A neural network for online spike classification that improves decoding accuracy

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Abstract

Objective. Separating neural signals from noise can help to improve brain-computer interface

performance and stability. However, "spike-sorting" suffers from a lack of a ground truth for classifying

waveforms, and most algorithms for separating neural action potentials from noise are not suitable for use

in real time and have shown mixed effects on decoding performance. Our goals were to automate the

intuition of human spike-sorters to operate in real time with an easily tunable parameter governing the

stringency with which spike waveforms are classified. *Approach*. We trained an artificial neural network

with one hidden layer on neural waveforms that were hand-labeled as either spikes or noise. The network

output was a likelihood metric for each waveform it classified, and we tuned the network's stringency by

varying the minimum likelihood value for a waveform to be considered a spike. Using the network's

labels to exclude noise waveforms, we decoded remembered target location from electrode arrays

implanted in prefrontal cortex (PFC) during a memory-guided saccade task in two rhesus macaque

monkeys. We assessed our network's performance by measuring how it influenced decoding accuracy.

Main results. The network classified waveforms in real time, and its classifications were qualitatively

similar to those of a human spike-sorter. Compared to decoding with all waveforms (i.e. threshold

crossings), we found that we could improve decoding performance, or at worst cause only a small change,

if we removed waveforms with low spike likelihood values. Furthermore, decoding with our network's

classifications became more beneficial as time since the array was implanted increased. *Significance*. Our

classifier serves as a feasible preprocessing step that could be applied to both offline neural data analyses

and online decoding. Additionally, we demonstrated that using our network's classifications for noise

removal could improve decoding performance with little risk of harm.

Introduction

Brain computer interfaces (BCIs) have been used as both research tools to understand neural

phenomena (1-4) and devices to improve patient control of prosthetics (5-7). BCIs interpret neural signals

arising from electrodes implanted in the cortex using real-time decoding algorithms; however, their

performance is limited by the difficulty of segregating individual neurons from extracellular voltage

signals ("spike-sorting"), the characteristics of the information present in individual neurons and multi-

unit activity (their "tuning"), as well as the nuances of interactions among neurons (8). Refining the raw

data is necessary to improve BCI performance, but identifying waveforms that contain relevant

information is challenging.

One common noise removal method is to use all waveforms crossing a minimum voltage

threshold for decoding, where all waveforms on a particular channel are considered to be from a single

neural "unit" (9, 10). Using threshold crossings also serves to reduce the amount of stored data. However,

threshold crossings often capture noise and the summed activity of multiple nearby neurons, called multi-

unit activity, on each channel (11, 12). Alternatively, since waveforms with the characteristic shape of an

action potential are the focus of most offline neural data analyses, a different approach is to isolate well-

defined spike waveforms from the threshold crossings via spike-sorting, and use only those waveforms

for decoding (9, 12). Unfortunately, most spike-sorting techniques are unsuitable for BCI use because

they are time-intensive and require manual refinement.

Furthermore, even with the development of real-time, automated spike-sorting algorithms (13,

14), there is no clear ground truth for what constitutes a spike waveform based solely on extracellular

recordings (12, 15-17). While some studies have found spike-sorted units improved decoding

performance (18-21), others have found that threshold crossings resulted in comparable or better decoding

that was more stable over time (10, 22-25). These conflicting results highlight the gap in our

understanding of what information in neural recordings is most valuable for BCIs. Even within studies,

there are inconsistent effects of spike-sorting on decoding, both between subjects and over time (10, 22).

It is also difficult to compare the effects of spike-sorting on decoding performance across studies due to

the inherent variability in spike-sorting techniques (12, 15-17) and differences in brain regions,

parameters being studied, and decoding algorithms that may influence which waveforms are the most

decodable (9, 22, 23, 26, 27).

52   Given the subjective nature of spike-sorting and its history of variable effects on decoding

performance, we sought to avoid explicitly sorting our data and instead developed an easily tunable spike

classifier that could objectively and efficiently assign waveforms to one of two classes: spike or noise. To

achieve this goal, we sought a classification approach that would (1) take advantage of the spike-sorted

waveforms from previous analyses that formed a readily available, labeled training data set; (2) output a

likelihood of being a spike for each waveform so we could simply adjust the minimum cutoff for binary

spike classification; (3) once trained could operate in real time. We built a neural network classifier that

satisfied these three criteria. To evaluate our classifier's performance, we decoded the activity of

electrode arrays implanted in prefrontal cortex (PFC), and compared decoding accuracy of task condition

with all threshold crossings to decoding accuracy with the waveforms selected by our classifier. We also

assessed how decoding performance changed as a function of the stringency of our classifier (i.e. the

minimum cutoff for spike classification) and explored how decoding accuracy with our classifier changed

as a function of time since the array implantation (array implant age).

65   By removing waveforms that the network identified as unlikely to be spikes, we improved

decoding performance for most sessions relative to decoding with all threshold crossings. Moreover, in

the remaining sessions there was no substantial detriment to decoding accuracy, meaning we could apply

our method with little risk of harming decoding performance. Our network classifications continued to

improve decoding accuracy in sessions recorded long after array implantation, even though the overall

decoding accuracy decreased and the array recordings became noisier. Thus, our real-time, tunable

waveform classifier demonstrates promise for long-term BCI applications and our findings highlight the

value of spike-sorted data as a training set to inform waveform classification.

74   **Materials and Methods**

We trained a neural network, using a database of spike-sorted waveforms, to assign waveforms to

a spike or noise class. We then tested the network's classifications with another data set that was

independent from the training set. To evaluate the network's classifications, we decoded task location

using only waveforms labeled as signal by the network and compared the accuracy to decoding accuracy

using all recorded waveforms. All experimental procedures were approved by the Institutional Animal

Care and Use Committee of the University of Pittsburgh and were performed in accordance with the

United States National Institutes of Health's *Guidelines for the Care and Use of Laboratory Animals*.

Neural recordings

We analyzed neural recordings from six adult male rhesus macaques (*Macaca mulatta*) which

had previously been spike-sorted for ongoing experiments in the laboratory. Data from four subjects were

used to train our spike-classifying neural network, and data from the remaining two subjects (Monkey Pe

and Monkey Wa) were used to assess decoding accuracy using the network's classifications. Raw

recordings from both 96-electrode 'Utah' arrays (Blackrock Microsystems, Salt Lake City, UT) and 16-

channel linear microelectrode arrays (U-Probe, Plexon, Dallas, TX) were band-pass filtered from 0.3 to

7,500 Hz, digitized at 30 kHz, and amplified by a Grapevine system (Ripple, Salt Lake City, UT). For

each recording session, a threshold was defined for each channel independently based on the root-mean-

squared voltage of the waveforms (V_{RMS}) recorded on that channel at the beginning of the session (i.e.

V_{RMS}*X, where X was a multiplier set by the experimenter). Each time the signal crossed that threshold, a

52-sample waveform segment was captured (i.e. a threshold crossing), with 15 samples prior to and 36

samples following the sample in which the threshold excursion occurred.

Offline spike-sorting

Waveform segments were classified as spike or noise waveforms using a custom, offline

MATLAB spike-sorting algorithm that used an automated competitive mixture decomposition method

(28). These automated classifications for each recording session were subsequently refined manually by a

researcher. While only a single researcher spike-sorted the waveforms from any particular session, the

data in the training set (7 sessions total) collectively consisted of data spike-sorted by four different

researchers. If multiple unique spike waveform shapes were present on a channel, the sorter would label

those as different units.

Training data set

We sought a diverse neural waveform training set that captured a variety of subjects, brain

regions, implanted array ages (i.e. number of days since the array was implanted) and recording devices

(Table 1). The four training data set subjects each had a 96-electrode 'Utah' array implanted in right or

left hemisphere visual area V4. Two of those subjects also had 16-channel U-probe recordings from the

frontal eye field (FEF), located in the anterior bank of the arcuate sulcus. Data from two sessions were

used for three of the subjects: one session recorded shortly after the array implant, and one session

recorded at least four months later. For the fourth subject, only a single recording session recorded shortly

after the array implant was used because there were no later recordings from that array. These recordings

were collected and sorted as part of previous studies which describe the experimental preparations in

detail (29-32). Data in the training set were assigned a label of 0 or 1. For each session, all waveforms

labeled as noise during spike-sorting were included in the training set (labeled as 0). For the spike

waveforms in the training set, only waveforms that were both labeled as spikes during spike-sorting and

from a channel with very distinct spike waveforms, defined as a channel with a signal-to-noise ratio

greater than 2.5 (33), were included in the training set (labeled as 1). The aim of these selections was to

emphasize relatively well-isolated single unit action-potential shapes while also exposing the network to a

variety of noise waveforms. If there were multiple single-units identified by the sorter on a particular

channel, they were not distinguished in the training set and all were treated as spike waveforms (labeled

as 1).

Overall, the network training set consisted of 24,810,795 waveforms from four monkeys, two

brain regions (V4 and FEF), two recording devices (Utah array and U-probe), and different array implant

ages. These waveforms were classified as spikes or noise using the offline spike-sorting technique

described above (Fig. 1a).

Table 1. Training data set information, including time since implant (array age), brain area (RH-right hemisphere, LH-left hemisphere), and number of recording sessions. Utah arrays contained 96 channels and U-Probes had 16 channels.

	Recording device	Brain area	Time since implant	# of sessions
Monkey Wi	Utah array	RH V4	~1 month	1
	Utah array	RH V4	~11 months	1
	U-Probe	RH FEF	-	1
Monkey Ro	Utah array	LH V4	<1 month	1
	Utah array	LH V4	~8 months	1
	U-Probe	LH FEF	-	1
Monkey Bo	Utah array	RH V4	<1 month	1
	Utah array	RH V4	~4 months	1
Monkey Bu	Utah array	RH V4	~1 month	1

Testing data set

We used recordings from two additional subjects, not included in the training set, for testing to

ensure that the trained network was generalizable to subjects not included in its training. The two testing

data set subjects (Monkey Pe and Monkey Wa) each had two 96-electrode 'Utah' arrays implanted, one in

visual area V4 and one in dorsolateral PFC (on the prearcuate gyrus just medial to the principal sulcus,

area 8Ar). Both arrays were implanted in the right hemisphere for Monkey Pe and in the left hemisphere

for Monkey Wa. Only data recorded from PFC were used for analyses (Table 2) because this provided a

matched data set from two monkeys for the purpose of assessing the impact of our method on decoding

and because it allowed us to test our network's generalizability in situations where the data were recorded

from a different region than the training set. However, we repeated some of the analyses in held out data

from V4 and included these results in the supplement (Supplementary Figure 4). Recordings within 50

days of the array implant were considered early sessions and recordings from over 50 days after the

implant were considered late sessions (see Table 2 for session counts by subject). For Monkey Pe, the

thresholds across channels were similar for all recording sessions (90% of channels had thresholds

between -39 and -20 µV, median: -29 µV). For Monkey Wa, the thresholds were more permissive for the

later recording sessions (> 6 months after the array implant) as the experiments sought to extract the

maximum remaining signal in arrays that were decreasing in recording quality. The median of Monkey

Wa's early sessions was -32 µV (90% between -43 and -23 µV), and the median of the later sessions was

-15 µV (90% between -29 and -12 µV). The waveforms in the testing data set were also spike-sorted

offline via the technique described above for a comparative analysis between our network classifications

and offline spike-sorting.

Table 2. Testing data set information, including time since implant (array age), brain area (RH-right hemisphere, LH-left hemisphere), and number of recording sessions.

	Recording device	Brain area	Time since implant	# of sessions
Monkey Pe	Utah array	RH PFC	11-43 days	14
		RH PFC	80-147 days	22
Monkey Wa	Utah array	LH PFC	27-40 days	5
		LH PFC	180-224 days	11

Neural Network

Using TensorFlow (34) and Keras (35), we developed a neural network to classify data segments

as spike or noise waveforms (Fig. 1b). The network accepts a waveform segment with '*s*' samples (*s*=52),

which passes through a hidden layer with '*n*' units (*n*=50) that uses a rectified linear unit (ReLU)

activation function. The product of the hidden layer passes through one output unit and then the network

applies a sigmoid function to its output.

With the aforementioned training waveforms, the network was trained to maximize accuracy

based on binary labels (0 for noise, 1 for spikes) using an Adam optimization algorithm (36) and a binary

cross-entropy loss function in batch sizes of 100.

For a single waveform input run through the trained network, the output was a value between 0

(likely not a spike) and 1 (likely a spike), which represented the network's assessment of the likelihood

that the input waveform was a spike. While this output value was not a conventional probability, we refer

to it as the probability of being a spike or P(spike) because it was a likelihood metric scaled between 0

and 1. Supplementary Figure 1 provides more intuition regarding the hidden layer units and how the

network assessed waveforms. In order to classify the waveforms using the network's output probability,

we set a minimum P(spike) for a waveform to be classified as a spike. We referred to this minimum

probability as the γ threshold. A γ threshold of 0 classified all of the waveforms that were captured for the

session as spikes; this is often referred to as 'threshold crossings' in the literature. Increasing γ resulted in

fewer waveforms classified as spikes because the P(spike) cutoff was higher.

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169   **Figure 1. Methods of classifying waveforms: classic (manual) spike-sorting and our neural network spike**
170   **classifier, Not A Sorter (NAS). (a) Spike-sorted channels with waveforms that were manually classified as**
171   **spikes (green) and noise (grey). The network was trained on 24,810,795 spike-sorted neural waveforms from**
172   **four monkeys. The waveforms were recorded on arrays implanted in V4 as well as U-probes placed in FEF.**
173   **(b) Neural network structure and output for three sample waveforms. Each s=52 waveform input (I) was**

passed through a hidden layer (II) with n=50 units. The resultant linear weighting of the waveform voltages was passed through a rectified linear unit (ReLU) non-linearity (III). The output was again passed through a weighted sum (IV) followed by a sigmoid non-linearity (V). The resulting value was the network's assessment of the likelihood that the input waveform was a spike waveform (VI). We referred to this value as P(s) or the "probability" of being a spike.

Custom Python and MATLAB scripts used to train the neural network and classify waveforms, as well as sample training and testing data, will be available on GitHub at the time of publication. We refer to our neural network as Not A Sorter (NAS), because it classifies waveforms as spikes or noise but does not attempt to sort the spikes into separable single units.

Memory-guided saccade task

The two testing data set subjects (Monkey Pe and Monkey Wa) performed a memory guided saccade task (Fig. 2a). Each subject fixated on a central point for 200 ms. Then, a target stimulus flashed at a set amplitude and direction for 50 ms, followed by a 500 ms delay. There were a total of 40 possible conditions: the radius could be one of five eccentricities and the target direction could be one of eight (spaced in 45° steps). After the delay, the fixation point disappeared instructing the subject to make a saccade to the location where the target flashed. For some sessions, during the saccade the target reappeared to help the subject; however, for all analyses we only used data from the beginning of the delay period (i.e. prior to the saccade). Subjects were rewarded with water or juice for making a saccade to the correctly remembered target location. For all decoding analyses, the decoded condition was the target location. Trials at a single eccentricity (Monkey Pe: 9.9°, Monkey Wa: ~7.6°) were used to maximize the number of recording sessions that could be compared. Thus, there were eight unique target locations to decode.

Only rewarded trials were analyzed. The number of trial repeats per condition varied over sessions. Monkey Pe had an average of 43 repeats per session (range: 25-75, sd: 17) and Monkey Wa had an average of 62 repeats per session (range: 51-83, sd: 11) for each target condition.

Decoding

To classify target direction prior to movement from PFC waveforms, we used a Poisson Naïve Bayesian decoder (Fig. 2b). All decoding analyses were performed offline. Waveforms classified as

spikes from 300 ms to 500 ms after fixation (i.e. 50 ms after stimulus offset) were used for decoding,

while waveforms classified as noise were discarded. Each channel was considered a single decodable unit

with one exception: to calculate decoding accuracy using manual spike-sorting, if there were multiple

single-units identified by the manual spike-sorting method on a channel, each single-unit on that channel

remained separate for the purposes of decoding. Units with an average firing rate across trials below 0.25

spikes per second were discarded. We allocated training and test trials using two different methods. For

one analysis, we assigned 80% of the trials from each session to the training set and the remaining 20% to

the test set (Fig. 4a,b). For the remaining analyses, we wanted two test sets, so we used 60% of the trials

for training and 20% for each of the two test sets. The two test sets allowed us to search for an optimal γ

threshold using the first test set and cross validate this value with the second test set. Training and test

data were rotated such that each trial was used for a test set only once.

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Figure 2. Offline decoding of planned direction from a memory guided saccade task. (a) Memory guided
saccade task. The monkey fixated on a central point. A brief stimulus flash occurred followed by a delay
period. When the fixation point turned off, the subject was required to make a saccade to the location of the
previously flashed stimulus. (b) Decoding paradigm. The stimulus flash could occur at 8 different angles
around the fixation point. A Poisson Naïve Bayes classifier was used to decode stimulus direction offline using
spikes recorded from a 96-electrode Utah array in PFC 50 ms after target offset. For each recording session,
we used 5-fold cross validation, where for each fold the data were split (step 1) into a training set to create
model distributions for each direction (step 2) and an independent testing set to test the accuracy of the

model's predictions (step 3). Note that the curves in step (2) do not depict actual distributions and simply
represent how a Poisson decoder could use spike counts from trials in the training set to distinguish between
different target conditions.

Our decoding algorithm created a Poisson distribution model for each target location (θ) using

the average spike count for each unit (n_{spike}) in the training set. For each test trial, the target location

with the maximum prediction probability, $P(\theta|n_{spike})$, was the predicted label. In equation (1),

$P(n_{spike}|\theta)$ was calculated using the Poisson model developed with the training trials:

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$$P(\theta|n_{spike}) = \frac{P(n_{spike}|\theta) * P(\theta)}{P(n_{spike})}$$

230   Decoding accuracy was calculated as the ratio of the number of correct prediction labels to the

231 total number of predictions. We used 5-fold cross validation and computed the average decoding accuracy

232 across folds.

234   **Results**

We trained a one-layer neural network called Not a Sorter (NAS) to evaluate the likelihood that a

neural waveform was a spike. We used a diverse set of waveforms in the training set from different brain

regions, subjects, and array implant ages (time since array implant) in order to expose the network to a

variety of waveform types. Each waveform in the training set was assigned a binary label of noise (0) or

spike (1) via offline spike-sorting with manual refinement by researchers. The network was also exposed

to variability in spike classification due to manual sorting because different researchers sorted different

sets of waveforms in the training data. The network learned to assess how spike-like a waveform was

based on binary labels, but itself output a continuous value between 0 and 1 for each waveform allowing

for tunable classification. We referred to this output as the probability of being a spike or P(spike). The

network classified 1000 waveforms in less than one millisecond on average (computed on a 2011 iMac

with a 2.8 GHz Intel Core processor). In a 10 ms bin, 1000 waveforms would be the expected output of

1000 channels each with a firing rate of 100 spikes/s. Thus, our network could be easily integrated to

operate within real-time computing constraints.

Qualitative assessment of NAS classifications

To assign each NAS-classified waveform a binary spike or noise label, we set a parameter called

the γ threshold, which was the minimum P(spike) a waveform needed to be considered a spike waveform.

We found that even a low γ threshold, such as $\gamma=0.20$ (Fig 3a), assigned most waveforms to classes that a

human spike-sorter would deem appropriate. Most spike-sorters and spike-sorting algorithms search for

waveforms with a canonical action potential shape – an initial voltage decrease followed by a sharp

increase, a narrow peak, and a return to baseline. Increasing the γ threshold to 0.70 (i.e. only waveforms

assigned a P(spike) > 0.70 were considered spikes) mimicked the effect of a more selective spike-sorter

(Fig 3b), where the percentage of spike waveforms on the channel decreased since more waveforms were

placed in the noise class and the remaining spike waveforms had a clearer single-unit shape. Considering

all of the waveforms from this same channel, the range of P(spike) values coincided well with our

subjective impression of the match of individual waves with a canonical action potential shape (Fig 3c),

and this was also true across all channels from this array (Fig 3d). Thus, tuning the γ threshold from low

values (near 0) to high values (near 1) resulted in a shift from a more permissive to a more restrictive

regime.

Figure 3. Classifying waveforms based on their probability of being a spike, P(spike), by setting γ, a tunable parameter. (a-b) Waveforms classified as spikes (green) or noise (grey) for a sample channel in Monkey Pe. The network outputs a value between 0 and 1, referred to as P(spike), for each waveform, where a value close to 1 means the network identifies that waveform as very likely to be a spike waveform. After running waveforms through the network, we set the minimum P(spike) to classify waveforms as spikes and referred to this value as the γ threshold. In (a) only waveforms assigned a spike probability greater than 0.20 (i.e. γ threshold = 0.20) and in (b) greater than 0.70 (i.e. γ threshold = 0.70) were classified as spikes. Increasing the γ threshold by definition results in a smaller percentage of waveforms captured in the spike class. (c) Waveforms from the same channel in (a) and (b) colored based on their network assigned P(spike) value. For a waveform where $\gamma_1 <$ P(spike) $< \gamma_2$, the waveform would be classified as a spike when the threshold is γ_1, but would be labeled as noise for γ_2. (d) Similar to (c) except the average of waveforms across all channels within the indicated P(spike) ranges. Modifying the γ threshold tuned the stringency of the spike classifications.

Quantitative assessment of NAS classifications with decoding

Although by eye the network appeared to classify waveforms reasonably well, there was no available ground truth data with which to assess its performance. Instead, we sought to objectively assess its performance by measuring its effect on offline decoding accuracy. Specifically, we hypothesized that decoding using spikes classified by our network would be better than using threshold crossings. We

applied our network to a set of data recorded from PFC on which the network had not been trained. First,

we set the γ threshold and discarded any waveforms with a P(spike) below the threshold. Next, we split

the data into training and testing sets. We trained the decoder using the training set to decode the task

condition (the remembered location, out of 8 possibilities) during the delay period of a memory-guided

saccade task on each trial (see Methods). Then, we used the trained decoder to assess decoding accuracy

in the test set(s). We repeated this process for the same data set using multiple γ thresholds between 0

(i.e., all threshold crossings were considered spikes) and 0.95 (i.e., only waveforms assigned a 0.95 or

greater probability by the network were considered spikes). Since only waveforms classified as spikes

were used for decoding, as the γ threshold increased, fewer waveforms remained for decoding.

290   Initially, we used 80% of trials to train the decoder and the remaining to test it. We analyzed

cross-validated decoding accuracy as a function of γ (sample recording sessions in Fig. 4a,b). The

decoding accuracy using threshold crossings (γ = 0) for the sample recording sessions in Fig. 4a and 4b

was close to chance levels (1 out of 8, 12.5%) for Monkey Pe (12.8%) and was also low for Monkey Wa

(20.1%). Using the network's classifications improved decoding accuracy relative to using threshold

crossings. The peak decoding benefit of using the network's spike classifications occurred at a low γ

threshold (Monkey Pe: γ = 0.16, Monkey Wa: γ = 0.16). Being very selective about the spikes used for

decoding (by setting a high γ threshold) did not have a large impact on decoding accuracy, which

appeared to plateau after the peak. We found that increments of the γ threshold did not remove the same

number of waveforms. A substantial proportion of waveforms were removed at the lowest γ threshold

tested (especially in Monkey Pe), and small increments of the γ threshold as it approached a value of 1

could also result in large increases in the proportion of removed waveforms (especially notable in Figure

4b).

303   Ideally, for the sake of simplicity during online BCI experiments, we could select a single γ

threshold and use it for all recording sessions. We wanted to find the lowest γ threshold that improved

decoding accuracy for the majority of recording sessions. We focused on lower γ thresholds because it

was a conservative approach, and because our intuition from single channels and results in example

sessions (Figure 4) showed it could have the greatest benefit for decoding. In order to ensure the γ

threshold we selected was cross-validated, for the remaining analyses we created two test sets for

decoding (training set: 60% of trials, test set 1: 20% of trials, test set 2: 20% of trials). The first test set

was used to determine the optimal value of the γ threshold and the second to determine its impact on

decoding. For each session, using the first test set, we found the maximum cross-validated decoding

accuracy at any γ threshold, and then searched for the lowest γ threshold that resulted in a decoding

accuracy within 99% of that maximum (Fig. 4c, d).

Since the optimal γ threshold varied between sessions, we computed the median across sessions

of these γ thresholds chosen with our cross-validation approach and rounded it to the nearest tested value

(Monkey Pe: $\gamma = 0.2$, Monkey Wa: $\gamma = 0.08$). We then used that median value as the γ threshold for all

sessions in the second test set and analyzed the change in decoding accuracy from decoding using

threshold crossings in that same test set, which we termed Δ % decoding accuracy (Fig. 4e, f). Combining

sessions from both animals, the average improvement in decoding accuracy was 3.9% (2-tailed, Wilcoxon

signed rank test; across subjects: $p < 0.0001$, Monkey Pe: $p < 0.0001$, Monkey Wa: $p = 0.057$). Thus, our

network's classifications tuned to the previously described γ thresholds resulted in a net benefit for

decoding performance across sessions compared to using threshold crossings. Although we could tune the

γ threshold for each session to maximize the decoding, our choice of a fixed γ threshold was more

consistent with the use of our network in an online decoding context, where it would be desirable to set a

constant γ threshold at the start of each session rather than tuning it as a free parameter. However, an

alternative strategy would be to collect a small data set at the start of each day to find the optimal gamma

value, and then continue experiments for the remainder of that day using the chosen value.

Given the ability of our network to improve decoding accuracy beyond that observed with

threshold crossings, we took advantage of our longitudinal recordings in a fixed paradigm to understand

how our network's performance varied as a function of time. Since there is mixed evidence in the

literature regarding decoding stability with long-term array implants, we were specifically interested in

how the passage of time since array implant (array implant age), and concomitant degradation of

recording quality, could influence our network's performance.

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Figure 4. Using NAS spike classifications improved decoding accuracy in many sessions. (a-b) Percent
decoding accuracy (black line) and percent of waveforms removed (maroon line) at different γ thresholds for
an example recording session in Monkey Pe and Wa. The decoding accuracy increased for low γ thresholds
and then reached an asymptote as γ increased. At the highest γ thresholds, decoding accuracy fell to chance
(black, horizontal dotted line). Other labeled relevant metrics include: decoding accuracy with threshold
crossings (blue dot) maximum decoding accuracy (green dot), γ threshold with the maximum decoding
accuracy (grey dotted line), and Δ % decoding accuracy (the difference for a given session between the

decoding accuracy using the network classifications at a particular gamma threshold and the decoding
accuracy with threshold crossings). (c-d) Distribution of γ thresholds across sessions (Monkey Pe: N =36,
Monkey Wa: N = 16) that resulted in the maximum decoding accuracy for each session. (e-f) Distribution of Δ
% decoding accuracy across all sessions where the γ threshold was the median from the distributions in (c-d).
Mean Δ % decoding accuracy shown in red. For both monkeys, using spikes classified by the network
improved decoding accuracy on average across sessions; however, this improvement was only statistically
significant for Monkey Pe (2-tailed, Wilcoxon signed rank test, Monkey Pe: p < 0.0001, Monkey Wa: p =
0.057).

Impact of array age

First, we assessed how the overall quality of our neural data changed as a function of time. As the

time since the array implant increased, the percentage of waveforms that were assigned a very low

probability of being a spike (P(spike) < 0.02) also increased (Fig 5a; Spearman's rank order, Monkey Pe:

$\rho = 0.87$, $p<0.0001$; Monkey Wa: $\rho = 0.90$, $p<0.0001$). Conversely, the percentage of waveforms assigned

higher probabilities of being a spike (P(spike) > 0.70) decreased over time (Fig 5b; Monkey Pe: $\rho = -0.87$,

$p<0.0001$; Monkey Wa: $\rho = -0.94$, $p<0.0001$). These classification changes were consistent with our

qualitative observations that the arrays showed an increasing proportion of multi-unit activity (relative to

single-unit activity) and apparent noise on the array over time. We further found that the ratio of the

percentage of waveforms assigned a high P(spike) to the percentage of waveforms assigned a low

P(spike) served as a proxy for a signal to noise ratio metric, and it was highly correlated with the median

signal to noise ratio (across channels) of the array in each session (Supplementary Figure 2). Both signal

to noise metrics decreased over time, and the decoding accuracy with threshold crossings also decreased

the longer the array had been implanted (Fig 5c; Monkey Pe: $\rho = -0.86$, $p<0.0001$; Monkey Wa: $\rho = -0.69$,

$p=0.004$).

Given the change in distribution of the types of waveforms present during the session over time,

we tried to optimize the γ threshold based on time since array implant. We used the same method as in

Fig. 4c-d to compute the γ thresholds with approximately maximum decoding (within 1%), except we

calculated the median of the early sessions (recorded 0-50 days post implant) and the median of the late

sessions (recorded >50 days post implant) separately. This did not prove to be a useful optimization as the

median values were similar between early and late sessions in both Monkey Pe ($\gamma_{early} = 0.25$, $\gamma_{late} = 0.18$)

and Monkey Wa ($\gamma_{early} = 0.06$, $\gamma_{late} = 0.08$). Decoding accuracy compared to threshold crossings (Δ %

decoding accuracy) was significantly increased in Monkey Pe (Figure 5d, 2-tailed, Wilcoxon signed rank

test, $p < 0.0001$) and was not significantly helped or hurt in Monkey Wa ($p = 0.09$). In line with the trends

in signal and noise over time, the decoding accuracies from later sessions (> 50 days post implant) were

helped more by the network classifications in both subjects than those from earlier sessions (Figure 5d,

Wilcoxon rank sum, Monkey Pe: $p = 0.01$; Monkey Wa: $p = 0.01$). When we normalized the decoding

accuracy at each γ threshold by the decoding accuracy using threshold crossings for each session and

separately averaged across early and late sessions, the increased benefit of using the network in the later

sessions compared to the earlier sessions was clear (Figure 5e).

Altogether, these results provide additional evidence that factors such as time since the array was

implanted and signal to noise ratio influence decoding accuracy and affect how noise removal impacts

decoding performance. Despite decreasing signal quality and increasing noise waveforms, the effect of

using our network classifications for noise removal prior to decoding was consistent in that it was most

often beneficial for decoding and at worst minimally detrimental. We also confirmed that these results

were not affected by poor γ threshold selection by using the maximum decoding accuracy regardless of γ

threshold to calculate Δ % decoding accuracy (Supplementary Figure 3a), nor were the results

substantially influenced by variability in the number of trials across sessions (Supplementary Figure 3b).

Lastly, we assessed whether the benefits of our network also held in another brain region (V4) recorded

from different Utah arrays but in the same sessions from the same animals in the test data set, and found

similar results (Supplementary Figure 4).

Figure 5. Noise on the array increased with the age of the array while decoding accuracy decreased in
Monkey Pe (blue) and Monkey Wa (orange). (a) Percentage of waveforms with a P(spike) < 0.02 over time. A
waveform with a P(spike) less than 0.02 was one that the network found very unlikely to be a spike. The
percentage of these unlikely spike waveforms increased as the array became older (Spearman's rank-order,
Monkey Pe: ϱ = 0.87, p < 0.0001; Monkey Wa: ϱ = 0.90, p < 0.0001). (b) Percentage of waveforms with a
P(spike) > 0.70 over time. The percentage of waveforms that the network found to be strongly spike-like
decreased as the array became older (Spearman's rank-order, Monkey Pe: ϱ = -0.87, p < 0.0001; Monkey
Wa: ϱ = -0.94, p < 0.0001). (c) Decoding accuracy with threshold crossings (i.e. γ = 0) decreased as the array
aged (Spearman's rank-order, Monkey Pe: ϱ = -0.86, p < 0.0001; Monkey Wa: ϱ = -0.69, p = 0.004). (d)
Change in percent decoding accuracy using network-classified spikes relative to decoding accuracy with

threshold crossings (Δ % decoding accuracy). We computed a distribution of the maximum γ threshold (similar to Fig. 4c-d) for sessions that were 0-50 days post array implant and used the median to set the γ threshold before computing decoding accuracy for those sessions. We repeated this for sessions more than 50 days post array implant. In Monkey Pe, using NAS classifications improved decoding accuracy (2-tailed, Wilcoxon signed rank test, p < 0.0001) and in Monkey Wa, using the classifications neither hurt nor helped decoding significantly (p = 0.09). In both subjects, the network helped decoding more in the late array sessions (>50 days post implant) compared to the early sessions (2-tailed, Wilcoxon rank sum, Monkey Pe: p = 0.01; Monkey Wa: p = 0.01). (e) Mean normalized decoding accuracy across early sessions (open circles) and late sessions (filled circles) as a function of γ threshold. Using the network classifications at any γ threshold greater than zero was more helpful for late sessions than it was for early sessions.

Comparison to spike-sorting

In light of the mixed benefits of spike-sorting on decoding in the literature (10, 18, 22, 25), we sought to evaluate the relative merits of manual spike-sorting and our network classifier on decoding accuracy in our data. We took advantage of the offline spike-sorting that had already been applied to these data and calculated the decoding accuracy of spike-sorted data (Fig. 6, data aggregated across subjects in the marginal histograms). Decoding using spike-sorted waveforms was better than using threshold crossings (Fig. 6 right marginal; 2-tailed, Wilcoxon signed rank test, $p < 0.0001$) similar to decoding with our network classifications (Fig. 6 top marginal, data from Fig. 5d aggregated across subjects; $p < 0.0001$). Our network's classifications were at least as helpful as manual spike-sorting for decoding relative to using threshold crossings (Fig. 6 center; paired, 2-tailed, Wilcoxon signed rank test, Monkey Pe: $p = 0.75$; Monkey Wa: $p = 0.09$). Furthermore, the automated real-time operation of our network confers a distinct advantage over manual spike-sorting.

Together, our analyses demonstrate that our network was able to produce classifications of spikes and noise that matched our qualitative expectations and could be tuned quantitatively to adjust its permissiveness. Compared to using threshold crossings, our network's classifications most often led to an improvement or minimally harmful change in decoding accuracy, indicating that there was usually little downside to the application of our network. These outcomes are valuable particularly in the context of a BCI where real time operation and maximal decoding accuracy are prized.

Figure 6. Using NAS classifications was comparable to using spike-sorted data for decoding. Δ % decoding
accuracy was calculated for both NAS classifications and spike-sorted waveforms as the change from
decoding accuracy with threshold crossings. Top: Distribution of Δ % decoding accuracy using NAS
classifications aggregated across subjects (γ threshold selected as in 5d). Using NAS classifications improved
decoding accuracy from that with threshold crossings (2-tailed, Wilcoxon signed rank test, p < 0.0001). Right:
Distribution of Δ % decoding accuracy using manual spike-sorting aggregated across subjects. Using spike-
sorted classifications also improved decoding accuracy from that with threshold crossings (2-tailed, Wilcoxon
signed rank test, p < 0.0001). Center: The joint distribution of Δ % decoding accuracy with spike-sorting and
NAS classified spikes. Using our network's classifications was at least as helpful as spike-sorting for decoding
(paired, 2-tailed, Wilcoxon signed rank test, Monkey Pe: p = 0.75; Monkey Wa: p = 0.09).

Discussion

By leveraging our pool of previously spike-sorted data, we trained a neural network classifier to

separate spikes from noise in extracellular electrophysiological recordings in real time. To assess the

value of this method, we used an objective criterion – the ability to decode remembered location from

prefrontal cortex in a memory-guided saccade task. We found that we could set a fixed γ threshold that

resulted in an improvement or at worst a small change in decoding accuracy for the majority of sessions,

and this effect persisted even as recording quality decreased over time. Compared to spike-sorting

(performed offline with substantial manual labor involved), classifying spikes with our network had

similar effects on decoding accuracy relative to threshold crossings and was less tedious.

Training the network

Developing a neural network trained on spike-sorted data allowed us to capture the essence of the

rules by which experienced researchers distinguish spikes from noise without explicitly defining all the

variables and exceptions, a standard challenge in spike classification (15). We found that a simple neural

network was quite good at capturing the nuances of the spike-sorted training set. In fact, the network was

sensitive enough to capture a researcher's specific spike-sorting tendencies, such that networks trained on

a particular individual's spike-sorted data performed better on classifying test data from that individual

than from other individual spike-sorters. Lee *et al*. (13) similarly demonstrated the promise of using

spike-sorted data as a training set for retinal recordings with dense arrays. Our findings extend this work

in a context where the training data were less extensively curated (spike-sorting performed by multiple

researchers), the data were noisier (cortical recordings in an alert macaque monkey) and the recording

electrodes were spatially sparse (at least 150 μm apart for U-probes and 400 μm for Utah arrays), a

situation that is common in numerous electrophysiology laboratories. A recent related effort (37) used a

neural network trained on hand-sorted spiking data to determine whether channels contained any spikes

for the purpose of rejecting channels with only artifacts. Our work is synergistic with their findings, and

extends them to demonstrate that removal of noise from single channels can lead to a positive impact on

decoding accuracy.

There were potentially misclassifications in our training set, as human spike-sorting is both

subjective and prone to error (16, 17). However, the network was trained on many waveforms to

minimize the impact of any confounders, including waveforms from multiple subjects, brain regions, and

human spike-sorters. An ideal and even more generalizable training set might include waveforms from

different research groups, artificially simulated neural waveforms and noise in which ground truth could

be established, and/or pre-defined quantities of certain types of spikes (e.g. shifted spikes, slow and fast

spike waveforms) and noise (e.g. electrical artifacts) to target what the network learns. However, no

matter how much training data is used, an ideal training set could only be defined in the context of

objective metrics to assess the trained network's performance.

Evaluating the network with decoding

Since there is no obvious ground truth for classifying spikes from extracellular array recordings,

one objective measure with practical benefit for BCI performance is how well we could decode

information from neural activity. We used independent test data to assess the effect of classifications from

our trained network on decoding accuracy. Although the expectation might be that removing noise from

neural data would improve decoding, historically doing so with spike-sorted classifications has had mixed

effects, in some cases hurting performance (18). Nevertheless, many BCIs currently operate with some

amount of online noise removal or visual inspection (such as disabling visibly noisy channels) to pre-

process neural data before decoding (1, 2, 38, 39). The question of whether to spike-sort BCI data prior to

decoding or simply use all threshold crossings has been a source of debate with no clear resolution.

Our findings provide additional support, across multiple sessions over time, that removing certain

noise waveforms prior to decoding provides an advantage over using all threshold crossings, consistent

with findings from Fraser *et al.* (22) and Christie *et al.* (10). Contrary to our results, another study by

Todorova *et al.* (18) found that using spike-sorted waveforms hurt decoding when noise waveforms were

discarded. It is difficult to speculate why removing noise waveforms hurt their result but helped ours

given that our methods of noise removal were different. Todorova *et al*. also found assigning noise

waveforms from spike-sorting to a new unit, rather than discarding them, improved decoding relative to

threshold crossings (18). Yet, when we assigned our noise waveforms to separate units, decoding

accuracy was worse or unchanged compared to discarding noise waveforms both with our spike-sorted

classifications and our network classifications (Supplementary Figure 5). Importantly, we defined 'noise'

operationally as any waveform that did not exceed the γ threshold of our network setting. This definition

of noise surely included both electrical artifacts as well as indistinct shapes that were actually neural in

origin. Removal of waveforms from the former class could only help decoding, but removal of the latter

class might actually harm decoding if the multiunit activity contained in those waveforms has information

about the condition of interest for decoding.

Apparent noise in neuronal activity, in addition to changing the activity of single electrode

channels, can also be correlated across channels. In the activity of pairs of single neurons, such

fluctuations in trial-to-trial correlated variability are sometimes termed "noise correlation" (also known as

spike count correlation, or r_{sc}). Noise correlation can have a substantial impact on the ability of a decoder

to extract information from neuronal populations (Averbeck et al, 2006, Kohn et al, 2016), and the

activity of multi-unit groups of neurons has a higher noise correlation than that of the constituent pairs of

neurons (Cohen and Kohn, 2011). Since our network acted on threshold crossings, which typically

contain the activity of many individual neurons, it likely also impacted the magnitude of the noise

correlation between channels. Our decoder assumed that the noise on each unit was independent, a choice

common to BCI, which permitted training the decoder with a smaller quantity of trials than is necessary to

learn the covariance structure in the population. The effect of our network on decoding surely depends on

not only the structure of the network and the choice of γ threshold, but also the structure of noise present

in the population and the sensitivity of the decoder to that noise.

In addition to understanding how the type of noise present in the data impacts decoding, it may

also be useful to evaluate the types of neural waveforms that contribute to decoding. Different levels of

single and multi-unit activity are captured during a neural recording depending on the threshold set by the

researcher (i.e. the minimum voltage level at which the neural activity is marked as a spike). By adjusting

this threshold, we change the candidate waveforms available for decoding. Oby *et al.* (26) found that the

optimal threshold depended on the type of information being extracted from neural activity. Another

study found that higher thresholds, which captured less multi-unit activity, resulted in worse decoding of

direction from M1 (10). These studies in the context of our own findings highlight the need to identify

specific types of waveforms that contribute to the decodable information in different brain regions and

task contexts.

Other variables that impact decoding accuracy

A key concern for practical implementation of BCIs is decoding stability. Array recordings often

get visibly noisier over time, potentially as a result of an immune response to the implant (40) or a

physical shift in position (41). In our study, as the time since array implant increased there were more

"low probability" waveforms and fewer "high probability" waveforms, indicating the array had fewer

well-defined single-unit waveforms. In line with this, several other studies with chronic array implants

have found that the number of single-units decreased over time (42-45). Nevertheless, there are mixed

findings on the stability of decoding performance in chronic array implants. Some studies have found

decoder performance with threshold crossings remained stable over time (23, 24, 46, 47); however, other

studies, including our own, observed a decrease in decoding accuracy over time (48, 49). Unlike the

previous literature investigating the effects of threshold crossings and spike-sorting on decoding

performance, we used neural activity from PFC, where decoding accuracy is typically lower (50-56) than

in motor cortex (7, 57-59). Stability of decoding accuracy with array age may also depend on the brain

region of the implant, the subject's task, initial decoding performance, and the choice to train the decoder

daily (23, 24) or hold it constant (46). The beneficial effects of our network on decoding were particularly

salient in the case of older array implants, where decoding accuracy had decreased over time.

Extensions for NAS

Our neural network-based spike sorter is a promising tool for both offline spike-sorting and

improving online decoding performance. However, in designing our network we only scratched the

surface of many potential avenues to address these challenges. We found that using networks with

different numbers of hidden units and layers did not substantially alter decoding accuracy even though

there were some differences in how these different sized networks classified waveforms. Given that our

network was relatively simple, it would be possible to implement similar operations with alternative

algorithms such as logistic regression. We chose a neural network because it was easily trainable from

existing data and there are many ways to modularly build upon its complexity. Although we opted to use

50 hidden units and one hidden layer, it is possible that a more complex network with additional filtering

operations and more categories of waveform classification may result in improved decoding performance

and could help to create a more robust spike-sorter or a pre-processing step for offline analyses. However,

our work had the value of exploring how a smaller network trained on spike-sorted data could identify

features that were most valuable for decoding. While training on spike-sorted data was an efficient choice

because of its easy availability, it may not be the best choice in regimes where canonical spike-shapes,

which spike-sorted classifications prioritize, do not necessarily carry the most decodable information

compared to less well-defined multi-unit activity (11, 23). If the goal is to improve decoding performance,

it might be valuable to train a network to select waveforms or units that positively contribute to decoding.

Ventura and Todorova (19) developed a method for identifying the information contribution of units for

decoding. Such a method could be used to develop a training set of neural waveforms that are assigned a

label based on how much they positively or negatively contributed to decoding.

Overall, we developed a new tool for pre-processing BCI data that classified threshold crossings

in a tunable manner that was beneficial for decoding. Furthermore, we showed that spike-sorted

waveform data are an underused training set with potential for building spike classification networks. A

neural network-based spike classifier has the potential to reduce the need for human intervention in spike-

sorting while maintaining sensitivity to nuances that many automated sorting algorithms miss. Our

tunable classifier is a step toward pre-processing methods that both optimize and stabilize online decoding

performance.

569   **Acknowledgements**

D.I. was supported by National Science Foundation (NSF) DBI 1659611, National Institutes of Health

(NIH) T32 GM008208, and the ARCS foundation Thomas-Pittsburgh Chapter Award. M.A.S. was

supported by NIH R01EY022928, R01MH118929, R01EB026953, and P30EY008098; NSF NCS

1734901; a career development grant and an unrestricted award from Research to Prevent Blindness; the

Eye and Ear Foundation of Pittsburgh. R.C.W. was supported by a Richard King Mellon

Foundation Presidential Fellowship in the Life Sciences (RCW). S.B.K. was supported by T32

EY017271-08. We would like to thank Samantha Schmitt for assistance with data collection and Ben

Cowley, Byron Yu, and Steve Chase for their valuable insight on this project.

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References

1. Sadtler PT, Quick KM, Golub MD, Chase SM, Ryu SI, Tyler-Kabara EC, et al. Neural constraints
on learning. Nature. 2014;512(7515):423-6.
2. Chase SM, Kass RE, Schwartz AB. Behavioral and neural correlates of visuomotor adaptation
observed through a brain-computer interface in primary motor cortex. J Neurophysiol. 2012;108(2):624-
44.
3. Golub MD, Yu BM, Chase SM. Internal models for interpreting neural population activity during
sensorimotor control. Elife. 2015;4.
4. Schafer RJ, Moore T. Selective attention from voluntary control of neurons in prefrontal cortex.
Science. 2011;332(6037):1568-71.
5. Velliste M, Perel S, Spalding MC, Whitford AS, Schwartz AB. Cortical control of a prosthetic
arm for self-feeding. Nature. 2008;453(7198):1098-101.
6. Hochberg LR, Bacher D, Jarosiewicz B, Masse NY, Simeral JD, Vogel J, et al. Reach and grasp
by people with tetraplegia using a neurally controlled robotic arm. Nature. 2012;485(7398):372-5.
7. Collinger JL, Wodlinger B, Downey JE, Wang W, Tyler-Kabara EC, Weber DJ, et al. High-
performance neuroprosthetic control by an individual with tetraplegia. Lancet. 2013;381(9866):557-64.
8. Averbeck BB, Latham PE, Pouget A. Neural correlations, population coding and computation.
Nat Rev Neurosci. 2006;7(5):358-66.
9. Lewicki MS. A review of methods for spike sorting: the detection and classification of neural
action potentials. Network. 1998;9(4):R53-78.
10. Christie BP, Tat DM, Irwin ZT, Gilja V, Nuyujukian P, Foster JD, et al. Comparison of spike
sorting and thresholding of voltage waveforms for intracortical brain-machine interface performance. J
Neural Eng. 2015;12(1):016009.
11. Stark E, Abeles M. Predicting movement from multiunit activity. J Neurosci. 2007;27(31):8387-
94.
12. Rey HG, Pedreira C, Quian Quiroga R. Past, present and future of spike sorting techniques. Brain
Res Bull. 2015;119(Pt B):106-17.
13. Chung JE, Magland JF, Barnett AH, Tolosa VM, Tooker AC, Lee KY, et al. A Fully Automated
Approach to Spike Sorting. Neuron. 2017;95(6):1381-94 e6.
14. Chaure FJ, Rey HG, Quian Quiroga R. A novel and fully automatic spike-sorting implementation
with variable number of features. J Neurophysiol. 2018;120(4):1859-71.
15. Wood F, Black MJ, Vargas-Irwin C, Fellows M, Donoghue JP. On the variability of manual spike
sorting. IEEE Trans Biomed Eng. 2004;51(6):912-8.
16. Pedreira C, Martinez J, Ison MJ, Quian Quiroga R. How many neurons can we see with current
spike sorting algorithms? Journal of Neuroscience Methods. 2012;211(1):58-65.
17. Rossant C, Kadir SN, Goodman DFM, Schulman J, Hunter MLD, Saleem AB, et al. Spike sorting
for large, dense electrode arrays. Nat Neurosci. 2016;19(4):634-41.
18. Todorova S, Sadtler P, Batista A, Chase S, Ventura V. To sort or not to sort: the impact of spike-
sorting on neural decoding performance. J Neural Eng. 2014;11(5):056005.
19. Ventura V, Todorova S. A computationally efficient method for incorporating spike waveform
information into decoding algorithms. Neural Comput. 2015;27(5):1033-50.
20. Kloosterman F, Layton SP, Chen Z, Wilson MA. Bayesian decoding using unsorted spikes in the
rat hippocampus. J Neurophysiol. 2014;111(1):217-27.
21. Santhanam G, Sahani M, Ryu S, Shenoy K. An extensible infrastructure for fully automated spike
sorting during online experiments. Conf Proc IEEE Eng Med Biol Soc. 2004;6:4380-4.

22.   Fraser GW, Chase SM, Whitford A, Schwartz AB. Control of a brain-computer interface without
spike sorting. J Neural Eng. 2009;6(5):055004.
23.   Chestek CA, Gilja V, Nuyujukian P, Foster JD, Fan JM, Kaufman MT, et al. Long-term stability
of neural prosthetic control signals from silicon cortical arrays in rhesus macaque motor cortex. J Neural
Eng. 2011;8(4):045005.
24.   Gilja V, Nuyujukian P, Chestek CA, Cunningham JP, Yu BM, Fan JM, et al. A high-performance
neural prosthesis enabled by control algorithm design. Nat Neurosci. 2012;15(12):1752-7.
25.   Dai J, Zhang P, Sun H, Qiao X, Zhao Y, Ma J, et al. Reliability of motor and sensory neural
decoding by threshold crossings for intracortical brain-machine interface. J Neural Eng.
2019;16(3):036011.
26.   Oby ER, Perel S, Sadtler PT, Ruff DA, Mischel JL, Montez DF, et al. Extracellular voltage
threshold settings can be tuned for optimal encoding of movement and stimulus parameters. J Neural Eng.
2016;13(3):036009.
27.   Bishop W, Chestek CC, Gilja V, Nuyujukian P, Foster JD, Ryu SI, et al. Self-recalibrating
classifiers for intracortical brain-computer interfaces. J Neural Eng. 2014;11(2):026001.
28.   Shoham S, Fellows MR, Normann RA. Robust, automatic spike sorting using mixtures of
multivariate t-distributions. J Neurosci Methods. 2003;127(2):111-22.
29.   Khanna SB, Snyder AC, Smith MA. Distinct Sources of Variability Affect Eye Movement
Preparation. J Neurosci. 2019;39(23):4511-26.
30.   Snyder AC, Morais MJ, Kohn A, Smith MA. Correlations in V1 are reduced by stimulation
outside the receptive field. J Neurosci. 2014;34(34):11222-7.
31.   Snyder AC, Morais MJ, Willis CM, Smith MA. Global network influences on local functional
connectivity. Nat Neurosci. 2015;18(5):736-43.
32.   Snyder AC, Smith MA. Stimulus-dependent spiking relationships with the EEG. J Neurophysiol.
2015;114(3):1468-82.
33.   Kelly RC, Smith MA, Samonds JM, Kohn A, Bonds AB, Movshon JA, et al. Comparison of
recordings from microelectrode arrays and single electrodes in the visual cortex. J Neurosci.
2007;27(2):261-4.
34.   Abadi M, Agarwal A, Barham P, Brevdo E, Chen Z, Citro C, et al. TensorFlow: Large-Scale
Machine Learning on Heterogeneous Distributed Systems. 2016.
35.   Chollet F. Keras. 2015.
36.   Diederik P. Kingma JB. Adam: a Method for Stochastic Optimization. International Conference
on Learning Representations2015.
37.   Saif-Ur-Rehman M, Lienkamper R, Parpaley Y, Wellmer J, Liu C, Lee B, et al. SpikeDeeptector:
A deep-learning based method for detection of neural spiking activity. J Neural Eng. 2019.
38.   Homer ML, Nurmikko AV, Donoghue JP, Hochberg LR. Sensors and decoding for intracortical
brain computer interfaces. Annu Rev Biomed Eng. 2013;15:383-405.
39.   Hochberg LR, Serruya MD, Friehs GM, Mukand JA, Saleh M, Caplan AH, et al. Neuronal
ensemble control of prosthetic devices by a human with tetraplegia. Nature. 2006;442(7099):164-71.
40.   Ward MP, Rajdev P, Ellison C, Irazoqui PP. Toward a comparison of microelectrodes for acute
and chronic recordings. Brain Res. 2009;1282:183-200.
41.   Perge JA, Homer ML, Malik WQ, Cash S, Eskandar E, Friehs G, et al. Intra-day signal
instabilities affect decoding performance in an intracortical neural interface system. J Neural Eng.
2013;10(3):036004.
42.   Dickey AS, Suminski A, Amit Y, Hatsopoulos NG. Single-unit stability using chronically
implanted multielectrode arrays. J Neurophysiol. 2009;102(2):1331-9.
43.   Downey JE, Schwed N, Chase SM, Schwartz AB, Collinger JL. Intracortical recording stability in
human brain-computer interface users. J Neural Eng. 2018;15(4):046016.
44.   Fraser GW, Schwartz AB. Recording from the same neurons chronically in motor cortex. J
Neurophysiol. 2012;107(7):1970-8.

45. Tolias AS, Ecker AS, Siapas AG, Hoenselaar A, Keliris GA, Logothetis NK. Recording
chronically from the same neurons in awake, behaving primates. J Neurophysiol. 2007;98(6):3780-90.
46. Nuyujukian P, Kao JC, Fan JM, Stavisky SD, Ryu SI, Shenoy KV. Performance sustaining
intracortical neural prostheses. J Neural Eng. 2014;11(6):066003.
47. Flint RD, Scheid MR, Wright ZA, Solla SA, Slutzky MW. Long-Term Stability of Motor Cortical
Activity: Implications for Brain Machine Interfaces and Optimal Feedback Control. J Neurosci.
2016;36(12):3623-32.
48. Perge JA, Zhang S, Malik WQ, Homer ML, Cash S, Friehs G, et al. Reliability of directional
information in unsorted spikes and local field potentials recorded in human motor cortex. J Neural Eng.
2014;11(4):046007.
49. Wang D, Zhang Q, Li Y, Wang Y, Zhu J, Zhang S, et al. Long-term decoding stability of local
field potentials from silicon arrays in primate motor cortex during a 2D center out task. J Neural Eng.
2014;11(3):036009.
50. Meyers EM, Freedman DJ, Kreiman G, Miller EK, Poggio T. Dynamic population coding of
category information in inferior temporal and prefrontal cortex. J Neurophysiol. 2008;100(3):1407-19.
51. Rizzuto DS, Mamelak AN, Sutherling WW, Fineman I, Andersen RA. Spatial selectivity in
human ventrolateral prefrontal cortex. Nat Neurosci. 2005;8(4):415-7.
52. Tremblay S, Doucet G, Pieper F, Sachs A, Martinez-Trujillo J. Single-Trial Decoding of Visual
Attention from Local Field Potentials in the Primate Lateral Prefrontal Cortex Is Frequency-Dependent. J
Neurosci. 2015;35(24):9038-49.
53. Jia N, Brincat SL, Salazar-Gomez AF, Panko M, Guenther FH, Miller EK. Decoding of intended
saccade direction in an oculomotor brain-computer interface. J Neural Eng. 2017;14(4):046007.
54. Boulay CB, Pieper F, Leavitt M, Martinez-Trujillo J, Sachs AJ. Single-trial decoding of intended
eye movement goals from lateral prefrontal cortex neural ensembles. J Neurophysiol. 2016;115(1):486-
99.
55. Parthasarathy A, Herikstad R, Bong JH, Medina FS, Libedinsky C, Yen SC. Mixed selectivity
morphs population codes in prefrontal cortex. Nat Neurosci. 2017;20(12):1770-9.
56. Spaak E, Watanabe K, Funahashi S, Stokes MG. Stable and Dynamic Coding for Working
Memory in Primate Prefrontal Cortex. J Neurosci. 2017;37(27):6503-16.
57. Sadtler PT, Ryu SI, Tyler-Kabara EC, Yu BM, Batista AP. Brain-computer interface control
along instructed paths. J Neural Eng. 2015;12(1):016015.
58. Masse NY, Jarosiewicz B, Simeral JD, Bacher D, Stavisky SD, Cash SS, et al. Non-causal spike
filtering improves decoding of movement intention for intracortical BCIs. J Neurosci Methods.
2014;236:58-67.
59. Koralek AC, Jin X, Long JD, 2nd, Costa RM, Carmena JM. Corticostriatal plasticity is necessary
for learning intentional neuroprosthetic skills. Nature. 2012;483(7389):331-5.
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Supplementary Figure 1. Weights for three sample hidden units (n1, n2, n3) of the neural network hidden layer and their outputs. To demonstrate how the network classified waveforms, we used 1000 example input waveforms from each of three different categories (bottom three panels: mean and standard deviation of the waveforms in each category). The first category included very spike-like waveforms (i.e. those that were assigned a P(spike) value between 0.95 and 1 by the network, green), the second included moderately spike-like waveforms (0.50 < P(spike) < 0.55, blue), and the third included noise (0<P(spike)<0.05, grey). To visualize the hidden layer, we selected three out of the 50 hidden units and plotted their weights (left panels, $W_{hidden\ layer}$). Each waveform was linearly scaled by the weight vector of units n1, n2, and n3 and summed (middle panels: mean and standard deviation of the scalar outputs of $W_{hidden\ layer}$ x waveform; waveform category indicated by bar color and column, and unit # indicated by row). The output of the hidden layer was passed through a ReLU non-linearity before being multiplied by an output weight (right, $W_{output\ layer}$). The resultant values were summed and passed through a sigmoid non-linearity to produce a P(spike) value (not shown).

Overall, the network's hidden units were difficult to characterize; however, we observed that certain hidden units tended to have a larger, positive response to very spike-like waveforms (e.g. n1 green bar) while others had a larger, positive response to noise waveforms (e.g. n3 grey bar). The positive $W_{output\ layer}$ for n1 would then weigh the final summed output towards a P(spike) of 1 and the negative $W_{output\ layer}$ for n3 would weigh the output towards a P(spike) of 0. Unlike n1 and n3, n2 had variable responses to different types of waveforms, indicating that not every unit had a well-defined spike or noise preference. Overall, we found that different units in the hidden layer focused on features that were associated with different types of spikes in the training data. Note that in the network, the contribution of bias values was not shown in this visualization.

Supplementary Figure 2. SNR metrics decreased as the array aged for Monkey Pe (blue) and Monkey Wa (orange). **(a)** SNR was computed for each channel on the array using the method described in Kelly et al. (2007). The median SNR of the array decreased over time in both subjects (Spearman's rank-order, Monkey Pe: ϱ = -0.77, p < 0.0001; Monkey Wa: ϱ = -0.72, p = 0.002). **(b)** SNR_{NAS} (i.e. the ratio of the percentage of waveforms with a P(spike) > 0.70 to the percentage of waveforms with a P(spike) < 0.02) served as an alternate measure of SNR as it was correlated with median SNR over time (Spearman's rank-order, Monkey Pe: ϱ = 0.78, p < 0.0001; Monkey Wa: ϱ = 0.75, p = 0.01).

Supplementary Figure 3. Change in percent decoding accuracy from that with threshold crossings (Δ % decoding accuracy) over time, as in Figure 5 except **(a)** using the maximum decoding accuracy for each session and **(b)** keeping the number of trials used to decode constant across all sessions. In (a) the maximum decoding accuracy across all γ thresholds greater than 0 on each session was used to show the best possible network performance if γ was chosen separately in each session (and tested in a cross-validated fashion). Δ % decoding accuracy was significantly greater than zero for both subjects across all sessions (2-tailed, Wilcoxon signed rank test, Monkey Pe: $p < 0.0001$; Monkey Wa: $p = 0.01$). There was a significant difference between the early array (0-50 days post implant) and the late array (> 50 days post implant) sessions in Monkey Wa, but not in Monkey Pe (2-tailed, Wilcoxon rank sum, Monkey Pe: $p = 0.09$; Monkey Wa: $p = 0.01$). In (b), decoding accuracy was computed for each session with 25 trials for Monkey Pe and 50 trials for Monkey Wa. The γ threshold was selected using the same method described for Fig. 4d. Δ % decoding accuracy again was greater than zero for both subjects across all sessions (2-tailed, Wilcoxon signed rank test, Monkey Pe: $p < 0.0001$; Monkey Wa: $p = 0.002$). Again, there was a significant difference between the early array (0-50 days post implant) and the late array (> 50 days post implant) sessions in Monkey Wa, but not in Monkey Pe (2-tailed, Wilcoxon rank sum, Monkey Pe: $p = 0.68$; Monkey Wa $p = 0.001$).

Supplementary Figure 4. The network had a similar effect on decoding from V4 96-electrode Utah array implants in each animal as it did on decoding from the PFC array in the same sessions from the same animals (Monkey Pe- blue, Monkey Wa- orange). This analysis was done using the same steps as in Figure 5 except using waveforms from the V4 array instead of the PFC array. Monkey Wa did not have any usable late session (> 50 days post implant) data from the V4 array. As a result, only Monkey Pe was tested for trends over time.

(a) Percentage of waveforms with a P(spike) < 0.02 over time. A waveform with a P(spike) less than 0.02 was one that the network found very unlikely to be a spike. The percentage of these unlikely spike waveforms only slightly but significantly increased as Monkey Pe's array became older (Spearman's rank-order, $\varrho = 0.43$, $p = 0.01$). **(b)** Percentage of waveforms with a P(spike) > 0.70 over time. The percentage of waveforms that the network found to be strongly spike-like decreased as Monkey Pe's array became older ($\varrho = -0.48$, $p = 0.003$). The magnitude of these changes over time was much smaller in V4 compared to PFC. **(c)** Decoding accuracy with threshold crossings (i.e. $\gamma = 0$) decreased by a small amount as the array aged in Monkey Pe ($\varrho = -0.41$, $p = 0.01$). **(d)** Change in percent decoding accuracy using network classified spikes relative to decoding accuracy with threshold crossings (Δ % decoding accuracy). We computed a distribution of the maximum γ threshold (using the same method as in Figure 4c-d) for sessions that were 0-50 days post array implant and used the median to set the γ threshold before computing decoding accuracy for those sessions. We repeated this for sessions more than 50 days post array implant. In Monkey Pe, using NAS classifications improved decoding accuracy (2-tailed, Wilcoxon signed rank test, $p < 0.0001$) and in Monkey Wa, using the classifications neither hurt nor helped decoding significantly ($p = 0.81$). There was a significant difference between the early array (0-50 days post implant) and the late array (> 50 days post implant) sessions in Monkey Pe (2-tailed, Wilcoxon rank sum, $p = 0.0496$).

Thus, despite differences in the array quality between PFC and V4 (as demonstrated by the higher threshold crossing decoding accuracies in V4 compared to PFC in Fig. 5), using our network classifications still proved beneficial in one subject and had no significant impact on the other subject.

Supplementary Figure 5. Assigning waveforms classified as noise on each electrode to a new unit rather than discarding them did not improve decoding performance in most sessions for Monkey Pe (blue) or Monkey Wa (orange). In the main text analyses, any waveform classified as noise via our network or via spike-sorting was discarded. However, Todorova et al. (2014) found that creating a separate noise unit for the noise waveforms from each unit actually led to better decoding performance than discarding the noise. We sought to assess whether this result held for our data. **(a)** Using the manually spike-sorted classifications, we added a new unit to accompany each spike unit that contained all waveforms classified as noise (plotted on the y-axis). Percent decoding accuracy was worse with noise units for Monkey Pe (paired, 2-tailed, Wilcoxon signed rank test- $p < 0.0001$) and not significantly different for Monkey Wa ($p = 0.19$). **(b)** Similar to (a), except using NAS classifications. For each session, we discarded the noise units and found the γ threshold that resulted in the maximum decoding accuracy. We classified waveforms as spikes or noise using that threshold and then computed decoding accuracy with and without the noise units for each session (for the latter, the test set was independent from the set used to select the γ threshold). Discarding noise resulted in a higher decoding accuracy for most sessions in both subjects (Monkey Pe: $p < 0.0001$, Monkey W: $p = 0.049$). Thus, unlike Todorova et al. (2014), we did not find preserving the noise as separate units to be beneficial for decoding. In fact, the noise removed from most of the later recording sessions (>50 days post implant) was typically harmful for decoding.