Out of the blue: Phototropins of the leaf vascular bundle sheath mediate the regulation of leaf hydraulic conductance by blue light

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Short title: Blue-light regulation of leaf hydraulics

One-Sentence summary: Blue light regulates the leaf hydraulic conductance via the bundle-sheath cells' blue light PHOT receptors which, via an autonomous signaling pathway, activate the BSCs' AHA2 H+-pump.

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ABSTRACT

The leaf veins bundle sheath cells (BSCs) constitute a selective barrier to water and solutes entering the mesophyll and play a major role in regulating the leaf radial hydraulic conductance (K_{leaf}). BSCs' plasma membrane H^+-ATPase, AHA2, increases K_{leaf} by acidifying the xylem sap. We hypothesized, based on the expression of PHOT1 and PHOT2 in Arabidopsis BSCs and on the known blue-light (BL)-induced increase of K_{leaf}, that similar to the guard cells BL signal transduction pathway, BL perception by the BSCs' phots activates its H^+-ATPase and upregulate K_{leaf}. Indeed, under BL, the K_{leaf} of the knockout mutant lines *phot1-5*, *phot2-1*, *phot1-5phot2-1*, and *aha2-4* was lower than that of the WT. BSC-only-directed complementation of *phot1-5* or *aha2-4* by *PHOT1* and *AHA2*, respectively, restored the BL-induced increase in K_{leaf}. BSC-specific silencing of *PHOT1* or *PHOT2* prevented such K_{leaf} increase, and we replicated this in WT plants with a xylem-fed kinase inhibitor (tyrphostin 9). In the *PHOT1*-complemented *phot1-5*, as in WT, but unlike in *phot1-5*, white light acidified the xylem sap (relative to darkness). These results, supported by assays of BSC protoplasts' water permeability,

hyperpolarization and cytosolic pH, highlight the BSCs' role in the BL-K_{leaf} link as mediators that regulate the xylem-to-mesophyll water flux.

Introduction

Light, the energy source for photosynthesis, also regulates plant growth and development (Kronenberg and Kendrick, 1986) as well as physiological traits such as stomatal conductance (g_s; Zeiger and Helper, 1977; Karlsson, 1986; Talbott et al., 2003) and leaf hydraulic conductance (K_{leaf}; Voicu et al., 2008; Ben Baaziz et al., 2012; Aasamaa and Sõber, 2012; Prado and Maurel, 2013). One of the most conserved and well-studied light-sensing mechanisms is the sensing of blue light (BL, 390–550 nm) that causes stomata to open (Grondin et al., 2015). In this signal transduction pathway in the guard cells (GCs), BL is perceived by the protein kinases phot1 and phot2 (Kinoshita et al., 2001). Their immediate phosphorylation substrate is BLUS1 (Blue Light Signaling1, At4g14480; Takemiya et al., 2013), which is a part of the earliest stomata-opening BL signaling complex with phot. Another part of this complex is the Raf-like kinase, BHP (BL-dependent H^+-ATPase phosphorylation, At4g18950; Hayashi et al., 2017; reviewed by Inoue and Kinoshita, 2017). While none of these directly phosphorylates the guard-cell H^+-ATPase (Hayashi et al., 2017), one of the other Raf-like kinases downstream of phot could be the target of tyrphostin 9, a kinase inhibitor that has been shown to inhibit the BL-induced phosphorylation of the penultimate threonine of the plasma membrane H^+-ATPase (AHA) in Arabidopsis GCs (ibid). This BL-induced phosphorylation activates the AHA (Kinoshita and Shimazaki, 1999; Svennelid et al., 1999), which, in turn, hyperpolarizes the GCs and acidifies their surrounding apoplast (Kinoshita and Shimazaki, 1999; Ueno et al., 2005; Den Os et al., 2007; Elmore and Coaker, 2011, reviewed by Inoue and Kinoshita, 2017). Conversely, at night, in the dark, the guard cells' H^+-ATPases are deactivated, the GCs are depolarized, and their apoplast becomes alkaline (Kinoshita et al., 2001; Kinoshita and Shimazaki, 1999). These pH and membrane potential changes cause ion and water fluxes across the GCs membranes causing their osmotic swelling during the day and shrinking at night, which underlies stomata opening and closure (Assmann, 1993; Roelfsema and Hedrich, 2005; Shimazaki et al., 2007).

Both BL and white light (WL) have been shown to increase the hydraulic conductance (K_{leaf}) of the entire leaf in several plant species [(Voicu et al., 2008 (bur oak, *Quercus macrocarpa*); Voicu et al., 2009; Sellin et al., 2011 (silver birch, *Betula pendula* Roth); Aasamaa and Sber, 2012 (deciduous trees); Ben Baaziz et al., 2012 (walnut, *Juglans regia* L.)]. Interestingly, K_{leaf} responds more quickly to light than g_S does (Guyot

et al., 2012). However, the molecular mechanism by which K_{leaf} increases in response to light quality is not yet fully understood.

In the past decade, it has been established that bundle sheath cells (BSCs), which form a parenchymal layer that tightly enwraps the entire leaf vasculature, can act as a dynamic and selective xylem-mesophyll barrier to water and ions, and participate in K_{leaf} regulation (Shapira et al., 2009; Shatil-Cohen and Moshelion, 2012). Moreover, we recently discovered that the BSCs' H^+-ATPase proton pump, AHA2, participates in regulating K_{leaf} via changes in the xylem pH and that the positive correlation observed between AHA2-driven xylem acidification and K_{leaf} is due to an increase in the osmotic water permeability of BSCs' membranes (Grunwald et al., 2021). AHA2 is particularly abundant in BSCs (present at a >3-fold higher level than that seen in neighboring mesophyll cells; Wigoda et al., 2017) which explains how such acidification is possible. In addition, the same BSC transcriptome analysis (ibid.) revealed substantial expression of the BL receptors phot1 and phot2 in BSCs, as well as substantial expression of the kinases BHP and BLUS1. These findings led us to hypothesize that the vascular BSCs possess a BL signal transduction pathway that is similar to the one that opens stomata and that plays a role in the regulation of K_{leaf}. In confirmation of this hypothesis, we describe here a few of the physiological and molecular mechanisms underlying the blue light-dependent regulation of K_{leaf} by BSCs.

Materials and Methods

Plant material

Plant genotypes. We used WT (wild type) *Arabidopsis thaliana* plants ecotype Colombia, Col-0 (aka Col), the T-DNA insertion AHA2 mutants *aha2-4* (SALK_ 082786) (Col) and *aha2-4* complemented with *SCR: AHA2* (line 56) (Col), and *SCR:GFP* (Col) as described previously (Grunwald et al., 2021). In addition, we used WT Arabidopsis (Col) with the glabrous mutation (WT Col-gl), *phot1-5* (*nph1-5*) (Col-gl), *phot 2-1* (*npl1-1* or *cav1-1*) (Col-gl) and the double mutant *phot1-5phot2-1* (*nph1-5 npl1-1*) (Col-gl) (Kagawa et al., 2001), which were kindly provided by the Ken-Ichiro Shimazaki lab (Tokyo, Japan). The *blus1-4* (Col) seed was a kind gift from the Takemiya lab (Yamaguchi, Japan).

Construction of SCR:amiR-PHOT1 plants. The premiR-*PHOT1*or *PHOT2* and synthetic genes were synthesized by Hylabs (Rehovot, Israel), based on a premiR164 backbone (Alvarez et al., 2006). We used the Web-based MicroRNA Designer (WMD,

http://wmd3.weigelworld.org) to produce a premiRNA gene MIR319a as described in WMD. After sequence verification, the premiR-*PHOT1*or premiR-*PHOT2* was cloned into the pDONR™ 221 and the SCR promoter into pDONRP4P1r (Invitrogen) vectors which are Gateway® compatible by BP reactions, and later cloned into the pB7M24GW (Invitrogen) two-fragment binary vector using an LR r reaction, according to the manufacturer's instructions. Each binary vector was transformed into *Agrobacterium tumefaciens* by electroporation and transformed into WT Col-0 using the floral-dip method (Clough and Bent, 1998). Transformants were selected based on their resistance to BASTA herbicide (glufosinate ammonium, Sigma cat # 45520), grown on plates with MS (Murashige and Skoog, Duchefa cat# M222.0050) basal medium + 1 % sucrose and 20 µg/ml BASTA. DNA insertion was verified in selected lines by PCR targeting the junction of the premiR-gene and the 35S terminator with the forward primer located about 1000 bp from the 3' end of premiR-gene and reverse primer on the 35S terminator (see primer list in Supplemental Table S1). PCR fragments were then sequenced and verified.

Construction of SCR: PHOT1 plants. Binary vectors were constructed with the *PHOT1* gene as described above and then transformed into *phot1-5* (Col-gl1) plants. Successful transformation was verified by PCR.

Plant growth conditions. Plants were grown in 250 ml pots, 2-3 plants per pot filled with soil (Klasmann686, Klasmann-Deilmann, Germany) + 4g/L Osmocote® 6M in a growth chamber kept at 22°C and 70% relative humidity, with an 8-h light (9:00 am–5:00 pm) /16-h dark photoperiod. During the day, the illumination, humidity and vapor pressure deficit (VPD) changed in a pattern resembling natural conditions, as in (Negin and Moshelion, 2017). The illumination intensity provided by LED lights strips [EnerLED 24 V-5630, 24 W/m, 3000 K (50%)/6000 K (50%)] reached 150–200 µmol m^{-2} sec^{-1} light at the plant level at midday. The plants were irrigated twice a week.

Preparation of detached leaves for gas-exchange measurements. Fully expanded leaves from 7- to 8-week-old plants were excised at the petiole base using a wet, sharp blade under a drop of water. Petioles were immediately submerged in 0.5-ml Eppendorf tubes with artificial xylem solution [AXS; 3 mM KNO_3, 1 mM $Ca(NO_3)_2$, 1 mM $MgSO_4$, 3 mM $CaCl_2$, 0.25 mM NaH_2PO_4, 90 µM EDFS (Sigma)]. The leaves were excised, shortly before the "lights off" transition (around 5:00 pm) on the evening preceding the measurements and placed in gas-sealed, transparent 25 cm × 20 cm × 15 cm plastic boxes with water-soaked tissue paper on the bottom to provide ~90% humidity. The transparent boxes were then

inserted into a larger light-proof plastic box overnight and kept in total darkness until the start of light treatments the next morning.

Light treatments of detached leaves

All gas-exchange experiments were conducted in a dark room (<1 umol m^{-2} s^{-1}) at a constant temperature of 22°C, between 10:00 am and 1:00 pm. The excised leaves were taken out of the light-sealed box and placed in one of two custom-made light chambers. The total illumination intensity in both light chambers was set to roughly 220 µmol m^{-2} s^{-1}. In the red-light chamber, the leaves were illuminated only with red light (RL, 660 nm) and in the RL+BL chamber they were illuminated with approximately 90% RL and 10% BL (450 nm). Leaves were exposed to either RL or RL+BL for 15 min at an ambient VPD of 1.3–1.5 kPa. A fan circulated the air in each light chamber for uniform and constant temperature and VPD. Next, leaves were transferred to a LI-COR 6400XT for an additional 2–3 min, under the same illumination conditions with VPD of about 1.3kPa, until stabilization of the gas-exchange measurements, after which a measurement was recorded. D-LED lighting with adjustable current source (DR-SD6/12) was used as light sources (http://www.d-led.net/). Light intensity was verified daily using a LI-250A light meter with a LI-190SA quantum sensor (LI-COR, Lincoln, NE, USA).

Determination of gas exchange, E and g_S, and hydraulic conductance, K_{leaf}

Following the light treatments, the leaves were immediately placed in a LI-COR 6400XT for measurements of stomatal conductance (g_S) and transpiration (E) similar to Grunwald et al. (2021), with the following changes: All of the experiments were conducted in the dark at 22°C and the illumination in the LI-COR cuvette was adjusted to match the preceding light treatments. Then, the water potential (Ψ_{leaf}) was determined in a pressure chamber (ARIMAD-3000; MRC Israel) and K_{leaf} was calculated (as in Grunwald et al., 2021) as:

Eq. 1: $K_{leaf} = -E / (\mathbf{\Psi}_{leaf} - \mathbf{\Psi}_{AXS}) \approx -E / \mathbf{\Psi}_{leaf}$

where AXS denotes artificial xylem sap (solution).

Additionally, g_S was measured at the same time as above using LI-COR 6400XT on intact leaves of whole plants left under the growth-room illumination for 2–4 h after the

lights were turned on. Illumination settings in the instrument's cuvette were set to the same conditions as in the growth room.

Inhibition of light transduction in detached leaves

Tyrphostin 9 (ENZO, cat. # BML-EI21, 100 mM in DMSO (Sigma, cat # W387520), kept in aliquots at -18 °C) was added to the AXS to a final concentration of 10 μM. AXS with DMSO at a concentration of 100 μl/L (vol./vol.) served as a control in this set of experiments. Excised leaves were kept overnight until the measurements as described above. For surface application, 0.05% Tween 20 (J.T. Baker cat# X251-07) was added to the tyrphostin 9 and control solutions and sprayed prior to overnight perfusion with unmodified AXS. Boxes holding the sprayed leaves did not contain the damp tissue paper, and leaves were patted dry prior to placing them in the light chamber to ensure the leaf surface was dry when the leaves were placed in the LI-COR 6400xt cuvettes.

Determination of xylem sap pH in detached leaves

Leaf preparation for imaging. On the eve of the experiment, leaves from 6- to 7-week-old plants, approximately 2.5 cm long and 1 cm wide, were excised with a sharp blade and perfused (as described) with AXS containing 100 μM of the dual-excitation fluorescent pH probe, FITC-D [fluorescein isothiocyanate conjugated to 10 kD dextran, (Sigma cat. # FD10S), added from a 10 mM stock in double-distilled water]. The excised leaves were then placed in the "double box" setup (i.e., a sealed plastic box inside a dark box) and kept in the dark until measurements the next morning.

Dark-treated leaves: Leaves were taken out of the dark boxes and immediately prepared for vein imaging on a microscope slide under low light conditions (<5 μmol m^{-2} sec^{-1}).

Light-treated leaves: Leaves were taken out of the dark boxes and placed in the growth chamber inside transparent gas sealed boxes (boxes as in K_{leaf}) for 30 min of growth room white-light illumination (see above under "Plant growth conditions") and were subsequently imaged.

The leaves were imaged using an inverted microscope (Olympus-IX8, as detailed by Grunwald et al., 2021). Image capture and image analysis of the intra-xylem pH were as already described (ibid.).

Determination of the BSCs' membrane potential (E_M)

Protoplasts preparation. To evaluate the protoplast membrane potential (E_M), we used a fluorescent (E_M-sensitive dual-excitation ratiometric dye, Di-8-ANEPPS (Pucihar et al., 2009), and an inverted epifluorescence microscope (Eclipse Ti-S, Nikon, Tokyo, Japan) coupled to an IXON Ultra 888 camera (Andor, UK). The experiments were performed between 10 am and 4 pm. BSC protoplasts were isolated as described (Shatil-Cohen et al., 2014) from 6- to 8-week-old SCR:GFP (Col) plants and kept in an Eppendorf vial at room temperature (22 ± 2°C) in the dark until use. A drop with a few protoplasts was added to a bath solution in an experimental chamber, containing 30 μM di-8-ANEPPS and 1% pluronic acid, with or without 10 μM tyrphostin 9 (see "Solutions" below), and protoplasts were allowed to settle for 10 min on a glass coverslip bottom. A Nikon 40X objective (Plan Fluor 40x/ 0.75, OFN25 DIC M/N2) was used for protoplast viewing and imaging.

GFP-based selection. An individual, perfectly round BSC (25–32.5 μm diameter) was selected for imaging based on its GFP fluorescence (excitation of 490/15 nm) delivered by a xenon lamp monochromator, Polychrome II (Till Photonics, Munich, Germany), under the control of IW6.1 (Imaging Workbench 6.1) software (Indec Biosystems, Santa Clara, CA), and the emitted fluorescence was filtered via a 515 nm dichroic mirror and a 525/50 nm band-pass barrier filter.

Size-based selection. In a single set of experiments, we used a *phot1-5* (Col-gl) mutant and WT (Col-gl) plants that were not labeled with GFP. BS protoplasts are about 50% smaller than mesophyll cells and contains about 50% chloroplast of compared to the mesophyll cells (Shatil-Cohen et al., 2011) and are relatively easy to detect under transmitted light. Thus, we based the BSCs selection on rejecting as "non-BSCs" protoplasts outside the above-mentioned size range (25–32.5 mm diameter). In a separately conducted experiment on protoplasts from GFP-labeled WT (Col), such size-based guesses were confirmed by GFP fluorescence in 68 out of 101 cases.

Light treatments and imaging. The selected BSC protoplast in the experimental chamber was exposed to the dark for 10-20 min (control), and a first pair of di-8-ANEPPS fluorescence images was recorded (excitation by a pair of 50 ms pulses, 438 nm then 531 nm, 3 ms apart, was delivered by the Polychrome II; the emitted fluorescence was filtered via a dichroic mirror of 570 nm and 585/20 nm emission band-pass filter; Chroma Technology Corp., Bellows Falls, VT, USA). Subsequently, the protoplast was illuminated for 5-10 min (by a home-made illuminator of crystal clear LEDs from Tal-Mir Electronics,

Israel) with either RL (660 nm, 220 µmol m^{-2} s^{-1}) or RL + BL (BL: 450 nm, 10–15% of the total intensity of 220 µmol m^{-2} s^{-1}) and a second pair of fluorescence images was recorded as above. The images were processed using FIJI (Abràmoff et al., 2004; Schindelin et al., 2012) to yield a pixel-by-pixel fluorescence ratio, as described previously (Wigoda et al., 2017).

***Fluorescence ratio calibration*.** BSC protoplasts incubated in di-8-ANEPPS as above were subjected to 5-s-long voltage pulses in the range of +17 to -223 mV using a patch-clamp pipette in a whole-cell configuration. A pair of images was recorded during the second half of each pulse, at a steady state. The patch-clamp imaging setup was as described in detail by Wigoda et al. (2017), except for the substitution of the abovementioned Eclipse microscope and IXON camera for the original image-acquisition instruments. This calibration confirmed the previously published positive correlation between the di-8-ANEPPS fluorescence ratio and depolarization (Supplemental Figure S1). This definitively positive correlation allows us to draw a clear conclusion about the direction of the experimentally induced change in the E_M, even if the relatively wide data distribution suggests caution in accepting the absolute E_M values.

***Solutions*.** Di-8-ANEPPS (Molecular Probes, Life Technologies, cat. # D3167, Oregon, USA): a 10 mM stock solution in DMSO was stored in 10 µL aliquots at -20°C.

Pluronic F-127 (Molecular Probes, Life Technologies, cat. # P6867, Oregon, USA), a 20% stock solution in DMSO was stored in 100 uL aliquots at room temperature.

Bath solution (in mM): 5 KCl, 1 $CaCl_2$, 4 $MgCl_2$, 10 MES; pH 5.6; osmolality: 435 mOsm, adjusted w/ D-sorbitol.

Patch-clamp pipette solution (in mM):112 K-gluconate, 28 KCl. 2 $MgCl_2$, 10 HEPES, 328 sorbitol, pH 7.5. Osmolarity: 480 mOsm.

Tyrphostin-9 was added to the bath solution from aliquots used in the detached-leaves experiments.

Determination of the BSCs' cytosolic pH (pH_{CYT})

The experiments were performed between 10 am and 4 pm. BSC protoplasts were isolated according to (Shatil-Cohen et al., 2014) from 6- to 8-week-old SCR:GFP (Col) plants and kept in an Eppendorf vial at room temperature (22 ± 2°C) in the dark until use. The protoplasts were treated as described previously (Torne et al., 2021), with the following modifications: a 10 min treatment was applied, either dark or RL+BL or RL (the

illumination setting was as described above for E_M determination), the bath solution was used for flushing out the external SNARF-AM during the last 1 min of the dark/light treatments and the cells were imaged within the subsequent 5 min. Image acquisition and analysis to determine the pH_{CYT} values were performed as described previously (Torne et al., 2021).

Solutions. Bath solution for protoplasts' pH_{CYT} determination: as for E_M determination. SNARF1-AM: (Molecular Probes, Thermo Fisher cat. #: C-1272); a 50 µg prepackaged portion was dissolved into a 10 mM stock in DMSO, then stored in 10 µL aliquots at -20°C. Pluronic F-127: as for E_M determination.

Measuring leaf vein density

Vein density was measured as described previously (Grunwald et al., 2021).

Determination of the osmotic water permeability coefficient (P_f) under different light regimes

Protoplast isolation and P_f determination. These were performed on *SCR:GFP* (Col) as described previously (Shatil-Cohen et al., 2014), with the following modifications: Both wash solutions (isotonic and hypotonic, the latter, 150 mOsm less than the isotonic solution) were buffered to pH 5.7.

After their isolation, the protoplasts were divided into two: the "light-treated" protoplasts were kept in a vial under the lights of the growth chamber for the duration of the experiment (between 10 AM and 2 PM), the "dark-treated" protoplasts were kept in a vial in a light-proof box. The two types were sampled alternatingly preserving their respective conditions of illumination (using green safe light while sampling in the dark). The sampled "light-treated" protoplasts were placed in the experimental chamber and left to settle down for 10-40 min on the bench under a cool-white-led illumination, and then, for 5-7 min on the microscope stage under the unfiltered microscope light (6 V / 30 W halogen lamp, PHILIPS 5761) during preparations and the 1 min videotaping of swelling during the osmotic challenge. The sampled "dark-treated" protoplasts were placed, under green "safe light", in the experimental chamber, left to settle for about 10 min and after the brief location of a GFP-positive BSC they remained in total darkness for 10 min. The 1-min videotaping of the protoplast's swelling during the hypoosmotic challenge in the

dark necessitated an illumination by the microscope lamp filtered (to block out the blue light) via a red plastic band-pass filter (~600-700 nm / 30 µmol m^{-2} sec^{-1}).

Determination of the pH of guttation drops

Whole, intact WT (Col) plants were covered overnight with black plastic bags to increase the relative humidity in their atmosphere and to prevent transpiration. The following morning, immediately after the growth room lights went on, guttation droplets were collected from the tips of the leaves. Ten to twenty guttation droplets were collected and pooled to reach a sample volume of approximately 20 µL in a 200-µL vial, which was immediately sealed. pH was measured within 10 min of sample collection using a micro-pH electrode MI-410 (Microlectrode, Inc., New Hampshire, USA).

RESULTS

phot1 and phot2 blue light receptors are involved in the regulation of K_{leaf}

Following the GC model in which the photoreceptors phot1 and phot2 transduce the blue light (BL) stomatal-opening signal, and encouraged by the substantial expression of the BL receptor genes *PHOT1* and *PHOT2* in the transcriptome of Arabidopsis BSCs (Wigoda et al., 2017), we explored the roles of these two light receptors in the regulation of leaf hydraulic conductance (K_{leaf}). We compared the K_{leaf} of WT plants to the K_{leaf} of knockout mutants lacking one or both light receptors (*phot1-5*, *phot 2-1* and *phot1-5 phot 2-1*; Figure 1) under two light regimes, red light (RL) and RL+BL. Under RL+BL, the K_{leaf} of WT leaves was more than 3-fold higher than it was under RL treatment and more than 2-fold higher than the K_{leaf} values of the three mutants treated with RL+BL. Also, under RL+BL, the K_{leaf} of the *phot2-1* mutant was about 40% higher than it was under the RL treatment. In contrast, BL did not seem to increase the K_{leaf} of the *phot1-5* mutant or that of the double mutant. Under RL, there were no differences between the K_{leaf} levels of the different mutants. However, while the WT's K_{leaf} was no different from that of the single-mutants, it was about double that of the double mutant (Figure 1). The BL response was particularly depressed in the double mutant: only 13% of the WT's K_{leaf} BL response remained in the double mutant, as compared to 32% and 40% of the WT's K_{leaf} in *phot1-5*

and in *phot2-1*, respectively (Figure 1), and only 35% of the WT's g_S BL response remained in the double mutant, as compared to 57% and 71% of the WT's g_S in *phot1-5* and *phot2-1*, respectively (Supplemental Figure S2A).

The higher calculated K_{leaf} value of the WT under BL (as compared to RL) was due to a greatly increased (less negative) leaf water potential, Ψ_{leaf}, and an appreciably higher transpiration rate, E (Eq. 1, Materials and methods). This invites the interpretation that the BL induced a highly conductive water pathway into the leaf, enabling higher radial water influx which, in turn, was able to compensate for the BL's enhancement of E (i.e., water efflux from the leaf via the BL-increased g_S; Supplemental Figure S2).

BSC-specific silencing of phot1 and phot2 receptors decreased K_{leaf}

To examine the specific participation of the BSCs' light receptors in the BL-induced increase in K_{leaf}, we silenced either the *PHOT1* or the *PHOT2* gene using amiRNA (artificial microRNA) under the BSC-specific promoter, *SCARECROW* (*SCR*, see Materials and Methods). Under RL+BL illumination, the K_{leaf} values of leaves of both types of transgenic plants, *SCR:amiR-PHOT1* and *SCR:amiR-PHOT2*, were lower than the K_{leaf} values of the WT leaves (by 40–50%; Figure 2). In contrast, all of the *SCR:amiR-PHOT* plants had WT-like g_S and E, while their Ψ_{leaf} levels were lower than that of the WT (by

50–90%), suggesting a reduction in the radial influx of water into the leaf (Supplemental Figure S3).

BSC-specific complementation of the *phot1-5* mutant with *PHOT1* restored K_{leaf}

We complemented the *phot1-5* (Col-gl) mutant plants with *SCR*:*PHOT1* to restore phot1 activity exclusively in the BSCs (Materials and methods) and compared them to WT (Col-gl) plants and to *phot1-5* plants. *SCR*:*PHOT1* plants illuminated with RL+BL had their K_{leaf} restored from *phot1-5* values to WT levels (Figure 3). While, as expected, BL increased the WT's g_S, the mutant's g_S and the g_S of the complemented mutant did not change under BL, also as expected from the main BL-receptor-devoid guard cells (Supplemental Figure S4). Under BL, the Ψ_{leaf} of the *SCR*:*PHOT1*-complemented mutant was restored to the WT level and both of those values were more than 50% greater than the mutant's Ψ_{leaf} (Supplemental Figure S4). Thus restored Ψ_{leaf} was the major contribution to the restored high K_{leaf} of the complemented mutant, suggesting a restoration of radial water influx to the leaf.

The kinase inhibitor, tyrphostin 9, prevented the increase in K_{leaf} under blue light

The kinase inhibitor tyrphostin 9 suppresses BL-dependent H^+-ATPase phosphorylation (of the penultimate threonine) in guard cells (Hayashi et al., 2017). To examine further whether the vascular phot receptors initiate a similarly sensitive phosphorylating event in the BL-dependent K_{leaf} regulation pathway, we perfused detached WT leaves with AXS, with or without 10 μM tyrphostin 9, and exposed them to RL or RL+ BL. The K_{leaf} values of RL+BL-illuminated leaves perfused with tyrphostin 9 were about 50% lower than those of RL+BL -illuminated control leaves (without tyrphostin 9), and were no different than the K_{leaf} of leaves illuminated with only RL (with or without exposure to tyrphostin 9; Figure 4A, left panel, Supplemental Figure S5). However, g_S was not affected by the petiole-delivered tyrphostin 9 (Figure 4B, left panel). In contrast to the perfused tyrphostin 9, spraying the inhibitor on the leaf surface did not interfere with the almost 3-fold increase in K_{leaf} that was induced by BL (Fig. 4A, right panel), but it did prevent the BL-induced increase in g_S (Figure 4B, right panel). The impaired stomatal response as a result of the application of tyrphostin 9 directly to the leaf surface is in agreement with tyrphostin 9's previously reported inhibition of BL-induced stomatal opening in epidermal peels (Hayashi et al., 2017). Leaf transpiration, E, behaved in a

pattern similar to g_S (Supplemental Figure S6A). Petiole-fed tyrphostin 9 abolished the BL-induced increase in Ψ_{leaf}, but tyrphostin 9 sprayed on the leaf surface did not prevent it (Supplemental Figure S6B).

Blue light hyperpolarizes the BSCs and raises their cytosolic pH

To investigate further whether the BSCs' BL signal transduction pathway stimulates their plasma-membrane H^+-ATPases (as in GCs), we monitored the membrane potential of BSC protoplasts using the potentiometric dual-excitation dye di-8-ANEPPS. Indeed, 5–10 min of RL+BL illumination hyperpolarized the BSC protoplasts relative to RL alone or to the dark (absence of illumination), while 10 µM tyrphostin 9 added in the bath depolarized the BSCs irrespectively of RL+BL illumination or its absence (Figure 5A, Supplemental Figure S1), hence, apparently independently of the inhibition of AHA activity.

To further explore our hypothesis of H^+-ATPase activation by BL, we monitored the cytosolic pH (pH_{CYT}) using the ratiometric, dual-emission fluorescent pH-sensitive dye, SNARF1. The pH_{CYT} of BSCs illuminated for 10 min with RL+BL (following dark treatment) was significantly more alkaline (by 0.72 pH units) than the pH_{CYT} of dark-treated BSCs and 1.06 pH units higher than the pH_{CYT} of the BSCs that were treated with only RL (Figure 5B), consistent with BL-induced activity of H^+-ATPases, which move protons from the BSCs into the xylem.

AHA2 plays a role in the blue light-induced regulation of K_{leaf}

Recently, we reported that AHA2 plays a major role in K_{leaf} regulation by acidifying the xylem sap. To further examine whether the BSCs' H^+-ATPase AHA2 participates in the *BL-initiated* K_{leaf} regulation, we examined the AHA2 knockout mutant *aha2-4* and its BSC-complemented transgenic plant line *SCR*:*AHA2* [i.e., the *aha2-4* mutant with AHA2 expressed only in its BSCs (Grunwald et al., 2021)] under the two light regimes (RL and RL+BL). The K_{leaf} of *aha2-4* leaves did not respond to the RL+BL illumination and was only about 40% of the K_{leaf} of BL-treated WT leaves. However, similar illumination increased the K_{leaf} of the AHA2-complemented (*SCR*:*AHA2*) plants to almost 65% of the WT's K_{leaf} (Figure 6), indicating that the BL-induced increase in K_{leaf} is dependent on AHA2. In contrast, under RL+BL, the E and g_S of *SCR*:*AHA2* did not differ from the E and g_S of the *aha2-4* mutant, and they remained unaffected by BL and lower than the E and g_S

of the WT (Supplemental Figure S7A,B). Notably, the Ψ_{leaf} of *SCR*:*AHA2* was about 40% higher than the Ψ_{leaf} of *aha2-4* and did not differ from that of the WT, suggesting a restoration of radial water influx to the leaf (Supplemental Figure S7C).

Morning light after night darkness acidifies the xylem sap

To delve further into the acidification-mediated link between light and K_{leaf}, we explored the diurnal transition between the xylem sap pH in the night and in the morning. We hypothesized that the xylem sap pH would be reflected in the pH of guttation drops exuded via hydathodes from the ends of the xylem conduits. Therefore, to take a preliminary peek at the xylem sap pH at night, we measured the pH of guttation drops collected from the tips of WT leaves before the lights were turned on and found that their mean pH was 7.2 ± 0.04 (±SE, 15 biological repeats collected over 3 days). Next, we measured the pH of the xylem sap in detached leaves, comparing leaves before dawn to leaves after dawn (i.e., leaves at the end of an overnight dark treatment to leaves after a 30 min of illumination with white light in the growth chamber). This comparison included leaves of WT, *phot1-5* and *SCR:PHOT1* plants. Illumination acidified the xylem sap in WT plants by approx. 0.6 pH units (from 6.0 to 5.4) and in the *SCR: PHOT1* plants by approx. 0.8 pH units (from 6.1 to 5.3), while the xylem pH in *phot1-5* remained unchanged, at about 5.9 (Figure 7), suggesting that phot1 in the BSCs is necessary for light-activated xylem acidification, which, in turn, increases K_{leaf} (Grunwald et al., 2021).

Light increases the osmotic water permeability, P_f, of the BSCs

To explore the link between light and K_{leaf} at the cellular level, we tested whether light affects the osmotic water permeability (P_f) of the BSC membrane. We determined the P_f of isolated BSC protoplasts from videotapes of their swelling during a hypotonic challenge after illumination with white light or after 10 min in the dark. The illuminated BSCs had higher P_f than the dark-treated ones (Figure 8), suggesting that the morning increase in the movement of water from the xylem to the BSCs (and then, presumably, further into the mesophyll) is due to increased permeability of the BSC membranes to water.

Leaf vein density is not affected by the expression of phot1 or phot2

To resolve whether the diminished K_{leaf} of the phot receptor mutants that we observed under BL was due to differences in leaf vein density, we quantified the vein density of all of the genotypes that we studied and found no differences among them (Supplemental Figure S8).

DISCUSSION

Blue-light signal transduction in BSCs is reflected in the K_{leaf} increase

We demonstrate here an autonomous BL signaling mechanism in Arabidopsis BSCs that underlies the light-induced increase of K_{leaf}, resembling the "classical" short-term, BL-induced signaling pathway for stomatal opening mediated by the photoreceptors phot1 and phot2 (Zeiger et al., 1983; Sakai et al., 2001; Kinoshita et al., 2001). To explain the underlying mechanism, we built on our earlier findings that the receptor genes *PHOT1* and *PHOT2* are substantially expressed in BSCs (Wigoda et al., 2017; Supplemental Table 2). Here, we link these findings using the phot receptor mutants *phot1-5* and *phot2-1* and the double mutant *phot1-5 phot2-1*. Based on the complete abolishment of the BL-K_{leaf} response (relative to RL alone) in the *phot1-5* mutant, which is presumed to possess phot2 activity, phot2 would appear completely redundant, were it not for (a) the strong depression of the BL-K_{leaf} response – compared to WT response – in the *phot2-1* mutant, presumed to possess phot1 activity, and (b) even more severe impairment of the double mutant's BL-K_{leaf} response compared to the responses of both single mutants (Figure 1).

Also the BL-g_S responses that we observed in the three phot mutants were depressed to different relative degrees resembling the gradation of BL-K_{leaf} responses (Supplemental Figure S2A) and suggesting a similar dependence of BL signaling on phots in the two systems (BSCs and GCs). Interestingly, after 2-4 h of illumination (well beyond our usual 20 min of illumination) the *phot* single mutants's g_S re-acquired the WT-like sensitivity to BL (Supplemental Figure S9), similar to an earlier report on stomatal aperture after 2-4 h of illumination with BL (Kinoshita et al., 2001). This could perhaps reflect a gradually accumulating effect of the GCs' AHA activity, overcoming the partially impaired phot-to-AHA signal transduction in the single *phot* mutants.

That the phots (and particularly phot1) are indispensable to BL signaling in the BSCs, as they are in GCs (Kinoshita et al., 2001), is additionally emphasized by our two

BSCs-delimited genetic phot manipulations: (a) the severe impairment of the BL-induced K_{leaf} increase in WT plants in which either phot1 or phot2 was silenced by the respective *SCR:amiR-PHOT* (Figure 2) and (b) the full restoration of K_{leaf} sensitivity to BL by *phot1-5* complementation with *SCR:PHOT1* (Figure 3). These manipulations, which did not affect g_S (Supplemental Figures S3A, S4A), also emphasize the independence of the BSCs' BL signal transduction from that of the GCs.

Prompted by finding BLUS1, the phots' indispensable partner kinase in the BL-induced early-signaling complex of GCs (Takemiya et al., 2013; Hayashi et al., 2017) also in the BSCs transcriptome (Wigoda et al., 2017; Supplemental Table S2), we found that the *blus1* mutant's K_{leaf} does not respond to BL (Supplemental Figure S10), underscoring further the resemblance in BL signaling between the two systems.

Additionally, the tissue-localized effects of the kinase inhibitor, tyrphostin 9, which presumably inhibited a BL-signaling kinase downstream of phot-BLUS1, demonstrated both the similarity between the BSCs' and the GCs' BL signaling pathways in WT plants, and, again, the independence of the two systems. Both signaling pathways were inhibited by tyrphostin 9, but only when it was applied in close proximity to the target tissue: The BSC system was affected only by the petiole-fed tyrphostin 9, which inhibited the BL-induced increase of K_{leaf} and Ψ_{leaf} without affecting g_S; whereas the GC system was affected only when tyrphostin 9 was sprayed on the leaf surface (i.e., on GCs), which abolished the BL-induced g_S increase without affecting K_{leaf} (Figure 4, Supplemental Figure S6). Notwithstanding the above, the depolarization of BSCs by tyrphostin 9 both under RL+BL and in the dark (Fig. 5A), suggests an additional or an alternative underlying mechanism *outside* the AHA-activating BL-signaling pathway.

Recently, we reported that the BSC H^+-ATPase AHA2 plays a major role in the acidification of the xylem sap, which, in turn, increases K_{leaf} (Grunwald et al., 2021). Indeed, the H^+-ATPase mutant, *aha2-4* had a diminished (compared to WT) BL-induced increase in K_{leaf} (Figure 6; likely due to a more alkaline xylem pH; Grunwald et al., 2021), as well as a diminished BL-induced increase in g_S (Supplemental Figure S7). In contrast, *aha2-4* plants complemented with the *SCR:AHA2* – even though directed only to BSCs – had their BL-induced K_{leaf} increase restored (Figure 6), while their g_S remained unaffected (Supplemental Figure S7), affirming the importance of the BSC AHA2 in mediating the BL-induced increase in K_{leaf} independently of g_S (i.e., of GCs).

Thus, with the genetic manipulation of the BL receptors phot1 and phot2 in the BSCs, the tissue-differential responses to pharmacological treatment and genetic manipulation of the AHA2 we demonstrated the independence of the BSCs' BL signaling from the GCs' BL signaling. This notion of independence is particularly important since the BSC layer constitutes a hydraulic valve between the xylem and the leaf, regulating water loss to the atmosphere *in series* with stomatal apertures.

Blue-light signal transduction in BSCs is reflected in their H^+-ATPase activation

Four of our findings focus on the activation of H^+-ATPase as the end-point of BL signaling: at the level of single isolated BSCs, (a) the BL-induced alkalinization of the BSCs cytosol (relative to dark or to RL; Figure 5B), (b) the BL-induced hyperpolarization in WT (Col) BSC protoplasts (relative to dark or to RL, Figure 5A), and also in WT (Col-gl) BSC protoplasts (Supplemental Figure S11A, Materials and Methods), (c) the absence of hyperpolarization under RL+BL treatment in the BSCs protoplasts of the *phot1-5* mutant and its restoration in the *phot1-5* mutant complemented with *PHOT1* solely in its BSCs (Supplemental Figure S11B); at the level of the detached leaf, (d) acidification of the xylem sap by white light (relative to dark) in WT, the absence of the white light response in the *phot1-5* mutant and a restoration of this response in the abovementioned *PHOT1*-complemented mutant (Figure 7). Taken together, both types of the pH responses (which occur by proton extrusion from the cytosol) and the hyperpolarization (which could also result from proton extrusion via the plasma membrane, resembling the hyperpolarization of GCs in response to BL (Assmann et al., 1985; Roelfsema et al., 2001), are strong evidence linking the phot receptors with the BL-induced activation of the BSCs' AHA2 in a similar way to BL activation of the GCs' AHA (Hayashi et al., 2017).

Light induces the increase of BSCs' membrane water permeability, K_{leaf} and K_{ros}

Here, we report that, in addition to external (experimentally imposed and constant) low pH, light also increases the P_f of isolated BSCs (Figure 8). Evidence is accumulating that P_f level reflects the underlying activity of aquaporins. For example, frog oocytes expressing AtPIP2;1 had higher P_f induced by co-expressing two of the Arabidopsis 14-3-3 protein isoforms which bound to the AtPIP2;1 (Prado et al., 2019). Thus, it would be tempting to speculate that light increased the BSCs' P_f (Figure 8) by enhancing the activity of one or more of the 11 BSCs' aquaporins (Supplemental Table S2).

Alternatively, or in addition, the light-induced increase in BSC P_f could be due to the alkalization of the BSCs' cytosol (Figure 5B), sensed by a histidine (His 197, conserved in all Arabidopsis PIP aquaporins; assayed in AtPIP2;2; Tournaire-Roux et al., 2003) and a leucine (Leu206 in BvPIP2;2 Fortuna et al., 2019) in the cytosol-facing D loop of the aquaporin protein. Moreover, the concurrent hyperpolarization of BSCs by BL (Figure 5A) could perhaps also contribute to the activation of the BSCs' aquaporins, as suggested by *in silico* simulations (Hub et al., 2010).

The role of aquaporins in controlling K_{leaf} was identified previously [in walnut, Ben Baaziz et al., 2012; in grapevine (*Vitis vinifera*), Vitali et al., 2016; in poplar (*Populus trichocarpa*), Muries et al., 2019; in Arabidopsis BSCs, Sade et al., 2014; (Prado et al., 2013)]. In particular, Prado et al. (2019) demonstrated a contribution of the abundant – also in the leaf veins – aquaporin AtPIP2;1 to the diurnal and circadian rhythms of the hydraulic conductance of the Arabidopsis rosette, K_{ros}, which peaked before midday. Thus, in addition to the likely P_f-promoting effect of the slightly acidic pH (6) perfusion solution (a link established by Grunwald et al., 2021), that Prado et al., (2019) used for their K_{ros} measurements, the changes that they observed in K_{ros} also reflected the effects of light and circadian rhythms (Prado et al., 2019). Thus, light-induced changes in aquaporin activity underlying the effects of light on K_{ros} are consistent with light-induced increase of P_f that we observed.

BL signaling increases the K_{leaf} - a model

Whatever the exact underlying molecular mechanism, our results support the following model of K_{leaf} increase by BL, in a GC-like, BL signal transduction pathway: the BSCs' phot1 and phot2 perceive the BL, which, via BLUS1 and unknown downstream protein kinases (the first of which could perhaps be BHP, (Hayashi et al., 2017; Hosotani et al., 2021) activates the plasma membrane H^+-ATPase (AHA2), resulting in the hyperpolarization of the BSC, the alkalization of the BCSs' cytosol and the acidification of the xylem sap, leading to an increase in the BSCs' P_f and a subsequent increase in K_{leaf}.

Expressed schematically:

BL → activation of BSC *phots* → phosphorylation* & activation of *blus1* and downstream kinases → activation of BSC H^+-ATPase (AHA2) → hyperpolarization and cytosol alkalization of BSCs and acidification of xylem → increase of P_f of BSCs → K_{leaf} increase

*We hypothesize that the abovementioned phosphorylation steps are similar to those seen in GCs (Figure 9), but this has not yet been explored in BSCs.

An early K_{leaf} response to blue light - what is the advantage?

Both the opening of the stomata and the increase in K_{leaf} occur in the early morning hours (e.g., Brodribb and Holbrook, 2004; Domec et al., 2009; Locke and Ort, 2015) when the irradiance spectrum includes relatively high levels of BL (Matthews et al., 2020, and references therein). *What is the advantage of the early [blue-]light response of stomata?* We suggest that it allows the acquisition of CO_2 under sub-maximum VPD (vapor pressure deficit, a measure of the driving force for leaf water evaporation), thereby increasing water-use efficiency.

Interestingly, K_{leaf} responds to light even faster than g_S does (Guyot et al., 2012) – in an ultimate manifestation of the independence of the BSCs' BL signal transduction from that of the GCs'. One possible advantage of this almost-simultaneity is matching the stomatal opening with the increased water flux into the leaf. Were it not for this accompanying K_{leaf} increase, the BL-induced stomata opening, which peaks in the morning hours (the "golden hour"; (Gosa et al., 2019), could lead to an imbalance in leaf water, even in the presence of a relatively low VPD, leading to a drop in the leaf water potential (Ψ_{leaf}), which, in turn, could close the stomata (Raschke, 1970; Guyot et al., 2012; Klein, 2014; Scoffoni et al., 2018) and thereby limit photosynthesis. Instead, an early morning increase in K_{leaf} elevates Ψ_{leaf}, thereby increasing and prolonging stomatal opening (ibid). Hypothetically, under more extreme conditions (e.g., with high VPD even in the early morning), BL-increased K_{leaf} would prevent a hydraulic pathway failure when stomatal conductance is at its peak (Brodribb and Holbrook, 2004; Halperin et al., 2017; Gosa et al., 2019).

Another possible advantage of the early morning, BL-induced increase in K_{leaf} may be an increased CO_2 permeability of aquaporins. A few lines of evidence converge to support this hypothesis; (a) CO_2 can cross cell membranes through aquaporins, particularly aquaporins of the PIP1 family (Kaldenhoff and Fischer, 2006), (b) the transcripts of PIP1 family aquaporins in walnut have been shown to be upregulated concomitantly with the increase in K_{leaf} induced by a brief (15 min) exposure to white light (Ben Baaziz et al., 2012), (c) in soybean (*Glycine max*), the transcript of a PIP1 family aquaporin (PIP1;8) was found to be more abundant in the morning than at midday, in correlation with a higher K_{leaf}

in the morning (Locke and Ort, 2015), (d) aquaporins control K_{leaf} (Shatil-Cohen and Moshelion, 2012; Sade et al., 2014), (e) CO_2 transported by xylem from the roots may be a source for the CO_2 assimilated into the bundle sheath and mesophyll (Janacek et al., 2009; Hubeau et al., 2019). Thus, assuming the K_{leaf} increase with the first light of day parallels the increase in the permeability of BSC aquaporins to CO_2, the passage from the xylem to the mesophyll of the root-originated CO_2 will be enhanced even before full stomatal opening. The increased availability of CO_2 at this time, when the photosynthetic light is already sufficient, would enhance CO_2 assimilation – a great advantage for the plant. This hypothesis is strengthened by the fact that (Stutz and Hanson, 2019) have shown that the xylem-transported CO_2 constitutes a varying proportion of the total CO_2 assimilated throughout the day depending on the availability of light and CO_2. Notably, photosynthetic CO_2 uptake reaches a maximum within 10 min of exposure to BL (Doi et al., 2015), which is over 3-fold faster than the full opening of stomata (Grondin et al., 2015).

The advantage of diminished AHA activity and P_f at night

The fact that the BSC P_f is low in the dark (Figure 8) may explain how the mesophyll does not get flooded at night when root pressure increases. At night, the P_f and K_{leaf} reduction likely prevent water influx into the leaf, while the buildup of root pressure is relieved via guttation. A malfunction of this mechanism is perhaps what underlies leaf flooding by the xylem sap (Shatil-Cohen and Moshelion, 2012). The inactivity of the BSCs H^+-ATPases, manifested here in the nightly alkalization of the xylem sap and in guttation drops, (as noted earlier, for example, also in tomatoes (Urrestarazu et al., 1996) and poplars (Aubrey et al., 2011) , could be advantageous for the plant as a means of energy conservation, as ATPases operate at a high energetic cost (Palmgren, 2001).

Our findings expand the understanding of the molecular basis of the regulation of leaf water influx. The fact that BL increases K_{leaf} in several other species imparts an even more general importance to our results. A focus on the hydraulic valve in series with the stomata should provide new directions for the study of plant water relations in different environments.

AUTHOR CONTRIBUTIONS

YG and SG constructed the BSCs-directed transgenic lines: mutant complementation and *PHOT* silencing., planned and performed the K_{leaf} and vein pH experiments, and wrote the

paper. YG planned and performed the P_f experiments. TT planned and performed the membrane potential and cytosolic pH experiments. NM and MM conceptualized and supervised the project and wrote the paper.

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FIGURE LEGENDS

Figure 1 The effect of blue light on the K_{leaf} of PHOT receptor mutants. K_{leaf} of fully expanded, excised leaves of WT (Col-gl) and *PHOT* mutants [*phot1-5* (Col-gl), *phot2-1* (Col-gl) and a double mutant, *phot1-5 phot2-1* (Col-gl)] determined immediately after 15 min of illumination with red light (RL; 220 µmol m^{-2} s^{-1}) or red + blue light (RL+BL; 220 µmol m^{-2} s^{-1}, consisting of 90% RL and 10% BL). The columns are mean values (±SE) of *n* biological repeats (leaves) from three independent experiments (the *n* values noted inside each column). Different letters denote significantly different values (ANOVA, post hoc Tukey's test; $P < 0.05$). Note the reduced response to RL+BL in the mutant lines.

Figure 2 amiR-silencing of phot1 or phot2 (*SCR:amiR-PHOT* plants) in the BSCs reduces K_{leaf} under RL+BL light. K_{leaf} was determined in fully expanded leaves of WT, *SCR:amiR-PHOT1* (two independent lines 1,2) and *SCR:amiR-PHOT2* (two independent lines 1,2) plants treated with RL or RL+BL (as in Figure 1) . Other details also as in Figure 1.

Figure 3 BSC-directed *PHOT1* complementation of the *phot1-5* mutant restores the normal K_{leaf}. Fully expanded leaves of WT (Col-gl), *phot1-5* (Col-gl) and *phot1-5* (Col-gl) plants complemented with *SCR:PHOT1* were subjected to illumination treatments, as in Figure 1. Other details also as in Figure 1.

Figure 4 The kinase inhibitor tyrphostin 9 either abolished the BL-induced K_{leaf} increase or prevented the BL-induced stomatal opening - depending on the treatment route. Fully expanded leaves of WT (Col-gl) were pre-incubated (petiole deep) in AXS without or with tyrphostin 9 (10 µm) or sprayed with AXS with or without tyrphostin 9 (10 µm), and kept overnight in dark boxes. Immediately after dark, leaves were illuminated for 15 min with RL or RL+BL, as indicated. (A) K_{leaf}. (B) Stomatal conductance (g_S). Other details as in Fig. 1. Note that K_{leaf} was reduced when tyrphostin 9 was fed via the petiole, but not when it was sprayed on the plants, and that the opposite was true for g_S.

Figure 5 Blue light hyperpolarizes the isolated BSC protoplasts and alkalinizes their cytosol relative to red light or darkness, and tyrphostin 9 depolarizes them. **(**A) Membrane potential (E_M) of BSC protoplasts, with or without bath-added tyrphostin 9 (10 μM), determined using the dual-excitation, ratiometric fluorescent dye di-8-ANEPPS. After at least 10 min in the dark, the cells were immediately imaged (dark treatment) or illuminated for an additional 5–10 min either with RL or RL+BL and then imaged (Materials and Methods). The columns are the mean fluorescence ratio (F_{438}/F_{531}) values (±SE) of the indicated number of biological repeats (protoplasts), derived from analysis of the images captured at the indicated excitation wavelengths in three to eight independent experiments. The positive correlation between E_M and the F_{438}/F_{531} (with depolarization indicated by the arrow) was verified in separate experiments using a patch clamp (Supplemental Figure S1). Different letters denote significantly different values (ANOVA, post hoc Tukey's test; $P < 0.05$). (B) Cytosolic pH of BSC protoplasts determined using the fluorescent pH probe SNARF1. Protoplasts were dark-treated for 10 min and then further treated with 10 min of either dark, RL or RL+BL, then imaged. Cytosolic pH values were derived from the analysis of the images captured; the emitted fluorescence ratio (F_{640}/F_{585}) values were converted to pH values based on a calibration curve (as detailed in Torne et al., 2021). The data are from at least three independent experiments. Other details are as in Figure 5A.

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Figure 7 Illumination with white light for 30 min acidified the xylem pH of WT plants and *SCR*:*PHOT1*-complemented mutants, but not that of the *phot1-5* mutant. The leaves of WT (Col-gl) plants, *phot1-5* (Col-gl) mutants and the *SCR:PHOT1*-complemented *phot1-5* mutant (as in Figure 3) were illuminated with white light in the growth room (150–200 μmol m^{-2} sec^{-1}). Other details as in Figure 1.

Figure 8 White light increases the BSCs' membrane osmotic water permeability coefficient (P_f). (A) Time course of the swelling of bundle sheath protoplasts from *SCR:GFP* plants upon exposure to a hypotonic solution under the same illumination conditions (white light or darkness) as during the light treatment preceding the assay (Materials and Methods). The arrow indicates the onset of the bath flush. (B) Time course of the osmotic concentration change in the bath (C_{out}) during the hypotonic challenge. (C) Mean (±SE) initial P_f values of the indicated numbers (*n)* of BSC protoplasts under two light regimes from five independent experiments. The asterisk indicates a significant difference ($P < 0.05$; Student's two-tailed, non-paired *t*-test).

Figure 9 Proposed BSC-autonomous blue-light signal transduction pathway - an artist's rendering. (A) A leaf radial water path, from xylem to mesophyll (K_{leaf}, hollow blue arrow) via the bundle sheath cells (BSCs), which tightly envelop the xylem. GC, stomatal guard cell; MC, mesophyll cell. (B) Blue-light (BL) signaling pathway (blue arrowheads) in a bundle sheath cell, from BL perception by the phototropin receptors (phot) complexed with the kinases, BLUS1 and BHP (Hayashi et al., 2017; Inoue and Kinoshita, 2017), through an intermediate tyrphostin 9 (TP9)-inhibitable phosphorylation (P) of an unknown substrate X by an unknown Raf-like kinase, down to the ultimate AHA2 activation by its phosphorylation (P). AHA2 activation results in proton extrusion via the pump and xylem sap acidification, presumably, at the expense of ATP breakdown to ADP with a transient phosphorylation (P) of the catalytic site on the pump protein, as expected from a P-type H^+-ATPase.

SUPPLEMENTAL FIGURES AND TABLES

Supplemental Figure S1. The relationship between the di-8-ANEPPS fluorescence ratio (F-ratio, F_{438}/F_{531}) and the BSC membrane potential (E_M).

Supplemental Figure S2. phot receptor mutants do not show any effect of BL on their g_S, E or Ψ_{leaf}.

Supplemental Figure S3. *SCR:amiR*-silencing of the *PHOT1* or *PHOT2* genes in the BSCs reduces Ψ_{leaf} under red + blue light, but does not reduce the stomatal conductance (g_S) or the rate of transpiration (E).

Supplemental Figure S4. BSC-directed *PHOT1* complementation of the *phot1-5* mutant elevates the leaf water potential (Ψ_{leaf}), but not stomatal conductance (g_S) or the transpiration rate (E).

Supplemental Figure S5. Tyrphostin 9 perfused via the petiole does not affect the examined physiological parameters under red-light-only illumination.

Supplemental Figure S6. Xylem-fed kinase inhibitor tyrphostin 9 does not affect the blue light-induced increase in the transpiration rate (E), but does prevent any blue light-induced increase in leaf water potential (Ψ_{leaf}).

Supplemental Figure S7. BSC-directed *AHA2* complementation of the *aha2-4* mutant restores its blue light-induced leaf water potential (Ψ_{leaf}) increase, but not the blue light-induced increases in stomatal conductance (g_S) or in the transpiration rate (E).

Supplemental Figure S8. Leaf vein density does not depend on the expression of phot1 or phot2.

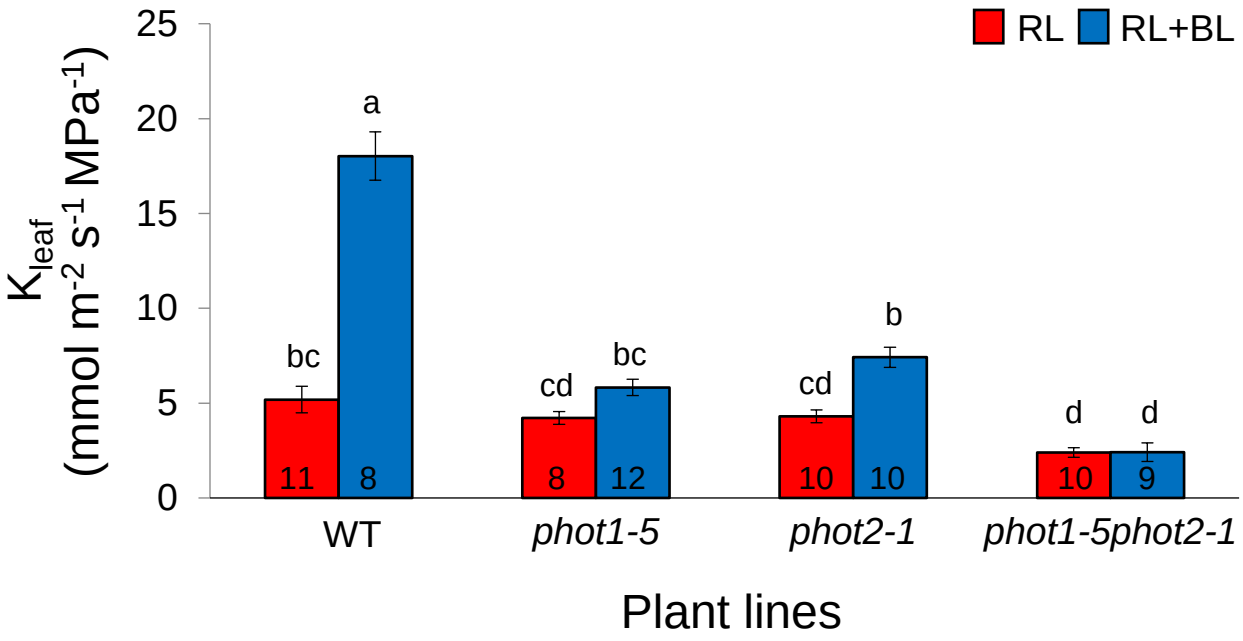
Supplemental Figure S9. After prolonged illumination, stomatal conductance (g_S) of intact leaves of whole plants did not differ among the different genotypes (WT, *phot* mutants and *SCR:PHOT1*).

Supplemental Figure S10. The BLUS1 kinase mutation abolishes the BL-induced K_{leaf} increase.

Supplemental Figure S11. The hyperpolarization of BSCprotoplasts by BL is prevented by the phot1 mutation.

Supplemental Table S1. List of primers used in this study

Supplemental Table S2. Expression levels of some genes found in the transcriptomes of bundle-sheath (BS) cells and mesophyll (MES) cells (Wigoda et al., 2017, GEO repository Experiment GSE85463) that are or may be related to blue-light signaling, encoding receptors, kinases, H^+-ATPases and aquaporins.



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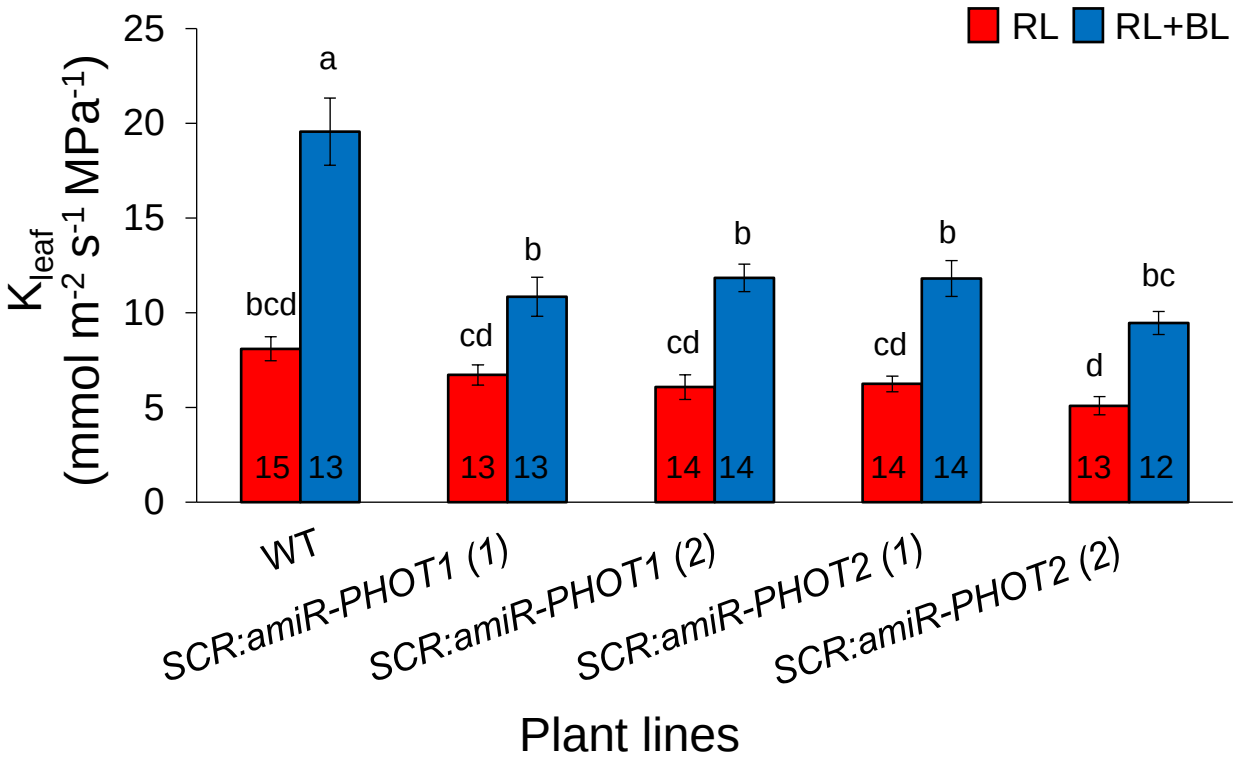
F1. Grunwald, Gosa et al., 2021



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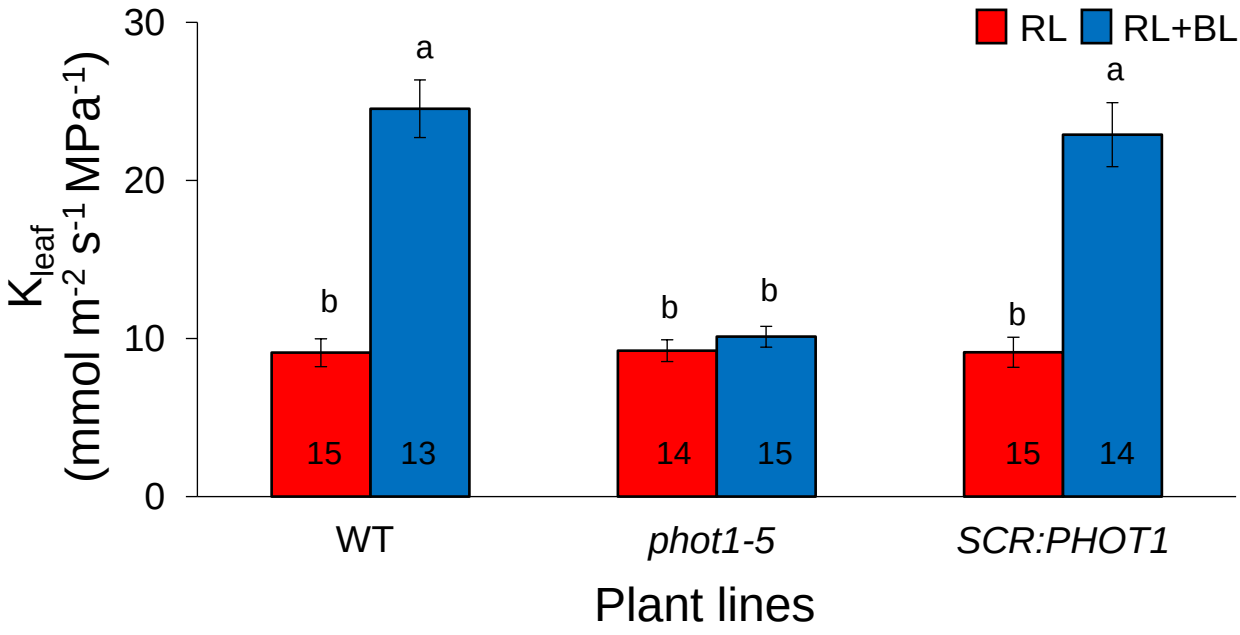
F2. Grunwald, Gosa et al., 2021



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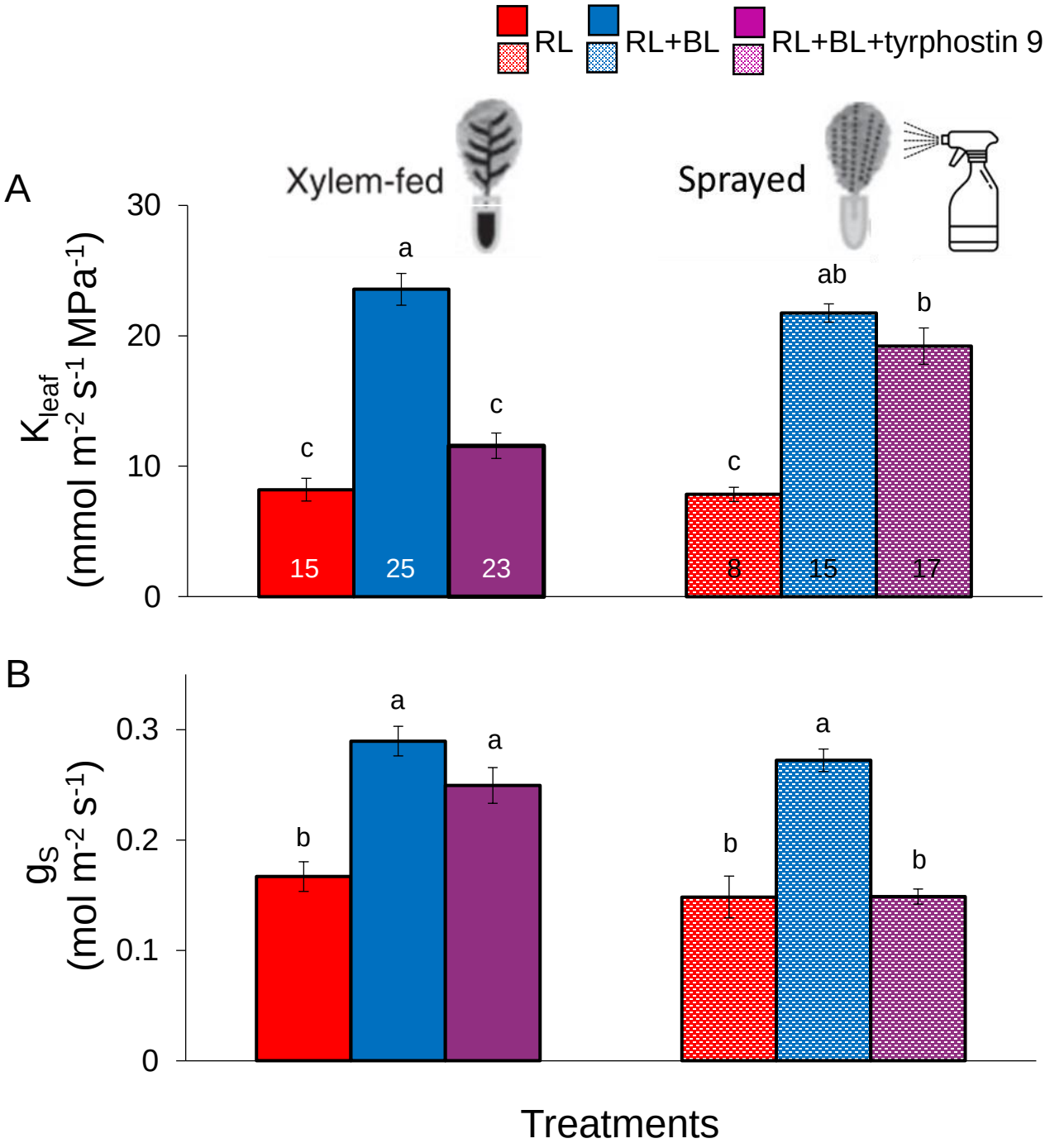
F3. Grunwald, Gosa et al., 2021



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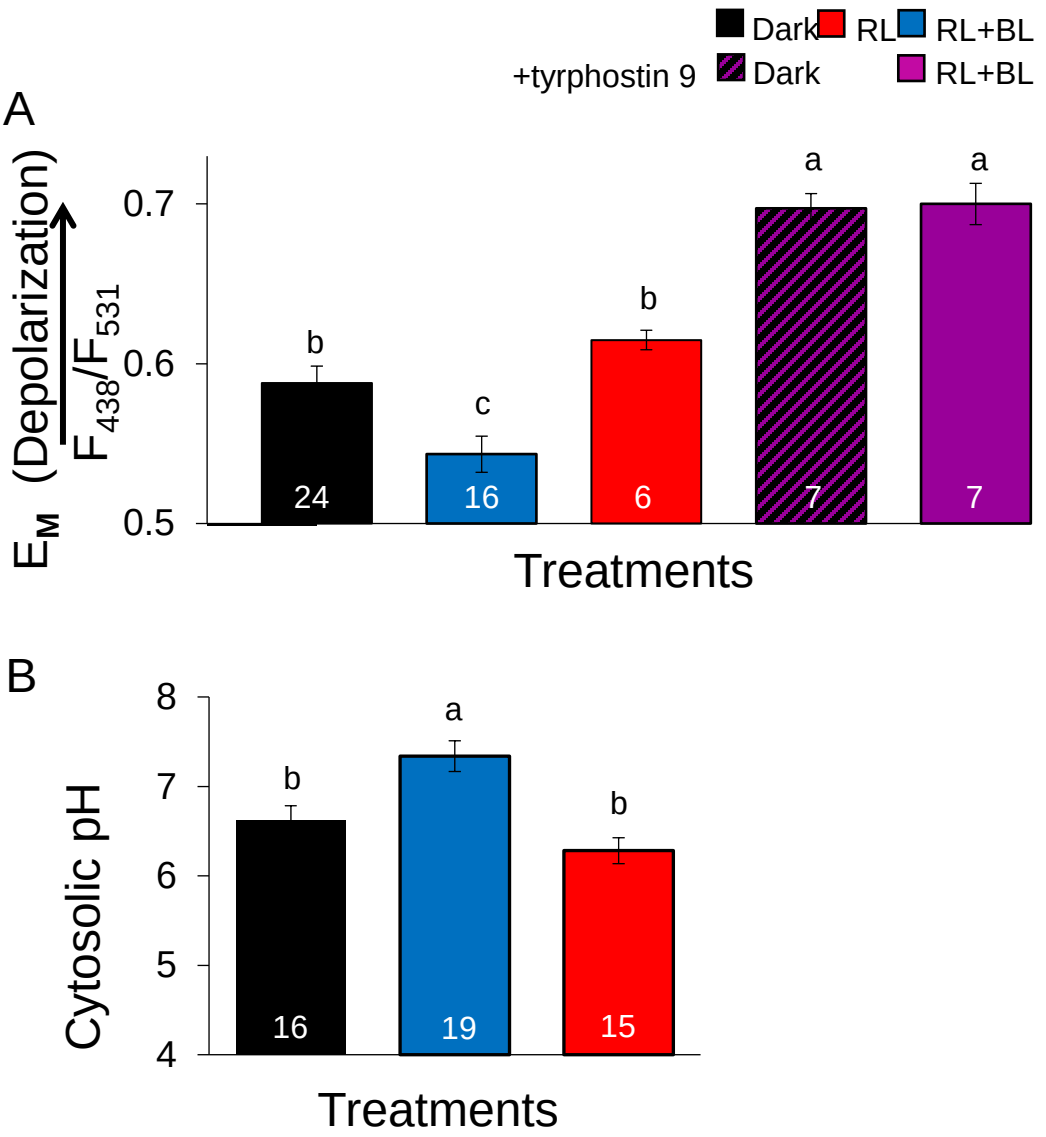
F4. Grunwald, Gosa et al., 2021



F5. Grunwald, Gosa et al., 2021

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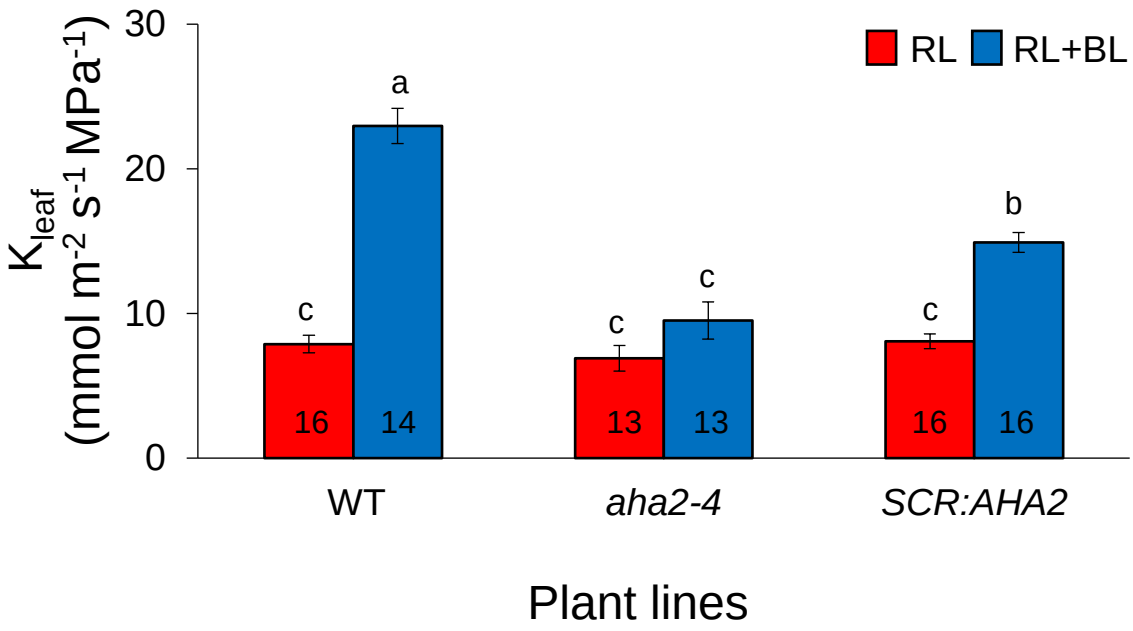

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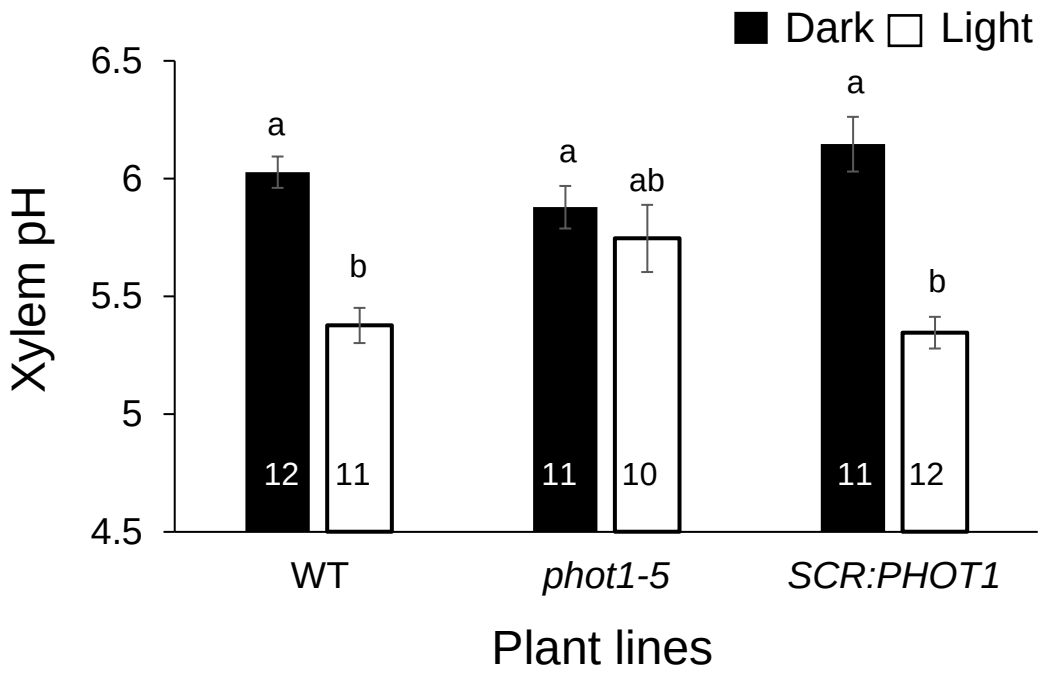
F6. Grunwald, Gosa et al., 2021



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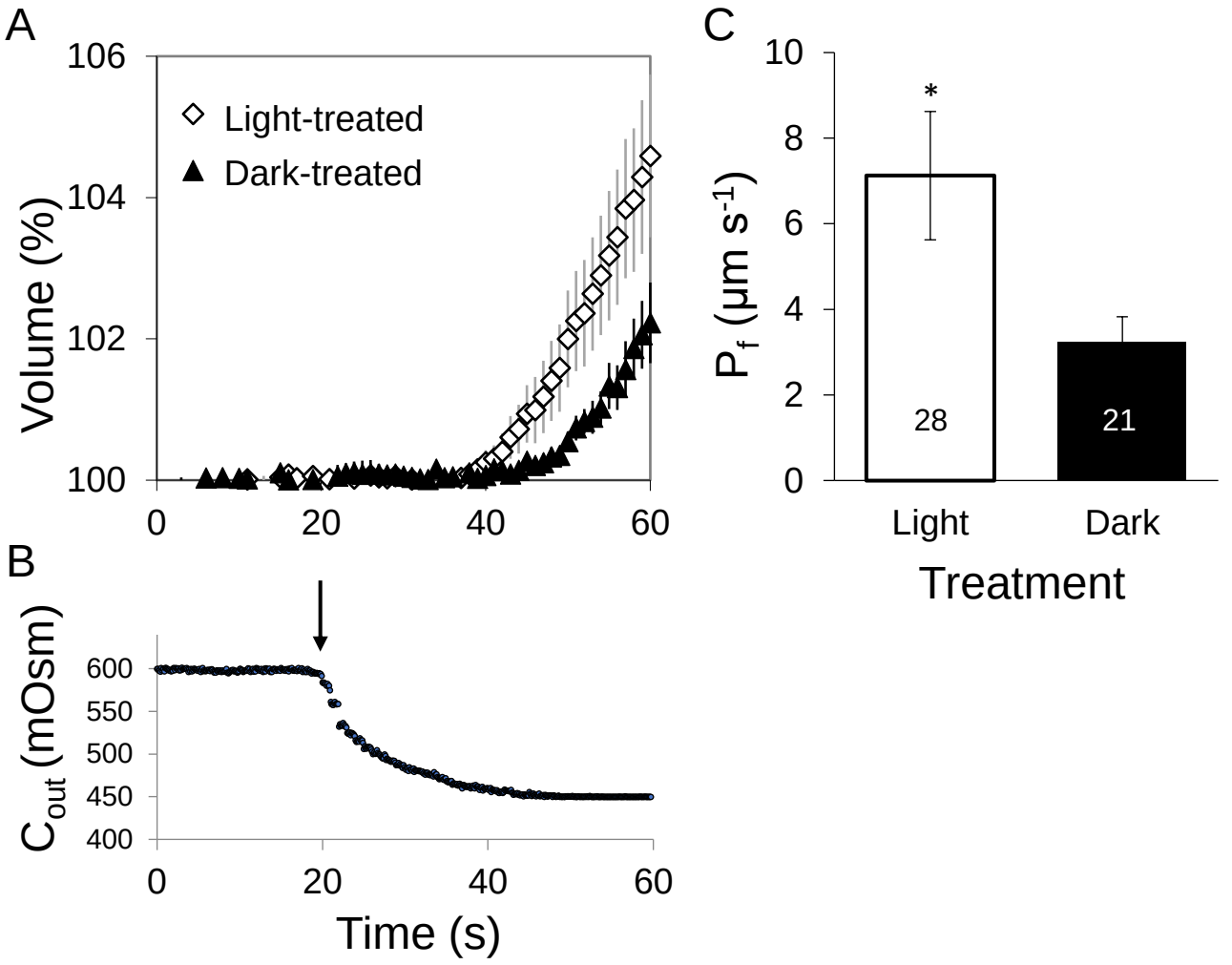

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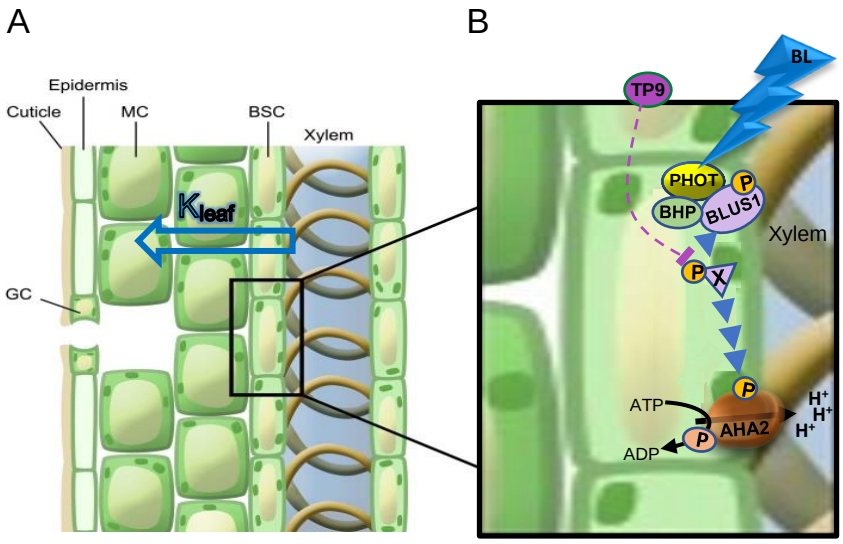
F8. Grunwald, GOsa et al, 2021



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F9. Grunwald, Gosa et al., 2021