

1 **Pervasive introgression of MHC genes in newt hybrid zones**

2 Running title: Pervasive MHC introgression

3 K. Dudek<sup>1</sup>, T. S. Gaczorek<sup>1</sup>, P. Zieliński<sup>1</sup>, W. Babik<sup>1\*</sup>

4 *<sup>1</sup>Institute of Environmental Sciences, Faculty of Biology, Jagiellonian University,*

5 *Gronostajowa 7, 30-387 Kraków, Poland*

6

7 \*Correspondence:

8 [wieslaw.babik@uj.edu.pl](mailto:wieslaw.babik@uj.edu.pl)

9

10 **Abstract**

11 Variation in the vertebrate major histocompatibility complex (MHC) genes is crucial for  
12 fighting pathogen assault. Because new alleles confer a selective advantage, MHC should  
13 readily introgress between species, even under limited hybridization. Using replicated  
14 transects through two hybrid zones between strongly reproductively isolated newts, we  
15 demonstrated recent and ongoing MHC introgression. Its extent correlated with the age of  
16 contact. In the older zone, MHC similarity between species within transects exceeded that of  
17 between transects within species, implying pervasive introgression – a massive exchange of  
18 MHC genes, not limited to specific variants. In simulations, the observed pattern emerged  
19 under the combined action of balancing selection and hybridization, but not when these  
20 processes acted separately. Thus, massive introgression at advanced stages of divergence can  
21 introduce novel and restore previously lost MHC variation, boosting the adaptive potential of  
22 hybridizing taxa. In consequence, MHC genes may be the last to stop introgressing between  
23 incipient species.

24

## 25 **Introduction**

26 Speciation is usually a prolonged process, because reproductive isolation between  
27 differentiating taxa evolves gradually (Coyne & Orr, 2004). Speciation commonly involves  
28 independent evolution in allopatry (Kisel & Barraclough, 2010). However, due to the dynamic  
29 nature of species ranges, periods of allopatry are often accompanied by episodes of contact,  
30 which set the stage for natural hybridization (Abbott et al., 2013; Harrison & Larson, 2016). If  
31 hybridization leads to introgression, some introgressed variants may be beneficial in the  
32 recipient species. Introgression of such variants will be favored by natural selection, even if  
33 overall introgression is constrained due to low fitness of hybrids and genome-wide barriers to  
34 neutral introgression (Barton, 1979). Such adaptive introgression may occur at many loci,  
35 perhaps even providing variation that fuels adaptive radiations (Meier et al., 2017), or may be  
36 restricted to particular genes or genomic regions (Hedrick, 2013; Huerta-Sánchez et al.,  
37 2014). While examples of adaptive introgression have been documented and theory  
38 underlying adaptive introgression is available, general properties of the process and the  
39 resulting empirical patterns are not sufficiently understood.

40       Some genes or classes of genes may be particularly prone to adaptive introgression due  
41 to the evolutionary processes affecting them. However, paradoxically, convincing  
42 demonstration of introgression of these genes may be the most challenging. Genes involved in  
43 fighting pathogen assault, which often evolve under various forms of selection that maintain  
44 and promote diversity (collectively termed balancing selection, BS), are prime candidates for  
45 adaptive introgression (Fijarczyk, Dudek, Niedzicka, & Babik, 2018; Schierup, Vekemans, &  
46 Charlesworth, 2000). This is because novel alleles may confer selective advantage and  
47 selection would favor their introgression (Phillips et al., 2018). Vertebrate Major  
48 Histocompatibility Complex (MHC) genes are a textbook example of long-term BS (Spurgin

49 & Richardson, 2010) driven in a large part by Red Queen dynamics (Ejmond & Radwan,  
50 2015). Adaptive MHC introgression between hybridizing species has been inferred in several  
51 systems (Abi-Rached et al., 2011; Fijarczyk et al., 2018; Grossen, Keller, Biebach, Croll, &  
52 Consortium, 2014; Nadachowska-Brzyska, Zielinski, Radwan, & Babik, 2012), with varying  
53 strength of supporting evidence. However, studying introgression of targets of balancing  
54 selection is challenging because BS itself generates patterns of variation and differentiation  
55 resembling those due to introgression, which makes distinguishing between these two  
56 processes difficult (Fijarczyk & Babik, 2015). Furthermore, MHC is a dynamically evolving  
57 multigene family characterized by frequent duplications and pseudogenizations, resulting in  
58 copy number variation and the mixture of functional genes and pseudogenes in the genome  
59 (Kelley, Walter, & Trowsdale, 2005). These genomic complexities impede both molecular  
60 analyses and realistic modeling and testing of competing evolutionary scenarios.

61 Hybrid zones – areas where incipient species meet and hybridize – are considered  
62 natural laboratories and windows on evolutionary processes, because they allow the outcomes  
63 of multiple generations of hybridization under natural conditions to be studied (Harrison &  
64 Larson, 2016). Hybrid zones allow direct observation of introgression, comparison between  
65 genomic regions, estimation of the strength of genome-wide reproductive isolation, and  
66 rigorous testing of adaptive introgression and alternative explanations (Gompert, Mandeville,  
67 & Buerkle, 2017; Payseur, 2010). Inferences are especially strong if multiple transects  
68 through a hybrid zone are available, which allows for: i) testing the repeatability of patterns  
69 between transects, ii) in the case of polymorphic genes, comparing intraspecific divergence  
70 between transects with interspecific divergence within transects. The latter constitutes a  
71 powerful test for introgression, as increased genetic similarity in contact zones is not expected  
72 under any scenario other than introgression (Fraïsse, Belkhir, Welch, & Bierne, 2016;

73 Nadachowska-Brzyska et al., 2012). In the context of testing adaptive introgression of genes  
74 under BS, such as the MHC, hybrid zones between strongly reproductively isolated taxa are  
75 excellent tools, because theory predicts a pronounced contrast between targets of adaptive  
76 introgression and the rest of the genome.

77 In this study we investigated introgression of MHC genes through hybrid zones between  
78 two European salamanders: Carpathian (*Lissotriton montandoni*) and smooth (*L. vulgaris*)  
79 newts. These newts hybridize in a narrow contact zone along the Carpathians, but  
80 hybridization is currently limited: even at the centre of the hybrid zone, parental genotypes  
81 predominate, resulting in a bimodal distribution of genotypes, that indicates strong  
82 reproductive isolation (Babik, Szymura, & Rafiński, 2003; Zieliński et al., 2013). However *L.*  
83 *montandoni* and *L. vulgaris* have experienced a long and apparently complex history of  
84 genetic exchange (Zieliński, Nadachowska-Brzyska, Dudek, & Babik, 2016), exemplified by  
85 a complete replacement of *L. montandoni* mtDNA by that derived from several  
86 phylogeographic lineages of *L. vulgaris* (Pabijan, Zielinski, Dudek, Stuglik, & Babik, 2017;  
87 Zieliński et al., 2013). Introgression of MHC classes I and II between *L. montandoni* and *L.*  
88 *vulgaris* has previously been inferred using continent-scale comparisons of differentiation and  
89 allele sharing between multiple evolutionary lineages within the *L. vulgaris* species complex  
90 (Fijarczyk et al., 2018; Nadachowska-Brzyska et al., 2012). However, such large scale  
91 analyses provide only limited insight into the temporal and spatial dynamics of introgression.  
92 Analyses of MHC introgression in hybrid zones are more informative in this respect, as they  
93 allow testing for recent or ongoing introgression and estimation of its strength.

94 We analyzed zones of hybridization between *L. montandoni* and two evolutionary  
95 lineages of *L. vulgaris*: i) *L. montandoni* x *L. v. ampelensis* (the IN zone) and ii) *L.*  
96 *montandoni* x *L. v. vulgaris* (the OUT zone). The two lineages within *L. vulgaris* diverged

97 more than 1 Mya (Pabijan et al., 2017; Zieliński et al., 2016), and each of them has  
98 experienced a distinct history of hybridization with *L. montandoni* (Zieliński et al., 2016). In  
99 particular, the IN zone appears older (Zieliński et al., 2019). Within each zone we sampled  
100 two transects separated by a distance of 100-200 km, which is two orders of magnitude larger  
101 than the per generation dispersal distances of these newts. We took advantage of two  
102 hierarchically sampled transects through each hybrid zone to compare introgression of MHC  
103 genes to that of more than 1100 other protein-coding genes scattered throughout the newt  
104 genome.

105

## 106 **Materials and Methods**

### 107 *Experimental design and sampling*

108 Hierarchical sampling comprised two hybrid zones with two transects within each: i) the IN  
109 zone (*L. montandoni* x *L. vulgaris ampelensis*) inside the Carpathian Basin, with the transects  
110 Lypcha (L) and Remetea (R) separated by ca 200 km, and ii) the OUT zone (*L. montandoni* x  
111 *L. v. vulgaris*) outside the Carpathian Basin, with the transects Suceava (S) and Tazlau (T)  
112 separated by ca. 100 km (Fig. 1A). Note that the compound R transect consists of three  
113 contact zones, as the two mountain ranges where *L. montandoni* occurs are separated by an  
114 upland inhabited by *L. vulgaris*. The description of all transects, including environmental  
115 characteristics, is given in Zieliński et al. (2019). We attempted to sample all water bodies  
116 where newts were present, with special emphasis on the centre of each zone, where both  
117 species occurred in syntopy and individuals of intermediate morphology were found. Each  
118 water body was considered a distinct locality and a distinct breeding population, although the  
119 distances between localities were often < 1 km, within the newt dispersal ranges. Newts were

120 captured by dip netting during breeding season, tailtip biopsies were taken and preserved in  
121 96% ethanol; animals were released immediately after sampling. DNA was extracted using  
122 the Wizard Genomic DNA purification kit (Promega). Data on genome-wide admixture,  
123 available for 96% of individuals genotyped in MHC, based on 2849 SNPs (minor allele  
124 frequency,  $MAF \geq 0.05$ ) in ca. 1100 protein-coding genes are from Zieliński et al. (2019).

### 125 *MHC genotyping, haplotypes and linkage*

126 The variable 2<sup>nd</sup> exon of both MHC class I and class II genes was PCR amplified, amplicons  
127 were sequenced on the Illumina platform and genotyping was performed using the adjustable  
128 clustering method implemented in AmpliSAS (Sebastian, Herdegen, Migalska, & Radwan,  
129 2016). Laboratory and bioinformatics procedures used for MHC class I genotyping were  
130 described in detail in Fijarczyk et al. (2018). MHC class II was amplified with primers based  
131 on those used in Nadachowska-Brzyska et al. (2012), modified to include additional variation  
132 in the regions of primer annealing, as identified by transcriptome resequencing and genome  
133 walking. The class II primers (see Supplementary Materials) target a longer fragment than  
134 that studied by Nadachowska-Brzyska et al. (2012) (244 vs 204 bp) and we verified,  
135 genotyping 9 individuals from that previous study, that the modified primers amplified all  
136 alleles previously detected in these individuals. Details of class II genotyping, analyses of  
137 diversity and tests of selection are in Supplementary Materials.

138 Eight MHC class I haplotypes were reported in Fijarczyk et al. (2018). To determine  
139 MHC class II haplotypes we used the same individuals: parents and offspring from two  
140 families of interspecific *L. montandoni* x *L. v. vulgaris* crosses (11 and 13 F1 offspring). To  
141 estimate linkage between the two MHC classes we compared class I and II genotypes in one

142 of the families (parents and 113 offspring) used previously for the construction of a linkage  
143 map (Niedzicka, Dudek, Fijarczyk, Zieliński, & Babik, 2017).

#### 144 *Classical MHC alleles and supertypes*

145 Although the primers were designed from transcriptome sequences, they amplified both  
146 highly expressed, putative classical MHC alleles, and alleles expressed at lower level, that  
147 may be pseudogenes or nonclassical MHC. Because almost all these alleles segregate as  
148 stable haplotypes (Fijarczyk et al., 2018, see also Results), the loci are tightly linked.  
149 Therefore all should be useful markers for detecting introgression of the MHC region.  
150 Nevertheless, classical genes may undergo different dynamics than nonclassical/nonfunctional  
151 genes. It is also possible that alleles within functional classes may be exchangeable and thus  
152 affected mainly by drift. Therefore we performed also additional analyses: i) excluding  
153 putative nonclassical/nonfunctional MHC class I alleles, and ii) grouping MHC alleles into  
154 supertypes (details in Supplementary Materials).

#### 155 *Statistical analysis*

156 Both MHC class I and class II are duplicated in *Lissotriton* newts and exhibit copy number  
157 variation. Assignment of alleles to loci was not possible, therefore we decided to perform  
158 statistical analyses on data with each allele encoded as an indicator binary variable (presence-  
159 absence). To assess differentiation between species and between transects visually and define  
160 main directions of the overall variation we used Principal Component Analysis (PCA) in  
161 adegenet. To assess overall MHC differentiation as well as differentiation between species  
162 within transects we used multidimensional scaling (MDS) ordination of pairwise  $F_{ST}$   
163 distances. To estimate the fraction of variation explained by consecutive levels of hierarchical



164 structuring we employed the AMOVA in Arlequin (Excoffier & Lischer, 2010) with the  
165 number of pairwise differences between individuals as the distance measure.

166 Comparison of the width and location of the centre of allele frequency clines is useful to  
167 compare introgression between markers. Here we compared MHC clines to the average  
168 genome-wide (SNP) clines using the following approach. For each individual the proportion  
169 of the genome derived from each species estimated with Admixture (Alexander, Novembre,  
170 & Lange, 2009) was taken from Zieliński et al. (2019). The proportion of MHC alleles  
171 derived from each species was estimated using Structure (Falush, Stephens, & Pritchard,  
172 2007) with MHC alleles encoded as binary dominant data. For transects L, S and T, two  
173 genetic clusters were strongly supported. In the compound transect R (Fig. 1A) three clusters  
174 were identified – one for *L. montandoni* and two for *L. vulgaris*, as populations of this species  
175 from an upland between two mountain ranges were assigned to a separate cluster. For the  
176 purpose of further analyses Q values from the two *L. vulgaris* clusters were summed.

177 Geographic clines were then fitted to the average population Q values for both genome-wide  
178 and MHC data using HZAR (Derryberry, Derryberry, Maley, & Brumfield, 2014). For both  
179 MHC and SNPs we fitted the same cline model, which estimated the location of cline center  
180 ( $c$ ) and width ( $w$ ,  $1/\text{maximum slope}$ ) as well as the allele frequencies at the ends of the cline.

181 We then used the parameters from the genome-wide cline to test whether clines estimated for  
182 MHC data fit the data better, which would indicate differences between clines. The fit was  
183 compared using the Akaike information criterion score corrected for small sample size  
184 (AICc), with  $\Delta\text{AICc} \geq 5$  was considered a significant support for an MHC-specific model.

185 Isolation by distance between interspecific population pairs within each transect was  
186 tested using a matrix permutation procedure (10 000 permutations), in which rows and  
187 columns were permuted independently. We tested for an increased allele sharing closer to the

188 centre of the zone in each transect separately. For each species 1-3 (depending on the  
189 distribution of localities in the transect) populations at the ends of the transect were classified  
190 as allopatric, while the remaining allotopic populations were classified as parapatric. For each  
191 species populations within groups were pooled, the fraction of alleles shared between species  
192 was calculated for each group and significance of the difference between groups was tested by  
193 permuting individuals within species between groups 10 000 times. Because in the OUT zone  
194 many syntopic populations were available and most individuals in these populations showed  
195 little genome-wide admixture, we also tested for an increased allele sharing in syntopy.

### 196 *Simulations*

197 A multiallelic gene under negative frequency dependent selection was simulated according to  
198 the infinite allele model in a linear stepping stone (Fig. S1) using Selector (Currat, Gerbault,  
199 Di, Nunes, & Sanchez-Mazas, 2015). Although newt MHC is multilocus, its evolutionary  
200 dynamics should be reflected adequately by a single, multiallelic locus, because most likely  
201 entire haplotypes are subject to selection. To verify this, additional simulations with  
202 multilocus MHC haplotypes were performed, as described in Supplementary Methods. An  
203 ancestral species split into two equally sized, completely isolated descendant species, which  
204 thus initially shared a pool of alleles (Fig. S1I). Following the split, each species colonized a  
205 world of demes conforming to a one-dimensional stepping stone model arranged into a  
206 horseshoe shape that approximated the two transects of the IN zone, i.e. the distance between  
207 transects was 5 times larger than the length of a transect within each species (Fig. S1II). No  
208 mutations were allowed, so differentiation between species occurred only via drift,  
209 maximizing the retention of ancestral polymorphism. Following prolonged evolution in  
210 isolation, secondary contact and hybridization ensued (Fig. S1III). A single deme hybrid zone

211 acted as a partial barrier to gene flow – immigration into the zone was high but emigration  
212 from the zone was strongly reduced as in classical tension zone models.

213 We investigated the effect of the following parameters: strength of selection ( $s = 0 -$   
214  $0.3$ ), deme size ( $N = 100 - 1000$ ), initial number of alleles ( $n_a = 15 - 500$ ), migration between  
215 demes ( $Nm = 0.1 - 2.5$ ), strength of barrier to introgression formed by the hybrid zone  
216 (emigration from the zone as a fraction of that from demes within species  $N_{hm} = 0.01 -$   
217  $0.1Nm$ ) and time of hybridization ( $0.01 - 0.1$  of time in isolation). A single value of time in  
218 isolation ( $t = 16\ 000$  generations) was used; because differentiation depends both on time of  
219 isolation and population sizes, by varying deme size we also implicitly varied time of  
220 isolation. The total number of alleles and the number of alleles within deme stabilized  
221 quickly, within ca. 2500 generations, regardless of the deme size.

222 At the end of each simulation 16 individuals from each deme within each transect  
223 were sampled (Fig. S1IV) to calculate: i) fractions of variation (AMOVA) accounted for by  
224 the between species and between transect within species levels, ii) the numbers and fractions  
225 of alleles shared exclusively (those shared by two focal groups but absent from all other  
226 groups) between species within transect and between transects within species. To investigate  
227 the temporal and spatial dynamics of introgression we followed the frequency of  
228 heterospecific genes through time in a single transect. Details of simulations are in  
229 Supplementary Materials.

230

## 231 **Results**

### 232 *MHC variation and evidence for positive selection*

233 MHC was genotyped in 1672 individuals from 128 localities in four transects (Fig. 1A, Table  
234 S1). Genotyping repeatability was 95% (class I) and 98% (class II). Overall variation was  
235 high: 920 MHC class I alleles (25 supertypes) in 1679 individuals and 263 class II alleles (22  
236 supertypes) in 1674 individuals (Table S1). The number of MHC alleles per individual ranged  
237 from 5 to 25 (class I: 4 – 21, class II: 1 – 7, Fig. S2), indicating extensive copy number  
238 variation. Eight haplotypes identified by genotyping two newt families contained 1-3 class II  
239 genes, with 5-9 genes per haplotype reported earlier for class I (Fijarczyk et al., 2018).

240 A strong signal of positive selection was detected in class II ( $\Delta AIC = 74.6$  for  
241 comparison of PAML models with (M8) and without (M7) positive selection), similar to that  
242 reported for class I (Fijarczyk et al., 2018). Only two class II codons were identified as targets  
243 of positive selection, but amino acid variation in positions corresponding to the human  
244 Antigen Binding Sites was extremely high (Table S2, Fig. S3).

### 245 *Comparison between zones and transects: strong MHC introgression in the IN zone*

246 Based on Principal Component (PC) ordination of the genome-wide single nucleotide  
247 polymorphism (SNP) data, species were well separated along PC 1, which explained five  
248 times more variation than PC 2 (Fig. 1B). In contrast, species were poorly separated on the  
249 MHC's PC1-PC2 plane, even when syntopic populations (defined as localities where both  
250 species were identified based on morphology) were excluded from the analyses, and the  
251 extent of separation appeared to differ between the IN and OUT zones (Fig. 1C). This  
252 interpretation did not change if the MHC classes were analysed separately (Fig. S4), or if  
253 MHC supertypes were used instead of alleles. In analyses performed for each zone

254 independently, no separation by species but some separation by transect was observed along  
255 PC 1 in the IN zone, while clear separation of species along PC 1 was evident in the OUT  
256 zone (Fig. 2A, B). Patterns of allele sharing were consistent with the above picture (Fig. 2C,  
257 D). In the IN zone a higher fraction of alleles were shared exclusively (shared by two focal  
258 groups but absent from other groups) between species within transect (L: 7.0%, R: 16.2%)  
259 than between transects within species (*L. montandoni*: 3.6%, *L. vulgaris*: 2.1%), while in the  
260 OUT zone exclusive allele sharing between transects within each species (*L. montandoni*:  
261 17.3%, *L. vulgaris*: 9.7%) was much higher than between species within transect (S: 0.5%, T:  
262 3.2%, Breslow-Day test for homogeneity of odds ratios, IN:  $P = 2.6 \times 10^{-4}$ , OUT:  $P = 1.9 \times$   
263  $10^{-4}$ ).

264 Analysis of Molecular Variance (AMOVA) revealed no differentiation between  
265 species, but significant differentiation between transects within species in the IN zone. In the  
266 OUT zone, the between species component accounted for a substantial fraction of variation  
267 (11.6%, Table 1A). The results for supertypes were virtually identical (Table S3). Thus, in the  
268 IN zone MHC variation was structured mainly by geography, while in the OUT zone by  
269 species (Fig. 1D, 2A, B). In both zones, the genome-wide differentiation between species was  
270 much higher than that between transects within species (Table 1A).

271

272 *The extent of introgression within transects*

273 Despite strong evidence for introgression in the IN zone, at local scales, i.e. within transects,  
274 interspecific differentiation in MHC was visible everywhere, though more so in the OUT zone  
275 (Table 1B, S3, Fig. 2E, F). The width or location of MHC clines did not differ from those of  
276 genome-wide clines in any transect (Fig. 1A, Table S4). Thus, a sharp transition in MHC,  
277 coinciding with the transition in genome-wide ancestry, occurs in each transect. Hence, at  
278 local scales, MHC gene flow is currently more restricted between species than between  
279 populations within species. However, the magnitudes of MHC and genome-wide  
280 differentiation on both sides of the zone are difficult to compare, as Admixture and Structure  
281 analyses that we used to estimate genome-wide and MHC ancestry enforce both scales to run  
282 from 0 to 1.

283         If introgression decreased with the distance from the centre of the zone, we would  
284 expect significant isolation by distance between heterospecific population pairs within  
285 transects. No such pattern was detected in any transect (matrix randomization test, all  $P >$   
286 0.05). A complementary, potentially more powerful test, compared the fraction of alleles  
287 shared between species closer and further from the centre (Table 2). Importantly, this  
288 approach allowed testing for MHC introgression in the OUT zone: although other analyses  
289 showed that MHC introgression in the OUT zone was not as strong as the IN zone, they did  
290 not provide decisive evidence for or against introgression. In the IN zone, no increased allele  
291 sharing was detected closer to the centre of the L transect, but a highly significant increase  
292 was found in the R transect. In the OUT zone allele sharing increased towards the centre,  
293 providing evidence for MHC introgression (Table 2). Because both species often co-occur in  
294 the same ponds in the OUT zone, we were also able to test for increased allele sharing in  
295 syntopy, including only newts with little or no genome-wide admixture ( $< 3\%$ ). In the S

296 transect, increased allele sharing was detected only in syntopy, while in the T transect the  
297 fraction of shared alleles in allotopy close to the centre was very similar to that in syntopy,  
298 and in both transects there was significantly more allele sharing close to the centre than far  
299 from the centre (Table 2).

### 300 *Simulations*

301 Spatially explicit, individual-based forward simulations (Fig. S1) were performed to help in  
302 interpretation of the results. First, we tested whether the observed patterns of MHC similarity  
303 between species and between transects could be due to BS alone, without introgression.  
304 Second, we checked whether under BS, introgression following secondary contact and  
305 hybridization was capable of producing the observed patterns. Third, we explored the  
306 temporal and spatial dynamics of introgression under BS and compared them to neutral  
307 introgression.

308         The number of alleles maintained within species depended on the deme size and  
309 strength of selection, but the effect of intraspecific migration rate was small, except for the  
310 largest deme size (Fig. S5A). Higher migration rate increased the fraction of alleles shared  
311 between transects (Fig. S5B). We therefore fixed migration at  $Nm = 2.5$  which, by increasing  
312 similarity between transects within species, made our simulations conservative, as stronger  
313 introgression would be required for similarity between species within transect to exceed  
314 similarity between transects within species. For the same reason we used the initial number of  
315 alleles,  $n_a = 500$ , which limited similarity between species at the onset of hybridization,  
316 because a large fraction of alleles was lost from each species, independently, during evolution  
317 in isolation. Considering allele sharing and AMOVA results simultaneously, balancing  
318 selection without hybridization did not produce patterns similar to those observed (Fig. 3).

319 Most importantly, the fraction of alleles shared exclusively between transects within species  
320 was always much higher than between species within transect (Fig. 3A). AMOVA results  
321 were more complex, as the between transects within species AMOVA component could be  
322 smaller or larger than the between species component, depending on the parameter values  
323 (Fig. 3B). With increasing deme size and increasing selection both components decreased as  
324 more and more variation was distributed within populations.

325 Scenarios allowing both BS and hybridization produced patterns similar to the  
326 observed under a relatively broad range of parameter values (Fig. 4A, B). The fraction of  
327 alleles exclusively shared between species within transects increased with time of  
328 hybridization, and after several hundred generations exceeded the fraction of alleles  
329 exclusively shared between transects within species (Fig. 4A). Similarly, with increasing time  
330 of hybridization, the between transects within species AMOVA component exceeded the  
331 between species component, and, as in the observed data, the between species component was  
332 negative for the smallest deme size (Fig. 4B). Importantly, introgression of genes under BS  
333 was strong even when hybridization was too short or too limited to cause noticeable  
334 introgression of neutral genes (Fig. 4C). However, several hundreds of generations were  
335 needed for introgression to occur when the hybrid zone constituted a strong barrier (Fig. 4C).  
336 Results of simulations assuming multilocus MHC haplotypes were very similar to those for a  
337 single multiallelic locus (Fig. S6).

338



339 **Discussion**

340 *Evidence for introgression and alternative explanations*

341 This work is, to our knowledge, the first analysis of MHC introgression through hybrid zones.  
342 Introgression was detected in both zones, but its extent differed considerably. In the IN zone  
343 MHC similarity between species within transects exceeded similarity between transects  
344 within species. This implies pervasive introgression: a massive exchange of MHC alleles  
345 between species, not limited to specific haplotypes or to a restricted geographic region. The  
346 evidence is convincing because the two alternative explanations for the observed pattern –  
347 chance and parallel selection – are unlikely. The first was rejected by simulating negative  
348 frequency dependence under isolation. The other explanation would require: i) the long-term  
349 maintenance of a very large number of alleles across the ranges of both species, and ii) locally  
350 similar pathogen communities leading to parallel selection, and parallel changes in allele  
351 frequencies. Such alleles should, however, be maintained only at low frequencies, otherwise  
352 they would be detected as shared between transects within species. This is extremely unlikely,  
353 as the number of alleles maintained under BS, although substantial, is limited (Schierup,  
354 1998, our simulations). An additional argument against parallel selection is that there was  
355 only a modest increase in local allele sharing in the OUT zone.

356 *MHC differentiation within transects*

357 Despite pervasive introgression, some MHC differentiation between species was maintained  
358 within each transect. The similar location and width of the MHC and the genome-wide clines  
359 are remarkable. This apparent similarity may however mask important differences between  
360 the two clines. While there is little doubt that the genome-wide cline is secondary, i.e. has  
361 been maintained since the formation of the zones after the secondary contact, the origin of the

362 currently observed MHC clines appears more complex. At regional scales, introgression  
363 increases both MHC allele sharing and similarity of allele frequencies between species. With  
364 limited hybridization, a steep transition between the two MHC pools has probably been  
365 maintained since secondary contact, but the nature of this transition may have been evolving.  
366 First, the magnitude of differentiation has probably decreased with time since contact, but is  
367 still detectable due to limited hybridization and the excellent resolution of highly polymorphic  
368 MHC (cf. Fig. 2E, F). Second, at different times, different alleles would contribute to the  
369 differentiation. The MHC cline can thus be viewed as both secondary, maintained since  
370 contact, but also as primary, being constantly re-formed *de novo* because of a turnover of  
371 alleles responsible for the differentiation at any given moment. With time, an increased  
372 similarity of allele pools could reduce the selective advantage of introgressed alleles and, in  
373 consequence, slow down introgression. This, though, would not affect the above scenario  
374 qualitatively.

#### 375 *Differences between the two hybrid zones*

376 MHC introgression was detected in all transects, but it was more limited in the OUT zone,  
377 despite broad syntopy there. The following factors, not mutually exclusive, may differ  
378 between the zones and contribute to the observed pattern: i) frequency of hybridization; ii)  
379 age of contact; iii) strength of selection favoring MHC introgression; iv) strength of the  
380 genomic barrier to MHC introgression.

381 There could simply be too little hybridization in the OUT zone to allow extensive MHC  
382 introgression, even if favored by selection. This is supported by our simulations, and by our  
383 empirical data: the presence of admixed individuals, though rare even in syntopy, indicates  
384 non-zero recent or ongoing hybridization (Zieliński et al., 2019). Introgression from the *L.*

385 *vulgaris* lineage inhabiting the OUT zone has been strong enough to cause a regional mtDNA  
386 replacement in *L. montandoni* (Zieliński et al., 2013). Furthermore, extensive hybridization  
387 was detected in a hybrid zone in southern Poland, where the same *L. vulgaris* lineage  
388 hybridizes with *L. montandoni* (Babik et al., 2003). Thus, a negligible frequency of  
389 hybridization as the factor limiting introgression is unlikely. Analyses of mtDNA (Pabijan et  
390 al., 2017; Zieliński et al., 2013) and nuclear (Pabijan et al., 2017; Zieliński et al., 2016)  
391 sequences revealed differences in the history of hybridization between *L. montandoni* and the  
392 two *L. vulgaris* lineages. Direct evidence for longer hybridization in the IN zone comes from  
393 the observation of shorter genomic heterozygosity tracts in diagnostic markers than in the  
394 OUT zone (Zieliński et al., 2019). Within the OUT zone, MHC introgression in the more  
395 northern S transect was detectable only in syntopy, while in the more southern T transect also  
396 in parapatry. Phylogeographic evidence suggests postglacial colonization of the OUT zone by  
397 *L. vulgaris* from the south (Zieliński et al., 2013), so the contact in the S transect could be  
398 more recent. However, if the OUT zone originated during postglacial colonization several  
399 thousand generations ago, it is difficult to imagine large differences in the age of the contact  
400 between the two areas separated by ca. 100 km, and without noticeable barriers to dispersal.  
401 The differences in the age of contact in different transects, could, however, be due to  
402 anthropogenic habitat alterations during the last thousand years. It is not known how stable  
403 the locations of the zones have been, but we have not detected signs of major hybrid zone  
404 movement (cf. Wielstra et al., 2017; Zieliński et al., 2019). At first sight longer history of  
405 hybridization would make introgression easier to detect. It may, however, not necessarily be  
406 so, as alleles derived from older introgression may be lost, or have spread through the species  
407 range. The latter would enter the pool of alleles shared between species globally, not locally,  
408 and the local sharing was used to infer introgression in the IN zone. Indeed, continent-scale

409 comparisons indicate considerable MHC introgression involving the *L. vulgaris* lineages from  
410 both IN and OUT zones (Fijarczyk et al., 2018). Therefore, although old MHC introgression  
411 is likely, results from the IN zone support recent and/or ongoing introgression. Differences in  
412 the strength of selection or in the distribution of the barrier loci surrounding MHC are also  
413 plausible explanations for differences in the extent of introgression between the two hybrid  
414 zones. Unfortunately, we do not have enough information to assess the contribution of these  
415 factors.

#### 416 *The nature of selection driving introgression*

417 Pervasive MHC introgression in the face of strong genome-wide isolation implies an adaptive  
418 process. There is strong theoretical (Muirhead, 2001) and empirical (Phillips et al., 2018;  
419 Pierini & Lenz, 2018) support for novel MHC variants conferring selective advantage. Such  
420 advantage may be a major contributor to selection that can overcome strong genome-wide  
421 barriers and facilitate adaptive introgression. Adaptive introgression may, in turn, be a major  
422 contributor to the complex suite of selective pressures that drive spatial patterns of MHC  
423 diversity. For example, simple variant-preserving balancing selection predicts lower-than-  
424 neutral structure in the loci of interest, a prediction supported by studies of the S-genes of  
425 plants (Glémin et al., 2005). The MHC, in contrast, shows diverse outcomes (reviewed in  
426 Spurgin & Richardson, 2010), of which the considerable within-species differentiation by  
427 population/region in our study is an example. Also simulations of host-pathogen coevolution  
428 showed a rapid turnover of MHC alleles, not consistent with pure negative frequency  
429 dependence (Ejsmond & Radwan, 2015). Explicit geographic population structure with some  
430 degree of gene exchange may be required to reconcile these properties and processes.  
431 Therefore, to obtain a more realistic picture of selection on MHC it is necessary to model

432 MHC-pathogen coevolution in geographically structured populations (Ejmsmond, Phillips,  
433 Babik, & Radwan, 2018; Ejmsmond & Radwan, 2015; Thompson, 2005). Though the pure  
434 negative frequency dependence and spatial structure of our simulations were certainly an  
435 oversimplification, our settings were likely conservative, and probably made retention of  
436 ancestral polymorphism easier and introgression more difficult than in natural systems. We  
437 thus conclude that adaptive interspecies introgression may be an important component of the  
438 evolution of the MHC.

#### 439 *Implications*

440 Although the number of studies reporting MHC introgression is still small, several cases  
441 described from diverse taxa (Abi-Rached et al., 2011; Grossen et al., 2014; Nadachowska-  
442 Brzyska et al., 2012) suggest that the phenomenon may be widespread. Signals of  
443 introgression in MHC appears stronger than in other targets of BS (Fijarczyk et al., 2018),  
444 though such processes may be more detectable at the MHC. An exciting possibility emerges  
445 that MHC may universally be among the last genes to stop flowing between incipient species  
446 as reproductive isolation becomes complete, as suggested previously for S genes in plants  
447 (Castric, Bechsgaard, Schierup, & Vekemans, 2008). A comparative analysis of multiple  
448 vertebrate hybrid zones between strongly isolated but still hybridizing taxa could be used to  
449 test this hypothesis. If confirmed, it would support a previously suggested idea (Fijarczyk et  
450 al., 2018) of a shared pool of adaptive variation available for hybridizing species that may  
451 boost adaptive potential.

452 Widespread introgression may also contribute to an explanation of why trans-species  
453 polymorphism (TSP) seems unusually common in MHC. While TSP has traditionally been  
454 attributed to a long-term retention of ancestral polymorphism caused by balancing selection

455 (Klein, Sato, & Nikolaidis, 2007), this explanation is not entirely convincing. Both rapid  
456 turnover of MHC alleles observed in empirical surveys and recent simulation results  
457 (Ejsmond et al., 2018; Ejsmond & Radwan, 2015), suggest that long term survival of allelic  
458 lineages required to explain TSP without introgression may not be as common as previously  
459 thought.

460

#### 461 **Acknowledgments**

462 We thank S. Bury, D. Cogălniceanu, A. Fijarczyk, M. Liana, M. Niedzicka, M. Pabijan, O.  
463 Zinenko for help in sampling. A. Fijarczyk, M. Koncezal, M. Pabijan, G. Palomar, K. Phillips,  
464 J. Radwan, B. Wielstra and members of our Genomics and Experimental Evolution group  
465 provided valuable comments. This work was supported by the Polish National Centre grant  
466 2016/21/N/NZ8/00922 to K.D. and partially supported by the Polish National Centre grant  
467 2014/15/B/NZ8/00250 to P.Z and by the Jagiellonian University (DS/WB/INoS/762/18).

468

#### 469 **References**

- 470 Abbott, R., Albach, D., Ansell, S., Arntzen, J. W., Baird, S. J. E., Bierne, N., . . . Zinner, D.  
471 (2013). Hybridization and speciation. *Journal of Evolutionary Biology*, 26(2), 229-  
472 246.
- 473 Abi-Rached, L., Jobin, M. J., Kulkarni, S., McWhinnie, A., Dalva, K., Gragert, L., . . .  
474 Parham, P. (2011). The shaping of modern human immune systems by multiregional  
475 admixture with archaic humans. *Science*, 334(6052), 89-94.
- 476 Alexander, D. H., Novembre, J., & Lange, K. (2009). Fast model-based estimation of ancestry  
477 in unrelated individuals. *Genome Research*, 19(9), 1655-1664.
- 478 Babik, W., Szymura, J. M., & Rafiński, J. (2003). Nuclear markers, mitochondrial DNA and  
479 male secondary sexual traits variation in a newt hybrid zone (*Triturus vulgaris* x *T.*  
480 *montandoni*). *Molecular Ecology*, 12(7), 1913-1930.
- 481 Barton, N. H. (1979). Gene flow past a cline. *Heredity*, 43(3), 333-339.
- 482 Castric, V., Bechsgaard, J., Schierup, M. H., & Vekemans, X. (2008). Repeated adaptive  
483 introgression at a gene under multiallelic balancing selection. *PLoS Genetics*, 4(8),  
484 e1000168.
- 485 Coyne, J. A., & Orr, H. A. (2004). *Speciation*. Sunderland: Sinauer.

- 486 Currat, M., Gerbault, P., Di, D., Nunes, J. M., & Sanchez-Mazas, A. (2015). Forward-in-time,  
487 spatially explicit modeling software to simulate genetic lineages under selection.  
488 *Evolutionary Bioinformatics*, *11*, EBO. S33488.
- 489 Derryberry, E. P., Derryberry, G. E., Maley, J. M., & Brumfield, R. T. (2014). HZAR: hybrid  
490 zone analysis using an R software package. *Molecular Ecology Resources*, *14*(3), 652-  
491 663.
- 492 Ejsmond, M. J., Phillips, K. P., Babik, W., & Radwan, J. (2018). The role of MHC supertypes  
493 in promoting trans-species polymorphism remains an open question. *Nature*  
494 *Communications*, *9*(1), 4362.
- 495 Ejsmond, M. J., & Radwan, J. (2015). Red Queen processes drive positive selection on major  
496 histocompatibility complex (MHC) genes. *PLOS Comput. Biol.*, *11*(11), e1004627.
- 497 Excoffier, L., & Lischer, H. E. (2010). Arlequin suite ver 3.5: a new series of programs to  
498 perform population genetics analyses under Linux and Windows. *Molecular Ecology*  
499 *Resources*, *10*(3), 564-567.
- 500 Falush, D., Stephens, M., & Pritchard, J. K. (2007). Inference of population structure using  
501 multilocus genotype data: dominant markers and null alleles. *Molecular Ecology*  
502 *Notes*, *7*(4), 574-578.
- 503 Fijarczyk, A., & Babik, W. (2015). Detecting balancing selection in genomes: limits and  
504 prospects. *Molecular Ecology*, *24*(14), 3529-3545.
- 505 Fijarczyk, A., Dudek, K., Niedzicka, M., & Babik, W. (2018). Balancing selection and  
506 introgression of new immune-response genes. *Proceedings of the Royal Society of*  
507 *London B: Biological Sciences*, *285*(1884), 20180819.
- 508 Fraïsse, C., Belkhir, K., Welch, J. J., & Bierne, N. (2016). Local interspecies introgression is  
509 the main cause of extreme levels of intraspecific differentiation in mussels. *Molecular*  
510 *Ecology*, *25*(1), 269-286.
- 511 Glémin, S., Gaude, T., Guillemin, M.-L., Lourmas, M., Olivieri, I., & Mignot, A. (2005).  
512 Balancing selection in the wild: testing population genetics theory of self-  
513 incompatibility in the rare species *Brassica insularis*. *Genetics*, *171*(1), 279-289.
- 514 Gompert, Z., Mandeville, E. G., & Buerkle, C. A. (2017). Analysis of population genomic  
515 data from hybrid zones. *Annual Review of Ecology, Evolution, and Systematics*, *48*,  
516 207-229.
- 517 Grossen, C., Keller, L., Biebach, I., Croll, D., & Consortium, I. G. G. (2014). Introgression  
518 from domestic goat generated variation at the major histocompatibility complex of  
519 alpine ibex. *PLoS Genetics*, *10*(6), e1004438.
- 520 Harrison, R. G., & Larson, E. L. (2016). Heterogeneous genome divergence, differential  
521 introgression, and the origin and structure of hybrid zones. *Molecular Ecology*,  
522 *25*(11), 2454-2466.
- 523 Hedrick, P. W. (2013). Adaptive introgression in animals: examples and comparison to new  
524 mutation and standing variation as sources of adaptive variation. *Molecular Ecology*,  
525 *22*(18), 4606-4618.
- 526 Huerta-Sánchez, E., Jin, X., Bianba, Z., Peter, B. M., Vinckenbosch, N., Liang, Y., . . . Ni, P.  
527 (2014). Altitude adaptation in Tibetans caused by introgression of Denisovan-like  
528 DNA. *Nature*, *512*(7513), 194.
- 529 Kelley, J., Walter, L., & Trowsdale, J. (2005). Comparative genomics of major  
530 histocompatibility complexes. *Immunogenetics*, *56*(10), 683-695.
- 531 Kisel, Y., & Barraclough, T. G. (2010). Speciation has a spatial scale that depends on levels  
532 of gene flow. *the american naturalist*, *175*(3), 316-334.



- 533 Klein, J., Sato, A., & Nikolaidis, N. (2007). MHC, TSP, and the origin of species: From  
534 immunogenetics to evolutionary genetics. *Annual Review of Genetics*, *41*, 281-304.
- 535 Meier, J. I., Marques, D. A., Mwaiko, S., Wagner, C. E., Excoffier, L., & Seehausen, O.  
536 (2017). Ancient hybridization fuels rapid cichlid fish adaptive radiations. *Nature*  
537 *Communications*, *8*, 14363.
- 538 Muirhead, C. A. (2001). Consequences of population structure on genes under balancing  
539 selection. *Evolution*, *55*(8), 1532-1541.
- 540 Nadachowska-Brzyska, K., Zielinski, P., Radwan, J., & Babik, W. (2012). Interspecific  
541 hybridization increases MHC class II diversity in two sister species of newts.  
542 *Molecular Ecology*, *21*(4), 887-906.
- 543 Niedzicka, M., Dudek, K., Fijarczyk, A., Zieliński, P., & Babik, W. (2017). Linkage map of  
544 *Lissotriton* newts provides insight into the genetic basis of reproductive isolation. *G3:  
545 Genes, Genomes, Genetics*, *7*(7), 2115-2124.
- 546 Pabijan, M., Zielinski, P., Dudek, K., Stuglik, M., & Babik, W. (2017). Isolation and gene  
547 flow in a speciation continuum in newts. *Molecular Phylogenetics and Evolution*, *116*,  
548 1-12.
- 549 Payseur, B. A. (2010). Using differential introgression in hybrid zones to identify genomic  
550 regions involved in speciation. *Molecular Ecology Resources*, *10*(5), 806-820.
- 551 Phillips, K. P., Cable, J., Mohammed, R. S., Herdegen-Radwan, M., Raubic, J., Przesmycka,  
552 K. J., . . . Radwan, J. (2018). Immunogenetic novelty confers a selective advantage in  
553 host-pathogen coevolution. *Proceedings of the National Academy of Sciences*, *115*,  
554 1552-1557.
- 555 Pierini, F., & Lenz, T. L. (2018). Divergent allele advantage at human MHC genes: signatures  
556 of past and ongoing selection. *Molecular Biology and Evolution*, *35*, 2145-2158.
- 557 Schierup, M. H. (1998). The number of self-incompatibility alleles in a finite, subdivided  
558 population. *Genetics*, *149*(2), 1153-1162.
- 559 Schierup, M. H., Vekemans, X., & Charlesworth, D. (2000). The effect of subdivision on  
560 variation at multi-allelic loci under balancing selection. *Genetical Research*, *76*(1), 51-  
561 62.
- 562 Sebastian, A., Herdegen, M., Migalska, M., & Radwan, J. (2016). amplisas: a web server for  
563 multilocus genotyping using next-generation amplicon sequencing data. *Molecular  
564 Ecology Resources*, *16*(2), 498-510.
- 565 Spurgin, L. G., & Richardson, D. S. (2010). How pathogens drive genetic diversity: MHC,  
566 mechanisms and misunderstandings. *Proceedings of the Royal Society B: Biological  
567 Sciences*, *277*(1684), 979-988.
- 568 Thompson, J. N. (2005). *The geographic mosaic of coevolution*. Chicago: University of  
569 Chicago Press.
- 570 Wielstra, B., Burke, T., Butlin, R. K., Avci, A., Üzümlü, N., Bozkurt, E., . . . Arntzen, J. W.  
571 (2017). A genomic footprint of hybrid zone movement in crested newts. *Evolution  
572 Letters*, *1*(2), 93-101.
- 573 Zieliński, P., Dudek, K., Arntzen, J. W., Palomar, G., Niedzicka, M., Fijarczyk, A., . . . Babik,  
574 W. (2019). Differential introgression across newt hybrid zones - evidence from  
575 replicated transects. *Molecular Ecology*, *submitted*.
- 576 Zieliński, P., Nadachowska-Brzyska, K., Dudek, K., & Babik, W. (2016). Divergence history  
577 of the Carpathian and smooth newts modelled in space and time. *Molecular Ecology*,  
578 *25*(8), 3912-3928.
- 579 Zieliński, P., Nadachowska-Brzyska, K., Wielstra, B., Szkotak, R., Covaciu-Marcov, S. D.,  
580 Cogălniceanu, D., & Babik, W. (2013). No evidence for nuclear introgression despite



581 complete mtDNA replacement in the Carpathian newt (*Lissotriton montandoni*).  
582 *Molecular Ecology*, 22(7), 1884-1903.

583

#### 584 **Data Accessibility**

585 Individual genotypes and sequences of all MHC alleles will be deposited in Dryad upon  
586 acceptance.

587

#### 588 **Author contributions**

589 K.D, T.S.G. and W.B. designed research, K.D. collected the data, K.D. P.Z., and W.B.  
590 analyzed the data, T.S.G. and W.B. designed simulations, T.S.G. performed simulations,  
591 W.B. and K.D wrote the paper. All authors read and approved the final version.

592

593 **Tables and Figures**

594

595 **Table 1. Analysis of Molecular Variance.** Percentages of SNP and MHC variation explained  
 596 by hierarchical levels of population structure are given. Syntopic populations were excluded  
 597 from the analysis.

598 <sup>1</sup>significance of the between species component could not be tested because only two transects were available in each region  
 599

A. AMOVA for each zone: MHC and SNP						
	IN			OUT		
	MHC	SNP		MHC	SNP	
Between species	-1.19	50.13	<sup>1</sup>	11.61	68.79	<sup>1</sup>
Between transects within species	7.70	13.18	***	2.44	1.99	***
Within transects	93.49	36.69	***	85.94	29.22	***

B. AMOVA for each transect: MHC						
	IN				OUT	
	L		R		S	T
Between species	6.66	***	3.55	***	15.01	**
Between populations within species	2.74	***	6.16	***	0.81	**
Within populations	90.6	***	90.29	***	84.18	***

600

601

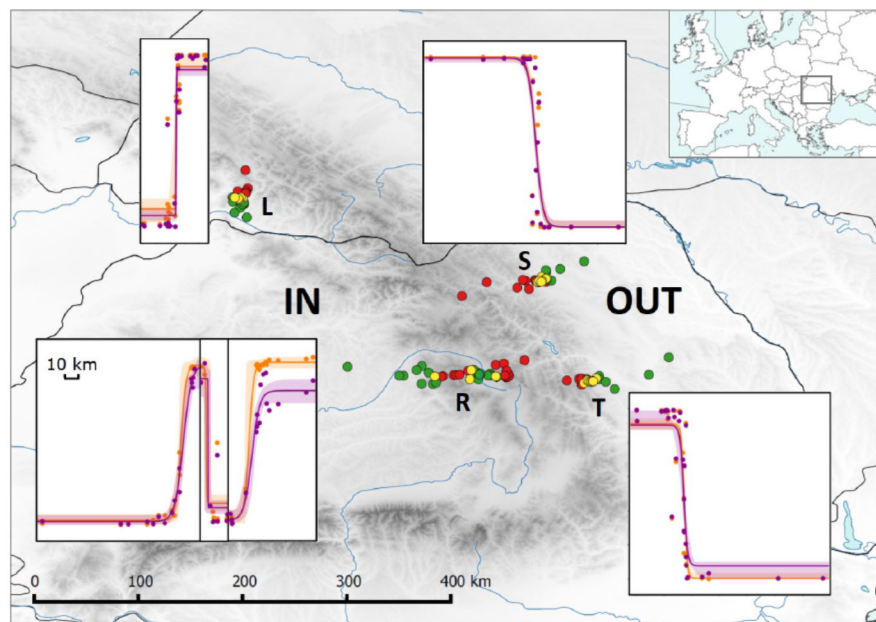
602 **Table 2. Allele sharing between species closer and farther from the centre of the hybrid**  
 603 **zone.** For each comparison gClose is the group closer and gFar is the group farther from the  
 604 centre (Syntopy < Parapatry < Allopatry). In syntopic populations individuals with > 3%  
 605 genome-wide admixture were excluded from analysis. One sided *P* values (sharing gClose >  
 606 sharing gFar, 10 000 permutations of individuals within species between groups) are reported.  
 607 Lm – *Lissotriton montandoni*, Lv – *L. vulgaris*, Allo – allopatry, Para – parapatry, Syn –  
 608 syntopy, Tr – transect.  
 609

Zone	Tr	Compared groups		Percent alleles shared		<i>P</i>	Number of MHC alleles		Sample size			
		gClose	gFar	gClose	gFar		gClose	gFar	Lm gClose	Lm gFar	Lv gClose	Lv gFar
IN	L	Para	Allo	24.2	19.2	0.80	413	328	145	33	82	33
IN	R	Para	Allo	39.1	18.3	<0.0001	529	290	222	23	201	25
OUT	S	Para	Allo	17.3	13.7	0.23	324	299	102	48	41	19
OUT	S	Syn	Allo	25.5	13.7	0.0090	271	299	40	48	52	19
OUT	S	Syn	Para	25.5	17.3	0.0092	271	324	40	102	52	41
OUT	T	Para	Allo	26.8	11.0	0.0026	399	218	148	15	62	16
OUT	T	Syn	Allo	26.9	11.0	0.0009	349	218	80	15	60	16
OUT	T	Syn	Para	26.9	26.8	0.74	349	399	80	148	60	62

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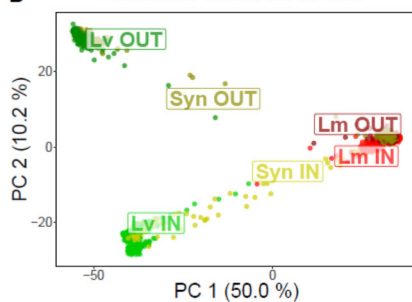
613 **Fig. 1. Sampling, clines and differentiation.** A) Sampling and clines. Location of the two  
 614 hybrid zones (IN, OUT), the four transects (L, R, S, T) and localities within transects are  
 615 shown: green – *L. vulgaris*, red – *L. montandoni*, yellow – syntopic *L. montandoni* & *L.*  
 616 *vulgaris*. For each transect genome-wide (orange, mean population Q value from Admixture  
 617 based on 2849 SNP) and MHC (purple, mean population Q value from Structure based on  
 618 MHC alleles coded as dominant markers) clines are shown. Note the compound nature of the  
 619 R transect, which is composed of three contact zones. B) Principal Component (PCA)  
 620 ordination of individuals based on genome-wide SNP data, light green – *L. vulgaris* inside the  
 621 Carpathian Basin (IN zone), dark green – *L. vulgaris* outside the Carpathian Basin (OUT  
 622 zone), light red – *L. montandoni* IN, dark red – *L. montandoni* OUT, yellow – individuals  
 623 from syntopic populations IN, olive – individuals from syntopic populations OUT; C) PCA  
 624 based on presence/absence of MHC class I and II alleles (individuals from syntopic  
 625 populations were excluded); D) Multidimensional scaling ordination of populations based on  
 626 pairwise  $F_{ST}$  in MHC; populations from a single transect are indicated with the same letter.  
 627

A



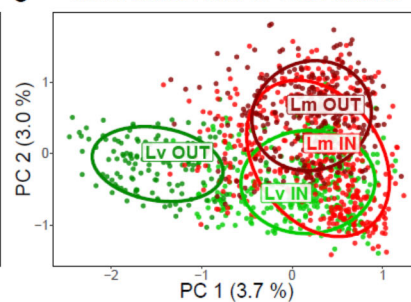
B

Genome-wide: 2849 SNP



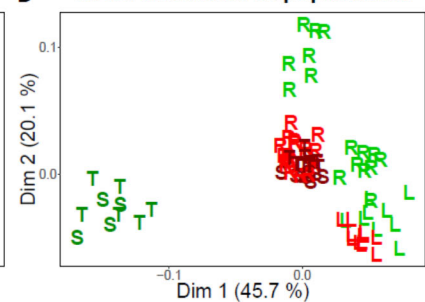
C

MHC class I and II: 1062 alleles



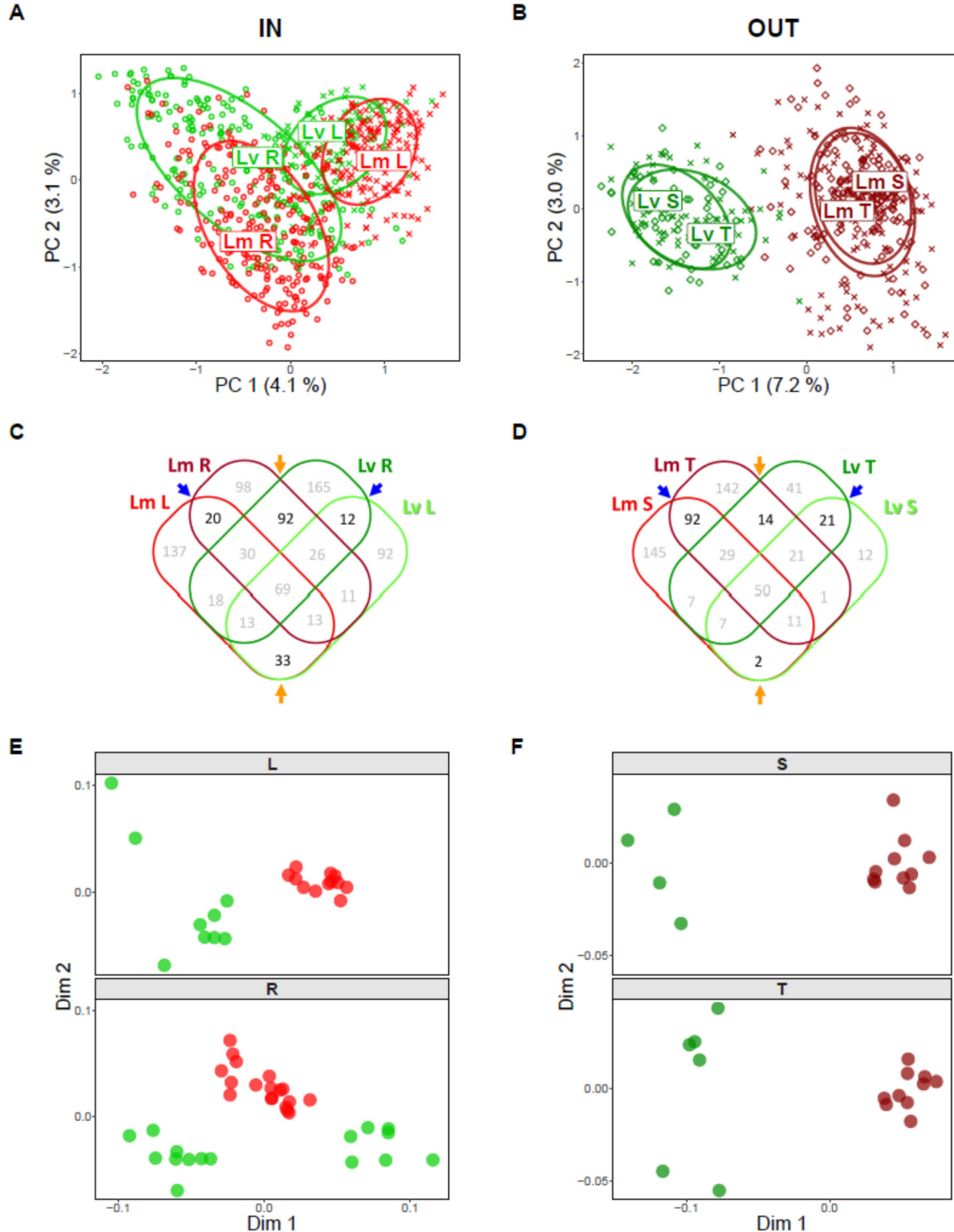
D

MHC: ordination of populations



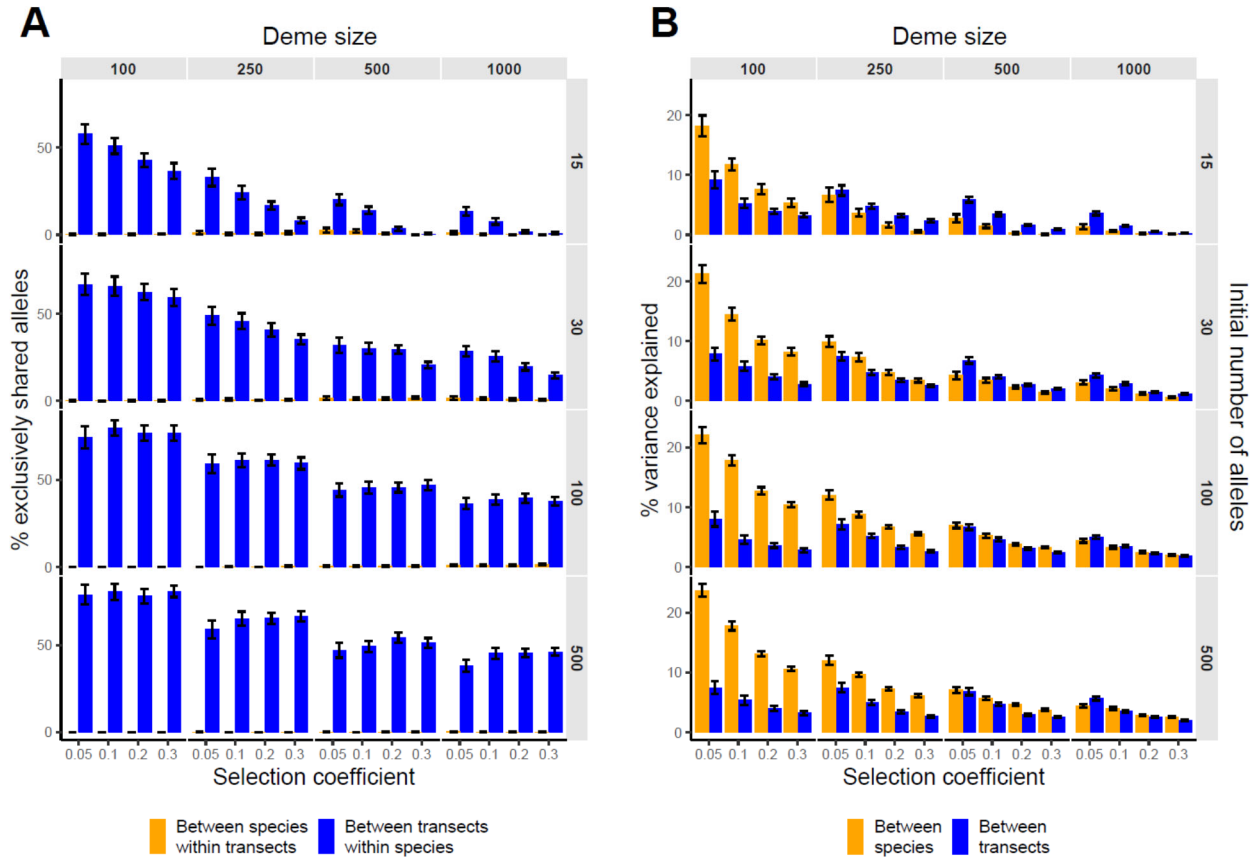
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631 Fig. 2. **MHC differentiation between species and transects in the IN (A, C, E) and OUT (B, D, F) zones.** A) and B) PCA based on presence/absence of MHC class I and II alleles,  
 632 transects within zones indicated with ellipses and different symbols. C) and D) Venn  
 633 diagrams showing in black the number of alleles shared between species within transects  
 634 (orange arrows) and between transects within species (blue arrows). E) and F)  
 635 Multidimensional scaling ordination of populations within transects based on pairwise  $F_{ST}$  in  
 636 MHC.  
 637



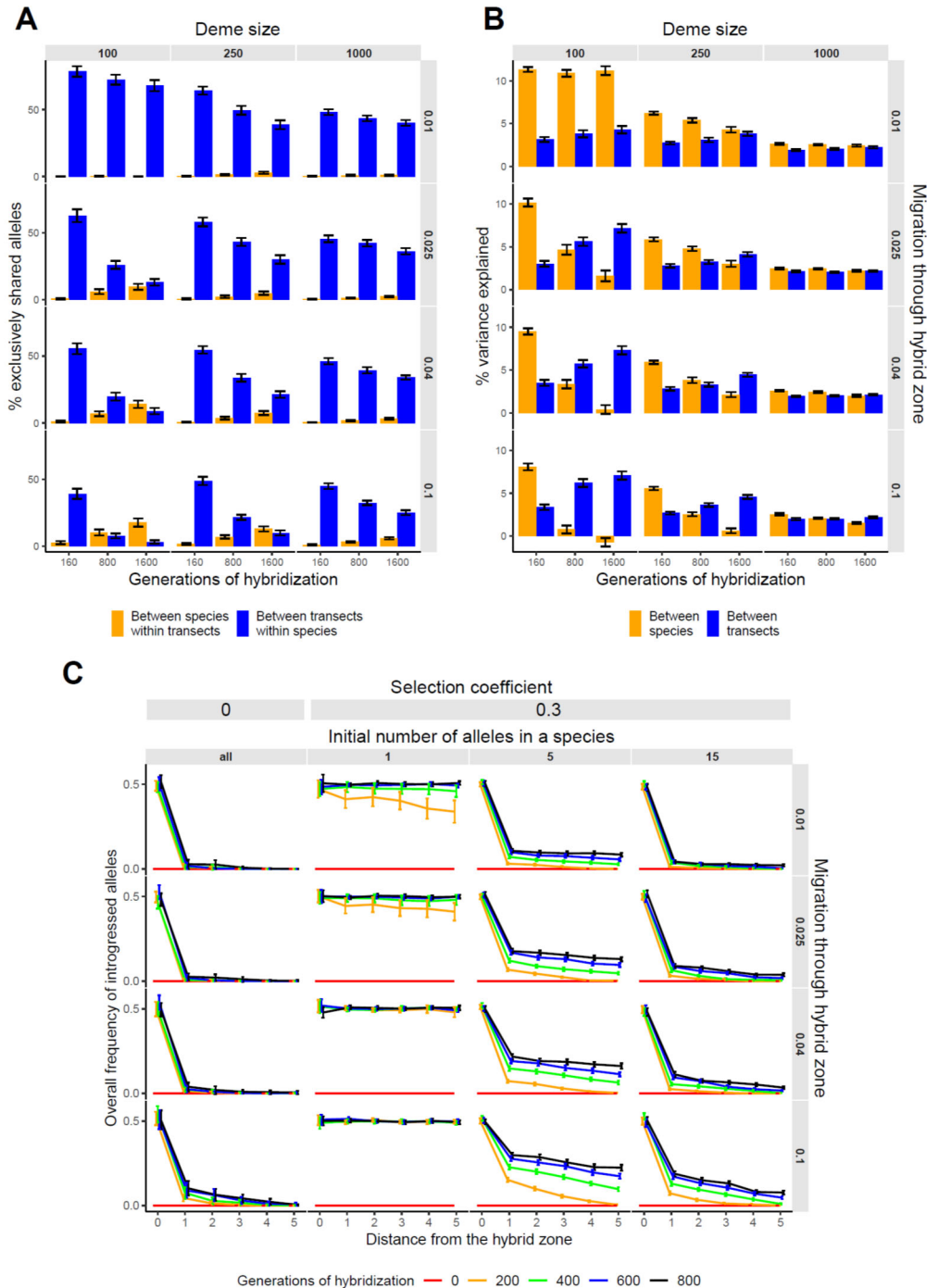
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639 Fig. 3. **Simulations: Isolation.** In all simulations migration rate between adjacent demes  
 640 within species was  $Nm = 2.5$ . A) Percentage of alleles shared exclusively (those shared by  
 641 two focal groups but absent from all other groups) between species within transect and  
 642 between transects within species, B) Percentage of total variance explained by between  
 643 species and between transect within species AMOVA components. Means and 95%  
 644 confidence intervals from 50 simulations are shown.



645  
 646

647 Fig. 4. **Simulations: Hybridization.** In all simulations migration rate between adjacent demes  
 648 within species was  $Nm = 2.5$ . A) Percentage of alleles shared exclusively (those shared by  
 649 two focal groups but absent from all other groups) between species within transect and  
 650 between transects within species, B) Percentage of total variance explained by between  
 651 species and between transect within species AMOVA components. C) Dynamics of  
 652 introgression in a single transect – frequency of introgressed alleles along the transect at  
 653 different times after hybridization. As expected, identical results were obtained for neutral  
 654 introgression, regardless of the initial number of alleles (all). Means and 95% confidence  
 655 intervals from 50 simulations are shown.



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