Population genomics and pathotypic evaluation of the bacterial leaf blight

pathogen of rice

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Abstract

Knowledge about population structure and virulence dynamics is critical to prevent outbreaks of bacterial diseases. *Xanthomonas oryzae* pv. *oryzae* (*Xoo*) causes bacterial blight disease, leading to substantial losses in rice production during the last century in Asia. For this concern, we used whole-genome sequences and large-scale virulence tests to explore the diversity and evolution of *Xoo* population during the past 30 years in the main rice-planting areas of China. Six separate lineages were revealed by phylogenomic analysis, two of which (CX-5 and CX-6) predominated in the population. Four and 5 sub-lineages from these two lineages respectively persisted in different areas for decades. Many recent sporadic outbreaks were caused by *Xoo* derived from the local sub-populations of these sub-lineages. The lineage and sub-lineage distribution of isolates strongly correlated to their geographical origin, which was found to be determined by the planting of the two major rice subspecies. Large-scale virulence tests indicated rapid dynamics of pathogenicity for *Xoo*. The *Xoo* isolates from lineages and sub-lineages with stricter geographical distribution showed less compatibility with resistance genes of rice than the wider distributed ones. Genetic background of *Xoo*, rice resistance genes and the planting environment of rice contributed to the rapid dynamics of virulence. This study provided a good model to understand the evolution and dynamics of plant pathogens in the context of interaction with their hosts which are influenced by both geographical conditions and farming practices.

Significance

Xanthomonas oryzae pv. *oryzae* (*Xoo*) which causes bacterial blight disease, was one

of the major serious threats to food security in East Asian countries during last century.

The introduction of several resistance genes into rice tremendously reduced the damage.

Through population genomics and large-scale pathotypic evaluation of more than 240

Xoo strains isolated in China during the past 30 years, we found that the genome

evolution and virulence dynamics of *Xoo* are extraordinary rapid, indicating its ability

to overcome the resistance conferred by the current resistance genes and the potential

for large-scale outbreaks in the future. It is therefore prudent to continue surveillance

of disease outbreaks caused by this pathogen, and to develop novel strategies for its

control.

Introduction

Population genomics is a powerful tool for understanding the formation and evolution of bacterial pathogens of human and some important domestic animals (1-6). It has also provided new strategies for the detection, prevention and control of important human pathogens (7, 8). Although plant pathogens are serious threat to food security worldwide, only a few studies have used genome-based methods to study their population structure, evolution and transmission in the context of interaction between pathogens and plant hosts (9-13).

Xanthomonas oryzae pv. *oryzae* (*Xoo*) causes bacterial blight (BB) disease of rice (*Oryza sativa*) and has been considered one of the top 10 plant bacterial pathogens based on scientific/economic importance (14). It is a notorious destructive pathogen which can cause a considerable reduction in rice production in both temperate and tropical regions, especially in Asia (15). On average, BB can lead to 20%~30% overall yield reduction with some severe cases even causing up to 50% loss (16). As the world's largest producer and consumer of rice, China has suffered serious BB outbreaks and huge food loss since the 1930s (17), with the most serious damage occurring between 1950s and 1980s (18). Breeding resistant cultivars has been proved to be the most effective method in controlling this pathogen. Since 1980s, BB-resistant rice varieties have been widely planted in China and have significantly reduced the loss caused by BB (19-21). However, sporadic outbreaks have occurred in different areas of China after the year of 2000 (22, 23). It is unknown whether the recent outbreaks relate to changes in the genetics and virulence of the past *Xoo* populations in China, allowing it

to overcome host defenses. Moreover, very little is known about the evolution, spread

and dynamics of this important plant pathogen involved in different outbreaks during

the past decades around the whole country in the context of rapid development of

modern agricultural science and technology.

The virulence differentiation among different strains of *Xoo* was studied by defining

different races or pathotypes. Strains of the same race share a common pathogenic

phenotype in a set of tested host cultivars. Rice lines carrying different resistance genes

(R genes) determine the race of strains of *Xoo*. Up to now, more than 100 races of *Xoo*

were identified (24-29). Although the pathogenic diversity of *Xoo* in China have been

studied previously (22, 30, 31), these studies classified *Xoo* isolates only based on their

pathogenic patterns on several rice lines, while the genetic basis for the pathogenicity

of these isolates is still unknown.

In this study, a comprehensive population genomic study was combined with a large-

scale determination of virulence of *Xoo* that have been isolated in the past 30 years in

China, to obtain a framework of genetic dynamics and virulence diversity over time

and space. Results from this work shed new light on the evolution of this important

plant pathogen.

Results

Population structure *Xoo* from China

To understand the population structure of *Xoo* in China, genomes of the 247 strains

were sequenced, including 237 isolated from China, 4 and 6 representative ones from

Japan and Philippines, respectively (Table S1). Reads were mapped to the genome of

PXO99[A], SNPs were called by Samtools. After the prophage region, repeat sequences

and recombinant regions were filtered; the core SNPs contained 5,271 variable sites.

Two methods were used to outline the population structure based on the core SNPs,

Maximum likelihood (ML) phylogenetic analysis and a tree-independent hierarchical

Bayesian clustering (BAPS).

All *Xoo* from China were clustered into 6 lineages (CX-1 to CX-6), with more than 70%

strains belonging to CX-5 and CX-6 (Figure 1). The three major rice production areas,

including South China, Yangtze Valley area and North China, were displayed in Figure

2a. We annotated the lineage-specific tree with the time and space information of the

isolated *Xoo* (Figure 1, 2b and 2c). CX-1 and CX-2 were most frequently represented

in South China at the year of 2003, of which most isolates were from Yunnan province.

The *Xoo* belonging to CX-3 were isolated from North China in 1984, 2003 and 2014,

with most of which from Northeast China in 2014. CX-5 and CX-6 were nationally

distributed during the past 30 years; while most members in CX-5 were dominant in

South China and Yangtze Valley, and CX-6 were more frequently from North China

and Yangtze Valley.

Phylogeography of CX-5 and CX-6

To investigate more details about evolution and dynamics of *Xoo* in China, we focused

on the two major lineages, CX-5 and CX-6. Phylogenetic analysis based on core SNP

alignment (Figures 3a and 3b) and pairwise SNP distance between isolates was

performed for both lineages (Figure 3c). Though no correlation between root-to-tip

branch lengths and the known years of isolation of the sequenced was observed for both

lineages, it's worth mentioning that some important points about the dynamics of these

population were illuminated, especially those for the recent outbreaks.

Four sub-lineages were identified in CX-5, CX-5.1 to CX-5.5 (Figure 3a). CX-5.1, CX-

5.2, and CX-5.5 were restricted in South China, while CX-5.3 and CX-5.4 can be found

in some places of both South China and Yangtze Valley. When time was considered, we

found that all these sub-lineages persisted at these places at least for decades. Then we

focused on the recent sporadic outbreaks being attributed to this lineage. Besides

clustered together on the tree, we considered epidemiologically and genomically linked

outbreak with an average SNP pairwise distance less than 5. In 2003, the outbreak in

Hunan province of Yangtze Valley was caused by CX-5.4 (HN-2003), while the

outbreak in Guangdong province of South China ascribed to two sub-population of CX-

5.5 (GD-2003-1 and GD-2003-2). One cluster of CX-5.3 caused outbreak in Guangxi

province of South China in 2014 (GX-2014).

CX-6 was divided into 5 sub-lineages, CX-6.1 to CX-6.5 (Figure 3b). Seven out of 10

isolates in CX-6.1 were from different places of South China. Actually, this sub-lineage

was the most diverse one in CX-6 with long branches. In CX-6.2, *Xoo* from North China

and Yangtze Valley were dominant on different branches, respectively. CX-6.3 and CX-

6.4 were most frequently found in Yangtze Valley, while *Xoo* from North China

predominated in CX-6.5. Similar as CX-5, all these sub-lineages have contributed to

BB on rice for a long time in China. These sub-lineages totally caused at less 4 local

outbreaks in 2003 and 2014. In 2003, the local outbreaks in two provinces of Yangtze

Valley (Henan, and Jiangsu) were caused by CX-6.2 (HeN-2003) and CX-6.4 (JS-2003),

respectively, while the one in North China (Jilin province) was ascribed to CX-6.5 (JL-

2003). Outbreak in Jiangsu province (Yangtze Valley) in 2014 was caused by one sub-

population of CX-6.4 (JS-2014).

Chinese *Xoo* diversity and dynamics in the context of rice host

There are two major domesticated rice subspecies *Oryza sativa japonica* and *indica*

existing in different areas of China; and most of rice varieties were derived from these

two subspecies (32, 33). Rice planted in high-altitude areas of Yunnan province from

South China and North China belong to subspecies *japonica* (http://www.ricedata.cn).

Xoo isolated from these areas were respectively clustered into 3 lineages, CX-1, CX-2,

CX-3 and one sub-lineage of CX-6 (CX-6.5) (Figure 1 and Figure 3b). Lineage CX-5

mainly contained *Xoo* isolated from rice in South China where subspecies *indica* is

planted (Figure 3a). CX-6 also contained *Xoo* isolated from rice of *japonica* from

Yangtze Valley (CX-6.4), *indica* from Yangtze Valley (CX-6.3) and some *indica* from

South China (CX-6.1), indicating the widest adaptation to different rice varieties of *Xoo*

in this lineage (Figure 3b). In CX-6.2, old *Xoo* were dominant on indica, while recent

isolated ones were almost from *japonica*, suggesting a putative host switch in this sub-

lineage.

Taken together, the distribution of Chinese *Xoo* lineages appeared to be impaired by

both biogeography and rapid dynamics over isolated time, which may be mainly due to

the distribution and dynamics of the two major subspecies of rice, *japonica* and *indica*.

Diversity of *Xoo* from different counties of Asia

To place *Xoo* from China into a global context, phylogenomic analysis was performed on the core genome SNPs of 109 *Xoo* genomes available in Genbank (including 100 from India, 8 from Philippines and 1 from Japan), and the 247 sequenced ones in this study. Phylogenetic analysis revealed a similar topological structure of Asian *Xoo* to that of Chinese *Xoo*, with some small differences in some branches (Figure S1). Eleven genetic lineages were identified for the *Xoo* population from Asia. *Xoo* from India and Philippines were respectively assigned into PX-A to PX-C and IX-I to IX-V according to previous studies (34, 35). Eight lineages displayed strict geographic distribution features, most of them being specific to one or at least a limited amount of rice-planting areas. Besides the three Chinese lineages, CX-1, CX-2 and CX-3, PX-B and PX-C were mainly represented in Philippines, and IX-I to IX-III were from India. CX-4 contained old *Xoo* from China and some recently isolated ones from India belonging to IX-V. For CX-5 and CX-6, besides both represented the major *Xoo* from China, they also contained a few *Xoo* from Japan, India and Philippines, with PX-A of Philippines being nested within the former one and IX-IV from India being encompassed among the members of the last one, indicating frequently transmission of these 2 lineages not only among different places of China but also among different Asian countries.

Rapid virulence dynamics of *Xoo* strains isolated in China during the past 30 years

To determine the pathogenic diversity of *Xoo* strains, virulence assays were conducted on six rice lines. Based on the interactions between *Xoo* strains and rice lines, most of the tested *Xoo* were classified into 9 pathogenic races (Table S1and Figure 4a). The top two races with most members, R5 and R8, contain 26% and 18% of the isolates,

respectively.

The three major rice-planting regions had different race compositions (Figure 4b).

Yangtze River Valley mainly contained *Xoo* from races R5 and R8, with the respective

percentage of the total population being 50% and 30%, respectively. *Xoo* isolates from

North China mostly belongs to R7 and R5, together accounts for about two third of the

total population. Five of the nine *Xoo* races were well represented in the *Xoo* population

from South China, with each of them account for more than 10% of the total population.

The virulence dynamics of *Xoo* during the last 30 years was hypothesized by analyzing

race distribution in China at different period (Figure 4c). Strains isolated before the year

of 2000 were mainly of the races of R5, R4, R8 and R1, while most *Xoo* isolated in

2003–2004 were R8, R7, R5 and R6; and about 45% of the isolates in 2014 were R5.

We also found different lineages have different race compositions, indicating genetic

background playing important roles in the interaction between *Xoo* and rice hosts

(Figure 4d). Compared to other lineages, the CX-6 *Xoo* isolates fall mostly within the

races of R5, R8 and R4, while have reduced representation in R1, R2 and R6. Moreover,

the rapid dynamics of pathogenic features can be inferred by focusing on the races of

Xoo isolated from the same places at the same time and belonging to the same lineage

(Figure 1). For example, most *Xoo* from Yunnan, one province of South China, in 2003

were clustered in CX-1 and CX-2. The 19 isolates in CX-1 represented 8 of the 9 races

described above, and the 14 strains in CX-2 belonged to 6 races. Even *Xoo* isolates from

the same outbreak were allocated into different races, with 16 *Xoo* in JL-2003 belonging

to 3 different races and 10 isolates in GD-2003-2 representing 2 races.

Resistance gene of rice contributes to the interaction between *Xoo* and rice

As each of the near-isogenic rice lines used in this study contains one known resistance (R) gene, we studied the interaction between *Xoo* and rice host by focusing on the capacity of *Xoo* to overcome the resistances conferred by these genes (Figure 1, Tables 1 and 2). When the time of isolation was considered, we found that *Xoo* from different times have different capacity to overcome different R genes (Table 1). R gene *xa5* showed the most effective resistance against *Xoo* all the time. At different periods, more than 90% *Xoo* could not infect the rice line IRBB5, which contained gene *xa5*. The resistance mediated by R gene *Xa3* was effective during the period of 1990 to 2004. However, rice containing this R gene only showed resistance against 5% of *Xoo* in 2014. Rice cultivars carrying R gene *Xa4* are resistant to more than 60% of *Xoo* isolated from 1970s to 2000 and 2014. However, they are only resistant to about 30% of the isolates recovered in 2003 and 2004.

To determine whether the *Xoo* genetic background contributes to the interaction between *Xoo* and rice lines, we studied the resistance capacity of R genes against *Xoo* from different lineages (Table 2). We found that resistance provided by R genes against *Xoo* are indeed depended on the genetic background of *Xoo*. R genes *Xa2* and *Xa14* effectively confer resistance against *Xoo* from Lineages CX-1, CX-2 and CX-3, preventing infection by more than 70% of *Xoo* strains from these lineages. Similarly, R gene *Xa3* and *Xa4* showed potential resistance against *Xoo* from CX-1 and CX-2, and CX-2 and CX-3, respectively. No R gene except *xa5* conferred resistance to more than 50% of *Xoo* strains from CX-5 and CX-6, which is consistent with the situation that

CX-5 and CX-6 had much more extensive distribution across different areas of China

during the past decades, comparing to the other Chinese *Xoo* lineages. Among the sub-

lineages of CX-5 and CX-6, *Xa2* was only effective against *Xoo* of CX-6.5, with

effectiveness of 90%, while *Xa4* could confer the resistance against more than 60% *Xoo*

from these two lineages except for CX-5.1 and CX-6.5; *Xa14* showed potential medium

level of resistance against *Xoo* from CX-6.1 and CX-6.5 (Table S2).

Discussion

RFLP (Restriction Fragment Length Polymorphism) was one of the successfully used

molecular techniques for diversity studies of plant pathogens before genomic methods

emerging (36-38). It was frequently used for genetic diversity of *Xoo* in several

important rice-planting countries (28, 39-44). These studies had provided some new

insights into the evolution and dynamics of *Xoo* population; however, many crucial

details were overlooked due to the intrinsic low sensitivity and stability of the typing

method. As the *Xoo* population structure were studied overly dependent on the marker

genes chosen, the definite genetic relationships among *Xoo* isolates from the same and

different outbreaks, the particular evolutionary history of the different clusters and

different isolates, and the sophisticated connection between genetic diversity and

virulence dynamics cannot be will well elaborated through these probe-dependent

methods. Genome-based population genetics methods have been proved more powerful

to study the diversity, evolution and dynamics of many pathogens of animals and plants

(1-6, 9-13). Previous report focusing on *Xoo* from India through population genomics

defined 5 lineages, with each of them showing a restricted-region distribution (35).

Here, we focused on genomic epidemiology of *Xoo* and provided some novel insights

into the diversity, evolution and dynamics of this notorious plant pathogen during the

past 30 year in the main rice-planting areas of China. We also found that the diversity

and dynamics of virulence features of this pathogen population are affected by the

population structure, genetic background, environmental factors and human activities.

These findings will benefit for development of new strategies and methods for

surveillance and controlling this destructive rice pathogen.

Some *Xoo* clusters found by traditional molecular typing methods from several Asian

countries showed strict geographical distribution, but others were widely distributed in

these countries (28, 39, 42). In this study, based on population genomics, we found that

there is a general correspondence between the areas of *Xoo* isolated and most Asian

lineages and some sub-lineages in CX-5 and CX-6 (Figures 1, 3 and S2). Moreover,

Asian *Xoo* was reported to be very different from those of Africa and North America

regions (45, 46). This suggests that the geographical factors as affected by climate, soil

types, or the choice of different rice cultivars and varieties may influence on the

diversity of *Xoo*. Information of the *Xoo* host subspecies in China indicates that variety

of rice was one of the major factors forces on the formation and dynamics of Chinese

Xoo population. Rice varieties planting in different counties and different areas of a

country are determined by geographical environment, climate factors and local

agricultural policy.

Though resistance breeding of rice had effectively prevented the unprecedented

outbreak of BB since 1990s, sporadic outbreaks occurred in different areas of China

after the year 2000 (22, 23). Genomic epidemiology analysis indicates that the recent

outbreaks in different places were contributed by different lineages or sub-lineages,

even some outbreaks from the same places caused by different lineages. There was no

special lineage or sub-lineage spreading nationwide, partially because of the great

diversity of rice varieties planting in China (http://www.ricedata.cn). All *Xoo* strains

causing the recent epidemic outbreaks except those in CX-1 were derived from the ones

leading to heavy losses in the history. On the other hand, only the minor lineage CX-4

was not represented by the recent outbreaks, so the diversity of the *Xoo* population was

not reduced due to wide planting of BB-resistant rice varieties. For the current practice

of rice production, the major BB threat in the three rice-planting areas of China is still

mainly from CX-5 and CX-6 as they were present in history, which call for different

detection and prevention strategies for different areas and rice varieties.

The virulence diversity of *Xoo* in China against rice was well investigated during the

past 30 years, however, the relationships between population genetic structure and

pathotype of *Xoo* were not explored well (22, 30, 31, 44, 47). All the tested *Xoo* strains

in this study were classified into nine major pathogenic races (Figure 4a). Races of *Xoo*

from South China were more diverse than those from North China and Yangtze Valley

areas. In line with this, phylogenetic analysis revealed that *Xoo* from South China are

more diverse compared to other places of China (Figure 1). Some rice growing regions

in South China, particularly in the Yunnan province, have three harvests per year and

use different rice cultivars for each harvest (48, 49). The greater genetic and virulence

diversity of *Xoo* thus could be caused by a greater diversity of rice cultivars in use. Our

result also showed that there were diverse races in the same lineage, for example, 85%

Xoo from lineage CX-6 could be assigned to 8 races (Figure 4d), indicating rapid

virulence dynamics in the same lineage. Moreover, the isolates from the same local

outbreak even belonged to different races. Although *Xoo* strains isolated in China were

consistently assigned to one of the 6 Chinese lineages, the dominating population of

Xoo races in specific areas displayed rapid dynamics and high level of diversity during

the past 30 years (Tables 1 and 2). Rapid virulence dynamics with relative stability of

the standing genetic variability of *Xoo* population indicates fast adaptation to host

during interaction for this plant pathogen.

Earlier isolates were more susceptible to resistances conferred by rice resistance genes

than strains isolated in later years after the resistance genes had been introduced to

control BB (39, 43). In China, since the 1980s, *indica* hybrid rice (especially Shanyou

63) had been planted on a large scale (50). Later, bacterial blight resistant rice varieties,

mainly the *india* rice carrying *Xa4* and *japonica* rice carrying *Xa3*, were grown on a

large scale for a long time (51). Soon after these rice varieties widely used for rice

production, the effectiveness of these R genes conferring resistance against infection

causing by *Xoo* was significantly compromised (Table 1). Actually, the resistance

abilities of these two R genes were also overcome frequently by *Xoo* population in other

Asian countries (40, 42, 43). Long-term cultivation of rice varieties carrying a single

resistance gene can be overcome by *Xoo*, and breeding rice varieties with multiple

resistance loci can provide prolonged resistance (52-54). R gene *xa5* showed very good

resistance (above 95%) to *Xoo* from all lineages. Previously, isolates of *Xoo* having

compatibility with *xa5* were only reported from Indian Lineages IX-I and IX-III, and

some Philipps and Korea isolates (35, 55-57). The only 2 Chinese *Xoo* from Yunnan

province (YN24 and YN04-5) belonging to IX-I and IX-III showed good infection

ability to rice with *xa5*, suggesting putative transmission of *Xoo* from India to South

China. Besides, several isolates from CX-5 and CX-6, especially those obtained

recently, can overcome the resistance contributed by *xa5*, indicating different origins of

these resistance compatibilities (Tables 1 and 2). Since rice variants with gene *xa5* were

less used in China, the development of its compatibility could not be due to a direct

selection pressure of this R gene.

The genetic background of *Xoo* is crucial for the interaction between *Xoo* and its host.

Isolates from different lineages and sub-lineages showed distinct capabilities to fight

against the same and different R genes. Overall, lineages and sub-lineages with stricter

geographical distribution can be more easily defeated by R genes than the wider

distributed ones (Tables 2 and S2). The underlying genetic determinations of these

differences call for advanced comparative genomics and wet experiments. However,

there is no lineage or sub-lineage with all isolates incompatible with one of the tested

R genes, which means that *Xoo* with different genetic backgrounds can adapt to

different rice hosts with variable resistances. This suggests that the introduction of new

R genes could reduce the loss causing by *Xoo*, but can't change the genetic diversity of

the *Xoo* population, as no lineage or sub-lineage can be eliminated thoroughly. This

partially because R genes usually restrict the pathogens to the initial infection site, but

have no killing abilities (58).

Materials and Methods

Genome sequencing

Xoo strains from 3 countries were used in this study (Table S1). The major strains were

isolated from China. And the represented *Xoo* strains for 4 different races from Japan

and 7 different races from Philippines were also sequenced.

The bacterial DNA of overnight cultures (30 °C) was extracted and purified using the

Easy-DNA kit (Invitrogen, USA) following the manufacturer's protocol. Total DNA

was sequenced by with Illumina HiSeq 2000, 2500, or 4000 to produce pair-end reads

with lengths of 100, 125 and 150 bp. For each *Xoo*, raw reads were assessed with the

FastQC tool (https://github.com/s-andrews/FastQC) and quality filtered using

Trimmomatic (https://github.com/timflutre/trimmomatic). The filtered reads were

error-corrected by library with Quake to produce clean reads (59).

Mapping and SNP calling

To study the population structure of *Xoo* from China, reads from each strain sequenced

was mapped onto the reference genome PXO99^A, which is used as a model strain for

studying the interaction between *Xoo* and rice host and is the first *Xoo* with complete

genome sequence (60), using BWA (61). Variant detection was performed using

Samtools mpileup (https://github.com/samtools/samtools) and filtered with a minimum

mapping quality of 30. SNP was excluded if its coverage was less than 10% or more

than 200% of the average coverage, if it was not supported by at least 5 reads on each

strand. Phage regions and repetitive sequences of the PXO99[A] genome were predicted

by PHASTER (http://phaster.ca) and RepeatScout (https://bix.ucsd.edu/repeatscout),

respectively. Snippy (https://github.com/tseemann/snippy) was used to generate the

whole genome alignment of all studied *Xoo*, and then the recombination was detected

by Gubbins (https://sanger-pathogens.github.io/gubbins) on the core genome alignment

after prophage and repeat regions were filtered. SNPs located within phage regions,

repetitive sequences or recombinant regions were excluded.

Assembly and whole genome alignment

To study the population structure of Asian *Xoo*, genomes from different locations were

downloaded from the Genbank as follows: 100 from India, 9 from Philippines, 1 from

South Korea and 1 from Japan (34, 35). We performed genome assembly from all the

strains sequenced in this study. For each strain, the pair-end clean reads were assembled

using Spades (62). After few evaluations, different sets of k-mer values were chosen

for Spades according to different read lengths. The final assemblies were obtained by

filtering out contigs with few reads supported or with lengths lower than 200 bp (Table

S1). Whole genome alignment was carried out on all the genomes obtained from public

database and assembled here by Parsnp (63) using the genome of PXO99[A] as reference.

Core SNPs were extracted from the core genome alignment by snp-sites

(https://github.com/sanger-pathogens/snp-sites). Recombination of the core genome

alignment was inferred by Gubbins. The final SNPs were obtained by filtering out the

prophage region and repeat sequences mentioned above, and the recombined regions

predicted here.

Phylogenetic analysis

The SNPs-based maximum likelihood (ML) phylogenies were built by RAxML with the generalized time-reversible model and a Gamma distribution to model site-specific rate variation (64). Bootstrap support values were calculated from 500 replicates. *Xoo* population structure of China was defined with the hierBAPS module of the BAPS software, which delineates the population structure by nested clustering (65). Three independent iterations with upper population sizes of 5, 10, and 15 were used to obtain optimal clustering of the population. Phylogenetic trees were annotated and visualized by iTOL (https://itol.embl.de/).

Virulence evaluation of *Xoo* strains

Virulence of *Xoo* was assesses by inoculating 6 near-isogenic rice lines, each carrying a specific R gene, IRBB2 (*Xa2*), IRBB3 (*Xa3*), IRBB4 (*Xa4*), IRBB5 (*xa5*), IRBB14 (*Xa14*) and IR24 (*Xa18*). IR24 was used as a susceptible check. Seeds of all lines in this study were obtained from the China National Rice Research Institute (CNRRI). Three weeks after sowing on a seedbed, seedlings were uprooted and transplanted into the field. The management of the rice plants in the field was proceeded as usual. Bacterial strain was cultivated on NA plates at 28 °C for 48 h. Then the bacterial colonies were suspended in sterile ddH$_2$O with concentration adjusted to 3×10^8 cfu/ml before inoculation. The leaf-clipping method was used to test the virulence of all the *Xoo* strains (66). Fifteen top fully expanded leaves per rice plant were inoculated with each strain. The lesions length was measured after 21 days post inoculation. The ratio of the lesion length compared to the whole leaf length was calculated as described

previously (31). When the ratio of the rice line was lower than 1/4, the rice line was

classified as resistant (R); when the ratios were between 1/4 and 1, the rice line was

rated as susceptible (S). The lesion length 14 days following inoculation was also

measured and used to compare with the data obtained 7 days later.

Data access

Sequence data generated for this study have been submitted to the NCBI Sequence Read

Archive under BioProject accession PRJNA350904.

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Conflict of Interest

The authors declare no conflict of interest.

Reference

1. Didelot X, Walker AS, Peto TE, Crook DW, & Wilson DJ (2016) Within-host evolution of bacterial pathogens. *Nat. Rev. Microbiol.* 14(3):150-162.

2. Baker S, Thomson N, Weill FX, & Holt KE (2018) Genomic insights into the emergence and
spread of antimicrobial-resistant bacterial pathogens. *Science* 360(6390):733-738.

3. Feasey NA*, et al.* (2016) Distinct *Salmonella Enteritidis* lineages associated with enterocolitis
in high-income settings and invasive disease in low-income settings. *Nat. Genet.* 48(10):1211-
1217.

4. Okoro CK*, et al.* (2012) Intracontinental spread of human invasive *Salmonella Typhimurium*
pathovariants in sub-Saharan Africa. *Nat. Genet.* 44(11):1215-1221.

5. Weill FX*, et al.* (2017) Genomic history of the seventh pandemic of cholera in Africa. *Science*
358(6364): 785-789.

6. Coll F*, et al.* (2018) Genome-wide analysis of multi- and extensively drug-resistant
Mycobacterium tuberculosis. *Nat. Genet.* 50(2):307-316.

7. Gardy JL & Loman NJ (2018) Towards a genomics-informed, real-time, global pathogen
surveillance system. *Nat. Rev. Genet.* 19(1):9-20.

8. Quainoo S*, et al.* (2017) Whole-Genome Sequencing of Bacterial Pathogens: the Future of
Nosocomial Outbreak Analysis. *Clin. Microbiol. Rev.* 30(4):1015-1063.

9. Zhong Z*, et al.* (2018) Population genomic analysis of the rice blast fungus reveals specific
events associated with expansion of three main clades. *ISME J* 12(8):1867-1878.

10. Baltrus DA, McCann HC, & Guttman DS (2017) Evolution, genomics and epidemiology of
Pseudomonas syringae: Challenges in Bacterial Molecular Plant Pathology. *Mol. Plant Pathol.*
18(1):152-168.

11. Brader G*, et al.* (2017) Ecology and Genomic Insights into Plant-Pathogenic and Plant-
Nonpathogenic Endophytes. *Annu. Rev. Phytopathol.* 55:61-83.

12. Hartmann FE & Croll D (2017) Distinct Trajectories of Massive Recent Gene Gains and Losses
in Populations of a Microbial Eukaryotic Pathogen. *Mol. Biol. Evol.* 34(11):2808-2822.

13. Hartmann FE, Sanchez-Vallet A, McDonald BA, & Croll D (2017) A fungal wheat pathogen
evolved host specialization by extensive chromosomal rearrangements. *ISME J* 11(5):1189-
1204.

14. Mansfield J*, et al.* (2012) Top 10 plant pathogenic bacteria in molecular plant pathology. *Mol.*
Plant Pathol. 13(6):614-629.

15. Mew T, Alvarez A, Leach J, & Swings J (1993) Focus on bacterial blight of rice. *Plant Dis.*
77:5-12.

16. Mew T (1987) Current status and future prospects of research on bacterial blight of rice. *Annu.*
Rev. Phytopathol. 25:359-382.

17. Fang Z (1963) *Rice Bacterial Blight* (Jiangsu People Press).

18. Zhang Q (2009) Genetics and Improvement of Bacterial Blight Resistance of Hybrid Rice in
China. *Rice Science* 16(2):83-92.

19. Pradhan SK*, et al.* (2015) Pyramiding of three bacterial blight resistance genes for broad-
spectrum resistance in deepwater rice variety, Jalmagna. *Rice (N Y)* 8(1):51.

20. Kottapalli KR, Lakshmi Narasu M, & Jena KK (2010) Effective strategy for pyramiding three
bacterial blight resistance genes into fine grain rice cultivar, Samba Mahsuri, using sequence
tagged site markers. *Biotechnol. Lett.* 32(7):989-996.

21. Huang N*, et al.* (1999) Pyramiding of bacterial blight resistance genes in rice: marker-assisted
selection using RFLP and PCR. *Theor. Appl. Genet.* 95(3):313-320.

22. Liu H, Yang W, Hu B, & Liu F (2007) Virulence analysis and race classification of *Xanthomonas*

472   *oryzae* pv. *oryzae* in China. *J. Phytopathol.* 155(3):129-135.

473   23.   Chen XL*, et al.* (2012) Elevational Variation in Diversity of *Xanthomonas oryzae* pv. *oryzae* in
474   South-West China. *J. Phytopathol.* 160(6):261-268.

475   24.   NODA T, YAMAMOTO T, KAKU H, & HORINO O (1996) Geographical Distribution of
476   Pathogenic Races of *Xanthomonas oryzae* pv. *oryzae* in Japan in 1991 and 1993. *Ann*
477   *Phytopathol Soc Jpn* 62:549–553.

478   25.   Liu H, Yang W, Hu B, & Liu F (2007) Virulence Analysis and Race Classification of
479   *Xanthomonas oryzae* pv. *oryzae* in China. *J Phytopathol* 155(3):129-135.

480   26.   Li G, Song C-F, Pang X-M, Yang Y, & Wang J-S (2008) Analysis of Pathotypic and Genotypic
481   Diversity of *Xanthomonas oryzae* pv. *oryzae* in China. *J Phytopathol* 154(4):208-218.

482   27.   Jeung JU, Heu SG, Shin MS, Vera Cruz CM, & Jena KK (2006) Dynamics of *Xanthomonas*
483   *oryzae* pv. *oryzae* Populations in Korea and Their Relationship to Known Bacterial Blight
484   Resistance Genes. *Phytopathol.* 96(8):867-875.

485   28.   Adhikari TB, Mew TW, & Leach JE (1999) Genotypic and pathotypic diversity in *Xanthomonas*
486   *oryzae* pv. *oryzae* in Nepal. *Phytopathol.* 89(8):687-694.

487   29.   Mishra D*, et al.* (2013) Pathotype and genetic diversity amongst Indian isolates of *Xanthomonas*
488   *oryzae* pv. *oryzae*. *PLoS One* 8(11):e81996.

489   30.   Li G, Song CF, Pang XM, Yang Y, & Wang JS (2009) Analysis of Pathotypic and Genotypic
490   Diversity of *Xanthomonas oryzae* pv. *oryzae* in China. *J. Phytopathol.* 157(4):208-218.

491   31.   Fang Z*, et al.* (1990) Studies on pathotypes of *Xanthomonas campestris* pv. *oryzae* in China.
492   *Acta Phytopathol Sin* 20:81-88.

493   32.   Huang X*, et al.* (2012) A map of rice genome variation reveals the origin of cultivated rice.
494   *Nature* 490(7421):497-501.

495   33.   Kovach MJ, Sweeney MT, & McCouch SR (2007) New insights into the history of rice
496   domestication. *Trends Genet.* 23(11):578-587.

497   34.   Quibod IL*, et al.* (2016) Effector Diversification Contributes to *Xanthomonas oryzae* pv. *oryzae*
498   Phenotypic Adaptation in a Semi-Isolated Environment. *Sci. Rep.* 6:34137..

499   35.   Midha S*, et al.* (2017) Population genomic insights into variation and evolution of *Xanthomonas*
500   *oryzae* pv. *oryzae*. *Sci. Rep.* 7:40694.

501   36.   Tzeng TH, Lyngholm LK, Ford CF, & Bronson CR (1992) A Restriction-Fragment-Length-
502   Polymorphism Map and Electrophoretic Karyotype of the Fungal Maize Pathogen
503   Cochliobolus-Heterostrophus. *Genetics* 130(1):81-96.

504   37.   Hulbert SH*, et al.* (1988) Genetic-Analysis of the Fungus, Bremia-Lactucae, Using Restriction
505   Fragment Length Polymorphisms. *Genetics* 120(4):947-958.

506   38.   Lazo GR, Roffey R, & Gabriel DW (1987) Pathovars of *Xanthomonas-Campestris* Are
507   Distinguishable by Restriction-Fragment-Length-Polymorphism. *Int. J. Syst. Bacteriol.*
508   37(3):214-221.

509   39.   Mishra D*, et al.* (2013) Pathotype and Genetic Diversity amongst Indian Isolates of
510   *Xanthomonas oryzae* pv. *oryzae*. *PLoS One* 8(11).

511   40.   Leach JE*, et al.* (1992) Assessment of Genetic Diversity and Population-Structure of
512   *Xanthomonas-Oryzae* Pv *Oryzae* with a Repetitive DNA Element. *Appl. Environ. Microbiol.*
513   58(7):2188-2195.

514   41.   George MLC, Bustamam M, Cruz WT, Leach JE, & Nelson RJ (1997) Movement of
515   *Xanthomonas oryzae* pv *oryzae* in southeast Asia detected using PCR-based DNA fingerprinting.

Phytopathol. 87(3):302-309.

42.   Adhikari TB*, et al.* (1995) Genetic Diversity of *Xanthomonas-Oryzae* Pv *Oryzae* in Asia. *Appl.*
Environ. Microbiol. 61(3):966-971.

43.   Nelson RJ*, et al.* (1994) Relationship between Phylogeny and Pathotype for the Bacterial-Blight
Pathogen of Rice. *Appl. Environ. Microbiol.* 60(9):3275-3283.

44.   Zhang Q, Leach JE, Nelson RJ, & Mew TW (1997) Genetic Structure of Rice Bacterial Blight
Pathogen Population in China. *Acta Agronomica Sinica* 23(02):150-158.

45.   Triplett LR*, et al.* (2011) Genomic analysis of *Xanthomonas oryzae* isolates from rice grown in
the United States reveals substantial divergence from known *X. oryzae* pathovars. *Appl. Environ.*
Microbiol. 77(12):3930-3937.

46.   Gonzalez C*, et al.* (2007) Molecular and pathotypic characterization of new *Xanthomonas*
oryzae strains from West Africa. *Mol. Plant. Microbe Interact.* 20(5):534-546.

47.   Yang SQ*, et al.* (2013) Molecular and pathogenic characterization of new *Xanthomonas oryzae*
pv. *oryzae* strains from the coastline region of Fangchenggang city in China. *World J. Microbiol.*
Biotechnol. 29(4):713-720.

48.   Tu M, Lu BR, Zhu Y, & Wang Y (2007) Abundant within-varietal genetic diversity in rice
germplasm from Yunnan Province of China revealed by SSR fingerprints. *Biochem. Genet.*
45(11-12):789-801.

49.   Xiong ZY*, et al.* (2010) Differentiation and distribution of indica and japonica rice varieties
along the altitude gradients in Yunnan Province of China as revealed by InDel molecular
markers. *Genet. Resour. Crop Evol.* 57(6):891-902.

50.   Chen S, Lin XH, Xu CG, & Zhang QF (2000) Improvement of bacterial blight resistance of
'Minghui 63', an elite restorer line of hybrid rice, by molecular marker-assisted selection. *Crop*
Sci. 40(1):239-244.

51.   Xu Z*, et al.* (2004) Race monitoring of rice bacterial blight (*Xanthomonas oryzae* pv. *oryzae*) in
China. *Chin J Rice Sci* 18:469-472.

52.   Joseph M*, et al.* (2004) Combining bacterial blight resistance and Basmati quality
characteristics by phenotypic and molecular marker-assisted selection in rice. *Mol. Breed.*
13(4):377-387.

53.   Sundaram RM*, et al.* (2008) Marker assisted introgression of bacterial blight resistance in
Samba Mahsuri, an elite indica rice variety. *Euphytica* 160(3):411-422.

54.   Basavaraj SH*, et al.* (2010) Marker-assisted improvement of bacterial blight resistance in
parental lines of Pusa RH10, a superfine grain aromatic rice hybrid. *Mol. Breed.* 26(2):293-305.

55.   Carpenter SCD*, et al.* (2018) A Strain of an Emerging Indian *Xanthomonas oryzae* pv. *oryzae*
Pathotype Defeats the Rice Bacterial Blight Resistance Gene xa13 Without Inducing a Clade III
SWEET Gene and Is Nearly Identical to a Recent Thai Isolate. *Front. Microbiol.* 9:2703.

56.   Huang S*, et al.* (2016) The broadly effective recessive resistance gene *xa5* of rice is a virulence
effector-dependent quantitative trait for bacterial blight. *Plant J.* 86(2):186-194.

57.   Jeung JU, Heu SG, Shin MS, Cruz CMV, & Jena KK (2006) Dynamics of *Xanthomonas oryzae*
pv. *oryzae* populations in Korea and their relationship to known bacterial blight resistance genes.
Phytopathol. 96(8):867-875.

58.   Shen YW & Ronald P (2002) Molecular determinants of disease and resistance in interactions
of *Xanthomonas oryzae* pv. *oryzae* and rice. *Microb. Infect.* 4(13):1361-1367.

59.   Kelley DR, Schatz MC, & Salzberg SL (2010) Quake: quality-aware detection and correction

of sequencing errors. *Genome Biol.* 11(11):R116.

60. Salzberg SL*, et al.* (2008) Genome sequence and rapid evolution of the rice pathogen
Xanthomonas oryzae pv. *oryzae* PXO99A. *BMC Genomics* 9:204.

61. Li H & Durbin R (2009) Fast and accurate short read alignment with Burrows-Wheeler
transform. *Bioinformatics* 25(14):1754-1760.

62. Bankevich A*, et al.* (2012) SPAdes: A New Genome Assembly Algorithm and Its Applications
to Single-Cell Sequencing. *J. Comput. Biol.* 19(5):455-477.

63. Treangen TJ, Ondov BD, Koren S, & Phillippy AM (2014) The Harvest suite for rapid core-
genome alignment and visualization of thousands of intraspecific microbial genomes. *Genome*
Biol. 15(11):524.

64. Stamatakis A (2014) RAxML version 8: a tool for phylogenetic analysis and post-analysis of
large phylogenies. *Bioinformatics* 30(9):1312-1313.

65. Cheng L, Connor TR, Siren J, Aanensen DM, & Corander J (2013) Hierarchical and Spatially
Explicit Clustering of DNA Sequences with BAPS Software. *Mol. Biol. Evol.* 30(5):1224-1228.

66. Kauffman H, Reddy A, Hsieh S, & Merca S (1973) An improved technique for evaluating
resistance of rice varieties to *Xanthomonas oryzae*. *Plant Dis Rep* 57:737–741.

Figure 1 Population structure of *X. oryzae* pv. *oryzae*. The maximum likelihood tree was inferred

by RAxML with the generalized time-reversible model and a Gamma distribution to model site-

specific rate variation based on the core SNPs of all the *Xoo* genome sequences. The six lineages

were decorated with different colors. From inner to outer, the first circle refers to major rice planting

areas, the second one describes isolated times of these strains and the third one represents the rice

subspecies which the *Xoo* strain isolates. The outmost 6 squares represent the virulence analysis of

these *Xoo* against 6 different rice near-isogenic lines with one known resistance gene in each line,

with solid square for resistance and hollow one for sensitivity. Bootstrap support values were

calculated from 500 replicates, and only values of > 70% were labelled.

Figure 2 Distribution of the 6 major Chinese *X. oryzae* pv. *oryzae* lineages. Pathotypes of *Xoo*

were determined on the following 13 near-isogenic lines, each carrying a specific R gene, IRBB2

(*Xa2*), IRBB3 (*Xa3*), IRBB4 (*Xa4*), IRBB5 (*xa5*), IRBB14 (*Xa14*). IR24 was used as a susceptible

check. (a) The map of the 3 major rice producing areas in China. (b) Distribution of the 6 *Xoo*

lineages among the 3 rice planting areas. (c) Dynamics of the 6 *Xoo* lineages during the past 30

years.

Figure 3 Phylogeography of CX-5 and CX-6. Maximum likelihood phylogeny of Lineage CX-5

(a) and CX-6 (b). Different sub-lineages were displayed by different branch colors. Bootstrap

support values were calculated from 500 replicates. The recent outbreaks are represented by

different backgrounds. Isolated information of *Xoo* including location, time and rice subspecies

were shown in three colored strips. (c) Mean Pairwise SNP Distances (MPSDs) of *Xoo* from the

outbreak, and between one in an outbreak and the other one out of the outbreak but in the same sub-

lineage were displayed by two different boxes beside the tree.

Figure 4 Virulence analysis and race classification of *X. oryzae* pv. *oryzae* China. Pathogenicity

profile of each race corresponding to was shown in Table S1. (a) Proportions of the 9 *Xoo* races. (b)

Distribution of the 9 races among the 3 major rice planting areas in China. (c) Dynamics of the 9

Xoo races during the past 40 years. (d) Distribution of the 9 races in Lineage CX-6 and others.

Table 1 Resistance of different rice lines to *Xoo* strains isolated at different times.

Shown is the percentage of *Xoo* strains isolated at different periods that are unable to

cause disease.

Time (# of strains)	IRBB2 (*Xa2*)	IRBB3 (*Xa3*)	IRBB4 (*Xa4*)	IRBB5 (*xa5*)	IRBB14 (*Xa14*)	IR24 (*Xa18**)
<1990 (35)	37	0	97	94	25	20
<2000 (69)	25	58	64	100	28	12
2003/2004 (97)	49	35	34	100	56	7
2014 (42)	36	5	79	90	33	5

*: Recurrent parent *Xa18*

620 **Table 2 Resistance of different rice lines to *Xoo* strains of different lineages.** Shown

621 is the percentage of *Xoo* strains of different lineages that are unable to cause disease.

Lineage (# of strains)	IRBB2 (*Xa2*)	IRBB3 (*Xa3*)	IRBB4 (*Xa4*)	IRBB5 (*xa5*)	IRBB14 (*Xa14*)	IR24 (*Xa18**)
CX-1 (19)	89	79	42	100	95	21
CX-2 (15)	73	93	80	100	100	7
CX-3 (20)	100	20	70	100	99	10
CX-5 (53)	11	32	47	96	17	11
CX-6 (123)	29	25	62	98	28	6

622 *: Recurrent parent *Xa18*. CX-4 was not included because of few isolates it contained.

624 **Supplementary Information**

625 Figure S1

626 Tables S1 and S2

Figure S1 Population structure of Asian *Xanthomonads oryzae* pv. *oryzae*. For phylogenetic
analysis, parsnp was used to call SNPs, and RAxML was used to build the ML tree. Background
colors of nodes represent different lineages. The outmost circle refers to different countries where
Xoo were isolated.

636 **Table S2 Resistance of different rice lines to *Xoo* strains of different sub-lineages**
637 **of CX-5 and CX-6.** Shown is the percentage of *Xoo* strains of different lineages that
638 are unable to cause disease.

Lineage (# of strains)	IRBB2 (*Xa2*)	IRBB3 (*Xa3*)	IRBB4 (*Xa4*)	IRBB5 (*xa5*)	IRBB14 (*Xa14*)	IR24 (*Xa18**)
CX-5.1 (5)	0	20	20	100	0	0
CX-5.3 (12)	25	25	67	92	17	17
CX-5.4 (8)	25	50	75	100	28	25
CX-5.5 (18)	0	28	28	100	28	0
CX-6.1(10)	30	30	90	100	50	20
CX-6.2 (47)	13	38	79	98	23	4
CX-6.3 (18)	10	17	68	100	11	0
CX-6.4 (18)	0	0	61	94	0	0
CX-6.5(31)	90	32	29	97	74	10

639 *: Recurrent parent *Xa18.* CX-5.2 was not included because of few isolates it contained.
640
641