Running Title: Dissociation of lipid transfer activities of MTTP

A missense mutation dissociates triglyceride and phospholipid transfer activities in zebrafish and human microsomal triglyceride transfer protein

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SUMMARY

Microsomal triglyceride transfer protein (MTP) transfers triglycerides and phospholipids and is essential for the assembly of Apolipoprotein B (ApoB)-containing lipoproteins in the endoplasmic reticulum. We have discovered a zebrafish mutant (*mttp*^{c655}) expressing a C-terminal missense mutation (G863V) in Mttp, one of the two subunits of MTP, that is defective at transferring triglycerides, but retains phospholipid transfer activity. Mutagenesis of the conserved glycine in the human MTTP protein (G865V) also eliminates triglyceride but not phospholipid transfer activity. The G863V mutation reduces the production and size of ApoB-containing lipoproteins in zebrafish embryos and results in the accumulation of cytoplasmic lipid droplets in the yolk syncytial layer. However, *mttp*^{c655} mutants exhibit only mild intestinal lipid malabsorption and normal growth as adults. In contrast, zebrafish mutants bearing the previously identified *mttp*^{stl} mutation (L475P) are deficient in transferring both triglycerides and phospholipids and exhibit gross intestinal lipid accumulation and defective growth. Thus, the G863V point mutation provides the first evidence that the triglyceride and phospholipid transfer functions of a vertebrate MTP protein can be separated, arguing that selective inhibition of the triglyceride transfer activity of MTP may be a feasible therapeutic approach for dyslipidemia.

Keywords: MTP, MTTP, ApoB, lipid transfer, zebrafish, yolk

INTRODUCTION

In vertebrates, Apolipoprotein B-containing lipoproteins (B-lps) are produced by the intestine and liver and transport lipid and fat-soluble vitamins to the peripheral tissues through the circulation. Lipoproteins are composed of a neutral core of triglyceride (TG) and cholesteryl esters surrounded by a monolayer of phospholipid, free cholesterol and sphingomyelin. B-lps contain one apolipoprotein B (ApoB) scaffold protein embedded in the phospholipid monolayer as well as other exchangeable lipoproteins (Hussain et al., 1996; Schumaker et al., 1994). B-lp assembly occurs in the endoplasmic reticulum (ER) and requires the activity of microsomal triglyceride transfer protein (MTP) (Hussain et al., 2003b; Patel and Grundy, 1996; Wetterau and Zilversmit, 1986). As ApoB is translated and translocated into the lumen of the ER, MTP physically interacts with and transfers lipids to ApoB to form primordial lipoproteins (Bradbury et al., 1999; Hussain et al., 2003b; Patel and Grundy, 1996; Wu et al., 1996). MTP may also aid in the formation of TG-rich lumenal lipid droplets that are believed to fuse to the primordial lipoproteins, thus increasing the size of nascent lipoproteins (Alexander et al., 1976; Boren et al., 1994; Hamilton et al., 1998; Kulinski et al., 2002; Raabe et al., 1999; Wang et al., 1997).

MTP is a heterodimer of the large 97-kDa M subunit (microsomal triglyceride transfer protein, MTTP) and the small 58-kDa P subunit protein disulfide isomerase (PDI)(Wetterau et al., 1990). Mutations in the *MTTP* gene that prevent lipid transfer and ApoB secretion cause the disease Abetalipoproteinemia (OMIM 200100), characterized by a virtual absence of plasma B-lps (Kane, 1995; Sharp et al., 1993; Shoulders et al., 1993; Wetterau et al., 1992). Patients exhibit fat malabsorption, low plasma triglyceride, and cholesterol levels as well as fat-soluble vitamin deficiencies (Kane, 1995; Lee and Hegele, 2014; Walsh and Hussain, 2016). Without adequate supplementation of essential fatty acids and fat-soluble vitamins, these patients can develop a variety of complications including neurological, opthalmological, and hematological disorders (Kane, 1995; Lee and Hegele, 2014).

Vertebrate MTP can transfer triacylglycerol, diacylglycerol, phospholipid, cholesteryl ester, ceramide, and sphingomyelin between vesicles *in vitro* (Athar et al., 2004; Iqbal et al., 2015; Jamil et al., 1995; Rava et al., 2005; Wetterau and Zilversmit, 1984, 1985). Kinetic studies suggest that MTP transiently interacts with membranes, acquires lipids, and then delivers these

lipids to an acceptor membrane. The transfer of lipids occurs down a concentration gradient and does not require energy (Atzel and Wetterau, 1993, 1994). While vertebrate MTP predominantly transfers TG (Rava et al., 2005; Wetterau and Zilversmit, 1985), the Drosophila orthologue of MTP lacks TG transfer activity (Rava et al., 2006), has phospholipid transfer activity and supports secretion of vertebrate ApoB (Khatun et al., 2012; Rava and Hussain, 2007; Rava et al., 2006; Sellers et al., 2003). A further analysis of MTTP orthologues in divergent species, including nematodes, insects, fish, and mammals, indicates that all orthologues bind PDI, localize to the ER, and support human ApoB secretion (Rava and Hussain, 2007). However, only vertebrate MTP orthologues exhibit TG transfer activity, suggesting that phospholipid transfer activity was the original function of MTP orthologues and that neutral lipid transfer first evolved in fish (Rava and Hussain, 2007).

Here we describe a hypomorphic missense mutation in the C-terminal domain of zebrafish *mttp* (G863V) that decreases the production and size of B-lps *in vivo*, but has minimal effects on lipid malabsorption in the intestine and no effect on growth. Biochemical characterization of the G863V allele indicates that it is defective in triglyceride transfer activity, but retains phospholipid transfer activity. Further, we show that mutation of the conserved glycine at position 865 in human *MTTP* also selectively abolishes triglyceride transfer activity. Taken together, these data provide the first evidence that the lipid transfer functions of a vertebrate MTP can be biochemically dissociated and argues that phospholipid transfer activity is sufficient for absorption of dietary lipid.

RESULTS

The c655 allele is a missense mutation in mttp. During routine screening, we identified zebrafish embryos with opaque yolks (Figure 1A). In contrast to the translucent yolks of wild-type fish, when illuminated by transmitted light, the mutant yolks appear dark (Figure 1A) and, with incident light, appear off-white (Figure 1— figure supplement 1A). The phenotype was found to be present in Mendelian ratios (Figure 1A), suggesting the presence of a homozygous recessive mutation. This new allele is identified as Carnegie allele *c655*. To map the location of the causative mutation, we did RNA sequencing of 23 opaque-yolk *c655* mutants and 23 translucent-yolk siblings and performed a Euclidean distance mapping analysis using the Mutation Mapping Analysis Pipeline for Pooled RNA-seq (MMAPPR)(Hill et al., 2013). The

Loess fit to the mapping scores (Euclidean Distance[4]) (Figure 1B, top) indicated the c655 mutation was located on Chromosome 1 and lay in the region between 9-20 MB (Figure 1B, bottom). Single nucleotide variants (SNVs) present in this 11MB region in *c655* mutant embryos were assessed for their effect on annotated genes using the Ensembl Variant Effect Predictor (McLaren et al., 2016), including using the Sorting Intolerant from Tolerant algorithm (SIFT) (Sim et al., 2012), to predict the impact of changes on protein-coding sequence (tolerated or deleterious). We extracted variants that alter the protein-coding sequence as candidates for the causal mutation (223 variants in 64 genes, of which 42 are missense variants predicted to be deleterious; Supplementary File 1).

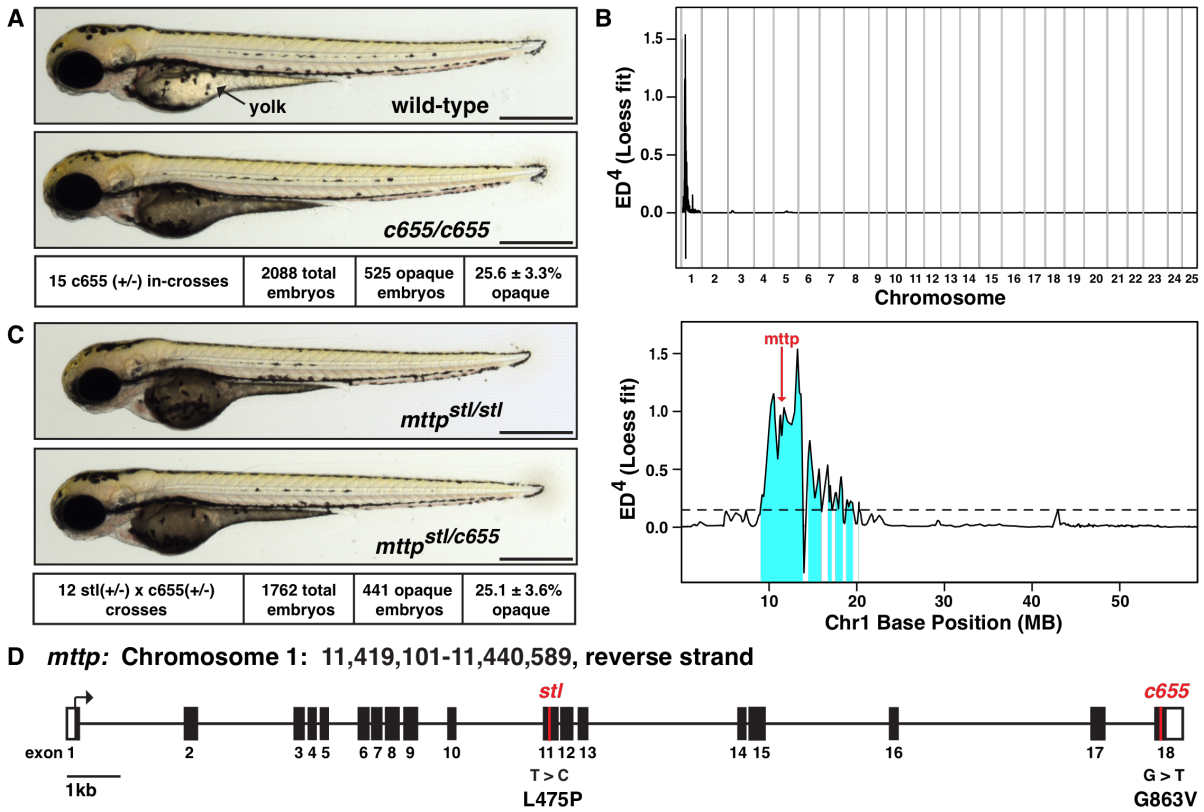

Figure 1: The *c655* allele is a missense mutation in the M-subunit of microsomal triglyceride transfer protein. (A) Representative images of wild-type (top) and homozygous mutant (middle) c655 zebrafish embryos at 3 days post fertilization (dpf); Scale = 500 μm. The dark/opaque yolk phenotype segregated with a Mendelian ratio consistent with a homozygous recessive mutation (bottom), mean +/- SD. (B) Euclidean distance mapping analysis plots

128 produced by MMAPPR (Hill et al., 2013), showing the likely genomic region of the *c655*
129 mutation: plot of the LOESS fit to the mapping scores (Euclidean Distance[4]) across all 25
130 chromosomes (top) and expanded view of chromosome 1(GRCz10: CM002885.1)(bottom).
131 Red arrow shows the position of the G>T missense mutation in *mttp* (ENSDARG00000008637,
132 position 11,421,261(GRCz10). (C) Representative images of a homozygous mutant zebrafish
133 embryo carrying the previously described *stalactite* (*stl*) missense mutation in *mttp* (Avraham-
134 Davidi et al., 2012)(top) and a trans-heterozygous $mttp^{stl/c655}$ embryo (middle); 3 dpf, scale = 500
135 μm. The dark/opaque yolk phenotype is present at expected ratios (bottom) and genotyping
136 confirms only embryos with opaque yolks are trans-heterozygous for the *mttp* alleles. (D)
137 Depiction of the *mttp* gene structure highlighting the locations of the *stl* (L475P) (position
138 11,431,645, GRCz10, transcript mtp-204 (ENSDART00000165753.2) nucleotide 2588) and
139 *c655* (G863V) (nucleotide 1424) missense alleles in exon 11 and 18, respectively.
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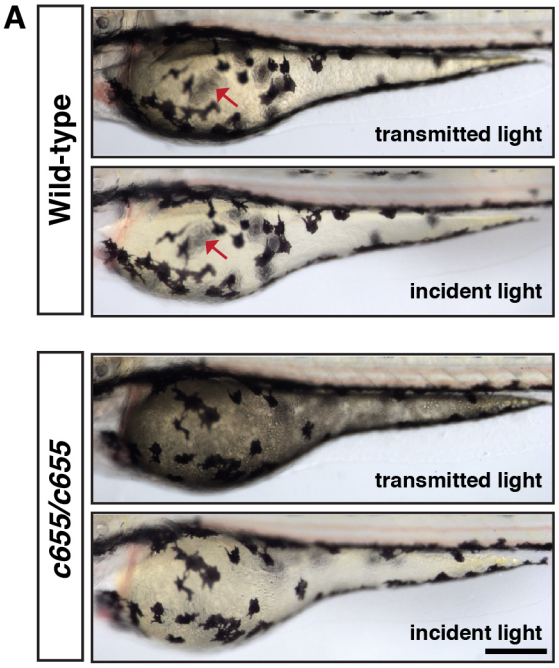

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144 **Figure 1 – figure supplement 1: Lipid droplets block light transmission through the yolk**
145 **of the embryo.** (A) Wild-type and *c655* mutant embryos were imaged at 3 dpf using either
146 transmitted light (illumination below the fish) or incident light (illumination from above the fish).
147 The wild-type embryos are translucent; the pigment cells on the opposite side of the embryo
148 (red arrow) are visible through the yolk with both light sources. The yolk is opaque in the

149 mutants; it appears dark with transmitted light and white with incident light. Pigment cells on the
150 opposite side of the embryo are barely visible in mutant embryos, regardless of light source.
151 Scale = 200 μM.
152
153
154 One of the SNVs linked to the *c655* phenotype was a missense mutation predicted to be
155 deleterious in exon 18 of the microsomal triglyceride transfer protein gene
156 (ENSDARG00000008637, Chr1:11,421,261 GRCz10). A previously identified missense
157 mutation in exon 11 of zebrafish *mttp*, stalactite (*stl*), also presents with an opaque yolk
158 phenotype (Figure 1C, top) (Avraham-Davidi et al., 2012), suggesting that the *c655* opaque yolk
159 phenotype might result from this newly identified missense mutation in *mttp*. To test this
160 hypothesis, we performed complementation crosses between $mttp^{c655/+}$ heterozygous fish and
161 $mttp^{stl/+}$ heterozygous fish. The *c655* mutation failed to complement the $mttp^{stl}$ mutation, as one-
162 quarter of the embryos in these crosses displayed the opaque yolk phenotype (Figure 1C,
163 bottom). All of the embryos exhibiting opaque yolks were heterozygous for both the $mttp^{stl}$ and
164 the *c655* mutation in *mttp*. This strongly argues that the G/T SNV in exon 18 of *mttp* is the
165 causative allele for the *c655* opaque yolk phenotype. This was further confirmed by rescuing
166 the *c655* opaque yolk phenotype with injections of a wild-type mttp-FLAG plasmid at the 1-cell
167 stage (Figure 1— figure supplement 2).
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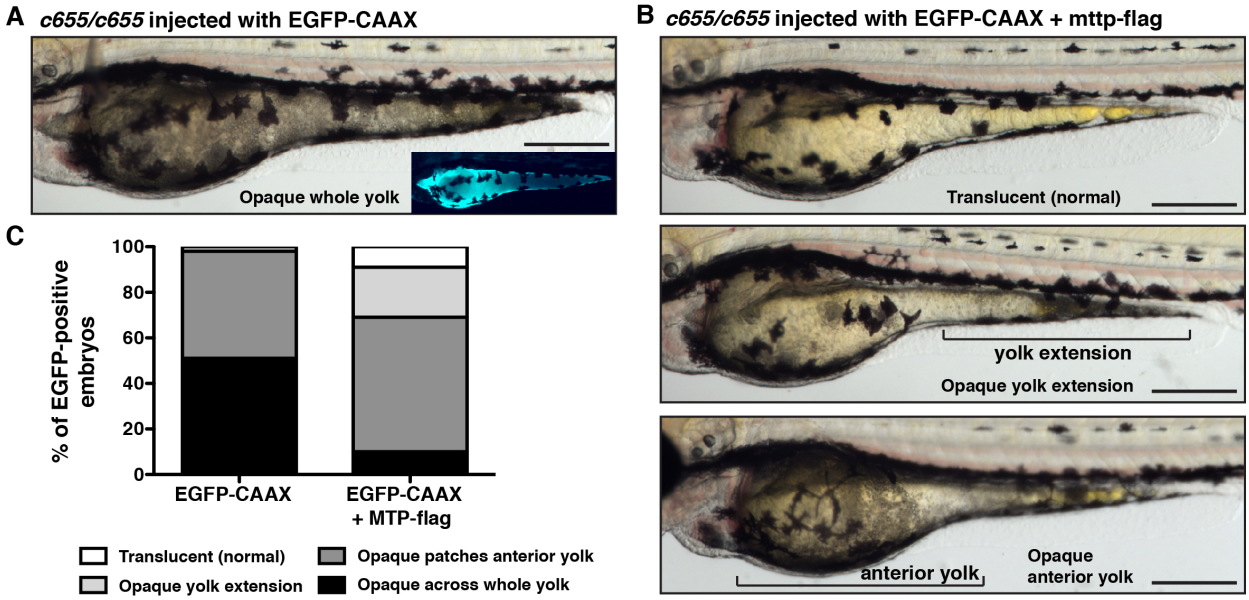

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172 **Figure 1 – figure supplement 2: Expression of wild-type zebrafish *mttp*-FLAG rescues**
173 **the opaque yolk phenotype in *mttp*$^{c655/c655}$ embryos.** One-cell stage *mttp*$^{c655/c655}$ embryos
174 were co-injected with CMV: *mttp*-FLAG and the CMV: *eGFP-CAAX* plasmid, or CMV: *eGFP-*
175 *CAAX* alone as a control. Embryos expressing eGFP in the YSL were imaged at 3 dpf, and
176 images were blinded and scored for the degree of yolk opacity. (A) Representative image of an
177 *mttp*$^{c655/c655}$ mutant embryo expressing eGFP-CAAX in the YSL and a fully opaque yolk. (B)
178 Examples of injected embryos with varying degrees of yolk opacity (normal translucent yolk,
179 opaque region in the yolk extension, opaque patches in the anterior yolk with or without opaque
180 yolk extension). (C) Images were binned into the four noted categories of yolk opacity. Results
181 represent pooled data from 3 independent experiments, n = 91 control and 102 Mttp-FLAG
182 eGFP-positive embryos total. Chi-square test, $p<0.001$. Scale = 500 μM.
183
184
185 Both the *mttp*^{stl} allele and *mttp*^{c655} allele are missense mutations. The *stl* allele results in the
186 conversion of a leucine to a proline at residue 475 and the *c655* mutation is a glycine to valine
187 mutation in the C-terminus of the protein at residue 863 (total length = 884 residues) (Figure
188 1D). An additional SNV in *mttp* at position Chr1:11,421,300 GRCz10 (T/C) causing a missense
189 mutation (M850T) was identified in *c655* mutants; however, this SNP was not predicted to be
190 deleterious and has been previously noted in the Ensembl zebrafish genome database.
191 Furthermore, no change in mRNA expression was noted for *mttp* in the *mttp*^{c655} mutants in our
192 RNAseq data-set (log2[fold change] = 0.18, adj. p-value = 0.19).
193
194 Although the *mttp*$^{stl/stl}$, *mttp*$^{c655/c655}$, and trans-heterozygous *mttp*$^{stl/c655}$ fish all exhibit opaque
195 yolks, the *mttp*$^{stl/stl}$ mutants have a more severe phenotype, in that their yolks are darker and
196 they retain the opaque phenotype longer during development. The *mttp*$^{c655/c655}$ mutant
197 phenotype is the least severe and the trans-heterozygotes exhibit an intermediate phenotype
198 (Figure 1— figure supplement 3). Encouraged by the differences in embryonic phenotype, we
199 hypothesized that these mutations would provide an opportunity to further dissect the molecular
200 details of MTP function *in vivo*.
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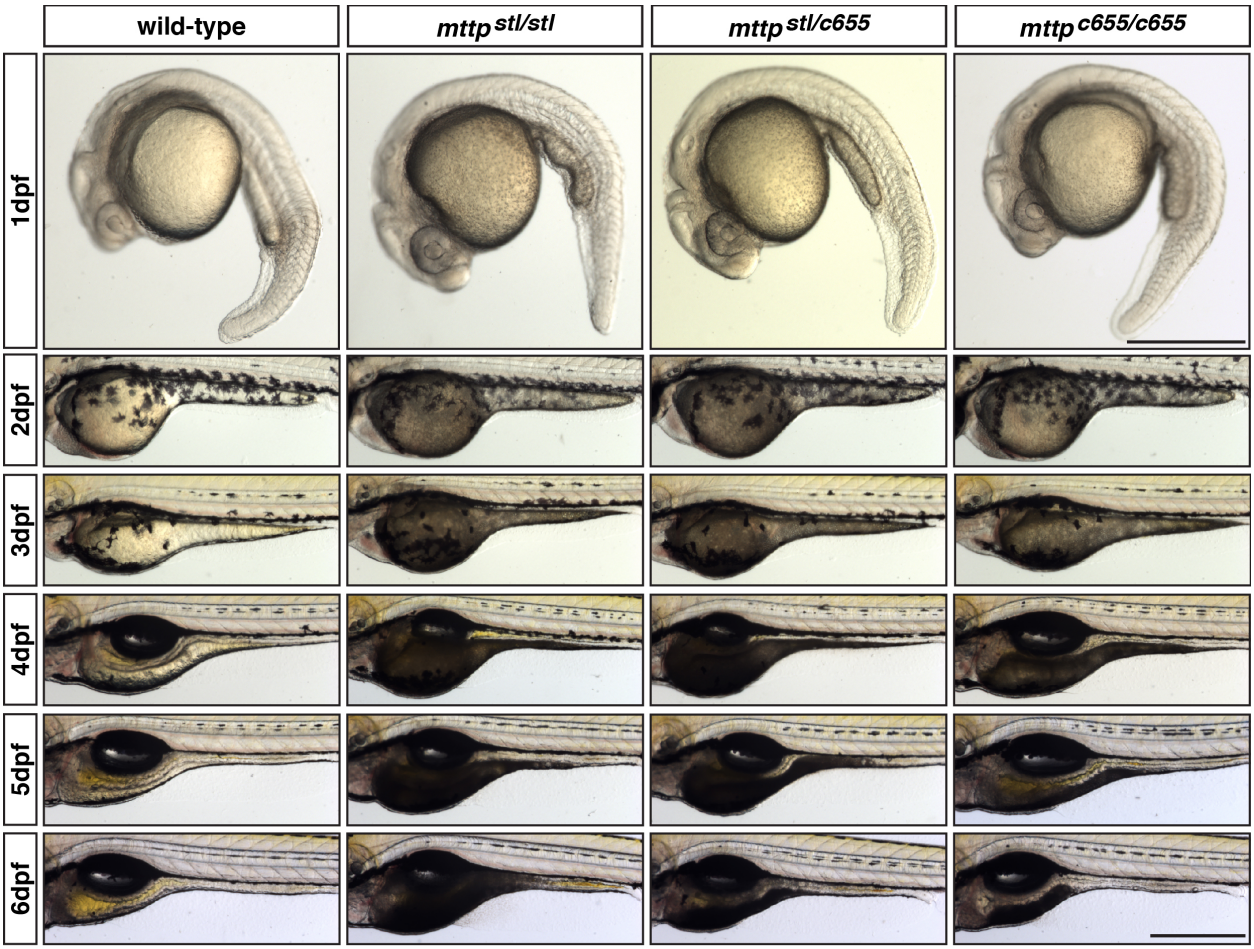

Figure 1 — figure supplement 3: The *stl* and *c655* mttp mutations have differential effects on the degree of yolk opacity during embryonic development. Representative images of wild-type, *mttp*$^{stl/stl}$, *mttp*$^{c655/c655}$ and trans-heterozygous *mttp* $^{stl/c655}$ mutants from 1 dpf to 6 dpf. The *mttp*$^{stl/stl}$ mutants are visibly opaque at 1 dpf and the area of opacity is retained for longer than in *mttp*$^{stl/c655}$ or *mttp*$^{c655/c655}$ mutants. 3 dpf images are the same fish shown in Figure 1. Scale = 500 μM.

Mttp mutants accumulate cytoplasmic lipid droplets in the yolk syncytial layer. As a lecithotrophic organism, zebrafish rely on their maternally-derived yolk as the source of nutrients and building blocks for embryogenesis (Hiramatsu et al., 2015; Mani-Ponset et al., 1996; Vernier and Sire, 1977). The yolk is rich in lipids (Fraher et al., 2016; Miyares et al., 2014; Wiegand, 1996) and following lipolysis and re-esterification, the lipids are packaged into lipoproteins in the ER of the yolk syncytial layer (YSL), a multi-nucleated cytoplasm that

219 surrounds the yolk mass (Figure 2A)(Carvalho and Heisenberg, 2010; Kimmel and Law, 1985;
220 Walzer and Schonenberger, 1979a, b). The zebrafish YSL expresses apolipoprotein B (Otis et
221 al., 2015) and microsomal triglyceride transfer protein (Marza et al., 2005; Schlegel and Stainier,
222 2006). The YSL produces B-lps (Thierer et al., In Press; Vernier and Sire, 1977; Walzer and
223 Schonenberger, 1979a), similar to the intestine (Glickman et al., 1976; Kessler et al., 1970),
224 liver (Mahley et al., 1970), placenta (Madsen et al., 2004), and embryonic yolk sac in mammals
225 (Farese et al., 1996; Plonne et al., 1992). In the mammalian intestine, ApoB mRNA is edited,
226 resulting in a truncated ApoB molecule (ApoB48)(Davidson and Shelness, 2000; Kane et al.,
227 1980); however, there is no evidence for editing in zebrafish, so all B-lp-producing tissues,
228 including the YSL, secrete lipoproteins containing ApoB100 (Thierer et al., In Press).
229
230 When MTP is mutated or absent, B-lp production is reduced or absent and TG accumulates in
231 cytoplasmic lipid droplets (LDs) instead (Khatun et al., 2012; Raabe et al., 1999). We have
232 previously shown that accumulation of LDs in intestinal enterocytes of zebrafish larvae fed a
233 high-fat meal causes the gut to be opaque (Otis and Farber, 2016)(Figure 2 – figure supplement
234 1), most likely due to the lipid droplets' ability to scatter light (Hwang et al., 2018; Michels et al.,
235 2008). Therefore, we hypothesized that the yolk opacity in the *mttp* mutant embryos is due to
236 aberrant accumulation of LDs in the cytoplasm of the YSL. Using transmission electron
237 microscopy, we found that the YSL in the wild-type embryos contains very few, if any, canonical
238 YSL LDs, whereas the $mttp^{stl/stl}$, $mttp^{c655/c655}$ and trans-heterozygous $mttp^{stl/c655}$ embryos
239 accumulate substantial numbers of cytoplasmic LDs (Figure 2B). LDs in $mttp^{stl/stl}$ mutants are
240 more numerous and more uniform in size, whereas the $mttp^{c655/c655}$ mutants often had very large
241 LDs in addition to small droplets (Figure 2C). As a result, the number of LDs per area of the
242 YSL is reduced in the $mttp^{c655/c655}$ mutants compared to $mttp^{stl/stl}$ mutants (Figure 2D). The trans-
243 heterozygous fish had LDs that were more similar in size to the $mttp^{stl/stl}$ mutants and had a trend
244 toward fewer lipid droplets per YSL area, although this was not significant.
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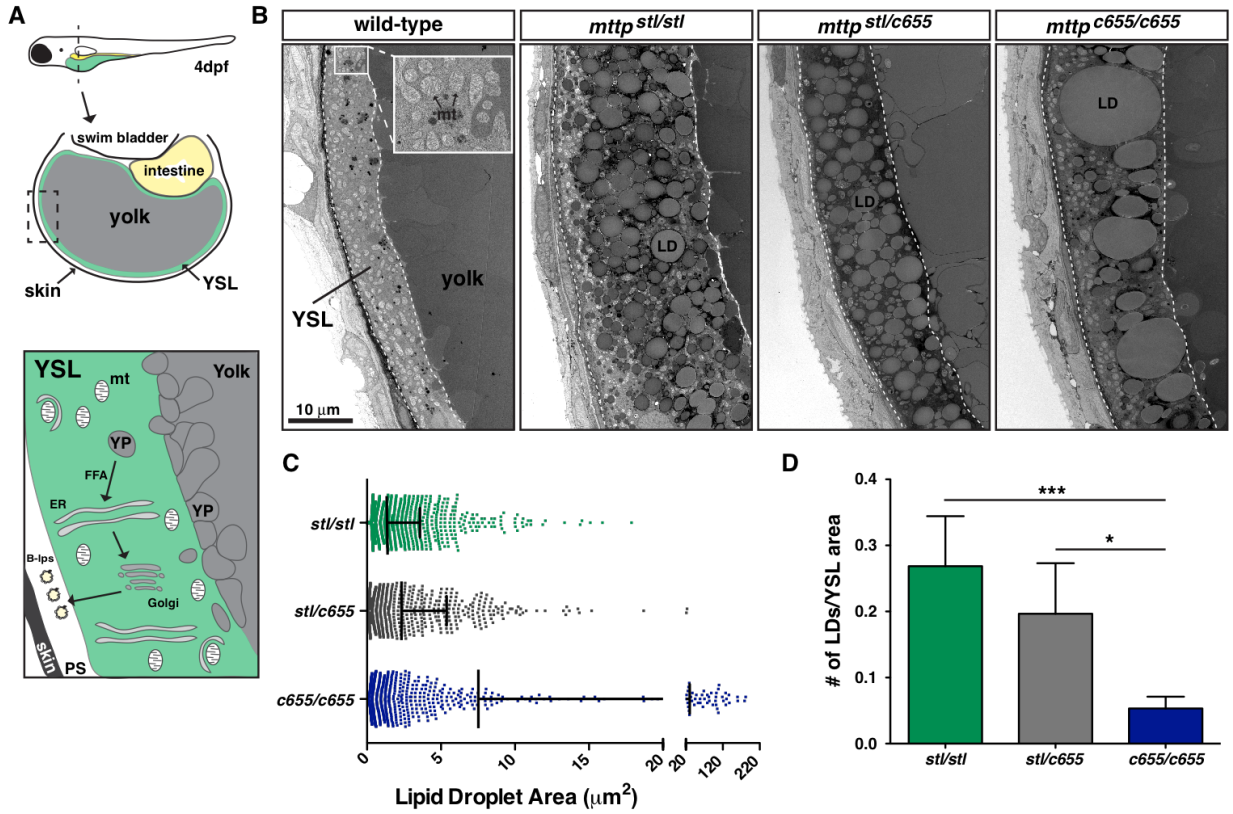

Figure 2: The opaque yolk phenotype results from the accumulation of aberrant cytoplasmic lipid droplets in the yolk syncytial layer. (A) (Top) Cartoon depicting the cross-sectional view of a 4-dpf zebrafish embryo. The yolk syncytial layer (YSL) surrounds the yolk mass and serves as the embryonic digestive organ. The dashed box indicates the view expanded in the bottom panel and in panel B. (Bottom) Stored yolk lipids undergo lipolysis in yolk platelets (YP), presumably releasing free fatty acids into the YSL. These fatty acids are re-esterified in the ER bilayer to form triglycerides, phospholipids, and cholesterol esters. The lipids are packaged into ApoB-containing lipoproteins in the ER with the help of MTP and are further processed in the Golgi before being secreted into the perivitelline space (PS) and then circulation. (B) Representative transmission electron micrographs of the yolk and YSL from wild-type and *mttp* mutants; dashed lines delineate the YSL region, mt = mitochondria, scale = 10 μm. (C) Quantification of lipid droplet size in *mttp* mutants, n ≥ 700 lipid droplets in 2 fish per genotype; mean +/- SD. (D) Quantification of the number of lipid droplets per YSL area, n = 7-9 YSL regions per genotype (3–5 regions per fish, 2 fish per genotype); mean +/- SD, Kruskall-Wallis with Dunn's Multiple Comparison test, vs. $mttp^{c655/c655}$ * $p < 0.05$, *** $p < 0.001$.

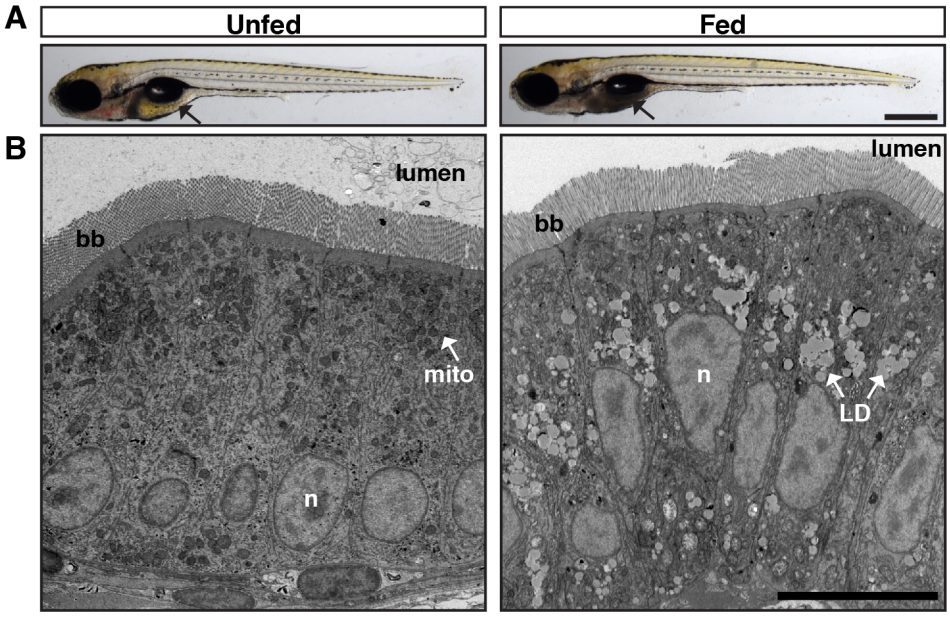

Figure 2 – figure supplement 1: Lipid droplets block light transmission through the larval intestine. (A) Wild-type fish at 6 dpf were fed a high-fat meal for 1h as described previously (Otis and Farber, 2016). Unfed fish have translucent intestines (black arrow, left) when imaged with transmitted light, whereas fed fish have opaque intestines (black arrow, right). Scale = 500 μM. (B) Electron microscopy following a 1 h high-fat feed reveals an accumulation of cytoplasmic lipid droplets in the intestinal enterocytes. By scattering light and blocking light transmission through the intestine, the accumulation of cytoplasmic lipid droplets causes the intestine to appear opaque. Nucleus (n), mitochondria (mito), brush border (bb), lipid droplet (LD). Scale = 10 μM.

The stl and c655 mutations have differential effects on adult size and steatosis. Patients with Abetalipoproteinemia often present in infancy with growth retardation, diarrhea, fat malabsorption, and failure to thrive (reviewed in (Lee and Hegele, 2014)), and whole body deficiency of MTP in a murine model is embryonic lethal (Raabe et al., 1998). In agreement with the mouse phenotype, in the original characterization of the zebrafish *mttp*$^{stl/stl}$ phenotype it was noted that the fish did not survive past 6 days post fertilization (dpf)(Avraham-Davidi et al., 2012). Therefore, we were surprised to find that some of the *stl* mutants were able to survive past larval stages. While these fish are generally much smaller in length and mass (Figure 3A,B) and their viability is reduced relative to their siblings (expected 25%, observed 3.8%

285 [5/131 fish] at 7.5 months), some of these fish have survived to be at least 18 months old.
286 Survival rates are better when the mutants are reared separately and are not competing with
287 siblings for food. Although these fish can reproduce, this is rare. In contrast, the *mttp*$^{c655/c655}$
288 mutants do not exhibit reduced viability and we did not find any reduction in size or fertility of the
289 *c655* mutants compared with siblings (Figure 3A,B). No difference in length or mass was noted
290 in fish trans-heterozygous for *mttp*$^{stl/c655}$ (Figure 3A,B).
291

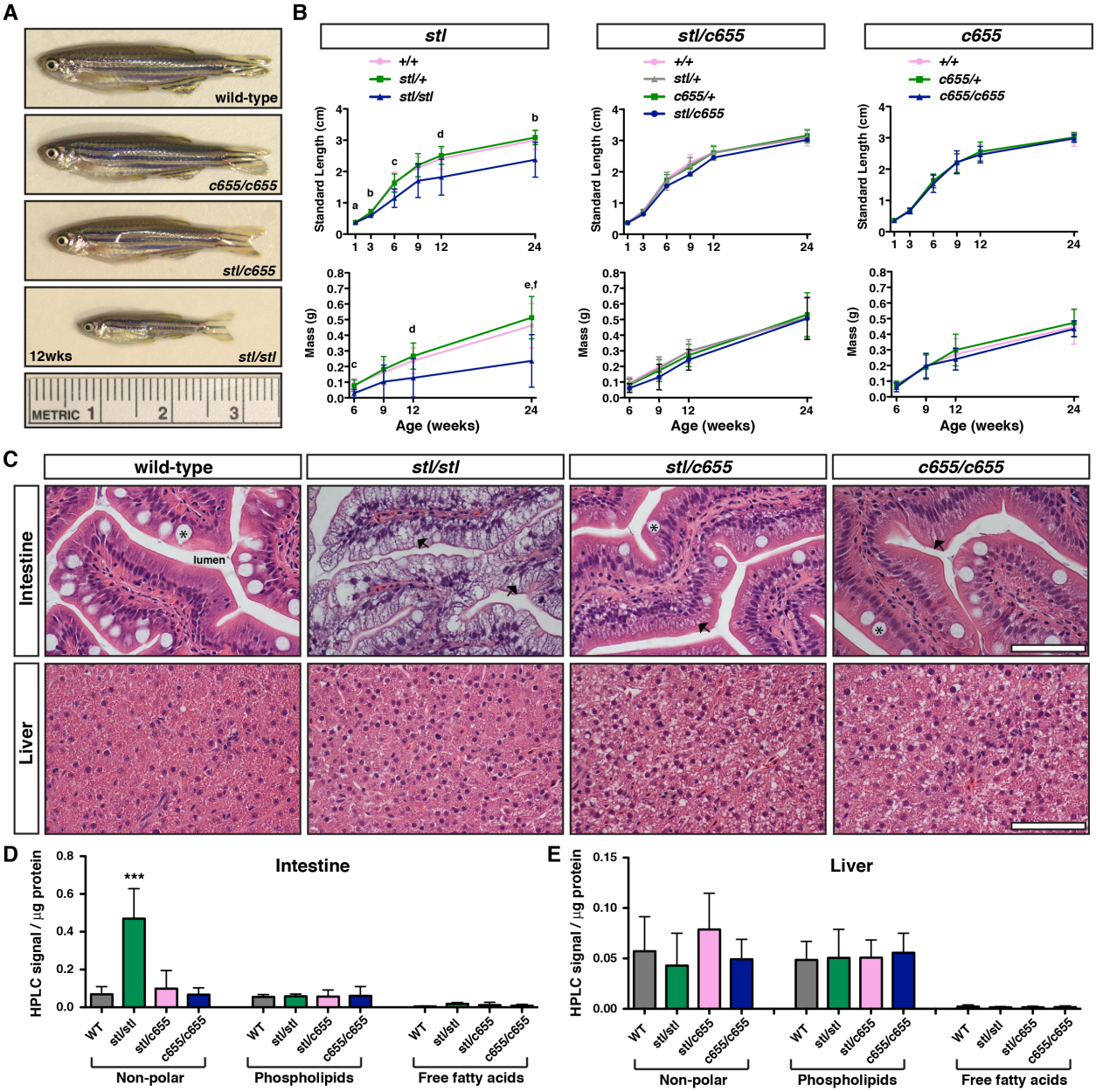

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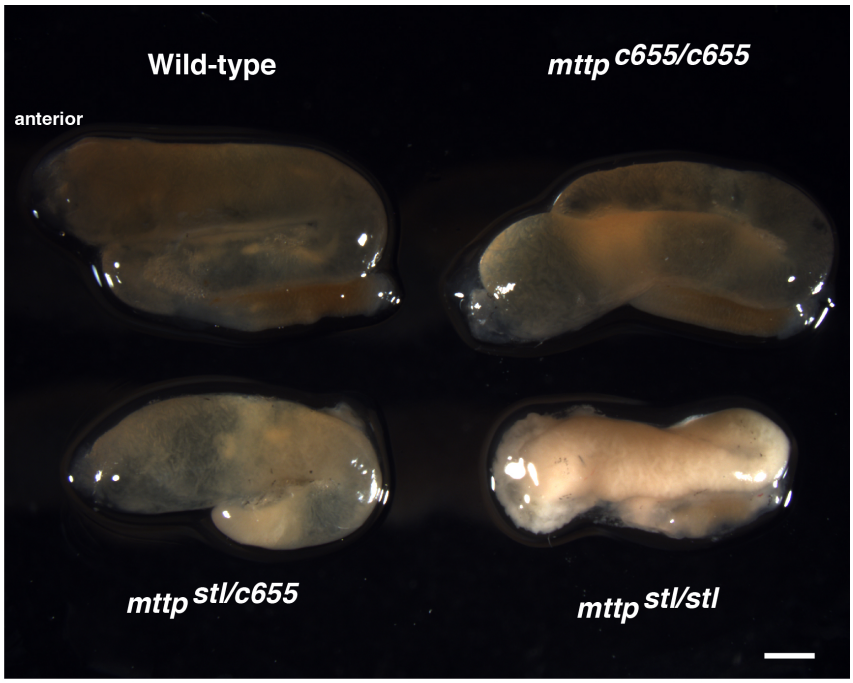
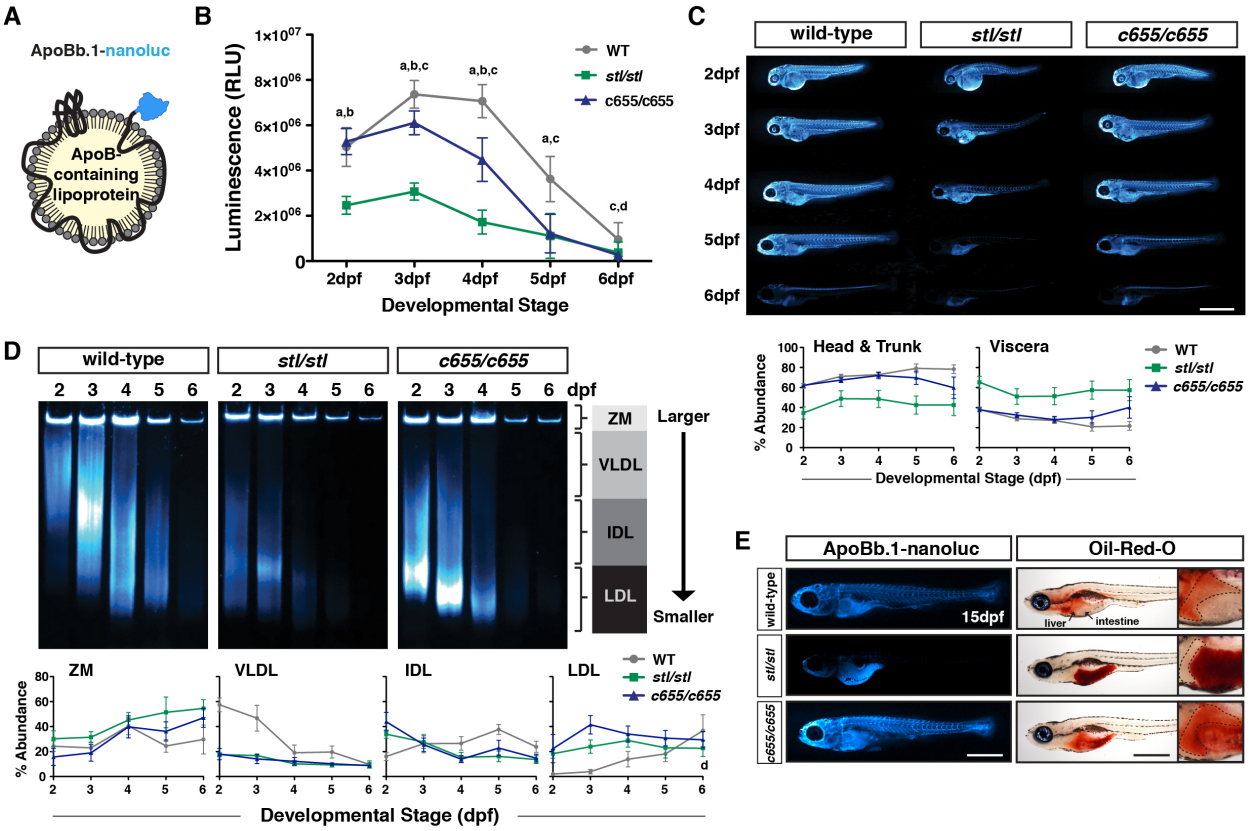
Figure 3: The *stl* and *c655 mttp* mutations have differential effects on the growth and accumulation of lipid in intestine. (A) Representative images of WT and *mttp* mutant fish at 12 weeks of age. (B) Developmental time-course of standard length and mass measurements of *mttp* mutant fish and siblings. Results are representative of pooled data from two independent experiments, n = 7–80 fish/genotype/time-point, mean +/- SD. Significance was determined with a Robust ANOVA and Games-Howell post-hoc tests were used to make pair-wise comparisons at each time point. Using a Bonferroni correction, p-values were adjusted to control for multiple comparisons (6 length or 4 mass comparisons), a: *stl/+* vs. *stl/stl*, $p < 0.01$; b: *+/+* vs. *stl/stl* and *stl/+* vs. *stl/stl*, $p < 0.05$; c: *+/+* vs. *stl/stl* and *stl/+* vs. *stl/stl*, $p < 0.001$; d: *stl/+* vs. *stl/stl*, $p < 0.05$; e: *+/+* vs. *stl/stl*, $p < 0.01$; f: *stl/+* vs. *stl/stl*, $p < 0.001$. (C) Representative images of H&E stained intestine and liver from adult WT and *mttp* mutant fish (7.5 months), scale = 50 μm, * indicate goblet cell, arrows indicate representative lipid accumulation in enterocytes. (D & E) Quantification of non-polar lipids, phospholipids, and free fatty acids in intestine and liver tissue using high-performance liquid chromatography; n = 5–6 fish per genotype, mean +/- SD, One-way ANOVA with Bonferroni post-hoc tests were performed separately for each lipid class; *** $p < 0.001$ *stl/stl* vs. all other genotypes.



314 **Figure 3 – figure supplement 1: Significant lipid accumulation in the intestine of *stl/stl***
315 **but not in *c655/c655* mutants.** Representative images of isolated intestines from adult WT
316 and *mttp* mutant fish (7.5 months), scale = 1 mm.
317
318
319 To assess why the *mttp*$^{stl/stl}$ mutants, but not the *mttp*$^{c655/c655}$ mutants, have defects in growth, we
320 examined whether the mutations have differential effects on fat malabsorption. H&E staining of
321 intestinal tissue from fasted adults revealed gross accumulation of lipid in the cytoplasm of
322 enterocytes in the *mttp*$^{stl/stl}$ fish (Figure 3C, Figure 3 – figure supplement 1). The *mttp*$^{c655/c655}$
323 mutants were largely protected from this abnormal lipid retention. While the trans-heterozygous
324 fish exhibited more visible lipid retention than either wild-type or *mttp*$^{c655/c655}$ fish, this was not as
325 profound as in the *mttp*$^{stl/stl}$ fish. Retention of non-polar lipid (TG and cholesterol ester),
326 phospholipid, and free fatty acids in the intestine were also quantified using HPLC and the
327 *mttp*$^{stl/stl}$ mutants contain ~7 times more neutral lipid than *c655* mutants (Figure 3D). This data
328 suggests the growth defects in *mttp*$^{stl/stl}$ mutants result from defects in dietary lipid absorption in
329 the intestine.
330
331 Besides accumulating lipids in the intestine, Abetaliporoteinemia patients sometimes develop
332 hepatic steatosis (reviewed in (Lee and Hegele, 2014)). Similarly, hepatocyte-specific deficiency
333 of MTTP in mice causes TG and cholesterol to accumulate in the liver (Khatun et al., 2012;
334 Raabe et al., 1999). Therefore, we hypothesized that the adult zebrafish *mttp* mutants would
335 also exhibit liver steatosis. However, mutants were not different than wild-type (Figure 3C,D).
336 Moreover, there was no significant difference in measured lipids in the livers of the different fish
337 (Figure 3E). While we were surprised that the *mttp*$^{stl/stl}$ mutants had very little accumulation of
338 lipid in their livers, this is in agreement with findings that combined intestinal and liver deficiency
339 of Mttp in mice results in accumulation of TG in the intestine, but not in the liver (Iqbal et al.,
340 2015).
341
342 ***Mttp mutations reduce the size and number of ApoB-containing lipoproteins in vivo.*** To
343 understand how the *mttp* mutations affect the production and size of B-lps during embryonic
344 development, we crossed the *mttp*^{stl} and *mttp*^{c655} mutations into our LipoGlo reporter line
345 (Thierer et al., In Press). These fish express an in-frame fusion of the engineered luciferase
346 reporter Nanoluc at the C-terminus of the Apolipoprotein Bb.1 gene (Figure 4A). Since ApoB is

347 an obligate structural component of B-lps with only one copy per lipoprotein particle (Elovson et
348 al., 1988), the relative number of tagged lipoprotein particles can be quantified using the Nano-
349 Glo assay in extracts from transgenic fish (Thierer et al., In Press). B-lp levels were measured
350 in whole fish lysate throughout embryonic development from 2–6 dpf. During this time, the fish
351 are relying solely on yolk lipids and the ApoB quantity measurements reflect lipoprotein particles
352 in the secretory pathway in the YSL, in the circulation, and in cells prior to degradation of
353 endocytosed lipoproteins. Wild-type embryos exhibit an increase in B-lp particle number from
354 2–3 dpf as yolk lipid is packaged into lipoproteins, and then numbers decline as the yolk is
355 depleted and the lipids in the lipoproteins are taken up by target tissues and lipoprotein particles
356 are degraded (Figure 4B). The *mttp*$^{c655/c655}$ embryos have the same relative number of ApoB
357 particles as wild-type embryos at 2 dpf, but from 3-6 dpf the numbers of particles never reach
358 wild-type levels and decline more rapidly. In contrast, *mttp*$^{stl/stl}$ embryos have profound defects
359 in B-lp production, since amount of ApoB is significantly lower than wild-type siblings at 2 dpf
360 (Figure 4B) (Thierer et al., In Press).
361
362



363
364

365 **Figure 4: The *stl* and *c655* mttp mutations have differential effects on ApoB-containing**
366 **lipoprotein number, size and distribution *in vivo*.** (A) LipoGlo fish express the Nanoluc®
367 luciferase enzyme as a C-terminal fusion on ApoBb.1 as a result of TALEN-based genomic
368 engineering. (B) LipoGlo signal (RLU: relative luminescence units) in WT, *mttp*$^{stl/stl}$ and
369 *mttp*$^{c655/c655}$ fish throughout embryonic development (2–6 dpf). Results represent pooled data
370 from 3 independent experiments, n = 22–34 fish/genotype/time-point. Significance was
371 determined with a Robust ANOVA, Games-Howell post-hoc tests were performed to compare
372 genotypes at each day of development, and p-values were adjusted to control for multiple
373 comparisons, a = WT vs. *mttp*$^{stl/stl}$, $p<0.001$, b = *mttp*$^{c655/c655}$ vs. *mttp*$^{stl/stl}$, $p<0.001$, c = WT vs.
374 *mttp*$^{c655/c655}$, $p<0.001$, d = WT vs. *mttp*$^{stl/stl}$, $p<0.05$. (C) Representative whole-mount images of
375 B-lp localization using LipoGlo chemiluminescent microscopy in WT, *mttp*$^{stl/stl}$, and *mttp*$^{c655/c655}$
376 fish throughout development; scale = 1 mm. Graphs represent pooled data from 3 independent
377 experiments, n = 13–19 fish/genotype/time-point; *mttp*$^{stl/stl}$ had a significantly different ApoB
378 localization from WT and *c655/c655*, $p<0.001$, Robust ANOVA. Post-hoc analysis reveals
379 statistical differences at all developmental stages $p<0.05$–0.001 (Games-Howell). (D)
380 Representative LipoGlo PAGE gels and quantification of B-lp size distribution from whole
381 embryo lysates during development. B-lps are divided into four classes based on mobility,
382 including zero mobility (ZM), and three classes of serum B-lps (VLDL, IDL and LDL). Graphs
383 show subclass abundance for WT, *mttp*$^{stl/stl}$, and *mttp*$^{c655/c655}$ fish at each day of embryonic
384 development as described in (Thierer et al., In Press). Results represent pooled data from n = 9
385 samples/genotype/time-point; at each particle class size, there were statistically significant
386 differences between genotypes (Robust ANOVA, $p<0.001$). Games-Howell post-hoc analysis
387 revealed numerous differences between genotypes at each developmental stage, see figure
388 supplement 2. (E) Representative whole-mount images of LipoGlo microscopy and Oil Red O
389 imaging in 15-dpf embryos chow-fed for 10 days and fasted ~18hrs prior to fixation; scale = 1
390 mm. Livers (outlined) are magnified for clarity in insets on right. Results represent pooled data
391 from 3 independent experiments, n = 15 fish/genotype/time-point.
392
393
394 To assess the localization of the B-lps throughout the embryos during development, we fixed
395 the embryos expressing ApoBb.1-nanoluc and performed chemiluminescent whole-mount
396 imaging (Figure 4C). Wild-type and *mttp*$^{c655/c655}$ embryos exhibit a similar distribution pattern of
397 LipoGlo throughout 2–4 dpf, but consistent with the quantitative assay, the signal in the head

398 and trunk decline more rapidly in *mttp*$^{c655/c655}$ fish. By 6 dpf, both wild-type and *mttp*$^{c655/c655}$ fish
399 show an accumulation of ApoB in the liver and the spinal cord (Figure 4C) (Thierer et al., In
400 Press). The LipoGlo in the *mttp*$^{stl/stl}$ embryos shows a very different pattern, in that it is
401 predominantly localized to the YSL/viscera at all stages and is at very low levels throughout the
402 rest of the body (Figure 4C). This is consistent with the prolonged retention of the opaque yolk
403 phenotype (Figure 1 – figure supplement 3) and suggests that the *mttp*$^{stl/stl}$ mutants are more
404 defective at secreting B-lps from the yolk than the *mttp*$^{c655/c655}$ mutants.
405
406 To determine whether the *mttp* mutations alter the size distribution of B-lps, we performed
407 native polyacrylamide gel electrophoresis of larval homogenates expressing the LipoGlo
408 reporter. Following electrophoretic separation and chemiluminescent imaging of the gels, B-lps
409 were classified into four different classes based on their migration distance (zero mobility, very
410 low-density lipoproteins (VLDL), intermediate-density lipoproteins (IDL), or low-density
411 lipoproteins (LDL)(Thierer et al., In Press). During development, the pattern of B-lps in wild-type
412 embryos is initially defined by VLDL (2 dpf), but expands to include IDL and LDL by 3–4 dpf as
413 the VLDL particles produced by the YSL are lipolyzed by the body tissues (Figure 4D, Figure 4
414 – figure supplement 1, (Thierer et al., In Press)). By 5–6 dpf, the yolk is depleted and any
415 remaining small particles are degraded. In contrast, both *mttp*$^{stl/stl}$ and *mttp*$^{c655/c655}$ embryos
416 produce very few VLDL particles (Figure 4D, 2 dpf), and instead, produce predominantly IDL
417 and LDL-sized particles.
418

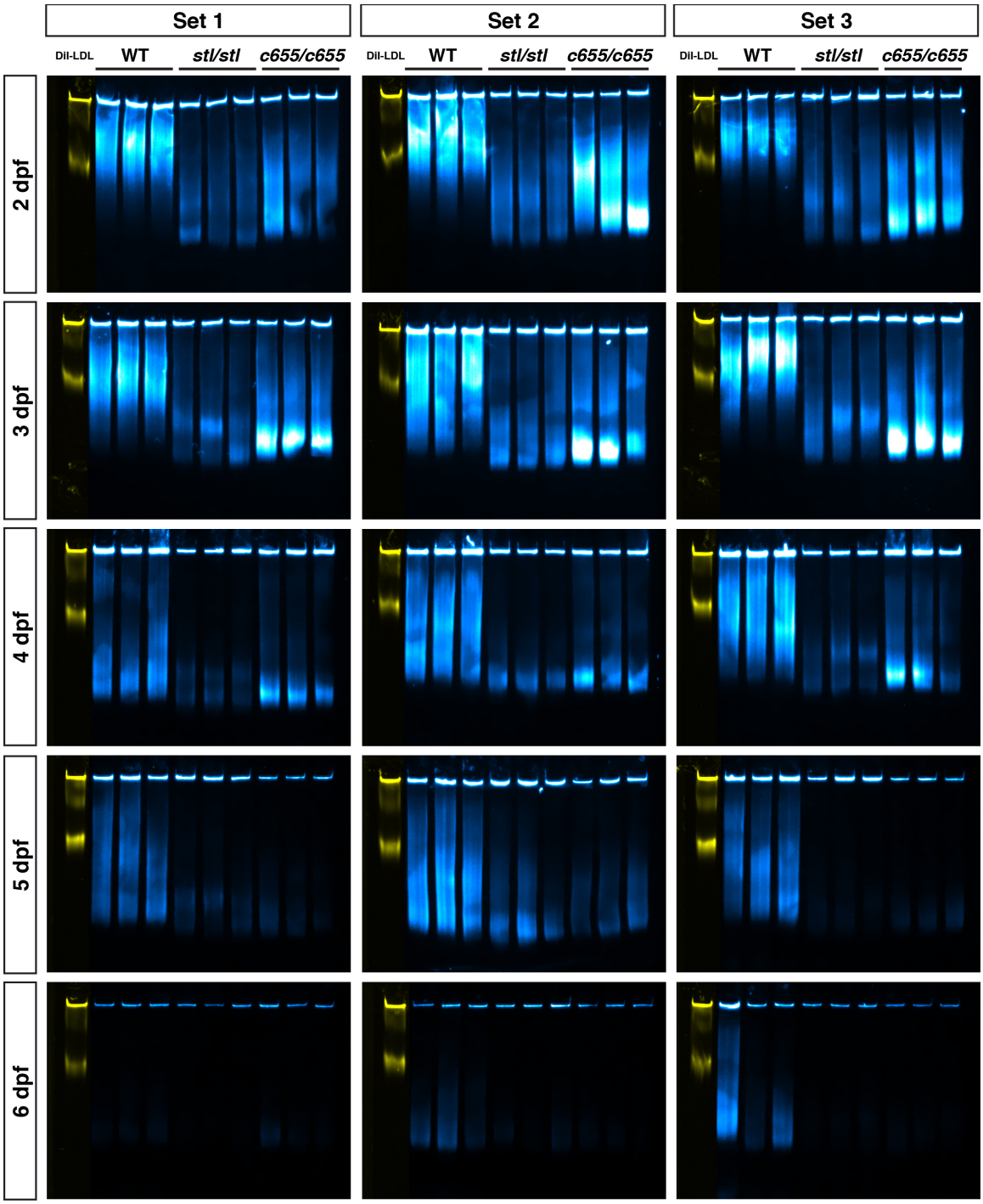


Figure 4 – figure supplement 1: LipoGlo lipoprotein gel primary data. Original gels corresponding to the data in Figure 4D. Each gel shows a composite image of the fluorescent DiI-LDL migration standard (yellow) and LipoGlo emission chemiluminescent exposure (blue) from WT, *mttp*$^{stl/stl}$ and *mttp*$^{c655/c655}$ fish. Gels were analyzed as detailed in (Thierer et al., In Press) and lipoprotein particles were binned into four classes based on migration relative to the

426 DiI-LDL standard, including zero mobility (ZM), and three classes of serum B-lps (VLDL, IDL
427 and LDL).
428
429 **Figure 4 – figure supplement 2:**

	Zero Mobility Band	VLDL Fraction	IDL Fraction	LDL Fraction
2 dpf				
WT vs. *stl/stl*	n.s.	p < 0.0005	p < 0.0005	p < 0.0005
WT vs. *c655/c655*	n.s.	p < 0.0005	p < 0.0005	p = 0.01
stl/stl vs. *c655/c655*	p = 0.005	n.s.	n.s.	
3 dpf				
WT vs. *stl/stl*	p = 0.015	p < 0.0005	n.s.	p < 0.0005
WT vs. *c655/c655*	n.s.	p < 0.0005	n.s.	p < 0.0005
stl/stl vs. *c655/c655*	p = 0.005	n.s.	n.s.	p < 0.0005
4 dpf				
WT vs. *stl/stl*	n.s.	p = 0.003	p = 0.005	P < 0.005
WT vs. *c655/c655*	n.s.	n.s.	p < 0.0005	P < 0.005
stl/stl vs. *c655/c655*	n.s.	n.s.	n.s.	n.s
5 dpf				
WT vs. *stl/stl*	p < 0.0005	p < 0.0005	p < 0.0005	n.s.
WT vs. *c655/c655*	p = 0.025	p = 0.005	p < 0.0005	p = 0.005
stl/stl vs. *c655/c655*	n.s.	n.s.	n.s	n.s.
6 dpf				
WT vs. *stl/stl*	p < 0.0005	n.s.	p < 0.0005	n.s.
WT vs. *c655/c655*	p = 0.03	n.s.	p = 0.005	n.s.
stl/stl vs. *c655/c655*	n.s.	n.s.	n.s.	n.s.

430 n.s. = not significant
431
432
433 Taken together, the LipoGlo data from embryos indicates that the *mttp*$^{stl/stl}$ and *mttp*$^{c655/c655}$
434 embryos are producing and secreting fewer, smaller B-lps from the yolk, although the effect is
435 more severe in *stl* mutants. Given the gross accumulation of lipid in the intestines of the adult
436 *mttp*$^{stl/stl}$ mutants, compared to the *mttp*$^{c655/c655}$ fish (Figure 3C), we hypothesize that the *mttp*$^{stl/stl}$
437 mutants are also less effective at secreting chylomicrons from the enterocytes. To test this
438 hypothesis, we performed chemiluminescent imaging using the LipoGlo reporter in 15-dpf larvae
439 fed a chow diet for 10 days and then fasted overnight. Wild-type LipoGlo fish have abundant
440 ApoB throughout their circulation and tissues (73.1+/- 4.0% in head and trunk vs. 26.9 +/-4.0%
441 in viscera, mean +/- SD, n = 15 fish) (Figure 4E). The *mttp*$^{c655/c655}$ mutation does not prevent
442 secretion of ApoB to the body tissues (73.1 +/- 3.7% in head and trunk vs. 26.9 +/- 3.7% in
443 viscera). In contrast, the *mttp*$^{stl/stl}$ fish have abundant LipoGlo signal in their intestine, and much
444 less in other tissues compared to WT (41% +/- 11% in head and trunk vs. 59 +/- 11% in viscera,
445 p<0.001, Kruskall-Wallis & Dunn's Multiple Comparisons Test) (Figure 4E). In agreement,
446 staining the neutral lipid with Oil Red O indicates *mttp*$^{stl/stl}$ mutants retain substantial lipid in their

447 intestines, whereas *mttp*$^{c655/c655}$ mutant fish have less lipid remaining in their intestine, but do
448 accumulate some lipid in their livers (Figure 4E). This data argues that the *stl* mutation severely
449 reduces B-lp secretion, not only from the yolk, but also from the enterocytes, whereas the *c655*
450 mutation only mildly decreases ApoB secretion in both embryos and larvae.
451
452 ***The c655 mutation in zebrafish mttp disrupts the triglyceride transfer activity but not the***
453 ***phospholipid transfer activity of the MTP complex.*** The dissimilar phenotypes of *in vivo* B-lp
454 secretion between the *stl* and *c655* mutations suggest that the two mutations are differentially
455 affecting MTP function. To provide molecular explanations for the phenotypes observed in the
456 different zebrafish mutants and to dissect further how each of the mutations affects MTP
457 function, we used cell and *in-vitro*-based assays of MTP function. COS-7 cells expressing
458 human ApoB48 were co-transfected with either an empty vector (pcDNA3), or a vector
459 containing wild-type zebrafish *mttp*, *mttp-stl* or *mttp-c655*, all with C-terminal FLAG-tags. We
460 found that *stl* and *c655* expressing COS-7 cells have significantly reduced concentrations of
461 ApoB in the conditioned media compared to wild-type Mttp expressing cells (Figure 5A). ApoB
462 levels in the media of the *stl*-expressing cells were similar to cells transfected with empty vector,
463 indicating background secretion levels. The concentration of ApoB48 was significantly higher in
464 the media of *c655*-expressing COS-7 cells compared to the *stl*-expressing cells and cells
465 transfected with empty vector (Figure 5A). ApoB48 concentrations in the cell lysate of cells
466 expressing *stl* were significantly higher than wild-type Mttp and *c655*-expressing cells (Figure
467 5B). These data suggest that the *stl* mutation does not support ApoB48 secretion whereas the
468 *c655* mutation does support ApoB48 secretion, but with reduced efficacy compared to wild-type
469 Mttp.
470

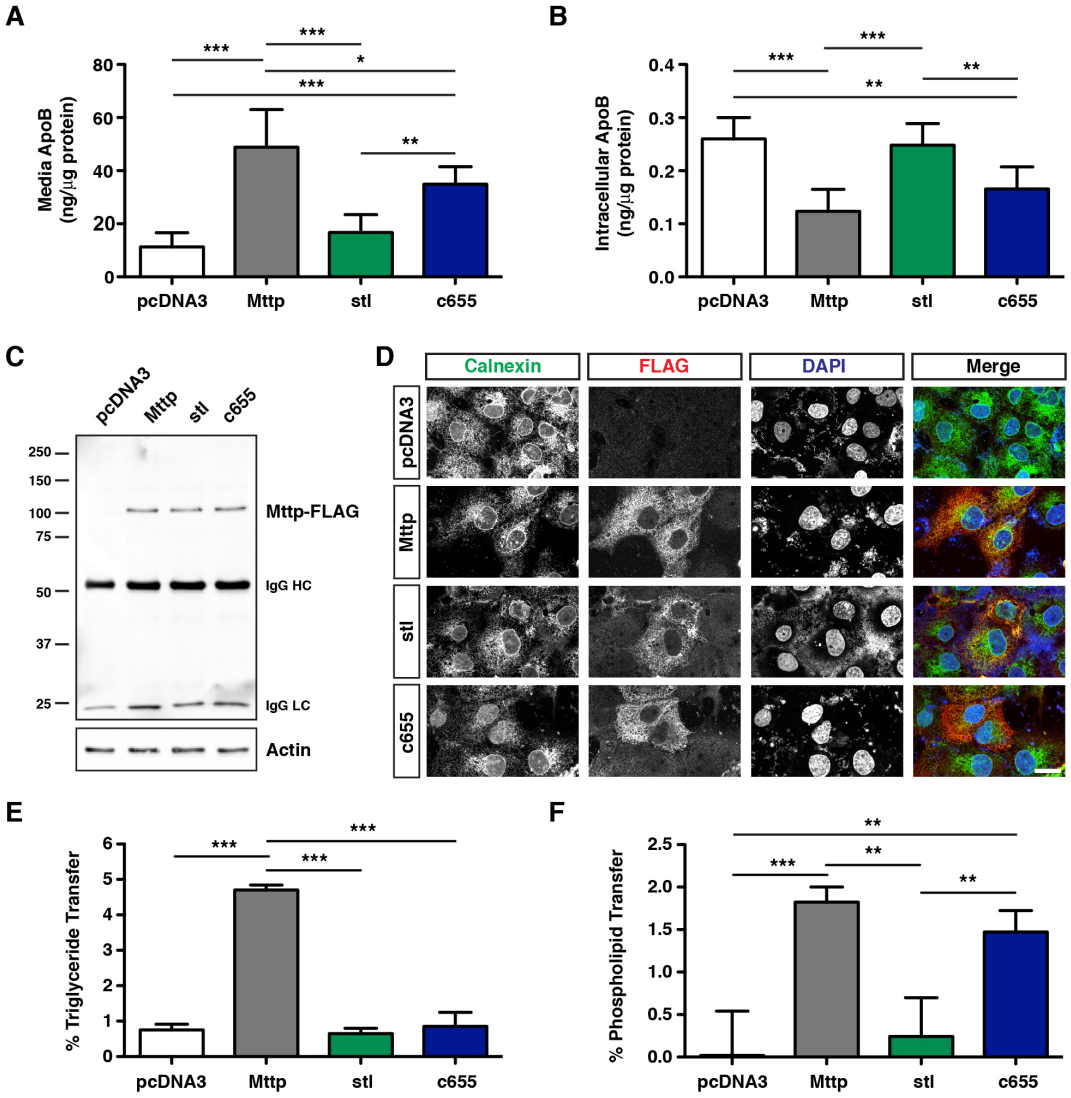
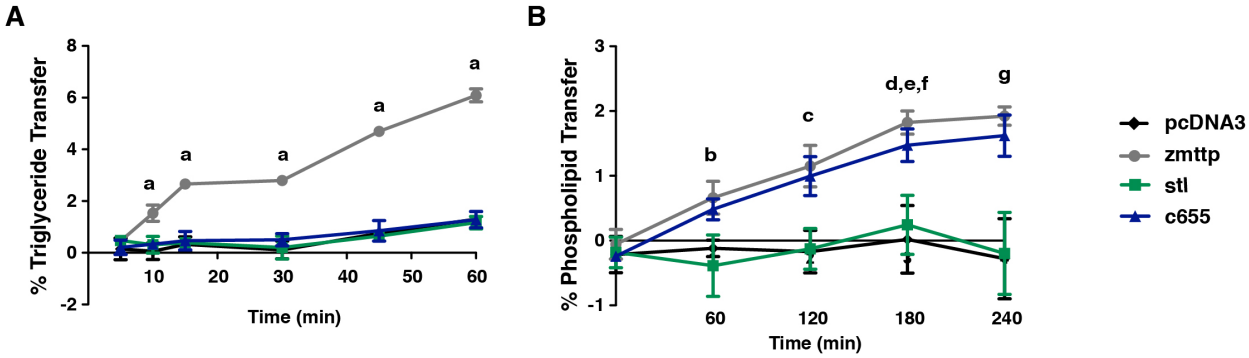


Figure 5: The *c655* mutation disrupts the triglyceride transfer activity, but not the phospholipid transfer activity of the zebrafish MTP complex. (A-B) COS-7 cells were first transfected with an expression vector for human apoB48 (5 μg), distributed equally in 6-well plates, and subsequently transfected with plasmids expressing either wild-type zebrafish *mttp*-FLAG, *mttp*^{stl}-FLAG, *mttp*^{c655}-FLAG, or empty vector (pcDNA3) (3 μg). After 72 h, ApoB48 was measured via ELISA in media (A) or in the cell (B). Data are representative of 7 independent experiments (each data point is the mean of 3 technical replicates), mean +/- SD, One-Way ANOVA with Bonferroni post-hoc tests, * $p<0.05$, ** $p<0.01$, *** $p<0.001$. (C) Cos-7 cells were transfected as described above and FLAG-tagged proteins were immunoprecipitated from cell lysates using anti-FLAG antibodies and eluted with FLAG peptides. Representative Western

484 blot on eluted fractions indicates equal concentrations of the various Mttp-FLAG proteins; actin
485 blot indicates equal loading of cell lysate. (D) Representative immunofluorescent staining using
486 anti-FLAG (red) and anti-Calnexin (green) antibodies in COS-7 cells expressing wild-type or
487 mutated *mttp*-FLAG constructs; scale = 25 μm. (E) COS-7 cells were transfected with plasmids
488 expressing pcDNA3, wild-type *mttp-FLAG*, or mutant *mttp*-FLAG constructs. Cells were lysed
489 and 60 μg of protein was used to measure the % triglyceride transfer of NBD-triolein from donor
490 to acceptor vesicles after 45 minutes; n = 3 (each n is the mean of three technical replicates
491 from independent experiments), mean +/- SD, One-way ANOVA with Bonferroni post-hoc tests,
492 *** $p<0.001$. (F) Wild-type and mutant Mttp proteins were purified using anti-FLAG antibodies
493 and used to measure the % transfer of NBD-labeled phosphoethanolamine triethylammonium
494 from donor to acceptor vesicles after 180 minutes; n = 3 (each n is the mean of 3 technical
495 replicates from independent experiments), mean +/- SD, randomized block ANOVA with
496 Bonferroni post-hoc tests, ***$p<0.001$.
497
498 To eliminate the possibility that the *stl* mutation did not support ApoB48 secretion due to low
499 expression, Mttp, *stl*, and *c655* were precipitated using anti-FLAG antibodies from cell lysates
500 and were subjected to Western blot analysis. We found that there was no difference in the
501 expression of wild-type Mttp and Mttp mutants in COS-7 cells (Figure 5C). Another reason for
502 the *stl* mutation to be deficient in supporting ApoB48 secretion could be due to protein mis-
503 localization. To check this, we immunostained Mttp-expressing COS-7 cells with anti-Calnexin
504 and anti-FLAG antibodies. Confocal imaging shows that the wild-type Mttp and both mutant
505 proteins were properly localized in the endoplasmic reticulum (Figure 5D). Additionally, the
506 percentage of cells expressing the FLAG-tagged proteins were similar among all groups (Mttp-
507 FLAG 37%, Mttp-*stl* 31%, Mttp-*c655* 41% transfection efficiency). These studies suggest that
508 *stl* and *c655* mutant Mttp proteins are expressed to similar levels and are in the ER, where
509 lipoprotein assembly occurs.
510
511 Next, we hypothesized that the *stl* mutant protein may not support ApoB48 secretion because it
512 might be defective in lipid transfer activity. Triglyceride transfer assays performed using cell
513 lysates showed that both of the mutant forms of Mttp (*stl* and *c655*) have significantly decreased
514 triglyceride transfer activity compared to wild-type zebrafish Mttp (Figure 5E, Figure 5 – figure
515 supplement 1a). The *stl* mutant form was also found to be defective in phospholipid transfer

516 activity compared to wild-type (Figure 5F, Figure 5 – figure supplement 1b). Contrary to this,
517 the *c655* mutant form had similar phospholipid transfer activity to wild-type Mttp (Figure 5F,
518 Figure 5 – figure supplement 1b). These data suggest that the *c655* mutation in *mttp* impairs
519 triglyceride transfer, but not phospholipid transfer activity. In contrast, the *stl* mutation is
520 defective in both transfer activities. These differences in lipid transfer activities between the *stl*
521 and *c655* mutations provide a biochemical explanation for their differential abilities to support
522 ApoB48 secretion and the different phenotypes observed in the fish.
523
524



a; pcDNA3 vs. all others, p<0.001
b; zmttp vs. stl, p<0.05
c; pcDNA3 & stl vs. zmttp & c655, p<0.01
d; pcDNA3 vs. zmttp & c655, p<0.001
e; stl vs. zmttp, p<0.001
f: stl vs. c655, p<0.01
g; pcDNA3 & stl vs. zmttp & c655, p<0.001

525
526
527 **Figure 5 – figure supplement 1: Triglyceride and phospholipid transfer assay time-**
528 **course data.** Measurements for triglyceride (A) and phospholipid transfer (B) by zebrafish
529 *mttp*-FLAG and mutants over a time-course. The single time-points depicted in the bar graphs
530 of figures 5E & F, correspond to the 45 min & 180 min (triglyceride and phospholipid transfer,
531 respectively) time-points in the curves shown. For both, n = 3 (each n is the mean of 3 technical
532 replicates from independent experiments), mean +/- SD, Repeated Measures ANOVA with
533 Bonferroni post-hoc tests, significance as noted in figure.
534
535
536 ***An orthologous c655 mutation in human MTTP (G865V) also disrupts the triglyceride***
537 ***transfer activity but not phospholipid transfer activity.*** The M subunit of MTP shares
538 homology with lipovitellin, a lipid transfer protein in the yolk of oviparous animals (Banaszak et
539 al., 1991). Homology modeling with the crystal structure of lamprey lipovitellin (PDB ID:

540 1LSH)(Anderson et al., 1998; Raag et al., 1988; Thompson and Banaszak, 2002) predicts three
541 major structural domains in human MTTP: an N-terminal beta-barrel, a middle alpha-helical
542 domain, and a C-terminal domain composed of two beta-sheets that form a hydrophobic lipid-
543 binding cavity (Hussain et al., 2003a; Mann et al., 1999; Read et al., 2000). The amino acid
544 sequence of the zebrafish Mttp is 54% identical (72% similar) to that of human MTTP and the
545 predicted secondary and tertiary structures are highly conserved (Figure 6A).
546
547 The leucine residue mutated in the *stl* mutant fish (L475P) is conserved in human MTTP (L477)
548 and lies within a highly conserved stretch of amino acids located in the middle alpha-helical
549 domain of MTTP (Figure 6A). Analysis of mutations in patients with Abetalipoproteinemia has
550 shown that missense mutations in the alpha-helical domain often prevent lipid transfer activity
551 and result in loss of ApoB secretion (Reviewed in (Walsh and Hussain, 2017)). Therefore, it is
552 likely that the *stl* L475P mutation alters the structure of the lipid-binding cavity in a similar
553 manner to these patient mutations.
554
555 The glycine residue mutated in the C-terminus of *c655* mutants (G863V) is also conserved in
556 the human sequence (G865) (Figure 6A). Unfortunately, it is unknown where the G865 residue
557 is located in relation to the lipid-binding cavity because the C-terminal sequence of MTTP
558 diverges from that of lipovitellin, so it is not modeled in the predicted MTTP structure. However,
559 because the predicted tertiary structure of zebrafish Mttp is very similar to that of human MTTP
560 and the sequence in the C-terminus is highly conserved (Figure 6A), we hypothesized that
561 introduction of the *c655* mutation into the human MTTP protein would also cause loss of
562 triglyceride transfer, but retention of phospholipid transfer activity. To test this hypothesis, we
563 performed site-directed mutagenesis on the human *MTTP*-FLAG (hMTTP) plasmid and
564 assessed the function of the MTTP (G865V) mutant protein.
565

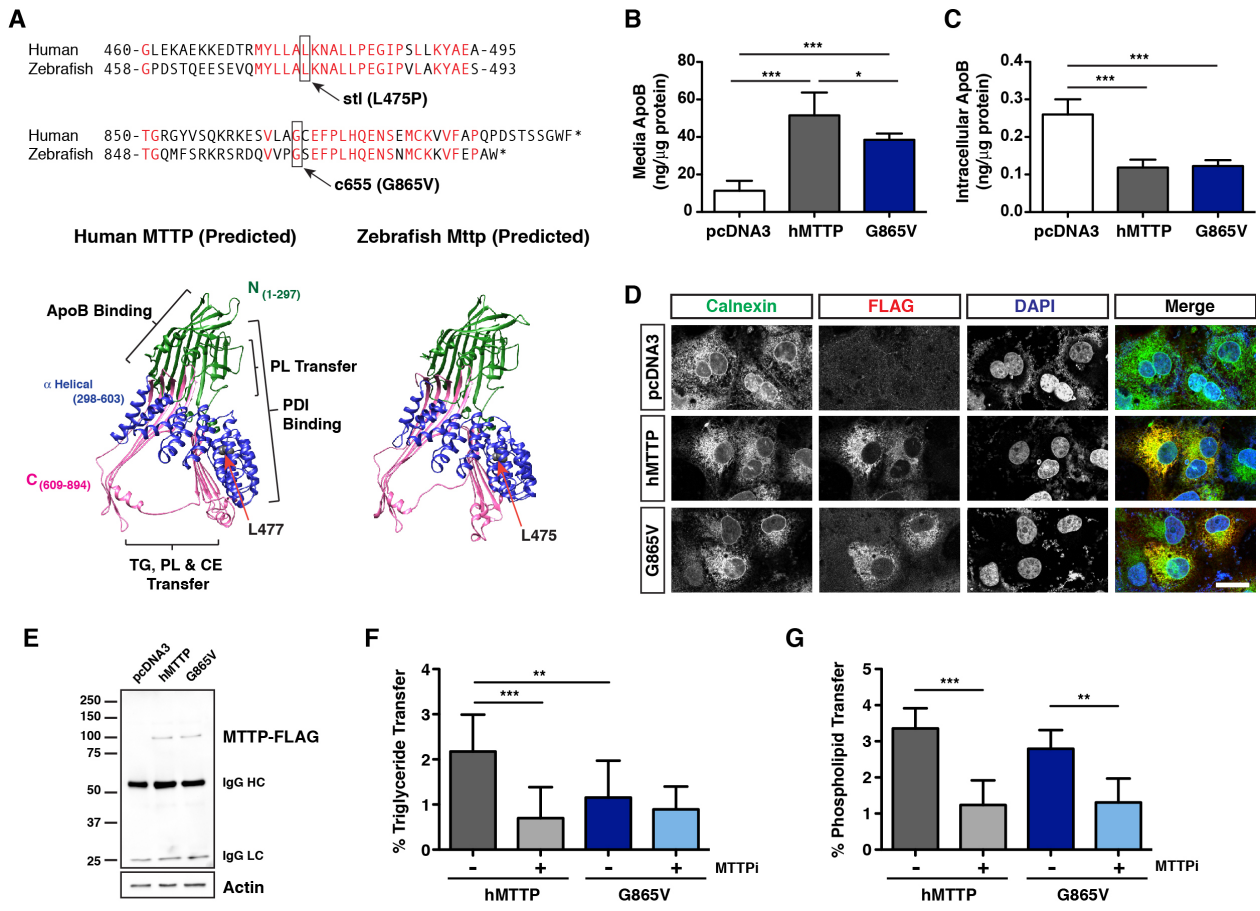


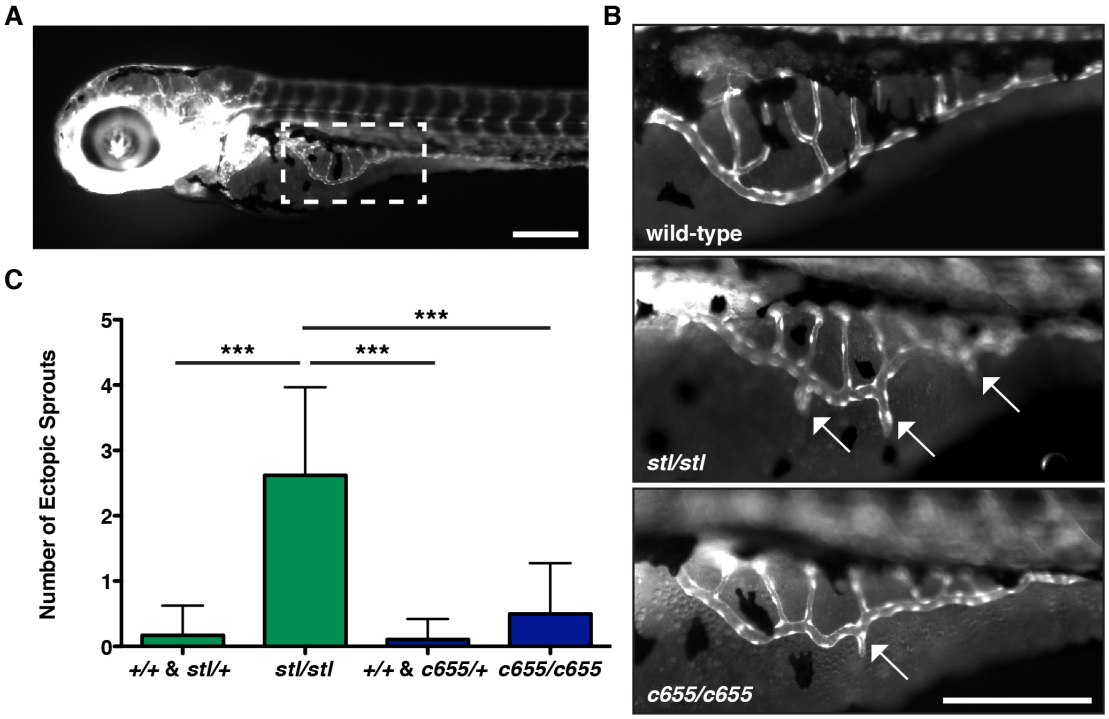
Figure 6: The *c655* mutation in human MTTP also disrupts the triglyceride transfer activity, but not the phospholipid transfer activity of the MTP complex. (A) Alignment of human and zebrafish Mttp amino acid sequences surrounding the *stl* or *c655* mutations. Ribbon diagrams of the predicted tertiary structures of human MTTP and zebrafish Mttp modeled using Phyre2 (Kelley et al., 2015) based on the lamprey lipovitellin structure (Anderson et al., 1998). The predicted phospholipid transfer sites in the N-terminal region (green) and the C-terminal region (pink, composed of two beta-sheets that form a hydrophobic lipid-binding pocket) are labeled. The location of the conserved human residue (L477) corresponding to the zebrafish *stl* (L475P) mutation is noted. The C-terminal region containing the *c655* mutation diverges from the lipovitellin structure and is not reliably modeled. (B,C) The *c655* mutation (G865V) was introduced into the human *MTTP*-FLAG plasmid. COS-7 cells were co-transfected with human ApoB48 and either wild-type human *MTTP*-FLAG, *MTTP*(G865V)-FLAG or empty pcDNA3 plasmids. After 72 h, apoB48 was measured via ELISA in media (B) or in the cell (C). Data are representative of 7 independent experiments (each data point is the mean of 3 technical replicates), pcDNA3 control data is re-graphed from figure 5A & 5B (data for 5A, 5B, 6B & 6C

583 were generated together); mean +/- SD, One-Way ANOVA with Bonferroni post-hoc tests, *
584 p<0.05, *** p<0.001. (D) Immunofluoresence in COS-7 cells expressing wild-type human
585 MTTP-FLAG or human MTTP(G865V)-FLAG using anti-FLAG (red) and anti-Calnexin (green)
586 antibodies; scale = 25 μm. (E) COS-7 cells were transfected and FLAG-tagged human MTTP
587 proteins were immunoprecipitated from cell lysates using anti-FLAG antibodies and eluted with
588 FLAG peptides. Representative Western blot on eluted fractions indicates equal concentrations
589 of the various MTTP-FLAG proteins; actin blot indicates equal loading of cell lysate. (F) COS-7
590 cells were transfected with plasmids expressing human wild-type or *MTTP*(G865V)-FLAG
591 constructs. Cells were lysed and 60 μg of protein was used to measure triglyceride transfer
592 activity in the presence or absence of the MTTP inhibitor lomitapide (1 μM)(% after 45 minutes);
593 n = 9 (3 measurements from each of 3 independent experiments), mean +/- SD, One-way
594 ANOVA with Bonferroni post-hoc tests, ** p<0.01, ***p<0.001). (G) Wild-type and mutant MTTP
595 proteins were purified using anti-FLAG antibodies and used to measure phospholipid transfer in
596 the presence or absence of lomitapide (1 μM) (180 minutes); n = 9 (3 measurements from each
597 of 3 independent experiments), mean +/- SD, randomized block ANOVA with Bonferroni post-
598 hoc tests, ** p<0.01, ***p<0.001.
599
600
601 COS-7 cells expressing the G865V mutant protein secreted slightly lower amounts (~75%) of
602 ApoB48 into the media compared to wild-type hMTTP, but this was significantly higher than the
603 background levels seen in cells transfected with the empty vector (pcDNA3) (Figure 6B).
604 ApoB48 concentrations in the cell lysate of cells expressing hMTTP and G865V mutant proteins
605 were similar (Figure 6C). These data suggest that the G865V mutation supports ApoB48
606 secretion albeit at lower efficiency. Attempts were made to explain the mechanisms for lower
607 efficiency in supporting ApoB secretion. Wild-type hMTTP-FLAG and G865V-FLAG were both
608 localized to the ER (Fig. 6D), transfection efficiency was similar (36% hMTTP, 35% G865V),
609 and the proteins were immunoprecipitated at similar levels from cell lysates (Figure 6E).
610 Triglyceride transfer assays with cell lysates of COS-7 cells expressing hMTTP-FLAG and
611 G865V-FLAG plasmids showed that the mutant G865V protein has significantly decreased
612 (~53%) triglyceride transfer activity compared to wild-type hMTTP, comparable to the activity
613 level of the WT hMTTP in the presence the MTTP inhibitor lomitapide (1 μM) (Figure 6F). In
614 contrast, the phospholipid transfer activity of hMTTP and G865V were not significantly different,

615 and the activity of both alleles were inhibited to equal extent by lomitapide (Figure 6G). These
616 data suggest that the *c655* mutation in both the zebrafish Mttp (G863V) and human MTTP
617 protein (G865V) results in significant loss of triglyceride transfer activity, but has no effect on
618 phospholipid transfer activity.
619
620
621
622 **DISCUSSION**
623
624 The characterization of the zebrafish *mttp c655* mutation provides the first evidence that the
625 triglyceride and phospholipid transfer functions of a vertebrate microsomal triglyceride transfer
626 protein can be dissociated and identifies the putative region responsible for triglyceride transfer
627 activity. Previous sequence comparisons of invertebrate and vertebrate orthologues of MTTP
628 strongly suggested that acquisition of triglyceride transfer activity during evolution was the result
629 of many changes in the lipid-binding cavity (Rava and Hussain, 2007), so it was unexpected that
630 one missense mutation in the C-terminus selectively eliminated triglyceride transfer activity.
631 Additionally, all of the characterized missense mutations from patients with
632 Abetalipoproteinemia cause MTP to be either absent or deficient in both phospholipid and
633 triglyceride transfer activities (Berthier et al., 2004; Di Filippo et al., 2012; Di Leo et al., 2005;
634 Khatun et al., 2013; Miller et al., 2014; Narcisi et al., 1995; Rehberg et al., 1996; Ricci et al.,
635 1995; Walsh et al., 2016; Walsh and Hussain, 2017; Walsh et al., 2015; Wang and Hegele,
636 2000).
637
638 While we do not know where the C-terminus is located in relation to the lipid-binding cavity
639 (Figure 6A), one of the Abetalipoproteinemia mutations (G865X) results in a C-terminal
640 truncation of 30 amino acids that prevents binding to PDI, thus resulting in loss of MTP protein
641 (Ricci et al., 1995). Subsequent analysis of constructs *in vitro* indicated that deletion of the last
642 20 amino acids (Δ20, S875X), or mutating the cysteine at position 878 (C878S), reduced the
643 expression of the protein to <15% of wild-type MTTP levels and abolished triglyceride transfer
644 activity (Narcisi et al., 1995). This cysteine residue is conserved in MTTP orthologues from
645 human to *C. elegans*, suggesting that it forms a disulfide bond essential for the tertiary structure
646 of the protein. The authors argue that the residual protein produced must be binding to PDI,
647 otherwise, no protein would be detected; however, it is unclear whether either of these mutated
648 proteins are still capable of transferring phospholipid (Narcisi et al., 1995). In contrast to these

649 mutations, the mutation of glycine to valine at position 865 in human MTTP (or 863 in zebrafish
650 mttp) does not reduce protein expression or alter the localization of MTTP in the ER of COS-7
651 cells (Figure 5D, 6D), indicating that the mutation does not interfere with PDI binding. This
652 suggests that the proposed disulfide bond is intact and that the G865V mutation may be more
653 directly affecting lipid transfer. Our data show that this mutation lacks the ability to transfer
654 triglycerides *in vitro*, but it is unclear whether the mutation also prevents binding of triglyceride in
655 the lipid-binding cavity. Perhaps binding can still occur, which would be consistent with the
656 evolutionary data, and maybe the C-terminus is necessary for transfer activity. Analyses of
657 Drosophila, zebrafish, or human MTTP chimeric proteins may help test this hypothesis in the
658 future. Additionally, in order to better understand how the *c655* mutation specifically inhibits
659 triglyceride transfer, a crystal structure including the C-terminus will likely be necessary.
660
661 The production of B-lps in the ER of the intestine and liver is thought to occur in two steps. In
662 the first step, MTP transfers lipids to ApoB as it is translated to form small primordial particles.
663 In the second step, it has been suggested that fusion of ApoB-free lipid droplets in the lumen of
664 the ER fuse to expand the lipoprotein core ("core expansion") (Alexander et al., 1976; Boren et
665 al., 1994; Hamilton et al., 1998; Wang et al., 1997). There is evidence to suggest that MTP is
666 also responsible for producing these ER-lumenal lipid droplets (Kulinski et al., 2002). Using our
667 LipoGlo assays, we have shown that the *c655* mutant fish produce small, homogenous,
668 particles, whereas the wild-type embryos form VLDL-sized lipoproteins in the YSL at 2–3 dpf
669 (Figure 4D). We have made similar observations in liver-specific *Mttp* KO mice expressing
670 Drosophila *Mttp*, which has robust phospholipid transfer activity, but is deficient in triglyceride
671 transfer (Khatun et al., 2012). Expression of fly Mttp resulted only in production of small B-lps,
672 but human MTTP rescued the particle size (Khatun et al., 2012). Therefore, the phospholipid
673 transfer activity of MTP may be crucial in the generation of the small homogenous particles
674 representative of the first step of lipoprotein assembly, whereas triglyceride transfer might be
675 primarily responsible for core expansion.
676
677 Although the effect of the *c655* mutation on the molecular function of the protein was
678 unexpected, the lack of intestinal or hepatic steatosis is also consistent with our previous data
679 with Drosophila Mttp expression in the livers of liver-specific Mttp-null mice. The phospholipid-
680 rich high-density B-lps produced by the fly Mttp in hepatocytes partially restore plasma lipid
681 levels and reduce liver steatosis (Khatun et al., 2012). Similarly, transfer of phospholipid and

682 production of small B-lps in the *c655* mutant fish is not only sufficient for moving lipid from the
683 liver, but is also capable of moving enough dietary lipid and fat-soluble vitamins from the
684 intestine to prevent intestinal steatosis and support normal growth (Figure 3,4). Furthermore,
685 retention of phospholipid transfer may also improve the health of the fish in ways that are
686 independent of lipoprotein production. For example, MTP-dependent phospholipid transfer has
687 been shown to be important for biogenesis and cell surface expression of CD1d and possibly
688 other lipid-antigen-presenting molecules (Dougan et al., 2005).
689
690 In contrast to the health and viability of *c655* mutants, the *stl* mutant fish that survive have gross
691 lipid accumulation in their intestine and severe growth defects (Figure 3,4), which is reminiscent
692 of patients with Abetalipoproteinemia. The mutant protein localizes normally to the ER, but is
693 deficient in triglyceride transfer, phospholipid transfer, and ApoB secretion (Figure 5). In the
694 predicted model of the tertiary structure of Mttp, this mutation is located in the middle alpha-
695 helical domain, not facing the lipid-binding cavity. The location and characterization of this
696 mutation is very similar to that of the described Y528H and S590I Abetalipoproteinemia
697 mutations, and it is likely that all three mutations prevent lipid transfer by altering the tertiary
698 structure of the M subunit (Khatun et al., 2013; Miller et al., 2014).
699
700 Counter to the original characterization of *stl* mutants (Avraham-Davidi et al., 2012), we show
701 that the *stl* mutants can survive to adulthood, especially when they are not competing with
702 siblings for resources. We hypothesize that during the maintenance of this mutant line since its
703 original characterization, a modifier has been eliminated that, when present in the *stl*
704 background, was incompatible with life. In support of this hypothesis, the excessive sprouting
705 angiogenesis defect for which the *stalactite* mutation was named (Avraham-Davidi et al., 2012),
706 was also not as severe as originally described (Supplementary Figure S1). Whether the
707 proposed modifier directly affects the secretion of B-lps, or some other aspect of development,
708 is currently unclear. While the *stl* mutation severely decreases B-lp number and particle size
709 (Figure 4D), some ApoB is still noted in the body of the $mttp^{stl/stl}$ embryos and larvae (Figure 4C,
710 4E). This suggests that there is enough transport of necessary lipids and fat-soluble vitamins by
711 the small numbers of B-lps produced by the YSL and intestine to support development.
712



713
714 **Figure S1: *c655/c655* embryos exhibit fewer ectopic angiogenic segments extending**
715 **from the subintestinal vessels than *stl/stl* embryos.** (A) The developing vasculature is
716 visualized in the *Tg(fli:eGFP)*^{y1} transgenic zebrafish line (Lawson and Weinstein, 2002). The
717 subintestinal vessels (boxed region) grow bilaterally onto the dorsolateral surface of the yolk
718 sac. Scale = 200 μM. (B) Representative wide-field images of *Tg(fli:eGFP)*^{y1} in wild-type,
719 *mttp*$^{stl/stl}$ or *mttp*$^{c655/c655}$ embryos at 3.5 dpf. Ectopic sprouts extending ventrally from the
720 subintestinal vein are more common in *mttp*$^{stl/stl}$ mutant embryos than in *mttp*$^{c655/c655}$ mutant
721 embryos. Scale = 200 μM. (C) Quantification of the average number of ectopic sprouts in *mttp*
722 mutants and siblings on 3.5 dpf. Results represent pooled data from 3 independent
723 experiments, n = 28-36 total embryos/genotype group; mean +/- SD, Kruskall-Wallis with Dunn's
724 Multiple Comparison test, *** $p<0.001$.
725
726
727 The *c655* mutants were initially identified due to the abnormal opaque appearance of their yolks,
728 which is also a component of the *stl* mutant phenotype (Figure 1). We show that this opacity is
729 due to the abnormal accumulation of lipid droplets in the yolk syncytial layer (Figure 2). As the
730 lipids are liberated from the yolk, they are rapidly re-esterified to triglyceride and phospholipid
731 and packaged into B-lps in the endoplasmic reticulum of the YSL. Because the *mttp* mutant fish
732 are defective at producing (*stl*) and/or expanding (*c655*) B-lps, the re-esterified lipids are

733 packaged instead into cytoplasmic lipid droplets. In contrast, cytoplasmic lipid droplets are only
734 very rarely noted in the YSL of wild-type embryos, suggesting that the rate of yolk lipid break-
735 down and the rate of B-lp production and secretion from the YSL are exquisitely coupled. We
736 also noted that the size range of the cytoplasmic lipid droplets in the YSL of the *c655* mutants
737 was much larger than in the *stl* mutants, with some LDs reaching 180 μm^2. As the surface area-
738 to-volume ratio decreases with increasing size spheres, the larger lipid droplets store more
739 neutral lipid relative to the surface of the phospholipid coat. Because the *c655* mutants can
740 transfer phospholipid to ApoB and secrete greater numbers of small dense B-lps than *stl*
741 mutants, we hypothesize the retained neutral lipid is stored in larger lipid droplets because the
742 phospholipid is being secreted and less available to coat lipid droplets. However, it is unclear
743 whether the larger LDs form due to local production of triglycerides and cholesterol esters
744 directly on their surface, or due to lipid droplet fusion, or both mechanisms (reviewed in
745 (Olzmann and Carvalho, 2019; Walther et al., 2017)). The differences in the concentration and
746 size of LDs between the mutants may result in differential effects on the degree of light
747 scattering, which could explain the differences in opacity noted between mutants (Figure 1 –
748 figure supplement 3).
749
750 None of the missense mutations identified in patients with Abetalipoproteinemia have been
751 found to dissociate the lipid transfer activities of MTP (reviewed in (Walsh and Hussain, 2017)).
752 However, given that the adult *c655* mutant zebrafish are indistinguishable from wild-type
753 siblings, it is entirely possible that humans carrying a missense mutation that results in retention
754 of phospholipid transfer exist in the population. While these people would likely have low
755 plasma triglycerides, we would expect transport of fat-soluble vitamins to be normal. A thorough
756 search of publicly available large human GWAS databases (Global BioBank Engine, T2D
757 Knowledge Portal, GTEx Portal) did not reveal any coding variants near G865 other than the
758 G865X mutation discussed above. However, since one copy of wild-type MTTP is sufficient to
759 prevent fat malabsorption when faced with an oral fat load (Di Filippo et al., 2019), individuals
760 heterozygous for a mutation similar to *c655* may not present with any changes in plasma lipid
761 profiles.
762
763 Abnormally elevated levels of ApoB-containing lipoproteins and remnants promote
764 atherosclerosis, the leading cause of death in the United States (CDC, 2018). Inhibition of MTP
765 has long been considered a possible therapeutic target for lowering disease risk by inhibiting the

766 production of VLDL and chylomicrons (Jamil et al., 1996; Wetterau et al., 1998)(for review see
767 (Hussain and Bakillah, 2008; Walsh and Hussain, 2017)). Currently, the only MTP inhibitor
768 approved for use in patients is lomitapide (Juxtapid®), which inhibits triglyceride and
769 phospholipid transfer and reduces ApoB secretion (Robl et al., 2001)(Figure 6F,G). While this
770 drug effectively reduces LDL cholesterol, total cholesterol, and plasma ApoB levels, it is only
771 approved for patients with homozygous familial hypercholesterolemia, whose plasma
772 cholesterol and triglyceride levels are up to four times the normal levels resulting in premature
773 cardiovascular disease (Cuchel et al., 2014; Cuchel et al., 2013; FDA, 2012). While lomitapide
774 effectively lowers circulating lipid levels and reduces cardiovascular disease risk in these
775 patients, side effects include fat accumulation in the liver and adverse gastrointestinal events
776 including reflux, indigestion, abdominal pain, constipation, and diarrhea (Blom et al., 2017;
777 Cuchel et al., 2007; Cuchel et al., 2013).
778
779 The lack of intestinal and hepatic steatosis in the *c655* mutant fish suggests that an MTP
780 inhibitor that selectively targets triglyceride transfer activity could potentially lower plasma lipids
781 while preventing these gastrointestinal and liver side effects. This would not only improve the
782 quality of life for patients currently taking lomitapide, but may also expand MTP inhibitor use to
783 patients other than those with familial hypercholesterolemia. While one of the original MTP
784 inhibitors discovered, BMS-200150, was very effective at inhibiting triglyceride transfer, but less
785 effective (~30%) at inhibiting phospholipid transfer in vitro (Jamil et al., 1996), later studies on
786 purified MTP protein indicated the compound inhibits transfer of both lipid classes (Rava et al.,
787 2006) and that it was not effective in animal models (Wetterau et al., 1998). Now that we
788 appreciate that the triglyceride and phospholipid transfer functions of MTTP can be dissociated,
789 we argue that it may be worth re-evaluating the phospholipid transfer activity of any previously
790 identified compounds that inhibited triglyceride transfer activity of MTP, but failed to inhibit ApoB
791 secretion *in vitro*. Perhaps new compounds could be specifically designed to target the C-
792 terminal region of MTTP, although this will likely require a crystal structure of the MTP complex.
793
794 However, before a selective triglyceride transfer inhibitor could be considered as a therapeutic
795 for a wide range of patients with hyperlipidemia, it will be important to determine whether the
796 small lipoprotein particles produced by selective inhibition of triglyceride transfer are not
797 atherogenic. Small dense LDL has been found to have greater potential for causing
798 atherosclerosis than larger LDL sub-fractions and is a better predictor of cardiovascular disease

799 than total LDL-cholesterol (Austin et al., 1988; Bjornheden et al., 1996; Ivanova et al., 2017). In
800 future work, we want to evaluate whether the *mttp*$^{c655/c655}$ mutants are protected from
801 atherosclerosis because they produce fewer B-lps or are at higher risk because the B-lp
802 particles secreted are small and dense.
803
804 In conclusion, the unexpected discovery of the c655 missense mutation in *mttp* has provided
805 novel insight into the structure-function relationship of MTP, underlining the importance of
806 forward-genetic screening approaches to reveal aspects of biology that may otherwise be
807 missed. Our work provides the first evidence that the triglyceride and phospholipid transfer
808 functions of vertebrate MTP can be separated and that selective retention of phospholipid
809 transfer is sufficient for dietary fat absorption and normal growth. These results argue that
810 selective pharmacologic inhibition of triglyceride transfer may be a feasible therapeutic
811 approach to treat disorders of lipid metabolism.
812
813 **Acknowledgments**
814 We gratefully acknowledge Michael Sepanski for electron microscopy, Andrew Rock and
815 Carmen Tull for fish husbandry, Matthew Bray for assistance using R, Amy Kowalski for
816 synthesis of the pDESTTol2pA2-CMV: eGFP-CAAX plasmid, and Jennifer Anderson for help
817 editing the manuscript. We would also like to especially thank Philip Ingham, who provided the
818 *kif7* mutant strain in which we identified the *c655* mutation. This work was supported by NIH
819 grants R01 DK093399 (Farber, PI; Busch-Nentwich, Co-PI), R01 GM63904 (The Zebrafish
820 Functional Genomics Consortium; Ekker, PI, Farber, Co-PI), HL-137202-01A1 (Hussain, PI),
821 R56 DK046900-17A1 (Hussain, PI) and F32DK109592 to M.H.W., as well as G. Harold & Leila
822 Y. Mathers Foundation (Farber, PI), VA Merit Award BX004113-01A1 (Hussain, PI), AHA
823 Postdoctoral Fellowship 19POST34410063 to S.R., and the Wellcome Trust [098051 and
824 206194] to E. Busch-Nentwich.
825
826 **Declaration of Interests**
827 Authors declare no competing interests.
828
829 **Supplementary Files**
830
831 Supplementary File 1: Single nucleotide variants present in *c655* mutant embryos.
832
833

834 **METHODS & MATERIALS**
835
836 ***Zebrafish husbandry and maintenance***
837 Adult zebrafish were maintained at 27°C on a 14:10h light:dark cycle and fed once daily with
838 ~3.5% body weight Gemma Micro 500 (Skretting USA). Embryos were obtained by natural
839 spawning and were raised in embryo medium at 28.5°C and kept on a 14:10 h light:dark cycle.
840 All embryos used for experiments were obtained from pair-wise crosses and were staged
841 according to (Kimmel et al., 1995). Exogenous food was provided starting at 5.5 days post
842 fertilization (dpf) unless otherwise noted. Larvae were fed with GEMMA Micro 75 (Skretting) 3x
843 a day until 14 dpf, GEMMA Micro 150 3x a day + Artemia 1x daily from 15 dpf–42 dpf and then
844 GEMMA Micro 500 daily supplemented once a week with Artemia. Zebrafish sex is not
845 determined until the juvenile stage, so gender is not a variable in experiments with embryos and
846 larvae. Sex of adult fish included in analyses is noted in Figure legends. All zebrafish protocols
847 were approved by the Carnegie Institution Department of Embryology Animal Care and Use
848 Committee (Protocol #139).
849
850 *Stalactite* (*stl*) *mttp* mutant zebrafish in the *Tg(fli1:eGFP)*^{y1} background (Avraham-Davidi et al.,
851 2012; Lawson and Weinstein, 2002; Yaniv et al., 2006) were provided by Karina Yaniv
852 (Weizmann Institute of Science, Israel) and out-crossed to the AB wild-type strain. The *stl*
853 mutation was maintained in both the presence and absence of the *fli1:eGFP* transgene. The
854 *c655* phenotype was identified in the Farber laboratory in the background of a *kif7* mutant strain
855 that was obtained from Philip Ingham (Lee Kong Chian School of Medicine, Singapore). The
856 *c655 mttp* mutation was isolated from the *kif7* mutation by out-crossing to the AB wild-type
857 strain. The *c655* mutation was crossed into the *Tg(fli1:eGFP)*^{Y1} reporter line. Both *stl* and *c655*
858 *mttp* mutations were crossed into the *ApoBb.1-NanoLuc* LipoGlo reporter line (Thierer et al., In
859 Press).
860
861
862 ***Positional Cloning***
863 To map the location of the mutation responsible for the c655 phenotype, 23 embryos with
864 normal yolks and 23 embryos with opaque yolks (3 dpf) were processed for RNA-seq (White et
865 al., 2017). RNA was extracted from embryos by mechanical lysis in RLT buffer (Qiagen, 79216)
866 containing 1 µL of 14.3M beta-mercaptoethanol (Sigma, M6250). The lysate was combined with
867 1.8 volumes of Agencourt RNAClean XP (Beckman Coulter, A63987) beads and allowed to bind

868 for 10 minutes. The plate was applied to a plate magnet (Invitrogen) until the solution cleared
869 and the supernatant was removed without disturbing the beads. This was followed by washing
870 the beads three times with 70% ethanol. After the last wash, the pellet was allowed to air dry for
871 10 mins and then resuspended in 50 µl of RNAse-free water. RNA was eluted from the beads
872 by applying the plate to the magnetic rack. RNA was quantified using the Quant-iT 610 RNA
873 assay (Invitrogen, Q33140). Total RNA from individual embryos was DNase treated for 20 mins
874 at 37°C followed by addition of 1 µL 0.5M EDTA and inactivation at 75°C for 10 mins to remove
875 residual DNA. RNA was then cleaned using 2 volumes of Agencourt RNAClean XP (Beckman
876 Coulter, A63987) beads under the standard protocol. Strand-specific RNA-seq libraries
877 containing unique index sequences in the adapter were generated simultaneously following the
878 dUTP method using 700 ng total RNA and ERCC spike mix 2 (Ambion, 4456740). Libraries
879 were pooled and sequenced on Illumina HiSeq 2500 in 75bp paired-end mode. Sequence data
880 were deposited in European Nucleotide Archive under accession ERP023267. FASTQ files
881 were aligned to the GRCz10 reference genome using TopHat2 (Kim et al., 2013) (v2.0.13,
882 options: --library-type fr-firststrand). Ensembl 88 gene models were supplied to TopHat2 to aid
883 transcriptome mapping. MMAPPR (Hill et al., 2013) was used to determine the location of the
884 causal mutation. Variants were called from the pooled data using the GATK HaplotypeCaller
885 (Van der Auwera et al., 2013). Variants inside the regions output by MMAPPR were selected
886 and filtered for ones where the mutant sample was called as being homozygous alternate and
887 the siblings were heterozygous. The consequences of these variants on annotated genes was
888 calculated using the Ensembl Variant Effect Predictor (McLaren et al., 2016) and SIFT (Sim et
889 al., 2012). Variants with the following consequences were selected as candidates for the causal
890 mutation: stop_gained, splice_donor_variant, splice_acceptor_variant, transcript_ablation,
891 frameshift_variant, stop_lost, initiator_codon_variant, missense_variant, inframe_insertion,
892 inframe_deletion, transcript_amplification, splice_region_variant,
893 incomplete_terminal_codon_variant.
894
895 ***DNA Extraction and Genotyping***
896 Genomic DNA was extracted from embryos or adult fin clips using a modified version of the
897 HotSHOT DNA extraction protocol (Meeker et al., 2007). Embryos/tissues were heated to 95°C
898 for 18 minutes in 100 µL of 50 mM NaOH. The solution was cooled to 25°C and neutralized
899 with 10 µL of 1M Tris-HCl pH 8.0. Genotyping primers for the stalactite allele were designed

900 using the dCAPS Finder 2.0 program (Neff et al., 2002) and synthesized by Eurofins Genomics.
901 The *stalactite* locus was amplified using the forward primer 5'-GTC TGA GGT TCA GAT GTA
902 CCT GTT AGG AC-3' and reverse primer 5'-CTC TGC TGT GAT GAG CGC AGG-3' (0.5 μM
903 primer, T_a = 60°C, extension time 30"). The forward primer introduces an AvaII restriction site
904 into the mutant amplicon, such that following digestion (5 units of AvaII (New England BioLabs,
905 R0153) at 37°C, 4 h) the WT band is 157bp, homozygous mutants have bands at 129 bp and 28
906 bp, and heterozygotes have all three bands. The *c655* locus was amplified using the forward
907 primer 5'-AGAGACGGTGTCCAAGCAGG-3' and reverse primer 5'-GCTCAAAGACTTTCTTGC-
908 3' (0.25 μM primer, Ta = 50°C, extension time 30"). The *c655* mutation introduces a BsrI
909 restriction site into the amplicon, such that following digestion (3 units of BsrI (New England
910 BioLabs, R0527) in NEB Buffer 3.1 (B7203), 65°C, 3.5 h) the WT band is 137 bp, homozygous
911 mutants have bands at 76 bp and 61 bp, and heterozygotes have all three bands. For the
912 *ApoBb.1-NanoLuc* genotyping protocol, see (Thierer et al., In Press).
913
914 ***Generation of mttp-FLAG and ApoB48 plasmids***
915 The wild-type zebrafish *mttp* coding sequence with a FLAG-tag prior to the termination codon at
916 the C-terminus was generated by custom gene synthesis and cloned into the pcDNA3.1+ vector
917 (*mttp*-FLAG)(Gene Universal Inc., Newark, DE). The *stl* and *c655* mutations were subsequently
918 introduced to this plasmid by site-directed mutagenesis (Gene Universal Inc.) to generate
919 $mttp^{stl}$-FLAG and $mttp^{c655}$-FLAG plasmids. The human pcDNA3.1-*MTTP*-FLAG plasmid was
920 synthesized as described previously (Rava et al., 2006; Sellers et al., 2003). The human
921 equivalent of the *c655* mutation (G865V) was introduced into this plasmid using the Q5 Site-
922 directed mutagenesis kit (New England Biolabs), with the following primer pair: Forward 5'-
923 CGTATTAGCAgtaTGTGAATTCC-3', Reverse 5- CTTTCTTTTCTTTTCTGAGAG-3'. The
924 human ApoB48 sequence (Hussain et al., 1995) was cloned into the pcDNA3 under control of
925 the CMV promoter.
926
927 ***Rescue of c655 opaque yolk phenotype***
928 $mttp^{c655/c655}$ embryos were injected at the 1-cell stage with 20pg of zebrafish *mttp*-FLAG plasmid
929 and 20 pg of CMV:*eGFP-CAAX* (synthesized using the Tol2kit Gateway cloning system using
930 the p5E-CMV/SP6, pME-*eGFP-CAAX*, and p3E-polyA entry clones (Kwan et al., 2007)) as a
931 marker of successful injections. Embryos were raised to 3 dpf and screened for eGFP

932 expression in the yolk sac. Images of eGFP+ control and experimental embryos were blinded
933 and scored for yolk opacity by another member of the lab.
934
935 ***Ectopic sprout analysis***
936 *mttp*$^{stl/stl}$, *mttp*$^{c655/c655}$ and WT zebrafish in the *Tg(fli1:eGFP)*^{y1} background were imaged at 3 dpf
937 with a Zeiss Axiozoom V16 microscope equipped with a Zeiss PlanNeoFluar Z 1x/0.25 FWD
938 56mm objective, AxioCam MRm camera, and Zen 2.5 software. The length of ectopic
939 angiogenic segments that extend from the subintestinal vessels were analyzed in Fiji
940 (Schindelin et al., 2012)(ImageJ V2.0.0, National Institutes of Health, USA) as described by
941 (Avraham-Davidi et al., 2012).
942
943 ***Transmission Electron Microscopy***
944 Wild-type, *mttp*$^{stl/stl}$, *mttp*$^{c655/c655}$, and *mttp*$^{stl/c655}$ mutant zebrafish embryos were fixed at 4 dpf in a
945 3% glutaraldehyde, 1% formaldehyde, 0.1 M cacodylate solution for 1-3 h. Embryos were
946 trimmed and swim bladders were deflated before embedding in 2% low melt agarose and
947 processed as described in (Zeituni et al., 2016). Post-fixation was performed for 1 h with 1%
948 osmium tetroxide + 1.25% potassium ferricyanide in cacodylate solution. Following 2 x 10 min
949 washes with water, samples were incubated with 0.05M maleate pH 6.5 for 10 min. Samples
950 were stained en bloc with 0.5% uranyl acetate in maleate for 4°C overnight. Following 2 x 15
951 min washes with water, samples were dehydrated through graded EtOH dilution (35%, 2 x 15
952 min; 50%, 15 min; 75%, 15 min; 95%, 15 min; 100% 4 x 15 min). Samples were washed with
953 propylene oxide 4 x 15 min before incubation with 1:1 propylene oxide/resin (Epon 812 epoxy,
954 Ladd Research Industries, Williston, VT) for 1 h and evaporated overnight. This was followed
955 by 2 x 1 h washes in 100% resin and a final embedding in 100% resin at 55°C overnight
956 followed by 70°C for three days. Sections were made on a Reichert Ultracut-S (Leica
957 Microsystems), mounted on naked 200 thin mesh grids, and stained with lead citrate. Images
958 were obtained with a Phillips Technai-12 electron microscope (FEI, Hillsboro, OR) and 794
959 Gatan multiscan CCD camera (Gatan, Pleasanton, CA) using Digital Micrograph software. Lipid
960 droplet number and area was quantified with Fiji.
961
962
963

964 ***Growth Time-Course***

965 Unsorted embryos from pair-wise in-crosses of *stalactite* or *c655* heterozygous fish and pair-
966 wise crosses of *mttp*$^{stl/+}$ x *mttp*$^{c655/+}$ were raised and were analyzed for standard length at 1, 3, 6,
967 9, 12, and 24 weeks post fertilization. At 1 week, fish were imaged using a Nikon SMZ1500
968 microscope with HR Plan Apo 1x WD 54 objective, Infinity 3 Lumenera camera and Infinity
969 Analyze 6.5 software. Standard length (Parichy et al., 2009) was measured using Fiji (NIH).
970 Starting at three weeks, standard length was measured with a ruler. Mass of the fish was also
971 measured starting at 6 weeks. At 1 and 3 weeks, gDNA was obtained from whole fish for
972 genotyping. At later time-points, genotyping was performed on fin clips. Images of fish at 12
973 weeks post fertilization were taken with a Canon T6 camera with a Canon EF 100mm Macro
974 Lens.

975

976 ***Tissue Histology***

977 Adult zebrafish (7.5 mo; 2 males, 1 female per genotype) were placed individually into mating
978 tanks and fasted overnight (~24 h). Fish were euthanized by submersion in ice-water. A piece
979 of the anterior intestine and the liver were dissected from each animal and fixed in neutral-
980 buffered formalin (Sigma, F8775) at 4°C for 48 h. Sectioning and hematoxylin & eosin staining
981 was performed by the Johns Hopkins University Oncology Tissue Services. Slides were imaged
982 with a Nikon E800 microscope with 60×/1.4 oil Plan Apo Nikon objective and Canon EOS T3
983 camera using EOS Utility image acquisition software.

984

985 ***Tissue Lipid Extractions, HPLC & Analysis***

986 Adult zebrafish (1 yr; 2–3 males, 2–3 females per genotype) were fasted overnight (~24 h) and
987 euthanized by submersion in ice-water. Similar size pieces of the anterior intestine and the liver
988 were dissected from each animal and frozen on dry ice. Tissues were sonicated in 550 μL of
989 homogenization buffer (20 mM Tris-HCl, 1 mM EDTA), and the protein concentration of each
990 sample was measured using the BCA protein assay kit (Pierce, 23225). Lipids were extracted
991 from the remaining sample volume by a modified Bligh-Dyer procedure (Carten et al., 2011),
992 dried under vacuum, and re-suspended in 50 μL of HPLC-grade isopropanol as the HPLC
993 injection solvent. Injection volumes were optimized for each sample to produce peak shapes
994 appropriate for quantitation (1–25 μL). The lipid components of each sample were separated
995 and detected by an HPLC-CAD system using a LPG-3400RS quaternary pump, WPS-3000TRS

996 autosampler (maintained at 20°C), TCC-3000RS column oven (maintained at 40°C), Accucore
997 C18 column (150 x 3.0 mm, 2.6 μm particle size), FLD-3100 fluorescence detector (8 mL flow
998 cell maintained at 45°C), and a Dionex Corona Veo charged aerosol detector (Thermo Fisher
999 Scientific). Lipids were separated over an 80 min time range in a multi-step mobile phase
1000 gradient as described in (Quinlivan et al., 2017). The lipid class and area of each analyte peak
1001 were determined using Chromeleon 7.2 (Thermo-Fisher Scientific) for chromatogram
1002 visualization and manual integration as described in (Otis et al., 2017). For quantitative
1003 comparisons between samples, each lipid peak area was normalized to the protein
1004 concentration of the homogenized tissue.
1005
1006 ***LipoGlo Assays***
1007 All LipoGlo assays were performed with fish carrying a single copy of the LipoGlo
1008 *(apoBb.1$^{NLuc/+}$)* reporter. For detailed LipoGlo methods see (Thierer et al., In Press); Nano-Glo
1009 reporter system reagents are all from Promega Corp., (N1110; (Hall et al., 2012)). For
1010 quantitative assays and B-lp size analysis, individual embryos were dispensed into 96-well
1011 plates (USAScientific, #1402-9589) and homogenized in 100 μL of B-lp stabilization buffer (40
1012 mM EGTA, pH 8.0, 20% sucrose + cOmplete mini, EDTA-free protease inhibitor (Sigma,
1013 11836170001)) by sonication in a microplate-horn sonicator (Qsonica Q700 sonicator with a
1014 Misonix CL-334 microplate horn assembly). Homogenate was stored on ice for immediate use
1015 or frozen at -20°C for later use. ApoB-Nanoluc levels were quantified by mixing 40 μL of
1016 embryo homogenate with an equal volume of diluted Nanoluc buffer (1:3 NanoGlo buffer:PBS +
1017 0.5% NanoLuc substrate (furimazine)) in a 96-well opaque white OptiPlate (Perkin-Elmer,
1018 6005290), and the plate was read within 2 minutes of buffer addition using a SpectraMax M5
1019 plate reader (Molecular Devices) set to top-read chemiluminescent detection with a 500 ms
1020 integration time. To quantify the size distribution of B-lps, 12 μL of homogenate was combined
1021 with 3 mL of 5x loading dye (40% sucrose, 0.25% bromophenol blue, in Tris/Borate/EDTA (TBE)
1022 buffer), and 12.5 mL of the resulting solution (10% larval homogenate) was loaded per well on a
1023 3% native polyacrylamide gel. Each gel included a migration standard of Di-I-labeled human
1024 LDL (L3482, ThermoFisher Scientific). Gels were run at 50V for 30 min, followed by 125V for 2
1025 h. Following application of 1 mL of TBE supplemented with 2 μL of Nano-Glo substrate to the
1026 surface of the gel and incubating for 5 min, gels were imaged with an Odyssey Fc (LI-COR
1027 Biosciences) gel imaging system. Images were obtained in the chemiluminescent channel (2

1028 min exposure) and then the 600 nm channel (30 sec) for Nanoluc detection and Di-I LDL
1029 standard detection, respectively. Each lane on the gel was converted to a plot profile in Fiji and
1030 divided into LDL, IDL, VLDL and Zero Mobility bins based on migration relative to the Di-I LDL
1031 standard. Pixel intensity from the plot profile was summed within each bin for comparison
1032 between genotypes. To determine the localization of B-lps in the whole fish, intact embryos or
1033 larvae were anesthetized and fixed in 4% paraformaldehyde for 3 h at room temperature.
1034 Following rinses in PBS + 0.1% tween-20 (3 x 15 min), embryos were mounted in 1% low-melt
1035 agarose (BP160-100, Fisher Scientific) in TBE supplemented with 1% Nano-Glo substrate.
1036 Chemiluminescent images (10 and 30 sec exposures with no illumination) and a brightfield
1037 image were taken with a Zeiss Axiozoom V16 microscope equipped with a Zeiss Plan NeoFluar
1038 Z 1x/0.25 FWD 56mm objective, AxioCam MRm camera, and Zen 2.5 software, using 2x2
1039 binning and 2x gain. Images were quantified using Fiji; regions of interest (ROI) were drawn on
1040 the brightfield image (viscera, trunk, and head), and these ROIs were used to quantify the
1041 NanoLuc intensity on the 30sec exposure chemiluminescent images. ROIs of the same shape
1042 were used to calculate the background signal, which was subtracted from the intensity value for
1043 each ROI.
1044
1045 ***Oil Red O staining***
1046 15-dpf larval zebrafish were fixed with 4% paraformaldehyde in PBS for 3 h at room
1047 temperature and then overnight at 4°C. Fish were rinsed in 60% 2-propanol for 10 minutes,
1048 rocking and then put into 0.3% Oil Red O (Sigma-Aldrich, #O0625) to rock overnight at room
1049 temperature. Fish were rinsed 3 times with 60% 2-propanol for 15 minutes. Washed fish were
1050 equilibrated step-wise into glycerol and imaged with incident light using a Nikon SMZ1500
1051 microscope with HR Plan Apo 1x WD 54 objective, Infinity 3 Lumenera camera, and Infinity
1052 Analyze 6.5 software.
1053
1054 ***ApoB secretion assays***
1055 Monkey kidney COS-7 cells which do not express MTTP or ApoB were plated in 10 cm^2 cell
1056 culture dishes at a density of 9 x 10^5 cells per plate and grown in Dulbecco's modified Eagle's
1057 medium (DMEM) containing 10% fetal bovine serum, L-glutamine, and antibiotics at 37°C.
1058 COS-7 cells were transfected with 5 µg of plasmid expressing human ApoB48 cDNA under the
1059 control of CMV promoter using endofectin (Genecopoeia, EF014) according to the

1060 manufacturer's protocol. After 24 hours, cells from each dish were harvested, equally distributed
1061 in 6-well plates, and reverse transfected with 3µg of either pcDNA3, pcDNA3-*mttp*-FLAG,
1062 pcDNA3-*mttp*^{stl}-FLAG, pcDNA3-*mttp*^{c655}-FLAG, pcDNA3-*MTTP*-FLAG, or pcDNA3-
1063 *MTTP*(G865V)-FLAG plasmids. After 32 h cells were incubated overnight with 1 mL of DMEM
1064 containing 10%FBS. The overnight conditioned media were collected to measure ApoB by
1065 ELISA (Bakillah et al., 1997; Hussain et al., 1995). Cells were scraped in PBS and a small
1066 aliquot was used to measure total protein using a Coomassie protein assay (Thermo Scientific,
1067 #1856209). Cells were lysed in cell extract buffer (100 mM Tris, pH 7.4, 150 mM NaCl, 1 mM
1068 EGTA, 1mM EDTA, 1% Triton X-100, 0.5% sodium deoxycholate). Lysates were rotated for 1 h
1069 at 4 ºC to solubilize the membranes and centrifuged at 16,000g for 30 mins. ApoB was
1070 measured in the supernatant via ELISA. Briefly, high binding 96 well plates (Corning, #3366)
1071 were incubated with capture antibody anti-LDL (apoB), clone 1D1(MyBiosource, #MBS465020,
1072 1:1000 dilution) overnight at room temperature. The plate was washed 3x with PBS-T (PBS +
1073 0.05% Tween-20) and blocked with 3% BSA (Boston Bio Products, #P753) for 1 h and washed
1074 3x with PBS-T, before incubating with 100 µL of standards and experimental samples for 3 h.
1075 The plate was washed 3x with PBST and incubated with 100 µL of human ApoB antibody
1076 (Academy Bio-Medical Company, Inc., #20S-G2, 1:1000 dilution) for 1 h. After washing the
1077 plate 3x with PBS-T, 100 µL of alkaline phosphatase labeled anti-goat IgG (Southern Biotech,
1078 #6300-04, 1:3000 dilution) was added to each well and incubated for 1 h. The plate was
1079 washed 3x with Diethanolamine buffer, pH 9.5 and 100 µl of PNPP (Thermo Scientific, #34045,
1080 1 mg/mL) was added to each well before reading the plate at 405nm in a PerkinElmer Victor^3
1081 1420 multilabel counter. Data for zebrafish and human plasmids were obtained in the same
1082 experiments, but are graphed separately in figures 5A,B & 6B,C; the pcDNA control data is
1083 displayed in both sets of graphs.
1084
1085 ***Immunofluorescence***
1086 COS-7 cells were plated at a density of 50,000 cells on coverslips in 12-well dishes and
1087 transfected with 2 µg of plasmids expressing either zebrafish or human MTTP-FLAG plasmids.
1088 After 48 h, cells were fixed in paraformaldehyde and blocked with PBS supplemented with 1 mM
1089 $MgCl_2$, 0.5 mM $CaCl_2$, 3% BSA, 0.1% Triton X-100 and 1% horse serum. Cells were incubated
1090 with anti-FLAG M2 monoclonal antibody (Sigma # F3165, 1:250 dilution) and anti-calnexin
1091 antibody (Santa Cruz Biotechnology, # sc-11397, 1:250 dilution) overnight. Cells were washed

1092 three times with PBS and incubated with goat anti-mouse Alexa Fluor-594 (Invitrogen, #A11005,
1093 1:500 dilution) and donkey anti-rabbit Alexa-Fluor-488 (Invitrogen, # A21206, 1:500 dilution) for
1094 1 h. The cells were washed and mounted with Vectashield mounting medium (Vector
1095 Laboratories, #H-1000). Images were taken on a Leica SP5II confocal microscope with a
1096 63x1.4 HCX PL Apo oil immersion lens.
1097
1098 ***Immunoprecipitation and Western blotting***
1099 Transfected COS-7 cells were washed three times with ice cold PBS and scraped in buffer K (1
1100 mM Tris-HCl, 1 mM EGTA and 1 mM $MgCl_2$, pH 7.6) containing protease inhibitor cocktail
1101 (Sigma, # P2714). Cells were mechanically lysed by passing them 10 times with $30_{1/2}$-gauge
1102 needle and small fractions were used to measure the total protein using a Coomassie protein
1103 assay (Thermo Fisher Scientific, #1856209). Cell lysate was incubated with Anti-FLAG M2
1104 antibody for 1 h and immunoprecipitated (IP) using (protein A/G) agarose beads (Santa-Cruz
1105 Biotechnology, # SC2003). The supernatants were used to detect actin via Western blotting
1106 and served as loading controls. Both the supernatant and immunoprecipitated fractions were
1107 subjected to electrophoresis on an 8% SDS-PAGE gel. The weight separated proteins were
1108 transferred to nitrocellulose membranes and probed with either anti-FLAG M2 (1:1000) or anti-
1109 actin (Thermo Fisher Scientific, #PA1-183, (1:3000)) prepared in 2% BSA in TBS. The blots
1110 were washed and probed with HRP-conjugated corresponding secondary antibodies (goat anti-
1111 rabbit, Cell Signaling Technology, #7074, 1:5000 or goat anti-mouse, Thermo Fisher Scientific,
1112 #62-6520, 1:5000). The blots were developed in ChemiDoc™-Touch Imaging system from Bio
1113 Rad.
1114
1115 ***Triglyceride transfer assay***
1116 Following transfection with plasmids as described above, cell lysate (35 µg) prepared in buffer K
1117 containing protease inhibitor cocktail was incubated with donor vesicles containing NBD-labeled
1118 triolein (Setareh Biotech, LLC, #6285) and acceptor vesicles. Fluorescence was measured at
1119 different time intervals (5, 10, 15, 30, 45 and 60 mins). Percent triglyceride transfer was
1120 calculated after subtracting the blank and dividing it by the total fluorescence reading obtained
1121 by disrupting vesicles with isopropanol, as described previously (Athar et al., 2004; Rava et al.,
1122 2005). Where noted, assays also included the MTTP inhibitor Lomitapide (Aegerion
1123 Pharmaceuticals, #AEGR-733) at a concentration of 1 µM.
1124

1125 ***Phospholipid transfer assay***

1126 COS-7 cells were transfected with 9 µg of either zebrafish *mttp*-FLAG or human MTTP-FLAG
1127 plasmids in 10 cm^2 cell culture dishes. After 48 h, cell lysates were prepared in buffer K
1128 containing protease inhibitor cocktail (Sigma, #P2714). The cell lysates were centrifuged at
1129 12,000g for 10 mins at 4ºC. A small aliquot of cell lysate was used for measuring protein and
1130 kept for Western blotting to measure expression level. Equal concentrations of protein from
1131 each sample were incubated with 40 µL of M2 agarose beads (Sigma, #A2220) for 3 h at 4ºC.
1132 FLAG-tagged protein were eluted twice with 200 ng/µL FLAG peptide (Sigma, #F3290; 1 h at
1133 4ºC). PL transfer activity was assayed using NBD labeled Phosphoethanolamine,
1134 triethylammonium (Thermo Fisher Scientific, #N360). The purified FLAG-tagged proteins were
1135 incubated with donor vesicles containing NBD-Phosphoethanolamine and acceptor vesicles.
1136 The fluorescence was measured at different time intervals (1, 2, 3 and 4 h). The percentage
1137 transfer of phospholipid was calculated as the difference between the fluorescence reading at
1138 the 0 h time point and 3 h time point divided by the total fluorescence reading obtained by
1139 disrupting vesicles with isopropanol as described previously (Athar et al., 2004; Rava et al.,
1140 2005). Where noted, assays also included the MTTP inhibitor Lomitapide at a concentration of
1141 1 µM.
1142

1143 ***Modeling***

1144 Predicted models of human MTTP and zebrafish Mttp proteins were generated using Phyre2
1145 (Kelley et al., 2015) and the graphics were enhanced for clarity using Chimera (UCSF Chimera,
1146 developed by the Resource for Biocomputing, Visualization, and Informatics at the University of
1147 California, San Francisco, with support from NIH P41-GM103311 (Pettersen et al., 2004).
1148

1149 ***Statistical Analyses***

1150 Graphing and some statistics, including One-way, randomized block and Repeated Measures
1151 ANOVA with Bonferroni post-hoc tests, Kruskall-Wallis with Dunn's Multiple Comparison test
1152 and Chi-square tests were performed with GraphPad Prism (GraphPad Software). When
1153 sample sizes and variance between groups were significantly different, Robust ANOVA was
1154 performed using R to determine overall significance of noted datasets (Mair and Wilcox,
1155 2018)(https://cran.r-project.org/web/packages/WRS2/vignettes/WRS2.pdf),((Mangiafico, 2015),
1156 https://rcompanion.org/rcompanion/d_08a.html). When significant differences were present
1157 between genotypes, Games-Howell post-hoc tests were used to make pair-wise comparisons at

1158 each time point using SPSS Statistics (IBM), adjusting the significance level for multiple
1159 comparisons. Details of the statistical analyses can be found either in the figure legend or
1160 results sections. Sample sizes for each experiment are indicated in the figure legends for each
1161 experiment.
1162
1163 ***Additional Software***
1164 DNA, mRNA, and protein sequence alignments were performed with MacVector V15.5
1165 (MacVector, Inc.). Microsoft Word and Excel were used for manuscript preparation and data
1166 analysis, respectively, figures were assembled in Adobe Illustrator CS5 (Adobe Systems) and
1167 references were assembled with EndNote 8X.
1168
1169
1170
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