Symptoms expression of bakanae disease following seed treatment with 1 phytohormone and metabolites 2 3 4 5 6 Shireen A. Jahan Quazi^{1*}, Sariah Meon^{2¶}, Zainal Abidin B.M. Ahmad ^{2¶} and Hawa Jaafar ^{3¶} 7 ¹Plant Pathology Division, Bangladesh Rice Research Institute, Gazipur, Bangladesh 8 ²Department of Plant Protection, Universiti Putra Malaysia, Serdang, Selangor, Malaysia 9 ³Department of Crop Science, Universiti Putra Malaysia, Serdang, Selangor, Malaysia 10 11 12 *Corresponding author 13 14 Email address: shireenbrri@yahoo.com (QSAJ) 15 16 ¶ These authors contributed equally to this work 17 18 19

20 Abstract

21 Gibberellic acid (GA₃) phytohormone responsible for bakanae disease development is well-known and identified. But a number of secondary metabolites along with GA₃, produced by 22 the causal pathogen in relation to bakanae symptoms expression were unknown. Therefore, the 23 aims of this research were to evaluate the symptoms expression analysis of bakanae disease by 24 pre-seed treatment with pure (synthetic) phytohormones and metabolites in susceptible rice 25 variety MR 211. The typical bakanae symptoms were evaluated by applying pure GA₃, FA and 26 MON either singly or in mixtures. It was confirmed that higher concentration of GA₃ singly or 27 with higher concentration of GA₃ and MON in mixtures, caused unusual elongation of 28 internodes. Plants became stunted when high concentration of FA was applied. Browning of 29 leaves and stems, crown rot, root necrosis occurred and root length was decreased when mixtures 30 of higher concentration of FA *MON*GA₃ were used as pre-treatment. Similar observations 31 32 were noted in plants inoculated with F. proliferatum at different score levels. The mechanisms of bakanae disease development through different symptoms expression in susceptible variety
 infected with *F. proliferatum* were identified.

35

36 Introduction

Fusarium fujikuroi, causal agent of bakanae disease is known to produce gibberellic acid 37 38 (GA₃), fumonisin (FB1), monilifornin (MON), fusaric acid (FA) and beauvericin (BEA) in diseased plant [1-4]. GA₃ is a growth promoting phytohormone but abnormal production of GA₃ 39 has been identified as a break-through in disease development and disease resistance or 40 41 susceptibility in plants [5]. In addition, GA₃ has been identified as being responsible for increase in plant height, whereas FA is responsible for decrease in plant height. Fungal metabolites MON, 42 FB1 and BEA have been identified for causing phytotoxicity in plants rather than establishment 43 of pathogenicity or disease symptoms expression in bakanae diseased plants [1, 6-7]. Besides 44 Fusarium fujikuroi, Fusarium proliferatum is also identified as a causal agent of bakanae disease 45 46 [8] and GA_3 FB1, MON, FA were also isolated from bakanae diseased plants infected with F. proliferatum [9]. It was also assumed that GA₃ MON and FA had strong influence on bakanae 47 symptoms expression as a significant amounts of GA₃ MON and FA were isolated from the 48 49 infected susceptible variety (data not presented here). Therefore, this study was carried out to verify the role of GA₃, MON and FA on bakanae symptoms expression, following exogenous 50 51 pre-treatment on seeds of susceptible variety MR 211 instead of inoculation with F. proliferatum 52 over time.

53

54

55 Materials and methods

57 Soil mixture

Sand 40%, clay 30% and peat (PEATGRO) 30% were mixed well and sterilized at 120
°C for 90 min. Trays (28 cm x 21 cm x 6.5 cm) were filled with 2 kg of the sterilized soil
mixture and used for sowing pre-germinated seeds.

61 *Chemicals used*

Pure (Synthetic) GA_{3} , FA, and MON were used in this study. These chemicals were chosen based on positive relationship that were observed in relation to bakanae symptoms expression in susceptible variety MR 211 (data not presented here). Two concentrations of each chemical ($GA_{3}=15 \ \mu g/g$, $GA_{3}=10 \ \mu g/g$, FA= 400 $\mu g/g$, FA= 100 $\mu g/g$, MON= 120 ng/g, MON= 80 ng/g) were used in this study. The pure chemicals GA_{3} , FA, and MON were purchased from Sigma-Aldrich.

68

69 Experimental layout and design

70 Pre-germinated seeds (soaked in water for 48 h) of susceptible variety MR 211 were soaked for 12 h in the probation hormone (GA_3) and metabolite (FA and MON) solutions at two 71 concentrations singly to find out the effective concentration suitable for causing infection or 72 73 symptoms expression. This moderate pre-treatment period (12 h) was chosen in order to avoid the toxicity caused by the metabolites FA and MON treatment over prolonged periods at higher 74 concentration used and to avoid death of plants at the very early stages of application ^{6, 10, 11}. The 75 76 two concentrations of each treatment were determined based on the highest and the lowest concentrations derived from bakanae diseased plants (data not presented here). The pre-treated 77 seeds were sown in the sterilized soil in trays. Untreated pre-germinated seeds and pre-78 79 germinated seeds inoculated with F. proliferatum (10^6 conidia/mL) used as controls were sown

at the same time as phytohormone and metabolite pre-treated seeds. Seeds were sown in two rows per tray with 15 seeds per row. A total of 8 treatments along with 2 controls were used in this experiment. Each tray represented a single replication and trays were arranged in a completely randomized design with 3 replications per treatment (30 seeds per replication). All plants were maintained in a glasshouse with day and night temperatures of 30–35 °C and 23–30 °C, respectively, and watered daily. No fertilizer was applied to avoid any effects on the phytohormone and metabolites.

87 88

After the single pre-treatment trial, mixtures with two pre-treatment combinations and mixtures with three pre-treatment combinations were employed to observe the symptoms expression in mixtures with different treatment combinations. All procedures used were the same as in the single pre-treatment trial. Development of symptoms associated with the pre-treatments, either singly or in mixtures with two and three pre-treatment combinations was recorded weekly for 3 weeks.

95

96 Sampling procedure and data collection

97 Plants were uprooted carefully without damaging the root tissues at the different 98 sampling times. Sampling times were determined based on disease score levels as follows: level 99 1 (7 days after pre-treatment), level 3 (14 days after pre-treatment) and level 5 (21 days after 99 pre-treatment). The stems, leaves (uppermost second leaf) and roots were separated, and stem 91 and leaf lengths were measured using a measuring scale (1 m).

Roots were washed with water to remove soil and root length was recorded using the Root
Scanner (Winrhizo V700, 2012b). Data expressed as percentage (%) increase or decrease in stem
height and root length over the untreated control at each sampling time was analyzed (10 plants
per replicate).

107

Effects of pre-treatment with GA₃, FA, and MON on stem cell elongation (Histopathological study) 110

Random samples of stem sections from plants pre-treated singly with GA₃ (15 mg/L) FA 111 (400 mg/L), and MON (120 μ g/L), in mixtures with GA₃ (15 mg/L) * FA (400 mg/L)* MON 112 (120 μ g/L), the untreated control and the diseased control were observed after 14 days of pre-113 treatment applied. Plants showing typical symptoms of bakanae disease in relation to disease 114 score level 1 and 3 were collected and studied. Stems were cut into 1 cm sections and prepared 115 for scanning electron microscope analysis. Average cell lengths in the longitudinal stem sections 116 117 of the plants grown from pre-treated seeds and inoculated plants were compared with those in the untreated control plants. Average cell length was measured from 30 randomly selected cells from 118 3 sets of samples (10 cells/sample set) observed under SEM. 119

120

121 Analysis of symptoms expression in plants pre-treated with pure GA_{3} , FA and MON applied 122 singly or in mixtures

Symptoms expression were analysed in plants pre-treated with pure GA₃, FA, and MON
at two concentrations applied singly and in mixtures with two and three treatment combinations
at stipulated disease scoring levels (score 1, 3 and 5) on percent increase or decrease basis in
comparison with diseased control plants. Data were analysed using SAS software (Version 9.2).

129 **Results**

130

132

131 *Effects of GA*₃, *FA and MON applied singly*

Both GA₃ concentrations increased stem height, whereas the other pre-treatments 133 decreased stem height after 7 days of pre-treatment. Highest stem height decrease was observed 134 in pre-treatment FA (400 mg/L) after 7 days of pre-treatment. It was also observed that the 135 136 increasing trend in stem height in GA₃ pre-treatment declined after 7 days to after 21 days. In contrast, pre-treatment with MON (120 µg/L) showed increasing stem height after 14 days of 137 pre-treatment and was found to continue increasing after 21 days which was similar to pre-138 treatment with GA₃ (15 mg/L). Percent decrease in stem height declined in pre-treatment with 139 140 FA (400 mg/L) over time. Stem height increase/decrease following the treatment used were 141 observed in Figure 1.

142

Figure 1. Stem height increase or decrease in plants following pre-seed treatment with GA₃, FA,
and MON singly in comparison with disease free (control) and disease control (inoculated)
plants after 14 days.

146

These results were further confirmed by the histopathological study comparing cell lengths elongation with the different treatments. Stem cell length (average) was elongated (61.33 μ m) in plants pre-treated with higher concentration of GA₃ and was comparable to average stem cell length in diseased control plants (63.47 μ m) after 14 days of pre-treatment and inoculation, respectively (Figure 2 a and b). Average cell length of stems was elongated somewhat (47.6 μ m) when pre-treated with higher concentration of MON compared to average stem cell length of healthy control plants (45.33 μ m) after 14 days (Figure 2 c and d).

154 In contrast, pre-treatment with FA showed stunting in stem height, which may be due to the 155 shorter stem cell length. This percentage decrease in stem height observed was probably due to

the higher cell division that occurred in plants at disease score 1, which caused shorter cell

length (40 μ m) in stems as compared to the stem cell length of healthy control plants (45.33 μ m) observed in the histopathological study (Figure 2 e and d). From these result, it was evident that higher concentrations of GA₃ and FA were responsible for the stem height increase and decrease, respectively, whereas the higher concentration of MON was responsible for marginal stem height increase after 14 days and onwards.

162

163 *Effects of GA*₃, *FA and MON applied in combinations*

164

In mixtures with two treatment combinations, seeds pre-treated with GA₃ (15 mg/L)* MON (120 165 166 μ g/L) resulted in the highest (%) increase in stem height after 7 -14 days and was found to the next to the diseased control plants. All combinations with GA₃ and MON increased stem height 167 after 14 days. Therefore, it was apparent that both GA₃ and MON had a synergistic effect on 168 169 stem height increase. Thus, at the disease score level 3, stem height was increased after 14 days, and this increasing trend was observed in pre-treatment combination treatments with GA₃ (15 170 mg/L)* MON (120 µg/L). In contrast, there might be an antagonistic response between FA and 171 GA₃ and stem height increase or decrease was observed in relation to higher concentration of 172 involvement with GA₃ or FA vice versa Stem height increase in plants of pre-treated seeds with 173 174 GA_3 (10 mg/L)* FA (400 mg/L) and in inoculated plants (diseased control) were observed to be almost similar after 7 days. Thus, stunting observed in plants after 7 days of inoculation might be 175 176 due to the effect of higher concentration of FA present in plants at the disease score level 1 (after 177 7 days of inoculation).

178

The mixtures with three pre-treatment combinations of FA (400 mg/L)* MON (80 μ g/L)* GA₃ (15 mg/L) and FA (100 mg/L)* MON (120 μ g/L)* GA₃ (15 mg/L) increased stem height after 7

days. This is largely due to the effect of GA₃ (15 mg/L) singly as the treatment increased stem
height early at 7 days after pre-treatment. Although stem cell length was observed to be
shortened in this pre-treatment compared to control plants (Figure 2 f and d) but stem height
increase after 14 days in this pre-treatment was attributed to the effect of excessive cell divisions
in this pre-treatment.

Figure 2: SEM micrograph of stem cell length in GA₃ (15 mg/L), FA (400 mg/L) and MON

189 (120 μ g/L) pre-treated plants singly or in combination in comparison with diseased and 190 healthy control plants. [GA₃ pre-treated stem cells (a), Stem cells of diseased control plants 191 (b), MON pre-treated stem cell (c), Stem cells of healthy control plants (d), FA pre-treated 192 stem cell (e) and Stem cells in mixtures of pre-treatment with GA₃ (15 mg/L)* FA (400 193 mg/L)* MON (120 μ g/L) (f)]

194

195 Along with increases in stem height, leaf browning, stem browning, crown rot and root necrosis were observed in plants pre-treated with MON at both concentrations after 21 days. Browning 196 was found to be more prominent at the lower concentration of MON (80 μ g/L) applied singly as 197 pre-treatment, whereas crown rot and root necrosis were more prominent at the higher 198 concentration of MON (120 μ g/L) (Figure 3). The browning symptoms observed in leaves and 199 stems were similar to the symptoms on plants infected with F. proliferatum (diseased control), 200 201 and could easily be distinguished from the symptoms on untreated control (healthy) plants (Figure 3). 202

Figure 3: Plants pre-treated with MON singly and diseased control plants showing leaf browning, stem browning, crown rot and root necrosis in comparison with untreated (healthy) control plants after 21 days.

206

The highest decrease in root length was observed after 7 days, while it was found to increase after 14 days in all pre-treatments along with the diseased control. The highest decrease in root length was observed in pre-treatment with GA_3 (15 mg/L) followed by the diseased control and

GA₃ after 7 days. Both concentrations of MON increased root length after 21 days of pretreatment but with necrotic lesions.

213	In mixtures with two pre-treatment combinations the highest root length increase was observed
214	in mixtures with GA3 (15 mg/L)* MON (120 $\mu\text{g/L})$ after 14 days (Figure 4a). It was also
215	observed that both concentrations of MON caused crown rot and root necrosis in mixtures with
216	GA ₃ in plants pre-treated with two pre-treatment combinations (Figure 5 a). It was also apparent
217	that MON in mixtures either with GA_3 or with FA caused prominent crown rot and root necrosis
218	compared to that in mixtures with FA and GA ₃ (Figure 4 b and c).
219 220 221 222 223 224 225 226 227 228 228	Figure 4: Plant roots showing different architectures when seeds were pre-treated with two treatment mixtures. [mixtures with MON and GA ₃ (a) and mixtures with FA and GA ₃ and mixtures with FA and MON (b)] *G1= GA ₃ (10 mg/L), G2= GA ₃ (15 mg/L), F1= FA (100 mg/L), F2= FA (400 mg/L), M1= MON (80 µg/L), M2= MON (120 µg/L)]. *MON = moniliformtn, GA ₃ = gibberellic acid, FA= fusaric acid Crown rot and root necrosis was also observed to be severe in mixtures with three pre-treatment combinations in GA ₃ (15 mg/L)* MON (120 µg/L)* FA (400 mg/L) (Figure 5).
230	Figure 5. Crown rot and root necrosis due to the effect of MON in mixtures with three pre-
231	treatment combinations. {*G1= GA ₃ (10 mg/L), G2= GA ₃ (15 mg/L), F1= FA (100 mg/L), F2= FA (400
232	mg/L), M1= MON (80 μ g/L), M2= MON (120 μ g/L)}. *MON = moniliformtn, GA ₃ = gibberellic acid,
233	FA = fusaric acid
234 235	From the scenario in pre-treatments, either singly or in mixtures, it was observed that higher
236	concentrations of GA ₃ and MON had a direct influence on increase in stem height. In contrast,

FA singly in any concentration or at higher concentrations in mixtures with GA₃ and MON
decreased stem height.

239

240 **Discussion**

241

Stem height increase in bakanae diseased plants at the disease score level of 3 (14 days 242 after inoculation) was found to be dependent on higher amount of GA₃ singly or in mixtures with 243 MON. Although stem height increase (%) was observed to be lower in pre-treatment GA_3 (15) 244 mg/L)* MON (120 µg/L) compared to diseased control plants after 14 days but it was presumed 245 to be due to slow down of GA_3 activity with time. Moreover, increase in GA_3 concentration was 246 initiated in the diseased control plants after 7 days of pathogen inoculation and reached a 247 maximum concentration in infected plants after 14 days, whereas pre-treatment with GA₃ 248 responded with a maximum level after 7 days. Similar observations were reported by other 249 250 researchers, where GA₃ concentration was increased to its highest level in plants after 1 week of application and subsequently decreased with time [12]. 251

252

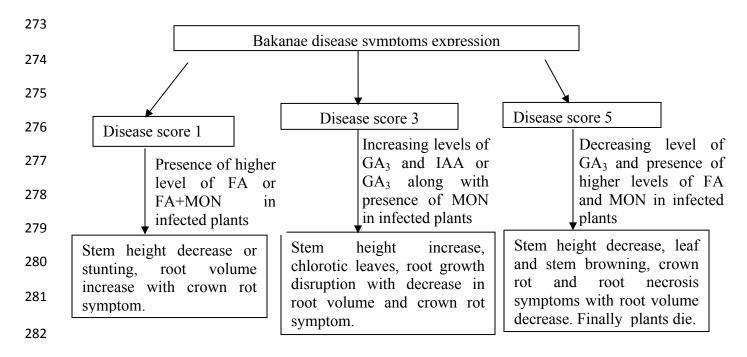
In contrast, pre-treatment with higher concentrations of FA singly or in mixtures with MON decreased stem height after 7 days and was reflected in bakanae diseased plants where stunting was observed at the disease score level of 1 after 7 days of inoculation. At the disease score level of 5, stem heights were found to decrease to some extent probably due to the effect of GA₃. A possible explanation for this was that the increasing rate of GA₃ was slowed down and FA was increased significantly due to antagonistic effect with each other with time in infected plants as observed after 21 days of pre-treatment. This is the first report that complex bakanae symptoms

associated with the combination effect of increased GA_3 levels along with metabolites FA and MON produced by the pathogen in infected plants, rather than solely dependent on increased levels of GA_3 in infected plants. bakanae symptoms expression due to phytohormonal and metabolites effect is as illustrated in the Figure 6.

264

Stem height increase (%) was reliant on elongation of cell length as well as an increase in cell division inside the plant tissues as observed by several researchers. Other researchers reported that cell division was mainly influenced by GA₃ application compared to cell elongation ¹¹. The authors assumed that IAA might have an influence on plant height increase after cell division occurred with GA₃ application. Later, other researchers explained that GA₃ pre-treated plants stimulated IAA synthesis in plant cells first and IAA had an effect on cell elongation and this cell elongation was increased in aged tissues compared to young tissues [14-15].

272



283 Figure 6: Flowchart of Bakanae symptoms expression due to phytohormonal

and metabolites effect (GA_3 = gibberellic acid, IAA= indole acetic acid, FA= fusaric acid, MON= moniliformin).

286 287

284

285

Thus, the increase in stem height in bakanae diseased plants was due to the combined effect of 288 cell division and cell length elongation. The mechanism is similar as observed in infected plants 289 with F. proliferatum which induced cell division first due to increase levels of GA₃ and thereby 290 stimulated IAA production in infected plants. Thus, IAA in infected plants influenced cell 291 292 enlargement in bakanae diseased plants. It was also established that IAA levels increased in bakanae diseased plants infected with F. proliferatum when GA₃ was increased [9]. Several 293 researchers have also supported that GA₃ and IAA play their role synergistically in plants [15-294 295 17]. This higher level of GA₃ along with increased IAA levels at the disease score level of 3, caused stem height increase compared to control plants after 14 days of inoculation. Although 296 297 some stem height increase and cell enlargement was observed in MON pre-treated stem cells, but 298 this response in relation to IAA increase has not been reported yet.

299

In the three pre-treatment combination FA (400 mg/L)* MON (120 µg/L)* GA₃ (15 mg/L) stem 300 height was increased (after 14 days) although shorter stem cell length (36.53 µm) was observed 301 compared to the control (45.33 µm) in the histopathological study. The shorter cell length in this 302 303 three pre-treatments mixture was probably due to an increase in cell division as a result of FA (400 mg/L) and/or GA₃ (15 mg/L) and observed in the histopathological study after 14 days of 304 inoculation. Moreover, it was reported that IAA was increased in plants at the second week after 305 306 GA₃ application ¹⁰, and that the increased IAA was reflected by a higher increase in stem height after 14 days with pre-treatment FA (400 mg/L)* MON (120 µg/L)* GA₃ (15 mg/L). Again the 307 shorter stem cell length (36.53 µm) was the average of 30 randomly selected stem cells pre-308

treated with FA (400 mg/L)* MON (120 μ g/L)* GA₃ (15 mg/L). Although some stem cells were initiated to elongate in this pre-treatment as observed in Figure 5.2e whereas, others were in initial stage of elongation and did not reflect individual cell enlargement in histopathological study where measurement was on average cell length. Additionally, stem height increase was slowed down after 7 days due to antagonistic effect of higher concentration of FA with GA₃ and after 14 days stem height increased due to the effect of higher concentration of MON and GA₃ combination.

316

317 In contrast, plant height stunting was found to be mainly due to the effect of FA rather than GA₃. This observation is supported by other researchers as well. Other researcher reported that plant 318 stunting or rossetting occurred due to low concentration of GA₃ in plants ¹⁶. It was also observed 319 320 that high levels of FA accumulation in plants resulted in stunted plant height and decreased root length in tomato [7, 19-20]. The lower amount of GA₃ (8.9 μ g/g fresh wt.) as determined in the 321 disease score level of 1 resulted in stunting of plants after 7 days of pre-treatment. Although 322 metabolic activity was not determined in this experiment but other researchers repoted that the 323 stunting mechanism is mainly due to changes in metabolic activity in plants, including speeding 324 up of the lipid peroxide activity, inhibition of ATP synthesis or decreased ATP levels [6, 19-20]. 325 compared to control plants. Moreover, higher concentrations of FA and MON also resulted in 326 327 plant height reduction as was observed in jimsonweed plants when pure FA and MON were 328 applied ⁷. Thus, lower amount of GA₃ and/or in mixtures with higher amounts of FA and MON contributed to the cessation and collapse of plants growth at the disease score level of 5 that was 329 330 observed after 21 days of pre-treatment.

331

Crown rot and root necrosis due to higher concentration of MON in infected plants at later 332 growth stages were observed. Similar symptoms have also been observed by other researchers in 333 different plant species [7, 21]. Brown to pink discoloration of leaves at a disease score level of 5 334 may be due to higher concentration of MON present in plants infected by Fusarium 335 proliferatum. It was also observed pink ear rot in maize when a higher concentration of MON 336 337 was associated with the ear rot causal pathogen *Fusarium suubgltinans* [22]. In addition, earlier researcher reported that MON had toxic effects with growth reductions of coleoptiles as well as 338 vein chlorosis and necrosis in corn and tobacco callus ¹⁹. MON was also found to cause 339 340 cytoplasmic disruptions and abnormal mitosis that resulted in "a disruption of the spindle apparatus" in infected plant cells ^{19, 21}. In contrast, reported that browning of stalk, leaf, and ear 341 of rice was due to the effect of fumonisin B1 when inoculated with F. proliferatum²². However, 342 the authors did not isolate other mycotoxins produced by the fungus in the rice plants infected 343 with F. proliferatum nor compared the symptoms treated with pure fumonisin B1. More recently 344 provided evidence that FB1 had no pathogenicity effects on bakanae symptoms development¹. 345 The browning or pinkish symptom associated with a disease score 5 was evaluated by applying 346 pure MON to germinating seeds of susceptible variety MR 211, and similar symptoms were 347 observed as in plants inoculated with F. proliferatum. Therefore, it was confirmed that browning 348 and pinkish discoloration of leaves and stems as well as crown rot and root necrosis were 349 associated with MON effect. Higher concentration of FA, MON and less GA₃ accumulation in 350 351 infected plants resulted in dead plants over time at a disease score level of 5, and this was attributed to inhibition of photosynthesis and toxic effects of MON and FA in plant cells. 352

353

355 References

381

- Cruz A, Marin P, González-Jaén M T, Aguilar KG, Cumagun CJR. Phylogenetic analysis, fumonisin production and pathogenicity of *Fusarium fujikuroi* strains isolated from rice in the Philippines. Jl of Sci Food and Agril. 2013; 93: 3032-3039.
- Zainudin NAIM, Razak AA, Salleh B. Secondary metabolite profiles and mating
 populations of *Fusarium* species in section *Liseola* associated with bakanae disease of
 rice. Malay J of Microbiol. 2008; 4(1): 6-13.
- 362 3. Glenn AE. Mycotoxigenic *Fusarium* species in animal feed. Animal Feed Sci and
 363 Technol. 2007; 137: 213–240.
- Desjardins AE, Manandhar HK, Plattner RD, Anandhar GG, Poling SM, Maragos CM.
 Fusarium species from Nepalese rice and production of mycotoxins and gibberellic acid
 by selected species. Appl and Environ Microbiol. 2000; 66(3): 1020–1025.
- 367 5. Bari R, Jones LDG. Role of plant hormones in plant defence responses. Plant Mole Biol.
 368 2009; 69:473–488.
- Bouizgarne B, El-Maarouf-Bouteau H, Frankart C, Reboutier D, Madiona K, Pennarun AM, et al. Early physiological responses of *Arabidopsis thaliana* cells to fusaric acid: toxic and signalling effects. New Phytologist. 2006; 169: 209–218.
- Abbas HK, Boyette CD, Hoagland RE. "Phytotoxicity of *Fusarium*, other fungal isolates, and of the phytotoxins fumonisin, fusaric acid, and moniliformin to jimsonweed".
 Phytoprotection. 1995; 76 (1): 17-25.
- 8. Quazi SAJ, Meon S, Jaafar H, Ahmad ZABM. Characterization of Fusarium proliferatum through species specific primers and its virulence on rice seeds. Int J Agri Biol. 2013;15:649-56.
- Quazi SAJ, Meon S, Jaafar H, Ahmad ZABM. The role of phytohormones in relation to bakanae disease development and symptoms expression. Physiol and Mole Plant Pathol. 2015; 90: 27-38
 - 10. Pavlovkin J, Mistrík I, Prokop M. Some aspects of the phytotoxic action of fusaric acid on primary *Ricinus* roots. Plant, Soil and Environ. 2004; 50 (9): 397–401.
- 383 11. Styer, C.H. and Cutler, H.G. (1984). Effects of moniliformin on mitosis in maize (*Zea mays* L.). Plant and Cell Physiol. 1996; 25(6): 1077-1082.
- 12. Kong L, Abrams SR,Owen SJ,Graham H, Aderkas PV. Phytohormones and their
 metabolites during long shoot development in Douglas-fir following cone induction by
 gibberellin injection. Tree Physiol. 2008; 28: 1357–1364.
- Arney SE, Mancinelli P. The basic action of gibberellic acid in elongation of 'meteor'
 pea stems. New Phytologist. 1965; 65(2): 161-175.
- I4. Gray WM, "Ostin A, Ran Sandberg G0, Romano CP, Estelle M. High temperature promotes auxin-mediated hypocotyl elongation in *Arabidopsis*. Proceedings of the National Academy of Sciences of the United States of the America. 1998; 95(12): 7197–7202. Available at : https://doi.org/10.1073/pnas.95.12.7197
- 15. Kazama H, Katsumi M. Auxin-gibberellin relationships in their effects on hypocotyl
 elongation of light-grown cucumber seedlings II. Effect of GA₃-pretreatment on IAA induced elongation. Plant and Physiol.1974; 15: 307-314.
- Bhattacharya S, Bhattacharya NC, Malik CP. Synergistic effect of gibberellic acid and
 indole-3-acetic acid on rooting in stem cuttings of *Abelmoschus esculentus* Moench.
 Planta. 1978; 138: 111-112.

- 400 17. Ockerse R, Galston AW. Gibberellin-Auxin Interaction in pea stem elongation. Plant
 401 Physiol. 1967; 42: 47-54.
- 402 18. Tanimoto E. Tall or short? Slender or thick? A plant strategy for regulating elongation
 403 growth of roots by low concentrations of gibberellin. Annals of Bot. 2012; 10(2): 373-81.
 404 doi: 10.1093/aob/mcs049. Epub 2012 Mar 21.
- 405 19. Diniz SPSS, Oliveira RC. Effects of fusaric acid on *Zea mays* L. seedlings. Int J of Exp
 406 Bot. 2009; 78: 155-160.
- 407 20. Asch VMAJ, Rijkenberg FHJ, Coutinho TA. Phytotoxicity of fumonisin B1, moniliformin and T-2 toxin to corn callus culture. Posthar Pathol and Mycotox. 1992;
 409 82(11): 1330-1332.
- 21. Lew H, Chelkowski J, Pronczuk P, Edinger W. Occurrence of the mycotoxin
 moniliformin in maize (*Zea mays* L.) ears infected by *Fusarium subglutinans* (Wollenw.
 & Reinking) Nelson *et al.*, Food Addit and Contamin. 13(3): 321-324.



Figure 1. Stem height increase or decrease in plants following pre-seed treatment with GA₃, FA, and MON singly in comparison with disease free (control) and disease control (inoculated) plants after 14 days.

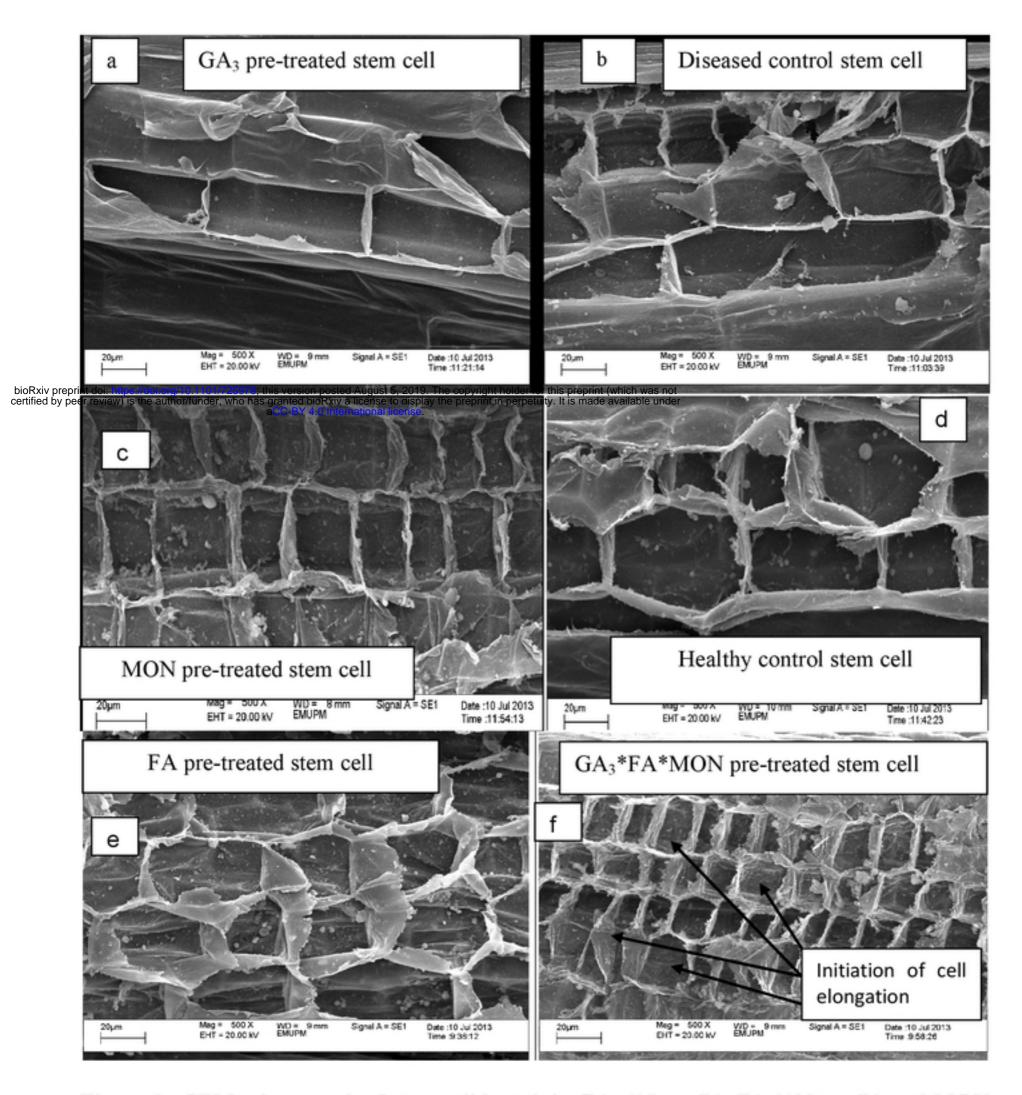
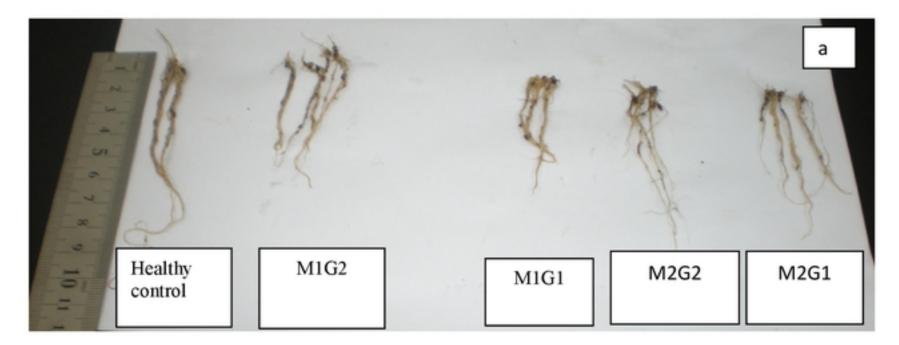
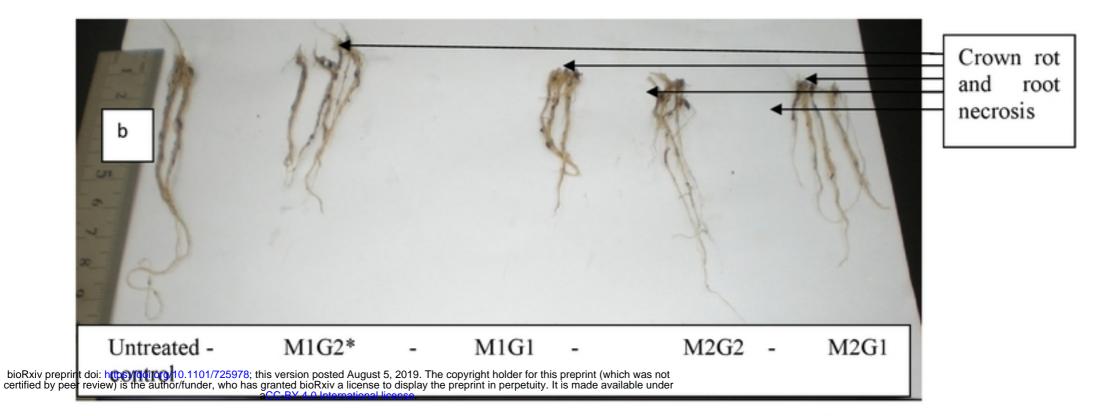


Figure 2: SEM micrograph of stem cell length in GA₃ (15 mg/L), FA (400 mg/L) and MON (120 μ g/L) pre-treated plants singly or in combination in comparison with diseased and healthy control plants. [GA₃ pre-treated stem cells (a), Stem cells of diseased control plants (b), MON pre-treated stem cell (c), Stem cells of healthy control plants (d), FA pre-treated stem cell (e) and Stem cells in mixtures of pre-treatment with GA₃ (15 mg/L)* FA (400 mg/L)* MON (120 μ g/L) (f)]



Figure 3: Plants pre-treated with MON singly and diseased control plants showing leaf browning, stem browning, crown rot and root necrosis in comparison with untreated (healthy) control plants after 21 days.





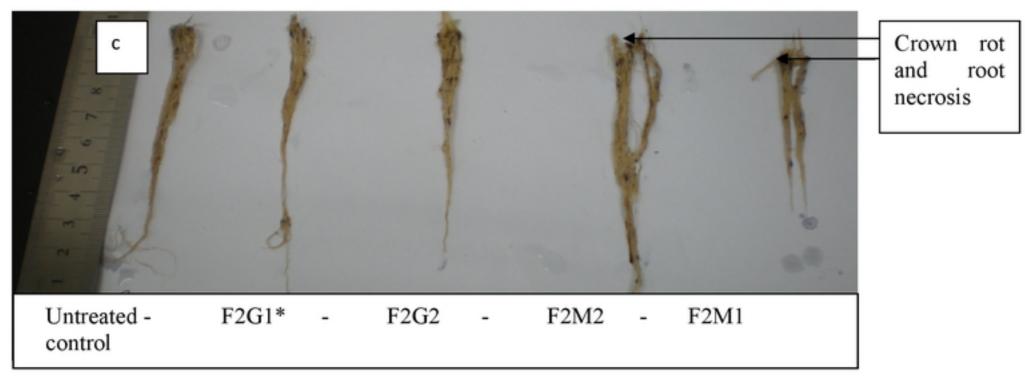


Figure 4: Plant roots showing different architectures when seeds were pre-treated with two treatment mixtures. [mixtures with MON and GA₃ (a) and mixtures with FA and GA₃ and mixtures with FA and MON (b)] *G1= GA₃ (10 mg/L), G2= GA₃ (15 mg/L), F1= FA (100 mg/L), F2= FA (400 mg/L), M1= MON (80 μ g/L), M2= MON (120 μ g/L)]. *MON = moniliformtn, GA₃ = gibberellic acid, FA= fusaric acid combinations in GA₃ (15 mg/L)* MON (120 μ g/L)* FA (400 mg/L) (Figure 5).

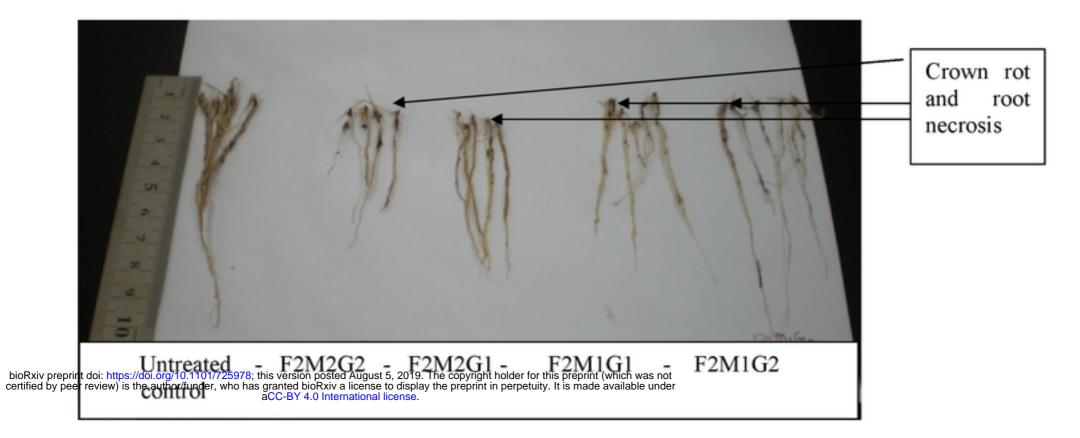


Figure 5. Crown rot and root necrosis due to the effect of MON in mixtures with three pretreatment combinations. {*G1= GA₃ (10 mg/L), G2= GA₃ (15 mg/L), F1= FA (100 mg/L), F2= FA (400 mg/L), M1= MON (80 μ g/L), M2= MON (120 μ g/L)}. *MON = moniliformtn, GA₃ = gibberellic acid, FA = fusaric acid