**Herbivore intraspecific variation and evolutionary divergence drive trophic cascade strength**

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**Abstract**

Trophic cascades—the indirect effect of predators on non-adjacent lower trophic levels—are important drivers of the structure and dynamics of ecological communities. However, the influence of intraspecific trait variation on trophic cascade strength remains largely unexplored, which limits our understanding of the mechanisms underlying ecological networks. Here we experimentally investigated how herbivore intraspecific genetic variation and evolutionary divergence related to host-plant specialization influences trophic cascade strength in a terrestrial tritrophic system. We found that the occurrence and strength of the trophic cascade are strongly influenced by herbivores' intraspecific variation and evolutionary divergence but are not associated with density-dependent effects mediated by herbivore population growth rate. Our findings stress the importance of intraspecific trait diversity and evolutionary adaptations as drivers of trophic cascade strength and underline that intraspecific variation should not be overlooked to decipher the joint influence of evolutionary and ecological factors on the functioning of multi-trophic interactions.

## Introduction

Predators strongly influence the structure and function of ecological communities by influencing prey density, distribution, and behavior which, in turn, have cascading effects on lower trophic levels (Sih *et al.* 1985; Beckerman, Uriarte & Schmitz 1997; Shurin *et al.* 2002; Schmitz, Krivan & Ovadia 2004; Suraci *et al.* 2016). This indirect effect of predators on non-adjacent lower trophic levels, the so-called "trophic cascades," are frequently observed in both aquatic and terrestrial ecosystems (Halaj & Wise 2001; Borer *et al.* 2005; Bruno & O'Connor 2005; Wu *et al.* 2011; Sanders, Kehoe & van Veen 2015). Trophic cascades are important drivers of the structure and dynamics of populations, communities, and ecosystems (Ripple *et al.* 2016) and have several implications for theoretical ecology, conservation biology, and ecosystem management (Post *et al.* 1999; Hulot *et al.* 2000; Estes *et al.* 2011). For instance, Delvin *et al.* (2015) showed that stocking fishes in fishless lakes decreases by a factor 10 the efflux rates of methane—an important greenhouse gas—by reducing zooplankton abundance, which in turn increases the abundance of methanotrophic bacteria. In another study, Schmitz *et al.* (2017) showed that the composition of the arthropod predator community and associated cascading effects on plant communities explain 41% of the variation in soil carbon retention across a human land-use gradient. Given the importance of trophic cascades, a major issue in ecology and conservation is to determine when and where trophic cascades occur, and what are the factors and mechanisms underpinning their strength.

Although the existence of trophic cascades has been widely demonstrated (Schmitz 2003; Romero & Koricheva 2011), most studies focused on whether cascades are more likely or stronger in some systems than others (Pace *et al.* 1999; Shurin *et al.* 2002), leading to a wealth of predictions about the relative strength of predator effects on plants among ecosystems with a particular focus on aquatic versus terrestrial ecosystems (Shurin *et al.* 2002; Borer *et al.* 2005). Although comparing cascade strength among systems is valuable, little is known about what causes variations in the magnitude of cascading effects within or among systems. In particular, the role of intraspecific trait variation for the occurrence and strength of trophic cascades remains largely unexplored (but see Post *et al.* 2008; Harmon *et al.* 2009; Jochum *et al.* 2012; Keiser *et al.* 2015; Royauté & Pruitt 2015; Start & Gilbert 2017). Most prior studies assumed that species identity or mean trait values adequately represent species interactions and their effects on community dynamics. This assumption is puzzling because it ignores the considerable intraspecific variation of traits (Benesh & Kalbe 2016), thereby overlooking a potentially important

determinant of species interactions, community structure and dynamics, as well as evolutionary responses to selective pressures (Roff 1997; Bolnick *et al.* 2011; Violle *et al.* 2012). Thus, knowledge of how intraspecific variation modulates bi- and tri-trophic interactions is crucial for better understanding and predicting the occurrence and strength of trophic cascades.

Different hypotheses have been proposed to explain variations in the occurrence and strength of trophic cascades in various types of ecosystems (Hulot *et al.* 2000; Polis *et al.* 2000; Borer *et al.* 2005). Hypotheses linked to habitat spatial heterogeneity, food web linearity or system productivity have received little support (Borer *et al.* 2005). On the contrary, interspecific variation in plant anti-herbivore defenses (Schmitz *et al.* 2000; Mooney *et al.* 2010), predator hunting mode and consumer efficiency (e.g., low metabolic costs, high consumption rate and population growth rate) can significantly affect trophic cascade strength (Romero & Koricheva 2011). In particular, high predator or herbivore efficiency increases cascade strength via high consumption rate of herbivores by predators, and plants by herbivores (Strong 1992; Polis 1999; Borer *et al.* 2005). Therefore, trophic cascade strength should depend on the herbivore's population growth rate in the absence of predators and on the predator efficiency in reducing herbivore density (Schmitz 1998; Borer *et al.* 2005). We thus expect density-mediated effects driven by intraspecific variation in herbivore population growth rate to be an important determinant of trophic cascade strength: the faster they growth, the stronger is the trophic cascade. Deviation from this prediction would stress the importance of considering intraspecific variation in other ecological traits and in evolutionary history of herbivores to better understand and predict cascade strength.

In this study, we experimentally investigated the effects of herbivore intraspecific trait variation on trophic cascade strength using a broad bean–aphid–ladybeetle system. The pea aphid *Acyrthosiphon pisum* Harris (Homoptera: Aphididae) feeds on many Fabaceae species and forms host-plant-associated populations ("host races" or "biotypes") that are genetically differentiated (Via 1999; Hawthorne & Via 2001; Peccoud *et al.* 2009) in a way that affects their performance on different host plants (Via 1999; Via, Bouck & Skillman 2000; Hawthorne & Via 2001). These different features make the pea aphid an interesting biological model to assess the influence of intraspecific trait variation and evolutionary divergence for trophic cascade strength. Here we conducted a full factorial laboratory experiment with six pea aphid clonal lineages (i.e. asexually reproducing aphid genetic lines) specialized either on alfalfa (*Alfalfa* biotype) or clover (*Clover* biotype) and exposed or not to a generalist predator. We first tested whether trophic cascade strength varies among aphid clonal lineages, and then investigated whether differences among lineages are best explained by density-mediated effects or by adaptive divergence

related to aphid host-plant specialization. Our study highlights the importance of accounting for intraspecific variation and evolutionary divergence to better understand and predict the strength of trophic cascades.

## Materials and methods

*Biological system*

The experimental system comprised a three level food chain: the predatory ladybeetle *Harmonia axyridis* Pallas (Coleoptera: Coccinellidae), the pea aphid *A. pisum*, and the broad bean *Vicia faba* L. cv. Aquadulce. The broad bean *V. faba* is the universal legume host on which all pea aphid biotypes can feed and successfully develop (Peccoud *et al.* 2009). Approximately 200 ladybeetle adults *H. axyridis w*ere collected in October 2015 near Auzeville Tolosane (43°32'N, 1°29'E, South of France), brought to the laboratory, reared in 5000-cm$^3$ plastic boxes, and fed three times a week an excess of pea aphids and pollen. Corrugated filter paper was added to each box to provide a suitable substrate for oviposition. *H. axyridis* eggs were collected three times a week and neonate larvae were reared in 175-cm$^3$ plastic boxes and fed pea aphids *ad libitum* until the beginning of the experiments. Stock colonies of 6 pea aphid clonal lineages (T9005, 10TV, T734, LL01, LSR1, and Oxford 683) were maintained in our laboratory at low density on broad bean grown from seeds (Ets Henrion s.a.; Belgium) in nylon cages (30 × 30 × 30 cm) for more than three months before the beginning of the experiments. All aphid lineages were free of any of the eight secondary symbionts reported in the pea aphid (Gauthier *et al.* 2015) (i.e. only harbour the obligate endosymbiont *Buchnera aphidicola*) to avoid potential confounding effects of variation in symbiont composition among aphid lineages. These lineages were selected from a large collection of clones maintained at INRA Rennes and their symbiotic status was checked using diagnostic PCR as described in Peccoud *et al.* (2015). Three lineages (T9005, 10TV, and T734) were of the *Clover* biotype and three (LL01, LSR1, and Oxford 683) of the *Alfalfa* biotype. We used a standard set of seven microsatellite loci to confirm that each lineage represented a unique genotype (clone) and that each belonged to the aphid biotype corresponding to the plant from which it was collected (Peccoud *et al.* 2009). All insects and plants were maintained in air-conditioned chambers (Dagard®) at 21 ± 1°C, 50–60% relative humidity, and under a 16L:8D photoperiod. These experimental conditions ensure the pea aphid reproduces only by apomictic parthenogenesis (i.e., offspring are clones of their mother).

*Experimental design*

In a full factorial laboratory experiment, we measured the effects of the 6 aphid clonal lineages and predators (presence or absence) on the fresh aboveground biomass and height of broad bean plants. At the onset of the experiment, three 8-day-old bean plants with two unfurled leaves were transplanted in 500 mL plastic pots containing 400 mL of fertilized soil substrate (®Jiffy substrates NFU 44-551), and then enclosed in transparent plastic cylinders (ø: 14 cm; h: 29 cm). They were watered every three days with 75 mL of tap water per pot. The top of the cylinder and two lateral openings were covered with mesh muslin for ventilation. Six parthenogenetic two-day-old adult female *A. pisum* were transferred to the upper leaves of the plants using a fine paintbrush, and allowed to acclimatize and reproduce for 24 h. Then, one second instar *H. axyridis* larva was introduced into each experimental cylinder of the predation treatment. Ten days later, all aphids were collected using a fine paintbrush and counted under a stereoscopic microscope. The ladybeetle larvae were isolated in small Petri dishes (50 × 9 mm) and starved for 24h to empty their gut before being weighed with a micro-balance ($10^{-7}$ g, SC2, Sartorius®). The plants were harvested, and their height and fresh aboveground biomasses measured. There were 20 replicates for each combination of aphid lineage and predator treatment (presence/absence) leading to a total of 240 replicates. 30 additional replicates without aphids and ladybeetles were performed as an insect-free control. As it was not possible to perform all replicates simultaneously, we conducted the experiment at three different dates with one third of the replicates of each treatment. For each date, we used the same methods and standardization of ladybeetle, aphid and plant age/stage/size. During the experiments, temperature and humidity were recorded continuously using Hobo U12 (Hobo®) units.

*Statistical analyses*

We performed the statistical analyses in two steps to (1) investigate whether trophic cascade strength differed among aphid lineages to test for the existence of intra-specific effects, and (2) determine whether the observed variations were linked to aphid biotype to test for the role of evolutionary divergence. For the first step, we analyzed the effects of predators, aphid lineage, and their interactions on plant fresh aboveground mass and height using Linear Mixed Models (LMMs) with experimental dates added as random effect. We next analyzed the effects of predators, aphid lineage, and their interactions on aphid density using a Generalized Linear Mixed Model (GLMM) with a Poisson distribution and a log link function, with experimental dates added as random effect. Finally, we analyzed the effects of aphid lineage on predator fresh body mass using a LMM with experimental dates added as random effect. The

significances of model fixed terms were assessed using Chi-tests from analyses of deviances, and post-

hoc Tukey tests were used to determine significant differences among means.

For the second step, we investigated the effects of aphid biotype, predators and their interaction on plant

biomass, plant height, and aphid density using LMM and GLMM models as described above, but adding

lineage identity as a random effect. We also analyzed the effects of aphid biotype on ladybeetle larva

body mass using LMM as described above, but adding lineage identity as a random effect. Aphid lineage

color or its interaction with the two other independent variables did not significantly affect the response

variables ($P > 0.05$) and was thus excluded from final analyses.

To better understand the links between plant response and aphid response to predators, we calculated,

for each aphid lineage, trophic cascade strength defined as the log response ratio of plants to predators:

$R_p = \ln(x_P/x_C)$, where $x_P$ and $x_C$ are the mean values of the plant trait (biomass or height) in the treatment

with and without predators, respectively (Hedges, Gurevitch & Curtis 1999). We also calculated this

ratio for aphid density ($R_a$) using the same formula. We next estimated the variance of each log ratio

estimate as $\text{var}(R) = s_P^2/(n_P x_P^2) + s_C^2/(n_C x_C^2)$, where $n$ and $s$ respectively denote the number of replicates

and the standard deviation in the treatments with predators $P$ and without predators $C$ (Hedges, Gurevitch

& Curtis 1999). We then calculated the 95% confidence intervals by multiplying $\text{var}(R)$ by 1.96 assuming

a normal distribution (Hedges, Gurevitch & Curtis 1999). A non-significant log ratio value (i.e. when its

95% confidence intervals overlap with zero) indicates the absence of predator effects whereas a positive

or negative log ratio value represents a positive or negative effect of predator on the lower trophic level

(aphids or plants), respectively (Hedges, Gurevitch & Curtis 1999).

To evaluate whether the effects of predators on plants and on aphids are positively related, we plotted

herbivore density log ratios against plant (biomass or height) log ratios. We next used a linear least

squares regression model to investigate the relationship between the direct effects of predators on aphids

and their indirect effects on plants. Finally, we investigated the effects of aphid lineage population growth

rate on the plant log ratios, aphid log ratios and average predator body mass using linear regression

models. The instantaneous population growth rate of each aphid population (in the absence of predators)

was calculated as $\ln(N_t/N_0)/t$ where $N_0$ is the initial aphid density (i.e. 6), $N_t$ is the final aphid density and

$t$ is the number of experimental days (i.e. 10).

To investigate the relationship between aphid biotype and the strength of predator effects on aphids and

plants, we calculated for each aphid biotype the mean aphid biotype population growth rate as well as

the plant and herbivore log ratios and their variances. We next plotted biotype log ratios against plant log

ratios, biotype population growth rate against plant log ratios, biotype population growth rate against

biotype log ratio, and biotype population growth rate against predator mass. We considered that log ratios

differ significantly if their 95% CIs do not overlap (Hedges, Gurevitch & Curtis 1999). LMMs and

GLMM were computed using the lme4 package (Bates *et al.* 2015), and analyses of deviance were

performed using the car package (Fox & Weisberg 2011). In cases where interaction terms of the LMMs

or GLMM were non-significant, these terms were removed prior to calculating significance values on

the main effects or other interactions. All analyses were performed using R 3.1.1 (R Development Core

Team 2017).

**Results**

*Influence of aphid lineage and biotype on predator body mass, herbivore density, and plant biomass*

*and height*

Ladybeetle body mass depended on aphid lineage ($\chi^2 = 55.14$, $df = 5$, $P < 0.0001$). Ladybeetle larvae

feeding on the aphid lineage LSR1 were about twice heavier than those feeding on the lineage T734 (Fig.

1a). Ladybeetle body mass significantly differed between aphid biotypes, with ladybeetles feeding on

aphids of the *Alfalfa* biotype being heavier than those feeding on the *Clover* biotype ($\chi^2 = 6.73$, $df = 1$, $P$

$= 0.009$, Fig 1b).

Aphid density varied strongly among lineages ($\chi^2 = 27370.6$, $df = 5$, $P < 0.0001$) with the highest density

for the LSR1 lineage and the lowest for the T734 lineage (Fig. 1c). Predators always significantly

decreased aphid density (Fig. 1c; $\chi^2 = 15175.4$, $df = 1$, $P < 0.0001$) although the strength of this effect

varied among lineages (significant interaction between lineage and predator treatment: $\chi^2 = 1827.9$, $df =$

$5$, $P < 0.0001$).

Aphid density significantly differed between biotypes ($\chi^2 = 10.99$, $df = 1$, $P = 0.0009$), and was affected

by the presence of predators ($\chi^2 = 15900.27$, $df = 1$, $P < 0.0001$) and by the interaction between predators

and biotype (Fig 1d; $\chi^2 = 1180.33$, $df = 1$, $P < 0.0001$). Aphid density was higher and predator effect on

aphid density stronger for the *Alfalfa* biotype than for the *Clover* biotype (Fig. 1d).

Plant biomass significantly varied among aphid lineages (Fig. 1e; $\chi^2 = 23.43$, $df = 5$, $P = 0.0003$), and

was affected by the presence of predators ($\chi^2 = 66.72$, $df = 1$, $P < 0.0001$) and by the interaction between

these two factors (Fig 1e; $\chi^2 = 13.57$, $df = 5$, $P = 0.0185$). Without predators, lineages 10TV and LSR1

had the strongest impact on plant fresh biomass, whereas with predators, lineages 10TV and T734 had

the weakest impact on plant fresh biomass (Fig. 1e). Predators indirectly increased plant biomass but this

increase depended upon the lineage with a large effect for 10TV and a weak non-significant one for

LSR1.

Plant biomass was significantly influenced by the predator treatments (Fig. 1f; $\chi^2 = 65.55$, $df = 1$, $P <$

$0.0001$), and by the interaction between predator treatments and aphid biotypes ($\chi^2 = 7.0221$, $df = 1$, $P =$

$0.0081$). Without predators, the *Clover* biotype had a stronger impact on plant biomass compared to the

*Alfalfa* biotype (Fig. 1f, blue dots). The positive indirect effect of predators on plant biomass was stronger

in plants exposed to the *Clover* than to the *Alfalfa* aphid biotypes (Fig. 1f, differences between red and

blue dots) indicating that the effect of predators on plant biomass depends on aphid biotypes. The effects

of aphid lineage or biotype, predators and their interactions on plant height were qualitatively similar

than their effects on plant biomass (see Fig. S1 and Text S1 for more details).

**Figure. 1. Influence of aphid lineage and biotype on the three trophic levels.** <u>Left column</u>: effects of the six aphid lineages (X axis) on mean (± SE) ladybeetle larva body mass (a), aphid density (c) and plant fresh biomass (e) (*n* = 20 replicates per treatment). Shaded area: aphid lineages of the *Clover* biotype; Non-shaded area: lineages of the *Alfalfa* biotype. <u>Right column</u>: effects of aphid biotype (X axis) on mean (± SE) ladybeetle larva biomass (b), aphid density (d) and plant fresh biomass (f) (*n* = 60 replicates per treatment). Red dots: with predators; Blue triangles: without predators. Within each panel, small or

capital letters denote significant differences ($P < 0.05$) among aphid lineages (panels a, c, e) or between aphid biotypes (panels b, d, f) within each predator treatment. For each lineage or each aphid biotype, an asterisk or "ns" denotes significant ($P < 0.05$) or non-significant ($P > 0.05$) predator effect (significance levels estimated with post hoc Tukey tests), respectively. Black lines of panels e and f represent mean ($\pm$ SE; dotted lines) plant fresh biomass in controls without aphids or ladybeetles.

*Relationship between the effects of predators on plants and their effects on aphids*

Predator direct effect on aphid density (i.e. herbivore density log ratio, $X$ axis in Fig. 2) was always significant as indicated by the non-overlap of log ratio confidence intervals with the intercept (plain vertical black line in Fig. 2a). The magnitude of this predator effect differed among aphid lineages and was minimal for the LSR1 lineage and maximal for the T734 lineage. Interestingly, aphid biotypes strongly influenced the predator direct effects on aphid density which was stronger for the *Clover* than the *Alfalfa* biotype (Fig. 2b).

Predator indirect effect on plant biomass varied significantly among lineages and did not always significantly differ from zero as for the LSR1 lineage ($Y$ axis in Fig. 2a). Moreover, predator indirect effect on plant biomass was significantly stronger for the *Clover* than for the *Alfalfa* biotype (Fig. 2b). The relationship between predator effects on plant biomass and on aphids was non-significant ($F_{(1,4)} = 3.80$, $P = 0.12$, $R^2 = 0.36$ Fig 2a). Finally, all data point cluster to the left of the 1:1 dotted line indicating strong attenuation of top-down effects at the plant level. The predator indirect effect on plant height as well as the influence of aphid lineages and biotype on this effect were qualitatively similar to these obtained for the plant biomass (see Fig. S2 and Text S2 for more details).

**Figure. 2. Relationship between the magnitude (log ratio ± 95% CI) of the predator effects on aphid density and on plant fresh biomass according to aphid lineage (a) and biotype (b).** Predator effect is significant if the confidence interval does not overlap zero (dark full lines). The dotted line shows the 1:1 relationship, representing equivalence of predator direct and indirect effects. If the data cluster to the left of the 1:1 line, then top-down effects are attenuating at the plant level; if they cluster to the right of the 1:1 line, then top-down effects are intensifying and, if they cluster along the 1:1 line, the effect magnitudes do not attenuate.

*Relationship between aphid population growth rate and predator effects on plants*

Although predator indirect effects on plant biomass tended to decrease with lineage population growth rate (Fig 3a), this relationship was non-significant (Fig. 3; $F_{(1,4)} = 3.68$, $P = 0.12$, $R^2 = 0.35$). Interestingly, predator indirect effects on plant biomass were stronger with *Clover* than with *Alfalfa* biotype despite the faster population growth rate of the later (Fig. 3b). The results for plant height were qualitatively similar to these for plant biomass (see Fig. S2 and Text S2 for more details).

Figure. 3. Relationship between aphid population growth rate (mean ± 95% CI) and the magnitude (log ratio ± 95% CI) of predator indirect effect on plant fresh biomass according to aphid lineage (a) and biotype (b). Predator effect is significant if the 95% CI does not overlap the X axis (dark full line).

*Influence of aphid population growth rate on herbivore density log ratio and on predator body mass*

Predator effect on aphid density (i.e. herbivore log ratio) was positively associated with aphid lineage population growth rate ($F_{(1,4)}$ = 132.96, $P$ = 0.000323, $R^2$ = 0.96; $y$ = 9.94$x$-6.62, Fig. 4a) showing that predators have a weaker effect on fast growing aphid lineages than on slow growing aphid lineages. Predator body mass was positively associated with aphid lineage population growth rate ($F_{(1,4)}$ = 20.29, $P$ = 0.01079, $R^2$ = 0.79; $y$ = 68.14 $x$-1.54, Fig. 4c) indicating that lineages with fast population growth result in larger predators than lineages with slow population growth. Interestingly, lineages of the *Clover* and *Alfalfa* biotypes clustered separately along the regression lines in figure 4a and c indicating that the influence of biotype on predator effect on aphids and on predator body mass are mainly linked to differences between biotype population growth rates. Grouping the data by aphid biotype (Fig. 4b and d) confirmed that population growth rate, predator effect on aphid density, and predator body mass differed between the two biotypes.

Figure. 4. Relationship between aphid population growth rate in absence of predators (mean ± 95% CI) and (first row) the direct effect of predators on aphid density (log ratio ± 95% CI) and (second row) predator body mass (mean ± 95% CI) according to aphid lineage (a, c) and biotype (b, d). In panel a and b, predator effect is significant if 95% CIs do not overlap the X axis (dark full line).

**Discussion**

Although intraspecific differences in organism phenotype and behaviour have clear implications for pairwise species interactions, their effects on higher order interactions remain largely unexplored (Bolnick *et al.* 2011; Toscano & Griffen 2014; Belgrad & Griffen 2016; Sanders *et al.* 2016). Here we quantified the impact of pea aphid clonal lineages (specialized on alfalfa or clover) on their universal legume host plant *Vicia fabae* in the presence or absence of a ladybeetle predator. We showed that trophic cascade strength strongly depends on intraspecific differences among herbivores and evolutionary divergence associated with host-plant specialization. Our study thus highlights the importance of intraspecific trait variation and evolutionary divergence as drivers of trophic cascade strength.

*Effects of herbivore intraspecific variation on trophic cascade strength and predator body mass*

We found that on average, predators decreased aphid population density by 66.03 % (± 0.24 %, 95 % CI), which, in turn, increased plant biomass by 16.29 % (± 3.72 %) and plant height by 20.18 % (±3.07 %). These values are within the range of values reported by previous meta-analyses on trophic cascades in terrestrial system (Schmitz *et al.* 2000; Shurin *et al.* 2002; Borer *et al.* 2005; Romero & Koricheva 2011) and confirm the previously described strong attenuation of the predator top-down effect down the food chain (Polis & Strong 1996; Borer *et al.* 2005). Nevertheless, our study indicates that considering only these average values limits our understanding of multi-trophic interactions as we found that trophic cascade strength strongly depends on intraspecific variations among herbivore lineages with cascading effect of predator on plants ranging from non-significant (0%) to an increase of 34% in plant biomass. These strong differences in trophic cascade strength mediated by herbivore intraspecific trait variation could contribute to explain why (1) previous studies had difficulties in assessing the strength and occurrence of trophic cascades (Schmitz *et al.* 2000; Halaj & Wise 2001), (2) the occurrence and strength of trophic cascades is so variable among studies, species, and habitats (Schmitz *et al.* 2000; Bell, Neill & Schluter 2003; Borer *et al.* 2005), and (3) no single hypothesis can explain variations in the magnitude of trophic cascades (Borer *et al.* 2005).

Interestingly, herbivore intraspecific variation did not only influence top-down effects but also climbed up the food chain and influenced the predator phenotype. Indeed, we found that predator body mass depends on which aphid clonal lineage and biotype they are feeding on. To the best of our knowledge, this is the first experimental evidence of a predator body mass being significantly influenced by the intraspecific herbivore prey specialization on particular host-plants. Different traits such as inter-biotype variations in defensive behaviour or palatability may explain the effects of aphid biotype on predator

body mass. Nevertheless, our results indicate that this effect is likely driven by the aphid population growth rate that strongly differs between aphid biotypes: fast growing aphid lineages support larger predators than slow growing lineages. Body size is a key trait that determines many ecological properties including fecundity, behaviour, population growth rate, trophic position, species interactions and community stability (Peters 1983; Brose *et al.* 2006; White *et al.* 2007). This implies that the effect of herbivore intraspecific variation and ecological specialization on predator body mass is likely to influence predator populations and thereby have long-term effects on the dynamics and structure of the community.

*Investigating the mechanisms underpinning the influence of intraspecific variation on trophic cascade*
An important step toward a better understanding of trophic cascade functioning is to explain how intraspecific differences at a given trophic level can influence adjacent trophic levels as well as predator indirect effects on plants (i.e. trophic cascade strength). As the conventional view is that trophic cascade strength strongly depends on the density of the interacting species (Schmitz *et al.* 2000; Schmitz, Krivan & Ovadia 2004; Borer *et al.* 2005), we hypothesized that differences in the population growth rate of aphid lineages would explain the intensity of lineages' impact on plant, predators, and trophic cascade strength. Accordingly, the predator direct effect on aphid density strongly depended on the lineages' population growth rate with fast growing lineages being less impacted by predators than slow growing lineages. Differential population growth rate among aphid biotypes thus explains why (1) predator direct effect on herbivore density is weaker for the *Alfalfa* than for the *Clover* biotype, and (2) ladybeetle larvae reach a larger body mass when feeding on the *Alfalfa* than on the *Clover* biotype (as mentioned above). We thus conclude that the ladybeetle-aphid interaction is strongly density-dependent and that the differential effects of aphid lineages or biotypes on this interaction are mainly linked to their differential population growth rate.

On the other hand, the direct effect of aphid lineages on plant biomass and height was not related to their population growth rate but was instead mainly linked to their host plant specialization. Surprisingly, plants were more impacted by the *Clover* than the *Alfalfa* biotype despite the faster population growth of the latter. This counter-intuitive result contradicts the herbivore efficiency hypothesis predicting that herbivores with the highest population growth rate should have the strongest effect on plants, which in turn should increase trophic cascade strength when predators are efficient in reducing herbivore populations (Borer *et al.* 2005). The differential effects of aphid biotype on plants could be linked to morphological, physiological and behavioural differences between aphid biotype (Via 1991; Kunert *et*

*al.* 2010) and/or linked to the plant defensive response against a given aphid biotype (Via 1991; Tétard-
Jones *et al.* 2007). For instance, biotype-specific aphid effectors injected while feeding may be
recognized differentially by the host plants and trigger more or less defense responses (Guy *et al.* 2016;
Sanchez-Arcos *et al.* 2016).

Whatever the exact mechanism driving the differential impact of aphid biotype on plants, we found that
the strength of trophic cascade strongly depends on herbivore biotypes and lineages, and is not directly
related to the predator effect on aphid density. Indeed, we found no straightforward relationship between
predator direct effect on herbivore density and its indirect effect on plant traits. This indicates that
herbivore intraspecific variation and evolutionary history of adaptive divergence play a stronger role in
determining trophic cascade strength than the density-dependent effects related to herbivore population
growth rate. More generally, herbivore intraspecific variation induced considerable changes in our
tritrophic system that could not be predicted from observations on a bitrophic system. We thus conclude
that going beyond pairwise interactions and considering the links between intraspecific trait variation
and evolutionary divergence is crucial to better understand multitrophic interactions.

*Ecological and evolutionary implications of herbivore intraspecific trait variation*
Herbivores are the hub between primary producers and higher trophic levels. Variations in herbivore
traits can thus have important consequences for the dynamics of ecological communities as shown by
previous studies focusing on pairwise interactions (Bolnick *et al.* 2011; Sentis, Morisson & Boukal
2015). Our results indicate that intraspecific variations in herbivore lineages and their ecological
specialization can also have important consequences for higher-order interactions and trophic cascade
strength. While the duration of our study was too short to measure feedback loops, we expect that the
contrasting effects of aphid biotype on plants and predators may feedback and have a long lasting effects
on predator and prey populations. For instance, smaller ladybeetles lay fewer (Dixon & Guo 1993; Bista
2013) and smaller eggs (Osawa 2005; Kajita & Evans 2010), which should in turn reduce top-down
pressure, thereby allowing for the larger growth of aphid populations. As a result, we would then expect
a stronger impact on plants which would then feedback on herbivore populations. We thus argue that
herbivore trait variation is likely to affect population dynamics on the longer term and should thus receive
more attention to better understand the structure and dynamics of ecological communities. More
generally, intraspecific variation at any trophic level might be influential and further studies are needed
to determine when and where intraspecific variation has the strongest influence on trophic cascade.

## Conclusion

Intraspecific variation is central to our understanding of evolution and population ecology, yet its consequences for community ecology are poorly delineated (Bolnick *et al.* 2011; Violle *et al.* 2012). Here, we showed that herbivore intraspecific variation influences the strength of trophic cascade. Interestingly, differences in trophic cascade strength were more related to aphid lineage and evolutionary divergence associated to host-plant specialization than to density-dependent effects mediated by aphid population growth rate. Our findings imply that intraspecific trait diversity and evolutionary adaptations are key drivers of trophic cascade strength and therefore they should not be overlooked to decipher the joint influence of evolutionary and ecological factors on the functioning of multitrophic interactions.

## Author contributions

A. S., R. B., E. D. and J-L. H. conceived and designed the experiments. A.S., R. B., N. D., and A. M. performed the experiments. A. S. analyzed the data and wrote the first draft of the manuscript. A. S., J-C. S., A. M., B. P., E. D., and J-L. H. contributed substantially to revisions.

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## Data accessibility statement

We confirm that, should this manuscript be accepted, the data supporting the results will be archived in an appropriate public repository such as Dryad or Figshare and the data DOI will be included at the end of the article.

## Additional information

Supplementary information is available for this paper

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**Supplementary information**

**Herbivore intraspecific variation and evolutionary divergence drive trophic cascade strength**

**Text S1. Influence of aphid lineage and biotype on plant height.**

Plant height varied among aphid lineages ($\chi^2 = 80.45$, $df = 5$, $P < 0.0001$), predator treatments ($\chi^2 = 49.94$,
$df = 1$, $P < 0.0001$), and the interaction between aphid lineage and predator treatments ($\chi^2 = 20.04$, $df = $
$5$, $P = 0.0012$). Without predators, lineages 10TV and LSR1 showed the strongest impact on plant height
whereas, with predators, lineage T734 had the weakest impact (Fig. S1a). Predator presence increased
plant height but the magnitude of this effect varied significantly among aphid lineages (Fig. S1a).

Plant height was significantly influenced by the interaction between predator treatments and biotypes ($\chi^2$
$= 13.71$, $df = 1$, $P = 0.0002$). Without predators, plant height diminished more for the *Clover* than the
*Alfalfa* biotypes (Fig. S1b). With predators, plant height did not significantly differ between biotypes
(Fig. S1b). Finally, the predator positive effect on plant height was stronger for plant exposed to the
*Clover* than to the *Alfalfa* biotypes (Fig. S1b).

**Figure S1. Plant height (mean ± SE) with (red dots) or without (blue triangles) predators according**
**to aphid lineage (a) and biotype (b).** Within each panel, letters denote significant differences ($P < 0.05$)
among aphid lineages (panel a) or aphid biotypes (panel b) with predators (small letters) or without
predators (capital letters). Asterisk or "ns" denote significant ($P < 0.05$) or non-significant ($P > 0.05$)
effect of predators within each lineage in (a) or aphid biotype in (b). Black lines represent mean (± SE;
dotted lines) plant height in controls without aphids or ladybeetles.

**Text S2. Relationship between the effects of predators on plant height and their effects on aphids**
Predator indirect effect on plant height varied significantly among lineages and was not always
significantly different from zero as for the OX683 and LL01 lineages (Fig. S2a). Moreover, predator
indirect effect on plant was significantly stronger for the *Clover* than for the *Alfalfa* biotype (Fig. S2b).
The relationship between predator effects on plants and on aphids was non-significant ($F_{(1,4)} = 1.55$, $P =$
$0.28$, $R^2 = 0.10$). Finally, all data point cluster to the left of the 1:1 dotted line indicating strong attenuation
of top-down effects at the plant level.

**Fig. S2. Relationship between the magnitude (log ratio ± 95% CI) of the predator effects on aphid**
**density and on plant height according to aphid lineage (a) and biotype (b).** Predator effect is
significant if the confidence interval does not overlap zero (dark full lines). The dotted line shows the
1:1 relationship, representing equivalence of predator direct and indirect effects. If the data cluster to the
left of the 1:1 line, then top-down effects are attenuating at the plant level; if they cluster to the right of
the 1:1 line, then top-down effects are intensifying and, if they cluster along the 1:1 line, the effect
magnitudes do not attenuate.

**Text S3. Relationship between aphid population growth rate and predator effects on plant height**

Although predator indirect effects on plant height tended to decrease with lineage population growth rate
(Fig S3a), this relationship was non-significant ($F_{(1,4)} = 1.20$, $P = 0.33$, $R^2 = 0.04$). Interestingly, predator
indirect effects on plant height were stronger with *Clover* than with *Alfalfa* biotype despite the faster
population growth rate of the later (Fig. S3b).

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659 **Fig. S3. Relationship between aphid population growth rate (mean ± 95% CI) and the magnitude**
660 **(log ratio ± 95% CI) of predator indirect effect on plant height according to aphid lineage (a) and**
661 **biotype (b)**. Predator effect is significant if the 95% CI does not overlap the X axis (dark full line).
662