curr Drug Metab.. **11**, 1-3

3 Wienkers, L. C. and Heath, T. G. (2005) Predicting in vivo drug interactions from in vitro drug discovery data. Nature Reviews Drug Discovery. **4**, 825-833

4 Zhang, H. F., Wang, H. H., Gao, N., Wei, J. Y., Tian, X., Zhao, Y., Fang, Y., Zhou, J., Wen, Q., Gao, J., Zhang, Y. J., Qian, X. H. and Qiao, H. L. (2016) Physiological Content and Intrinsic Activities of 10 Cytochrome P450 Isoforms in Human Normal Liver Microsomes. J. Pharm. Exp. Ther. **358**, 83-93

5 Davydov, D. R. (2011) Microsomal monooxygenase as a multienzyme system: the role of P450-P450 interactions. Expert Opinion on Drug Metabolism & Toxicology. **7**, 543-558

6 Reed, J. R. and Backes, W. L. (2012) Formation of P450·P450 complexes and their effect on P450 function. Pharm. Ther. **133**, 299-310

7 Reed, J. and Backes, W. (2016) The functional effects of physical interactions involving cytochromes P450: putative mechanisms of action and the extent of these effects in biological membranes. Drug Metab Rev. . **48**, 453-469

8 Davydov, D. R. (2016) Molecular Organization of the Microsomal Oxidative System: a New Connotation for an Old Term. Biochemistry Moscow-Supplement Series B-Biomedical Chemistry. **10**, 10-21

9 Davydov, D. R., Davydova, N. Y., Rodgers, J. T., Rushmore, T. H. and Jones, J. P. (2017) Toward a systems approach to the human cytochrome P450 ensemble: interactions between CYP2D6 and CYP2E1 and their functional consequences. Biochem. J. **474**, 3523 - 3542

10 Lieber, C. S. and Abittan, C. S. (1999) Pharmacology and metabolism of alcohol, including its metabolic effects and interactions with other drugs. Clinics in Dermatology. **17**, 365-379

11 Cederbaum, A. I. (1998) Ethanol-related cytotoxicity catalyzed by CYP2E1-dependent generation of reactive oxygen intermediates in transduced HepG2 cells. Biofactors. **8**, 93-96

12 Dupont, I., Lucas, D., Clot, P., Menez, C. and Albano, E. (1998) Cytochrome P4502E1 inducibility and hydroxyethyl radical formation among alcoholics. J. Hepatol. **28**, 564-571

13 Cederbaum, A. I. (2006) CYP2E1 - Biochemical and toxicological aspects and role in alcohol-induced liver injury. Mount Sinai J. Medicine. **73**, 657-672

14 Meskar, A., Plee-Gautier, E., Amet, Y., Berthou, F. and Lucas, D. (2001) Alcohol-xenobiotics interactions. Role of cytochrome P450 2E1. Pathologie Biologie. **49**, 696-702

15 Tan, Y., Patten, C. J., Smith, T. and Yang, C. S. (1997) Competitive interactions between cytochromes P450 2A6 and 2E1 for NADPH-cytochrome P450 oxidoreductase in the microsomal membranes produced by a baculovirus expression system. Arch Biochem Biophys. **342**, 82-91

16 Kelley, R. W., Cheng, D. M. and Backes, W. L. (2006) Heteromeric complex formation between CYP2E1 and CYP1A2: Evidence for the involvement of electrostatic interactions. Biochemistry. **45**, 15807-15816

17 Davydov, D. R., Davydova, N. Y., Sineva, E. V. and Halpert, J. R. (2015) Interactions among Cytochromes P450 in Microsomal Membranes: Oligomerization of Cytochromes P450 3A4, 3A5 and 2E1 and its Functional Consequences. J. Biol.Chem. **453**, 219-230

18 Dorian, P., Sellers, E. M., Reed, K. L., Warsh, J. J., Hamilton, C., Kaplan, H. L. and Fan, T. (1983) Amitriptyline and ethanol pharmaco kinetic and pharmacodynamic interaction. European J. Clin. Pharm.. **25**, 325-332

19 Tanaka, E. (2003) Toxicological interactions involving psychiatric drugs and alcohol: An update. J. Clin. Pharm. Ther. **28**, 81-95

20 Davydov, D. R., Davydova, N. Y., Sineva, E. V., Kufareva, I. and Halpert, J. R. (2013) Pivotal role of P450-P450 interactions in CYP3A4 allostery: the case of alpha-naphthoflavone. Biochemical J. **453**, 219-230

21 Spatzenegger, M., Liu, H., Wang, Q. M., Debarber, A., Koop, D. R. and Halpert, J. R. (2003) Analysis of differential substrate selectivities of CYP2B6 and CYP2E1 by site-directed mutagenesis and molecular modeling. J. Pharm. Exp. Ther. **304**, 477-487

22 Jang, H.-H., Dayydov, D. R., Lee, G.-Y., Yun, C.-H. and Halpert, J. R. (2014) The role of cytochrome P450 2B6 and 2B4 substrate access channel residues predicted based on crystal structures of the amlodipine complexes. Arch. Biochem. Biophys. **545**, 100-107

23 Smith, P. K., Krohn, R. I., Hermanson, G. T., Mallia, A. K., Gartner, F. H., Provenzano, M. D., Fujimoto, E. K., Goeke, N. M., Olson, B. J. and Klenk, D. C. (1985) Measurement of protein using bicinchoninic acid. Anal. Biochem. **150**, 76-85

24 Bartlett, G. R. (1959) Phosphorus assay in column chromatography. J. Biol. Chem. **234**, 466-468

25 Shen, A. L., Porter, T. D., Wilson, T. E. and Kasper, C. B. (1989) Structural analysis of the FMN binding domain of NADPH-cytochrome P-450 oxidoreductase by site-directed mutagenesis. J. Biol. Chem. **264**, 7584-7589

26 Massey, V. (1959) The microestimation of succinate and the extinction coefficient of cytochrome c. Biochim. Biophys. Acta. **34**, 225-256

27 French, J. S. and Coon, M. J. (1979) Properties of NADPH-cytochrome P-450 reductase purified from rabbit liver microsomes. Arch. Biochem. Biophys. **195**, 565-577

28 Guengerich, F. P., Martin, M. V., Sohl, C. D. and Cheng, Q. (2009) Measurement of cytochrome P450 and NADPH-cytochrome P450 reductase. Nature Protocols **4**, 1245-1251

29 Kennedy, C. J. (1994) CHAPTER 35 - Xenobiotics: designing an in vitro system to study enzymes and metabolism. In Biochemistry and Molecular Biology of Fishes (Hochachka, P. W. and Mommsen, T. P., eds.). pp. 417-430, Elsevier

30 Hood, C. A., Fuentes, G., Patel, H., Page, K., Menakuru, M. and Park, J. H. (2008) Fast conventional Fmoc solid-phase peptide synthesis with HCTU. J. Pept. Sci. **14**, 97-101

31 Fekkes, D. (1996) State-of-the-art of high-performance liquid chromatographic analysis of amino acids in physiological samples. J. Chromatogr. B-Biomed. Appl. **682**, 3-22

32 Reinke, L. A. and Moyer, M. J. (1985) Para-nitrophenol hydroxylation - a microsomal oxidation which is highly inducible by ethanol. Drug Metab. Dispos. **13**, 548-552

33 Wibo, M., Amar-Costesec, A., Berthet, J. and Beayfay, H. (1971) Electron microscope examination of subcellular fractions III. Quantitative analysis of the microsomal fraction isolated from rat liver. J. Cell. Biol. **51**, 52-71

34 Davydov, D. R., Deprez, E., Hui Bon Hoa, G., Knyushko, T. V., Kuznetsova, G. P., Koen, Y. M. and Archakov, A. I. (1995) High-pressure-induced transitions in microsomal cytochrome P450 2B4 in solution - evidence for conformational inhomogeneity in the oligomers. Arch. Biochem. Biophys. **320**, 330-344

35 Donato, M. T., Jimenez, N., Castell, J. V. and Gomez-Lechon, M. J. (2004) Fluorescence-based assays for screening nine cytochrome P450 (P450) activities in intact cells expressing individual human P450 enzymes. Drug Metab. Dispos. **32**, 699-706

36 Ghosal, A., Hapangama, N., Yuan, Y., Lu, X. W., Horne, D., Patrick, J. E. and Zbaida, S. (2003) Rapid determination of enzyme activities of recombinant human cytochromes P450, human liver microsomes and hepatocytes. Biopharm. Drug Dispos. **24**, 375-384

Table 1. Characterization of microsomal preparations used in this study.

Parameter	HLM-1	HLM-2
Total P450, nmol/mg protein	0.256±0.020	0.398±0.053
CYP2E1, nmol/mg protein	0.032	0.028
CYP1A2, nmol/mg protein	0.032	0.017
CYP3A4, nmol/mg protein	0.057	0.219
CPR , nmol/mg (activity assay)	0.058±0.010	0.051±0.012
CPR, nmol/mg (MS assay)	0.059	0.051
b_5 , nmol/mg protein	0.375±0.019	0.329±0.002
Phospholipid (PL), nmol/mg protein	724±136	650±136
PL/P450 molar ratio	2828	1633
P450/CPR molar ratio	4.4	7.7
CYP2E1 molar fraction[a], %	12.5	7.1
CYP1A2 molar fraction[a],%	12.7	4.3
CYP3A4 molar fraction[a],%	22.4	55.1

[*] The estimates obtained from spectroscopic and enzyme activity assays represent the averages of 2-8 individual measurements. The"±" values are the confidence intervals calculated for p=0.05. The values obtained with mass-spectroscopy are the results a single measurement.

[a] The molar fraction was calculated from the ratio of the content of the individual P450 determined with mass spectrometry to the total P450 content determined with absorbance spectroscopy.

Table 2. Effect of incorporation of CYP2E1-BODIPY on the degree of its homo-olegomerization explored with homo-FRET*

Microsomal preparation	$[CYP2E1]_{50\%}$[a], pmol/cm^2	$(Lipid/CYP2E1)_{50\%}$[b]	FRET Efficiency, %
SS(CPR)	0.081 ±0.020	4323 ±1113	72.9
SS(b5 and CPR)	0.143 ±0.038	2444 ±699	71.8
HLM	0.914 ±0.245	383 ±111	79.0
SS(1A2)	0.890 ±0.040	393 ±18	78.6
SS(3A4)	0.900 ±0.204	388 ±93	77.9
SS(2C19)	0.059 ±0.021	5891 ±2441	80.0

* The values given in the Table were obtained by fitting of the titration curves with the binary association equation (Eq. 1). The "±" values show the confidence interval calculated for p = 0.05.

[a] Surface density of P450 in microsomal membranes at which the amplitude of the titration curves reaches 50% of the maximal level. This value is equal to the value of K_D determine from the fitting of the titration curves with Eq. 1.

[b] Lipid-to-P450 ratio at which the amplitude of the titration curves reaches 50% of the maximal level.

Table 3. Effect of incorporation of CYP2E1 into HLM on the kinetic parameters of oxidation of *p*-Nitrophenol and Chlorzoxazone*

Molar ratio of added CYP2E1 to P450 present in HLM	p-Nitrophenol[a]			Chlorzoxazone[b]	
	S_{50}, μM	N	V_{max}, min^{-1}	K_M, μM	V_{max}, min^{-1}
0	42.3 ±7.8	1.5 ±0.5	5.0 ±1.9	54.4 ±16.1	11.8 ±2.7
0.25	25.8 ±2.7	1.4 ±0.6	9.9 ±3.2	78.5 ±29.0	41.9 ±11.3
0.5	33.5 ±8.7	1.4 ±0.1	22.8 ±11.8	54.2 ±4.4	41.6 ±4.3
1	33.0 ±9.7	1.9 ±0.8	17.5 ±8.6	65.2 ±20.2	77.9 ±8.2
1.75 – 2.0[c]	35.5 ±12.6	2.0 ±0.7	15.2 ±4.6	86.7 ±18.2	54.1 ±11.0

* The estimates given in the table represent the averages of 2-4 individual measurements. The"±" values are the confidence intervals calculated for p=0.05.

[a] The parameters of pNP metabolism given in this table were obtained with the HLM-1 preparation.

[b] The parameters of CLZ metabolism were obtained with the HLM-2 preparation.

[c] The values given in this row for the parameters of pNP hydroxylation were measured at a two-fold molar excess of CYP2E1 over the internal P450 content, while in the case of chlorzoxazone this ratio was equal to 1.75.

Table 4. Parameters of metabolism of 7-CEC and 7-ERR by recombinant human cytochromes P450*.

P450 species	CEC		ERR	
	K_M, μM	V_{max}, s^{-1}	K_M, μM	V_{max}, s^{-1}
CYP1A2	2.3 ±0.10	7.9 ±2.2	0.40 ±0.14	1.1 ±0.3
CYP2B6	8.2 ±2.4	1.0 ±0.2	No activity detected	
CYP2C8	No activity detected		0.26 ±0.14	0.02 ±0.01
CYP2C19	24 ±5	2.3 ±0.5	0.33 ±0.13	0.06 ±0.02
CYP2E1	No activity detected		33.0 ±9.2	0.05 ±0.02

* The estimates given in the table represent the averages of 2-4 individual measurements. The"±" values are the confidence intervals calculated for p=0.05.

Table 5. Parameters of CEC dealkylation in intact and CYP2E1-enriched HLM obtained from the fitting of titration curves with the Michaelis-Menten equation.

HLM sample	K_M, μM	V_{max}, min^{-1}
HLM-1	15.6 ± 3.0	3.4 ± 0.3
HLM-1 + CYP2E1[a]	9.4 ± 1.2 (0.004)	3.8 ± 0.3 (0.159)
HLM-2	32.2 ± 14.7	1.9 ± 0.5
HLM-2 + CYP2E1 [a]	6.1 ± 1.9 (0.009)	2.3 ± 0.8 (0.450)

[*] The estimates given in the table represent the averages of 2-4 individual measurements. The"±" values are the confidence intervals calculated for p=0.05. The values in parentheses represent the p-values of Student's t-test for the hypothesis of equality of the respective values with the value presented in the previous line (i.e., at no CYP2E1 incorporated).

[a] The results obtained with the samples containing CYP2E1 incorporated at 1:1 molar ratio to the microsomal P450 content (0.26 and 0.4 nmol/mg protein for HLM-1 and HLM-2, respectively).

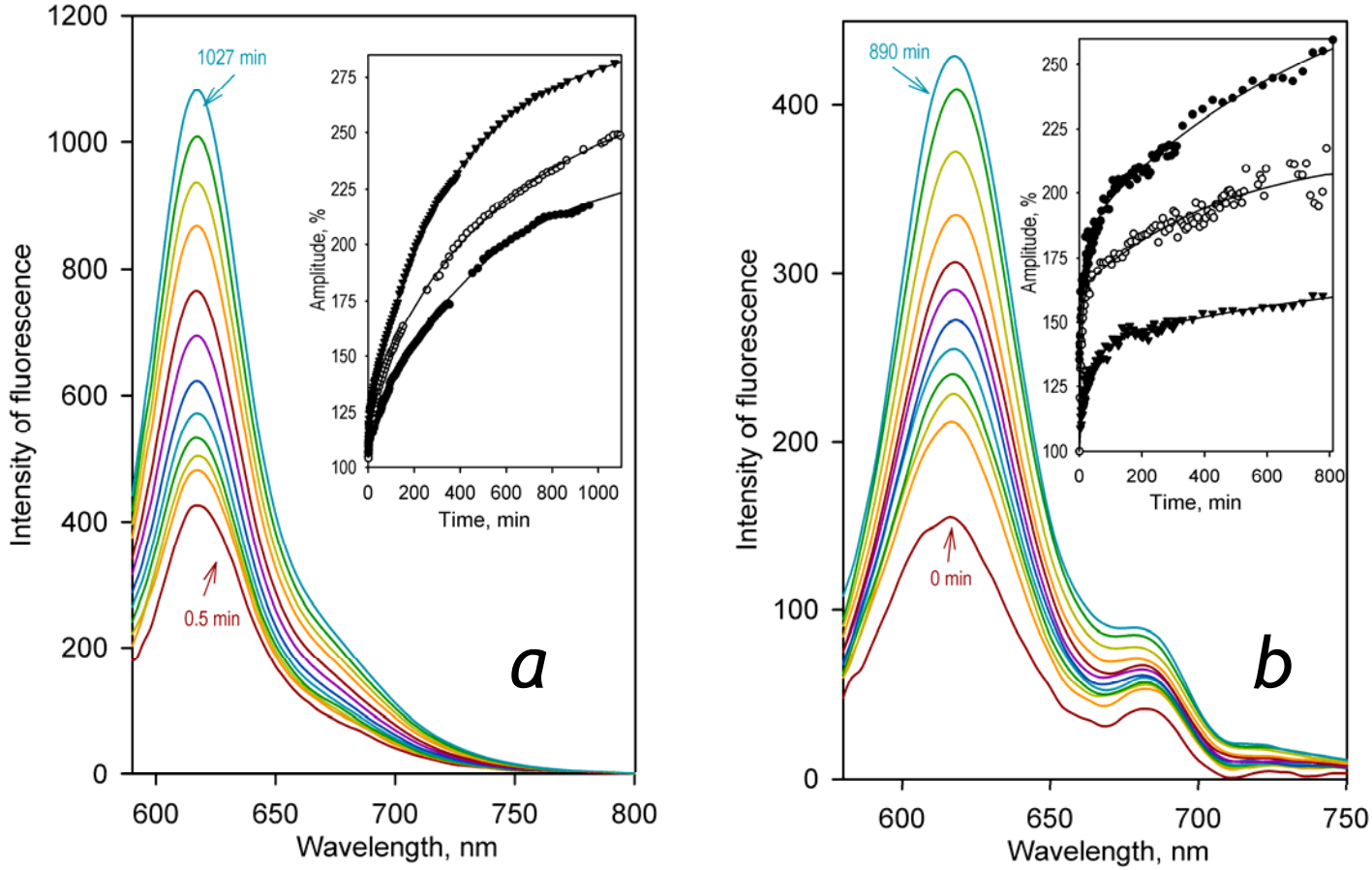


Fig.1. Effect of incorporation of CYP2E1-BODIPY into HLM-1 (***a***) and SS-RR (***b***) on the intensity of BODIPY fluorescence. The main panels show the series of spectra recorded in the process of incorporation and the inserts show the kinetic curves of the increase in the relative intensity of fluorescence of the probe. The spectra shown in panel ***a*** were recorded at 0.5, 2, 5, 16, 33, 67, 120, 230, 370, 530, 740 and 1027 min after addition of CYP2E1-BODIPY to HLM-1 to reach the CYP2E1-to-phospholipids molar ratio ($R_{L/P}$) of 560:1 (or 0.33 μM CYP2E1-BODIPY per mg of microsomal protein). Kinetic curves shown in the inset were recorded at $R_{L/P}$ ratios of 1:213 (filled circles), 1:560 (open circles) and 1:2240 (filled triangles). The spectra shown in panel ***b*** were recorded at 0.1, 0.6, 3,6, 11, 24, 49, 95, 190, 390,780 and 890 min after addition of CYP2E1-BODIPY to SS-RR to reach the $R_{L/P}$ ratio of 3600:1. Kinetic curves shown in the inset were recorded at $R_{L/P}$ ratios of 1:600 (filled triangles), 1:560 (open circles) and 1:2240 (filled circles).

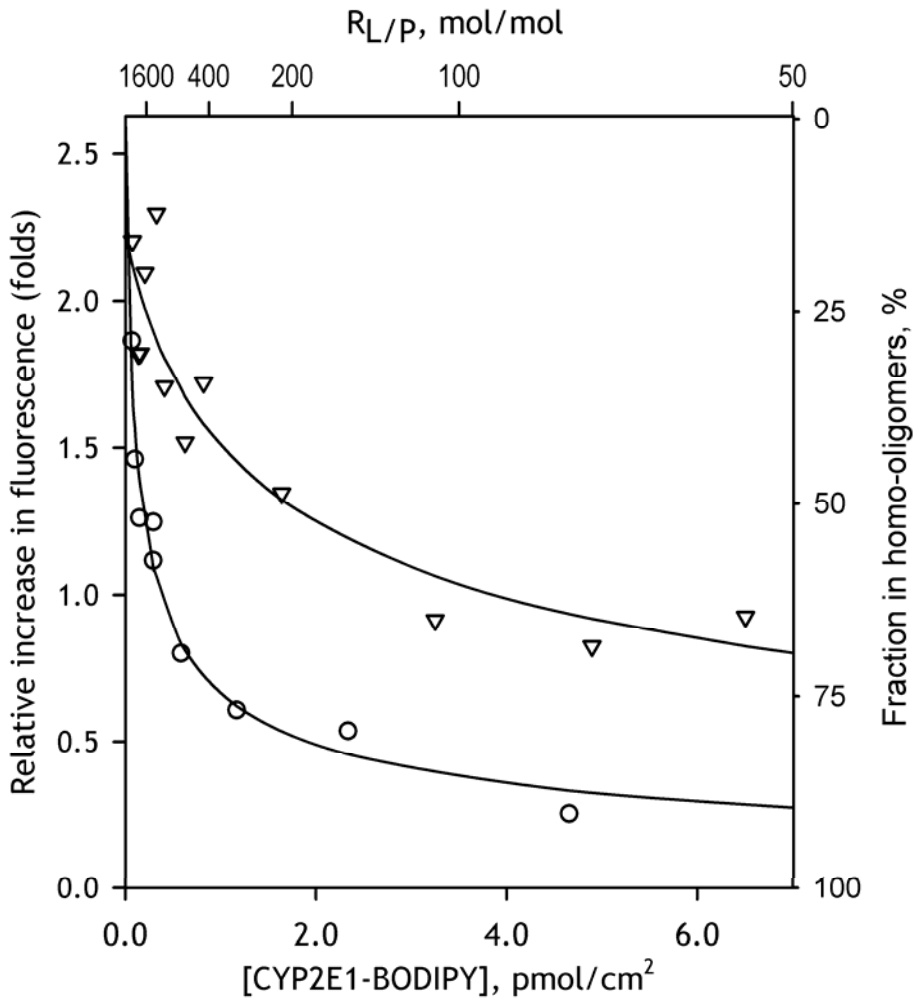


Fig. 2. The dependencies of the relative increase in fluorescence intensity caused by incorporation of CYP2E1-BODIPY into the membranes of HLM-1 (triangles) and SS-RR (circles) on the concentration (surface density) of CYP2E1-BODIPY in the membrane. Solid lines show the approximations of the data sets with Eq. 1. The scale shown on the right of the plot indicates the apparent fraction of CYP2E1 present in homo-oligomeric state estimated from the results of this fitting.

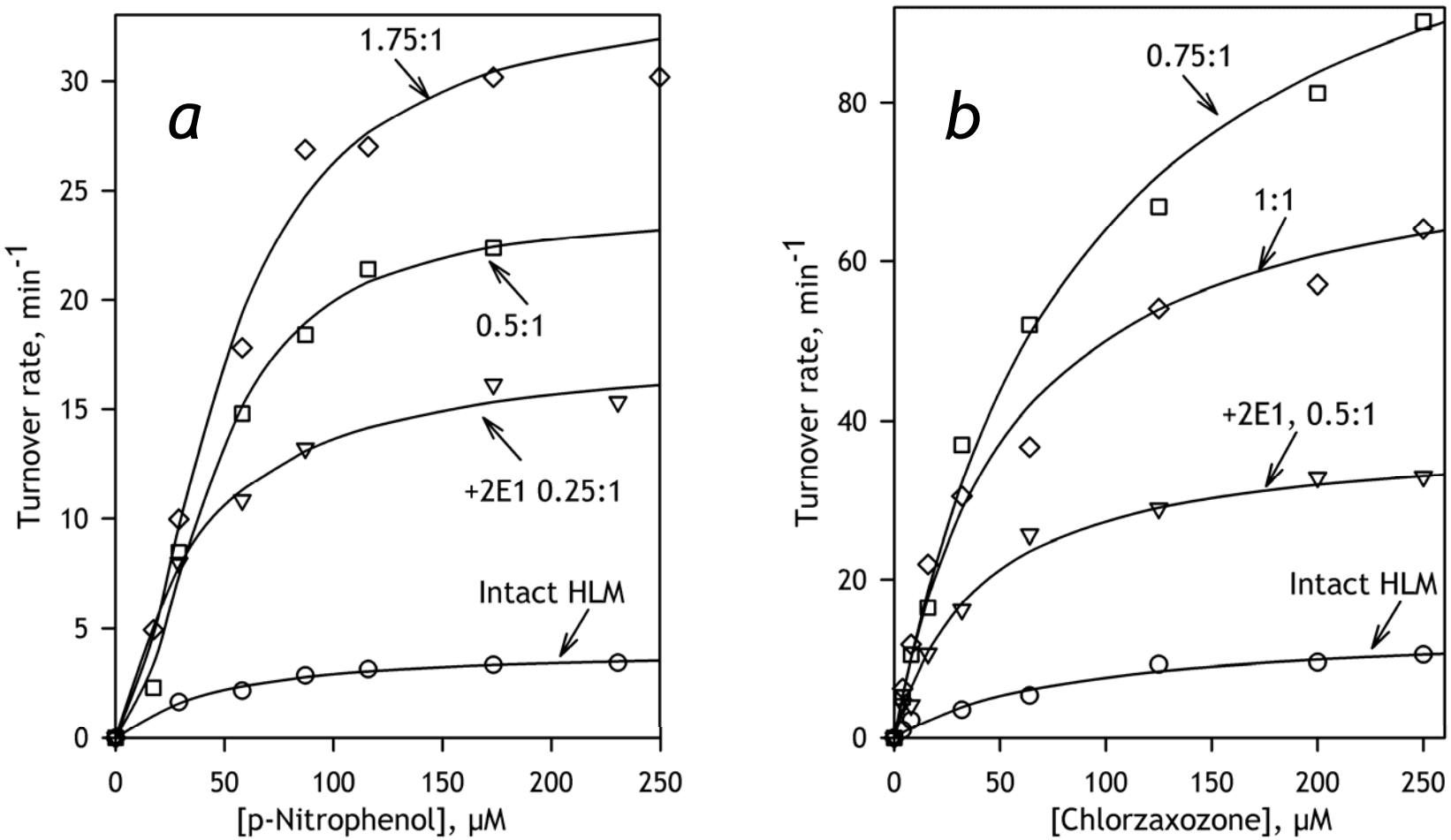


Fig. 3. The effect of incorporation of CYP2E1 into HLM-2 on CYP2E1-dependent hydroxylation of pNP (***a***) and CLZ (***b***). The lines represent the approximations of the data sets with the Hill (***a***) or Michaelis-Menten (***b***) equations. The ratios shown next to each curve indicate the molar ratio of added CYP2E1 to the internal cytochrome P450 content in HLM-2 (0.4 nmol/mg protein).

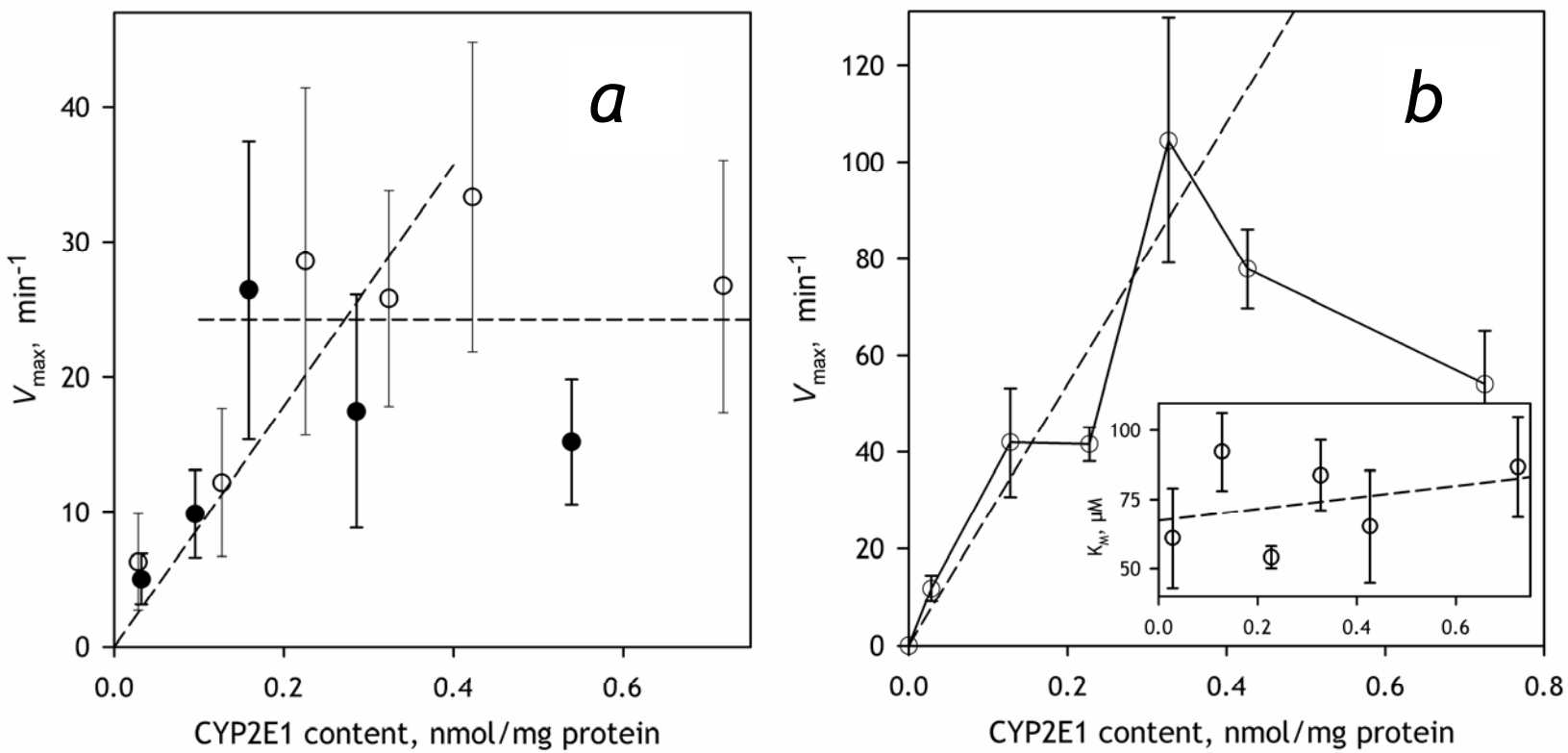
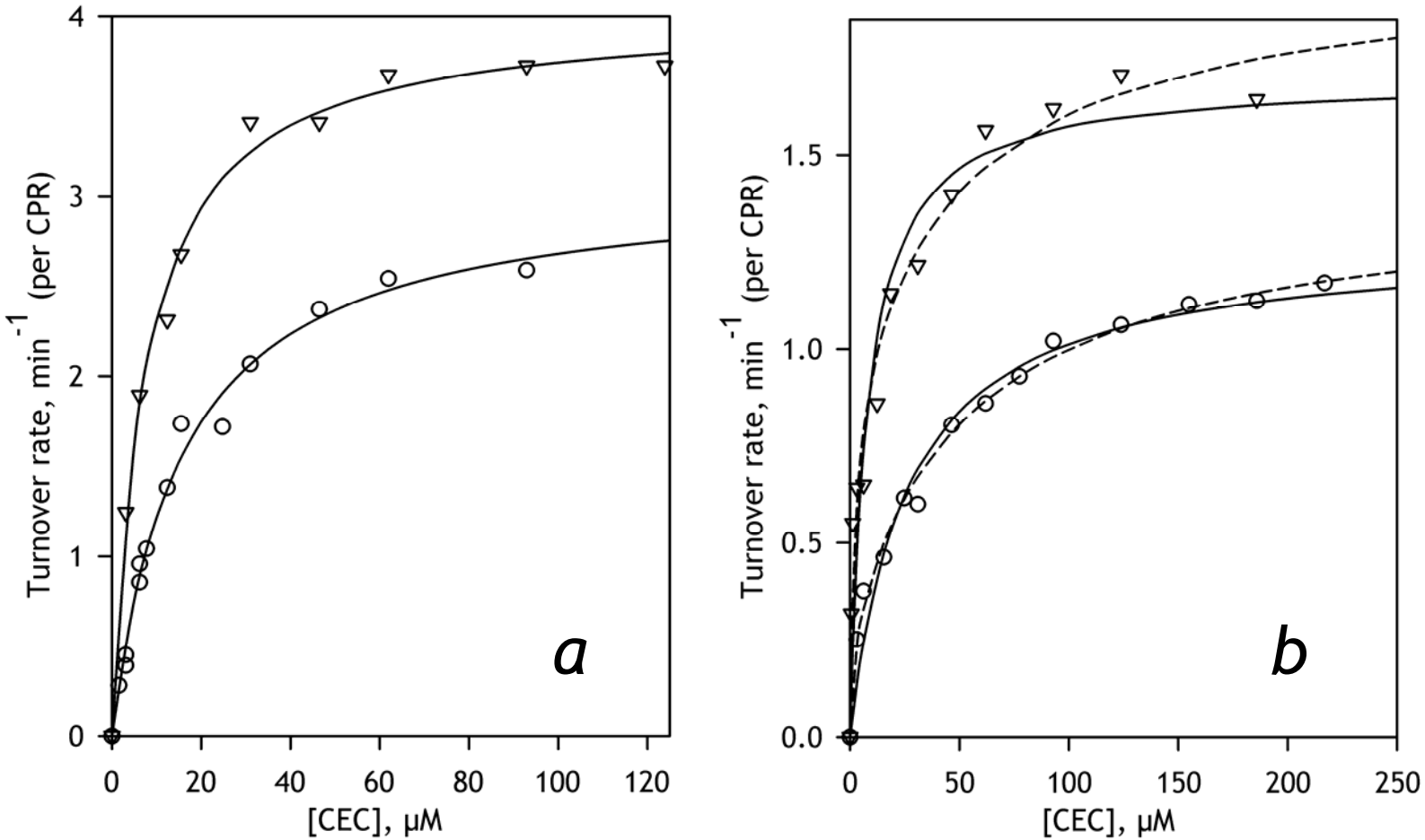


Fig. 4. The effect of incorporation of CYP2E1 into HLM on the metabolism of CYP2E1-specific substrates pNP (***a***) and CLZ (***b***). The data shown in open and filled circles in panel ***a*** were obtained with HLM-1 and HLM-2 preparations respectively. The results presented in the panel ***b*** were obtained with HLM-2. Error bars represent the confidence intervals calculated for p=0.05 on the basis of 2-5 measurements.



Fug. 5. The effect of incorporation of CYP2E1 into HLM on the dependencies of the rate of CEC metabolism on substrate concentration. The results obtained with HLM-1 and HLM-2 are shown in the panels ***a*** and ***b*** respectively. The data obtained with intact HLM are shown in circles while the triangles designate the results obtained with the samples containing CYP2E1 incorporated at 1:1 ratio to the microsomal P450 content (0.26 and 0.4 nmol/mg protein for HLM-1 and HLM-2, respectively). The solid limes represent the results of fitting of the datasets to the Michaelis-Menten equation. The dashed lines in the panel ***b*** shows the approximations with the equation of the sum of two Michaelis-Menten functions.

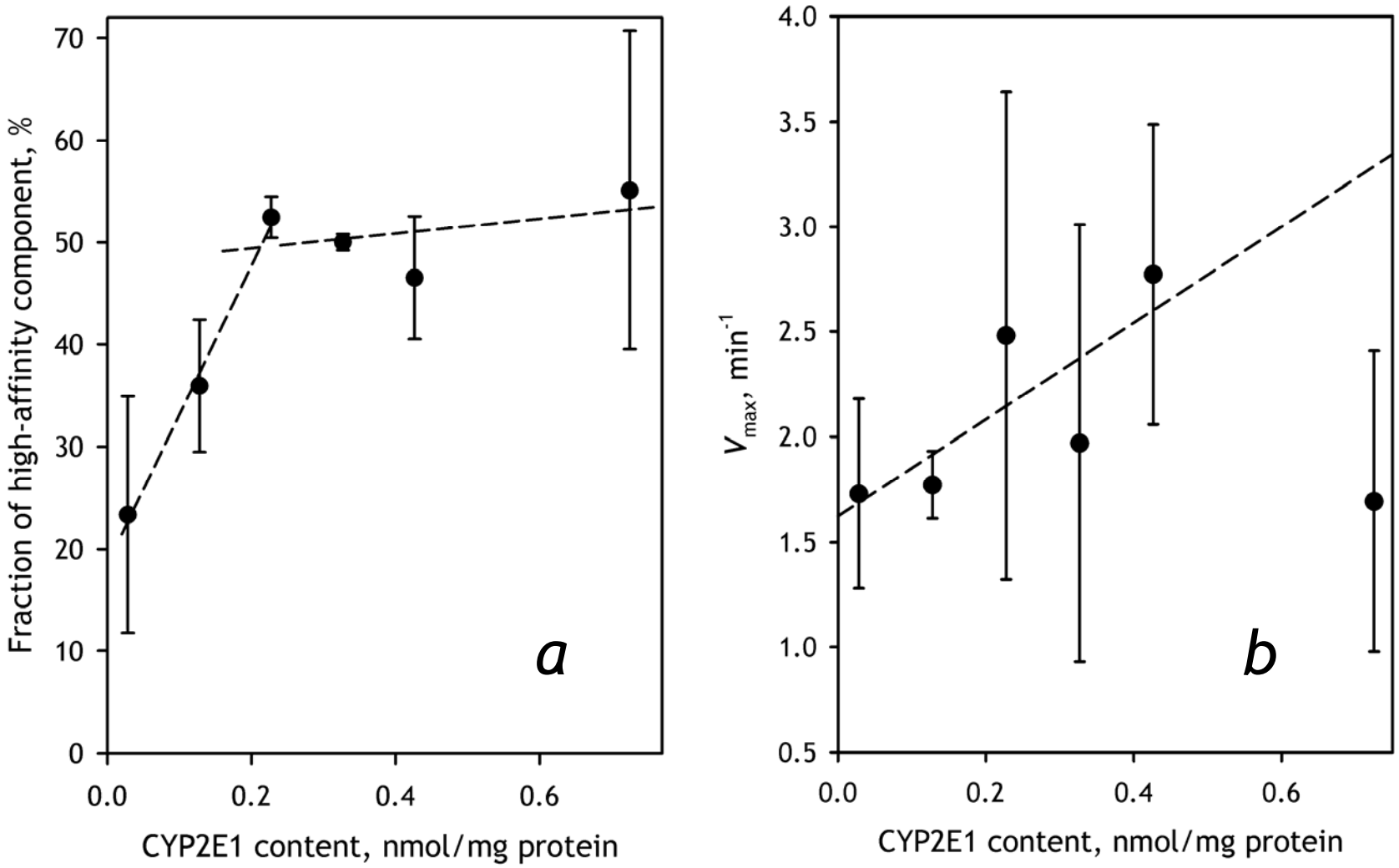


Fig. 6. The effect of incorporation of CYP2E1 into HLM-2 on the parameters of metabolism of CEC. Panel ***a*** shows the dependence of the fraction of the high-affinity component on CYP2E1 content. The dependence of the total maximal rate of metabolism is sown in panel ***b.*** Error bars represent the confidence intervals calculated for p=0.05 on the basis of 2-4 measurements.

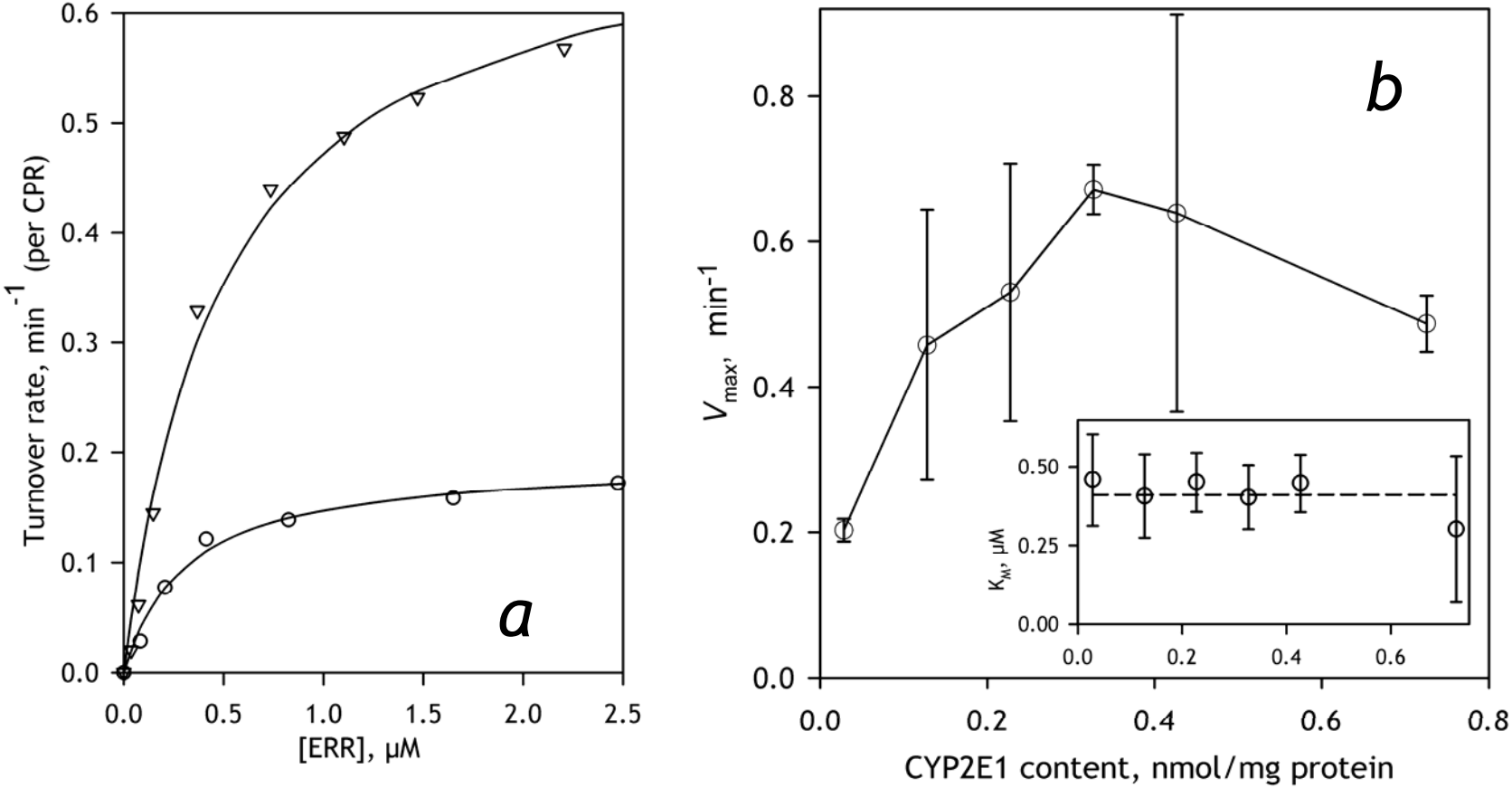


Fig. 7. The effect of incorporation of CYP2E1 into HLM-2 on the parameters of metabolism of ERR. Panel ***a*** exemplified the dependence of the rate of turnover on substrate concentration obtained with intact HLM (circles) and the sample containing CYP2E1 incorporated in the amount of 0.3 nmol per mg protein that corresponds to 0.75:1 ratio to the microsomal P450 content (triangles). Panel ***b*** shows the dependencies of V_{max} (main panel) and K_M on CYP2E1 content. Error bars represent the confidence intervals calculated for p=0.05 on the basis of 2-5 measurements.

Fig.7

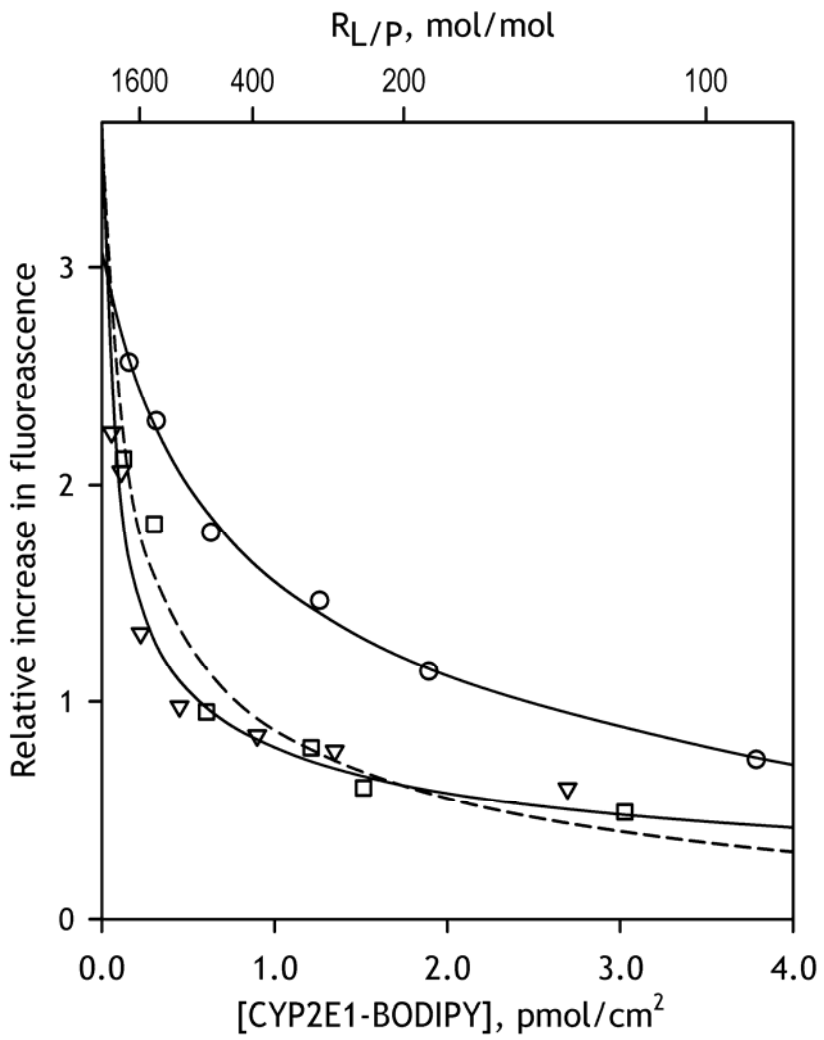


Fig. 8. Effect of increasing concentrations of CYP2E1-BODIPY on the relative increase in fluorescence intensity caused by incorporation of CYP2E1-BODIPY into the membranes of Supersomes™ containing recombinant CYP1A2 and CPR (circles, solid line), CYP2C19, CPR and cytochrome b_5 (triangles, solid line) or control Supersomes™ containing human CPR and cytochrome b_5 but bearing no cytochromes P450 (squares, dashed line). The lines show the approximations of the data sets with Eq. 1.