Ischemic Stroke Reduces Bone Perfusion and Alters Osteovascular Structure

Nicholas J. Hanne, PhD[a], Andrew J. Steward, PhD[a], Carla Geeroms[b], Elizabeth D. Easter[c], Hannah L. Thornburg[a], Greet Kerckhofs, PhD[b,d,e], Tatjana Parac-Vogt, PhD[f], Huaxin Sheng, MD[g], Jacqueline H. Cole, PhD*,[a]

[a] Joint Department of Biomedical Engineering, University of North Carolina, Chapel Hill, NC, and North Carolina State University, Raleigh, NC, USA

[b] Prometheus, Division of Skeletal Tissue Engineering, KU Leuven, Leuven, Belgium

[c] Materials Science and Engineering, North Carolina State University, Raleigh, NC, USA

[d] Institute of Mechanics, Materials and Civil Engineering, UC Louvain, Louvain-la-Neuve, Belgium

[e] Materials Engineering, KU Leuven, Leuven, Belgium

[f] Department of Chemistry, KU Leuven, Leuven, Belgium

[g] Department of Anesthesiology, Duke University Medical Center, Durham NC, USA

Short title: Stroke Alters Osteovascular Structure and Function

*Corresponding Author:
Jacqueline H. Cole
Joint Department of Biomedical Engineering
University of North Carolina and North Carolina State University
911 Oval Drive
Campus Box 7115
Raleigh, NC 27695-7115
Tel: 919-515-5955
Fax: 919-513-3814
jacquecole@ncsu.edu

Abstract

Rationale: Stroke patients lose bone mass and experience fracture at an elevated rate. Although functional intraosseous vasculature is necessary for skeletal maintenance, the effect of stroke on osteovasculature is unknown. Objective: To characterize changes to osteovascular function, structure, and composition following mild-to-moderate-severity ischemic stroke in mice, both with and without exercise therapy. Methods and Results: Twelve-week-old male mice (n=27) received either a stroke (middle cerebral artery occlusion) or sham procedure, followed by four weeks of daily treadmill or sedentary activity. Intraosseous perfusion, measured weekly in the proximal tibial metaphysis, was reduced by stroke for two weeks. In the second week of recovery, exercise nearly restored perfusion to sham levels, and perfusion tended to be lower in the stroke-affected limb. At the conclusion of the study, osteovascular structure was assessed with contrast-enhanced computed tomography in the distal femoral metaphysis. Stroke significantly increased osteovascular volume and branching but reduced the relative number of blood vessels close to bone surfaces (6-22 μm away) and increased the relative number more than 52 μm away. These differences in vessel proximity to bone were driven by changes in the stroke-exercise group, indicating compounded effects of stroke and exercise. Exercise, but not stroke, nearly reduced the amount of osteogenic Type H blood vessels in the proximal tibial metaphysis, quantified with immunofluorescence microscopy. Conclusions: This study is the first to examine the effects of stroke on osteovasculature. Stroke increased the amount of osteovasculature, but since blood vessels close to bone are associated with bone remodeling, the shift in osteovascular structure could play a role in bone loss following stroke. The exercise-induced reduction in the amount of Type H vessels and the stroke-exercise effect on osteovascular structure suggest moderate aerobic activity may have detrimental effects on bone remodeling during early stroke recovery.

Keywords

ischemic stroke, bone vasculature, intraosseous perfusion

Non-standard Abbreviations and Acronyms

Ad.D – Adipocyte density

Ad.Th – Adipocyte thickness

Ad.V/Ma.V – Adipocyte volume per marrow volume

EMCN - Endomucin

LDF – Laser Doppler flowmetry

Ves.D – Blood vessel density

Ves.Th – Blood vessel thickness

Ves.S-BS – Vessel surface to bone surface distance

Ves.V/Ma.V – Vessel volume per marrow volume

Introduction

Stroke is the leading cause of disability in the US, as well as one of the leading causes worldwide [1]. Stroke not only causes cognitive and motor impairments but also negatively affects skeletal health [2–8]. Stroke patients lose bone mass at an accelerated rate, fall more, and experience hip fracture 2-4 times more frequently than individuals of the same age who have not experienced stroke [2,3,5]. Because bone adapts to altered mechanical loading [9], and bone mineral density (BMD) loss and fracture preferentially affect the paretic limbs [6,7], hemiplegia and limb disuse are thought to be solely responsible for these skeletal deficits post-stroke [4,6,8]. In addition, rehabilitation activity and exercise are correlated with reduced fracture risk and improved skeletal health outcomes following stroke [3,10–13], further supporting loading as a primary driver. However, a study in severe stroke patients showed lower BMD in various sites on the stroke-affected side, relative to the unaffected side, despite complete bedrest [8]. Even after controlling for activity level and other factors, lower vascular elasticity, a metric of vascular health, has been associated with lower polar stress-strain index, a surrogate measure of bone torsional stiffness and strength [14], in the radial diaphysis [15]. Together these results suggest that the negative effects of stroke on bone extend beyond just mechanical unloading and that vascular deficits may also contribute.

Because vasculature is critically important for maintaining functional bone tissue, supplying not only oxygen and nutrients but also cell signaling factors [16], vascular dysfunction may contribute to the increased fracture risk experienced by stroke patients. Little is known about the effects of stroke on limb vasculature, in particular on vascular perfusion and structure, and no previous study has examined the effects on the vasculature within bone, or *osteovasculature*. Two previous studies reported decreased blood flow in the affected lower leg compared to the

unaffected leg in elderly chronic stroke patients with mild-to-moderate gait asymmetry at least 6

months after moderate ischemic stroke [17,18]. Changes to limb blood flow, however, may not be

indicative of changes to osteovasculature, which has more direct impacts on bone remodeling and

the marrow microenvironment. In osteoporotic individuals, increased intraosseous perfusion

measured by contrast-enhanced clinical imaging was strongly correlated with increased bone

formation rate in the iliac crest [19] and weakly correlated with higher BMD in the proximal femur

[20]. In addition, an increased number of capillaries have been observed within 50 μm of bone

surfaces at active remodeling sites, compared to non-remodeling bone surfaces, in both humans

[21] and rats [22]. Within long bones, endothelial cells that express both endomucin (EMCN) and

CD31 (*Type H cells*) have been shown to regulate osteogenesis in mice [23,24]. The effects of

stroke on these osteovascular metrics (perfusion, structure, and cellular composition) are unknown

and may provide insight into the relationship between bone vasculature and bone loss following

stroke. Although characterizing osteovasculature in humans is difficult, it can be examined in

animal models, such as the established rodent models of ischemic stroke involving middle cerebral

artery occlusion (MCAo) [25,26].

In our previous study using an MCAo mouse model, we showed that although stroke did

not cause detrimental changes to bone microstructure, it prevented the exercise-induced

microstructural gains observed in the sham group, even without changes to limb coordination

during gait [27]. Since bone and vascular health are tightly coupled, we hypothesized that stroke

negatively impacts osteovasculature and inhibits the positive effects of treadmill exercise. In this

study, we used the same bones to characterize changes to *in vivo* functional intraosseous perfusion,

blood vessel network microstructure, and endothelial cellular composition following ischemic

stroke.

Methods

Study Design

All procedures were approved by North Carolina State University's Animal Care and Use Committee. Animals were group housed 4-5 per cage with a 12-hour diurnal light cycle and free access to standard chow and water. Twenty-seven male C57Bl/6J mice (The Jackson Laboratory, Bar Harbor, ME) received either an ischemic stroke (n = 15) or sham surgery (n = 12) at twelve weeks of age (Fig. 1). Mice were housed individually with wetted food and hydrogel packs for four days following surgery during the acute recovery period. After this period, they were returned to their original group cages and split into either daily treadmill exercise groups (n = 6 sham-exercise, n = 8 stroke-exercise) or sedentary control groups (n = 6 sham-sedentary, n = 7 stroke-sedentary). Surgery and exercise groups were randomly assigned to cages. Body mass was measured twice a day for the four days following surgery and weekly thereafter. Stroke recovery and sensorimotor function were assessed daily for four days following stroke and weekly thereafter. Intraosseous perfusion was measured with laser Doppler flowmetry (LDF) during the stroke or sham surgery and then weekly thereafter. After four weeks of stroke recovery with treadmill or sedentary activity, the mice were euthanized with CO_2 asphyxiation followed by cervical dislocation. Femora and tibiae from both left (stroke-affected) and right (stroke-unaffected) hindlimbs were collected. A subset of both affected and unaffected femora from six mice (n = 1 sham-sedentary, n = 2 sham-exercise, n = 1 stroke-sedentary, n = 2 stroke-exercise) were fixed for 16 hours in 5% neutral buffered formalin at 4°C and then stored in 1X phosphate buffered saline (PBS) at 4°C until assessment of osteovascular structure. All other bones were fixed for 18 hours in 10% neutral buffered formalin at 4°C and then stored in 70% ethanol at 4°C for assessment of osteovascular composition.

Stroke Procedure

Ischemic stroke was induced with the middle cerebral artery occlusion (MCAo) procedure using aseptic technique [25,26,28]. Anesthesia was induced with 5% isoflurane in a 70:30 N_2:O_2 gas mixture, then maintained with about 2% isoflurane. Stroke was induced by occluding the middle cerebral artery with a thin 6-0 nylon monofilament occluder with a silicon-coated tip (Doccol Corporation, Redlands, CA) for 30 minutes. MCAo was monitored using LDF (moor VMS-LDF, Moor Instruments Ltd, Axminster, UK) with a monofilament probe (VP10M200ST, Moor Instruments), ensuring that cerebral blood flow was reduced by 80% relative to baseline and maintained throughout the procedure. The sham procedure was the same as the stroke procedure but without insertion of the occluder. For tibial perfusion, a small, 2-5 mm-long incision was made over the proximal anteromedial side of the left tibia near the metaphysis, avoiding underlying soft tissue and muscle. A small region of the periosteum was scraped away (about 0.5 mm^2), and a needle probe (VP4 Needle Probe, Moor Instruments) was held firmly against the bone with a micromanipulator (MM3-ALL, World Precision Instruments, Sarasota, FL). The severity of stroke impairments was examined weekly following surgery by assessing sensorimotor function with neuroscores [27].

Treadmill Exercise

Exercise groups were acclimated to the treadmill (Exer 3/6, Columbus Instruments, Columbus, OH) for two days prior to and two days following (on recovery days 4-5) the stroke or sham procedure. After acclimation, exercise groups performed a standard moderate treadmill exercise regimen (12 m/min, 30 min, 5 days/wk, 5° incline), while sedentary groups were placed on a stationary replica treadmill for a matched time period.

Bone Perfusion (Tibia)

During the four-week recovery period, intraosseous perfusion was measured weekly in

affected (left) and unaffected (right) proximal tibiae using laser Doppler flowmetry (Fig. 2A) [29].

LDF provides a functional measure of blood flow in long bones that is influenced by the amount

of blood vessels, blood flow velocity, blood vessel permeability, and blood vessel size [20]. We

previously showed that our modified, less invasive LDF procedure can be performed serially

without inducing inflammation or gait abnormalities [30]. Using the same method as during the

MCAo procedure, an incision was made over one tibia, the periosteum was gently removed, and

the LDF needle probe was placed firmly against bone with the micromanipulator. A 30-sec

measurement was recorded, the probe was removed and repositioned, and a second 30-sec

measurement was recorded. The procedure was repeated for the contralateral tibia. Each weekly

perfusion measurement was composed of the weighted mean of the two 30-sec long measurements

for that bone.

Osteovascular Structure (Femur)

Osteovascular structure was examined in the subset of affected and unaffected femora

stored in PBS using contrast-enhanced micro-computed tomography (CE-CT). Each femur was

cut in half transversely with a scalpel, and the distal halves were incubated in a 3.695 mmol/L

staining solution of hafnium-substituted Wells-Dawson polyoxometalate hydrate in 1 mL PBS for

48 hours at 4°C according to a previously described protocol [31]. Samples were scanned with a

phoenix nanotom® m (GE Measurement and Control Solutions, Wunstorf, Germany) using 60 kV

peak X-ray tube potential, 140 µA X-ray intensity, and 500-ms integration time, and scans were

reconstructed at an isotropic voxel size of 2 µm.

Blood vessel and adipocyte microstructure was assessed in a manually selected volume of

interest (VOI) that was 1.6-mm long (about 10% average femur length), starting from the distal

growth plate and extending proximally into the metaphysis, excluding cortical bone tissue, using

the CTAn software (Bruker MicroCT, Kontich, Belgium). Within this VOI, cancellous bone was

segmented, binarized via automatic 3D Otsu segmentation, and subtracted from the image, leaving

the marrow volume. A manual global thresholding method was used to segment blood vessels and

adipocytes. The vessel volume fraction (vessel volume/marrow volume, Ves.V/Ma.V, %), vessel

density (Ves.D, $\#/mm^3$), distribution of vessel thickness (Ves.Th, μm), adipocyte volume fraction

(adipocyte volume/marrow volume, Ad.V/Ma.V, %), adipocyte density (Ad.D, $\#/mm^3$), and

distribution of adipocyte thickness (Ad.Th, μm) were quantified using CTAn. Branching

parameters of the osteovascular network, including total number of branches, number of junctions,

number of triple points (e.g., junctions with three branches), and number of quadruple points (e.g.,

junctions with four branches), were quantified in BoneJ (FIJI v. 1.51n) [32,33]. The distribution

of distances between blood vessel surfaces and bone surfaces (Ves.S-BS distance) was calculated

in MATLAB® (R2018, The MathWorks, Natick, MA).

Osteovascular Composition (Tibia)

Osteovascular composition was examined using immunofluorescence confocal

microscopy in bone tissue sections from a subset of affected tibiae (n = 3 sham-sedentary, n = 2

sham-exercise, n = 5 stroke-sedentary, n = 4 stroke-exercise) labeled for markers of non-arterial

endothelial cells (EMCN) and endothelial cells in sinuses, arterioles, venules, and capillaries

(CD31) [23,34]. Blood vessels that are positive for both EMCN and CD31 (defined as *Type H*)

have been shown to regulate bone-vascular crosstalk and couple osteo- and angiogenesis in long

bones [24]. Samples were prepared and labeled using a previously described procedure [35],

producing 50-µm-thick longitudinal sections of the proximal tibial metaphysis labeled for EMCN

and CD31. Nuclear staining was performed with DAPI.

Immediately following staining, sections were imaged at 20X on a Zeiss Laser Scanning

Microscope 880 with Airyscan (Carl Zeiss Microscopy, Thornwood, NY), and the areas of CD31-

positive, EMCN-positive, and Type H vessels, relative to total area, were quantified in a

metaphyseal region of interest (ROI) equal to 10% of the tibia length. CD31-positive regions inside

the ROI were manually traced into masks in FIJI, while EMCN-positive regions were

automatically binarized with custom code in MATLAB®. The CD31 mask, EMCN mask, and the

intersection of both masks (Type H) were calculated in FIJI. Osteovascular composition

parameters were measured in two sections per bone, and the mean values were used for analysis.

Statistical Analyses

Statistical models were analyzed with SAS (SAS University Edition v. 9.4, SAS Institute

Inc., Cary, NC) with a significance level of 0.05. Statistical models were designed to determine

the following: 1) effect of ischemic stroke and exercise on intraosseous perfusion during stroke

recovery; 2) acute effect of ischemic stroke on intraosseous perfusion during surgery; 3) effect of

stroke and exercise on osteovascular structure and composition; and 4) whether stroke

differentially affects intraosseous perfusion or osteovascular structure in the affected vs.

unaffected limb.

For analysis #1, LDF measures of perfusion were compared between surgery and activity

groups at each timepoint and limb using a mixed hierarchical linear model (procedure GLIMMIX)

with interaction between all terms [36]. LDF was measured across four timepoints (Weeks 1-4) on

each limb of each mouse. Effect differences between surgery and activity groups were compared

within each timepoint (i.e., stroke-exercise vs. stroke-sedentary at Week 2) using least squares

means and Tukey-Kramer adjustments for multiple comparisons.

For analysis #2, the stability of the tibial perfusion measurements throughout the surgery

was examined using the slope of the 30-minute LDF measurement taken during the surgical

procedure. To determine if the slope for each group was different from zero, which would indicate

that the perfusion measurement varied and was not stable, an unpaired t-test was performed for the

sham group, and a Wilcoxon rank sum test was performed for the stroke group. Slopes were also

compared between the stroke and sham groups with a Wilcoxon two-sample test.

For analysis #3, distribution data from CE-CT (vessel thickness, blood vessel surface to

bone surface distance) were compared between surgery and activity groups using a mixed

hierarchical linear model (procedure GLIMMIX) with interaction between terms. Effect

differences were compared between groups within histogram bins (i.e., stroke-exercise vs. stroke-

sedentary within bin 'a') using least squares means with Tukey-Kramer adjustments for multiple

comparisons. All other CE-CT parameters (Ad.V/Ma.V, average Ad.Th, Ad.D, Ves.V/Ma.V,

average Ves.Th, Ves.D, branches, junctions, triple points, and quadruple points) were compared

between surgery and activity groups using a repeated measures factorial model (procedure

MIXED) but without the 'bin' repeated factor. Immunofluorescence parameters were compared

between surgery and activity groups using a standard two-factor general linear model (procedure

GLM) with interaction and Tukey adjustments for multiple comparisons.

For analysis #4, the same repeated measures models from analyses 1 and 3 were used, but

least squares means were compared between limbs within surgery group (i.e., affected vs.

unaffected side within the stroke group).

221   Immunofluorescence and LDF data during MCAo are presented as mean ± standard

222 deviation. Data analyzed using least squares means (repeated LDF measures, CE-CT data) are

223 presented as least squares mean ± 95% confidence interval. Repeated LDF and CE-CT data are

224 presented as the least squares mean for both limbs per mouse unless otherwise noted.

226 **Results**

227 Bone Perfusion

228   Tibial perfusion was reduced in both limbs for two weeks following stroke, with 23% lower

229 perfusion in the stroke groups relative to the sham groups at Week 1 ($p = 0.0064$) and 24% lower

230 perfusion at Week 2 ($p = 0.0061$), but perfusion levels were similar between surgery groups at

231 Week 3 ($p = 0.71$) and Week 4 ($p = 0.60$) (Fig. 2B). Perfusion was nearly greater in exercise

232 groups, relative to sedentary groups, at Week 2 (21%, main effect $p = 0.050$), primarily driven by

233 the nearly increased perfusion in stroke-exercise (30% relative to stroke-sedentary, $p = 0.054$), not

234 sham-exercise (relative to sham-sedentary, $p = 0.32$). Exercise did not affect perfusion in other

235 weeks ($p = 0.94$ Week 1, $p = 0.79$ Week 3, $p = 0.22$ Week 4). Compared to the unaffected tibia,

236 perfusion in the affected tibia was nearly decreased for the stroke group at Week 2 (21%, $p = $

237 0.055) but not in Week 1 ($p = 0.42$), Week 3 (16%, $p = 0.11$), or Week 4 ($p = 0.58$) (Fig. 2C).

238 Perfusion was similar between affected and unaffected limbs for the sham group at all timepoints

239 ($p = 0.61$-0.90).

240   During the surgeries (occluded for stroke, not occluded for sham), tibial perfusion

241 measurements remained stable throughout, with slopes that were not significantly different from

242 zero for either stroke ($p = 0.39$) or sham ($p = 0.14$). The slopes were similar between stroke (-

243 $0.0063 ± 0.0415$ PU/min) and sham ($0.0049 ± 0.0106$ PU/min) groups ($p = 0.98$). These results

demonstrate that cerebral ischemia does not directly impact blood supply to the tibia during the

occlusion, suggesting that any changes to osteovasculature during the recovery period result from

more systemic effects.

Osteovascular Structure

Blood vessel volume fraction in the distal femur was increased by 38% in the stroke groups

relative to sham (p = 0.0061) and decreased by 14% in the exercise groups relative to sedentary (p

= 0.031, Fig. 3A-B). Exercise mitigated the effect of stroke, with 17% lower Ves.V/Ma.V in

stroke-exercise than in stroke-sedentary (p = 0.027), but did not affect blood vessel volume fraction

in sham-exercise compared to sham-sedentary (p = 0.20, Fig. 3B). Neither stroke (p = 0.74) nor

exercise (p = 0.24) affected blood vessel density (stroke-exercise: 132.9 ± 215.8 mm^{-3}, stroke-

sedentary: 62.2 ± 305.2 mm^{-3}, sham-exercise: 188.2 ± 215.8 mm^{-3}, sham-sedentary: 54.1 ± 305.2

mm^{-3}). The vessel network was more branched following stroke, with 26% greater number of

branches (p = 0.0089, Fig. 4A), 27% more junctions (p = 0.015, Fig. 4B), 33% more triple points

(p = 0.040, Fig. 4C) and 43% more quadruple points (p = 0.017, Fig. 4D), for the stroke group

compared to sham. Exercise did not affect any of these network branching parameters.

Neither stroke (p = 0.31) nor exercise (p = 0.84) affected mean blood vessel thickness (Fig.

3C). However, the distribution of Ves.Th was shifted following stroke, with fewer small vessels

(6-22 μm thickness, p = 0.029 for 6-10 μm, p = 0.033 for 10-14 μm, p = 0.019 for 14-18 μm, p =

0.056 for 18-22 μm) and more larger vessels (greater than 66 μm thickness, p = 0.010) (Fig. 5A).

Exercise mitigated the stroke-induced increase in larger blood vessels (p = 0.0032 stroke-exercise

vs. stroke-sedentary), bringing vessel thickness for stroke-exercise to similar values as sham-

exercise (p = 0.72) (Fig. 5A). The proximity of blood vessels to bone surfaces was altered by both

stroke and exercise, as seen by the shifted distribution of Ves.S-BS distance (Fig. 5B). Compared

to sham, the stroke group had 38% fewer blood vessels positioned within 4 μm of bone surfaces

(p = 0.041), 19% fewer blood vessels positioned 4-8 μm from bone surfaces (p = 0.0010), and

16% more vessels positioned 52 μm or farther away from bone surfaces (p = 0.036). Exercise had

a similar effect as stroke, with 33% fewer blood vessels within 4 μm of bone surfaces (p = 0.017)

and 16% more vessels 52 μm or farther away from bone surfaces (p = 0.036) compared to

sedentary. The effect of stroke and exercise were driven primarily by changes in the stroke-

exercise group, which had significantly fewer vessels positioned within 4 μm and significantly

more vessels positioned farther than 52 μm from bone surfaces compared to all other groups (Fig.

5B).

Adipocyte microstructure was not significantly affected by stroke or exercise, with similar

values for adipocyte volume fraction, density, and thickness across groups (Table I). However,

adipocyte density was nearly increased for stroke-exercise relative to sham-exercise (109%, p =

0.086). Affected and unaffected side measurements were not significantly different in the stroke

or sham groups for any of the CE-CT parameters (vessel microstructure, vessel branching, or

adipocyte microstructure).

Osteovascular Composition

Neither stroke nor exercise affected the relative area of endomucin-expressing (p = 0.43

and p = 0.54, Fig. 6B) or CD31-expressing (p = 0.18 and p = 0.11, Fig. 6C) blood vessels in the

proximal tibial metaphysis of affected limbs. The relative area of Type H vessels expressing both

EMCN and CD31 (Fig. 6A) was nearly lower for exercise compared to sedentary (186%, p =

0.059, Fig. 6D) but was not significantly different between stroke and sham (p = 0.20).

Discussion

This study is the first to report the effects of ischemic stroke on osteovasculature. Middle cerebral artery occlusion in mice was associated with increased blood vessel volume within the distal femoral metaphysis, primarily due to increased vascular branching and a greater number of larger vessels. These changes followed reduced perfusion in the proximal tibia for two weeks following stroke, with relatively lower blood supply to the affected than unaffected bone in stroke groups but not sham groups. Furthermore, the effects of stroke and exercise on the proximity of vessels to bone surfaces were driven primarily by changes in the stroke-exercise group, yielding fewer blood vessels near to bone surfaces. Because previous studies have shown that active sites of remodeling are associated with increased concentration of capillaries within 50 µm of the remodeling surface compared to non-remodeling bone surfaces [21], these changes to osteovascular structure and function may play a role in bone loss in stroke patients.

Limb disuse via hindlimb suspension has also been shown to decrease bone perfusion in rodents [37,38]. However, in our study, mice remained ambulatory following the mild to moderate severity stroke with no alterations in limb coordination patterns [27], yet bone perfusion was still decreased post-stroke, even in the stroke-exercise group. Together, these results suggest that the osteovascular changes in this study primarily result from more systemic stroke effects rather than from limb disuse. Future work with this induced stroke model may improve understanding about the pathogenesis of skeletal fragility in stroke patients, in particular examining how osteovascular structure and function may contribute.

In this study, stroke decreased functional blood supply within the proximal tibia for two weeks. Perfusion was reduced in both limbs and tended to be lower in the affected side than in the unaffected side during the second and third week of recovery, mimicking the differential decreases

in lower leg blood flow observed in previous studies of chronic stroke patients with mild-to-moderate gait asymmetry [17,18]. The decreased perfusion post-stroke was transient, returning to sham levels by the third week of recovery in all stroke groups. Exercise accelerated this recovery, with perfusion in the stroke-exercise group returning to sham-sedentary levels by the second week, indicating that moderate exercise may be a viable strategy for increasing intraosseous blood supply following stroke. Although a number of studies in human stroke patients have shown increased limb perfusion with exercise following stroke [18,39], this study is the first to demonstrate that exercise restores perfusion for the microvasculature within bone, as well.

Perfusion measurements alone do not provide information about changes to the structure or composition of the vascular network, since perfusion could be reduced through decreased vascular density, increased vascular tone, reduced vessel diameter, decreased vascular permeability, or lower blood velocity. Stroke increased osteovascular volume, thickness, and branching in the distal femur by the end of the four-week recovery period. Considering the early transient reductions in bone perfusion in the proximal tibia, the increased osteovascular volume may be a compensatory response to offset perfusion deficits. However, a study examining the progression of these changes over time, and at the same site, is needed to assess the relative timing.

Ischemic stroke is associated with many conditions that could contribute to the reduced intraosseous perfusion observed, including increased vascular tone, increased vascular resistance, or decreased vasodilation. Vascular tone, which constricts blood vessels and decreases perfusion, was higher in cutaneous blood vessels in the paretic hands of patients with ischemic lesions [40] and in denervated arteries in rabbit ears [41], suggesting that stroke-related damage to the central nervous system may contribute to the reduced intraosseous perfusion. Stroke has been associated with increased vascular elasticity (a measure of vascular resistance) in the forearm [15], and

resistance arteries in cutaneous blood vessels in the paretic arms of stroke patients were also less

responsive to exogenous acetylcholine-induced vasodilation [42], which would increase vascular

resistance. However, the effect of stroke on the arteries that supply bone are unknown, and future

studies are required to determine whether increased vessel volume and branching are

compensatory for changes to vascular tone, resistance, or sensitivity to vasodilators following

stroke within long bones. Exercise restored the stroke-related perfusion deficits by the second

week of recovery but also reduced osteovascular volume relative to stroke-sedentary. Moderate

aerobic endurance training decreased peripheral vascular resistance in rats [43] and increased

vasodilator bioavailability in the leg in humans [44], suggesting that the exercise-related perfusion

increases may result, at least in part, from improved vascular function. Interestingly, exercise

mitigated the stroke-related increases in intraosseous vessel volume, further supporting the idea

that the increased vessel volume could be a pathological compensatory response and does not

necessarily reflect improved vascular function.

Although stroke increased the amount of blood vessels in bone, the structure and

composition of the osteovasculature were also affected, which may contribute to bone loss

following stroke. The combination of stroke and exercise decreased the relative number of blood

vessels near bone surfaces, which have been shown to be associated with sites of active bone

remodeling [21,22,45]. Exercise, in mitigating increased blood vessel volume post-stroke, may

have also reduced the number of branches in close contact with bone surfaces. Since exercise did

not reduce vessel-bone distance in the sham-exercise group, and since stroke did not decrease

vessel-bone distance in the stroke-sedentary group, the combined effect of exercise and stroke may

compound the detrimental changes in vessel proximity to bone surfaces. Although the relative area

of Type H endothelia was not affected by stroke in the tibia, stroke shifted the distribution of blood

vessel diameter in the femur, decreasing the amount of vessels between 6-22 μm in diameter,

which matches the size of Type H capillaries known to regulate osteogenesis [23]. Stroke causes

chronic inflammation [46], which stimulates angiogenesis [47] and thus may be responsible for

the increased vascular volume and branching following stroke, but inflammation also dysregulates

Notch signaling [48], the mechanism by which Type H capillaries regulate bone modeling [23,24].

Conversely, exercise nearly lowered the relative area of Type H endothelia in the proximal tibia

in both sham and stroke groups but did not affect the number of branches or vessel thickness

distribution in the distal femur. The reduction in relative area of Type H endothelial vessels could

be due to increased hypoxia-inducible factor (HIF) signaling, which, like Notch, is responsible for

Type H activity [24]. Since HIF is released by tissues experiencing hypoxia, HIF in the marrow

space may be increased with exercise following stroke if perfusion is negatively affected, and less

oxygen and fewer nutrients are able to reach bone tissue. Although some HIF signaling helps

recruit osteoblasts [24], compromised bone blood flow restricts oxygen availability to bone cells,

stimulating bone resorption [49], and reduced Type H activity could decrease osteogenesis [45].

Despite the increased amount of osteovasculature, the reduction in vessel-bone proximity,

resulting primarily from loss of small vessels, may explain the lack of exercise-induced gains in

bone microstructure following stroke, which we found in the same region using the same bones as

in this study [27].

In this study, we extended previous clinical findings that limb perfusion is reduced

following ischemic stroke and demonstrated for the first time, using a mouse model, that

intraosseous perfusion is also reduced during early stroke recovery, particularly in the affected

limb, and notably even in the absence of limb disuse. These functional deficits occurred in

conjunction with changes in osteovascular structure, including potentially compensatory increases

in vascular volume, branching, and the number of larger vessels. Further, although exercise

mitigated the negative effects of stroke on bone perfusion, the combination of stroke and exercise

altered the vessel proximity to bone to a less osteogenic arrangement with fewer small vessels

located near bone. These findings suggest that exercise like this moderate treadmill regime may

not be beneficial for osteovascular structure, although more studies are needed to examine if these

effects extend to other exercise therapies or rehabilitation strategies. Examining potential

mechanisms for these changes in osteovascular structure and function following stroke may

provide insight for mitigating skeletal fragility in human stroke patients.

Acknowledgments

We thank Dr. Eva Johannes and Dr. Mariusz Zareba for confocal microscopy support; and Dr.

Consuelo Arellano for statistical consulting. This work was performed in part at the Cellular and

Molecular Imaging Facility (CMIF) at North Carolina State University, which is supported by the

State of North Carolina and the National Science Foundation.

Sources of Funding

Research reported in this publication was supported by the Eunice Kennedy Shriver National

Institute of Child Health and Human Development (NICHD) of the NIH under award number

K12HD073945 and by the American Heart Association (AHA) under award number

7GRNT33710007. The content is solely the responsibility of the authors and does not necessarily

represent the official views of the NIH.

Disclosures

The authors have nothing to disclose.

References

[1] E.J. Benjamin, P. Muntner, A. Alonso, M.S. Bittencourt, C.W. Callaway, A.P. Carson, A.M. Chamberlain, A.R. Chang, S. Cheng, S.R. Das, F.N. Delling, L. Djousse, M.S.V. Elkind, J.F. Ferguson, M. Fornage, L.C. Jordan, S.S. Khan, B.M. Kissela, K.L. Knutson, T.W. Kwan, D.T. Lackland, T.T. Lewis, J.H. Lichtman, C.T. Longenecker, M.S. Loop, P.L. Lutsey, S.S. Martin, K. Matsushita, A.E. Moran, M.E. Mussolino, M. O'Flaherty, A. Pandey, A.M. Perak, W.D. Rosamond, G.A. Roth, U.K.A. Sampson, G.M. Satou, E.B. Schroeder, S.H. Shah, N.L. Spartano, A. Stokes, D.L. Tirschwell, C.W. Tsao, M.P. Turakhia, L.B. VanWagner, J.T. Wilkins, S.S. Wong, S.S. Virani, American Heart Association Council on Epidemiology and Prevention Statistics Committee and Stroke Statistics Subcommittee, Heart Disease and Stroke Statistics-2019 Update: A Report From the American Heart Association, Circulation. 139 (2019) e56–e528. doi:10.1161/CIR.0000000000000659.

[2] J. Kanis, A. Oden, O. Johnell, Acute and Long-Term Increase in Fracture Risk After Hospitalization for Stroke, Stroke. 32 (2001) 702–706. doi:10.1161/01.STR.32.3.702.

[3] L. Jørgensen, B.K. Jacobsen, T. Wilsgaard, J.H. Magnus, Walking after Stroke: Does It Matter? Changes in Bone Mineral Density Within the First 12 Months after Stroke. A Longitudinal Study, Osteoporos. Int. 11 (2000) 381–387. doi:10.1007/s001980070103.

[4] L. Jørgensen, B.K. Jacobsen, Functional status of the paretic arm affects the loss of bone mineral in the proximal humerus after stroke: a 1-year prospective study, Calcif. Tissue Int. 68 (2001) 11–15. doi:10.1007/s002230001165.

[5] A. Ramnemark, L. Nyberg, B. Borssén, T. Olsson, Y. Gustafson, Fractures after Stroke, Osteoporos. Int. 8 (1998) 92–95. doi:10.1007/s001980050053.

[6] A. Ramnemark, L. Nyberg, R. Lorentzon, U. Englund, Y. Gustafson, Progressive

Hemiosteoporosis on the Paretic Side and Increased Bone Mineral Density in the Nonparetic

Arm the First Year after Severe Stroke, Osteoporos. Int. 9 (1999) 269–275.

doi:10.1007/s001980050147.

[7] A. Fisher, W. Srikusalanukul, M. Davis, P. Smith, Poststroke Hip Fracture: Prevalence,

Clinical Characteristics, Mineral-Bone Metabolism, Outcomes, and Gaps in Prevention,

Stroke Res. Treat. 2013 (2013). doi:10.1155/2013/641943.

[8] G.S. Beaupré, H.L. Lew, Bone-Density Changes After Stroke, Am. J. Phys. Med. Rehabil.

85 (2006) 464–472. doi:10.1097/01.phm.0000214275.69286.7a.

[9] H.M. Frost, Bone's Mechanostat: A 2003 Update, Anat. Rec. A. Discov. Mol. Cell. Evol.

Biol. 275A (2003) 1081–1101. doi:10.1002/ar.a.10119.

[10]   J.J. Eng, M.Y.C. Pang, M.C. Ashe, Balance, falls, and bone health : role of exercise in

reducing fracture risk after stroke, J. Rehabil. Res. Dev. 45 (2008) 297–314.

[11]   M.Y.C. Pang, R.W.K. Lau, The Effects of Treadmill Exercise Training on Hip Bone

Density and Tibial Bone Geometry in Stroke Survivors: A Pilot Study, Neurorehabil. Neural

Repair. 24 (2010) 368–376. doi:10.1177/1545968309353326.

[12]   R.P.V. Peppen, G. Kwakkel, S. Wood-Dauphinee, H.J. Hendriks, P.J.V. der Wees, J.

Dekker, The impact of physical therapy on functional outcomes after stroke: what's the

evidence?, Clin. Rehabil. 18 (2004) 833–862. doi:10.1191/0269215504cr843oa.

[13]   K. Borschmann, M.Y.C. Pang, S. Iuliano, L. Churilov, A. Brodtmann, E.I. Ekinci, J.

Bernhardt, Changes to volumetric bone mineral density and bone strength after stroke: a

prospective study, Int. J. Stroke Off. J. Int. Stroke Soc. 10 (2015) 396–399.

doi:10.1111/ijs.12228.

[14]   A.M. Weatherholt, K.G. Avin, A.L. Hurd, J.L. Cox, S.T. Marberry, B.G. Santoni, S.J.

Warden, Peripheral quantitative computed tomography (pQCT) predicts humeral diaphysis

torsional mechanical properties with good short-term precision, J. Clin. Densitom. Off. J. Int.

Soc. Clin. Densitom. 18 (2015) 551–559. doi:10.1016/j.jocd.2014.10.002.

[15]   M.Y.C. Pang, F.Z.H. Yang, A.Y.M. Jones, Vascular Elasticity and Grip Strength Are

Associated With Bone Health of the Hemiparetic Radius in People With Chronic Stroke:

Implications for Rehabilitation, Phys. Ther. 93 (2013) 774–785. doi:10.2522/ptj.20120378.

[16]   S. Stegen, G. Carmeliet, The skeletal vascular system – Breathing life into bone tissue,

Bone. 115 (2018) 50–58. doi:10.1016/j.bone.2017.08.022.

[17]   F.M. Ivey, A.W. Gardner, C.L. Dobrovolny, R.F. Macko, Unilateral Impairment of Leg

Blood Flow in Chronic Stroke Patients, Cerebrovasc. Dis. Basel. 18 (2004) 283–9.

[18]   F.M. Ivey, C.E. Hafer-Macko, A.S. Ryan, R.F. Macko, Impaired Leg Vasodilatory

Function After Stroke: Adaptations With Treadmill Exercise Training, Stroke. 41 (2010)

2913–2917. doi:10.1161/STROKEAHA.110.599977.

[19]   J. Reeve, M. Arlot, R. Wootton, C. Edouard, M. Tellez, R. Hesp, J.R. Green, P.J.

Meunier, Skeletal Blood Flow, Iliac Histomorphometry, and Strontium Kinetics in

Osteoporosis: A Relationship Between Blood Flow and Corrected Apposition Rate, J. Clin.

Endocrinol. Metab. 66 (1988) 1124–1131. doi:10.1210/jcem-66-6-1124.

[20]   J.F. Griffith, D.K. Yeung, P.H. Tsang, K.C. Choi, T.C. Kwok, A.T. Ahuja, K.S. Leung,

P.C. Leung, Compromised Bone Marrow Perfusion in Osteoporosis, J. Bone Miner. Res. 23

(2008) 1068–1075. doi:10.1359/jbmr.080233.

[21]   H.B. Kristensen, T.L. Andersen, N. Marcussen, L. Rolighed, J.-M. Delaisse, Increased

473   presence of capillaries next to remodeling sites in adult human cancellous bone, J. Bone

474   Miner. Res. 28 (2013) 574–585. doi:10.1002/jbmr.1760.

475   [22]   R. Prisby, A. Guignandon, A. Vanden-Bossche, F. Mac-Way, M.-T. Linossier, M.

476   Thomas, N. Laroche, L. Malaval, M. Langer, Z.-A. Peter, F. Peyrin, L. Vico, M.-H. Lafage-

477   Proust, Intermittent PTH(1–84) is osteoanabolic but not osteoangiogenic and relocates bone

478   marrow blood vessels closer to bone-forming sites, J. Bone Miner. Res. 26 (2011) 2583–

479   2596. doi:10.1002/jbmr.459.

480   [23]   S.K. Ramasamy, A.P. Kusumbe, L. Wang, R.H. Adams, Endothelial Notch activity

481   promotes angiogenesis and osteogenesis in bone, Nature. 507 (2014) 376–380.

482   doi:10.1038/nature13146.

483   [24]   A.P. Kusumbe, S.K. Ramasamy, R.H. Adams, Coupling of angiogenesis and

484   osteogenesis by a specific vessel subtype in bone, Nature. 507 (2014) 323–328.

485   doi:10.1038/nature13145.

486   [25]   J. Koizumi, Y. Yoshida, T. Nakazawa, G. Ooneda, Experimental studies of ischemic

487   brain edema, I: a new experimental model of cerebral embolism in rats in which recirculation

488   can be introduced in the ischemic area, Jnp J Stroke. 8 (1986).

489   [26]   E.Z. Longa, P.R. Weinstein, S. Carlson, R. Cummins, Reversible middle cerebral artery

490   occlusion without craniectomy in rats., Stroke. 20 (1989) 84–91.

491   doi:10.1161/01.STR.20.1.84.

492   [27]   N.J. Hanne, A.J. Steward, M.R. Sessions, H.L. Thornburg, H. Sheng, J.H. Cole, Stroke

493   prevents exercise-induced gains in bone microstructure but not composition in mice,

494   BioRxiv. (2019) 708388. doi:10.1101/708388.

495   [28]   H. Sakai, H. Sheng, R.B. Yates, K. Ishida, R.D. Pearlstein, D.S. Warner, Isoflurane

496   provides long-term protection against focal cerebral ischemia in the rat, Anesthesiology. 106

497   (2007) 92–99; discussion 8-10.

498   [29]   N.J. Hanne, E.D. Easter, S. Stangeland-Molo, J.H. Cole, A Minimally Invasive

499   Technique for Serial Intraosseous Perfusion Measurements in the Murine Tibia Using Laser

500   Doppler Flowmetry, BioRxiv. (2019) 708453. doi:10.1101/708453.

501   [30]   N.J. Hanne, E.D. Easter, J.H. Cole, Validating Minimally Invasive Laser Doppler

502   Flowmetry for Serial Bone Perfusion Measurements in Mice, BioRxiv. (2019) 708412.

503   doi:10.1101/708412.

504   [31]   G. Kerckhofs, S. Stegen, N. van Gastel, A. Sap, G. Falgayrac, G. Penel, M. Durand, F.P.

505   Luyten, L. Geris, K. Vandamme, T. Parac-Vogt, G. Carmeliet, Simultaneous three-

506   dimensional visualization of mineralized and soft skeletal tissues by a novel microCT

507   contrast agent with polyoxometalate structure, Biomaterials. 159 (2018) 1–12.

508   doi:10.1016/j.biomaterials.2017.12.016.

509   [32]   M. Doube, M.M. Kłosowski, I. Arganda-Carreras, F.P. Cordelières, R.P. Dougherty, J.S.

510   Jackson, B. Schmid, J.R. Hutchinson, S.J. Shefelbine, BoneJ: Free and extensible bone

511   image analysis in ImageJ, Bone. 47 (2010) 1076–1079. doi:10.1016/j.bone.2010.08.023.

512   [33]   J. Schindelin, I. Arganda-Carreras, E. Frise, V. Kaynig, M. Longair, T. Pietzsch, S.

513   Preibisch, C. Rueden, S. Saalfeld, B. Schmid, J.-Y. Tinevez, D.J. White, V. Hartenstein, K.

514   Eliceiri, P. Tomancak, A. Cardona, Fiji: an open-source platform for biological-image

515   analysis, Nat. Methods. 9 (2012) 676–682. doi:10.1038/nmeth.2019.

516   [34]   M.P. Pusztaszeri, W. Seelentag, F.T. Bosman, Immunohistochemical expression of

517   endothelial markers CD31, CD34, von Willebrand factor, and Fli-1 in normal human tissues,

J. Histochem. Cytochem. Off. J. Histochem. Soc. 54 (2006) 385–395.

doi:10.1369/jhc.4A6514.2005.

[35]   A.P. Kusumbe, S.K. Ramasamy, A. Starsichova, R.H. Adams, Sample preparation for

high-resolution 3D confocal imaging of mouse skeletal tissue, Nat. Protoc. Lond. 10 (2015)

1904–1914. doi:http://dx.doi.org.prox.lib.ncsu.edu/10.1038/nprot.2015.125.

[36]   R.C. Littell, ed., SAS for mixed models, 2nd ed, SAS Institute, Inc, Cary, N.C, 2006.

[37]   J.N. Stabley, R.D. Prisby, B.J. Behnke, M.D. Delp, Chronic skeletal unloading of the rat

femur: Mechanisms and functional consequences of vascular remodeling, Bone. 57 (2013)

355–360. doi:10.1016/j.bone.2013.09.003.

[38]   P.N. Colleran, M.K. Wilkerson, S.A. Bloomfield, L.J. Suva, R.T. Turner, M.D. Delp,

Alterations in skeletal perfusion with simulated microgravity: a possible mechanism for bone

remodeling, J. Appl. Physiol. 89 (2000) 1046–1054.

[39]   Billinger Sandra A., Gajewski Byron J., Guo Lisa X., Kluding Patricia M., Single Limb

Exercise Induces Femoral Artery Remodeling and Improves Blood Flow in the Hemiparetic

Leg Poststroke, Stroke. 40 (2009) 3086–3090. doi:10.1161/STROKEAHA.109.550889.

[40]   A.G. Herbaut, J.D. Cole, E.M. Sedgwick, A cerebral hemisphere influence on cutaneous

vasomotor reflexes in humans., J. Neurol. Neurosurg. Psychiatry. 53 (1990) 118–120.

[41]   R.D. Bevan, A. Clementson, E. Joyce, J.A. Bevan, Sympathetic denervation of resistance

arteries increases contraction and decreases relaxation to flow, Am. J. Physiol.-Heart Circ.

Physiol. 264 (1993) H490–H494. doi:10.1152/ajpheart.1993.264.2.H490.

[42]   J.-S. Wang, C.F. Yang, M.-Y. Liaw, M.-K. Wong, Suppressed cutaneous endothelial

vascular control and hemodynamic changes in paretic extremities with edema in the

extremities of patients with hemiplegia, Arch. Phys. Med. Rehabil. 83 (2002) 1017–1023. doi:10.1053/apmr.2002.33235.

[43]   M.K. Karhunen, M.P. Rämö, R. Kettunen, L. Hirvonen, The cardiovascular effects of deconditioning after endurance training in rats, Acta Physiol. Scand. 133 (1988) 307–314. doi:10.1111/j.1748-1716.1988.tb08412.x.

[44]   J. Sugawara, H. Komine, K. Hayashi, M. Yoshizawa, T. Otsuki, N. Shimojo, T. Miyauchi, T. Yokoi, S. Maeda, H. Tanaka, Systemic α-adrenergic and nitric oxide inhibition on basal limb blood flow: effects of endurance training in middle-aged and older adults, Am. J. Physiol.-Heart Circ. Physiol. 293 (2007) H1466–H1472. doi:10.1152/ajpheart.00273.2007.

[45]   T.L. Andersen, T.E. Sondergaard, K.E. Skorzynska, F. Dagnaes-Hansen, T.L. Plesner, E.M. Hauge, T. Plesner, J.-M. Delaisse, A Physical Mechanism for Coupling Bone Resorption and Formation in Adult Human Bone, Am. J. Pathol. 174 (2009) 239–247. doi:10.2353/ajpath.2009.080627.

[46]   K.L. Lambertsen, K. Biber, B. Finsen, Inflammatory cytokines in experimental and human stroke, J. Cereb. Blood Flow Metab. 32 (2012) 1677–1698. doi:10.1038/jcbfm.2012.88.

[47]   C. Costa, J. Incio, R. Soares, Angiogenesis and chronic inflammation: cause or consequence?, Angiogenesis. 10 (2007) 149–166. doi:10.1007/s10456-007-9074-0.

[48]   T. Quillard, J. Devallière, S. Coupel, B. Charreau, Inflammation dysregulates Notch signaling in endothelial cells: Implication of Notch2 and Notch4 to endothelial dysfunction, Biochem. Pharmacol. 80 (2010) 2032–2041. doi:10.1016/j.bcp.2010.07.010.

561   [49]   T.R. Arnett, D.C. Gibbons, J.C. Utting, I.R. Orriss, A. Hoebertz, M. Rosendaal, S.

562   Meghji, Hypoxia is a major stimulator of osteoclast formation and bone resorption, J. Cell.

563   Physiol. 196 (2003) 2–8. doi:10.1002/jcp.10321.

Tables

Table I. Adipocyte Microstructure in the Distal Femoral Metaphysis

Trait	Sham		Stroke	
	Sedentary	**Exercise**	**Sedentary**	**Exercise**
Ad.V/Ma.V (%)	0.15 ± 0.77	0.16 ± 0.55	0.15 ± 0.77	0.45 ± 0.55
Ad.D (#/mm^3)	127.7 ± 166.4	113.5 ± 117.7	118.4 ± 166.4	237.0 ± 117.7*
Ad.Th (µm)	20.5 ± 10.3	20.7 ± 7.3	20.2 ± 10.3	23.1 ± 7.3

least squares mean ± 95% confidence interval

*: $p < 0.10$ stroke-exercise vs. sham-exercise

Figures and Legends

Figure 1. Study design. Mice received stroke or sham surgery, followed by four weeks of treadmill or sedentary activity. Intraosseous perfusion was measured weekly during recovery in affected and unaffected tibiae with laser Doppler flowmetry (LDF). After sacrifice, osteovascular structure and composition were characterized with contract-enhanced computed tomography (CT) and immunofluorescence (IF) microscopy.

Figure 2. A) Intraosseous perfusion in proximal tibiae measured by laser Doppler flowmetry. B) Perfusion decreased in both limbs at Weeks 1 and 2 post-stroke and was increased with exercise at Week 2 (least squares mean ± 95% confidence interval across both limbs). C) Comparing limbs for stroke, perfusion was nearly decreased in the affected (Aff) relative to unaffected (Un) limb during Week 2. a: $p < 0.05$ stroke vs. sham (main effect); b′: $p < 0.1$ exercise vs. sedentary (main effect); c′: $p < 0.1$ stroke-exercise vs. stroke-sedentary; d′: $p < 0.1$ Aff vs. Un limb within stroke group.

Figure 3. A) Osteovascular structure in the distal femoral metaphysis. B) Blood vessel volume fraction was increased with stroke, while exercise mitigated the effect of stroke. C) Mean vessel thickness was not altered by stroke or exercise. a: p < 0.05 stroke vs. sham (main effect); b: p < 0.05 exercise vs. sedentary (main effect); c: p < 0.05 stroke-exercise vs. stroke-sedentary.

Figure 4. Osteovascular network in the distal femoral metaphysis. A) Vascular branching, B) number of junctions, C) number of triple points, and D) number of quadruple points were increased with stroke. a: $p < 0.05$ stroke vs. sham (main effect); a′: $p < 0.10$ stroke vs. sham (main effect).

Figure 5. Osteovascular arrangement in the distal femoral metaphysis. A) Distribution of vessel thickness was shifted with stroke, with fewer smaller vessels and more larger vessels. Exercise mitigated the stroke-induced increase in large vessels. B) Both stroke and exercise reduced the relative number of blood vessels nearer to bone surfaces and increased the number farther from bone surfaces, especially for stroke-exercise. a: $p < 0.05$ stroke vs. sham (main effect); a′: $p < 0.10$ stroke vs. sham (main effect); b: $p < 0.05$ exercise vs. sedentary (main effect); c: $p < 0.05$ stroke-exercise vs. stroke-sedentary; e: $p < 0.05$ stroke-sedentary vs. sham-sedentary; e′: $p < 0.10$ stroke-sedentary vs. sham-sedentary; f: $p < 0.05$ stroke-exercise vs. sham-exercise; f′: $p < 0.10$ stroke-exercise vs. sham-exercise.

Figure 6. A) Representative longitudinal section in proximal tibia metaphysis, showing osteovascular composition with immunofluorescence. B) Endomucin (EMCN)$^+$ and C) CD31$^+$ areas were unaffected by stroke or exercise. D) Relative area of Type H endothelial cells (EMCN$^+$ and CD31$^+$) was nearly reduced by exercise. b′: p < 0.1 exercise vs. sedentary (main effect).