

Genetic paths to evolutionary rescue and the distribution of fitness effects along them

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ABSTRACT

The past century has seen substantial theoretical and empirical progress on the genetic basis of adaptation. Over this same period a pressing need to prevent the evolution of drug resistance has uncovered much about the potential genetic basis of persistence in declining populations. However, we have little theory to predict and generalize how persistence – by sufficiently rapid adaptation – might be realized in this explicitly demographic scenario. Here we use Fisher's geometric model with absolute fitness to begin a line of theoretical inquiry into the genetic basis of evolutionary rescue, focusing here on asexual populations that adapt through *de novo* mutations. We show how the dominant genetic path to rescue switches from a single mutation to multiple as mutation rates and the severity of the environmental change increase. In multi-step rescue, intermediate genotypes that themselves go extinct provide a 'springboard' to rescue genotypes. Comparing to a scenario where persistence is assured, our approach allows us to quantify how a race between evolution and extinction leads to a genetic basis of adaptation that is composed of fewer loci of larger effect. We hope this work brings awareness to the impact of demography on the genetic basis of adaptation.

KEYWORDS Antimicrobial drug resistance; Evolutionary escape; Fisher's geometric model; Genetic basis of adaptation; Mathematical theory

Our understanding of the genetic basis of adaptation is rapidly improving due to the now widespread use of experimental evolution and genomic sequencing (see examples in Bell 2009; Stapley *et al.* 2010; Dettman *et al.* 2012; Schlötterer *et al.* 2015). A recurrent observation, especially in asexual microbes, is that the more novel the environment and the stronger the selection pressure, the more likely it is that adaptation primarily proceeds by fewer mutations of larger effect (i.e., that adaptation is oligogenic *sensu* Bell 2009). An extreme case is the evolution of drug resistance, which is often achieved by just one or two mutations (e.g., Bataillon *et al.* 2011; Pennings *et al.* 2014).

However, drugs, and other sufficiently novel environments, will often induce not only strong selection but also population decline. Such declines hinder both the production and maintenance of genetic variation (Otto and Whitlock 1997), thus impeding evolution and threatening extinction. In this scenario, drug resistance evolution is a particular instance of the more general phenomenon of evolutionary rescue (Gomulkiewicz and

Holt 1995; Bell 2017), where persistence requires sufficiently fast adaptive evolution.

Most theory on the genetics of adaptation (reviewed in Orr 2005) assumes constant population size and therefore does not capture the characteristic 'race' between adaptation and extinction that occurs during evolutionary rescue. Many models have been created to describe this race (reviewed in Alexander *et al.* 2014) but so far largely focus on two extreme genetic bases, both already introduced in Gomulkiewicz and Holt (1995): rescue is either caused by minute changes in allele frequencies across many loci in sexuals (i.e., the infinitesimal model; Fisher 1918) or by the substitution of a single large effect 'resistance' mutation (e.g., one locus, two allele models). We therefore lack a theoretical framework for the genetic basis of evolutionary rescue that captures the arguably more realistic situation where an intermediate number of mutations are at play (but see exceptions below). The absence of such a framework prevents us from predicting the number of mutations that evolutionary rescue will take and the distribution of their effect sizes. The existence of a more complete framework could therefore provide valuable information to those investigating the genetic basis of drug resistance (e.g., the number and effect sizes of mutations to look for) and would

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55 extend our understanding on the genetic basis of adaptation to
56 cases of non-equilibrium demography (i.e., rapid evolution and
57 eco-evo dynamics).

58 Despite these gaps in the theory on the genetic basis of evolu-
59 tionary rescue, there is a wealth of data. For example, the genetic
60 basis of resistance to a variety of drugs is known in many species
61 of bacteria (reviewed in [MacLean et al. 2010](#)), fungi (reviewed
62 in [Robbins et al. 2017](#)), and viruses (reviewed in [Yilmaz et al.](#)
63 [2016](#)). This abundance of data reflects both the applied need
64 to prevent drug resistance and the relative ease of isolating the
65 genotypes that survive (hereafter "rescue genotypes"), e.g., in a
66 Luria-Delbrück fluctuation assay (reviewed in [Bataillon and](#)
67 [Bailey 2014](#)). Assaying fitness in the environment used to isolate
68 mutants (e.g., in the drug) then provides the distribution of fit-
69 ness effects of potential rescue genotypes. Additional data on
70 the genetic basis of drug resistance arise from the construction
71 of mutant libraries (e.g., [Weinreich et al. 2006](#)) and the sequenc-
72 ing of natural populations (e.g., [Pennings et al. 2014](#)). Together,
73 the data show that resistance often appears to arise by a single
74 mutation (e.g., [MacLean and Buckling 2009](#); [Lindsey et al. 2013](#);
75 [Gerstein et al. 2012](#)) but not always (e.g., [Bataillon et al. 2011](#);
76 [Pennings et al. 2014](#); [Gerstein et al. 2015](#); ?). The data also indi-
77 cate that the fitness effect of rescue genotypes is more often
78 large than small, creating a hump-shaped distribution of selec-
79 tion coefficients (e.g., [Kassen and Bataillon 2006](#); [MacLean and](#)
80 [Buckling 2009](#); [Gerstein et al. 2012](#); [Lindsey et al. 2013](#); [Gerstein](#)
81 [et al. 2015](#)) that is similar in shape to that proposed by [Kimura](#)
82 [\(1983\)](#) but with a lower bound that is often much greater than
83 zero.

84 Theory on evolutionary rescue (reviewed in [Alexander et al.](#)
85 [2014](#)) has primarily focused on the probability of rescue rather
86 than its genetic basis. However, a few studies have varied the
87 potential genetic basis enough to make some inference about
88 how evolutionary rescue is likely to happen. For instance, in the
89 context of pathogen host-switching, [Antia et al. \(2003\)](#) numeri-
90 cally explored the probability of rescue starting from a single
91 ancestral individual when n intermediate mutations (each occur-
92 ring from the previous with the same probability) are required.
93 They focused on the case of rescue by two mutations ($n = 1$),
94 when the intermediate genotype had a growth rate that was
95 either the same as the ancestor, below that of the ancestor, or
96 intermediate between the ancestor and the rescue genotype. In
97 all cases they found that rescue became less likely as the number
98 of intermediate mutations increased, suggesting that rescue will
99 generally proceed by the fewest possible mutations. This frame-
100 work was expanded greatly by [Iwasa et al. \(2004a\)](#) who derived
101 approximations for the probability of rescue with an arbitrary
102 number of mutational steps, considering arbitrary mutational
103 networks (i.e., allowing mutations between any two genotypes)
104 and standing genetic variation in an ancestral population. Assu-
105 ming the probability of mutation between any two genotypes
106 is of the same order, they showed that genetic paths with fewer
107 mutational steps contributed more to the probability of rescue,
108 again suggesting rescue will occur by the fewest possible muta-
109 tions. [Iwasa et al. \(2004a\)](#) also found that multiple simultaneous
110 mutations (i.e., arising in the same meiosis) can contribute more
111 to rescue than paths that gain these same mutations sequen-
112 tially (i.e., over multiple generations) when the growth rates of
113 the intermediate mutations are small enough, suggesting that
114 rare large mutations can be the most likely path to rescue when
115 the population is very maladapted or the rescue phenotype is
116 complex (i.e., there is a fitness valley separating the wildtype

117 and rescue genotype). This point was previously demonstrated
118 by [Alexander and Day \(2010\)](#), who emphasized that multiple
119 simultaneous mutations become the dominant path to rescue in
120 the most challenging environments. As a counterpoint, [Uecker](#)
121 [and Hermisson \(2016\)](#) explored a greater range of fitness values
122 in a two-locus two-allele model, showing that, with standing ge-
123 netic variation, rescue by sequential mutations at two loci (two
124 mutational steps) can be more likely than rescue by mutation
125 at a single locus (one simultaneous mutational step) when the
126 wildtype is very maladapted, as, in the former case, the single
127 mutants can act as a buffer in the face of environmental change.
128 In summary, current theory indicates that the genetic basis of
129 rescue hinges on the chosen set of genotypes, their fitnesses, and
130 the mutation rates between them. So far these choices have been
131 in large part arbitrary or chosen for mathematical convenience.

132 Here we follow the lead of [Anciaux et al. \(2018\)](#) by implicitly
133 choosing the range of genotypes, fitnesses, and mutation rates
134 from an empirically-justified fitness-landscape model ([Tenail-
135 lion 2014](#)). In particular, we use Fisher's geometric model to describe
136 adaptation following an abrupt environmental change that insti-
137 gates population decline. There are two key differences between
138 this approach and earlier models using Fisher's geometric model
139 (e.g., [Orr 1998](#)): here 1) the dynamics of each genotype depends
140 only on their absolute fitness (instead of only on their relative
141 fitness) and 2) multiple mutations can segregate simultaneously
142 (instead of assuming only sequential fixation), allowing multiple
143 mutations to fix – in our case, rescue the population – together as
144 a single haplotype (i.e., stochastic tunnelling, [Iwasa et al. 2004b](#)).
145 In this non-equilibrium scenario, variation in absolute fitness,
146 which allows population size to vary, can create feedbacks be-
147 tween demography and evolution, which could strongly impact
148 the genetic basis of adaptation relative to the constant popula-
149 tion size case. In contrast to [Anciaux et al. \(2018\)](#), our focus here
150 is on the genetic basis of evolutionary rescue and we also explore
151 the possibility of rescue by mutant haplotypes containing more
152 than one mutation. In particular, we ask: (1) How many muta-
153 tional steps is evolutionary rescue likely to take? and (2) What
154 is the expected distribution of fitness effects of the surviving
155 genotypes and their component mutations?

156 We first introduce the modelling framework before summar-
157 izing our main results. We then present the mathematical anal-
158 yses we have used to understand these results and end with a
159 discussion of our key findings.

160 **Data availability**

161 Code used to derive analytical and numerical results and pro-
162 duce figures (referred to here as File S1; Mathematica, ver-
163 sion 9.0; [Wolfram Research Inc. 2012](#)) and code used to run
164 individual-based simulations (Python, version 3.5; Python Soft-
165 ware Foundation), as well as simulation data and freely ac-
166 cessible versions of File S1 (CDF and PDF), are available at
167 <https://github.com/mmosmond/GeneticBasisOfRescue>.

168 **Model**

169 **Fisher's geometric model**

170 We map genotype to phenotype to fitness using Fisher's geo-
171 metric model, originally introduced by Fisher (1930, p. 38-41)
172 and reviewed by [Tenail-
173 lion \(2014\)](#). In this model each geno-
174 type is characterized by a point in n -dimensional phenotypic
175 space, \vec{z} . We ignore environmental effects, and thus the pheno-
type is the breeding value. Some phenotype, \vec{o} , has maximum

176 fitness and fitness declines monotonically as phenotypes de- 234
 177 part from \vec{o} . Without loss of generality, the n phenotypic axes 235
 178 are chosen and scaled such that fitness can be described by 236
 179 a multivariate normal distribution with variance 1 in each di- 237
 180 mension, no covariance, and height W_{max} . Thus the fitness 238
 181 of phenotype \vec{z} is $W(\vec{z}) = W_{max} \exp(-\|\vec{z} - \vec{o}\|^2/2)$, where 239
 182 $\|\vec{z} - \vec{o}\| = \sqrt{\sum_{i=1}^n (z_i - o_i)^2}$ is the Euclidean distance of \vec{z} from 240
 183 the optimum, \vec{o} . Here we are interested in absolute fitness; 241
 184 we take $\ln[W(\vec{z})] = m(\vec{z}) = m_{max} - \|\vec{z} - \vec{o}\|^2/2$ to be the 242
 185 continuous-time growth rate (m is for Malthusian fitness) of 243
 186 phenotype \vec{z} . We ignore density- and frequency-dependence 244
 187 in $m(\vec{z})$ for simplicity. The fitness effect, i.e., selection coeffi- 245
 188 cient, of phenotype z' relative to z in a continuous-time model 246
 189 is exactly $s = \log[W(z')/W(z)] = m(z') - m(z)$ (Martin and 247
 190 Lenormand 2015). This is approximately equal to the selection 248
 191 coefficient in discrete time ($(W(z')/W(z) - 1)$) when selection is 249
 192 weak ($(W(z') - W(z)) \ll 1$).

193 To make analytical progress we use the isotropic version of 247
 194 Fisher's geometric model, where mutations (in addition to sele- 248
 195 ction) are assumed to be uncorrelated across the scaled traits. 249
 196 Universal pleiotropy is also assumed, so that each mutation 250
 197 affects all scaled phenotypes. In particular we use the "class- 251
 198 ic" form of Fisher's geometric model (Harmand *et al.* 2017), 252
 199 where the probability density function of a mutant phenotype 253
 200 is multivariate normal, centred on the current phenotype, with 254
 201 variance λ in each dimension and no covariance. This implies 255
 202 a continuum-of-alleles (Kimura 1965), i.e., phenotype is contin- 256
 203 uous and each mutation is unique. Mutations are assumed to 257
 204 be additive in phenotype, which induces epistasis in fitness as 258
 205 fitness is a non-linear function of phenotype. We assume asexual 259
 206 reproduction, i.e., no recombination., which is appropriate for 260
 207 many cases of antimicrobial drug resistance.

208 An obvious and important extension would be to relax the 261
 209 simplifying assumptions of isotropy and universal pleiotropy, 262
 210 which we leave for future work. Note that mild anisotropy 263
 211 yields the same bulk distribution of fitness effects as an isotropic 264
 212 model with fewer dimensions (Martin and Lenormand 2006), 265
 213 but this does not extend to the tails of the distribution. There- 266
 214 fore, whether anisotropy can be reduced to isotropy with fewer 267
 215 dimensions in the case of evolutionary rescue, where the tails 268
 216 are essential, is unknown.

217 Given this phenotype to fitness mapping and phenotypic 269
 218 distribution of new mutations, the distribution of fitness effects 270
 219 (and therefore growth rates) of new mutations can be derived ex- 271
 220 actly. Let m be the growth rate of some particular focal genotype 272
 221 and m' the growth rate of a mutant immediately derived from it. 273
 222 Then let $s_o = m_{max} - m$ be the selective effect of a mutant with 274
 223 the optimum genotype and $s = m' - m$ the selective effect of 275
 224 the mutant with growth rate m' . The probability density func- 276
 225 tion of the selective effects of new mutations, s , is then given by 277
 226 equation 3 in Martin and Lenormand (2015). Converting fitness 278
 227 effects to growth rate ($m' = s + m$), the probability density func- 279
 228 tion for mutant growth rate m' from an ancestor with growth 280
 229 rate m is (cf. equation 2 in Anciaux *et al.* 2018)

$$f(m'|m) = \frac{2}{\lambda} f_{\chi_n^2} \left(\frac{2(m_{max} - m')}{\lambda}, \frac{2(m_{max} - m)}{\lambda} \right), \quad (1)$$

230 where $f_{\chi_n^2}(x, c)$ is the probability density function over positive 281
 231 real numbers x of $\chi_n^2(c)$, a non-central chi-square deviate with n 282
 232 degrees of freedom and noncentrality $c > 0$ (equation 26.4.25 in 283
 233 Abramowitz and Stegun 1972).

234 Lifecycle

235 We are envisioning a scenario where N_0 wildtype individuals, 236
 237 each of which have phenotype \vec{z}_0 , experience an environmental 238
 239 change, causing population decline, $m_0 \equiv m(\vec{z}_0) < 0$. Each 240
 241 generation, an individual with phenotype \vec{z} produces a Poisson 242
 243 number of offspring, with mean $\ln[m(\vec{z})]$, and dies. Each off- 244
 245 spring mutates with probability U , with mutations distributed 246
 247 as described above (see Fisher's geometric model).

242 Probability of rescue

243 Let p_0 be the probability that a given wildtype individual is 244
 245 "successful", i.e., has descendants that rescue the population. 246
 247 The probability of rescue is then one minus the probability that 248
 249 none of the initial wildtype individuals are successful,

$$P = 1 - (1 - p_0)^{N_0} \approx 1 - \exp(-N_0 p_0), \quad (2)$$

247 where the approximation assumes small p_0 and large N_0 . What 248
 249 remains is to find p_0 .

249 Summary of Results

250 We start with a heuristic explanation of our main results before 251
 252 turning to more detailed derivations in the next section.

252 Rescue by multiple mutations

253 A characteristic pattern of evolutionary rescue is a "U"-shaped 254
 255 population size trajectory (e.g., Orr and Unckless 2014). This is 256
 257 the result of an exponentially-declining wildtype genotype being 258
 259 replaced by an exponentially-increasing mutant genotype. On a 260
 261 log scale this population size trajectory becomes "V"-shaped 262
 263 (we denote it a 'V-shaped log-trajectory'). On this scale, the pop- 264
 265 ulation declines at a constant rate (producing a line with slope 265
 266 $m_0 < 0$) until the growing mutant subpopulation becomes rela- 267
 268 tively common, at which point the population begins growing 268
 269 at a constant rate (a line with slope $m_1 > 0$). This characteris- 269
 270 tic V-shaped log-trajectory is observed in many of our simulations 270
 271 where evolutionary rescue occurs (Figure 1A). In contrast, when 271
 272 the wildtype declines faster and the mutation rate is larger we 272
 273 sometimes see 'U-shaped log-trajectories' (e.g., the red and blue 273
 274 replicates in Figure 2A). Here there are three phases instead of 274
 275 two; the initial rate of decline (a line with slope $m_0 < 0$) is first 275
 276 reduced (transitioning to a line with slope $m_1 < 0$) before the 276
 277 population begins growing (a line with slope $m_2 > 0$).

278 As expected, V-shaped log-trajectories are the result of a sin- 279
 280 gle mutation creating a genotype with a positive growth rate 280
 281 that escapes loss when rare and rescues the population (Figure 281
 282 1B), i.e., 1-step rescue. U-shaped log-trajectories, on the other 282
 283 hand, occur when a single mutation creates a genotype with a 283
 284 negative (or potentially very small positive) growth rate, itself 284
 285 doomed to extinction, which out-persists the wildtype and gives 285
 286 rise to a double mutant genotype that rescues the population 286
 287 (Figure 2B), i.e., 2-step rescue. These two types of rescue com- 287
 288 prise the overwhelming majority of rescue events observed in 288
 289 our simulations, across a wide range of wildtype decline rates 289
 290 (e.g., Figure 3).

290 Of course, with sufficiently high mutation rates rescue by 291
 292 3 or more mutations comes to dominate (Figure S1). It has 292
 293 recently been suggested that when the mutation rate, U , is 293
 294 substantially less than a critical value, $U_C = \lambda n^2/4$, we are 294
 295 in a "strong selection, weak mutation" regime where essen- 295
 296 tially all mutations arise on a wildtype background (Martin 296
 297 and Lenormand 2015).

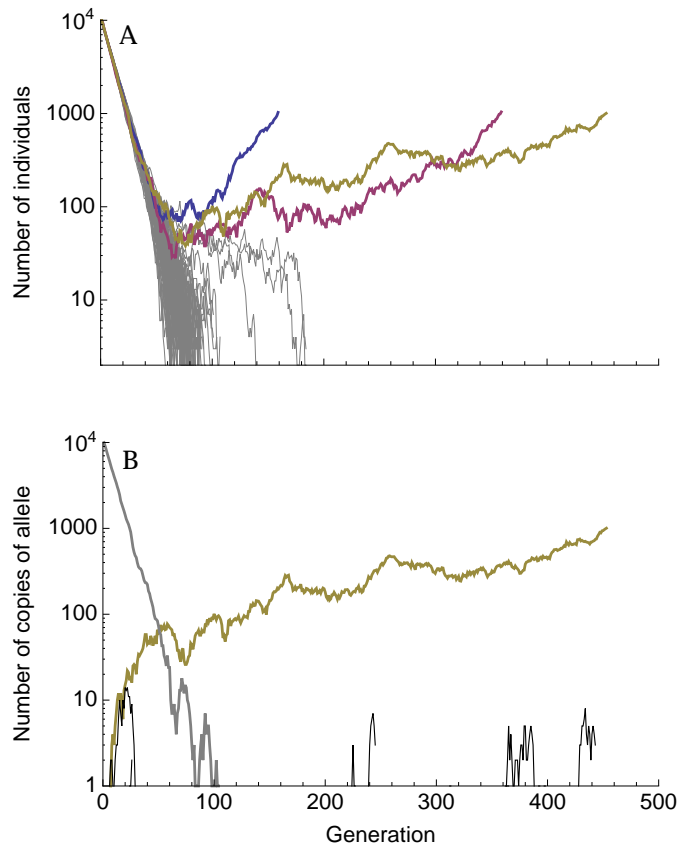


Figure 1 Typical dynamics with a relatively slow wildtype decline and a small mutation rate ($m_0 = -0.1$, $U = 10^{-4}$). **(A)** Population size trajectories on a log scale. Each line is a unique replicate simulation (100 replicates). Replicates that went extinct are grey, replicates that were rescued are in colour (and are roughly V-shaped). **(B)** The number of individuals with a given derived allele, again on a log scale, for the yellow replicate in A. The number of individuals without any derived alleles (wildtypes) is shown in grey, the rescue mutation is shown in yellow, and all other mutations are shown in black. Other parameters: $n = 4$, $\lambda = 0.005$, $m_{max} = 0.5$.

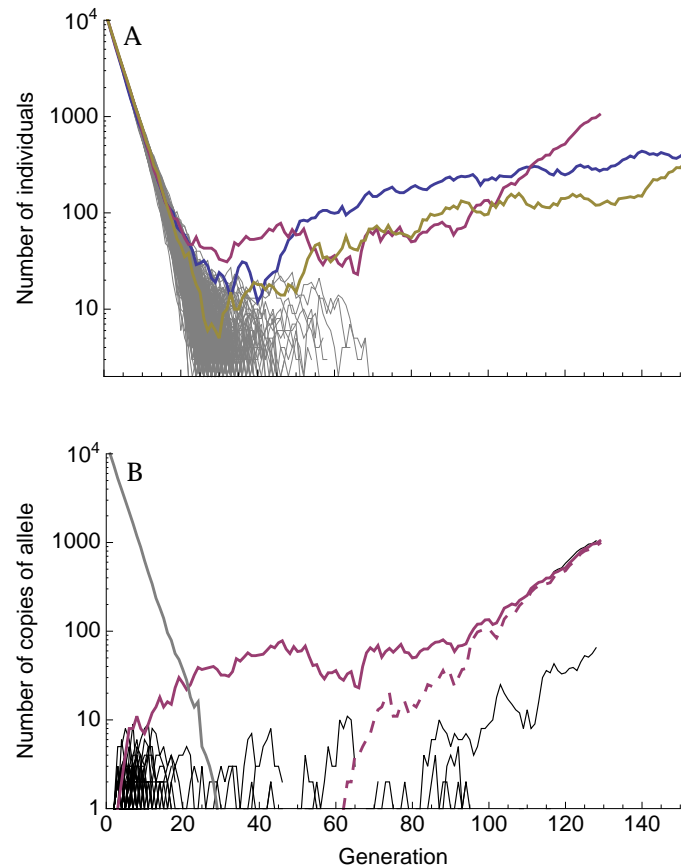


Figure 2 Typical dynamics with a relatively fast wildtype decline and a large mutation rate ($m_0 = -0.3$, $U = 10^{-2}$). **(A)** Population size trajectories on a log scale. Each line is a unique replicate simulation (500 replicates). Replicates that went extinct are grey, replicates that were rescued are in colour. Note that the blue and red replicates are cases of 2-step rescue (and roughly U-shaped), while the yellow replicate is 1-step rescue (and therefore V-shaped). **(B)** The number of individuals with a given derived allele, again on a log scale, for the red replicate in A. The number of individuals without any derived alleles (wildtypes) is shown in grey, the rescue mutations are shown in red, and all other mutations in black. Here a single mutant with growth rate less than zero arises early and outlives the wildtype (solid red). A second mutation then arises on that background (dashed red), making a double mutant with a growth rate greater than zero that rescues the population. Other parameters: $n = 4$, $\lambda = 0.005$, $m_{max} = 0.5$.

289 and Roques 2016), consistent with the House of Cards approx-
 290 imation (Turelli 1984, 1985), and thus rescue will occur by a
 291 single mutation of large effect (Anciaux et al. 2018). In the other
 292 extreme, when $U \gg U_C$, we are in a weak selection, strong
 293 mutation regime where many mutations arise on the same back-
 294 ground, creating a multivariate normal phenotypic distribution
 295 (Martin and Roques 2016), consistent with the Gaussian approx-
 296 imation (Kimura 1965; Lande 1980), and thus rescue will occur
 297 by many mutations of small effect (Anciaux et al. 2019). As
 298 shown in Figure 3 (where $U = U_C/10$) and Figure S1 (which
 299 spans $U_C = 0.02$), rescue by a small number of mutations (but
 300 more than one) can become commonplace in the transition zone,
 301 $U \sim U_C$, where there are often a considerable number of coseg-
 302 regating mutations (e.g., Figure 2B, where $U = U_C/2$).

303 The probability of k -step rescue

304 Approximations for the probability of 1-step rescue under the
 305 strong selection, weak mutation regime were derived by Anci-
 306 aux et al. (2018). Here we extend this study by exploring the
 307 contribution of k -step rescue, deriving approximations for the

308 probability of such events, as well as dissecting the genetic basis
 309 of both 1- and 2-step rescue in terms of the distribution of fitness
 310 effects of rescue genotypes and their component mutations.

311 Although requiring a sufficiently beneficial mutations to arise
 312 on a rare mutant genotypes doomed to extinction, multi-step
 313 evolutionary rescue can be the dominant form of rescue when
 314 the wildtype is sufficiently maladapted (Figures 3 and S1). In-
 315 deed, on this fitness landscape, the probability of producing
 316 a rescue genotype in one mutational step mutant drops very
 317 sharply with maladaptation (Anciaux et al. 2018); mutants with
 318 intermediate growth rates can thus be a “springboard” – albeit
 319 not always a very bouncy one – from which rescue mutants are
 320 produced. These intermediates contribute more as mutation

321 rates and the decline rate of the wildtype increase (Figures 3
322 and S1); the former because double mutants become more likely
323 and the latter because the springboard becomes more necessary.
324 With a large enough number of wildtype individuals or a high
325 enough mutation rate (Figure S1), multi-step rescue can not only
326 be more likely than 1-step, but also very likely in an absolute
327 sense.

328 **Alternative regimes of 2-step rescue**

329 2-step rescue can occur through first-step mutants with a wide
330 range of growth rates. As shown below, these first-step mutants
331 can be divided into three regimes: "sufficiently subcritical",
332 "sufficiently critical", and "sufficiently supercritical" (we will of-
333 ten drop "sufficiently" for brevity; Figure 4). Critical first-step
334 mutants are defined by having growth rates close enough to
335 zero that occasionally such a mutation will, by chance, persist
336 for long enough that it will almost certainly produce successful
337 double mutants. Subcritical first-step mutants are then defined
338 by having growth rates that are negative enough to prevent such
339 long persistence times. Their dynamics are roughly determin-
340 istic; mutations conferring growth rates closer to zero persist
341 longer but are less likely to arise from the wildtype. Similarly,
342 supercritical first-step mutants are defined by having positive
343 growth rates that are large enough to prevent long persistence
344 times once conditioned on extinction; conditioned on extinc-
345 tion these genotypes behave like subcritical mutations with a
346 growth rate of the same absolute value (Maruyama and Kimura
347 1974). As with subcritical first-step mutants, the dynamics of
348 supercritical mutants are relatively deterministic, but in this case
349 persistence times and mutational input from the wildtype both
350 favour growth rates closer to zero.

351 The relative contribution of each regime changes with both
352 the initial degree of maladaptation and the mutation rate (Fig-
353 ures 5 and S2). When the wildtype is very maladapted (relative
354 to mutational variance), most 2-step rescue events occur through
355 subcritical first-step mutants (Figure 5A), which are more abun-
356 dant than critical or supercritical mutants and yet persist longer
357 than the wildtype. If the initial maladaptation is milder, how-
358 ever, the slow decline of the wildtype buys time for critical and
359 supercritical mutations to arise and contribute to 2-step rescue,
360 and also increases the rate at which they are produced. The
361 rate of mutation also plays an interesting role in determining
362 the relative contributions of each regime (Figures 5B and S2).
363 When mutations are rare, only first-step mutations that are very
364 nearly neutral ($m \sim 0$) will persist long enough to give rise to a
365 2-step rescue mutation. As the mutation rate increases, however,
366 the range of first-step mutant growth rates that can persist long
367 enough to lead to 2-step rescue widens (because fewer individ-
368 uals carrying the first-step mutation are needed to produce a
369 successful double mutant).

370 **The distribution of fitness effects among rescue mutations**

371 Mutants causing 1-step rescue have growth rates that cluster
372 around small positive values ($m \gtrsim 0$; Figure 6). Consequently,
373 the distribution of fitness effects (DFE) is shifted to the right
374 relative to the constant population size case, beginning at $s =$
375 $m - m_0 \geq -m_0 > 0$. As a result of this increased threshold,
376 the 1-step rescue DFE has a smaller variance than both the DFE
377 of random mutations and the DFE of mutations that establish
378 in a constant population (Kimura 1983). Further, unlike these
379 other distributions, the variance in the DFE under 1-step rescue
380 decreases as the rate of wildtype decline increases, due to rescue

being restricted to a smaller proportion of the available mutants.

382 The DFE of genotypes that cause 2-step rescue (the combined
383 effect of two mutations) is also clustered at small positive values,
384 but it has a variance that is less affected by the rate of wildtype
385 decline (Figure 6). This is because double mutant rescue geno-
386 types are created via first-step mutant genotypes that have larger
387 growth rates than the wildtype (i.e., are closer to the optimum),
388 allowing them to create double mutants with a larger range of
389 positive growth rates.

390 Finally, we can also look at the distribution of growth rates
391 among first-step mutations that lead to 2-step rescue (Figures
392 7 and S2), i.e., 'springboard mutants'. Here there are two main
393 factors to consider: 1) the probability that a mutation with a
394 given growth rate arises on the wildtype background but does
395 not by itself rescue the population and 2) the probability that
396 such a mutation persists long enough for a sufficiently beneficial
397 second mutation to arise on that same background and together
398 rescue the population. As explained above, increasing the rate of
399 wildtype decline (or decreasing the rate of mutation) shifts the
400 contribution from critical first-step mutants to subcritical first-
401 step mutants, lowering the mode and increasing the variance.

402 Finally, note that, given 2-step rescue, the growth rate of both
403 the first and second mutation may be negative when alone in
404 the wildtype background. This potentially obscures empirical
405 detection of the individual mutations involved in evolutionary
406 rescue.

407 **Mathematical Analysis**

408 **The probability of k -step rescue**

409 Generic expressions for the probability of 1- and 2-step rescue
410 were given by Martin *et al.* (2013), using a diffusion approxima-
411 tion of the underlying demographics. The key result that we
412 will use is the probability that a single copy of a genotype with
413 growth rate m , itself fated for extinction but which produces
414 rescue mutants at rate $\Lambda(m)$, rescues the population (equation
415 S1.5 in Martin *et al.* 2013). With our lifecycle this is (c.f., equation
416 A.3 in Iwasa *et al.* 2004a)

$$417 \quad p(m, \Lambda(m)) = 1 - \exp \left[|m| \left(1 - \sqrt{1 + \frac{2\Lambda(m)}{m^2}} \right) \right]. \quad (3)$$

418 We can therefore use $p_0 = p(m_0, \Lambda(m_0))$ as the probability that a
419 wildtype individual has descendants that rescue the population
420 and what remains in calculating the total probability of rescue
421 (Equation 2) is $\Lambda(m_0)$. We break this down by letting $\Lambda_i(m)$ be
422 the rate at which rescue genotypes with i mutations are created;
423 the total probability of rescue is then given by Equation 2 with
424 $p_0 = p(m_0, \sum_{i=1}^{\infty} \Lambda_i(m_0))$.

425 In 1-step rescue, $\Lambda_1(m_0)$ is just the rate of production of res-
426 cue mutants directly from a wildtype genotype. This is the
427 probability that a wildtype gives rise to a mutant with growth
428 rate m (given by $Uf(m|m_0)$) times the probability that a geno-
429 type with growth rate m establishes. Again approximating our
430 discrete time process with a diffusion process, the probability
431 that a lineage with growth rate $m \ll 1$ establishes, ignoring
further mutation, is (Martin *et al.* 2013)

$$432 \quad p_{est}(m) \approx \begin{cases} 0 & m \leq 0 \\ 1 - \exp(-2m) & m > 0 \end{cases}. \quad (4)$$

433 This reduces to the $2(s + m_0)$ result in Otto and Whitlock (1997)
when $m = s + m_0$ is small, which further reduces to $2s$ in a

Symbol	Meaning
n	number of (scaled) phenotypic dimensions
λ	variance in mutant phenotypes along each dimension
m_{max}	maximum growth rate
$f(m' m)$	distribution of growth rates among mutants from a genotype with growth rate m (eq. 1)
U	per genome mutation probability
N_0	initial number of wildtype individuals
m_0	wildtype growth rate
p_0	probability that an initial wildtype individual has descendants that rescue the population
P	probability of rescue (eq. 2)
$p(m, \Lambda(m))$	probability a genotype with growth rate m , itself fated for extinction, has descendants that rescue the population (eq. 3)
$p_{est}(m)$	probability a genotype with growth rate m establishes, i.e., rescues the population (eq. 4)
$\Lambda(m)$	probability that an individual with growth rate m produces a mutant that has descendants that rescue the population
$\Lambda_i(m)$	probability that an individual with growth rate m produces a mutant that has descendants with $i - 1$ additional mutations that rescue the population
$\Lambda_2^{(-)}(m), \Lambda_2^{(0)}(m), \Lambda_2^{(+)}(m)$	probability that an individual with growth rate m produces sufficiently subcritical, critical, or supercritical first-step mutants that eventually lead to 2-step rescue (eq. 8)
ψ	$2(1 - \sqrt{1 - m/m_{max}})$
ψ_0	$2(1 - \sqrt{1 - m_0/m_{max}})$
ρ_{max}	m_{max}/λ
α	$\rho_{max}\psi_0^2/4$

Table 1 Frequently used notation.

434 population of constant size, $m_0 = 0$ (Haldane 1927). Using this,
435 the rate of 1-step rescue is

$$\Lambda_1(m_0) = U \int_0^{m_{max}} f(m|m_0) p_{est}(m) dm. \quad (5)$$

436 Taking the first order approximation of $p(m_0, \Lambda_1(m_0))$ with
437 $\Lambda_1(m_0)/m_0^2$ small gives the probability of 1-step rescue given in
438 equation 5 of [Anciaux et al. \(2018\)](#), which effectively assumes
439 deterministic wildtype decline. For completeness we rederive
440 their closed-form approximation in File S1 (and give the results
441 in the Appendix, see [Approximating the probability of 1-step
442 rescue](#)).

443 The probability of 2-step rescue is only slightly more compli-
444 cated. Here $\Lambda_2(m_0)$ is the probability that a mutation arising on
445 the wildtype background creates a genotype that is also fated
446 for extinction but persists long enough for a second mutation

447 to arise on this mutant background, creating a double mutant
448 genotype that rescues the population. We therefore have

$$\Lambda_2(m_0) = U \int_{-\infty}^{m_{max}} f(m|m_0) [1 - p_{est}(m)] p(m, \Lambda_1(m)) dm. \quad (6)$$

449 Following this logic, we can retrieve the probability of k -step
450 rescue, for arbitrary $k \geq 2$, using the recursion

$$\Lambda_k(m_0) = U \int_{-\infty}^{m_{max}} f(m|m_0) [1 - p_{est}(m)] p(m, \Lambda_{k-1}(m)) dm, \quad (7)$$

451 with the initial condition given by Equation 5.

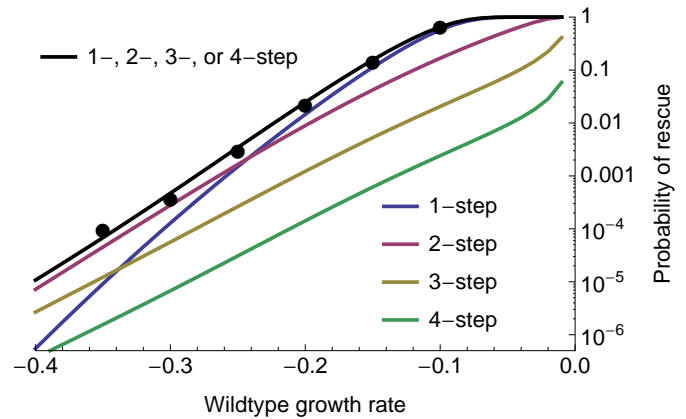


Figure 3 The probability of evolutionary rescue as a function of initial maladaptation. Shown are the probabilities of 1-, 2-, 3-, and 4-step rescue (Equations 2-7), as well as the probability of rescue by up to 4 mutational steps (using $\Lambda(m_0) = \sum_{i=1}^4 \Lambda_i(m_0)$). Dots are individual-based simulation results (ranging from 10^3 to 10^5 replicates per dot), allowing any number of mutations. Simulated populations were considered rescued when there were 1000 individuals with positive growth rates. Parameters: $N_0 = 10^4$, $U = 2 \times 10^{-3}$, $n = 4$, $\lambda = 0.005$, $m_{max} = 0.5$.

452 Approximating the probability of 2-step rescue

453 The probability of 2-step rescue is given by Equation 2 with
454 $p_0 = p(m_0, \Lambda_2(m_0))$ (Equations 3-6). We next develop some
455 intuition by approximating this for different classes of single
456 mutants.

457 First, note that when the growth rate of a first-step mutation
458 is close enough to zero such that $m^2 \ll \Lambda_1(m)$, we can approx-
459 imate the probability such a genotype leads to rescue before
460 itself going extinct, $p(m, \Lambda_1(m))$, with $\sqrt{2\Lambda_1(m)}$ (c.f. equation
461 A.4b in [Iwasa et al. 2004a](#)). As shown in the Appendix (see [Mutant lineage dynamics](#)), while $t < 1/|m|$ a mutant lineage with
462 growth rate m that is destined for extinction persists for t gener-
463 ations with probability $\sim 2/t$ (Equation 21) and in generation
464 t since it has arisen has $\sim t/2$ individuals (Equation 22). Thus,
465 while $T < 1/|m|$ a mutant lineage that persists for T generations
466 will have produced a cumulative number $\sim T^2/4$ individuals.
467 Such lineages will then lead to 2-step rescue with probability
468 $\sim \Lambda_1(m)T^2/4$ until this approaches 1, near $T = 2/\sqrt{\Lambda_1(m)}$.
469 Since the probability of rescue increases like T^2 while the proba-
470 bility of persisting to time T declines only like $1/T$, most rescue

472 events will be the result of rare long-lived single mutant geno-
 473 types. Considering only the most long-lived genotypes, the prob-
 474 ability a first-step mutation leads to rescue is then the probability
 475 it survives long enough to almost surely rescue, $\sim 2/\sqrt{\Lambda_1(m)}$
 476 generations. Thus, for first-step mutants with growth rates satis-
 477 fying $2/\sqrt{\Lambda_1(m)} < 1/|m|$, implying $m^2 \ll \Lambda_1(m)$, such that
 478 they can by chance persist for unusually long times, the probabil-
 479 ity they lead to rescue is $\sim \sqrt{\Lambda_1(m)}$, consistent with our Taylor
 480 series approximation of $p(m, \Lambda_1(m))$. This same reasoning has
 481 been used to explain why the probability a neutral mutation
 482 segregates long enough to produce a second mutation is $\sim \sqrt{U}$
 483 in a population of constant size (Weissman *et al.* 2009).

484 At the other extreme, when the growth rate of a first-step
 485 mutation is far enough from zero such that $m^2 \gg \Lambda_1(m)$, we
 486 can approximate $p(m, \Lambda_1(m))$ with $\Lambda_1(m)/|m|$ (c.f. equation
 487 A.4c in Iwasa *et al.* 2004a). As just discussed, conditioned on
 488 extinction such genotypes cannot persist long enough to almost
 489 surely lead to 2-step rescue. Instead, we expect such mutations
 490 to persist for at most $\sim 1/|m|$ generations (Equation 21) with
 491 a lineage size of ~ 1 individual per generation (Equation 22),
 492 and thus produce a cumulative total of $\sim 1/|m|$ individuals.
 493 The probability of 2-step rescue from such a first-step mutation
 494 is therefore $\Lambda_1(m)/|m|$, again consistent with our Taylor series
 495 approach. This is the same reasoning that explains why a rare
 496 mutant genotype with selection coefficient $|s| \gg 0$ in a constant
 497 population size model is expected to have a cumulative number
 498 $\sim 1/|s|$ individuals, given it eventually goes extinct (Weissman
 499 *et al.* 2009).

500 The transitions between these two regimes occur when
 501 $\Lambda_1(m)/|m| = \sqrt{2\Lambda_1(m)}$, i.e., when $|m| = \sqrt{\Lambda_1(m)}/2$. We
 502 call single mutants with growth rates $m < -\sqrt{\Lambda_1(m)}/2$ "suf-
 503 ficiently subcritical", those with $|m| < \sqrt{\Lambda_1(m)}/2$ "suffi-
 504 ciently critical", and those with $m > \sqrt{\Lambda_1(m)}/2$ "suffi-
 505 ciently supercritical". Given that U and thus $\Lambda_1(m)$ will generally be small, m
 506 will also be small at these transition points, meaning we can
 507 approximate them with $m^* = \sqrt{\Lambda_1(0)}/2$ and $-m^*$. We then
 508 have an approximation for the rate of 2-step rescue,

$$\begin{aligned} \Lambda_2(m_0) &= \Lambda_2^{(-)}(m_0) + \Lambda_2^{(0)}(m_0) + \Lambda_2^{(+)}(m_0) \\ \Lambda_2^{(-)}(m_0) &= U \int_{-\infty}^{-m^*} f(m|m_0) \Lambda_1(m)/|m| dm \\ \Lambda_2^{(0)}(m_0) &= U \int_{-m^*}^{m^*} f(m|m_0) [1 - p_{est}(m)] \sqrt{2\Lambda_1(m)} dm \\ \Lambda_2^{(+)}(m_0) &= U \int_{m^*}^{m_{max}} f(m|m_0) [1 - p_{est}(m)] \Lambda_1(m)/|m| dm \end{aligned} \quad (8)$$

509 where $\Lambda_2^{(-)}(m_0)$ is the rate of 2-step rescue through sufficiently
 510 subcritical first-step mutants, $\Lambda_2^{(0)}(m_0)$ is the rate of 2-step res-
 511 cue through sufficiently critical first-step mutants, and $\Lambda_2^{(+)}(m_0)$
 512 is the rate of 2-step rescue through sufficiently supercritical first-
 513 step mutants. A schematic depicting the 1- and 2-step genetic
 514 paths to rescue is given in Figure 4.

515 **Closed-form approximation for sufficiently critical rescue**
 516 When U is small m^* is also small, allowing us to use $m = 0$
 517 within the integrand of $\Lambda_2^{(0)}(m_0)$, giving

$$\begin{aligned} \Lambda_2^{(0)}(m_0) &\approx U f(0|m_0) \sqrt{2\Lambda_1(0)} 2m^* \\ &= 2U f(0|m_0) \Lambda_1(0). \end{aligned} \quad (9)$$

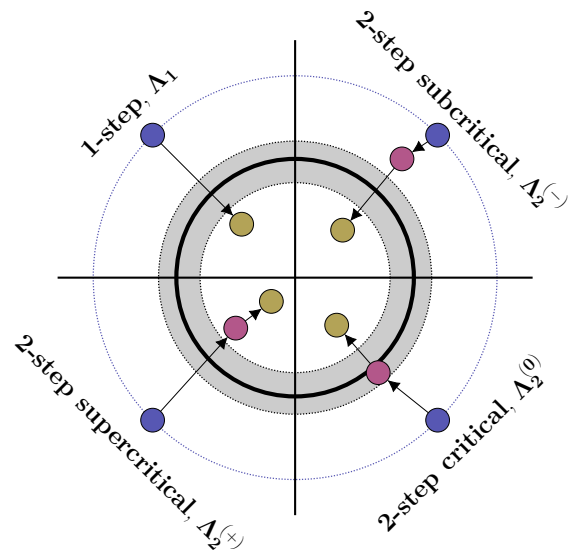


Figure 4 1- and 2-step genetic paths to evolutionary rescue. Here we show an $n = 2$ dimensional phenotypic landscape. Continuous-time (Malthusian) growth rate (m) declines quadratically from the centre, becoming negative outside the thick black line. The grey zone indicates where growth rates are "sufficiently critical" (see text for details). Blue circles show wildtype phenotypes, red circles show intermediate first-step mutations, and yellow circles show the phenotypes of rescue genotypes.

518 We can then approximate $\Lambda_1(m)$ with $\tilde{\Lambda}_1(m)$ (Equation 19) and
 519 take $m \rightarrow 0$ (Equation 20), giving a closed form approximation
 520 for the rate of 2-step rescue through critical single mutants in
 521 Fisher's geometric model

$$\Lambda_2^{(0)}(m_0) \approx 4U^2 f(0|m_0) \sqrt{m_{max}\lambda/\pi}. \quad (10)$$

522 This well approximates numerical integration of $\Lambda_2^{(0)}(m_0)$ (Equa-
 523 tion 8; see Figure 5 and File S1). In general, it will perform better
 524 as the range of critical growth rates, and thus $U\sqrt{m_{max}\lambda}$, be-
 525 comes smaller.

526 To get a better understanding of how the rate of 2-step critical
 527 rescue depends on the underlying parameters of Fisher's
 528 geometric model, we approximate $f(m|m_0)$, assuming $\rho_{max} =$
 529 m_{max}/λ is large, and convert this to a distribution over $\psi =$
 530 $2(1 - \sqrt{1 - m/m_{max}})$, a convenient rescaling (for details see File
 531 S1 and Anciaux *et al.* 2018). Evaluating this at $m = 0$ gives

$$\Lambda_2^{(0)}(m_0) \approx U^2 (1 - \psi_0/2)^{(1-n)/2} e^{-\alpha} \frac{2}{\pi}, \quad (11)$$

532 where $\psi_0 = 2(1 - \sqrt{1 - m_0/m_{max}})$ and $\alpha = \rho_{max}\psi_0^2/4$.

533 **Closed-form approximations for sufficiently non-critical res-**
 534 **cue** We can also approximate $\Lambda_1(m)$ in $\Lambda_2^{(-)}(m_0)$ and $\Lambda_2^{(+)}(m_0)$
 535 with $\tilde{\Lambda}_1(m)$ (Equation 19), leaving us with just one integral over
 536 the growth rates of the first-step mutations. We then replace
 537 $f(m|m_0)$ with its approximate distribution over ψ .

538 In the case of subcritical rescue we can then make two con-
 539 trasting approximations. First, when the ψ (and thus m) that
 540 contribute most are close enough to zero (meaning maladapt-
 541 ation is not too large relative to mutational variance) and we
 542 ignore mutations that are less fit than the wildtype, we have

$$\Lambda_2^{(-)}(m_0) \approx U^2 \frac{(1 - \psi_0/2)^{1-n}}{1 - \psi_0/4} e^{-\alpha} \frac{\log(\psi_0/\psi_-^*)}{\pi}, \quad (12)$$

543 where $\psi_-^* = 2(1 - \sqrt{1 + \tilde{m}^*/m_{max}})$ and $\tilde{m}^* = \sqrt{\tilde{\Lambda}_1(0)}/2$. Sec-
544 ond, when the mutational variance, λ , is very small relative to
545 maladaptation, implying that mutants far from $m = 0$ substan-
546 tially contribute, we find

$$\Lambda_2^{(-)}(m_0) \approx -U^2 \frac{(1 - \psi_0/2)^{1-n}}{1 - \psi_0/4} \left(e^{-\alpha} \frac{1}{(\alpha/2)^3 \pi} \right)^{1/2}. \quad (13)$$

547 These two approximations do well compared with $\Lambda_2^{(-)}(m_0)$
548 (Equation 8; see Figure 5 and File S1). As expected, we find
549 that Equation 13 does better under fast wildtype decline while
550 Equation 12 does better when the wildtype is declining more
551 slowly.

552 For supercritical 2-step rescue, only first-step mutants with
553 growth rates near m^* will contribute (larger m will rescue them-
554 selves and are also less likely to arise by mutation), and so we
555 can capture the entire distribution with a small m approximation
556 (following the same approach that led to Equation 12). As shown
557 in File S1, this approximation works well for $\psi < \sqrt{2/\rho_{max}}$, be-
558 yond which the rate of 2-step rescue through such first-step
559 mutants falls off very quickly due to a lack of mutational input.
560 Thus, considering only supercritical single mutants with scaled
561 growth rate less than $\sqrt{2/\rho_{max}}$, our approximation is

$$\Lambda_2^{(+)}(m_0) \approx U^2 \frac{(1 - \psi_0/2)^{1-n}}{1 - \psi_0/4} e^{-\alpha} \frac{\log(\psi_{max}/\psi_+^*)}{\pi}, \quad (14)$$

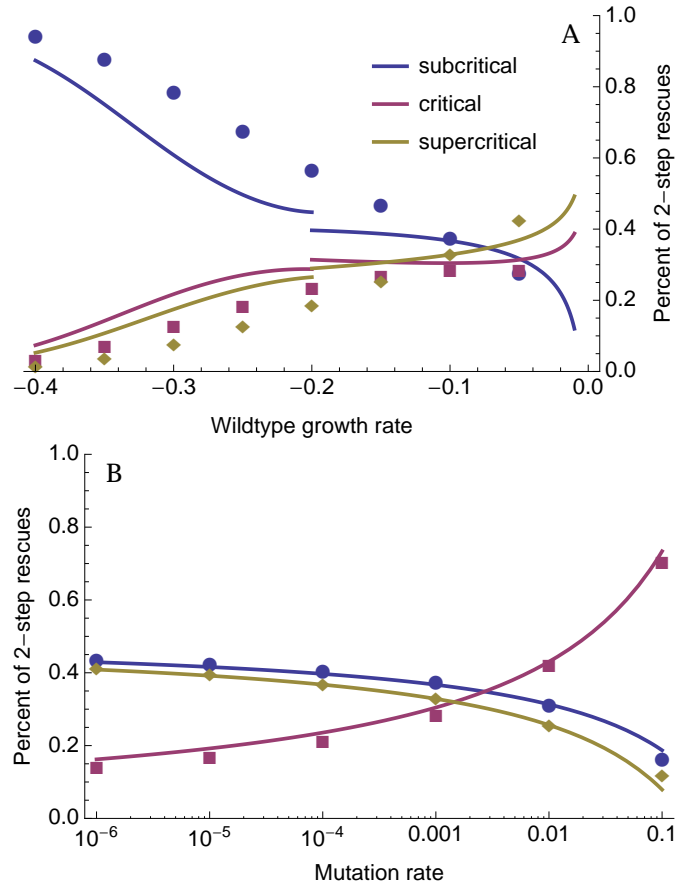
562 with $\psi_+^* = 2(1 - \sqrt{1 - \tilde{m}^*/m_{max}})$ and $\psi_{max} = \sqrt{2/\rho_{max}}$.
563 This approximation tends to provide a slight overestimate of
564 $\Lambda_2^{(+)}(m_0)$ (Equation 8; see Figure 5 and File S1).

565 **Comparing 2-step regimes** These rough but simple closed-form
566 approximations (Equations 11–14) show that, while the contri-
567 bution of critical mutants to 2-step rescue scales with U^2 , the
568 contribution of non-critical single mutants scales at a rate less
569 than U^2 due to an increase in the range of critical single mutants
570 (both ψ_-^* and ψ_+^* increase with U). This difference in scaling
571 with U is stronger when the wildtype is not very maladapted
572 relative to the mutational variance, i.e., when Equation 12 is the
573 better approximation for subcritical rescue. The approximations
574 also show that, when initial maladaptation is small the ratio of
575 supercritical to subcritical contributions (Equation 12 divided
576 by 14) primarily depends only on the range of growth rates in-
577 cluded in each regime, while with larger initial maladaptation
578 this ratio (Equation 13 divided by 14) begins to depend more
579 strongly on initial maladaptation and mutational variance (α).
580 The effect of maladaptation and mutation rate on the relative
581 contributions of each regime is shown in Figure 5.

582 The distribution of growth rates among rescue genotypes

583 We next explore the distribution of growth rates among rescue
584 genotypes, i.e., the distribution of growth rates that we expect
585 to observe among the survivors across many replicates.

586 We begin with 1-step rescue. The rate of 1-step rescue by
587 genotypes with growth rate m is simply $Uf(m|m_0)p_{est}(m)$. Di-
588 viding this by the rate of 1-step rescue through any m (Equation
589 5) gives the distribution of growth rates among the survivors



590 **Figure 5** The relative contribution of sufficiently subcritical,
591 sufficiently critical, and sufficiently supercritical single mutants
592 to 2-step rescue. The curves are drawn using Equations
593 10–14 (Equation 12 is used for $m_0 < 0.2$ while Equation 13
594 is used for $m_0 > 0.2$). The dots are numerical evaluations of
595 Equation 8. Parameters: $n = 4$, $\lambda = 0.005$, $m_{max} = 0.5$, (A)
596 $U = 10^{-3}$, (B) $m_0 = -0.1$.

$$g_1(m) = \frac{Uf(m|m_0)p_{est}(m)}{\Lambda_1(m_0)}, \quad (15)$$

597 where U cancels out. This distribution is shown in blue in Figure
598 6. The distribution has a mode at small but positive m as a
599 result of two conflicting processes: smaller growth rates are
600 more likely to arise from a declining wildtype but larger growth
rates are more likely to establish given they arise. As the rate
of wildtype decline increases, the former process exerts more
influence, causing the mode to move towards zero and reducing
the variance.

We can also give a closed form approximation here using the
same approach taken to reach Equation 19. On the ψ scale we
have

$$\tilde{g}_1(\psi) = \frac{\exp(\alpha)\sqrt{\alpha\rho_{max}}}{[\exp(\alpha)\sqrt{\pi\alpha}\text{Erfc}(\sqrt{\alpha}) - 1]}\psi_0 e^{-\rho_{max}(\psi-\psi_0)^2/4}\psi, \quad (16)$$

601 implying the ψ are distributed like a normal truncated below
602 $\psi = 0$ and weighted by ψ . This provides a very good approxi-
603 mation (see dashed blue curves in Figure 6).

604 In 2-step rescue, the rate of rescue by double mutants with
605 growth rate m_2 is given by Equation 6 with $\Lambda_1(m)$ replaced

606 by $Uf(m_2|m)p_{est}(m_2)$. Normalizing gives the distribution of
 607 growth rates among the double mutant genotypes that rescue
 608 the population

$$g_2(m_2) \approx \frac{A(m_2)}{\int_0^{m_{max}} A(m_2) dm_2}$$

$$A(m_2) = \int_{-\infty}^{m_{max}} f(m|m_0) [1 - p_{est}(m)] p(m, Uf(m_2|m)p_{est}(m_2)) dm. \quad (17)$$

609 This distribution, $g_2(m)$, is shown in red in Figure 6. Because the
 610 first-step mutants contributing to 2-step rescue tend to be nearer
 611 the optimum than the wildtype, this allows them to produce
 612 double mutant rescue genotypes with higher growth rates than
 613 in 1-step rescue (as seen by comparing the mode between blue
 614 and red curves in Figure 6). The fact that these first-step mutants
 615 are closer to the optimum also allows for a greater variance in
 616 the growth rates of rescue genotypes than in 1-step rescue. This
 617 also allows the 2-step distribution to maintain a more similar
 618 mode and variance across wildtype decline rate than the 1-step
 619 distribution. Note that because the U in $Uf(m_2|m)p_{est}(m_2)$ does
 620 not cancel, $g_2(m_2)$ depends on U and the buffering effect of
 621 first-step mutants depends on the mutation rate. In principle,
 622 decreasing the mutation rate disproportionately increases the
 623 contribution of first-step mutants with growth rates closer to
 624 zero, as these genotypes are more likely to persist long enough
 625 to produce rescue mutants (see [The distribution of growth rates among rescue intermediates](#) below for more discussion). This
 626 implies that the distribution of rescue genotype growth rates
 627 is more strongly buffered from the rate of wildtype decline at
 628 lower mutation rates.

630 **The distribution of growth rates among rescue intermediates**

631 Finally, our analyses above readily allow us to explore the distri-
 632 bution of first-step mutant growth rates that contribute to 2-step
 633 rescue. Analogously to Equation 15, we drop the integral in
 634 $\Lambda_2(m_0)$ (Equation 6) and normalize, giving

$$h(m) = \frac{Uf(m|m_0) [1 - p_{est}(m)] p(m, \Lambda_1(m))}{\Lambda_2(m_0)}, \quad (18)$$

635 where the first U cancels but the U within $\Lambda_1(m)$ does not. This
 636 distribution is shown in black in Figure 7. At slow wildtype
 637 decline rates the overwhelming majority of 2-step rescue events
 638 arise from first-step mutants with growth rates near 0. As in-
 639 dicated by Equation 8, the contribution of first-step mutants
 640 with growth rate m declines as $\sim 1/|m|$ as m departs from zero,
 641 due to shorter persistence times given eventual extinction. As
 642 wildtype growth rate declines the relative importance of muta-
 643 tional input, $f(m|m_0)$, grows, causing the distribution to flatten
 644 and first-step mutants with substantially negative growth rates
 645 begin to contribute (see also Figure 5A). Decreasing the muta-
 646 tion rate disproportionately increases the contribution of first-
 647 step mutants with growth rates near zero (while simultaneously
 648 shrinking the range of growth rates that are sufficiently critical;
 649 Figure 5B) making the distribution of first-step mutant growth
 650 rates contributing to 2-step rescue more sharply peaked around
 651 $m = 0$ (Figure S2). Correspondingly, with a higher mutation
 652 rate a greater proportion of the contributing single mutants have
 653 substantially negative growth rates.

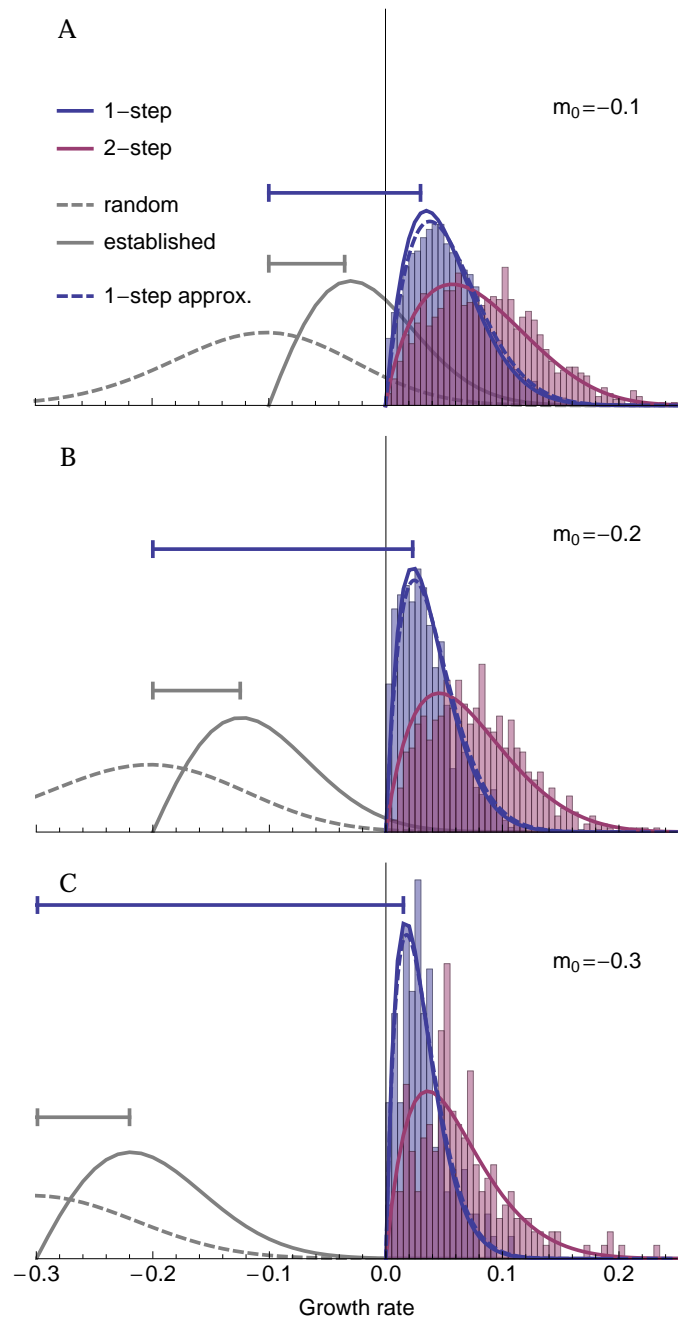


Figure 6 The distribution of growth rates among rescue genotypes under 1-step (blue; Equation 15 solid and 16 dashed) and 2-step (red; Equation 17) rescue for three different levels of initial maladaptation. For comparison, the distribution of random mutations (dashed; Equation 1) and the distribution of beneficial mutations that establish in a population of constant size (solid grey; Equation 1 times Equation 4 and normalized) are shown. Horizontal lines indicate the most common fitness effect ($s = m_0 - m$) in a population of constant size (grey) and in 1-step rescue (blue). The histograms show the distribution of growth rates among rescue genotypes observed across (A) 10^4 , (B) 10^5 , and (C) 10^6 simulated replicates. Populations were considered rescued once there were ≥ 100 individuals with positive growth rate. The most common genotype at this point was considered the rescue genotype, and the number of mutational steps to rescue was set as the number of mutations in that genotype. Other parameters: $N_0 = 10^4$, $U = 2 \times 10^{-3}$, $n = 4$, $\lambda = 0.005$, $m_{max} = 0.5$.

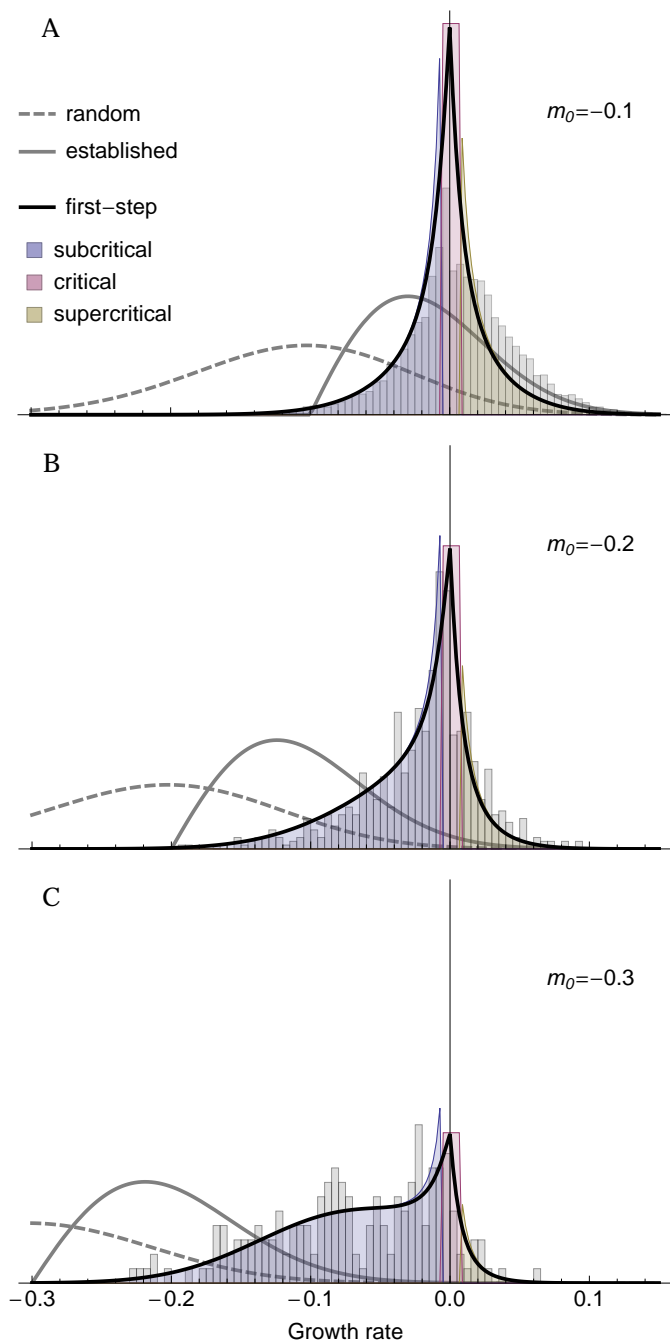


Figure 7 The distribution of growth rates among first-step mutations that lead to 2-step rescue (black; Equation 18) for three different levels of initial maladaptation. Shading represents our sufficiently subcritical approximation (blue; replacing $p(m, \Lambda_1(m))$ with $\Lambda_1(m)/|m|$ in the numerator of Equation 18), our sufficiently critical approximation (red; using $Uf(0|m_0)\sqrt{2\Lambda_1(0)}$ as the numerator in Equation 18), and our sufficiently supercritical approximation (yellow; replacing $p(m, \Lambda_1(m))$ with $\Lambda_1(m)/|m|$ in the numerator of Equation 18). The histograms show the distribution of growth rates among first-step mutations in rescue genotypes with 2 mutations observed across (A, B) 10^5 or (C) 10^6 simulated replicates. Note that the discrepancy in panel A is likely due to us sampling only the most common rescue genotype in each replicate. See Figure 6 for additional details.

654 Discussion

655 Here we've explored the possibility of evolutionary rescue by
 656 multiple mutations on a simple fitness landscape. We find that
 657 rescue by multiple mutations can be the most likely path to per-
 658 sistence under high mutation rates or when the population is
 659 initially very maladapted. Under these scenarios, intermediate
 660 genotypes that are declining less quickly provide a 'springboard'
 661 from which rescue genotypes emerge. In 2-step rescue these
 662 springboard single mutants come from one of three regimes:
 663 those that have growth rates near enough to zero (i.e., replace-
 664 ment) to, by chance, persist for unusually long periods of time
 665 and therefore reach unusually large subpopulation sizes and
 666 those with growth rates either too negative or too positive to do
 667 so. The relative contribution of each regime shifts with initial
 668 maladaptation and mutation rate; rare mutations that can occa-
 669 sionally reach unusually large subpopulation sizes play a larger
 670 role as maladaptation declines and mutation increases. In con-
 671 trast, when initial maladaptation is very high most first-step
 672 mutations are themselves also very maladapted and thus restricted
 673 in the subpopulation sizes they reach. All three regimes help
 674 maintain the variance in the distribution of fitness effects among
 675 rescue genotypes as initial maladaptation increases; meanwhile,
 676 in 1-step rescue the variance declines due to ever more extreme
 677 sampling of the tail of the mutational distribution.

678 How often rescue arises as a result of multiple mutations is
 679 an open question. It is clear that more than one mutation can
 680 contribute to adaptation to near-lethal stress, but experiments
 681 are often designed to avoid extinction (reviewed in Cowen *et al.*
 682 2002), and therefore greatly expand the scope for multiple mu-
 683 tations to arise on a single genotype. A few exceptions provide
 684 some insight. For example, populations of *Saccharomyces cerevisiae*
 685 that survived high concentrations of copper acquired multiple
 686 mutations (Gerstein *et al.* 2015) – in fact the authors argue for the
 687 'springboard effect' discussed above, where first step mutations
 688 prolong persistence and thereby allow further mutations to arise.
 689 In *Pseudomonas fluorescens*, fluctuation tests with nalidixic acid
 690 showed that nearly a third of the most resistant surviving strains
 691 were double mutants (Bataillon *et al.* 2011), which were able
 692 to tolerate 10x higher drug concentrations than single mutants,
 693 suggesting 2-step rescue might dominate at high drug concen-
 694 trations. It is unclear if our prediction – that rescue takes more
 695 mutational steps with greater initial maladaptation – holds true.
 696 Verification will require more experiments that allow extinction
 697 and uncover the genetic basis of adaptation at different severities
 698 of environmental change (e.g., drug concentration).

699 In describing the genetic basis of adaptation in populations
 700 of constant size, Orr (1998) showed that the mean phenotypic
 701 displacement towards the optimum scales roughly linearly with
 702 initial displacement. Converting phenotype to fitness, this im-
 703 plies the mean fitness effect of fixed mutations, $s = m - m_0$,
 704 increases as $\sim \exp(-m_0)$ as initial Malthusian fitness, m_0 ,
 705 declines, which is roughly linear when m_0 is small. Here we see
 706 that, under 1-step rescue, the mean fitness effect also increases
 707 roughly linearly as the initial growth rate declines (see hori-
 708 zontal blue lines in Figure 6). However, the rate of this linear
 709 increase in fitness effect is much larger under rescue than in a
 710 population of constant size (compare blue and grey horizontal
 711 lines in Figure 6), where declines in wildtype fitness not only
 712 allow larger mutations to be beneficial but also require larger
 713 mutations for persistence. Thus the race between extinction and
 714 adaptation during evolutionary rescue is expected to produce a
 715 genetic basis of adaptation with fewer mutations of larger effect.

716 While under 1-step rescue the fitness effect of the first mu- 778
717 tation increases roughly linearly as wildtype fitness declines, 779
718 below some wildtype fitness most rescue events will be 2-step 780
719 (e.g., at $m_0 \approx -0.25$ in Figure 3). At this junction the effect size 781
720 of the first mutation will no longer increase as quickly (and po- 782
721 tentially even decrease), as its expected fitness will itself begin 783
722 to decline substantially with the fitness of the wildtype (Figures 784
723 5 and 7). Thus as rescue switches from dominantly k -step to 785
724 dominantly $(k + 1)$ -step the genetic basis of adaptation becomes 786
725 more diffuse, with each mutation having a smaller individual 787
726 fitness effect as the contributing fitness effects spread over more 788
727 loci. In the limit of large k (due to large initial maladaptation or 789
728 high mutation rates), the genetic basis of adaptation should at 790
729 some point converge to many loci with small effect, as would 791
730 also be expected in a population of constant size. Indeed, at 792
731 very high mutation rates the rate of adaptation (the change in 793
732 mean fitness) is the same under rescue as it is in populations 794
733 of constant size (Anciaux *et al.* 2019), implying that the genetic 795
734 basis of adaptation no longer depends on demography. It is 796
735 therefore at intermediate levels of initial maladaptation and low 797
736 mutation rates, where rescue primarily occurs from a few large 798
737 effect mutations, that the race between adaptation and persis- 799
738 tence is predicted to have the largest effect on the genetic basis 800
739 of adaptation.

740 Fluctuation tests (?) provide a means to isolate rescue geno- 802
741 types, whose growth rates can then be measured under the 803
742 selective conditions. Consistent with our theory (Figure 6), the 804
743 resulting growth rate distributions in both bacteria and yeast 805
744 often find modes that are substantially greater than zero (as op- 806
745 posed to, say, an exponential distribution; Kassen and Bataillon 807
746 2006; MacLean and Buckling 2009; Gerstein *et al.* 2012; Lindsey 808
747 *et al.* 2013; Gerstein *et al.* 2015). A number of these conform even 809
748 more closely to our expected shape (Kassen and Bataillon 2006; 810
749 Gerstein *et al.* 2015) while the others appear to be substantially 811
750 more clumped around the mode, perhaps due to a very restricted 812
751 number of possible rescue mutations in any one circumstance, 813
752 the size of the experiment, or the way in which growth rates are 814
753 measured. Further, these experiments are designed to begin with 815
754 substantial standing genetic variation (at least across replicates), 816
755 which should increase the contributions of mutations with small 817
756 growth rates (?), although these could be outcompeted by muta- 818
757 tions with higher growth rates and/or be undersampled. The 819
758 role of standing genetic variance becomes particularly important 820
759 when we consider rescue by multiple steps, as the ability of the 821
760 intermediate genotypes to persist long enough to accumulate 822
761 further mutations will differ strongly between environments. 823
762 Finally, Gerstein *et al.* (2015) not only provides the distribution 824
763 of growth rates among rescue genotypes, but also the growth 825
764 rates of individual mutations that compose multi-step rescue 826
765 genotypes. In two of three cases they see that most component 827
766 mutations have growth rates near that of the multi-mutation 828
767 genotype while in the other case there appears to be multiple 829
768 mutations that provide intermediate benefits, consistent with 830
769 their hypothesis of multi-step rescue. These growth rates were 831
770 measured in a less stressful environment than the original selec- 832
771 tive environment, which unfortunately makes it impossible to 833
772 tell if these individual mutations had positive or negative growth 834
773 rates (i.e., were subcritical or supercritical) during rescue.

774 Pinpointing the mutations responsible for adaptation is ham- 836
775 pered by genetic hitchhiking, as beneficial alleles elevate the fre- 837
776 quency of linked neutral and mildly deleterious alleles (Barton 838
777 2000). The problem is particularly severe under strong selection 839

and low recombination, and therefore reaches an extreme in 840
the case of evolutionary rescue in asexuals, especially if many 841
neutral and deleterious mutations are segregating at the time 842
of environmental change. To circumvent this, mutations that 843
have risen to high frequency in multiple replicates are often 844
introduced in a wildtype background, in isolation and some- 845
times also in combination with a small number of other common 846
high-frequency mutations, and grown under the selective condi- 847
tions (e.g., Jochumsen *et al.* 2016; Ono *et al.* 2017). As we have 848
demonstrated above, however, under multi-step rescue there 849
may be no one mutation that individually confers growth in 850
the selective conditions. Thus, a mutation driving rescue may 851
go undetected or be mistaken as a hitchhiker if the appropri- 852
ate multiple-mutation genotypes are not tested. Unfortunately 853
reverse engineering all combinations of mutations quickly be- 854
comes unwieldy as the number of mutations grows, and thus 855
this approach will not be practical under severe initial maladap- 856
tation and high mutation rates, where we predict rescue to occur 857
by many mutations. Interestingly, our simulations show that 858
the population dynamics themselves may help differentiate how 859
many mutations contribute to rescue (e.g., V- vs. U-shaped log- 860
trajectories; Figures 1 and 2), and fitting models of k -step rescue 861
could produce estimates for the growth rates of the k genotypes.

862 Environmental change often selects for mutator alleles, which 863
elevate the rate at which beneficial alleles arise and subsequently 864
increase in frequency with them (Tenaillon *et al.* 2001). When 865
beneficial alleles are required for persistence, as in evolution- 866
ary rescue, mutator alleles can reach very high frequencies or 867
rapidly fix (e.g., Mao *et al.* 1997). Consistent with this, mutator 868
alleles are often associated with antibiotic resistance in clinical 869
isolates (see examples in Bell 2017). Further, the more beneficial 870
mutations available the larger the advantage of a mutator allele; 871
for a mutator that increases the mutation rate m -fold, its relative 872
contribution to the production of n beneficial mutations scales 873
as m^n (Tenaillon *et al.* 1999). Thus multi-step rescue can impose 874
much stronger selection for mutator alleles than 1-step rescue. 875
There are a number of examples where lineages with higher 876
mutation rates acquired multiple mutations and persisted at 877
higher doses of antibiotics (Couce *et al.* 2015; San Millan *et al.* 878
2017). The number of mutations required for persistence is, how- 879
ever, often unknown, making it difficult to compare situations 880
where rescue requires different numbers of mutations. Experi- 881
ments with a combination of drugs may provide a glimpse; for 882
instance, *Escherichia coli* populations only evolved resistance to a 883
combination of two drugs (presumably through the well-known 884
mutations specific to each drug) when mutators were present, 885
despite the fact that mutators were not required for resistance 886
to either drug in isolation (Gifford *et al.* 2019). In cases where 887
we have less information on the genetic basis of resistance, our 888
model suggests that mutators will be more advantageous when 889
initial maladaptation is severe (e.g., higher drug concentrations 890
or a larger number of drugs), as rescue will then be dominated 891
by genetic paths with more mutational steps.

892 Here we have investigated the genetic basis of evolution- 893
ary rescue in an asexual population that is initially genetically 894
uniform. Extending this work to allow for recombination and 895
standing genetic variation at the time of environmental change 896
will likely be interesting. The effect of standing genetic variation 897
on the probability of 1-step rescue is relatively straight-forward 898
to incorporate, depending only on the expected number of res- 899
cue mutations initially present and their mean establishment 900
probability (Martin *et al.* 2013). In the case of mutation-selection

840 balance, the probability of 1-step rescue from standing genetic
841 variance in Fisher's geometric model was given by [Anciaux *et al.*](#)
842 (2018), whose equations 3 and 5 immediately give the distribu-
843 tion of fitness effects among those that rescue. Allowing these
844 standing genetic variants to be springboards to multi-step res-
845 cue will help clarify the role of standing genetic variation on the
846 genetic basis of rescue more generally. Recombination can help
847 combine such springboard mutations into rescue genotypes but
848 will also break these combinations apart, as demonstrated in a
849 2-locus 2-allele model of rescue ([Uecker and Hermisson 2016](#)).
850 How recombination affects the genetic basis of evolutionary res-
851 cue when more loci can potentially contribute remains to be
852 seen.

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Appendix

Approximating the probability of 1-step rescue

The probability of 1-step rescue in this model has been derived by [Anciaux *et al.* \(2018\)](#). As replicated in File S1 and given by their equation 7, when $\rho_{max} = m_{max}/\lambda$ is large a closed-form approximation is

$$\Lambda_1(m_0) \approx \tilde{\Lambda}_1(m_0) \equiv -m_0 U \frac{(1 - \psi_0/2)^{(1-n)/2}}{1 - \psi_0/4} g(\alpha), \quad (19)$$

where $\psi_0 = 2(1 - \sqrt{1 - m_0/m_{max}})$, $g(\alpha) = \exp(-\alpha)/\sqrt{\pi\alpha} - \text{erfc}(\sqrt{\alpha})$, and $\alpha = \rho_{max}\psi_0^2/4$, with $\text{erfc}(\cdot)$ the complimentary error function. When the wildtype declines slowly m_0 and thus ψ_0 is small and $\Lambda_1(m_0) \approx U g(\alpha)$. In the limit $m_0 \rightarrow 0$, Equation 19 becomes

$$\tilde{\Lambda}_1(0) \equiv \lim_{m_0 \rightarrow 0} \tilde{\Lambda}_1(m_0) = 2U\sqrt{m_{max}\lambda/\pi}. \quad (20)$$

Mutant lineage dynamics

Here we follow the lead of [Weissman *et al.* \(2010\)](#) and [Uecker and Hermisson \(2016\)](#) in approximating our discrete-time process with a continuous-time branching process (see chapter 6 in [Allen 2010](#)). Consider a birth-death process, where individuals give birth at rate b and die at rate d . One can then obtain the probability generating function for the number of individuals at a given time, $n(t)$, given the initial number, $n(0)$. We are primarily interested in new mutant lineages, $n(0) = 1$. The generating

1076 function then allows us to calculate the probability that a lineage
 1077 persists at least until time t and the distribution of $n(t)$ given it
 1078 does so (see below).

1079 To convert between birth and death rates and our compound
 1080 Malthusian parameter we follow [Uecker and Hermisson \(2016\)](#)
 1081 in equally distributing the growth rate m between birth and
 1082 death, $b = (1 + m)/2$ and $d = (1 - m)/2$, such that $m = b - d$
 1083 and the continuous-time process exhibits the same amount of
 1084 drift as the discrete time process (and matches discrete-time
 1085 simulations well; [Uecker et al. 2014](#)). We can now report the
 1086 necessary results in terms of m (assuming $|m| < 1$).

1087 Denoting the extinction time as T , the probability a mutant
 1088 with growth rate m persists until time t is approximately (see
 1089 File S1 for derivation)

$$P(T > t) \approx \begin{cases} 2/t & t \ll |1/m| \\ -2m \exp(mt) & t \gg -1/m > 0 \end{cases} \quad (21)$$

1090 As pointed out in [Weissman et al. \(2010\)](#) (whose equation A2
 1091 differs from Equation 21 by a factor of 2 because they have
 1092 $b + d = 2$), the distribution of persistence times has a long
 1093 tail (like $1/t$) until being cut off (declining exponentially) at
 1094 $t = -1/m$.

1095 Given a lineage persists until t , the distribution of $n(t)$ is
 1096 roughly (see File S1 for derivation)

$$P(n(t) = n | n(t) > 0) \approx \begin{cases} 2(1/t)(1 + 2/t)^{-n} & t \ll |1/m| \\ -2m(1 + m)^{n-1} & t \gg -1/m > 0 \end{cases} \quad (22)$$

1097 As pointed out in [Weissman et al. \(2010\)](#) (whose equation A3
 1098 only differs from Equation 22 by constants), the distribution of
 1099 $n(t)$ is approximately geometric for small or large t , implying
 1100 $n(t)$ is very unlikely to be greater than the minimum of t and
 1101 $-1/m$.

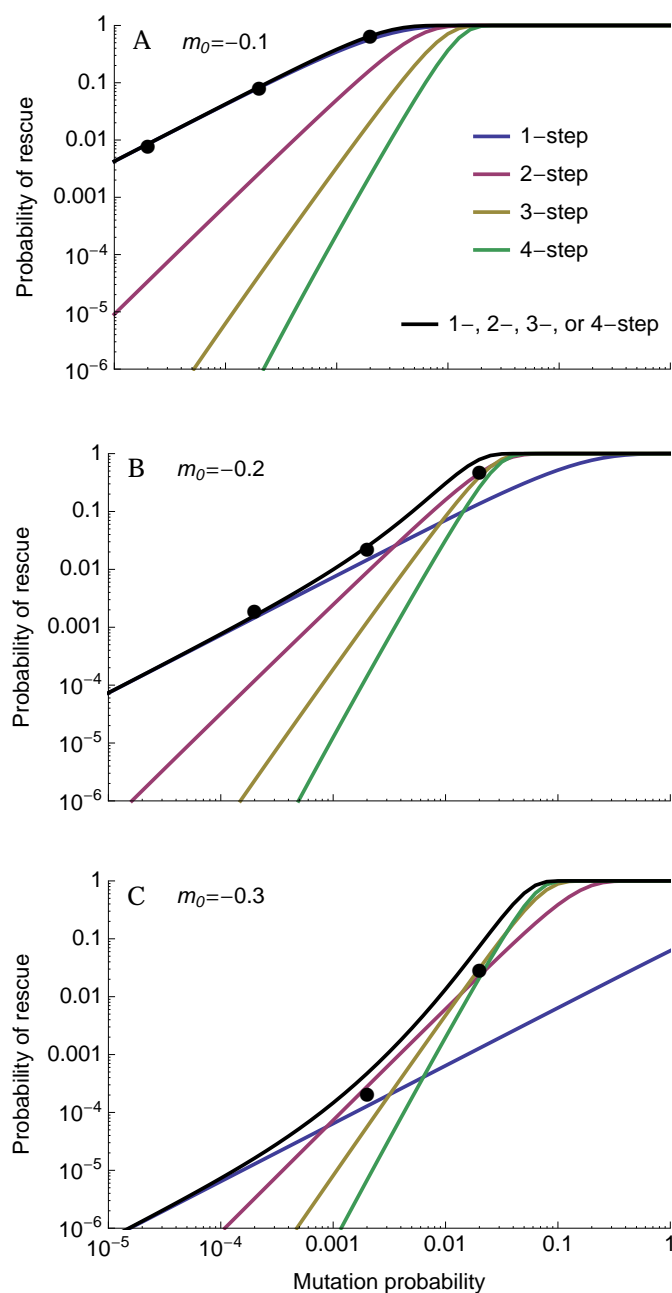


Figure S1 The probability of rescue as a function of mutation rate for three different levels of initial maladaptation. See Figure 3 for details. Other parameters: $n = 4$, $\lambda = 0.005$, $m_{max} = 0.5$, $N_0 = 10^4$.

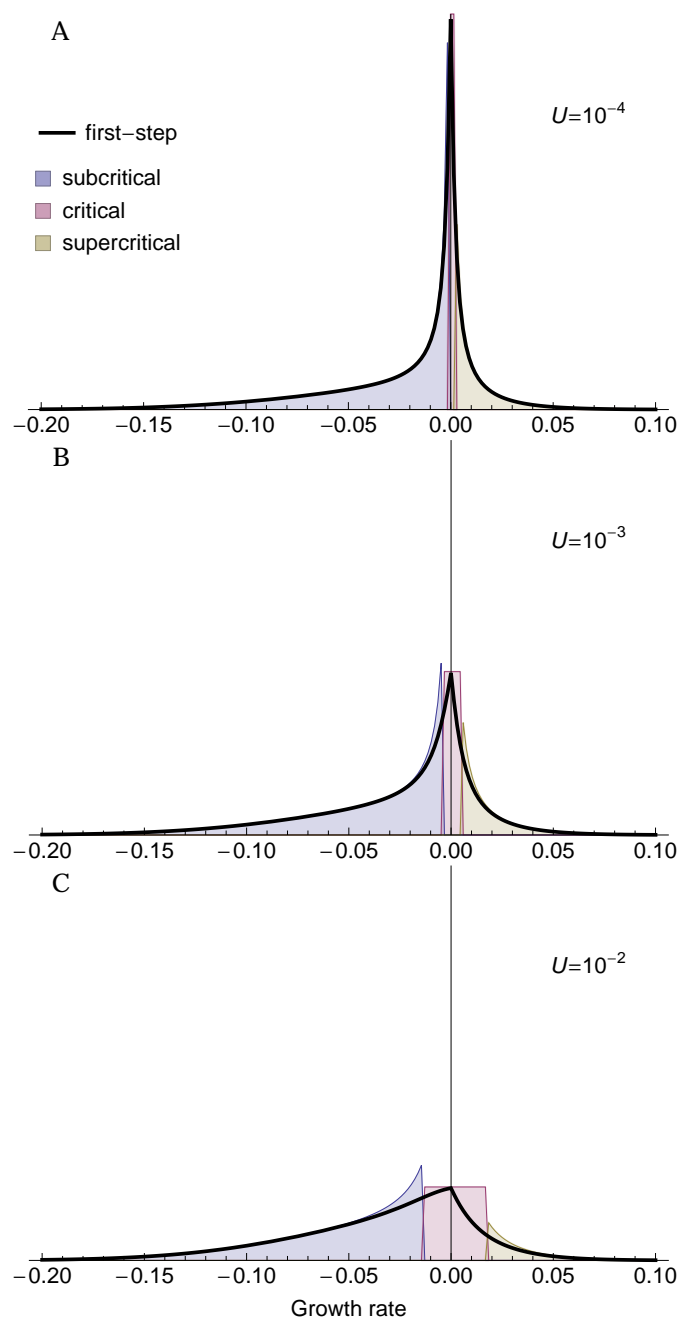


Figure S2 The distribution of first-step mutant growth rates given 2-step rescue under three mutation rates. See Figure 7 for details. Parameters: $n = 4$, $\lambda = 0.005$, $m_{max} = 0.5$, $m_0 = -0.2$.