Sexual selection does not increase the rate of compensatory adaptation to a mutation influencing a secondary sexual trait in *Drosophila melanogaster*

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Abstract

Theoretical work predicts that sexual selection can enhance natural selection, increasing the rate of adaptation to new environments and helping purge harmful mutations. While some experiments support these predictions, remarkably little work has addressed the role of sexual selection on compensatory adaptation—populations' ability to compensate for the costs of deleterious alleles that are already present. We tested whether sexual selection, as well as the degree of standing genetic variation, affect the rate of compensatory evolution via phenotypic suppression in experimental populations of *Drosophila melanogaster*. These populations were fixed for a spontaneous mutation causing mild abnormalities in the male sex comb, a structure important for mating success. We fine-mapped this mutation to an ~85 kb region on the X chromosome containing three candidate genes, showed that the mutation is deleterious, and that its phenotypic expression and penetrance vary by genetic background. We then performed experimental evolution, including a treatment where opportunity for mate choice was limited by experimentally enforced monogamy. Although evolved populations did show some phenotypic suppression of the morphological abnormalities in the sex comb, the amount of suppression did not depend on the opportunity for sexual selection. Sexual selection, therefore, may not always enhance natural selection; instead, the interaction between these two forces may depend on additional factors.

Introduction

Sexual selection has important impacts on many aspects of how organisms evolve, including on speciation rates and the degree of sexual dimorphism (e.g., Masta and Maddison 2002; Ellis and Oakley 2016). It was once thought that sexual selection may act independently or even antagonistically to other components of natural selection (e.g. viability and fecundity). However, sexual selection might also be concordant with, and represent a substantial portion of, the total selection on an allele. If so, sexual selection on males might also influence the overall mutation load or rate of adaptation, including in females. For instance, sexual selection may influence how organisms respond to selective pressures in the short term, influencing how quickly populations adapt to novel environments, in particular when the population begins at a distance from an optimum (Long *et al.* 2012). Additionally, some models predict that sexual selection should help populations filter out harmful mutations more rapidly than selection on other fitness components (viability and fecundity selection) alone (Agrawal 2001). This prediction is based partly on the observation that sexual displays are often correlated with overall condition. Any mutation that reduces an organism's nonsexual fitness is therefore also likely to affect its mating success (Rowe and Houle 1996; Chandler *et al.* 2013b) or even its success in sperm competition (Clark *et al.* 2012). In those cases, total selection against such mutations is stronger than it would be without sexual selection.

Empirical support for this scenario has been mixed. In some studies testing these predictions, evidence supported a role for sexual selection in purging deleterious mutations or accelerating adaptation (Radwan 2004; Sharp and Agrawal 2008; Hollis *et*

61 *al.* 2009; Jarzebowska and Radwan 2010; McGuigan *et al.* 2011; Long *et al.* 2012;
62 Almbro and Simmons 2014; Lumley *et al.* 2015; Grieshop *et al.* 2016; Jacomb *et al.*
63 2016). However, a handful of studies also contradict these predictions (Hollis and Houle
64 2011; Plesnar *et al.* 2011; Arbuthnott and Rundle 2012; Cabral and Holland 2014;
65 Chenoweth *et al.* 2015), perhaps because of the confounding effects of sexual conflict,
66 or because they used large-effect mutations or strong mutagens not representative of
67 natural variation.

68 Of course, deleterious mutations are not always purged by selection; they can
69 increase in frequency and occasionally become fixed via drift, hitchhiking, or
70 antagonistic pleiotropy, especially if their effects on fitness are only mildly deleterious
71 (and in populations with a small effective population size). In those cases where the
72 deleterious alleles are difficult for selection to purge, alleles at other loci that
73 compensate epistatically for the fitness costs of these fixed deleterious alleles may be
74 favored by selection. There is evidence of compensatory adaptation in both microbial
75 and multicellular organisms. For instance, alleles conferring antibiotic resistance are
76 sometimes costly in the absence of antibiotics, but compensatory mutations can reduce
77 those costs (Reynolds 2000; Maisnier- Patin *et al.* 2002; Comas *et al.* 2012). In the blowfly,
78 diazinon resistance via alleles at the *Rop-1* gene had negative pleiotropic effects,
79 increasing fluctuating asymmetry, but these effects were ultimately compensated by
80 modifiers (McKenzie and Clarke 1988; Davies *et al.* 1996). Additionally, sex
81 chromosome dosage compensation could also be considered a form of compensatory
82 adaptation, having likely evolved in response to loss-of-function mutations on Y or W
83 chromosomes (Charlesworth 1978). In addition, the phenotypic expression (penetrance

84 and expressivity) of many mutations can be strongly influenced by genetic background
85 (e.g., Chandler *et al.* 2013a, 2017; Mullis *et al.* 2018; Hou *et al.* 2019). Thus, selection
86 favoring suppressor alleles at other loci may also contribute to compensatory adaptation
87 by limiting the phenotypic expression of a deleterious mutation.

88 Although sexual selection has received a lot of attention as a possible influence
89 on the rate of purging of deleterious mutations, the role of sexual selection in
90 compensatory evolution remains largely unexplored. Nevertheless, we might similarly
91 predict that sexual selection can also accelerate compensatory adaptation, especially if
92 sexual displays are condition dependent. In one study (Pischedda and Chippindale
93 2005), the *nub*^{1} mutation, which drastically reduces the size of the wing, resulting in an
94 inhibition of males' ability to generate courtship songs, was fixed in experimental
95 populations of *Drosophila melanogaster*. As predicted, this mutation had greater fitness
96 costs in males than it did in females, but males also showed greater compensatory
97 fitness recovery over 180 generations (albeit without compensating for the effects on
98 wing morphology directly; A. Chippindale, personal communication), providing some
99 support that sexual selection may enhance compensatory adaptation. However, this
100 study was not replicated (only a single lineage), since the *nub*^{1} populations were
101 originally generated for other purposes. Clearly, more study is needed on whether
102 sexual selection can speed up compensatory adaptation.

103 In this study, we address the question of whether sexual selection can impact the
104 rate of compensatory evolution (via phenotypic suppression) using experimental
105 evolution in *Drosophila melanogaster*. We chose a mutation in the *sex combs distal*
106 gene (*scd*^{1}) (Boube *et al.* 1997; Randsholt and Santamaria 2008), a spontaneous,

107 partially penetrant mutation affecting the development of the male sex comb, a structure
108 critical for male mating success (Ng and Kopp 2008) and rapidly evolving across
109 *Drosophila* species (Atallah *et al.* 2009, 2012; Kopp 2011; Malagón *et al.* 2014). First,
110 we further mapped the mutation and characterized its effects across different wild type
111 genetic backgrounds; because we found abundant genetic variation in natural
112 populations modifying its penetrance and expressivity, we next focused on
113 compensatory adaptation via phenotypic suppression in experimentally evolved
114 populations. Despite a general compensatory response, we observed no evidence that
115 sexual selection influenced the rate of compensatory evolution via phenotypic
116 suppression of the sex comb phenotypes.
117

118 **Methods**

119 All data and scripts are available on Github (https://github.com/DworkinLab).

120 *Mapping scd^1*—

121 *sex combs distal^1* (*scd^1*) is a spontaneous X-linked allele resulting in ectopic sex comb
122 bristles on the second tarsal segment of the prothoracic leg in males (Boube *et al.*
123 1997). In the base stock (Bloomington Drosophila Stock Center strain #5070, *y^1 scd^1*
124 *ras^1 v^1 f^1*), it has incomplete penetrance, with only about 70% of males showing the
125 ectopic sex comb bristles (Figure 1), and no visible phenotype in homozygous or
126 heterozygous females. In our populations, males also sometimes exhibited minor
127 defects in the primary sex comb, such as a gap or partially untransformed bristles.
128 The identity of the gene and molecular lesion of this allele are unknown, although
129 some previous recombination mapping suggested it was near 1-30.6, and that a local

130 duplication of the 8C-9B region of the X chromosome onto the Y (*DP(1:Y)FF*), could
131 partially rescue the phenotype of *scd*^{1} (Santamaria and Randsholt 1995; Randsholt and
132 Santamaria 2008). We attempted to further fine map the gene through duplication
133 mapping. Virgin female flies of strain BDSC 5070 (y^{1} scd^{1} ras^{1} v^{1} f^{1}) were crossed to
134 males of strains carrying duplicated segments of the X chromosome translocated onto
135 the Y chromosome or chromosome III (Table 1; Cook *et al.* 2010; Venken *et al.* 2010),
136 and the male offspring were scored for the presence of mutant phenotypes, such as the
137 ectopic sex comb or disruptions in the primary sex comb. Assuming *scd*^{1} is a recessive
138 loss-of-function allele, if the duplicated segment contains a functional wild-type copy of
139 the *scd* gene, then the mutant phenotype would be rescued and no male offspring from
140 these crosses would show sex comb defects. Because *scd*^{1} is only partially penetrant,
141 we scored numerous male offspring from each cross.
142 *Influence of genetic background on penetrance and expressivity—*
143 To determine the extent of genetic variation for the penetrance and expressivity
144 of *scd*^{1}, we crossed virgin female y^{1} scd^{1} ras^{1} v^{1} f^{1} flies to males of a randomly chosen
145 subset of Drosophila Genetic Reference Panel lines (Mackay *et al.* 2012). When the
146 adult F1 offspring eclosed, we fixed specimens in 70% ethanol, and then mounted male
147 prothoracic legs in 70% glycerol and scored them for the presence of ectopic sex combs
148 on the second tarsal segment, abnormalities in the primary sex comb, and primary sex
149 comb tooth number. These crosses only examine each wild-type genetic background in
150 a heterozygous state, and thus it is expected that this will underestimate the actual
151 degree of background dependence, as recessive effects of alleles in each background
152 will not be captured.

153 To test for an effect of genetic background on penetrance, we fit a logistic model
154 testing for the effect of genotype on presence of an ectopic sex comb using glm() in
155 base R version 3.6.1. We also confirmed those results using glmer() in the lme4
156 package version 1.1-21.

157 *Introgression of scd^1*—

158 To generate populations for experimental evolution, we introgressed the scd^1
159 mutation into FVW, a domesticated lab-maintained population founded from flies
160 collected from Fenn Valley, MI in 2010. The FVW population was maintained in
161 population cages with 10 bottles for egg-laying each generation for approximately 10
162 generations prior to beginning introgressions (Chari *et al.* 2017).

163 To begin the introgression (Supplementary Figure 1A), virgin females of the 5070
164 progenitor strain (with the genotype y^1 scd^1 ras^1 v^1 f^1) were crossed to FVW males; this
165 stock strain carries visible X-linked genetic markers (y^1 causes yellow body
166 pigmentation, ras^1 and v^1 affect eye color, and f^1 produces forked bristles). The
167 heterozygous F1 females were then backcrossed to FVW males. From the F2 offspring,
168 we selected males showing the scd^1 phenotype, which were thus hemizygous for scd^1,
169 but with phenotypically wild-type eyes and normal bristles to eliminate the ras^1, v^1, and
170 f^1 mutations, which are all located to the right of scd^1 on the X chromosome. We then
171 crossed these males to virgin FVW females, to obtain female offspring heterozygous for
172 scd^1 in a partial FVW background. We then crossed these females to FVW males, and
173 selected males with ectopic sex combs, but not yellow bodies, to eliminate the y^1
174 mutation to the left of scd^1. Each male from these crosses thus carries an independently
175 derived X chromosome with scd^1 in a random FVW background.

176 We obtained eight such males and crossed them to FVW females. From these
177 crosses, we obtained virgin females heterozygous for *scd*^{1} in an FVW background. The
178 first of these virgin *scd*^{1}/*scd*$^{+}$ females to emerge were crossed with FVW males to obtain
179 more *scd*^{1} males with a mostly FVW background. The later-emerging *scd*^{1}/*scd*$^{+}$ females
180 were kept isolated at cooler temperatures (18˚C) until the *scd*^{1} males from the previous
181 cross emerged. We then crossed the *scd*^{1}/*scd*$^{+}$ females to the *scd*^{1} males. Finally, we
182 set up sib matings among the offspring of these crosses, using only males hemizygous
183 for *scd*^{1} (showing a sex comb phenotype) and females of unknown genotype (either
184 *scd*^{1}/*scd*$^{+}$ or *scd*^{1}/*scd*^{1}). Of those crosses, we kept those in which the mother was
185 inferred to be homozygous for *scd*^{1}, in which nearly all male progeny displayed *scd*^{1}
186 phenotypes. This allowed us to establish a homozygous *scd*^{1} line with an FVW genetic
187 background, which we designated as *scd**. *scd** carried at least four independently
188 derived X chromosomes with *scd*^{1} in an FVW background (Supplementary Figure 1A).
189 To introduce further genetic diversity (from the FVW population) into *scd**, *scd**
190 males were crossed to wild-type FVW females to obtain heterozygous *scd*^{1} females with
191 additional genetic material from the FVW background. These females were then
192 backcrossed to FVW males. Five replicate backcrosses were set up in culture bottles,
193 each with 25-30 FVW males and 25-30 females, heterozygous for *scd*^{1} and for alleles
194 from the FVW background. We then selected males with the *scd*^{1} phenotype, and
195 backcrossed them to virgin *scd** females, in six replicate bottles each containing 20-25
196 males and 20-25 females, to maintain *scd*^{1} while introducing additional genetic diversity
197 from the FVW population. This whole cycle was then repeated once to establish the
198 *scd*** base population for experimental evolution (Supplementary Figure 1B).

199 *Fitness effects of scd^1*—

200 To test whether the *scd*^{1} allele was deleterious, we tracked changes in the
201 frequency of the *scd*^{1} phenotype in polymorphic populations with the *scd*^{1} allele at 0.7
202 initial frequency. We initiated four replicate populations, each consisting of 70 *scd***
203 males, 70 *scd*** females, 30 FVW males, and 30 FVW females. Populations were
204 placed in population cages with four culture bottles for mating and oviposition for five
205 days, after which the flies were discarded and the bottles transferred to fresh cages at
206 24°C. After adult flies began emerging, they were allowed to mate for three to four days.
207 The old bottles were then removed, and fresh bottles were placed in the cage for egg
208 laying. After two days of egg laying, the flies were discarded and the bottles moved to
209 fresh cages. This cycle was repeated for a total of nine generations.

210 For each of the first five generations, and at generation nine, we scored male sex
211 comb phenotypes. 50 males were picked randomly, and the first prothoracic legs from
212 each male were mounted on glass slides in 70% glycerol/PBS to check for the presence
213 of an ectopic second sex comb and other abnormalities. While this does not give an
214 exact measurement of the frequency of the *scd*^{1} allele because of this allele's
215 incomplete penetrance (though penetrance is almost complete in the FVW background;
216 see below), it should provide a reasonable proxy. Even though reductions in the
217 frequency of the mutant phenotype could also be driven by selection for suppressor
218 alleles, this should still give an indication of whether or not the *scd*^{1} phenotype is
219 deleterious.

220 To test whether there was evidence that the *scd*^{1} allele was deleterious (and
221 decreased in frequency) we fit a logistic regression tracking number of *scd*^{1} and wild-

222 type males each generation. As the frequency of *scd*[1] at generation 0 was set at exactly
223 0.7, we utilized an offset and suppressed the model intercept. Additionally we checked
224 the results of this model using a logistic mixed model allowing for a variation in the slope
225 of the response by replicate lineage. Analyses were conducted in R using glm() and
226 glmer() from the lme4 package.

227 *Experimental evolution—*

228 To test whether sexual selection influences the rate of compensatory adaptation,
229 we set up two treatments. In the low sexual selection (LSS) treatment, we removed
230 sexual selection by enforcing monogamous mating. Each generation, we set up 100
231 vials, each containing one male and one virgin female. After a three-day interaction
232 period, we anesthetized the flies using CO_2, discarded males, and placed the females in
233 a population cage with four bottles containing culture media for egg laying. After four
234 days, the bottles were removed and incubated at 24˚C. When adult flies began eclosing,
235 we selected virgins for the next generation. Thus, while this treatment did preclude mate
236 choice, it still allowed for fecundity and viability selection (Arbuthnott and Rundle 2012).

237 In the high sexual selection (HSS) treatment, we followed a similar protocol
238 except allowed the opportunity for sexual selection. Each generation, 100 males and
239 100 virgin females were allowed to interact in a population cage, along with an open
240 culture bottle for food and moisture. After the three-day interaction period, we placed the
241 cage in a refrigerator to knock the flies out, and then we sorted males and females.
242 Males were discarded, and females were placed in fresh cages with four culture bottles
243 for a four-day egg-laying period. After egg laying, females were discarded, and the

244 bottles were placed in an environmental chamber at 24˚C until adults began emerging,
245 at which point we collected virgins for the next generation.
246 Additionally, we also set up a treatment with low levels of genetic variation (LV)
247 to test whether compensatory adaptation is limited when segregating genetic variation is
248 diminished; in other words, testing whether the mutational target size of the
249 compensatory response was large enough that *de novo* mutations could contribute in
250 the time frame of the experimental evolution regime. In this treatment, each population
251 was established from the offspring of a single-pair mating between a randomly chosen
252 virgin *scd*** female and a randomly chosen *scd*** male. These populations were kept
253 under the same regime as the HSS treatment.
254 Finally, we set up a wild-type control (WTC) treatment using wild-type FVW flies.
255 WTC populations were also maintained under the same regime as the HSS treatment.
256 These provide a control for lab domestication and unknown aspects of the experimental
257 protocol.
258 All populations were initiated using randomly selected *scd*** flies (see above),
259 except for the LV treatments as described. We set up three replicate populations of
260 each treatment except for WTC, in which we performed two replicates. Experimental
261 evolution was conducted for a total of 24 generations. We assayed male sex comb
262 phenotypes as described earlier at generations 1, 7, 13, 19, and 24, using 30 randomly
263 selected males from each population at each time point.
264 To test how male sex comb traits changed over the course of experimental
265 evolution, we fit generalized linear mixed models using the glmmTMB v0.2.3 (Hadfield
266 2010) and lme4 v1.1-21 (Bates *et al.* 2015) packages in R (v3.6.1). For sex comb tooth

267 number for both primary and ectopic/secondary combs, we assumed a Poisson
268 distribution and used a log link function. The model included generation, treatment, and
269 their interaction as fixed effects; we also included individual fly, and in some cases
270 replicate population nested within treatment, as random effects (some models failed to
271 converge when replicate nested within treatment was included as a random effect). We
272 also tested for lineage specific zero-inflation in the data, but found no evidence for this,
273 so excluded this to reduce number of parameters. To test whether the frequency of
274 defects in the primary sex comb changed over time, we fit a mixed logistic model (sex
275 comb defects present/absent), again with generation, treatment, and their interaction as
276 fixed effects, and replicate nested within treatment, as well as individual fly, as random
277 effects. Power simulations were performed using simr v1.0.5 (Green and MacLeod
278 2016).
279
280 **Results**
281 *Mapping scd^1*—
282 Though we could not map *scd^1* to a specific gene, we were able to further narrow
283 down its location to an ~85-kb (cytological region 8F8-9A1) region on the X
284 chromosome through duplication mapping (Table 1, Figure 2). This region contains only
285 two complete annotated protein-coding genes, *btd* and *Sp1* (both of which influence
286 aspects of leg development and morphogenesis), and three annotated long non-coding
287 RNAs, CR42657, CR44016, and CR53498. Interestingly, the two parent strains carrying
288 the duplications that rescued *scd^1* had smaller than average sex combs in the absence
289 of the *scd^1* mutation (Table 1), similar to a past study involving this mutation (Randsholt

290 and Santamaria 2008), suggesting that the *scd* gene product is a suppressor of sex
291 comb development.
292 *Influence of genetic background on penetrance and expressivity*—
293 When females of the original *scd*^{1} stock strain were crossed to males of various
294 wild-type strains to generate males that were hemizygous for *scd*^{1} and heterozygous for
295 different genetic backgrounds, the penetrance and expressivity of *scd*^{1} varied widely
296 (Figure 3) demonstrating segregating variation for them. A logistic model using
297 penetrance (presence/absence of ectopic sex comb) as the response variable with the
298 genetic background as a fixed effect was a significantly better fit than a null model not
299 accounting for genetic background (χ^2 = 127.6, df = 18, p = 5.3 x 10^{-19}), and when we fit
300 a model including genetic background as a random effect, there was substantial among-
301 strain variance (σ^2 = 4.19 on the link scale). Similarly, a model with number of ectopic
302 sex comb teeth as the response variable (expressivity) and genetic background as a
303 fixed effect was a significantly better fit than the null model (χ^2 = 259.0, df = 18, p < 1.0 x
304 10^{-10}). The progenitor *scd*^{1} strain from the Bloomington stock center had the lowest
305 penetrance (frequency of flies exhibiting an ectopic sex comb on the second tarsal
306 segment) and among the lowest expressivity (number of teeth in the ectopic sex comb).
307 Some of the other wild-type genetic backgrounds, even in a heterozygous state,
308 resulted in nearly complete penetrance for *scd*^{1}, including the outbred population (FVW)
309 used for experimental evolution (described below). Interestingly, the FVW outbred
310 population only showed intermediate levels of expressivity of the mutant phenotype,
311 consistent with segregating variation in this population. Overall this result suggests
312 genetic background has a strong impact on the phenotypic expression on *scd*^{1}. It also

313 suggests that the partial penetrance initially observed in the progenitor stock (strain
314 5070) may reflect the accumulation of suppressor/compensatory mutations in the base
315 stock center strain itself. These results suggest that a compensatory response could
316 potentially be due to the accumulation of segregating suppressor alleles.

317 *Fitness effects of scd^1*—

318 The frequency of male flies exhibiting the scd^1 phenotype (in an FVW genetic
319 background) decreased across five generations of experimental evolution in populations
320 polymorphic for scd^1 (Figure 4). With a starting allele frequency of 0.7 the frequency
321 decreased to an average frequency of 0.4 (across the multiple replicates) in males by
322 generation 9. To test this more rigorously, we fit a logistic model with an offset (starting
323 frequency of scd^1 = 0.7), and the effect of generation was significant (effect = -0.197 on
324 logit link scale, s.e. = 0.049, $p = 5.1 \times 10^{-5}$). These results are consistent with the
325 mutation having moderate deleterious effects.

326 *Experimental evolution*—

327 In the populations carrying the scd^1 allele, defects such as gaps in the sex comb
328 were observed occasionally, and at significantly higher frequencies in the High Sexual
329 Selection (HSS) populations, and marginally significant frequencies in the Low Sexual
330 Selection (LSS) populations, than in the wild-type populations. The frequency of these
331 gaps appeared to decrease in the HSS populations across the 24 generations of the
332 experiment, although the interaction between generation and treatment was not
333 significant (Figure 5B; Table 2). In addition, the ectopic sex combs induced by the scd^1
334 mutation became smaller on average in both the HSS and LSS treatments (Figure 5C;
335 Table 3), losing on average ~0.5 teeth across the 24 generations of experimental

evolution. This is consistent with some of the compensatory response being the result of the increase in frequency of naturally occurring suppressor alleles. Again, we saw no evidence for differences in rate of compensation between the HSS and LSS treatments, with the magnitude of the interaction term (change in slope relative to LSS) being ~1/10 the magnitude of the effect of generation. This suggests the additional compensatory effects of sexual selection were relatively weak in this experimental system. Using a power analysis, we confirmed that the power to detect such an effect would be very small (Supplementary Figure 2) unless we used a large number of independent replicate lineages (~30 per treatment), although the power to detect such an effect of such a magnitude (assuming it was real) would be approximately 80% with three replicates if the response continued for 40 generations of experimental evolution (Supplementary Figure 3).

For the Low genetic Variation (LV) treatment no significant change over time was observed, as expected (generation effect = 0.0017, SD = 0.0044, p = 0.70). No significant changes were observed in the primary sex comb tooth number across 24 generations in any of the experimental treatments (Figure 5A; Table 4).

Discussion

Some, but not all, previous work has found that sexual selection may facilitate populations in purging deleterious mutations (Radwan 2004; Hollis *et al.* 2009; Jarzebowska and Radwan 2010) or accelerating rate of adaptation (Jacomb *et al.* 2016; Parrett and Knell 2018). Few studies, however, have addressed whether sexual selection may facilitate compensatory adaptation, where populations evolve traits to

359 compensate for the fitness costs of deleterious mutations. While compensatory
360 adaptation itself is well documented in other systems (Reynolds 2000; Maisnier-Patin *et*
361 *al.* 2002; Estes *et al.* 2011; Chandler *et al.* 2012; Comas *et al.* 2012; Chari *et al.* 2017),
362 in this experiment, we found no effect of the sexual selection regime on the rate of
363 compensatory adaptation (at least with respect to the mutation's sex comb phenotypes).
364 This is perhaps surprising, because we found clear evidence of standing genetic
365 variation modulating the expression of this mutation, so genetic variation does not
366 appear to be a limiting factor here. Moreover, our experiments show that the scd^1
367 mutation is deleterious (Figure 4), and given the importance of the *Drosophila* sex comb
368 for male mating success (Ng and Kopp 2008), we expected that the fitness costs of this
369 mutation would involve male sexual fitness. Thus, we predicted that the costs of this
370 mutation would be higher in the HSS treatment, in which there was a high opportunity
371 for female mate choice, than in the LSS treatment, with reduced opportunity for sexual
372 selection. It is possible that this mutation has effects on other aspects of fitness in males
373 or females (viability, fecundity), but unfortunately our experiments did not directly
374 measure specific fitness components. Even so, theoretical work predicts that sexual
375 selection should act in concert with natural selection because of condition dependence
376 (Whitlock and Agrawal 2009); that is, mutations that reduce nonsexual fitness should
377 also reduce mating success, since sexual displays are often indicators of overall
378 condition. While some work has supported this prediction, our findings add to a growing
379 body of work suggesting that this is not always the case (Hollis and Houle 2011; Plesnar
380 *et al.* 2011; Arbuthnott and Rundle 2012, 2014; Cabral and Holland 2014; Power and
381 Holman 2015; Chenoweth *et al.* 2015). It is possible that the relatively minor degree of

382 phenotypic suppression observed here made it difficult to detect differences between
383 treatments. In fact, it is interesting that the largest difference between the HSS and LSS
384 treatments appears to be the decline in primary sex comb defects in the HSS population
385 (Figure 5), though this difference is non-significant, given that we might expect primary
386 sex comb defects to be more deleterious than the presence of an “extra” sex comb.

387 Even though sexual selection did not impact the rate of compensatory evolution,
388 we did observe evidence of weak compensatory adaptation via phenotypic suppression
389 of the *scd*^{1} mutation in both the HSS and LSS treatments. On average, the ectopic sex
390 combs lost about half a tooth (starting with a mean of ~ 3.5 teeth) over the course of 24
391 generations in these populations; in other words, the expressivity of the mutation
392 declined slightly. One possible explanation for the similar response in both the high and
393 low sexual selection treatment is simply that the compensatory response (in terms of
394 phenotypic suppression) was sufficiently weak that any subtle difference between these
395 treatments would be difficult to detect given our design. However, the power analyses
396 (Supplementary Figures 2 and 3) suggest that if an effect of this magnitude were real, it
397 is sufficiently small that it would require ~25 replicate lineages of each treatment to
398 detect or a doubling of the number of generations of experimental evolution.

399 As expected, we did not observe any significant trend in the LV treatment, in
400 which populations experienced genetic bottlenecks prior to beginning the experiment
401 (LV populations were treated the same way as HSS populations). Combined with the
402 observation that genetic background has strong influences on the penetrance and
403 expressivity of this mutation (Figure 3), this suggests that compensatory adaptation by
404 phenotypic suppression relies heavily on the presence of standing genetic variation,

405 rather than rapid accumulation of new mutations. An interesting side note is that the
406 initial strain (obtained from the Drosophila stock center) carrying *scd*^{1} has among the
407 lowest penetrance/expressivity for this mutation of all the genetic backgrounds that we
408 tested. This may suggest that the stock strain has already undergone compensatory
409 adaptation, and that alleles suppressing the phenotypic expression of *scd*^{1} had become
410 fixed throughout the maintenance of this stock (which were subsequently removed
411 when we outcrossed the mutation), though unfortunately we do not have any data on
412 how long the *scd*^{1} stock strain has been maintained.

413 While we were unable to map *scd*^{1} to a specific gene, we were able to localize it
414 to an ~85 kb region containing only a few candidates. The only protein-coding candidate
415 genes in this region, *Sp1* and *btd*, both have known roles in leg development (Estella
416 and Mann 2010), but are not specifically known to influence sex comb development.
417 There are also three long non-coding RNAs in this region (CR42657, CR44016, and
418 CR43498). Interestingly, all three of these RNAs show evidence of male-specific
419 expression in modENCODE RNA-seq data available on FlyBase (Graveley *et al.* 2011;
420 Brown *et al.* 2014). However, CR44016 shows expression only at very low levels and
421 only in adult males, not pupae or larvae, suggesting it is unlikely to be involved in the
422 development of sex combs. CR42657 and CR43498 both show expression in pupae
423 and/or larvae, as well as adult males (but not adult females); however, these RNAs
424 seem to be expressed in the testis and accessory gland and not other tissues (though
425 expression in legs specifically was not assessed in the modENCODE dataset). This
426 suggests that these male-specific reproductive tissues may be driving these sex-specific

427 expression patterns, not a role in sex comb development. Further work is necessary to
428 identify the molecular nature of *scd*^{1}.

429 There are a number of important limitations to point out about our study. First,
430 much of the focus was on compensation by suppression of the phenotypic effects of the
431 *scd*^{1} mutation on the sex combs directly. While we observed similar levels of phenotypic
432 compensation with both our high and low sexual selection treatments (LSS and HSS), it
433 is possible that compensatory evolution differed with respect to the fitness components
434 (viability, fecundity, and sexual/mating components), which were not evaluated. Thus
435 we limit our interpretation to the effects on morphological compensation/suppression,
436 recognizing that we cannot rule out differential patterns of compensatory response for
437 fitness *per se*. Indeed, this pattern has been observed previously (Pischedda and
438 Chippindale 2005; Chari *et al.* 2017). Additionally, this experiment was performed over a
439 relatively short time period (25 generations); if we continued the experiment over a
440 longer period, subtle differences in the rate of morphological compensation may have
441 become apparent (Supplementary Figure 3).

442 In summary, we found evidence of moderate compensatory adaptation to a
443 deleterious mutation by selection for modifier alleles that suppress the mutation's
444 phenotypic effects. However, while compensatory adaptation did depend on the
445 presence of standing genetic variation, it did not depend on the opportunity for sexual
446 selection, in spite of the affected phenotype's known role in mating. This adds to a
447 growing body of studies suggesting that sexual selection does not always enhance
448 natural selection. Future work should tease apart when and why sexual and natural

selection act in concert and when they are likely to operate differently (Martínez-Ruiz and Knell 2017).

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458 **Figure Legends**

459 **Figure 1.** (A) Wild-type *Drosophila* leg showing a normal male sex comb (black arrow).
460 (B) scd^1 leg showing the normal primary sex comb (black arrow) as well as a smaller
461 ectopic sex comb on the second tarsal segment (white arrow).

462 **Figure 2.** Duplication mapping of scd^1. Purple bars represent duplications that rescued
463 the scd^1 mutant phenotype; pink bars represent duplications that failed to rescue scd^1.
464 (A) Entire region of the X chromosome in which duplications were tested. (B) Close-up
465 of the putative scd^1-containing region (red box). Because the duplication carried by
466 strain 30333 is sufficient to rescue the scd^1 phenotype, we hypothesize that the *scd*
467 gene must lie entirely within this region; at the same time, the neighboring duplications
468 (30522 and 30334) did not rescue the scd^1 phenotype, so *scd* must lie at least partially
469 outside of these regions.

470 **Figure 3.** (A) Penetrance and (B) expressivity of the scd^1 mutation varies among
471 different wild type genetic backgrounds. In these experiments, stock scd^1 females were
472 crossed to males of different wild-type strains, and phenotypes were scored in F1 males
473 (which were hemizygous for scd^1 and heterozygous for a different wild genetic
474 background). Penetrance was measured as the proportion of males showing an ectopic
475 sex comb on the second tarsal segment of the prothoracic leg, while expressivity was
476 measured as the number of sex comb teeth on the ectopic sex comb. Error bars
477 indicate 95% confidence intervals.

478 **Figure 4.** The scd^1 mutation is deleterious. Average frequency of scd^1 over time, across
479 4 replicate populations, each initialized with 70% scd^1 males. Error bars indicate 95%

480 confidence intervals. The shaded region indicates the 95% confidence interval for the
481 best-fit line, with the starting frequency fixed at 70%.
482 **Figure 5.** Plots of model fits for sex comb traits in experimental evolution populations.
483 (A) Number of teeth in the primary sex comb across 24 generations of experimental
484 evolution in all experimental treatments. (B) Proportion of male flies with defects in the
485 primary sex comb in all four treatments. (C) Number of teeth in ectopic/secondary sex
486 combs in males in the HSS and LSS treatments. Shaded regions on plots indicate 95%
487 confidence intervals for the predicted values.
488
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642

Table 1. Duplication mapping of the *scd*^{1} allele. Homozygous female *scd*^{1} flies were crossed to males carrying a segment of the X chromosome duplicated onto either the Y chromosome or chromosome III. Assuming *scd*^{1} is a recessive loss-of-function allele, if the male progeny of these crosses show a mutant phenotype, then the duplication does not rescue *scd*^{1} and *scd* must lie at least partially outside the duplication. If none of the male offspring show a mutant phenotype, then *scd*^{1} is rescued and *scd* lies within the duplication.

Duplication Stock #	**Symbol**	**Duplicated region**	**Rescues *scd*^{1}?**
29759	Dp(1;Y)BSC35	X:8192725-8271204;X:9030055;Y & X:1;X:493529;Y	No; replicated 2x
30522	Dp(1;Y)BSC146	X:8714331-8897281;X:9686653;Y & X:1;X:493529;Y	No; replicated 2x
29782	Dp(1;Y)BSC58	X:9355691-9500067;X:10744934;Y & X:1;X:493529;Y	Yes; replicated 2x; note: duplication on its own has reduced sex comb teeth in primary comb (mean=8.2, sd=1.3, SE=0.4, N=11)
29783	Dp(1;Y)BSC59	X:10085520-10218380;X:10744934;Y & X:1;X:493529;Y	No
29784	Dp(1;Y)BSC60	X:10331363-10385547;X:10744934;Y & X:21572099-22456281;h28-h29;Y & X:1;X:493529;Y	No
29786	Dp(1;Y)BSC62	X:10460679-10601454;X:10744934;Y & X:1;X:493529;Y	No
29788	Dp(1;Y)BSC64	X:10601454-10738950;X:10744934;Y & X:1;X:493529;Y & X:21204834-21318903;h28-h29;Y	No
30333	Dp(1;3)DC212	X:9677341;X:9784700;3L:6442676 (r6, Dp)	Yes; note: duplication on its own has small sex combs for primary comb (3-5 teeth)
30334	Dp(1;3)DC213	X:9762229;X:9807047;3L:6442676 (r6, Dp)	No

8 **Table 2.** Effect estimates for logistic mixed model for gaps in the primary sex comb as response variable; generation,

9 treatment, and their interaction as fixed effects; and replicate nested within treatment, and individual fly as random effects.

	Estimate	Standard Error	*P*
Intercept	-4.96	1.2	4.6×10^{-5}
generation	0.040	0.07	0.57
treatment-HSS	3.0	1.3	0.017
treatment-LSS	2.3	1.3	0.071
treatment-LV	1.3	1.4	0.36
generation x treatment-HSS	-0.094	0.075	0.21
generation x treatment-LSS	-0.046	0.075	0.54
generation x treatment-LV	-0.038	0.081	0.64

10

11

12 **Table 3.** Effect estimates for generalized linear model (poisson) using number of teeth in the ectopic/secondary sex comb
13 as response variable; generation, treatment, and their interaction as fixed effects; and replicate nested within treatment,
14 and individual fly as random effects. This model included only the HSS and LSS populations; WTC populations were
15 excluded because they did not display ectopic sex combs, and because the purpose of this model was to test for a
16 difference between the HSS and LSS treatments specifically, we considered the LV treatment on its own separately.

	Estimate	Standard Error	*P*
Intercept	1.38	0.091	$< 2.0 \times 10^{-16}$
generation	-7.6×10^{-3}	3.9×10^{-3}	0.049
treatment-LSS	-0.12	0.13	0.36
generation x treatment-LSS	-1.5×10^{-3}	5.6×10^{-3}	0.79

17

18

19 **Table 4.** Effect estimates for generalized linear mixed model for primary sex comb tooth number as response variable;

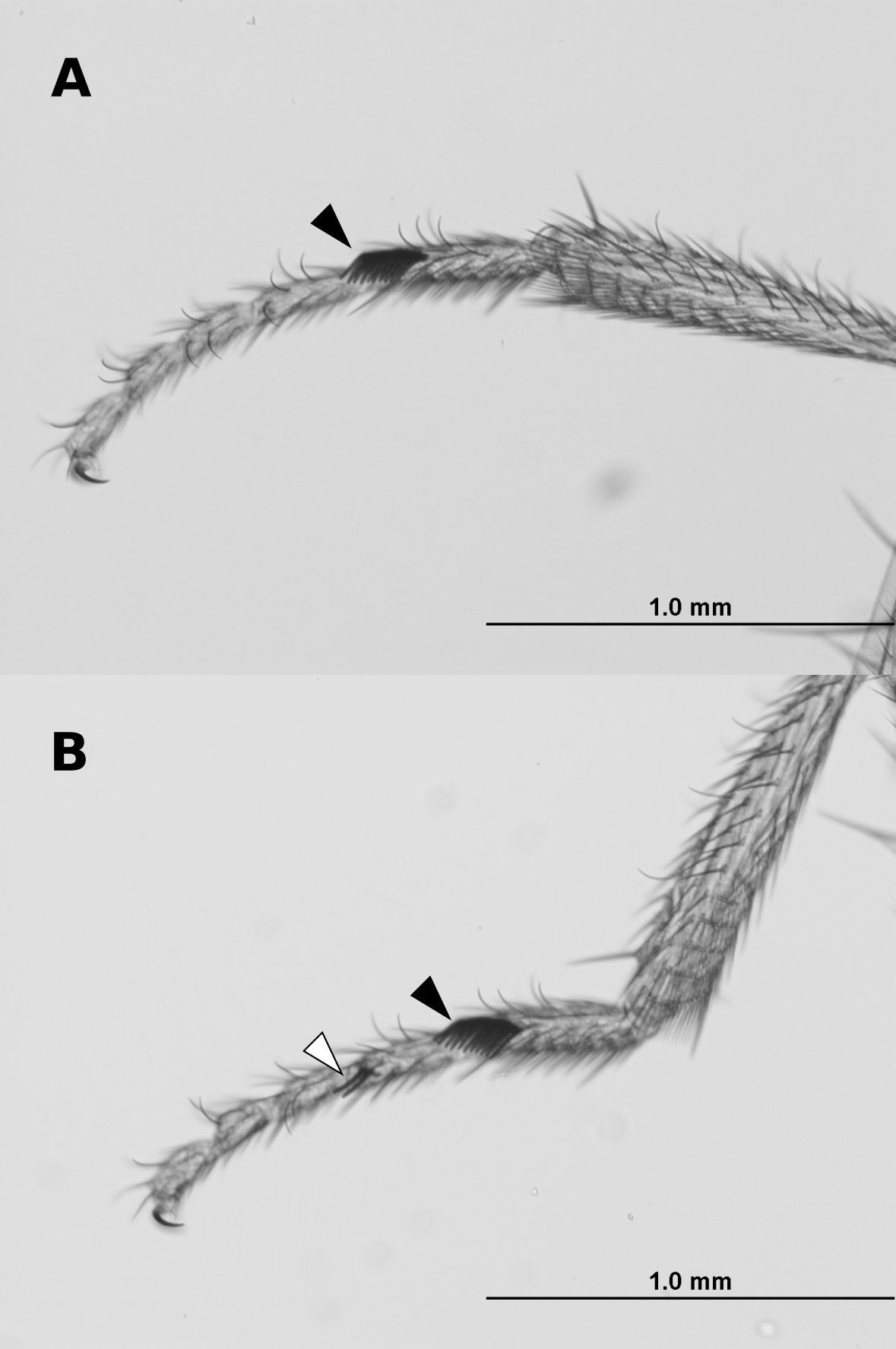
20 generation, treatment, and their interaction as fixed effects; and individual fly as a random effect (models including

21 replicate population nested within treatment failed to converge).

	Estimate	Standard Error	*P*
Intercept	2.41	0.035	$< 2 \times 10^{-16}$
generation	4.4×10^{-4}	2.2×10^{-3}	0.85
treatment-HSS	4.2×10^{-4}	0.046	0.99
treatment-LSS	0.017	0.045	0.71
treatment-LV	6.9×10^{-3}	0.047	0.88
generation x treatment-HSS	5.2×10^{-4}	2.9×10^{-3}	0.86
generation x treatment-LSS	1.3×10^{-3}	2.9×10^{-3}	0.66
generation x treatment-LV	1.4×10^{-3}	3.0×10^{-3}	0.64

22

23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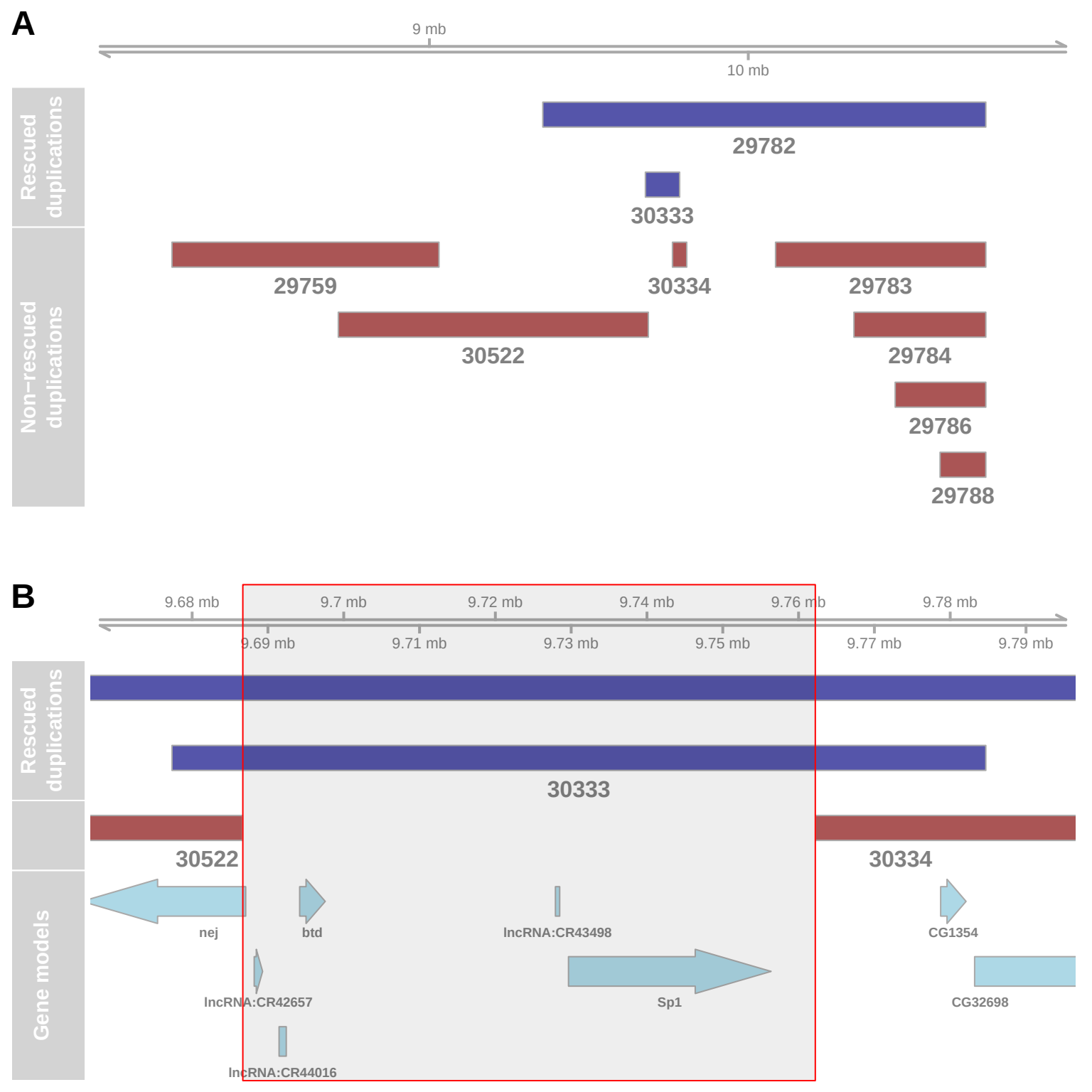
A
9 mb
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Rescued duplications
29782
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Non−rescued duplications
29759
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29786
29788
B
9.68 mb
9.7 mb
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9.78 mb
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9.71 mb
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Rescued duplications
30333
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CG32698
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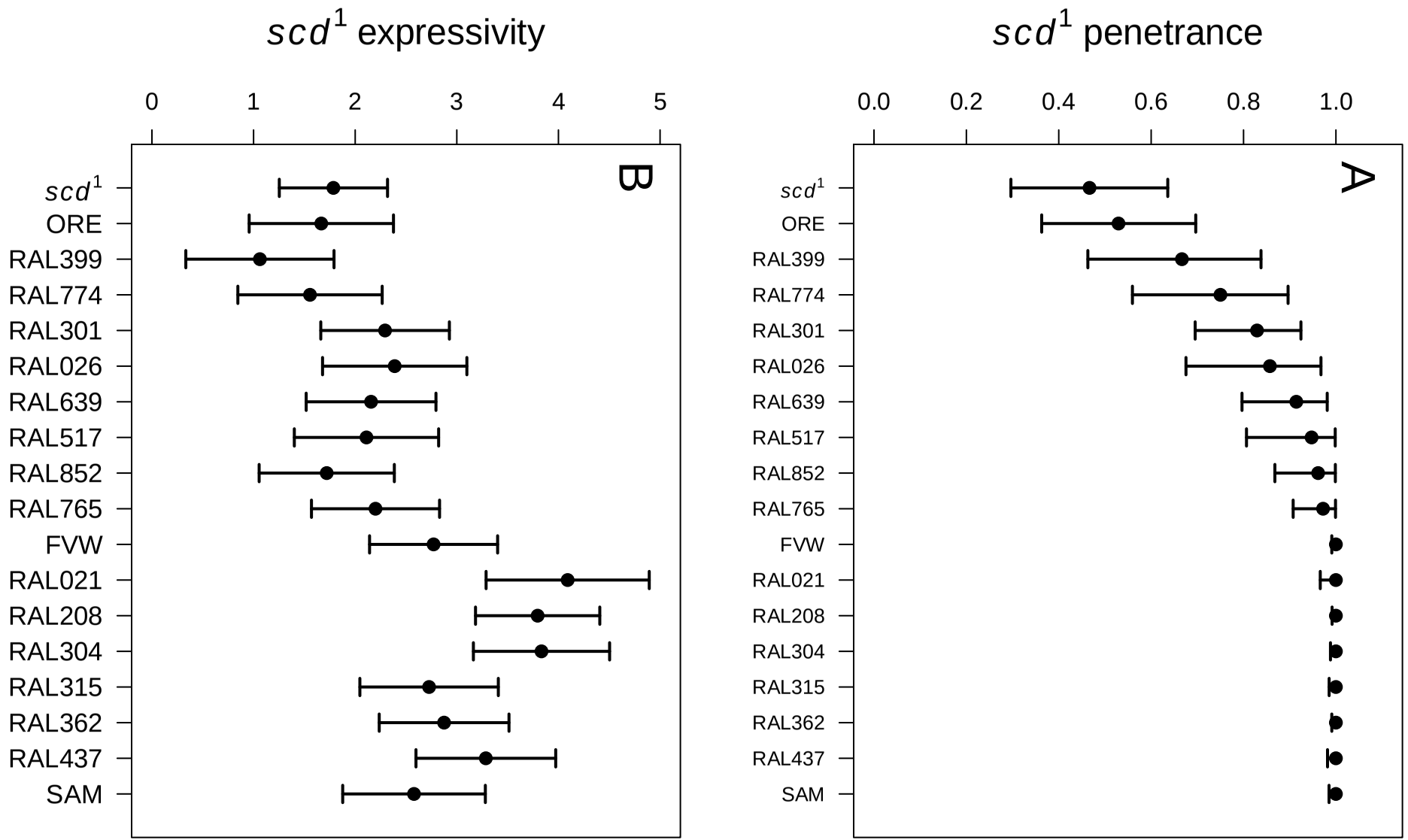
scd1 expressivity
0
1
2
3
4
5
B
scd1
ORE
RAL399
RAL774
RAL301
RAL026
RAL639
RAL517
RAL852
RAL765
FVW
RAL021
RAL208
RAL304
RAL315
RAL362
RAL437
SAM
scd1 penetrance
0.0
0.2
0.4
0.6
0.8
1.0
A
scd1
ORE
RAL399
RAL774
RAL301
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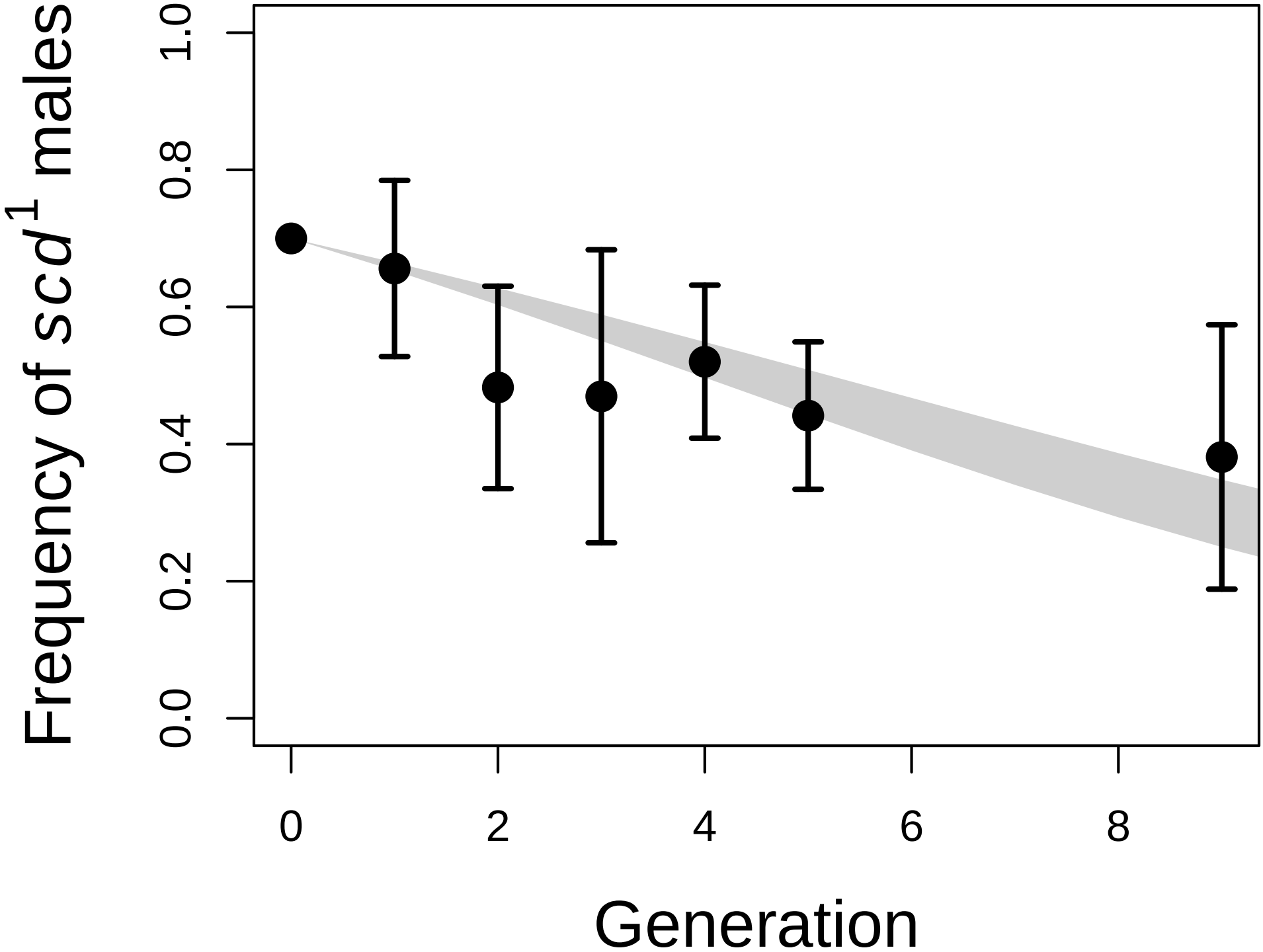
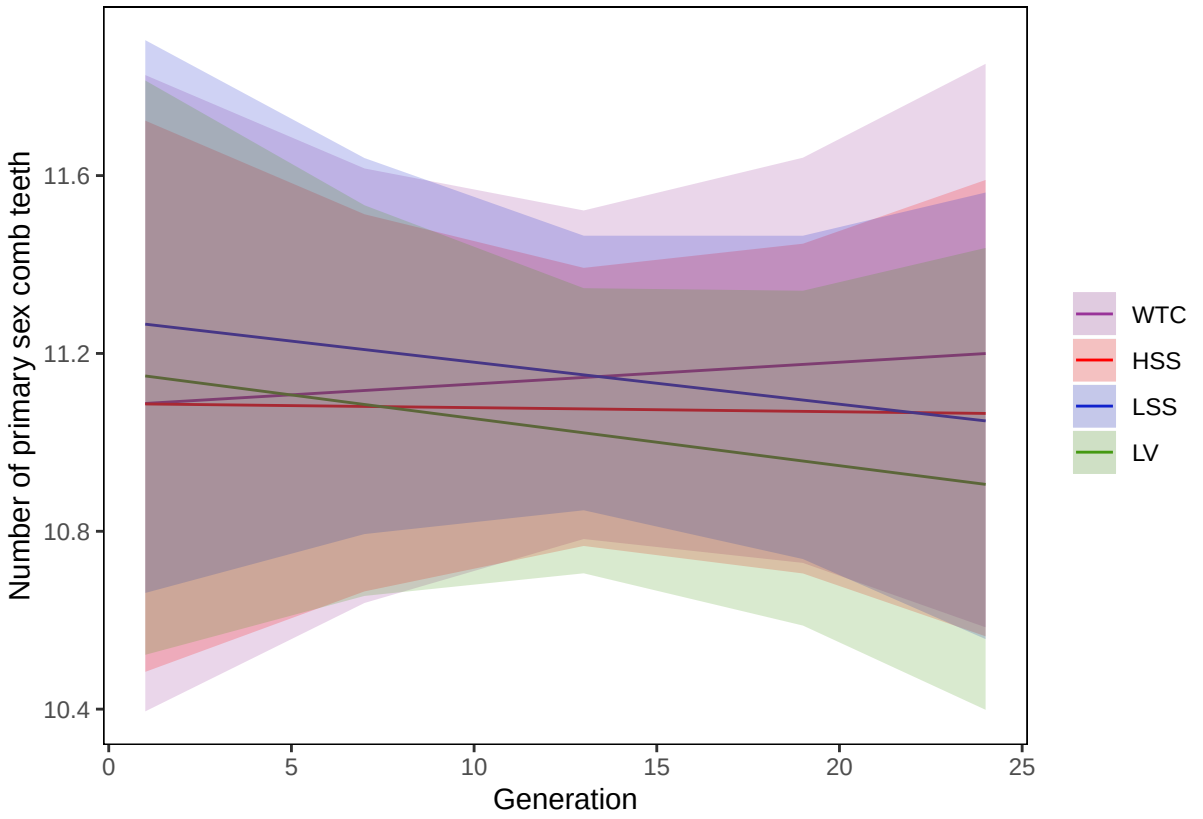
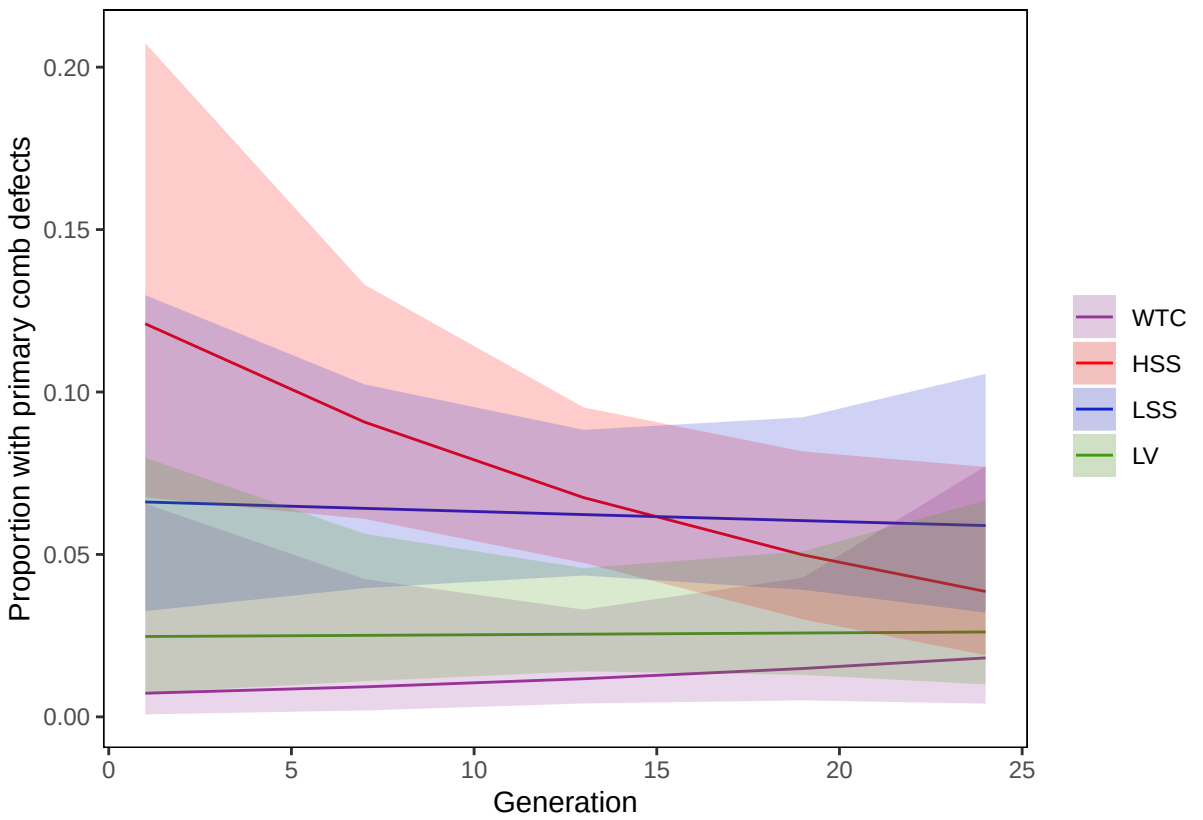

Frequency of scd^1 males
1.0
0.8
0.6
0.4
0.2
0.0
0
2
4
6
8
Generation

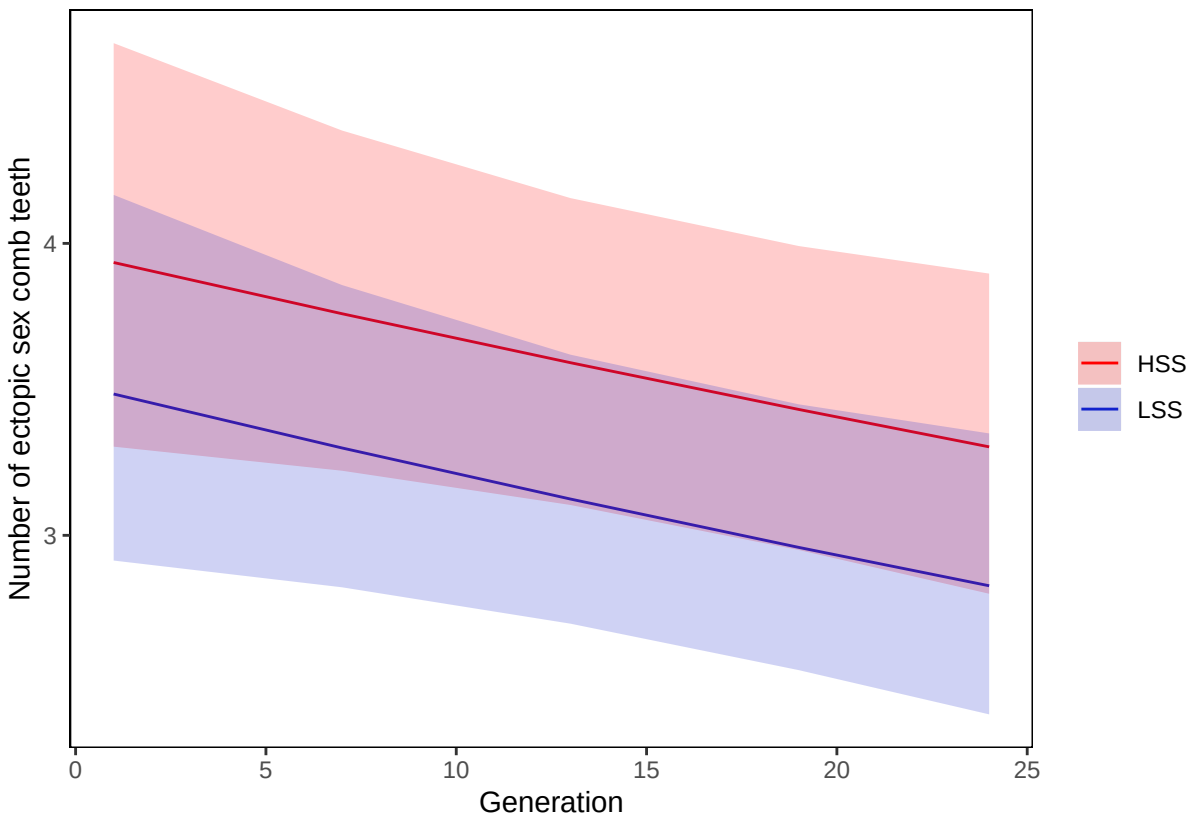
A


Number of primary sex comb teeth
11.6
11.2
10.8
10.4
0
5
10
15
20
25
Generation
WTC
HSS
LSS
LV

B


Proportion with primary comb defects
0.20
0.15
0.10
0.05
0.00
0
5
10
15
20
25
Generation
WTC
HSS
LSS
LV

C


Number of ectopic sex comb teeth
4
3
0
5
10
15
20
25
Generation
HSS
LSS