

## **Spatial arrangement drastically changes the neural representation of multiple visual stimuli that compete in more than one feature domain**

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## ABSTRACT

1           Natural scenes often contain multiple objects and surfaces. However, how neurons in the  
2 visual cortex represent multiple visual stimuli is not well understood. Previous studies have shown  
3 that, when multiple stimuli compete in one feature domain, the evoked neuronal response is biased  
4 toward the stimulus that has a stronger signal strength. Here we investigate how neurons in the  
5 middle temporal (MT) cortex of macaques represent multiple stimuli that compete in more than  
6 one feature domain. Visual stimuli were two random-dot patches moving in different directions.  
7 One stimulus had low luminance contrast and moved with high coherence, whereas the other had  
8 high contrast and moved with low coherence. We found that how MT neurons represent multiple  
9 stimuli depended on the spatial arrangement of the stimuli. When two stimuli were overlapping,  
10 MT responses were dominated by the stimulus component that had high contrast. When two  
11 stimuli were spatially separated within the receptive fields, the contrast dominance was abolished.  
12 We found the same results when using contrast to compete with motion speed. Our neural data and  
13 computer simulations using a V1-MT model suggest that the contrast dominance found with  
14 overlapping stimuli is due to normalization occurring at an input stage fed to MT, and MT neurons  
15 cannot overturn this bias based on their own feature selectivity. The interaction between spatially  
16 separated stimuli can largely be explained by normalization within MT. Our results revealed new  
17 rules on stimulus competition and highlighted the impact of hierarchical processing on  
18 representing multiple stimuli in the visual cortex.

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## SIGNIFICANCE STATEMENT

Previous studies have shown that the neural representation of multiple visual stimuli can be accounted for by a divisive normalization model. By using multiple stimuli that compete in more than one feature domain, we found that luminance contrast has a dominant effect in determining competition between multiple stimuli when they were overlapping but not spatially separated. Our results revealed that neuronal responses to multiple stimuli in a given cortical area cannot be simply predicted by the population neural responses elicited in that area by the individual stimulus components. To understand the neural representation of multiple stimuli, rather than considering response normalization only within the area of interest, one must consider the computations including normalization occurring along the hierarchical visual pathway.

## 30 **Introduction**

31

32 In natural scenes, multiple visual stimuli are often present in a local spatial region. While  
33 it is generally well understood how neurons in the visual cortex encode a single stimulus, how  
34 neurons encode multiple visual stimuli within their receptive fields (RFs) remains to be elucidated.  
35 Because visual perception depends critically on the integration and segregation of multiple visual  
36 stimuli (Braddick, 1993), understanding the neural representation of multiple stimuli is of  
37 significant importance.

38

39 The middle temporal (MT) cortex is an extrastriate brain area that is important for visual  
40 motion processing (Britten, 2003; Born and Bradley, 2005; Park and Tadin, 2018). Neurons in area  
41 MT receive feedforward inputs from direction-selective neurons in V1 (Movshon and Newsome,  
42 1996) and have RFs about ten times larger in size than those of V1 neurons at the same  
43 eccentricities (Gattass and Gross, 1981; Albright and Desimone, 1987). Previous studies have  
44 shown that neuronal responses in area MT elicited by multiple moving stimuli follow a sub-linear  
45 summation of the responses elicited by the individual stimulus components (Snowden et al., 1991;  
46 Qian and Andersen, 1994; Recanzone et al., 1997; Ferera and Lisberger, 1997; Britten and Heuer,  
47 1999; Heuer and Britten, 2002; McDonald et al., 2014), consistent with a model of divisive  
48 normalization (Simoncelli and Heeger, 1998; Britten and Heuer, 1999; Carandini and Heeger,  
49 2011).

50

51 Work in our laboratory has shown that the direction tuning curves of MT neurons to  
52 overlapping random-dot stimuli moving transparently in different directions can also be described  
53 as a weighted sum of the responses elicited by the individual stimulus components (Xiao et al.,  
54 2014; Xiao and Huang, 2015). When two stimulus components have different signal strengths in  
55 one feature domain, defined either by motion coherence or luminance contrast, MT neurons pool  
56 the stimulus component that has a stronger signal strength with greater weight (Xiao et al., 2014).  
57 The response bias in MT toward the stimulus component that has a stronger signal strength can be  
58 accounted for by a descriptive model of divisive normalization (Xiao et al., 2014), similar to the  
59 contrast normalization model used to describe neuronal responses in V1 (Carandini et al., 1997;  
60 Busse et al. 2009).

61

62           However, natural scenes contain multiple visual stimuli that often differ in more than one  
63 feature domain. For example, one stimulus may have a stronger signal strength in feature A but a  
64 weaker signal strength in feature B, whereas another stimulus may have a weaker signal strength  
65 in feature A but a stronger signal strength in feature B. In this case, it is unclear which stimulus  
66 has an overall stronger signal strength and, more generally, how visual stimuli with multiple  
67 competing features interact within neurons' RFs.

68  
69           One possibility is that, to neurons in a given brain area, the overall signal strength of a  
70 visual stimulus is reflected in the evoked responses of a population of neurons in that area. Due to  
71 divisive normalization within that area, a neuron may weigh a visual stimulus more strongly if the  
72 population neural response elicited by that stimulus is greater than the population response elicited  
73 by a competing stimulus. Alternatively, how neurons in a given brain area weigh multiple  
74 competing stimuli may be the result of neural computations occurring in multiple stages along the  
75 hierarchical visual pathway and may not be explained by simply considering the population neural  
76 responses elicited by the individual stimulus components in the area of interest.

77  
78           Here, we investigate the rule by which neurons in area MT encode multiple moving stimuli  
79 that compete in more than one feature domain. We found that MT responses to multiple stimuli  
80 changed drastically when the spatial arrangement of the visual stimuli was varied. Our results  
81 revealed how visual stimuli that differ in multiple feature domains interact within neurons' RFs  
82 and shed light on how the neuronal responses in a given cortical area are shaped by neural  
83 processing along the hierarchical visual pathway.

## 84 85 86 **Materials and Methods**

87  
88           Two male adult rhesus monkeys (*Macaca mulatta*) were used in the neurophysiological  
89 experiments. Experimental protocols were approved by the Institutional Animal Care and Use  
90 Committee of UW-Madison and conform to U.S. Department of Agriculture regulations and to the  
91 National Institutes of Health guidelines for the care and use of laboratory animals. Procedures for  
92 surgical preparation and electrophysiological recordings were routine and similar to those  
93 described previously (Xiao et al., 2015). A head post and a recording cylinder were implanted

94 during sterile surgery with the animal under isoflurane anesthesia. For electrophysiological  
95 recordings from neurons in area MT, we took a vertical approach and used tungsten electrodes (1-  
96 3 M $\Omega$ , FHC). We identified area MT by its characteristically large portion of directionally selective  
97 neurons, small RFs relative to those of neighboring medial superior temporal cortex (area MST),  
98 its location at the posterior bank of the superior temporal sulcus, and visual topography of the RFs  
99 (Gattass and Gross, 1981). Electrical signals were amplified and single units were identified with  
100 a real-time template-matching system and an offline spike sorter (Plexon). Eye position was  
101 monitored using a video-based eye tracker (EyeLink, SR Research) with a rate of 1000 Hz.

102

### 103 *Visual stimuli and experimental procedure*

104

105 Stimulus presentation and data acquisition were controlled by a real-time data acquisition  
106 program “Maestro” (<https://sites.google.com/a/srscicomp.com/maestro/home>). Visual stimuli  
107 were presented on a 25-inch CRT monitor at a viewing distance of 63 cm. Monitor resolution was  
108 1024  $\times$  768 pixels, with a refresh rate of 100 Hz. Stimuli were generated by a Linux workstation  
109 using an OpenGL application that communicated with an experimental control computer. The  
110 luminance of the video monitor was measured with a photometer (LS-110, Minolta) and was  
111 gamma-corrected.

112

113 Visual stimuli were achromatic random-dot patches presented within a circular aperture  
114 with a diameter of 3°. Individual dots were squares of 2 pixels extending 0.08° on each side, and  
115 each random-dot patch had a dot density of 2.7 dots/deg<sup>2</sup>. The dots had a luminance of either 79  
116 or 22 cd/m<sup>2</sup>, presented on a uniform background with a luminance of 10 cd/m<sup>2</sup>, which gives rise  
117 to a Michelson contrast of either 77.5% or 37.5%. Random dots in each patch moved within the  
118 stationary aperture in a specified direction. The motion coherence of each random-dot patch was  
119 set to either 100% or 60%. To generate a random-dot patch moving at N% of motion coherence  
120 (after Newsome and Pare 1988; Britten et al. 1992), N% of the “signal” dots were selected to move  
121 coherently, while the rest of the dots referred to as the “noise” dots were repositioned randomly  
122 within the aperture. Random selections of the “signal” and “noise” dots occurred at each monitor  
123 frame. Therefore, a given dot would switch back and forth between a signal dot and a noise dot.  
124 The lifetime of each dot was as long as the motion duration.

125

126           In each experimental trial, the monkey maintained fixation within a  $1^\circ \times 1^\circ$  electronic  
127 window around a small fixation point. After a neuron was isolated, we first characterized its  
128 direction selectivity by interleaving trials of a  $30^\circ \times 27^\circ$  random-dot patch, moving in different  
129 directions at a step of  $45^\circ$  and at a speed of  $10^\circ/\text{s}$ . The direction selectivity and preferred direction  
130 (PD) were determined on-line using MATLAB (MathWorks). We then characterized the speed  
131 tuning of the neuron using a random-dot patch moving at different speeds (1, 2, 4, 8, 16, 32, or  
132  $64^\circ/\text{s}$ ) in the neuron's PD. Using a cubic spline, the preferred speed (PS) of the neuron was taken  
133 as the speed that evoked the highest firing rate in the fitted speed tuning curve. Next, we used a  
134 series of  $5^\circ \times 5^\circ$  random-dot patches moving in the PD and at the PS of the neuron to map the  
135 neuron's RF. The location of the patch was randomized and the screen was tiled in  $5^\circ$  steps. The  
136 RF map was interpolated at  $0.5^\circ$  intervals, and the location giving rise to the highest firing rate  
137 was taken as the center of the RF.

138

139           In the main experiments, the visual stimuli appeared after the monkey maintained fixation  
140 for 200 ms. To separate the neuronal responses to the stimulus motion from those due to the  
141 stimulus onset, the visual stimuli were first turned on and remained stationary for 200 ms before  
142 they started to move for 500 ms. The visual stimuli were then turned off. The monkeys maintained  
143 fixation for an additional 200 ms after the stimulus offset. In some stimulus trials, two random-dot  
144 patches that moved in different directions, referred to as two stimulus components, were presented  
145 simultaneously. The direction separation between two stimulus components was fixed at  $90^\circ$ . We  
146 varied the vector averaged (VA) direction of the bi-directional stimulus around  $360^\circ$  to  
147 characterize the response tuning curve. The two stimulus components were either overlapping in  
148 one of two locations (site a or b) within the RF, or they were spatially separated within the RF, one  
149 centered at site a and the other at site b, with at least  $1^\circ$  gap between the borders of the two random-  
150 dot patches (illustrated in Fig. 1). In other trials, only one stimulus component was presented at  
151 either site a or site b and the direction was varied to characterize the tuning curve to the stimulus  
152 component. For the majority of the experiments, the VA and component directions were varied in  
153 a step of  $15^\circ$ . In a small set of experiments, the directions were varied in a step of  $30^\circ$ . The trials  
154 presenting bi-directional stimuli and individual stimulus components were randomly interleaved.

155

156           In the first experiment, one random-dot patch, referred to as the “low contrast & high  
157 coherence” component, had a luminance contrast of 37.5% and a motion coherence of 100%. The  
158 other random-dot patch, referred to as the “high contrast & low coherence” component, had a  
159 luminance contrast of 77.5% and a motion coherence of 60%. Both stimulus components moved  
160 at the same speed, which was set at the neuron’s PS if it was below 10°/s, or at 10°/s if the PS was  
161 at or greater than 10°/s. Note that when a random-dot patch moved at 60% coherence in a given  
162 direction, the visual stimulus was different from a situation where 60% of the dots always moved  
163 coherently and the rest of the 40% of dots always moved randomly. Because the random selection  
164 of signal and noise dots occurred at each monitor frame in our stimuli, a noise dot at one frame  
165 may turn into a signal dot in the next frame and move in the coherent direction. Perceptually, it is  
166 difficult to segregate the noise dots from the signal dots of the same stimulus component. The  
167 noise dots of the “high contrast & low coherence component” are not an independent entity and  
168 do not appear to interfere with the coherence of the “low contrast & high coherence” component  
169 perceptually.

170  
171           In the second experiment, we set the motion coherence of both random-dot patches to 100%  
172 but used different speeds for the two stimulus components. One random-dot patch, referred to as  
173 the “low contrast & faster speed” component, had a luminance contrast of 37.5% and moved at  
174 10°/s. The other random-dot patch, referred to as the “high contrast & slower speed” component,  
175 had a luminance contrast of 77.5% and moved at 2.5°/s.

## 176 177 Data analysis

178  
179           Response firing rate was calculated during the period of 500-ms stimulus motion and  
180 averaged across repeated trials. We fitted the raw direction tuning curves for the bi-directional  
181 stimuli and the individual stimulus components using splines at a resolution of 1°. We then  
182 rotated the spline-fitted tuning curve to the bi-directional stimuli so that the VA direction of 0°  
183 was aligned with the PD of each neuron. In the first experiment, the responses of each neuron to  
184 the bi-directional stimuli and individual stimulus components were normalized by the maximum  
185 response to the “low contrast & high coherence” component. In the second experiment, the  
186 responses of each neuron were normalized by the maximum response to the faster speed



187 component. We averaged the rotated and normalized tuning curves across neurons to obtain  
188 population-averaged tuning curves.

189

190 To quantify the relationship between the responses elicited by the bi-directional stimuli  
191 and those elicited by the individual stimulus components, we fitted the direction tuning curves  
192 using a summation plus nonlinear interaction (SNL) model (Eq. 1), which has been shown to  
193 provide a better fit of MT responses elicited by bi-directional stimuli than a linear weighted  
194 summation model (Xiao et al., 2014).

195

$$196 \quad R_{pred}(\theta_1, \theta_2) = w_1 R_1(\theta_1) + w_2 R_2(\theta_2) + b R_1(\theta_1) R_2(\theta_2), \quad (1)$$

197

198 where  $R_{pred}$  is the response to the bi-directional stimuli predicted by the model;  $\theta_1$  and  $\theta_2$  are the  
199 two component directions;  $R_1$  and  $R_2$  are the measured component responses elicited by the two  
200 stimulus components when presented alone;  $w_1$  and  $w_2$  are the response weights for  $R_1$  and  $R_2$ ,  
201 respectively; and  $b$  is the coefficient of multiplicative interaction between the component  
202 responses. To determine whether the response elicited by the bi-directional stimuli showed a  
203 significant bias toward one of the two stimulus components, we compared the response weights  
204  $w_1$  and  $w_2$  using either a paired t-test or a Wilcoxon signed-rank test.

205

206 We also fitted the response tuning curves to the bi-directional stimuli using a few variants  
207 of a divisive normalization model (Carandini and Heeger, 2011) (see Results). The model fits were  
208 obtained using the constrained minimization tool ‘fmincon’ (MATLAB) to minimize the sum of  
209 squared error.

210

211 To evaluate the goodness of fit of a model for the response tuning curve to the bi-directional  
212 stimuli, we calculated the percentage of variance (PV) accounted for by the model as:

213

$$214 \quad PV = 100 \times \left( 1 - \frac{SSE}{SST} \right), \quad (2)$$

215

216 where SSE is the sum of squared errors between the model fit and the neuronal data, and SST is  
217 the sum of squared differences between the data and the mean of the data (Morgan et al., 2008).

218 V1-MT Model

219

220 We adapted a computational model proposed by Simoncelli and Heeger (1998)  
221 (<http://www.cns.nyu.edu/~lcv/MTmodel/>) to reconstruct our visual stimuli and to simulate the  
222 neuronal response tuning to the bi-directional stimuli that were either overlapping or spatially  
223 separated. The model contained several consecutive stages, which can be interpreted as V1 simple,  
224 V1 complex, and MT (Simoncelli and Heeger, 1996; Rust et al., 2006). Based on the dimensions  
225 of video monitor and viewing distance in our neurophysiological experiments,  $1^\circ$  of visual angle  
226 corresponds to 21 pixels. The random-dot patch in our model simulations had a circular aperture  
227 with a diameter of 63 pixels (i.e.  $3^\circ$ ) and the same dot density as used in our experiments. Each  
228 dot had a size of  $2 \times 2$  pixels.

229

230 We set the RFs of model neurons by Gaussian convolutional filters (Table 1). We estimated  
231 the size of the RF for each neuron type by summing the lengths of the incorporated filters. For the  
232 spatially-separated stimuli, we set a blank gap between the two stimulus components as the RF  
233 size of the V1 complex neuron, which is  $1.2^\circ$ , to ensure that no V1 neuron would be driven by  
234 both stimulus components. We generated direction-selective neuron populations that  
235 approximately tiled a sphere in the frequency domain. We tuned the contrast response functions  
236 by adjusting  $C_{50}$  values for V1 and MT neurons. These  $C_{50}$  values were represented in the model  
237 as  $\sigma^2$  in the normalization equation (Eq. 3), which was applied to both V1 complex cell and MT  
238 stages of the model (adapted from Simoncelli and Heeger 1998 and Rust et al., 2006).

239

$$240 \quad R'_n(t) = \frac{|R_n(t)|}{K \sum_m |w_m \cdot R_m(t)| + \sigma^2}, \quad (3)$$

241

242 where  $R_n(t)$  represents the  $n$ th neuron's linear filter response;  $R'_n(t)$  represents the normalized  
243 response of either V1 complex cell or MT neuron;  $| \cdot |$  denotes half-wave rectification;  $K$  represents  
244 the strength of normalization, which was set as  $1 - \sigma^2$ ;  $m$  represents the  $n$ th neuron's normalization  
245 pool;  $w$  represents the Gaussian spatial weighting profile of the normalization pool, with a standard  
246 deviation of  $SD_{norm}$ . The model parameters for V1 and MT stages are defined in Table 1. We fitted  
247 the model contrast response functions to neural data from V1 and MT as described in Sclar et al.  
248 (1990). Similarly, we tuned coherence responses by varying the spatial scale of the normalization

249 pool ( $m$ ), the weighting profile within the pool ( $w$ ), and the size of the V1 linear RF. The MT  
 250 coherence response function was fitted to data replotted from Figure 1C in Britten and Newsome  
 251 (1998). We are not aware of published neural data on V1 coherence response function. So the  
 252 parameters for V1 model neurons were varied to simulate our MT responses to bi-directional  
 253 stimuli without a constraint on V1 coherence response function. The same model parameters were  
 254 used for the overlapping and spatially separated conditions.

255

256 **Table 1.** Model parameters for V1 and MT neurons

257

Model parameters	V1 stage	MT stage
RF size (pixels)	15 (simple cell) 25 (complex cell)	211
Standard Deviation (SD) of Gaussian RF profile (pixels)	0.73/2.19/6.57 (3 scales for simple cells) 7 (complex cell)	53
$\sigma^2 (C_{50})$	0.0016	0.000049
Size of normalization pool $m$ (pixels)	38	153
$SD_{norm}$ (pixels)	5	52
Baseline activity	0	0.1

258

259

260 We explored several variants of the model architecture. The model parameters were fitted  
 261 after each architectural manipulation. The following changes enabled the model to better capture  
 262 the trends of the stimulus competition found in our neural data. First, we used area-normalized  
 263 Gaussian functions to set the weights for the spatial pooling and local population normalization.  
 264 Second, multiple frequency scales for V1 simple cells were computed by tripling the standard  
 265 deviation of the underlying 3<sup>rd</sup> order derivative Gaussian, similar to the doubling suggested in  
 266 Simoncelli and Heeger (1998) – this change was made after spectral analysis of stimuli showed  
 267 that a wider range of scales was necessary to capture motion at lower coherence. Third, V1 afferent  
 268 weights were not adjusted to zero mean, allowing MT neurons to have variable proportions of  
 269 positive and negative inputs. Finally and importantly, rectification and static nonlinearity were  
 270 applied to the MT stage after spatial pooling and before normalization, which is physiologically  
 271 plausible and provides a better fit of our neural data.

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275

## 276 **Results**

277

278 We asked the question of how neurons in extrastriate area MT represent multiple visual  
279 stimuli that compete in more than one feature domain. To address this question, we conducted  
280 neurophysiological experiments and computer simulations. We recorded from isolated single  
281 neurons in area MT of two macaque monkeys while they performed a fixation task. Visual stimuli  
282 were two random-dot patches moving simultaneously in different directions within the RFs. In the  
283 first experiment, we used luminance contrast and motion coherence as two competing features.  
284 One stimulus had high contrast but moved with low coherence, whereas the other stimulus had  
285 low contrast but moved with high coherence (see Methods). We manipulated the spatial  
286 arrangement of the visual stimuli to investigate the contributions of earlier visual areas and area  
287 MT in mediating the competition between multiple stimuli. In a second experiment, we used  
288 luminance contrast and motion speed as two competing features. We first present the results from  
289 the neurophysiological experiments and then computer simulations.

290

### 291 Neurophysiological experiments

292

293 We measured the direction tuning curves of MT neurons in response to two stimuli that  
294 had competing visual features and moved simultaneously in different directions. Our dataset  
295 includes recordings from 76 MT neurons, 43 from monkey G and 33 from monkey B. We set the  
296 angular separation between the motion directions of two individual stimuli, referred to as the  
297 stimulus components, at 90° and varied the VA direction of the stimuli. In the first experiment,  
298 one stimulus component had a low contrast of 37.5% and moved at a high motion coherence of  
299 100%. The other component had a high contrast of 77.5% and moved at a low coherence of 60%.  
300 Figure 1 shows the direction tuning curves of two representative neurons. The red curve shows the  
301 neuronal response elicited when both stimulus components were present, as a function of the VA  
302 direction of the two stimulus components. The green and blue curves show the neuronal responses  
303 elicited by the individual stimulus components when presented alone. The tuning curves of the  
304 component responses are arranged such that, at each VA direction, the data points on the green  
305 and blue curves correspond to the responses elicited by the individual stimulus components of that  
306 VA direction (note the color-coded abscissas for the component directions in Fig. 1A2).

307

308 For the two example neurons, the peak response of the direction tuning curve to the “low  
309 contrast & high coherence” component alone (shown in blue) was greater than that to the “high  
310 contrast & low coherence” component (shown in green) (Fig. 1). This is expected since MT  
311 neurons are sensitive to motion coherence within a large coherence range (Britten et al., 1993),  
312 whereas their contrast response function saturates at a low luminance contrast (Sclar et al., 1990).  
313 Consequently, the average of the response tuning curves to the two stimulus components (shown  
314 in gray) was biased toward the “low contrast & high coherence” component. Surprisingly, we  
315 found that when the two stimulus components were overlapping, the neuronal responses elicited  
316 by the bi-directional stimuli were strongly biased toward the “high contrast & low coherence”  
317 component (Fig. 1A). This response bias was robust and occurred when we placed the overlapping  
318 stimuli at a different site within the RF (Fig. 1B).

319  
320 Two overlapping visual stimuli could stimulate not only the RFs of single MT neurons but  
321 also the RFs of single V1 neurons. The response bias toward the “high contrast & low coherence”  
322 component may be caused by the neural processes within area MT, or alternatively inherited from  
323 earlier visual areas such as V1. To determine the contribution of earlier visual areas to the response  
324 bias, we placed two stimulus components at different locations within the RF of a given MT  
325 neuron. The two stimulus components were separated by a gap of at least  $1^\circ$  (illustrated in Fig.  
326 1C). With this spatial arrangement, the RF of a single V1 neuron could only be stimulated by one  
327 of the two stimulus components, whereas the RF of an MT neuron could still be stimulated by both  
328 components. We found that the response tuning to the bi-directional stimuli changed drastically  
329 when stimulus components were spatially separated. MT responses elicited by the bi-directional  
330 stimuli no longer showed a bias toward the “high contrast & low coherence” component, but  
331 roughly followed a scaled average of the component responses (Fig. 1C).

332  
333 Figure 2 shows the tuning curves averaged across 70 MT neurons. The population-  
334 averaged response elicited by the “low contrast & high coherence” component moving in the PD  
335 of each neuron, aligned to  $0^\circ$ , was significantly greater than that elicited by the “high contrast &  
336 low coherence” component moving in the PD (one-tailed paired t-test,  $p = 4.1 \times 10^{-7}$ ). However,  
337 when the two stimuli were overlapping, the population response elicited by the bi-directional  
338 stimuli was almost completely biased toward the weaker “high contrast & low coherence”

339 component, regardless of the spatial location within the RF (Fig. 2A and 2B). The bias toward the  
340 “high contrast & low coherence” component at a given VA direction was in a manner of “higher-  
341 contrast-take-all”. For example, at a VA direction of 45° where the “low contrast & high  
342 coherence” component moved in the PD (0°) and the “high contrast & low coherence” component  
343 moved in 90° (indicated by a dotted line in Fig. 2A), the bi-directional response closely followed  
344 the much weaker response elicited by the “high contrast & low coherence” component. When the  
345 two stimulus components were spatially separated within the RF, the strong bias toward the “high  
346 contrast & low coherence” component was abolished (Fig. 2C). The population response to the  
347 bi-directional stimuli now showed roughly equal weighting of the responses elicited by the  
348 individual stimulus components.

349  
350 The SNL model (see Eq. 1 in Methods) provided an excellent fit of the MT responses  
351 elicited by the bi-directional stimuli, illustrated by the black curves in Figure 1. Across our neuron  
352 population the model fit accounted for, on average, 83% of the response variance (see Methods).  
353 Figure 3 compares the response weights for the two stimulus components obtained from the SNL  
354 model fits. In the overlapping condition, the mean response weight  $w_2$  for the “high contrast & low  
355 coherence” component was significantly greater than the weight  $w_1$  for the “low contrast & high  
356 coherence” component (one-tailed paired t-test,  $p = 1.9 \times 10^{-45}$  for *site a*,  $p = 2.5 \times 10^{-28}$  for *site b*)  
357 (Fig. 3A). Nearly all data points, each representing the result from one neuron, were below the  
358 unity line. The mean response weight for the “high contrast & low coherence” component was  
359 0.97 (std = 0.24), whereas the mean weight for the “low contrast & high coherence” component  
360 was 0.23 (std = 0.25), indicating a dominant effect of the “high contrast & low coherence”  
361 component in determining the neuronal response to the bi-directional stimuli.

362  
363 When the two stimulus components were spatially separated within the RF, the response  
364 weights changed significantly, becoming symmetrically distributed relative to the unity line (Fig.  
365 3B). The spread of weights in the spatially-separated condition is larger than that in the overlapping  
366 condition. The mean weight for the “high contrast & low coherence” component decreased to 0.66  
367 (std = 0.32), whereas the mean weight for the “low contrast & high coherence” component  
368 increased to 0.68 (std = 0.43). The mean weights for the two components were no longer different

369 (paired t-test,  $p = 0.8$ ) but were significantly greater than 0.5 of response averaging (t-test,  $p <$   
370 0.001).

371

372 To quantify the response bias toward an individual stimulus component, we calculated a  
373 bias index (BI) :

374

$$375 \quad BI = (w_2 - w_1)/(w_2 + w_1) \quad (4)$$

376

377 A positive value of the index indicates a bias toward the “high contrast & low coherence”  
378 component. Figure 3C shows how this bias index changes with the spatial arrangement of the  
379 visual stimuli. In the overlapping condition, the mean BI is 0.73 (std = 0.23), which is significantly  
380 greater than 0 (one-tailed t-test,  $p = 7.5 \times 10^{-35}$ ). In the spatially-separated condition, the mean BI is  
381 -0.01 (std = 0.95), which is not significantly different from 0 ( $p = 0.7$ ). The mean BI obtained in  
382 the overlapping condition is significantly greater than that in the spatially-separated condition  
383 (one-tailed paired t-test,  $p = 4.7 \times 10^{-9}$ ), indicating a change of the response bias when the spatial  
384 arrangement of the visual stimuli is altered.

385

386 We previously found that the tuning curves of some MT neurons to overlapping bi-  
387 directional stimuli can show a directional “side-bias” toward one of the two direction components  
388 (Xiao and Huang, 2015). A subgroup of neurons prefers the stimulus component at the clockwise  
389 side of two motion directions, whereas another group prefers the component direction at the  
390 counter-clockwise side. These response biases can occur even when both stimulus components  
391 have the same contrast and coherence. In the experiment shown in Figures 1-3, the “high contrast  
392 & low coherence” component always moved at the counter-clockwise side direction (Fig. 2A, 2B).  
393 Could the strong bias toward the “high contrast & low coherence” component in the overlapping  
394 condition be due to a biased neuron sample that happened to have a strong bias toward the direction  
395 component at the counter-clockwise side? To address this concern, we arranged the direction  
396 components differently.

397

398 Figure 4A and B show the averaged direction tuning curves of 15 MT neurons when the  
399 direction of the “high contrast & low coherence” component was placed at the counter-clockwise

400 side under the overlapping and spatially separated conditions, as in Figure 2. When the “high  
401 contrast & low coherence” component was placed at the clockwise side of the two component  
402 directions, the responses of the same 15 neurons to the bi-directional stimuli still showed a strong  
403 bias toward the “high contrast & low coherence” component under the overlapping condition (Fig.  
404 4C), and showed roughly equal weighting of the two components under the spatially-separated  
405 condition (Fig. 4D). Placing the “high contrast & low coherence” component at the clockwise or  
406 counter-clockwise side of the two component directions had no effect on the response bias, as  
407 measured by the bias index under the overlapping and spatially-separated conditions (Wilcoxon  
408 rank-sum test,  $p = 0.6$ ).

409 To shed light on the neural mechanisms underlying the response bias, we examined the  
410 timecourse of the neuronal responses in the overlapping and spatially separated conditions. Figure  
411 5 shows the PSTHs calculated using a 10-ms time bin when either the “high contrast & low  
412 coherence” component or the “low contrast & high coherence” component moved in the PD.  
413 When stimuli were overlapping, as soon as MT neurons started to respond to the onset of the static  
414 stimuli (see Methods), the response elicited by both stimulus components already closely followed  
415 the “high contrast & low coherence” component, even before the onset of the stimulus motion  
416 (Fig. 5A, B). After the onset of the motion response, the neuronal response to the bi-directional  
417 stimuli continued to follow the response elicited by the “high contrast & low coherence”  
418 component throughout the motion period, regardless of whether the component moved in the PD  
419 and elicited a strong response (Fig. 5A), or 90° away from the PD and elicited a weak response  
420 (Fig. 5B). Since the strong bias towards the “high contrast & low coherence” component in the  
421 overlapping condition occurred at the very beginning of the stimulus onset, it is unlikely that the  
422 bias was due to selective attention (see Discussion).

423  
424 When stimuli were spatially separated, MT neurons also followed the “high contrast & low  
425 coherence” component in response to the onset of the static stimuli (Fig. 5C, D). After the motion  
426 onset, when the “high contrast & low coherence” component moved in the PD, the motion response  
427 elicited by the bi-directional stimuli initially followed the “high contrast & low coherence”  
428 component for ~30 ms, and was then “pulled down” by the non-PD component (see the arrow in  
429 Fig. 5C). When the “high contrast & low coherence” component moved in the non-PD, the motion



430 response elicited by the bi-directional stimuli followed the “high contrast & low coherence”  
431 component for ~10 ms after the onset of the motion response to the PD component, and was then  
432 “pulled up” by the PD component (see the arrow in Fig. 5D). These results suggest that response  
433 normalization under the spatially separated condition takes 10~30 ms to occur.

434  
435         When two stimulus components overlap, the random dots from each component constitute  
436 only half of the total number of dots of the two moving surfaces. Could the strong response bias  
437 toward the “high contrast & low coherence” component be due to a reduction of the motion  
438 coherence of the “low contrast & high coherence” component when the stimuli overlapped? We  
439 think this is an unlikely explanation because overlapping reduces the percentage of the signal dots  
440 relative to the total number of dots for both stimulus components. In addition, our stimuli moved  
441 in two directions separated by 90°. Human observers can reliably segregate the two stimulus  
442 components at this angle separation and the “low contrast & high coherence” component still  
443 appears to move coherently. Overlapping does not change the relative coherence levels nor the  
444 perceived coherence of the two stimulus components. When overlapping random-dot stimuli have  
445 the same luminance contrast but move at different motion coherences, macaque MT response to  
446 both stimulus components is biased toward the high coherence component (Xiao et al., 2014),  
447 indicating that stimulus overlapping does not prevent the response bias toward the high coherence  
448 component given equal contrast.

449  
450         To determine whether the dominance by the high-contrast component on MT responses  
451 elicited by overlapping stimuli occurs only when luminance contrast and motion coherence  
452 compete with each other, we conducted a second experiment using visual stimuli that differ in  
453 luminance contrast and motion speed. We previously found that when two overlapping random-  
454 dot patches moved in the same direction at different speeds, within a range of low to intermediate  
455 speeds, the responses of MT neurons elicited by the bi-speed stimuli was biased toward the faster  
456 speed component (X. Huang et al., unpublished data). Motivated by this finding, we used motion  
457 speed to compete with luminance contrast. As in the main experiment, the visual stimuli contained  
458 two random-dot patches moving in two directions separated by 90° and we varied the VA direction  
459 to measure the direction tuning curves. One stimulus component had a high luminance contrast of  
460 77.5% and moved at a slower speed of 2.5°/s. The other stimulus component had a low luminance

461 contrast of 37.5% and moved at a faster speed of  $10^\circ/\text{s}$ . Both stimulus components moved at 100%  
462 coherence and were either overlapping or spatially-separated within the RF of a given MT neuron  
463 as in the first experiment. We also measured the direction tuning curves when the two stimulus  
464 components both had high luminance contrast (77.5%) and moved at  $2.5^\circ/\text{s}$  and  $10^\circ/\text{s}$ , respectively,  
465 at 100% coherence.

466

467 We recorded from 13 MT neurons using these visual stimuli. Figure 6 shows the  
468 population-averaged tuning curves. When both stimulus components had high contrast, the peak  
469 response elicited by the faster ( $10^\circ/\text{s}$ ) stimulus component moving in the PD (i.e.  $0^\circ$ ) was greater  
470 than that elicited by the slower ( $2.5^\circ/\text{s}$ ) component moving in the PD. The component responses  
471 are shown in green and purple in Figure 6A. When the two stimulus components were overlapping,  
472 the tuning curve elicited by both stimulus components (shown in red) is biased toward the faster  
473 stimulus component, more than what is predicted by the average of the component responses  
474 (shown in gray) (Fig. 6A). We fitted the direction tuning curves using the SNL model for each  
475 neuron (Eq. 1). The median response weight obtained by the model fit for the faster stimulus  
476 component (0.88) was significantly greater than the median weight (0.41) for the slower  
477 component (Wilcoxon signed-rank test,  $p = 7.3 \times 10^{-4}$ ). This result extended our previous finding  
478 of the response bias toward the faster stimulus component for stimuli moving in the same direction  
479 (unpublished results) to stimuli moving in different directions.

480

481 When the overlapping stimuli moving at different speeds had different luminance contrasts,  
482 the responses elicited by both stimulus components showed a strong bias toward the “high contrast  
483 & slower speed” component, even though the peak response to this component alone was  
484 significantly weaker than that to the “low contrast & faster speed” component (Fig. 6B). We found  
485 the same result when the two stimulus components overlapped at a different site within the RF  
486 (Fig. 6C). Under the overlapping condition, the median response weight for the “high contrast &  
487 slower speed” component was 0.81, which was significantly greater than the median weight for  
488 the “low contrast & faster speed” component (0.17) (Wilcoxon signed-rank test,  $p = 2.4 \times 10^{-4}$ ).  
489 Separating the two stimulus components spatially within the RF abolished the bias toward the  
490 “high contrast & slower speed” component (Fig. 6D). As the spatial arrangement of the stimulus  
491 components changed from overlapping to spatially separated, the median bias index (Eq. 4)

492 decreased significantly from 0.65 to -0.08 (Wilcoxon signed-rank test,  $p = 0.0012$ ). These results  
493 confirmed that luminance contrast has a dominant effect on MT responses elicited by overlapping  
494 stimuli, which is not unique to the competition between contrast and motion coherence. The spatial  
495 arrangement of visual stimuli can substantially change the competition between multiple stimuli  
496 within the RF.

497

#### 498 Fitting response tuning curve using the normalization model

499

500 Previous studies have shown that neuronal responses elicited by multiple stimuli in many  
501 brain areas can be described by a divisive normalization model (Carandini and Heeger, 2011). We  
502 asked whether our results could also be accounted for by response normalization. We first fitted  
503 the data using the following equation:

504

$$505 \quad R_{pred}(\theta_1, \theta_2) = \frac{S_1^n}{S_1^n + S_2^n + \sigma} R_1(\theta_1) + \frac{S_2^n}{S_1^n + S_2^n + \sigma} R_2(\theta_2) + c, \quad (5)$$

506

507 where  $R_1$  and  $R_2$  are the evoked direction tuning curves to the two stimulus components 1 and 2,  
508 respectively.  $\theta_1$  and  $\theta_2$  are the component directions.  $S_1$  and  $S_2$  represent the signal strengths of the  
509 “low contrast & high coherence” component and the “high contrast & low coherence” component,  
510 respectively.  $R_{pred}$  is the model-predicted response elicited by both stimulus components presented  
511 simultaneously.  $n$ ,  $\sigma$ , and  $c$  are model parameters with the constraints of  $n \geq 1$  and  $c > 0$ . Equations  
512 of the similar form have been used previously to describe normalization involving contrast, in  
513 which case the signal strength is simply the luminance contrast (Carandini et al., 1997; Busse et  
514 al., 2009; Xiao et al., 2014; Bao and Tsao, 2018). Since our visual stimuli competed in more than  
515 one feature domain, it was not obvious which stimulus component had an overall stronger signal  
516 strength. Because the brain has to make an inference of the signal strength based on the elicited  
517 neural responses, we assumed that the signal strength of a stimulus component, in the “eye” of MT  
518 neurons, is reflected in the neural responses elicited by that stimulus component moving in a fixed  
519 direction summed across a population of MT neurons that have different PDs evenly spanning  
520 360°. This summed population response is invariant to the direction of the stimulus component,  
521 which is suitable for representing signal strength. Equivalently, the summed population neural  
522 response in MT can be approximated by summing the responses of each neuron elicited by

523 stimulus component  $i$  moving in different directions spanning  $360^\circ$  and averaged across neurons  
524 in our data sample, e.g. to sum the population-averaged component responses across directions in  
525 Figure 2. This was how we calculated  $S_i$ , ( $i=1, 2$ ).

526  
527 This normalization model (Eq. 5) failed to capture the response tuning to overlapping bi-  
528 directional stimuli, accounting for only 33% of the response variance (34% for site  $a$ , 32% for site  
529  $b$ ). The model performed better when stimuli were separated, accounting for 66% of the variance.  
530 We found similar results when using this model to fit the data from our second experiment, in  
531 which luminance contrast competed with motion speed. The model accounted for an average of  
532 44% of the response variance (38% for site  $a$ , 50% for site  $b$ ) when stimuli were overlapping, and  
533 77% of the variance when stimuli were separated (Table 2).

534  
535 It has been suggested that response normalization can be tuned, such that individual  
536 stimulus components contribute differently to normalization (Ni et al., 2012; Rust et al., 2006; also  
537 see Carandini et al., 1997). We therefore fitted our data using a tuned normalization equation:

538  
539 
$$R_{pred}(\theta_1, \theta_2) = \frac{S_1^n}{S_1^n + \alpha S_2^n + \sigma} R_1(\theta_1) + \frac{S_2^n}{S_1^n + \alpha S_2^n + \sigma} R_2(\theta_2) + c, \quad (6)$$

540  
541 where  $\alpha$  is a positive parameter that scales the contribution of  $S_2$  with respect to  $S_1$  to normalization.  
542 We found that introducing tuned normalization did not improve the model performance at all when  
543 stimuli were overlapping, accounting for an average of 33% of the response variance (34% for site  
544  $a$ , 32% for site  $b$ ). When stimuli were separated, the tuned normalization model accounted for 68%  
545 of the variance. We found the same results when fitting the data collected when contrast competed  
546 with speed (Table 2).

547  
548 The poor fit of the responses under the overlapping condition by the standard normalization  
549 model (Eq. 5) can be understood because MT neurons showed a very strong bias toward the high  
550 contrast component, whereas  $S_1$  and  $S_2$  were similar. The tuned normalization was not able to  
551 improve the fit because, although it changed the relative contributions of the stimulus components  
552 to the normalization pool in the denominator, it kept the numerators in Equation 6 unchanged.  
553 Hence the relative weights for the two stimulus components did not change. To capture the strong

554 bias toward the high contrast component in the overlapping condition, a weighting parameter is  
 555 needed in the numerator. Accordingly, we fitted our results using the following equation:

556

$$557 \quad R_{pred}(\theta_1, \theta_2) = \frac{s_1^n}{s_1^n + \beta s_2^n + \sigma} R_1(\theta_1) + \frac{\beta s_2^n}{s_1^n + \beta s_2^n + \sigma} R_2(\theta_2) + c, \quad (7)$$

558

559 where  $\beta$  is a positive parameter and appears in both the numerator and the denominator. This  
 560 parameter allows the relative response weights for the two stimulus components to vary. When  $\beta$   
 561 is greater than one, the response weight for the high contrast component ( $R_2$ ) is greater than that  
 562 for the low contrast component ( $R_1$ ). As expected, this equation fitted the data well, accounting for  
 563 >80% of the response variance for both the overlapping and spatially separated conditions (Table  
 564 2). However, the normalization model itself does not provide an explanation for why the response  
 565 weight is greater for the high contrast component in the overlapping condition but not in the  
 566 spatially separated condition.

567

568 **Table 2.** Fitting the direction tuning curves using the normalization model

569

Visual Stimuli	$S_1$	$S_2$	Percentage of Variance Accounted for (mean $\pm$ std)		
			Normalization (Eq. 5)	Tuned Normalization (Eq. 6)	Normalization with weighted Numerators (Eq. 7)
Contrast vs. Coherence (N = 70)	Low contrast & high coherence	High contrast & low coherence			
Overlapping (site a)	122.3	126.0	34 $\pm$ 18	34 $\pm$ 18	86 $\pm$ 16
Overlapping (site b)	128.3	130.9	32 $\pm$ 19	32 $\pm$ 19	81 $\pm$ 19
Spatially Separated	130.4	130.3	66 $\pm$ 24	68 $\pm$ 25	83 $\pm$ 17
Contrast vs. Speed (N = 13)	Low contrast & faster speed	High contrast & slower speed			
Overlapping (site a)	128.1	83.6	38 $\pm$ 21	39 $\pm$ 21	88 $\pm$ 14
Overlapping (site b)	113.3	81.3	49 $\pm$ 20	49 $\pm$ 20	84 $\pm$ 15
Spatially Separated	128.1	81.3	77 $\pm$ 20	77 $\pm$ 20	90 $\pm$ 5

570

571

572 Computer simulations using a V1-MT model

573

574 Our spatially separated visual stimuli fall inside the RFs of single MT neurons, whereas  
 575 only one of the stimulus components would fall inside the RFs of single V1 neurons. Hence, our

576 spatially-separated visual stimuli can interact within the RFs of MT neurons but not V1 neurons.  
577 In contrast, the overlapping stimuli can interact within the RFs of both MT and V1 neurons. To  
578 explore the neural mechanisms underlying our physiological findings, we conducted computer  
579 simulations using a hierarchical feedforward model adapted from Simoncelli and Heeger (1998).  
580 This model consists of two processing stages corresponding to areas V1 and MT. Each stage carries  
581 out a series of computations including spatiotemporal filtering, spatial pooling, rectification, and  
582 divisive normalization. At the V1 stage, simple cells receive input directly from the visual stimulus  
583 and complex cells pool inputs from rectified and divisively normalized responses of V1 simple  
584 cells. At the MT stage, MT neurons pool inputs from V1 complex cells, followed by rectification  
585 and divisive normalization (Simoncelli and Heeger, 1998; Rust et al., 2006).

586  
587 We generated random-dot visual stimuli that are similar to those used in our physiological  
588 experiments and simulated the neuronal responses in areas MT and V1. The visual stimuli and a  
589 simplified architecture of the model are illustrated in Figure 7. The diameter of each random-dot  
590 patch was  $3^\circ$ , extending 63 pixels. The RF sizes of model V1 and MT neurons, set by the sizes of  
591 the convolution filters, were  $1.2^\circ$  and  $10^\circ$  in diameter, respectively (see Methods). The populations  
592 of model neurons in V1 and MT stages approximately tiled a sphere in the spatiotemporal  
593 frequency domain, as in Simoncelli and Heeger's model (1998). The RFs of V1 and MT neuron  
594 populations covered a region of the visual field that was  $17.3^\circ \times 17.3^\circ$ . In the overlapping  
595 condition, the apertures of two random-dot patches overlapped within the RFs (Fig. 7A). In the  
596 spatially-separated condition, the two random-dot patches were placed side by side, separated by  
597 a blank gap that was  $1.2^\circ$  wide, within the RFs of single MT neurons (Fig. 7B). In the overlapping  
598 condition, the V1 neurons whose RFs covered *site a* were activated by both stimulus components  
599 (Fig. 7A). In the spatially-separated condition, V1 neurons were activated by only one stimulus  
600 component, either at *site a* or *site b* (Fig. 7B).

601  
602 We tuned the model parameters (see Methods) to match the experimentally measured  
603 contrast response functions of V1 and MT neurons (Sclar et al., 1990) and the coherence response  
604 function of MT neurons (Britten and Newsome, 1998). The simulated contrast response functions  
605 of V1 and MT neurons fitted the experimental data almost perfectly, and the simulated coherence  
606 response function of MT neurons also matched the data well (Fig. 8A-C). As far as we know, an

607 experimentally measured coherence response function of V1 neurons has not been described  
608 previously. Our simulations show that V1 responses increased monotonically with the coherence  
609 level of moving random-dot stimuli (Fig. 8D). The model V1 neurons had slightly higher firing  
610 rates in response to low coherence stimuli and more trial-to-trial variability in comparison with the  
611 model MT neurons (Fig. 8C and D).

612

613 The MT responses elicited by our visual stimuli that competed between luminance contrast  
614 and motion coherence were well captured by the model. Consistent with our experimental data  
615 (Fig. 2), the tuning curve of model MT neurons to the “low contrast & high coherence” component  
616 had a greater peak response than that of the “high contrast & low coherence” component (Fig. 9A,  
617 B). In the overlapping condition, the simulated MT response elicited by the bi-directional stimuli  
618 was nearly completely biased toward the weaker “high contrast & low coherence” component (Fig.  
619 9A), as found in the neural data. The model also captured the change of MT response tuning when  
620 visual stimuli were rearranged spatially. In the spatially-separated condition, the tuning curve of  
621 model MT neurons elicited by the bi-directional stimuli was no longer dominated by the “high  
622 contrast & low coherence” component (Fig. 9B).

623

624 At the V1 stage of the model, the tuning curves of V1 complex cells showed a slightly  
625 greater mean peak response to the “high contrast & low coherence” component than to the “low  
626 contrast & high coherence” component (Fig. 9C). In the overlapping condition, the simulated V1  
627 response elicited by the bi-directional stimuli was strongly biased toward the “high contrast & low  
628 coherence” component (Fig. 9C), to the extent similar to that found in model MT neuron (Fig. 9A),  
629 as measured by the weights for the component responses using the SNL model fits. The bias index  
630 (Eq. 4) for the V1 model neuron was 0.90 and that for the MT model neuron was 0.93. These  
631 simulation results suggest that the strong bias toward the “high contrast & low coherence”  
632 component found in MT is inherited from V1.

633

634 In the spatially-separated condition, the V1 response elicited by the bi-directional stimuli  
635 was the same as that elicited by the single stimulus component placed within the RFs of V1 neurons  
636 (Fig. 9D, E). Although the V1 peak response elicited by the “high contrast & low coherence”  
637 component at site  $a$  was slightly stronger than that elicited by the “low contrast & high coherence”

638 component at site  $b$ , the MT response elicited by the bi-directional stimuli was skewed toward the  
639 “low contrast & high coherence” component, consistent with the average of the component  
640 responses (Fig. 9B). These simulation results suggest that MT response elicited by the bi-  
641 directional stimuli in the spatially-separated condition (Fig. 9B) may be due to feature competition  
642 within MT.

643

644 The response tuning curves of single MT neurons measured by varying the VA direction  
645 of the bi-directional stimuli can be mapped to the responses of a population of MT neurons that  
646 have different PDs, elicited by the bi-directional stimuli moving in a given VA direction. Figure 7  
647 summarizes the changes of the response distributions across neuron populations at V1 and MT  
648 stages, under the overlapping and spatially-separated conditions. These results reveal the  
649 importance of neural processing at different stages of the visual hierarchy on determining how  
650 multiple visual stimuli compete within neurons’ RFs in a given brain area.

651

652

## 653 **Discussion**

654

655 We have shown that how MT neurons represent multiple stimuli competing in more than  
656 one feature domain depends on the spatial arrangement of the visual stimuli. When two stimuli are  
657 overlapping, MT responses are dominated by the stimulus component that has high contrast. When  
658 two stimuli are spatially separated, the contrast dominance is abolished. Our neural data and model  
659 simulations suggest that the contrast dominance found with overlapping stimuli is due to  
660 normalization occurring at an input stage fed to MT, and MT neurons cannot overturn this contrast  
661 dominance based on their own feature selectivity. The interaction between spatially separated  
662 stimuli can largely be explained by normalization within area MT. By using multiple visual stimuli  
663 competing in more than one features domain, our study revealed how neural processing along the  
664 hierarchical visual pathway shapes neural representation of multiple visual stimuli in extrastriate  
665 cortex.

666

### 667 Consideration of the effect of attention

668 Attention can bias neuronal responses elicited by multiple stimuli in the RF in favor of the  
669 attended stimulus (Reynolds et al., 1999; Li and Basso, 2005; Treue and Maunsell, 1996; Ferrera



670 and Lisberger, 1997; Treue and Martinez-Trujillo, 1999; Recanzone and Wurtz, 2000; Lee and  
671 Maunsell, 2010). Although in this study the animals performed a fixation task without the need to  
672 engage goal-directed attention, could the high contrast component capture stimulus-driven  
673 attention (Corbetta and Shulman, 2002) and bias the neuronal response elicited by the overlapping  
674 stimuli? Several considerations argue against this possibility. While an abrupt stimulus onset  
675 captures attention (Yantis and Jonides, 1984), a visual stimulus that is brighter than other  
676 distractors does not automatically capture attention (Jonides and Yantis, 1988). The two stimulus  
677 components of our overlapping stimuli were turned on and started to move at the same time. The  
678 stimulus onset may automatically draw attention toward the spatial location of the overlapping  
679 stimuli, but it is unlikely to draw attention toward only the high contrast component. Furthermore,  
680 stimulus-driven attention occurs with a time delay (Nakayama and Mackeben, 1989) and its effect  
681 on neuronal responses in MT is transient, lasting for about 70 ms (Busse et al., 2008). In contrast,  
682 we found that the response bias toward the high contrast component is present in the very  
683 beginning of the neuronal responses following the onset of the static stimuli, and the bias is  
684 persistent throughout the motion period (Fig. 5). In addition, Wannig and colleagues (2007) have  
685 shown that attention directed to one of two overlapping surfaces can alter the responses of MT  
686 neurons. However, attention led to a response magnitude modulation of about 20% in MT between  
687 conditions when attention was directed to two different surfaces (Wannig et al., 2007). Even if, for  
688 some reason, the animals were consistently attending to the high contrast component throughout  
689 the stimulus presentation period in our study, the effect of attention would be insufficient to  
690 account for the nearly complete dominance by the high contrast component.

691

### 692 *Mechanisms underlying stimulus interactions*

693 The primate visual system is hierarchically organized (Maunsell and van Essen, 1983;  
694 Felleman and Van Essen, 1991). The response properties of neurons in a visual area are shaped by  
695 feedforward input, as well as intra-areal and feedback processes. To understand the mechanisms  
696 underlying neural encoding of multiple stimuli, it is important to determine how these processes  
697 contribute to the RF properties in a given visual area. However, it is often difficult to disentangle  
698 the contribution of feedforward input from other neural processes. We have previously found that,  
699 in response to overlapping stimuli, MT neurons show a bias toward the stimulus component that  
700 has a higher signal strength, defined by either luminance contrast or motion coherence (Xiao et

701 al., 2014). The response bias can be described by a model of divisive normalization. Because  
702 neurons in V1 also show a bias toward the stimulus component that has a higher contrast (Busse  
703 et al., 2009; MacEvoy et al., 2009) and divisive normalization may occur in both V1 and MT  
704 (Simoncelli and Heeger, 1998; Heuer and Britten, 2002), it was unclear how the feedforward input  
705 from V1 contributed to the response bias found in MT.

706

707 In this study, we are able to differentiate the impact of feedforward input from other neural  
708 processes on the response properties of MT neurons. Our results suggest that neurons in V1 may  
709 respond more strongly to the “high contrast & low coherence” component than to the “low contrast  
710 & high coherence” component used in our experiment, due to V1 neurons’ sensitivities to contrast  
711 and coherence. When two stimuli overlap, the responses of V1 neurons elicited by both stimulus  
712 components may already show a strong bias toward the “high contrast & low coherence”  
713 component due to divisive normalization in V1 (Fig. 9C). MT neurons are no longer able to remix  
714 the stimulus components according to their own sensitivities to contrast and coherence. In other  
715 words, MT neurons inherit the response bias toward the high contrast component from their input.  
716 When two visual stimuli are spatially separated, MT neurons receive inputs from two different  
717 pools of V1 neurons and each neuron pool responds to only one stimulus component (Fig. 7B).  
718 The neuronal responses elicited by the two stimulus components remain separated in V1. MT  
719 neurons can mix the responses elicited by the two stimulus components via spatial and directional  
720 pooling and divisive normalization within MT. As a result, the mixing in MT may well reflect the  
721 sensitivities of MT neurons to different stimulus features. Our model simulations make predictions  
722 regarding how V1 neurons respond to multiple competing stimuli (e.g. as shown in Fig. 9C), which  
723 can be tested in future physiological study.

724

### 725 *Implications on normalization and encoding of multiple visual stimuli*

726 Our finding that the response weighting for competing stimuli depends on the spatial  
727 arrangement provides a new perspective on the well-established normalization model (Carandini  
728 and Heeger, 2011). The basic form of normalization equations (Eqs. 5-6) predicts that the response  
729 weight for a stimulus component increases with its signal strength, but does not consider the spatial  
730 arrangement of the visual stimuli. We made a surprising finding that MT response to overlapping  
731 stimuli cannot be predicted by the population neural responses in MT elicited by the individual

732 stimulus components. One must consider the neural computations occurring along the hierarchical  
733 visual pathway.

734

735         Majaj, Carandini, and Movshon (2007) showed that pattern-direction selective neurons in  
736 MT characterized by overlapping drifting gratings (i.e. plaid) do not integrate the directions of the  
737 component gratings when they were spatially separated within the RF, suggesting that the  
738 computation underlying pattern-direction selectivity in MT is local. Different from the plaid, the  
739 overlapping random-dot stimuli used in our study elicit the percept of motion transparency. We  
740 showed that changing the spatial arrangement of visual stimuli can have a substantial impact not  
741 only on motion integration but also on the competition between multiple stimuli. Our results  
742 revealed that contrast has a dominant effect in determining stimulus competition within a local  
743 spatial region when multiple stimuli differ in more than one feature domain. When visual stimuli  
744 are spatially separated, the effect of contrast is substantially reduced.

745

746         A seminal model involving MT neurons pooling inputs from V1 and divisive normalization  
747 in both V1 and MT has been successful in explaining a range of experimental results of MT  
748 responses (Simoncelli and Heeger, 1998; Rust et al., 2006). However, the model in its original  
749 form does not specify how features are spatially integrated and it does not differentiate overlapping  
750 and spatially separated stimuli (Majaj et al., 2007). In our study, we adapted this model to simulate  
751 both overlapping and spatially separated conditions and showed that the framework can explain  
752 our main physiological findings. Also using this model, Busse, Wade, and Carandini (2009)  
753 previously demonstrated the impact of response normalization in V1 on neural response in MT.  
754 They showed that, by making the contrasts of two drifting gratings of a plaid to be unequal, the  
755 response of a model MT neuron changed from representing the pattern motion of the plaid to  
756 mostly representing the higher-contrast grating component, likely due to contrast normalization in  
757 V1 (Busse et al., 2009). However, the MT response elicited by the higher-contrast grating alone  
758 could also be greater than that elicited by the lower-contrast grating. The model-predicted response  
759 bias toward the higher-contrast component in MT may also be contributed by response  
760 normalization within MT, akin to our experimental result obtained using random-dot stimuli with  
761 unequal contrasts (Xiao et al., 2014). In comparison, our current study provides unequivocal new  
762 evidence on how responses in MT are shaped by the hierarchical network. By using two stimuli

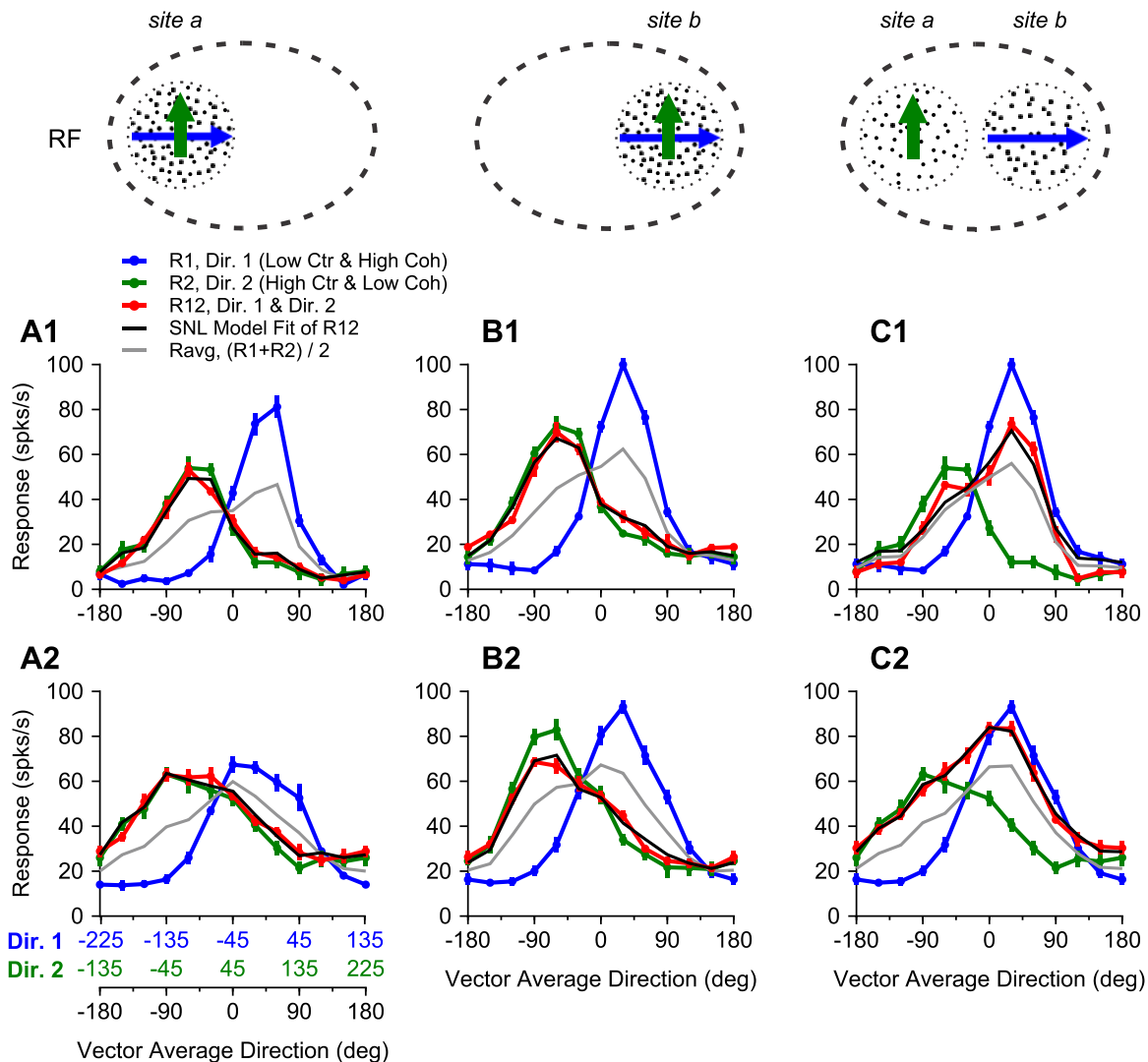
763 competing in more than one feature domain, we demonstrated neurophysiologically and  
764 computationally the substantial impact of stimulus competition in the input stage on the neuronal  
765 responses in MT and how that impact changes with the spatial arrangement of visual stimuli. Our  
766 finding may also apply to other visual areas in the hierarchical network, including those in the  
767 ventral visual stream where response normalization has been well documented.

768 **References**

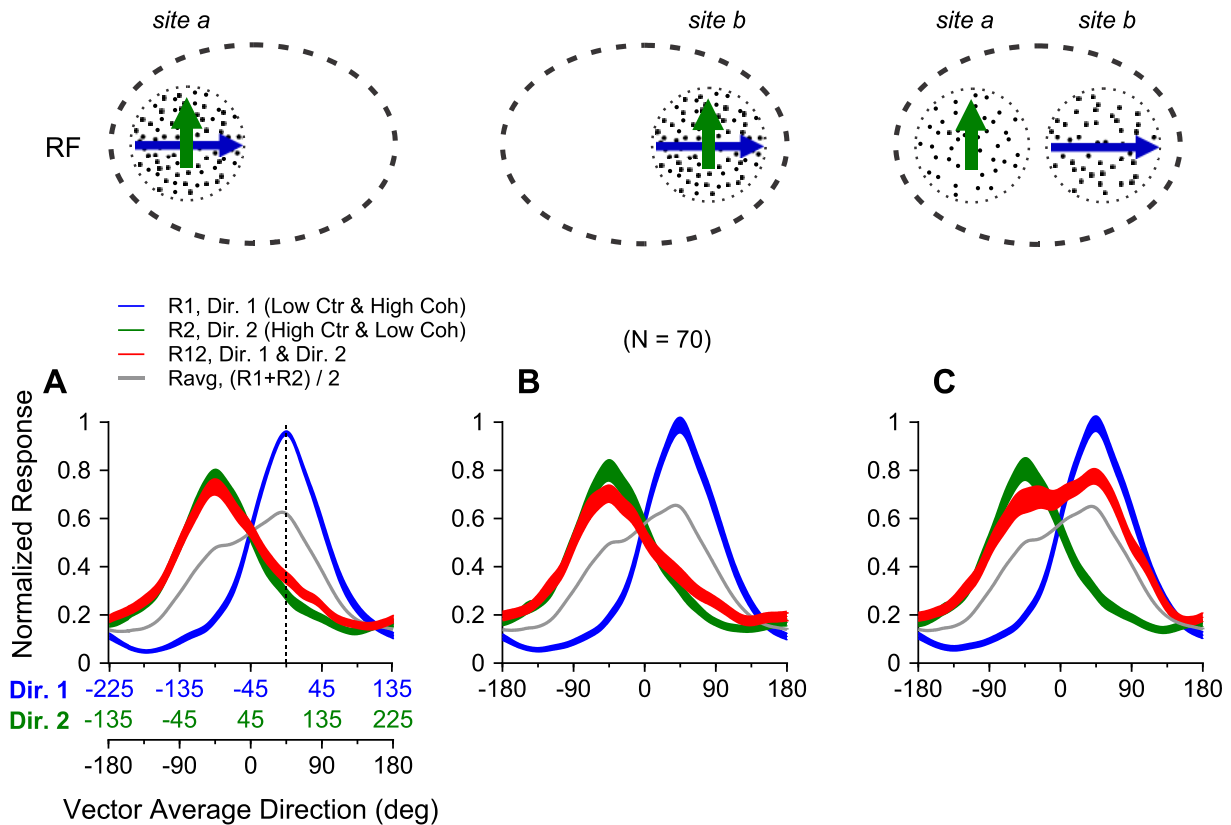
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## Figures and Figure Legends

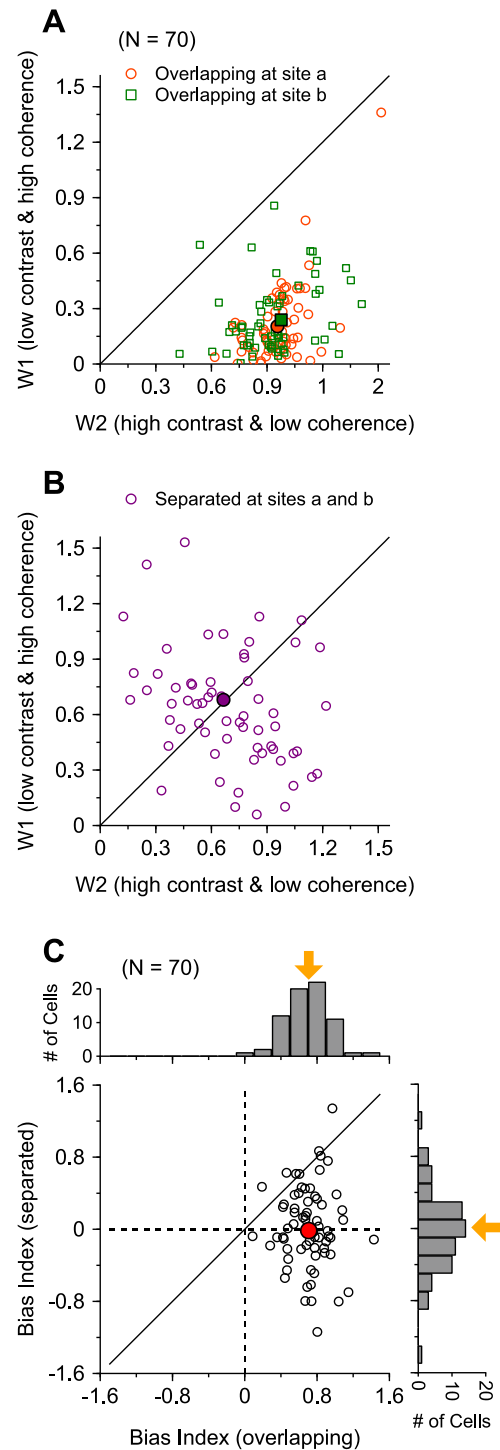


859 **Figure 1.** The response tuning curves of two example MT neurons to overlapping (A, B) and  
 860 spatially-separated stimuli (C). Visual stimuli were achromatic random-dot patches moving in two  
 861 directions separated by 90°. The “low contrast & high coherence” component (shown in blue  
 862 arrow) moved at the clockwise side of the two component directions, whereas the “high contrast  
 863 & low coherence” component (shown in green arrow) moved in the direction at the counter-  
 864 clockwise side. The X-axis labeled in black indicates the vector average direction of the bi-  
 865 directional stimuli. The X-axes labeled in blue and green (A2) indicate the direction of the “low  
 866 contrast & high coherence” component (Dir. 1) and the direction of “high contrast & low  
 867 coherence” component (Dir. 2), respectively. The three X-axes are aligned such that the  
 868 component directions shown in blue and green correspond to the directions of the two stimulus  
 869 components at each vector average direction. A1-C1: Response tuning curves from one neuron.  
 870 A2-C2: Response tuning curves from another neuron. The responses elicited by the bi-directional  
 871 stimuli are shown in red (R12). The SNL model fits of R12 are shown in black. Error bars represent  
 872 standard errors.

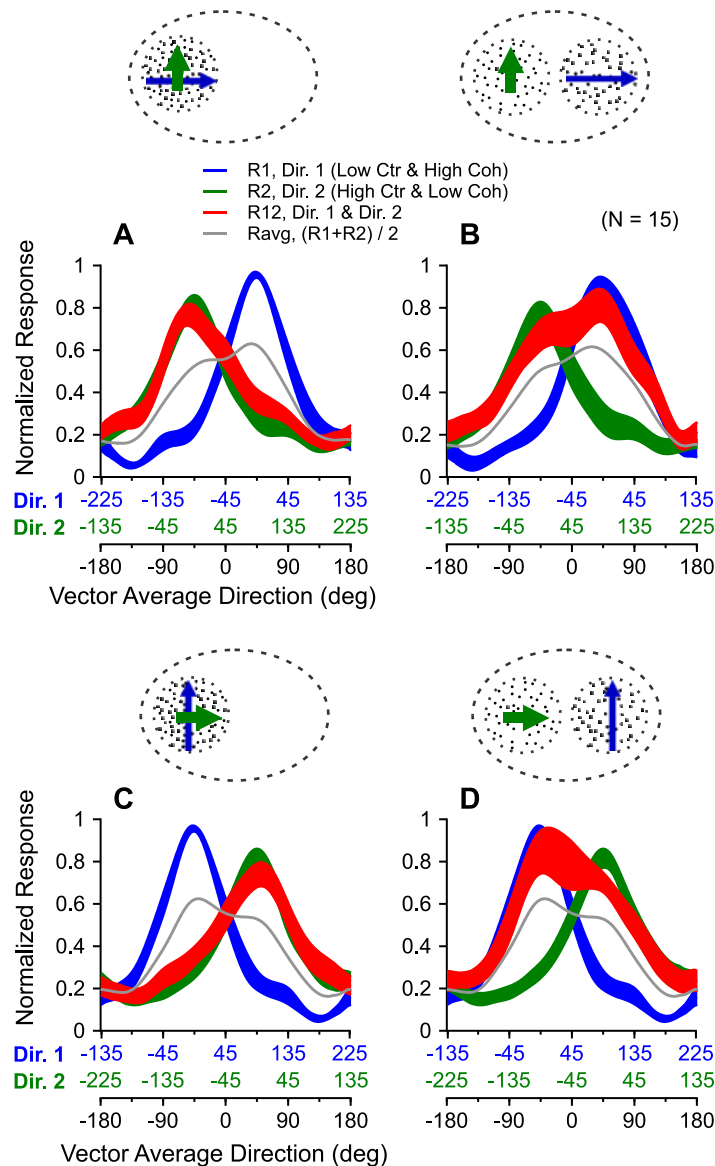


873 **Figure 2.** Population-averaged tuning curves to the bi-directional stimuli (red) and the  
 874 unidirectional stimulus components (blue and green). The vector average direction of the bi-  
 875 directional stimuli and the directions of individual stimulus components are labeled in the  
 876 corresponding X-axes (A), following the same convention as in Figure 1. The direction of 0° was  
 877 aligned with each neuron's PD before the tuning curves were averaged across neurons. The  
 878 stimulus components were overlapping at site *a* (A) or site *b* (B), or spatially separated (C)  
 879 within the RFs. The width of each tuning curve represents the standard error. The average of the responses  
 880 to the two stimulus components is shown in gray.

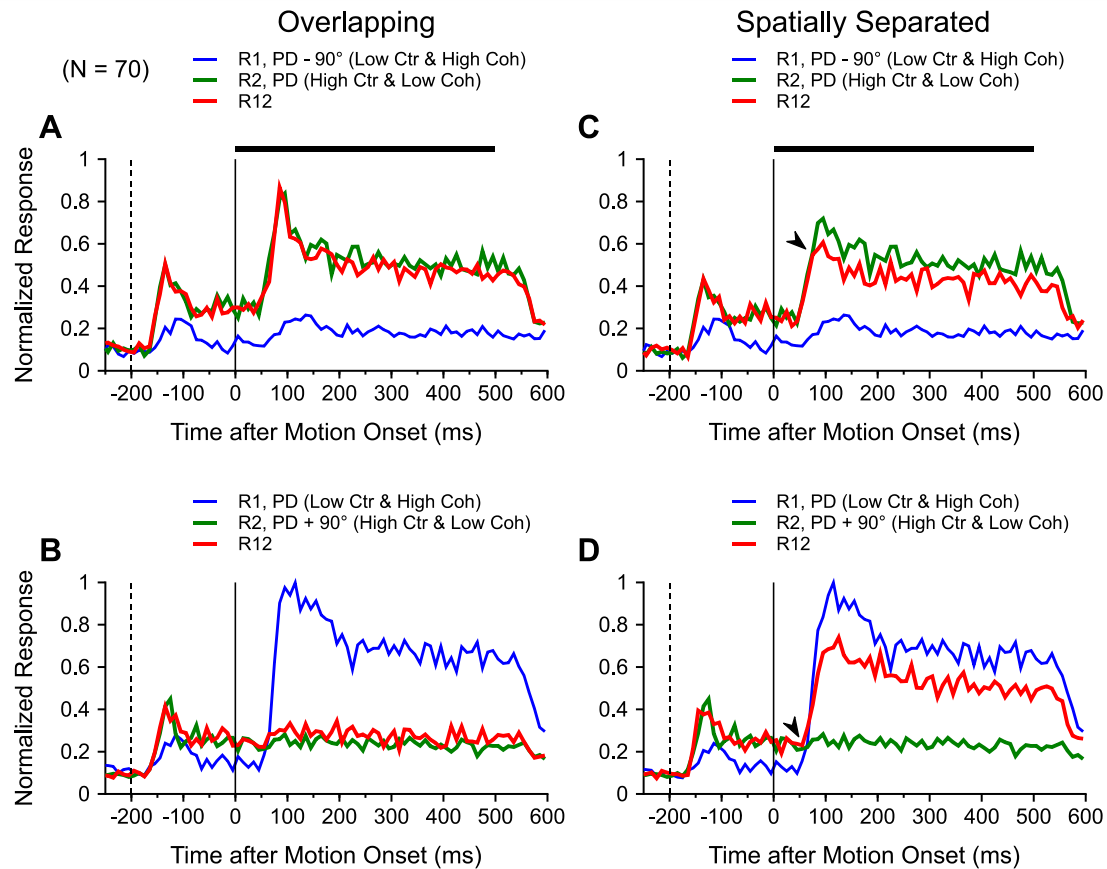




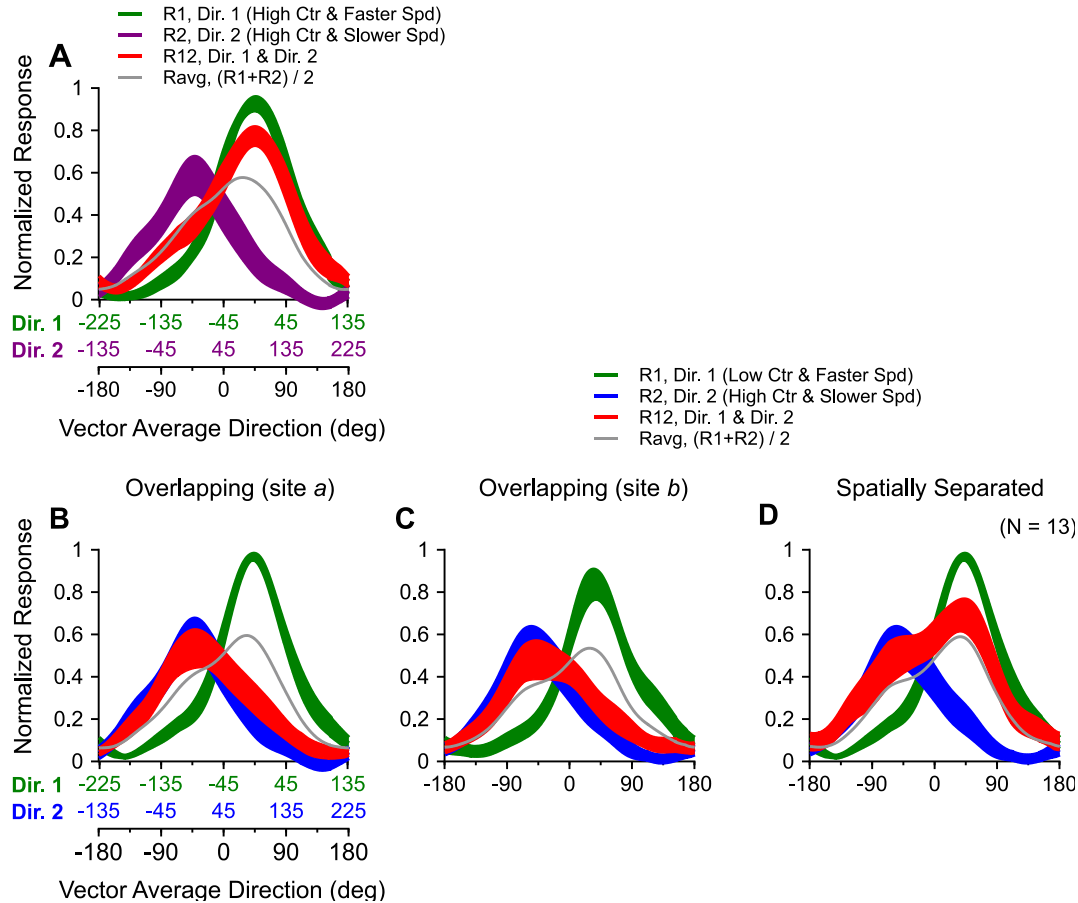
881 **Figure 3.** The effect of the spatial arrangement of the  
882 bi-directional stimuli on the response weights for the  
883 stimulus components. Each dot represents the result  
884 from one neuron. Comparing the response weights for  
885 the “low contrast & high coherence” component  
886 (ordinate) with the “high contrast & low coherence”  
887 component (abscissa) under the overlapping (A) and  
888 the spatially separated (B) conditions. C. Comparing  
889 the bias indices between the spatially separated  
890 (ordinate) and overlapping (abscissa) conditions. The  
891 histograms in C show the distributions of the bias index  
892 for the overlapping (top) and spatially separated (right)  
893 conditions.



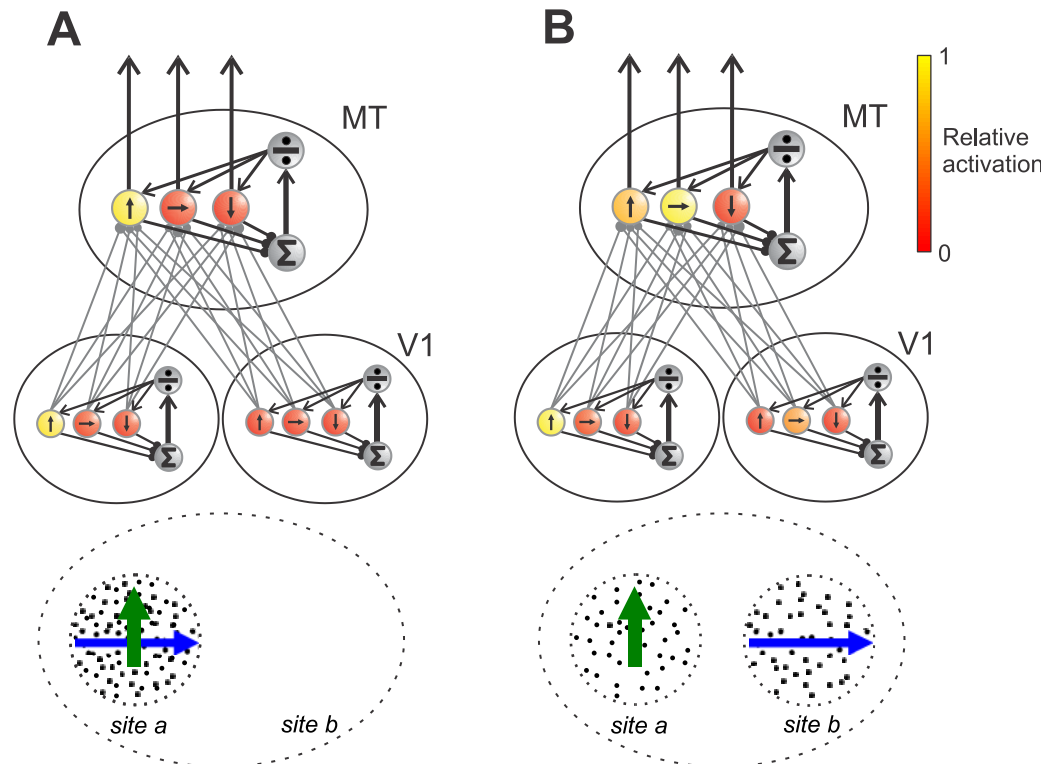
894 **Figure 4.** Control for the directional arrangement of the two stimulus components. **A, B.** Response  
 895 tuning curves averaged across 15 MT neurons to the bi-directional stimuli and the stimulus  
 896 components when the direction of the “high contrast & low coherence” component was placed at  
 897 the counter-clockwise side of the two component directions, as in Figures 1 and 2. **C, D.** Response  
 898 tuning curves averaged across the same 15 neurons when the direction of the “high contrast & low  
 899 coherence” component was placed at the clockwise side of the two component directions. **A, C.**  
 900 Overlapping condition. **B, D.** Spatially separated condition. Notice the switch of the values in the  
 901 X-axes of the component directions, shown in blue and green, between A and C, as well as between  
 902 B and D.



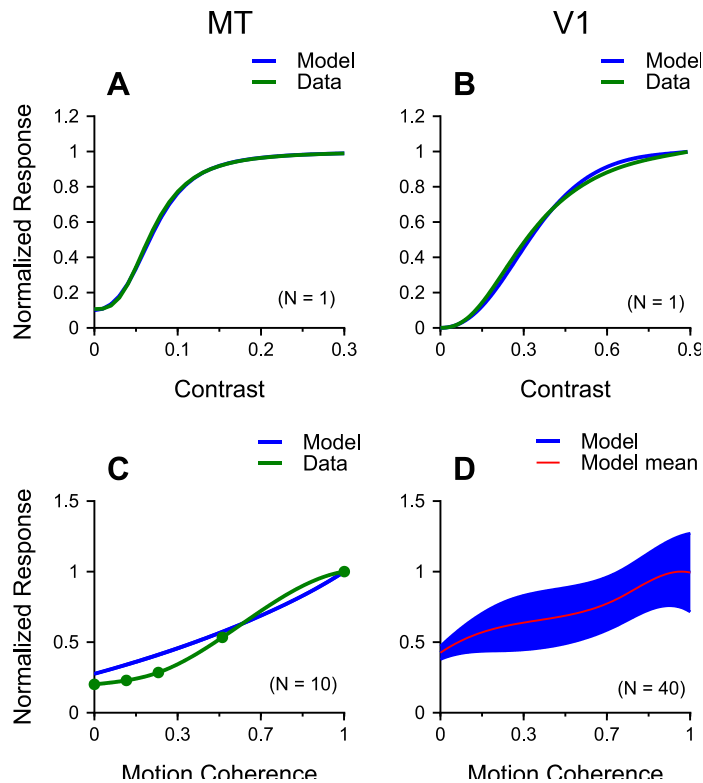
903 **Figure 5.** Timecourse of the neuronal responses to the bi-directional stimuli and the stimulus  
 904 components. Peristimulus time histograms (PSTHs) were calculated using a 10-ms time bin and  
 905 averaged across 70 neurons. **A, B.** The two stimulus components overlapped (at site *a*) within the  
 906 RF. **C, D.** The two stimulus components were spatially separated within the RF. The dashed  
 907 vertical lines at -200 ms indicate the onset of the static stimuli. The solid vertical lines at time 0  
 908 indicate motion onset. The solid horizontal bars shown in A and C indicate the stimulus motion  
 909 period. The “high contrast & low coherence” component moved in the PD in A and C, and moved  
 910 in a non-PD in B, D.



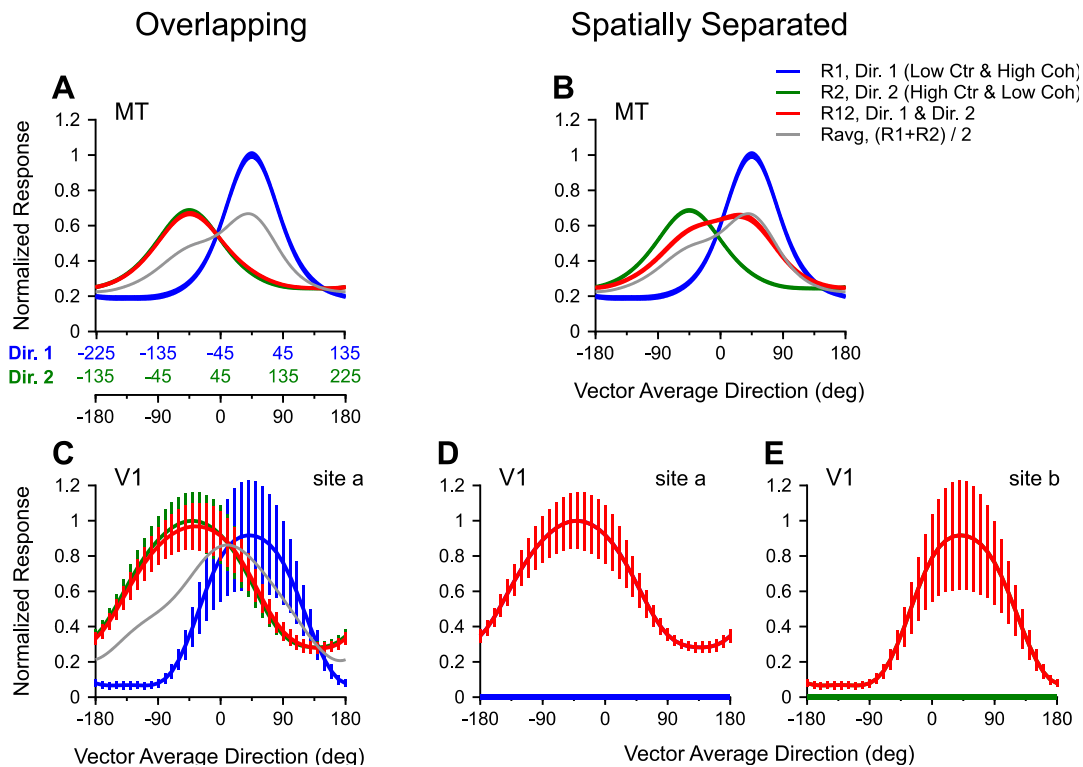
911 **Figure 6.** Averaged response tuning curves to two stimulus components that moved in different  
 912 directions and at different speeds. Both stimulus components moved at 100% coherence. The  
 913 response tuning to both stimulus components presented simultaneously is shown in red. The width  
 914 of each tuning curve represents the standard error. The average of the component responses elicited  
 915 by the individual stimulus components is shown in gray. **A.** Both stimulus components had high  
 916 luminance contrast and were overlapping. **B-D.** The two stimulus components competed in  
 917 luminance contrast and motion speed. The faster speed component had low luminance contrast,  
 918 whereas the slower speed component had high luminance contrast. The stimulus components were  
 919 overlapping at site *a* (**B**) or site *b* (**C**), or were spatially separated (**D**) within the RFs. The vector  
 920 average direction of the bi-directional stimuli and the directions of individual stimulus components  
 921 are labeled in the corresponding X-axes (**A, B**), following the same convention as in Figure 2.



922 **Figure 7.** Illustration of a simplified architecture for the V1-MT model. Each MT neuron receives  
923 feedforward inputs from multiple neurons at the V1 stage. Responses are divisively normalized by  
924 the sum of local population activity at both V1 and MT stages. Each small circle represents a  
925 neuron and the black arrow inside the circle indicates the PD. The color of each circle indicates  
926 the response magnitude of the neuron. Yellow means maximum response and red means minimum  
927 response. Visual stimuli are illustrated below neural circuit as the input to the V1 stage. The green  
928 and blue arrows represent the “high contrast & low coherence” component and the “low contrast  
929 & high coherence” component, respectively. Two pools of neurons at the V1 stage that respond  
930 only to site *a* or site *b* respectively are illustrated. The RFs of the MT neurons are illustrated by  
931 the dotted ellipse and cover both site *a* and site *b*. **A.** Overlapping condition. **B.** Spatially separated  
932 condition.



933 **Figure 8.** Contrast and coherence response functions of model V1 (**B, D**) and MT (**A, C**) neurons.  
934 **A, B.** Fitted contrast response functions to sinusoidal gratings for model neurons. Green curves  
935 are experimental data replotted from Sclar et al. (1990). **C.** Fitted coherence response function to  
936 high contrast random-dots for model MT neurons. Green dots are experimental data replotted from  
937 Britten and Newsome (1998). The green curve is the spline fit of the experimental data points. **D:**  
938 Coherence response to high contrast random-dots for model V1 complex cells. The widths of the  
939 blue curves in C and D represent the standard deviation. N indicates the number of repeats for  
940 simulations. The stimulus dots were regenerated randomly for each simulation in C and D.



941 **Figure 9.** Computer simulations of direction tuning curves of MT and V1 neurons to the bi-  
 942 directional stimuli used in the main physiological experiment. The visual stimuli are either  
 943 overlapping (**A, C**) or spatially separated (**B, D, E**) within the RFs of model MT neurons. The  
 944 two stimulus components compete in luminance contrast and motion coherence. The simulated  
 945 responses to the “low contrast & high coherence” component and the “high contrast & low  
 946 coherence” component are shown in blue and green, respectively. The responses to the bi-  
 947 directional stimuli are shown in red. The vector average direction and the directions of individual  
 948 stimulus components are labeled in the corresponding X-axes (**A**), following the same  
 949 convention as in Figure 2. **A, B.** Simulated responses of model MT neurons. **C-E.** Simulated  
 950 responses of model V1 complex cells. Widths of the tuning curves in A and B and the error bars  
 951 in C-E represent standard deviations.