Antibody Targeting of B7-H4 Enhances the Immune Response in Urothelial Carcinoma

Joseph R. Podojil[1], Alex P. Glaser[2,3, 4], Dylan Baker[5,6], Damiano Fantini[2,3], Yanni Yu[2,3], Valerie Eaton[1], Santhosh Sivajothi[5, 6], Paul Robson[5, 6], Stephen D. Miller[1,*], Joshua J. Meeks[2, 3*]

Author Affiliation:
[1]Department of Microbiology and Immunology, Feinberg School of Medicine, Chicago, IL
[2]Department of Urology, Feinberg School of Medicine, Chicago, IL
[3]Department of Biochemistry, and Molecular Genetics, Feinberg School of Medicine, Chicago, IL
[4]Division of Urology, Department of Surgery, NorthShore University Health Systems
[5]The Jackson Laboratory for Genomic Medicine, Farmington, Connecticut.
[6]Department of Genetics and Genome Sciences, Institute for Systems Genomics, University of Connecticut, Farmington, Connecticut.

[*]Co-Corresponding Author (these authors have contributed equally):
Joshua J. Meeks, joshua-meeks@northwestern.edu
Stephen D. Miller s-d-miller@northwestern.edu

Keywords: immunotherapy, bladder cancer, macrophage, T cell, checkpoint

Abstract

Locally advanced urothelial carcinoma has a poor survival despite a response to immunotherapy in approximately 25% of patients. To develop new therapies targeting other immune checkpoints, we identified increased expression of the T cell inhibitor protein, B7-H4 (VTCN1, B7S1, B7X), during tumor development of murine bladder cancer. Evaluation of B7-H4 in human bladder cancer by single-cell RNA-Seq and immune mass cytometry showed that B7-H4 is expressed by luminal tumors that are not normally responsive to PD-1 inhibitors. Additionally, overexpression B7-H4 was associated with significantly worse patient survival. In support of clinical translation, treatment of human monocyte and T cell co-cultures with a B7-H4 blocking antibody resulted in enhanced IFN-γ secretion by $CD4^+$ and $CD8^+$ T cell. While the study of B7-H4 in mouse cancer models has been difficult using tumor cell lines, our findings show that B7-H4 expression increased at 3 months after exposure to BBN on $CD11b^+$ monocytes/macrophages and delivery of anti-B7-H4 antibody resulted in decreased tumor size and increased CD8 immune cell infiltration with increased $CD8^+$/IFN-γ-producing T cells and a complimentary decrease in tumor infiltrating T regulatory cells (Tregs). Furthermore, treatment with a combination of anti-PD-1 and anti-B7-H4 antibodies resulted in a significant reduction in tumor stage. This data suggests that novel anti-B7-H4 antibodies maybe a viable target for bladder cancers unresponsive to PD-1 checkpoint inhibitors.

Introduction

The anti-tumor immune response is dynamic and evasion of the immune system by the tumor is hallmark of cancer. Due to the expression of neoantigens driven by tumor-derived mutations, cells of the innate and adaptive immune system are capable of mounting vigorous anti-tumor responses. Yet, many tumors are able to evade this response by upregulating checkpoint inhibitors that suppress immune activation. While CTLA-4 and PD-L1 are the most well-described checkpoint proteins, B7-H4 (B7S1, B7x, VTCN1), a B7 family member expressed on antigen presenting cells and/or tumors, and can inhibit T cell-mediated inflammatory responses. The majority of work focused on elucidating the functional immunoregulatory role of B7-H4 has involved assessment of the ability of an agonistic B7-H4-immunoglobulin fusion protein (B7-H4Ig) in the treatment of autoimmune disease, and the presence of B7-H4 expression in human cancers [as reviewed in (1)]. B7-H4 functionally decreases inflammatory $CD4^+$ T cell directly and indirectly via B7-H4-induced increases in both the number and function of regulatory $CD4^+$ T cells (2-4). However, evidence for the ability of B7-H4 blockade to decrease tumor burden is lacking due to the limited number of mouse tumors that natively express B7-H4, likely secondary to non-orthotopic placement of ex-vivo derived cell lines. However, it was reported that genetic ablation of *B7-h4 (Vtcn1)* in a model of liver cancer was associated with increased $CD8^+$ infiltration of the tumor and decreased exhaustion of $CD8^+$ T cells (5).

In contrast to the majority of mouse tumor models, the expression level of B7-H4 protein in human ovarian cancer has been related to cancer types, cancer stage, the numbers of Tregs, and patient survival (6-8). Additionally, more than 90% of endometrial and breast cancers express B7-H4 protein (9), and B7-H4 expression in renal cell carcinoma (10-12), melanoma (13), breast (14), lung (15,16), gastric (17), colorectal (18), pancreatic (19), and prostate (20) cancer are associated with one or more clinicopathological factors, including tumor size, cancer stage, metastasis, progression, prognosis, survival, the intensity of tumor infiltrating T cells, the

infiltration intensity of Tregs and tumor associated macrophages, disease recurrence or mortality. Therefore, targeting B7-H4 may be an alternative strategy to reinvigorate tumor-specific T cell responses.

Urothelial carcinoma is the fifth most common cancer in the US and has the second worst survival for patients with metastasis at only 5% at years (21). While systemic chemotherapy has been the standard of care for treatment of patients with metastatic urothelial carcinoma, in 2016 antibodies targeting immune checkpoint blockade (ICB), specifically PD1 and PDL1 were FDA approved (22). However, only 21% of patients with metastatic urothelial carcinoma refractory to chemotherapy respond to anti-PD1 antibodies (23). While the factors that determine clinical response are not completely known, features such as immune cell infiltration and high total mutation burden have been associated with an increased response (24). Not all studies have demonstrated that PDL1 expression is associated with improved survival following anti-PD-1 therapy, suggesting that multiple aspects of the regulation of immune responses to tumor remain unclear (25). Thus, most patients will progress and would benefit from additional therapies, targeting distinct, non-overlapping immune regulatory pathways.

Results

B7-H4 expression increases during bladder cancer development and is overexpressed in human bladder cancer

To investigate the interaction between bladder cancer initiation and progression we used the syngeneic, carcinogen-induced mouse model of bladder cancer derived from timed exposure to N-butyl-N-(4-hydroxybutyl)-nitrosamine (BBN). Our laboratory has previously validated the genomic credibility of this model that develops muscle-invasive bladder cancer (MIBC) between three and five months after initial exposure (37). Evaluation of the mutation signature most closely aligns with those caused by smoking, justifying the clinical relevance of BBN to bladder cancer, in which over 60% of cancers are attributed to smoking carcinogens (38). The BBN model of bladder cancer offers the distinct advantage that immune effector and regulatory processes can be assessed both in the bladder and peripheral immune organs from the initiation of tumorigenesis until the development of palpable tumors. Therefore, C57BL/6 (B6) mice were provided BBN and gene expression profiling was performed early (1, 2, and 4 weeks) after exposure to the carcinogen. We identified early expression of immune response-related genes that increased from low levels at one week to higher levels at two weeks of BBN exposure (Figure 1A). This included transcripts for CD8, CD4, and pathways associated with regulation of inflammation. For example, markers of T cell exhaustion (PD-L1, LAG-3 and Tim-3) and regulation (FoxP3 and Nrp-1) increased by 2 weeks, and B7-H4, a protein expressed by APCs that is known to inhibit T cell function via both direct and indirect mechanisms (4,39), was significantly increased by four weeks and corresponded to a decrease in CD8 and CD4 transcripts within the bladder (Figure 1A). To determine if B7-H4 could play a role in human bladder cancer, we evaluated the expression of B7-H4 in the cancer genome atlas for muscle invasive bladder cancer (MIBC). Patients with high expression of B7-H4 had significantly worse overall survival compared to those with lower expression (21.85 vs 34.03 months, p=0.03) (Figure 1B). Additionally, enriched B7-H4 expression positively correlated with B cells, iDCs, and macrophage

transcripts, however B7-H4 expression did not correlate with T cells and IFN-γ levels present from MIBCs (29) (Supplemental Figure 1). Unlike other immune checkpoints (such as PD-L1, PD-L1, and CTLA-4), expression of B7-H4 does not appear to correlate with neoantigens, total mutation burden, APOBEC mutations or gene signatures (Supplemental Figure 2). We also found significant B7-H4 enrichment in bladder tumor subtypes that had relatively low expression of PD-L1, PD-1, and CTLA-4 including luminal, luminal papillary and luminal infiltrated tumors using the 2017 TCGA molecular subtyping classification (Figure 1C). Since expression of PD-L1 is directly associated with response to anti-PD-L1 and anti-PD-1 therapy, treatment of bladder cancer via blockade with a checkpoint inhibitor expressed in a different tumor subtype could lead to improved overall outcomes for patients. Thus, we were interested in the location of B7-H4 expression in bladder cancers, including localization of the tumor and tumor microenvironment. Using immunohistochemistry, we found expression of B7-H4 localized to immune cells resident in normal bladder, with increased expression in low and higher grade papillary tumors, respectively (Figure 1D, Right, middle panels). Yet, using the BBN mouse carcinogen model, B7-H4 expression co-localized with $CD11b^{+}$ myeloid cells (Figure 1D, left) only. This suggests that expression of B7-H4 was likely localized to the myeloid lineage only and not expressed by the tumors in the BBN mouse model. To confirm this localization, we performed single-cell RNA-Seq of a stage Ta high grade bladder tumor (Figure 1E). *VTCN1* (B7H4) mRNA was detected on both urothelial cells (*EPCAM*+) and myeloid cells (*CD68*+) by scRNA sequencing. While *VTCN1* was expressed ubiquitously throughout the urothelial clusters, myeloid *VTCN1* was found on a subset of cells. Myeloid *VTCN1* shows scattered expression across the myeloid cluster without restriction to any one particular subcluster. To futher confirm the cellular localization of B7-H4, we performed imaging mass cytometry, that demonstrated localization of B7-H4 to CD68+ myeloid cells. Consistent with scRNA sequencing many CD68+ cells did not show expression of B7-H4 by imaging mass cytometry.

Temporal expression of PD-L1 and B7-H4 and their respective ligands in BBN-induced bladder cancer

Mice given BBN in drinking water, develop carcinoma *in situ* (CIS) by 2-3 months, and mice present with tumors by 3 months and bulky tumors by 4 months (40). We next went on to validate the transcript data presented in Figure 1, by determining how the number of $CD4^+$ T cells, $CD8^+$ T cells, and monocytes/macrophages changed over time during BBN-induced bladder cancer development. Bladders (Figure 2) were collected at 0.5, 1, 2, 4, and 5 months after the initiation of BNN-induced bladder cancer. The results show a bi-phasic pattern in the number of $CD4^+$ T cells, $CD8^+$ T cells, and monocytes within the bladder over time (Figure 2A-C). Similar to our RNA-Seq results, we identified a peak in the numbers of pro-inflammatory T cells ($CD4^+$ and $CD8^+$) at 1 month, but a decline in cell numbers by 2 months. The numbers of bladder infiltrating $CD4^+$ and $CD8^+$ cells did not increase until 3 months concomitant with the development of carcinoma. During BBN exposure, the gradual increase in $CD4^+$ and $CD8^+$ T cells was associated with increased expression of the exhaustion marker, PD-1 (Figure 2D-E). The numbers of $CD4^+$ T cells expressing the neuropilin-1/semaphorin3a (Nrp-1/Sema3a), B7-H4 receptor (39) was increased at 1 and 5 months (Figure 2G) while B7-H4 receptor-positive $CD8^+$ T cells increased at 0.5 and 1 months (Figure 2H). Monocytes ($CD11b^+/Ly6C^+/Ly6G^-$) increased sharply at 1 month and subsequently decreased concordant with the decrease in T cell numbers. The infiltrating monocytes expressed PD-L1 at 1 month with a second wave of increased expression at 5 months (Figure 2F). There was a small peak of B7-$H4^+$ monocytes at one month, followed by a decline, and a second peak concordant with tumor development and immune cell infiltration of bladder tumors at 5 months (Figure 2I). Additionally, the bi-phasic pattern of PD-L1 and B7-H4 expression correlated with both the numbers of $CD4^+$ Treg (Figure 2J) and effector $CD4^+$ T and $CD8^+$ T cells (Figure 2K and 2L). These findings confirm B7-H4 expression by bladder-infiltrating monocytes concordant with $CD4^+$ and $CD8^+$ TILs suggestive of induction of an anti-inflammatory regulatory

mechanism. Based on these results, we investigated the blockade of B7-H4 as a potential therapy to increase T cell responses by blocking B7-H4-mediated T cell regulation.

Anti-B7-H4 blocks the immunoregulatory function of B7-H4Ig and increases mouse and human $CD4^+$ and $CD8^+$ T cell function

To functionally demonstrate that anti-B7-H4 can inhibit the regulatory function of B7-H4, we tested the ability of anti-B7-H4 to block the immunoregulatory function of human B7-H4Ig. SJL/J lymph node cells ($5x10^5$ cells/well) were labeled with CFSE, and stimulated with anti-CD3 (1μg/ml) plus control Ig, human B7-H4Ig, and/or anti-B7-H4 (10μg/ml) as indicated (Figure 3A). On Day 3 of culture, the cells were collected and the level of $CD4^+$ T cell CFSE dilution assessed via flow cytometry. While anti-B7-H4 alone did not alter the extent of $CD4^+$ T cell proliferation, the addition of anti-B7-H4 inhibited hB7-H4Ig-induced suppression of $CD4^+$ T cell proliferation. As a secondary confirmation of the CFSE findings, replicate wells were pulsed with ^{3}H-TdR on Day 1 and cultures harvested on Day 3. The level of ^{3}H-TdR incorporation (counts per minute, CPM) confirmed that anti-B7-H4 blocked the inhibitory function of human B7-H4Ig (Figure 3A). As a further confirmation of the ability of anti-B7-H4 to block hB7-H4Ig function, SJL/J mice were primed with $PLP_{139-151}$/CFA, and treated 3 times per week for 2 weeks beginning at the time of priming (Day 0) with control Ig, hB7-H4Ig, or anti-B7-H4 as indicated. Mice were observed for paralytic disease symptoms. As we previously reported (4), hB7-H4Ig treatment significantly decreased disease severity (Figure 3B). While anti-B7-H4 treatment alone did not modulate the level of disease severity, treatment with anti-B7H4 blocked the hB7-H4Ig-induced decrease in disease severity (Figure 3B). In addition, anti-B7H4 blocked the hB7-H4Ig-induced decrease in *in vivo* $CD4^+$ T cell function as measured by delayed-type hypersensitivity (DTH) responses to ear challenge with $PLG_{139-151}$. Taken together, these findings show that anti-B7-H4 blocks hB7-H4Ig-induced regulatory function on mouse T cell responses both *in vitro* and *in vivo*.

To determine if anti-B7-H4 could block B7-H4-induced regulation of human T cells, $CD14^+$ monocytes from healthy donors were cultured for 3 days with IL-10, IL-6, and IL-10 plus IL-6 (10ng/ml). Adherent cells were collected and analyzed via flow cytometry. Upregulated expression of B7-H4, CD80, CD86, PD-L1, and PD-L2 expression was identified following stimulation with IL-10 and/or IL-6 (Figure 4A). Therefore, IL-10 + IL-6 stimulated $CD14^+$ APCs were co-cultured with autologous $CD4^+$ and $CD8^+$ T cells in anti-CD3-coated wells $\pm$ control Ig or anti-B7-H4 and proliferation and IFN-γ production measured on 3 d thereafter. While addition of anti-B7-H4 did not significantly enhance proliferation of $CD4^+$ (Figure 4B) or $CD8^+$ T cells (Figure 4D), anti-B7-H4 induced a significant increase in production of IFN-γ by both $CD4^+$ and $CD8^+$ T cells (Figure 4C and 4E). Thus, anti-B7-H4 blockade increased the function of both human $CD4^+$ and $CD8^+$ T cells stimulated with B7-H4 expressing monocytes.

Anti-B7-H4 treatment increases $CD8^+$ T cell infiltration into the bladder during BBN-induced bladder cancer and IFN-γ production by splenic $CD8^+$ T cells

After confirmation of the functional activity of our B7-H4 antibody, we tested its activity in the BBN carcinogen model of bladder cancer. Mice received BBN containing water for four months and received treatment with a species and isotype matched Control Ab, anti-PD-1, or anti-B7-H4 for once weekly for four total treatments. After treatment, animals were placed on normal drinking water for 30 days followed by tumor analysis (Figure 5A). Anti-B7-H4 treatment led to a decrease in the number of advanced tumors (70% compared to 50% stage 3 or greater), and a significant increase in the number of $CD8^+$ bladder-infiltrating T cells (Figure 5B, C). In contrast, anti-B7-H4 treatment decreased the number of $FoxP3^+$ Tregs (Fig 5B,D), indicating that anti-B7-H4 treatment had shifted the $CD8^+$ T cell to Treg ratio in favor of an anti-tumor response. While treatment of mice with anti-PD-1 or anti-B7-H4 did not alter the number of total splenic CD45hi cells, $CD4^+$ T cells, or $CD8^+$ T cells (Figure 5E), both anti-PD-1 and anti-B7-H4 treatments significantly

increased the number of $CD8^+$/IFN-γ^+ cells present within the spleen compared to control IgG-treated mice (Figure 5F). Additionally, total splenocytes from the treatment groups were activated *ex vivo* in the presence of anti-CD3 for 3 days to assess levels of cytokine secretion. Significantly, splenocytes from anti-B7-H4-treated mice secreted significantly higher levels of IFN-γ in response to anti-CD3 stimulation than did T cells from control IgG or anti-PD-1 treated mice (Figure 5G). Taken together, these findings indicate that anti-B7-H4 treatment of BBN mice induced an increase in $CD8^+$ T cell infiltration into tumors within the bladder and increased the level of IFN-γ secreted per splenic $CD8^+$ T cell.

We evaluated the gene expression profile of anti-B7H4 treated mice and found a re-establishment of normal gene expression associated with urothelium differentiation. These included increased expression of bladder muscle/detrusor-related pathways and urothelial differentiation (uroplakin 3b) and decreased expression of cell-cycle progression, mitosis, and cell cycle checkpoints (Supplemental Figure 3). Most prominent, response to B7-H4 was associated with decreased expression of oncogenes Hoxb9, H19 and Egln3. We also observed metabolic changes suggestive of aerobic pathways including Car2 (carbonic anhydrase), adh1 (alcohol dehydrogenase), and Pck1. Lastly, we showed that anti-B7H4 antibody therapy synergized with anti-PD1 therapy, the current standard of care for metastatic, and chemotherapy-resistant cancer, leading to a significant reduction in advanced tumors (40% increase in stage I tumors: 60% in combination treated mice compared to 20% in vehicle treated mice (Figure 5H) ($p=0.017$).

Discussion

Bladder cancer, similar to other solid tumors attributed to carcinogen exposure including lung cancer and melanoma, is driven by a high total mutation burden at approximately 7 mutations/MB (41). Systemic immunotherapy has changed the survival of patients with bladder cancer for those unresponsive to cisplatin (CIS) chemotherapy, or unable to received chemotherapy with a median progression free survival of 10.3 months (95% confidence interval [CI], 8.0 to 11.8) compared with 7.4 months (95% CI, 6.1 to 8.3) in those treated with chemotherapy (hazard ratio for death, 0.73; 95% CI, 0.59 to 0.91; P=0.002) (23). Recent application of anti-PD1 and anti-PDL1 therapy to the neoadjuvant setting have shown promising results to decrease pathologic stage and potentially improve recurrence free survival (42). Yet, less than 20% of patients treated with checkpoint immunotherapy have a clinical response and even in those with MIBC, choosing the right systemic therapy prior to cystectomy remains critical to limit toxicity prior to resection. Until the early reports of Phase III trials of Atezolizumab and Pembrolizumab in the first-line metastatic setting, the value to PD-L1 testing was unclear. Yet, inferior reports of response to PD-L1 low or non-expressing tumors in May of 2018 resulted in label changes for both Atezolizumab and Pembrolizumab confirming the critical association of PD-L1 immunohistochemical marker activity with response to each therapy. Thus, patients that are PD-L1 “negative” have a low likelihood of response and are unlikely to benefit from systemic therapy targeting PD-1/PD-L1. Due to the limitations of the current landscape of PD-1/PDL-1 therapy for bladder cancer, investigation of other immune checkpoints is a necessity for those patients unlikely to respond to PD-1/PD-L1. In our evaluation of MIBC, we found that B7-H4 is expressed in higher levels in luminal and luminal papillary tumors. This distinction separates B7-H4 from CTLA-4 and PD-L1 which are expressed at higher levels in basal cancers. While tumor subtyping has been performed in small series of patients treated with anti-PD-1/PD-L1, luminal and luminal papillary tumors are the poorest responder (22,43)

To identify potential immune regulatory mechanisms involved in the immune-editing of bladder cancer, we profiled the immune landscape of the mouse BBN bladder cancer model from 2 to 20 weeks. Our group has previously validated the genomic alterations that develop in this model (37). To our knowledge, we are the first to describe the immune activation that is bi-modal after BBN treatment. The present data show that there is an acute inflammatory phase that starts within two weeks, but then decreases between four and twelve weeks. While we observed invasion of the bladder by lymphocytes between two and four weeks post initiation of BBN treatment, expression of regulatory monocyte and T cell populations potentially decrease the effector $CD4^+$ and $CD8^+$ T cells until the tumor develops at three months. With the development of CIS and invasive bladder cancer, the number tumor infiltrating lymphocytes increases, but these cells express markers of exhaustion (PD-L1) indicating they are likely not effective in controlling tumor cell growth. Upregulation of B7-H4 expression was observed in monocytes in the BBN-treated mice, preceding both the decrease in $CD8^+$ cells and increase in Tregs. This led to further functional studies of B7-H4 in the BBN bladder cancer model.

Across solid tumors, expression of B7-H4 is associated with worse survival (6-8). High expression of B7-H4 has been described in ovarian, endometrial, gastric, renal cell, melanoma, breast, lung, colorectal, pancreatic, and prostate cancer. Mechanistic investigation of B7-H4 in liver cancer identified B7-H4 expression on tumor cells and antigen presenting cells of the liver that were co-expressed with PD-1 and Tim-3. Genetic targeting of B7-H4 resulted in increased proliferation of $CD8^+$ T cells with decreased levels of exhaustion transcription factors (5). In the current study, we confirmed increased $CD8^+$ TIL activation induced by treatment with anti-B7-H4 antibody as a mechanism of immune activation. Additionally, we found lower numbers of $FoxP3^+$ Tregs that can decrease the anti-tumor immune response.

In addition to effects on Tregs, receptor ligation by the immune modulatory molecules, such as

B7-H4, have been suggested to directly alter the phenotype of $CD4^+$ T cells (44,45). Co-culture of $CD4^+$ T cells with IL-10/TGF-β treated macrophages expressing B7-H4 decreased $CD4^+$ T cell proliferation and increased the number of $FoxP3^+$ $CD4^+$ T cells (46). Published data show that hB7-H4Ig binds to $CD4^+$ T cells in an activation-dependent manner, indicating that B7-H4 may function as a co-inhibitory (negative co-stimulatory) molecule for $CD4^+$ T cells (4,44,47). The present data show that B7-H4 also directly interacts with a functional B7-H4 receptor on CD8+ T cells, as the addition of anti-B7-H4 to monocyte/$CD8^+$ T cell co-cultures induced a significant increase in the level of secreted IFN-γ (Figure 4E). This is supported by our previous findings showing that hB7-H4Ig can down-regulate IL-17 and IFN-γ production of mouse T cells in the absence of Tregs (4). Further, our *in vivo* data show that treatment of C57BL/6 mice with BBN-induced bladder cancer have a significant increase in the number of $CD8^+$ T cells and a significant decrease in the number of Tregs present within the bladder (Figure 5C and D). This finding correlated with an increase in the percentage of IFN-γ^+/$CD8^+$ T cells present within the spleen and a significant decrease in tumor stage as compare to Control Ab treated mice. While the present data have shown the functionality of anti-B7-H4 treatment in the presence of monocyte expressed B7-H4, B7-H4 expression by tumor cells is also a putative mechanism by which these cells evade anti-tumor immune responses. In the majority of breast and ovarian cancers, B7-H4 mRNA is expressed at approximately 2-fold or greater than the level expressed within normal tissue (48), and B7-H4 protein is present in half of early stage and two-thirds of late stage ovarian tumors (49). Additionally, tumor tissues from breast, uterus, ovary, colon, and pancreas showed a statistically significant increase in the percentage of cells expressing B7-H4 (50). In the 4T1 metastatic breast cancer model, transfer of tumor cells into B7-$H4^{-/-}$ vs. wildtype mice resulted in fewer lung nodules, enhanced survival, and decreased tumor infiltration of immunosuppressive cells (51). Based on these findings expression of B7-H4 within the tumor microenvironment, either by the tumor cells and/or by infiltrating monocytes, is hypothesized to promote immune

evasion. In support of this hypothesis, we showed that stimulation of the B7-H4 receptor complex via hB7-H4Ig treatment of mice during EAE increased the number and function of Tregs (4).

Our investigation identified B7-H4 as a negative regulator of T cell activity in the bladder during cancer development. The present data show that treatment of C57BL/6 male mice receiving BBN containing water with anti-B7H4 increases the number of CD8+ T cells and decreases the number of Tregs present within the bladder. The anti-B7-H4 treatment-induced skewing of the $CD8^+$ T cell to Treg ratio within the bladder correlated with a decrease in the bladder cancer stage score. Additionally, co-treatment of mice with both anti-B7-H4 and anti-PD1 further decreased bladder cancer stage score, and such combination therapy may be considered a target for future therapeutic trials in patients with bladder cancer.

Materials and Methods

Tumor Preparation for Single-Cell RNA-Seq

Tumor specimen was minced and enzymatically dissociated DMEM supplemented with Liberase TM (0.0625 mg/ml) and DNase I (Sigma, D5025, 0.2 mg/mL) for 30 minutes. Every 10 minutes specimen was gently pipetted and enzyme mix was exchanged for freshly made enzyme mix. After dissociation tissue was spun down at 1300 RPM for 7 minutes and filtered to through a 100 um filter to yield a single cell suspension. Cells were spun down, resuspended in PBS supplemented with 0.5% BSA and 2 mmol/L EDTA and stained with PI (BD) and Calcein Violet (Invitrogen). Viable cells were sorted using BD FACS Aria Fusion instrument. Sorted cells were washed and resuspended in PBS containing 0.04% BSA. Cells were counted on Countess II automated cell counter (Thermo Fisher) 12,000 cells were loaded per lane onto a 10X Chromium microfluidic chip. Single-cell capture, barcoding, and library preparation were performed using the 10X Chromium version 2 chemistry according to the manufacturer's protocol (#CG00052). cDNA and libraries were checked for quality on Agilent 4200 Tapestation and quantified by KAPA qPCR before sequencing on a single lane of a HiSeq4000 (Illumina) to an average depth of 50,000 reads per cell.

Single-Cell Data Processing

The Cell Ranger pipeline (v1.2, 10X Genomics) was used to convert Illumina base call files to FASTQ files, align FASTQs to the GRCH38 reference (v3.0.0, 10X Genomics) for human samples to produce a digital gene-cell counts matrix. The resultant gene-cell matrix was filtered to remove cells with fewer than 500 transcripts and genes with fewer than two counts in two cells. The gene-cell matrices were then normalized such that the number of unique molecular identifiers (UMI) in each cell is equal to the median UMI count across the data set and log transformed. Expression at 1,000 highly variable genes in each data set, selected as the genes with the highest dispersion, was used to reduce the dimensionality of the data sets to three dimensions using Uniform Manifold Approximation and Projection (UMAP) and cells were clustered using Leiden based clustering in

the UMAP space. Genes of interest were plotted in UMAP space using adjusted values based on Markov Affinity-based Graph Imputation (MAGIC) of the raw gene-cell counts matrix.

Multiplexed Imaging by IMC

Formalin-fixed paraffin-embedded human Ta NMIBC tissues were cut into 5-µm sections and mounted on slides. Slides were incubated for 15 minutes at 55°C in a dry oven, deparaffinized in fresh histoclear, and rehydrated through a series of graded alcohols. Antigen retrieval was performed in a decloaking chamber (BioSB TintoRetriever) for 15 minutes at 95°C in Citrate Buffer. After blocking in buffer containing 3% BSA, slides were incubated overnight at 4°C with a cocktail of metal-conjugated IMC-validated primary antibodies. The following day, slides were washed twice in Dulbecco's Phosphate-Buffered Saline and counterstained with iridium intercalator (0.25 µmol/L) for 5 minutes at room temperature, to visualize the DNA. After a final wash in ddH20, the slides were air-dried for 20 minutes. The slides were then loaded on the Fluidigm Hyperion imaging mass cytometer. Regions of interest were selected using the acquisition software and ablated by the Hyperion. The resulting images were exported as 16-bit .tiff files using the Fluidigm MCDViewer software and analyzed using the open source Histocat++ toolbox

Mice and tumor staging: Mice were cared for in a fashion supervised by the ACUC at Northwestern University and all research was approved in an animal research protocol. Male C57BL/6 mice at least 6 weeks old received N-butyl-N-(4-hydroxybutyl)-nitrosamine (BBN) at a dose of 0.05% in drinking water *ad libitum*. After 20 weeks of exposure, mice were randomized to therapy which included treatment with IgG (200 µg/kg, BioXcell), anti-PD1 (200 µg/kg, BioXcell), and/or anti-B7-H4 (200 µg/mouse, from Amplimmune). Mice were injected with four weekly doses. At the end of treatment, animals were euthanized by CO_2 with secondary cervical dislocation.

Bladders were removed and divided in the sagittal plane, and either fixed in 10% formalin or snap-frozen in liquid nitrogen. Pathology was staged using human AJCC staging of degree of bladder wall invasion.

RNA extraction, Library Creation and RNA-Seq

Mouse tumors were ground using liquid nitrogen and a mortar and pestle, and then RNA was extracted using Trizol Reagent (Thermo Fisher Scientific) following manufacturer's instructions. RNA quality and concentration were assessed by NanoDrop and then Qubit Fluorometric Quantitation (Thermo Fisher Scientific). Library preparation and RNAseq were performed at the NuSeq core facility (Northwestern University) using an Illumina HiSeq 2000.

Immunohistochemistry

Bladders were fixed in 10% formalin and embedded in paraffin. 4 micron thick sections were used for IHC staining for CD8 (1:1000, #MA5-13263, Invitrogen) and FoxP3 (1:400, #12653, Cell Signaling). IHC was performed using a Dako Autostainer Plus instrument (Dako, CO, USA), and anti-rabbit Dako EnVision+System-HRP (Dako). For each tumor, the HRP positive cells were counted per low-power (10X objective) field. Each tumor was evaluated with at least six fields in two separate sections.

Human CD14$^+$ monocyte plus T cell co-cultures. PMBCs from healthy donors (n=5) (LifeSource; Evanston, IL) were collected and CD14$^+$ monocytes were purified via AutoMacs Magnetic Bead cell separation technology. The CD14- fractions were collected, and CD4+ and CD8$^+$ T cells purified via AutoMacs Magnetic Bead cell separation technology. The autologous CD4$^+$ and CD8+ T cells were cryopreserved for use in the monocyte/T cell co-cultures. The CD14$^+$ monocytes were cultured in the presence of medium alone, recombinant human IL-6, recombinant human IL-10, or recombinant human IL-6 plus recombinant human IL-10 (20ng/ml)

at $5x10^6$ cells/well in 6-well plates at a final volume of 2ml in cRPMI w/ 10% FCS for 3 days. On day 3 of culture, the non-adherent cells were removed, and the adherent monocytes collected following incubation in presence of 1ml of 1xDPBS containing 2mM EDTA, and incubate the plates at 37°C for 15min. The cryopreserved $CD4^+$ and $CD8^+$ T cells were thawed, washed, and stained with CFSE. For the macrophage/T cell co-cultures, the cells were cultured in flat-bottomed 96-well plates. The wells were coated with anti-CD3 (0.5µg/ml; 100µl/well; coating wells for 2 hours at 37°C; wells washed 3x with 1x DPBS), and the appropriate amount of Control Ab or anti-B7-H4 was added. For the assessment of T cells proliferation, the T cells were collected, washed in PBS, stained with LIVE/DEAD® Fixable Aqua Dead Cell Stain (Life Technologies; Grand Island, NY), blocked with anti-CD16/32 (ThermoFisher Scientific; San Jose, CA), and then stained with the indicated antibodies - anti-CD4 (clone OKT4) or anti-CD8 (clone 3B5). Results are expressed as the mean percentage of proliferated $CD4^+$ and $CD8^+$ T cells.. For cytokine analysis replicate wells were harvested on day +3 of co-culture and cytokine secretion determined via multiplex Luminex LiquiChip (Millipore).

Mouse lymph node cultures. Lymph nodes were collected from naïve SJL/J mice, and cells made into a single cell suspensions. CFSE-labeled total lymph node cells ($5x10^5$ cells/well) were cultured in the presence of anti-CD3 (1µg/ml), plus Control Ig, hB7-H4Ig, anti-B7-H4, or hB7-H4Ig plus anti-B7-H4 (5µg/ml). The cells were cultured for 5 days. For the assessment of T cells proliferation, the T cells were collected, washed in PBS, stained with LIVE/DEAD® Fixable Aqua Dead Cell Stain (Life Technologies; Grand Island, NY), blocked with anti-CD16/32 (ThermoFisher Scientific; San Jose, CA), and then stained with the indicated antibodies - anti-CD4 (clone RM4-5). Replicate wells were pulsed with 1µCi of tritiated thymidine at 24 h, and the cultures were harvested on day +5.

$PLP_{139-151}$/CFA Induced EAE and DTH. 6-7 week-old female SJL/J mice were immunized s.c. with 100µl of an emulsion containing 200µg of *M. tuberculosis* H37Ra (BD Biosciences; San Jose, CA) and 50µg of $PLP_{139–151}$ distributed over three sites on the flank. Mice were treated with Control Ig, hB7-H4Ig, and anti-B7-H4 beginning at the time of priming. Mice were treated with 100µg per dose injections 3x per week for 2 weeks via i.p. injection. Individual animals were observed at the indicated time points and clinical scores assessed in a blinded fashion on a 0–5 scale: 0, no abnormality; 1, limp tail; 2, limp tail and hind limb weakness; 3, hind limb paralysis; 4, hind limb paralysis and forelimb weakness; and 5, moribund. The data are reported as the mean daily clinical score. On day +9 post priming, mice were assayed for delayed type hypersensitivity (DTH). Mice were anaesthetized by inhalation of isoflurane and the thickness of both ears was measured using a dial thickness gauge. 10µg of $PLP_{139-151}$ (negative control) and OVA protein in 10µl of PBS was injected into the left and right ear, respectively. The increase in ear thickness was determined after 24 h.

Flow Cytometry. For cell analysis spleens were dissociated into a single cell suspensions and RBCs lysed. For the bladder leukocytes, Single cell suspensions were prepared by mincing the bladder tissue in 2ml of Accutase (MilliPore) plus 1mg/ml collagenase, and the samples were incubated at 37°C for 30 min. Following the enzyme digestion, the bladder samples were disrupted with a 100µm cell strainer, the cell strainer washed 2x with 10ml of HBSS+ 5% FCS, and the cells pelleted. The cells were washed in PBS, stained with LIVE/DEAD® Fixable Aqua Dead Cell Stain (Life Technologies; Grand Island, NY), blocked with anti-CD16/32 (ThermoFisher Scientific), and then stained with the indicated antibodies. 10^6 viable cells were analyzed per individual sample using a BD LSRFortessa (BD Bioscience), and the data analyzed using FloJo Version 9.5.2 software (Tree Star, Inc.; Ashland, OR).

CD4+ T cells	Color	CD8+ T cells	Color	CD4/CD8	Color	Monocytes	Color
VID	Live/Dead Blue Indo-violet	VID	Live/Dead Blue Indo-violet	VID	Live/Dead Blue Indo-violet	VID	Live/Dead Blue Indo-violet
CD45 (clone 30-F11)	V500	CD45 (clone 30-F11)	V500	CD45 (clone 30-F11)	V500	CD45 (clone 30-F11)	V500
CD3 (clone 145-2C11)	BV650	CD3 (clone 145-2C11)	BV650	CD3 (clone 145-2C11)	BV650	CD11b (clone M1/70)	BV650
CD4 (clone RM4-5)	PB	CD8 (clone 53-6.7)	PB	CD4 (clone RM4-5)	PB	Ly6C (clone AL-21)	Alexa Fluor 700
CD25 (clone PC61)	APC/Cy7	CD25 (clone PC61)	APC/Cy7	CD8 (clone 53-6.7)	APC/Cy7	Ly6G (clone 1A8)	APC/Cy7
CD44 (clone IM7)	BV605	CD44 (clone IM7)	BV605	Nrp-1 (polyclonal)	APC	CD80 (clone 16-10A1)	PB
FoxP3 (clone FJK-16s)	Alexa Fluor 700	PD-1 (clone RMP1-30)	PE-Cy7	Sema3a (mouse and human clone #215)	FITC	CD86 (clone GL1)	APC
PD-1 (clone RMP1-30)	PE-Cy7					PD-L1 (clone 10F.9G2)	PE/Cy7
						B7-H4 (clone MIH29)	PE

Bioinformatics

Clinical, mRNA seq, and mutation data from The Cancer Genome Atlas (TCGA) bladder urothelial carcinoma dataset was downloaded from the Broad Institute Genome Data Analysis Center (GDAC) using TCGA data version 2016_01_28 (http://gdac.broadinstitute.org) (26,27). Analysis

was performed with R v3.5.1 utilizing Bioconductor v3.9. RNA-seq mRNA expression levels are presented as RNA-seq by expectation-maximization (RSEM) values, and z-score >2 was used to define overexpression (28). Overall survival outcomes were calculated using the log-rank test and R packages survival v2.43-3 and survminer v0.4.3. Pan-cancer expression profile of B7-H4 (VTCN1) was extracted from the Broad GDAC Firehose v1.1.38 (http://firebrowse.org/). Spearman correlations between B7-H4, PD-1, PD-L1, and CTLA-4 expression and mutation, neoantigen, and APOBEC enrichment was performed using ggplot2 v3.1.0. Molecular subtypes (luminal, luminal-papillary, luminal-infiltrated, basal-squamous, and neuronal) were assigned as previously described (29). Comparison of expression between subtypes was performed using Kruskal-Wallis ANOVA. Immune signatures were calculated as previously described (30-32). Hierarchical clustering of immune signatures in among molecular subtypes was performed with multiClust v1.4.0 and gplots v3.0.1 (33). Correlation heatmap of immune signatures and B7-H4 expression was created using ggplot2 v3.1.0.

For murine tumors, following quality control and adapter trimming, reads were mapped to mm10 genome using STAR (34), and then counted using HTSeq (35). Gene counts were converted to CPM and further analyzed in R using the Bioconductor packages limma and edgeR. Limma was used to generate MDS plots and call differentially expressed (DE) genes. Top DE genes were visualized by Volcano plots, and used for Pathway Analysis that was conducted using the Bioconductor packages TopGO and ReactomePA (36). The most significant pathways (reactomeDB terms) enriched in DE genes were shown by horizontal barplots, where color intensity tracked with p-values (computed by Fisher Exact test).

Statistical Analysis Comparisons of the percentage of animals showing clinical disease were analyzed by X^2 using Fisher's exact probability, and two-way ANOVA with a Bonferroni post-test was used to determine statistical differences between mean clinical disease scores. Single comparisons of two means were analyzed by Student's t-test.

Disclosure of Potential Conflicts of Interest: None

Author Contribution

JRP- designing research studies, conducting experiments, acquiring data, analyzing data, and writing the manuscript, editing manuscript

APG- designing research studies, conducting experiments, acquiring data, analyzing data, and writing the manuscript, editing manuscript

DB designing research studies, conducting experiments, acquiring data, analyzing data, and writing the manuscript, editing manuscript

DF- designing research studies, conducting experiments, acquiring data, analyzing data, and writing the manuscript, editing manuscript

YY designing research studies, conducting experiments, acquiring data, analyzing data, editing manuscript

VE- designing research studies, conducting experiments, acquiring data, analyzing data, editing the manuscript.

SS designing research studies, conducting experiments, acquiring data, analyzing data, providing reagents, and writing the manuscript.

PR- designing research studies, conducting experiments, acquiring data, analyzing data, writing and editing the manuscript.

SDM- designing research studies, conducting experiments, acquiring data, analyzing data, writing and editing the manuscript.

JJM- designing research studies, conducting experiments, acquiring data, analyzing data, writing and editing the manuscript.

Acknowledgements

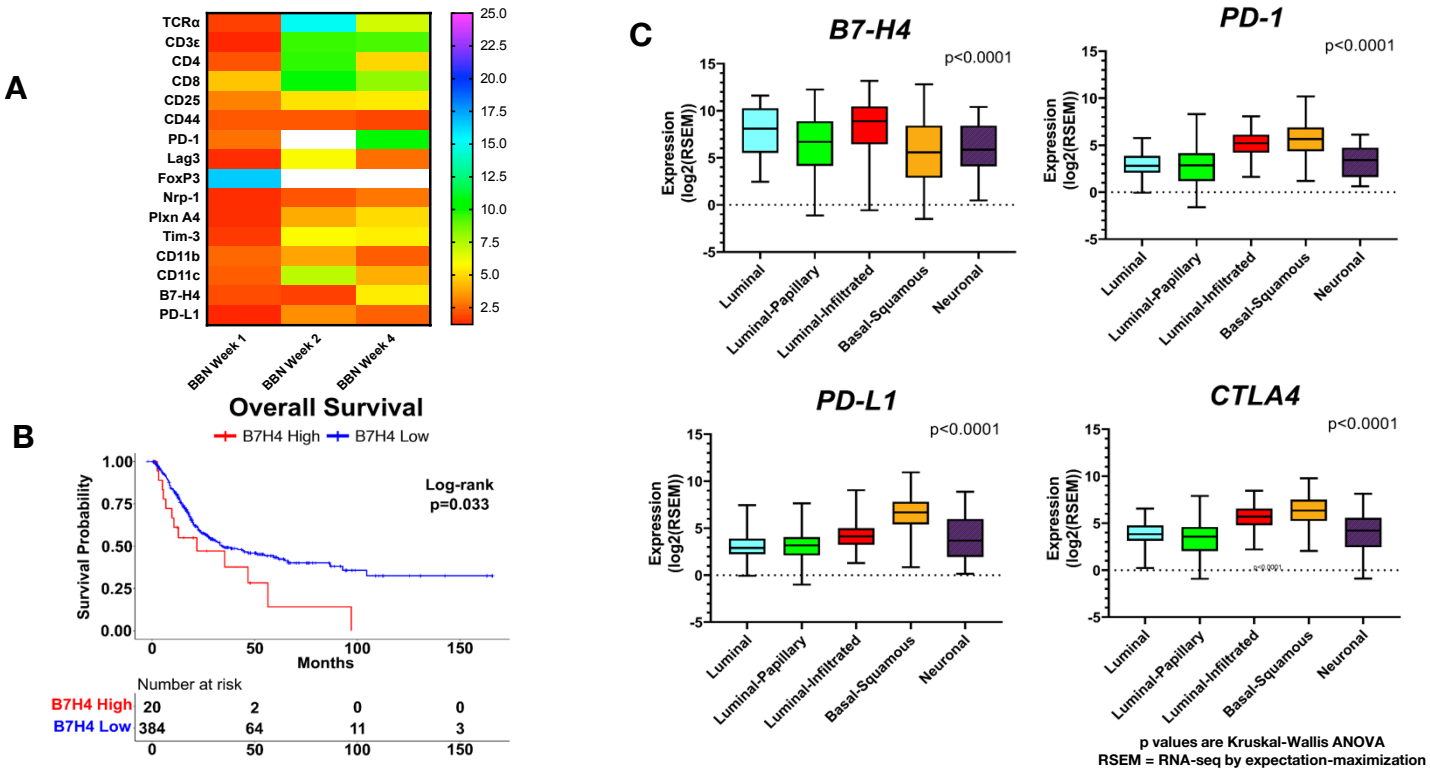
JJM and SDM were supported by a Translational Bridge Grant from the Robert H. Lurie Cancer Center Funded by the John P. Hanson Foundation.

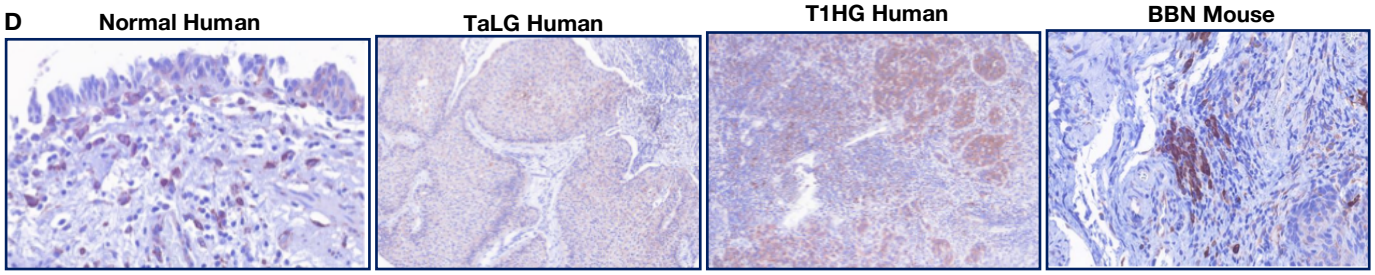
References

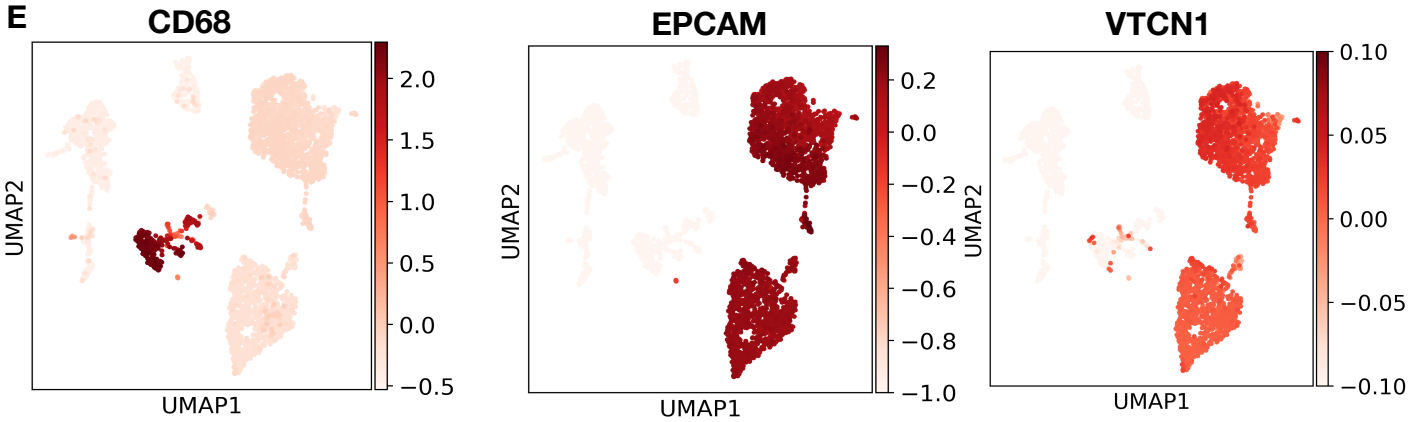
1. Podojil JR, Miller SD. Potential targeting of B7-H4 for the treatment of cancer. Immunological reviews 2017;276(1):40-51.
2. Zang X, Loke P, Kim J, Murphy K, Waitz R, Allison JP. B7x: a widely expressed B7 family member that inhibits T cell activation. Proceedings of the National Academy of Sciences of the United States of America 2003;100(18):10388-92.
3. Ichikawa M, Chen L. Role of B7-H1 and B7-H4 molecules in down-regulating effector phase of T-cell immunity: novel cancer escaping mechanisms. Front Biosci 2005;10:2856-60.
4. Podojil JR, Liu LN, Marshall SA, Chiang MY, Goings GE, Chen L*, et al.* B7-H4Ig inhibits mouse and human T-cell function and treats EAE via IL-10/Treg-dependent mechanisms. Journal of autoimmunity 2013;44:71-81.
5. Li J, Lee Y, Li Y, Jiang Y, Lu H, Zang W*, et al.* Co-inhibitory molecule B7 superfamily member 1 expressed by tumor-infiltrating myeloid cells induces dysfunction of anti-tumor CD8(+) T cells. Immunity 2018;48(4):773-86 e5.
6. Tringler B, Liu W, Corral L, Torkko KC, Enomoto T, Davidson S*, et al.* B7-H4 overexpression in ovarian tumors. Gynecologic oncology 2006;100(1):44-52.
7. Zhang LL, Shao SL, Wu Y. Expressions of osteopontin and B7-H4 in epithelial ovarian neoplasm and their significance. Chinese journal of cancer 2010;29(1):25-9.
8. Kryczek I, Wei S, Zhu G, Myers L, Mottram P, Cheng P*, et al.* Relationship between B7-H4, regulatory T cells, and patient outcome in human ovarian carcinoma. Cancer research 2007;67(18):8900-5.
9. Cheung SS, Ou D, Metzger DL, Meloche M, Ao Z, Ng SS*, et al.* B7-H4 expression in normal and diseased human islet beta cells. Pancreas 2014;43(1):128-34.
10. Xu Y, Zhu S, Song M, Liu W, Liu C, Li Y*, et al.* B7-H4 expression and its role in interleukin-2/interferon treatment of clear cell renal cell carcinoma. Oncology letters 2014;7(5):1474-8.
11. Krambeck AE, Thompson RH, Dong H, Lohse CM, Park ES, Kuntz SM*, et al.* B7-H4 expression in renal cell carcinoma and tumor vasculature: associations with cancer progression and survival. Proceedings of the National Academy of Sciences of the United States of America 2006;103(27):10391-6.
12. Zhang L, Wu H, Lu D, Li G, Sun C, Song H*, et al.* The costimulatory molecule B7-H4 promote tumor progression and cell proliferation through translocating into nucleus. Oncogene 2013;32(46):5347-58.
13. Quandt D, Fiedler E, Boettcher D, Marsch W, Seliger B. B7-h4 expression in human melanoma: its association with patients' survival and antitumor immune response. Clin Cancer Res 2011;17(10):3100-11.
14. Mugler KC, Singh M, Tringler B, Torkko KC, Liu W, Papkoff J*, et al.* B7-h4 expression in a range of breast pathology: correlation with tumor T-cell infiltration. Appl Immunohistochem Mol Morphol 2007;15(4):363-70.
15. Li ZY, Zhang XH, Chen Y, Guo JG, Sai K, Yang QY*, et al.* Clinical significance of B7-H4 expression in matched non-small cell lung cancer brain metastases and primary tumors. OncoTargets and therapy 2013;6:869-75.
16. Sun Y, Wang Y, Zhao J, Gu M, Giscombe R, Lefvert AK*, et al.* B7-H3 and B7-H4 expression in non-small-cell lung cancer. Lung cancer (Amsterdam, Netherlands) 2006;53(2):143-51.
17. Arigami T, Uenosono Y, Ishigami S, Hagihara T, Haraguchi N, Natsugoe S. Clinical significance of the B7-H4 coregulatory molecule as a novel prognostic marker in gastric cancer. World journal of surgery 2011;35(9):2051-7.

18. Zhao LW, Li C, Zhang RL, Xue HG, Zhang FX, Zhang F*, et al.* B7-H1 and B7-H4 expression in colorectal carcinoma: correlation with tumor FOXP3(+) regulatory T-cell infiltration. Acta histochemica 2014;116(7):1163-8.
19. Chen Y, Sun J, Zhao H, Zhu D, Zhi Q, Song S*, et al.* The coexpression and clinical significance of costimulatory molecules B7-H1, B7-H3, and B7-H4 in human pancreatic cancer. OncoTargets and therapy 2014;7:1465-72.
20. Zang X, Thompson RH, Al-Ahmadie HA, Serio AM, Reuter VE, Eastham JA*, et al.* B7-H3 and B7x are highly expressed in human prostate cancer and associated with disease spread and poor outcome. Proceedings of the National Academy of Sciences of the United States of America 2007;104(49):19458-63.
21. Zhang Y, Zhu C, Curado MP, Zheng T, Boyle P. Changing patterns of bladder cancer in the USA: evidence of heterogeneous disease. BJU international 2012;109(1):52-6.
22. Rosenberg JE, Hoffman-Censits J, Powles T, van der Heijden MS, Balar AV, Necchi A*, et al.* Atezolizumab in patients with locally advanced and metastatic urothelial carcinoma who have progressed following treatment with platinum-based chemotherapy: a single-arm, multicentre, phase 2 trial. Lancet (London, England) 2016;387(10031):1909-20.
23. Bellmunt J, de Wit R, Vaughn DJ, Fradet Y, Lee JL, Fong L*, et al.* Pembrolizumab as second-line therapy for advanced urothelial carcinoma. The New England journal of medicine 2017;376(11):1015-26.
24. Cristescu R, Mogg R, Ayers M, Albright A, Murphy E, Yearley J*, et al.* Pan-tumor genomic biomarkers for PD-1 checkpoint blockade-based immunotherapy. Science (New York, NY 2018;362(6411).
25. Fares CM, Van Allen EM, Drake CG, Allison JP, Hu-Lieskovan S. Mechanisms of resistance to immune checkpoint blockade: Why does dheckpoint inhibitor immunotherapy not work for all patients? American Society of Clinical Oncology educational book American Society of Clinical Oncology 2019;39:147-64.
26. Marx V. Drilling into big cancer-genome data. Nature methods 2013;10(4):293-7.
27. Center BITGDA. Analysis overview for bladder urothelial carcinoma (primary solid tumor cohort) - 28. Broad Institute of MIT and Harvard 2016 doi 10.7908/C19G5M58.
28. Li B, Dewey CN. RSEM: accurate transcript quantification from RNA-Seq data with or without a reference genome. BMC bioinformatics 2011;12:323.
29. Robertson AG, Kim J, Al-Ahmadie H, Bellmunt J, Guo G, Cherniack AD*, et al.* Comprehensive molecular characterization of muscle-invasive bladder cancer. Cell 2018;174(4):1033.
30. Bindea G, Mlecnik B, Tosolini M, Kirilovsky A, Waldner M, Obenauf AC*, et al.* Spatiotemporal dynamics of intratumoral immune cells reveal the immune landscape in human cancer. Immunity 2013;39(4):782-95.
31. Faruki H, Mayhew GM, Serody JS, Hayes DN, Perou CM, Lai-Goldman M. Lung adenocarcinoma and squamous cell carcinoma gene expression subtypes demonstrate significant differences in tumor immune landscape. J Thorac Oncol 2017;12(6):943-53.
32. Glaser AP, Fantini D, Wang Y, Yu Y, Rimar KJ, Podojil JR*, et al.* APOBEC-mediated mutagenesis in urothelial carcinoma is associated with improved survival, mutations in DNA damage response genes, and immune response. Oncotarget 2018;9(4):4537-48.
33. Lawlor N, Fabbri A, Guan P, George J, Karuturi RK. multiClust: An R-package for identifying biologically relevant clusters in cancer transcriptome profiles. Cancer informatics 2016;15:103-14.
34. Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S*, et al.* STAR: ultrafast universal RNA-seq aligner. Bioinformatics (Oxford, England) 2013;29(1):15-21.
35. Anders S, Pyl PT, Huber W. HTSeq--a Python framework to work with high-throughput sequencing data. Bioinformatics (Oxford, England) 2015;31(2):166-9.

36. Yu G, He QY. ReactomePA: an R/Bioconductor package for reactome pathway analysis and visualization. Molecular bioSystems 2016;12(2):477-9.
37. Fantini D, Glaser AP, Rimar KJ, Wang Y, Schipma M, Varghese N*, et al.* A Carcinogen-induced mouse model recapitulates the molecular alterations of human muscle invasive bladder cancer. Oncogene 2018;37(14):1911-25.
38. Fantini D, Seiler R, Meeks JJ. Molecular footprints of muscle-invasive bladder cancer in smoking and nonsmoking patients. Urologic oncology 2018.
39. Podojil JR, Chiang MY, Ifergan I, Copeland R, Liu LN, Maloveste S*, et al.* B7-H4 modulates regulatory CD4(+) T cell induction and function via ligation of a semaphorin 3a/plexin A4/neuropilin-1 complex. J Immunol 2018;201(3):897-907.
40. Shin K, Lim A, Odegaard JI, Honeycutt JD, Kawano S, Hsieh MH*, et al.* Cellular origin of bladder neoplasia and tissue dynamics of its progression to invasive carcinoma. Nature cell biology 2014;16(5):469-78.
41. Chalmers ZR, Connelly CF, Fabrizio D, Gay L, Ali SM, Ennis R*, et al.* Analysis of 100,000 human cancer genomes reveals the landscape of tumor mutational burden. Genome medicine 2017;9(1):34.
42. Necchi A, Anichini A, Raggi D, Briganti A, Massa S, Luciano R*, et al.* Pembrolizumab as neoadjuvant therapy before radical cystectomy in patients with muscle-invasive urothelial bladder carcinoma (PURE-01): An open-label, single-arm, phase II study. J Clin Oncol 2018:JCO1801148.
43. Kim J, Kwiatkowski D, McConkey DJ, Meeks JJ, Freeman SS, Bellmunt J*, et al.* The cancer genome atlas expression subtypes stratify response to checkpoint inhibition in advanced urothelial cancer and identify a subset of patients with high survival probability. European urology 2019;75(6):961-4.
44. Sica GL, Choi IH, Zhu G, Tamada K, Wang SD, Tamura H*, et al.* B7-H4, a molecule of the B7 family, negatively regulates T cell immunity. Immunity 2003;18(6):849-61.
45. Choi IH, Zhu G, Sica GL, Strome SE, Cheville JC, Lau JS*, et al.* Genomic organization and expression analysis of B7-H4, an immune inhibitory molecule of the B7 family. J Immunol 2003;171(9):4650-4.
46. Cao Q, Wang Y, Zheng D, Sun Y, Wang Y, Lee VW*, et al.* IL-10/TGF-beta-modified macrophages induce regulatory T cells and protect against adriamycin nephrosis. J Am Soc Nephrol 2010;21(6):933-42.
47. Wei J, Loke P, Zang X, Allison JP. Tissue-specific expression of B7x protects from CD4 T cell-mediated autoimmunity. The Journal of experimental medicine 2011;208(8):1683-94.
48. Salceda S, Tang T, Kmet M, Munteanu A, Ghosh M, Macina R*, et al.* The immunomodulatory protein B7-H4 is overexpressed in breast and ovarian cancers and promotes epithelial cell transformation. Experimental cell research 2005;306(1):128-41.
49. Simon I, Katsaros D, Rigault de la Longrais I, Massobrio M, Scorilas A, Kim NW*, et al.* B7-H4 is over-expressed in early-stage ovarian cancer and is independent of CA125 expression. Gynecologic oncology 2007;106(2):334-41.
50. Qian Y, Shen L, Cheng L, Wu Z, Yao H. B7-H4 expression in various tumors determined using a novel developed monoclonal antibody. Clinical and experimental medicine 2011;11(3):163-70.
51. Abadi YM, Jeon H, Ohaegbulam KC, Scandiuzzi L, Ghosh K, Hofmeyer KA*, et al.* Host b7x promotes pulmonary metastasis of breast cancer. J Immunol 2013;190(7):3806-14.


A
TCRα
CD3ε
CD4
CD8
CD25
CD44
PD-1
Lag3
FoxP3
Nrp-1
Plxn A4
Tim-3
CD11b
CD11c
B7-H4
PD-L1
BBN Week 1
BBN Week 2
BBN Week 4
25.0
22.5
20.0
17.5
15.0
12.5
10.0
7.5
5.0
2.5
B
Overall Survival
B7H4 High
B7H4 Low
Log-rank
p=0.033
Survival Probability
Months
Number at risk
B7H4 High 20 2 0 0
B7H4 Low 384 64 11 3
C
B7-H4
PD-1
PD-L1
CTLA4
p<0.0001
Expression (log2(RSEM))
Luminal
Luminal-Papillary
Luminal-Infiltrated
Basal-Squamous
Neuronal
p values are Kruskal-Wallis ANOVA
RSEM = RNA-seq by expectation-maximization


D
Normal Human
TaLG Human
T1HG Human
BBN Mouse


E
CD68
EPCAM
VTCN1
UMAP1
UMAP2

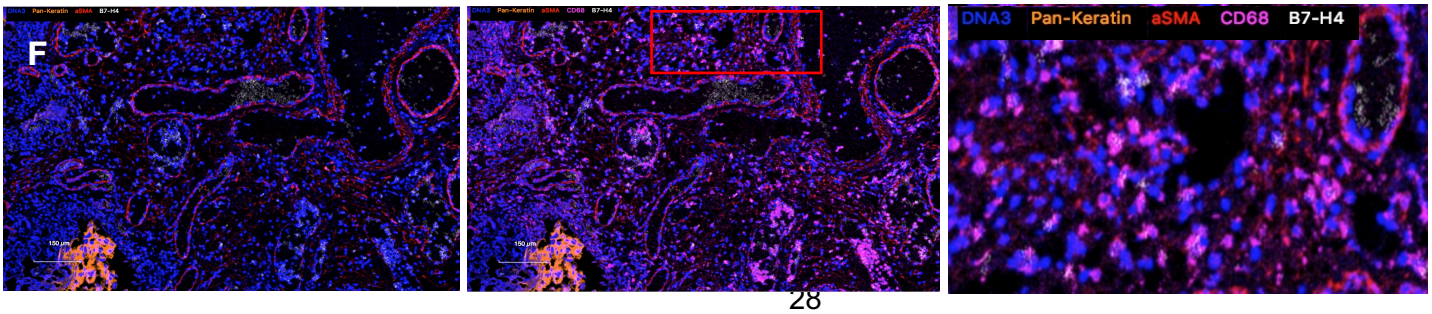

F
DNA3
Pan-Keratin
aSMA
CD68
B7-H4

Figure 1. B7-H4 is expressed by bladder cancer and myeloid cells in human and murine bladder cancer. A) Heat map of mRNAs from immune regulatory genes at 1, 2 and 4 weeks of BBN exposure. B) Overall survival of the muscle invasive bladder cancers (MIBC) from the TCGA identifying worse survival for patients with high levels of VTCN1 (B7-H4). C) In contrast to tumors that express high levels of other immune regulatory proteins, like PD-L1, PD1 and CTLA4, B7-H4 is expressed in luminal and luminal infiltrated tumors, D) Immunohistochemistry of B7-H4 in normal human bladder, low grade non-invasive (stage TaLG), invasive (Stage T1HG) and BBN mouse bladder tumor demonstrating differences in cell localization. These localizations in the bladder were confirmed by single-cell RNA-seq of a low grade Ta tumor (E) showing high expression in luminal cells (+EPCAM) and scattered expression in myeloid cells (CD68). Immune mass cytometry (F) of a human bladder confirms localization of B7-H4 with CD68+ cells.

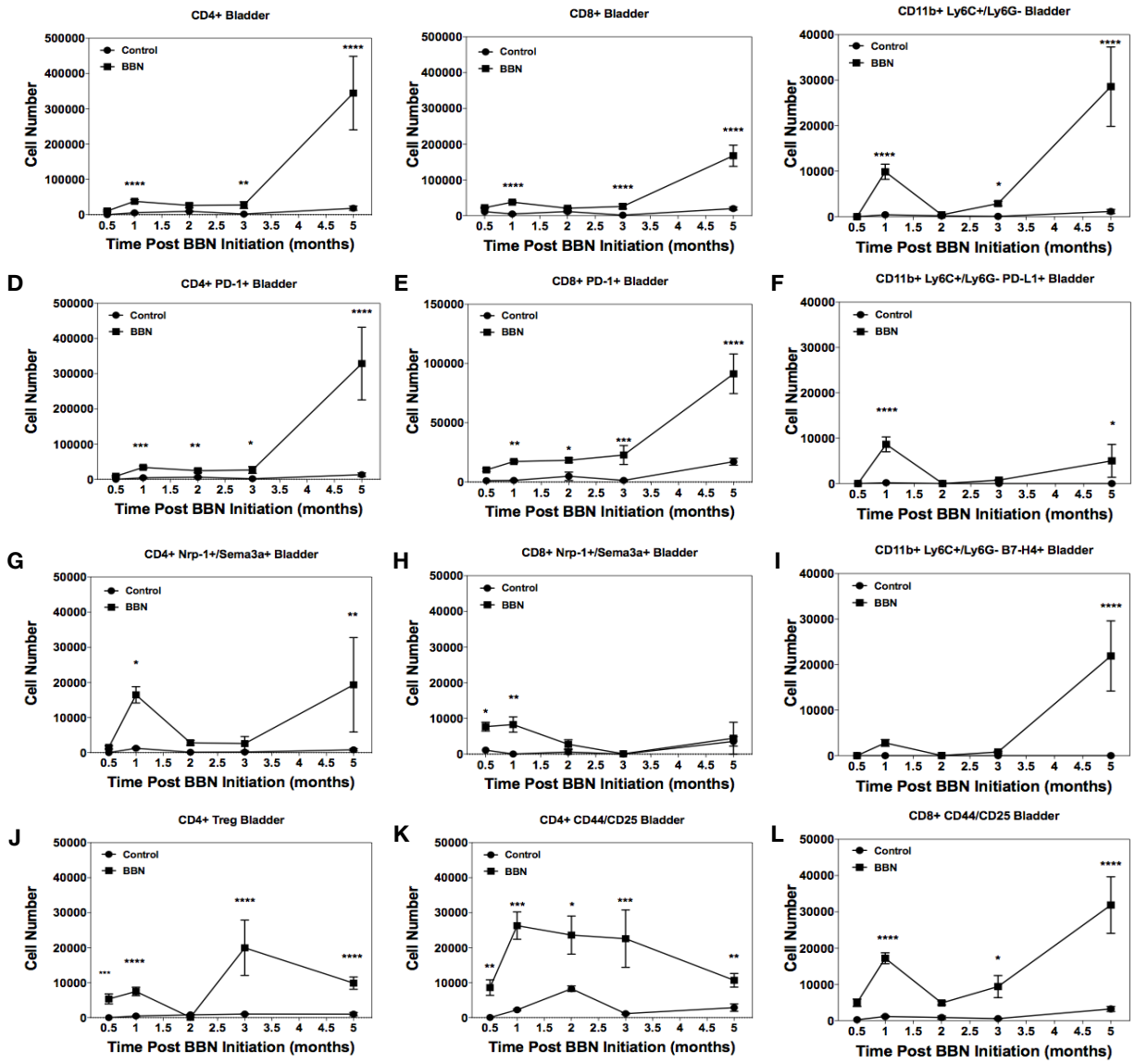


Figure 2. Bi-phasic pattern of immune bladder immune infiltration during BBN-induced cancer. Male C57BL/6 mice (n=10 mice per group for each time point) were supplied either sterile water or sterile water containing 0.05% BBN *ad libitum*. On the indicated time points, 10 representative naïve control and BBN treated mice were taken for analysis. The bladders were harvested and 2 random bladders each pooled to generate a total of 5 analytical samples for flow cytometric analysis. The cells samples were gated as follows; for T cells (singlets, cells, live/CD45^{hi}, CD3^{+}/CD4+ or CD3^{+}/CD8^{+} into flow plots for specified T cells markers), and for monocytes (singlets, cells, live/CD45^{hi}, CD11b^{+}, Ly6C^{+}/Ly6G^{-} into flow plots for specified monocyte markers).

The data are presented as the mean number of each specified cell population per sample ± S.E.M. Asterisks indicate a statistically significant difference in the number of cells present within the bladder of mice receiving BBN as compared to Control naïve mice analyzed at each time point - *$p<0.05$, **$p< 0.01$, ***$p< 0.001$, ****$p<0.0001$, respectively.

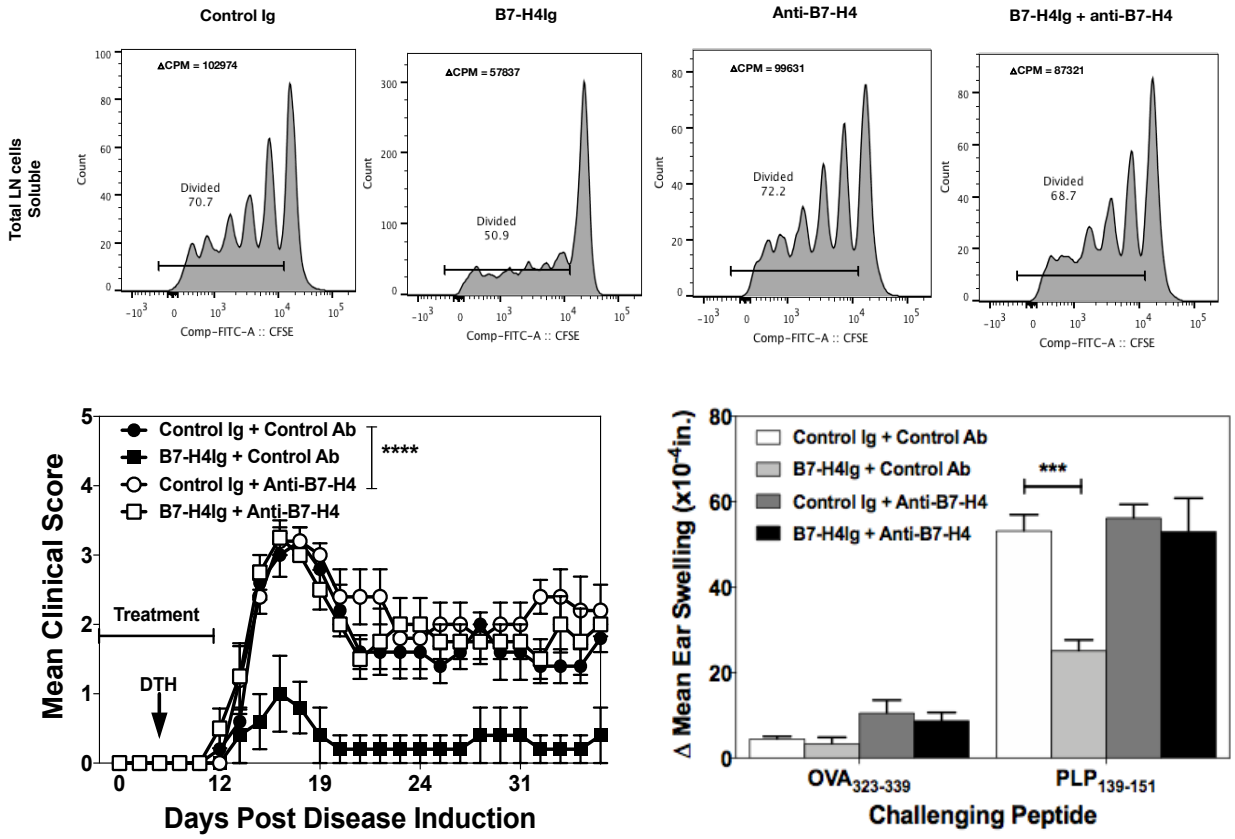


Figure 3. Anti-B7-H4 blocks the immunosuppressive function of B7-H4Ig. Total LN cells were collected from naïve SJL/J mice, and cells were labeled with CFSE. CFSE-labeled lymph node cells ($5x10^5$ cells/well) were cultured in the presence of anti-CD3 (1μg/ml), plus Control Ig, hB7-H4Ig, anti-B7-H4, or hB7-H4Ig plus anti-B7-H4 (5μg/ml). Replicate wells were pulsed with 1μCi of tritiated thymidine at 24 h, and the cultures were harvested on day +5. For the assessment of T cells proliferation, the percentage of live CD4+ T cells that diluted CFSE were gated. The level of cellular proliferation is also presented on each flow histogram as determined by tritiated thymidine incorporation (Δ CPM = CPM with anti-CD3 – CPM with PBS) (A). SJL/J mice (n=10/group) were primed with $PLP_{139-151}$/CFA and treated with species and isotype-matched Control Ig, hB7-H4Ig, anti-B7-H4, or hB7-H4Ig plus anti-B7-H4 (100μg/dose; 3x/wk; 2wks), and mice were followed for disease severity. The data is presented as the mean clinical score ± S.E.M.

(B). On day +9 of the disease course the $PLP_{139-151}$ *in vivo* DTH responses were assessed. The data are presented as the mean ear swelling $\pm$ S.E.M. for both control antigen challenge ($OVA_{323-339}$) and specific antigen challenge ($PLP_{139-151}$) (C). One representative experiment of three is presented. Asterisks (***) indicates a statistically significant decrease in the $PLP_{139-151}$-induced clinical EAE or DTH response in mice B7-H4Ig treated mice in comparison to mice treated with Control Ig, $p < 0.001$ respectively.

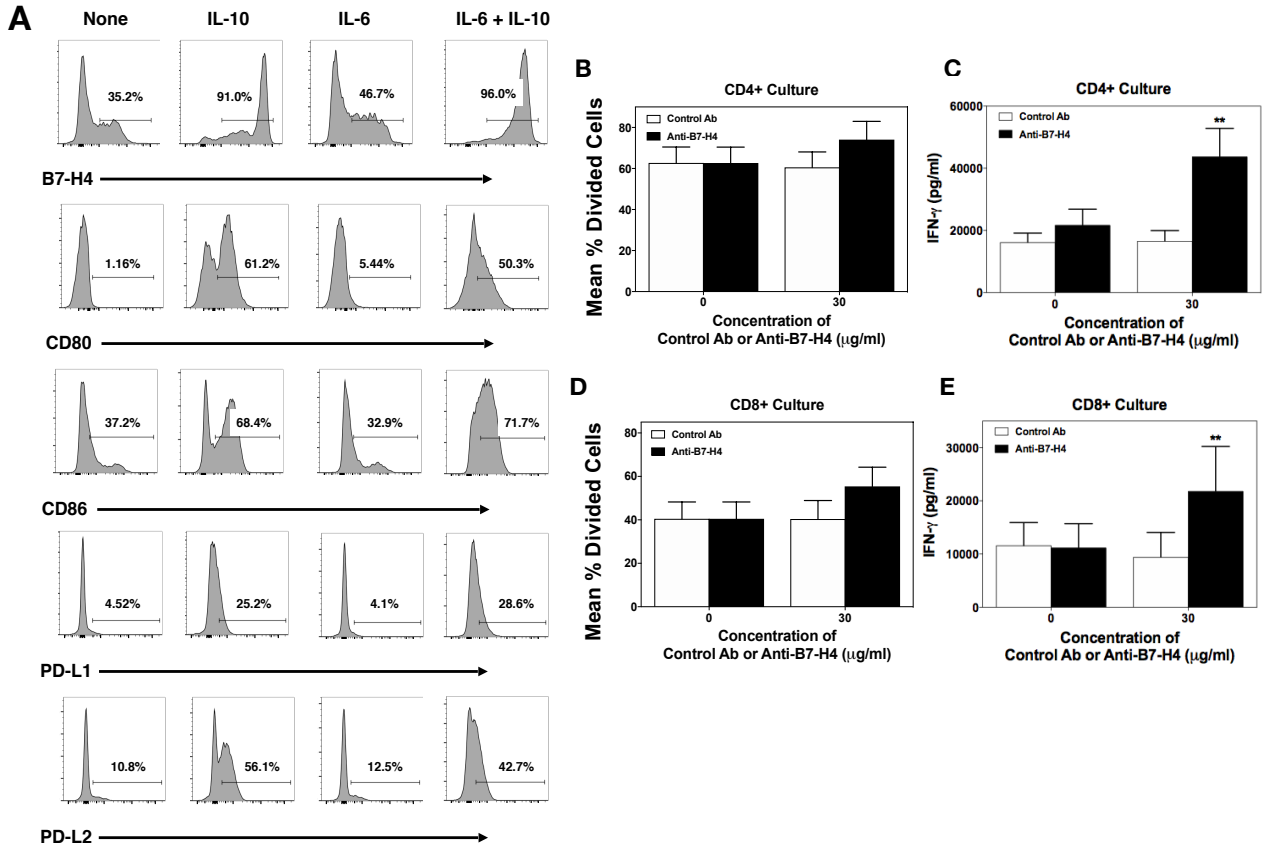


Figure 4. Anti-B7-H4 treatment increase IFN-γ secretion by T cell cultured in the presence of B7-H4+ monocytes. CD14^{+} monocytes were sort purified from healthy donor PBMCs (n=6) and cells were cultured in the presence of medium alone, IL-10, IL-6, or IL-10 plus IL-6 (20ng/ml) for 3 days. The percent of resultant monocytes expressing B7-H4, CD80, CD86, PD-L1, and PD-L2 was assessed (A). Sort purified autologous CFSE-labeled CD4^{+} T cells (B, C) and CD8+ T cells (D, E) were co-cultured with IL-10 plus IL-6 conditioned monocytes and anti-CD3 for 3 days, and the level of proliferation was assessed via flow cytometry (B, D) and the level of secreted IFN-γ measured (C, E). Proliferation data is presented as the mean percent divided cells and concentration of IFN-γ (pg/ml) present with the culture ± S.E.M. Asterisks (**) indicates a

statistically significant difference as compared to the Control Ab treated cultures, $p<0.01$, respectively.

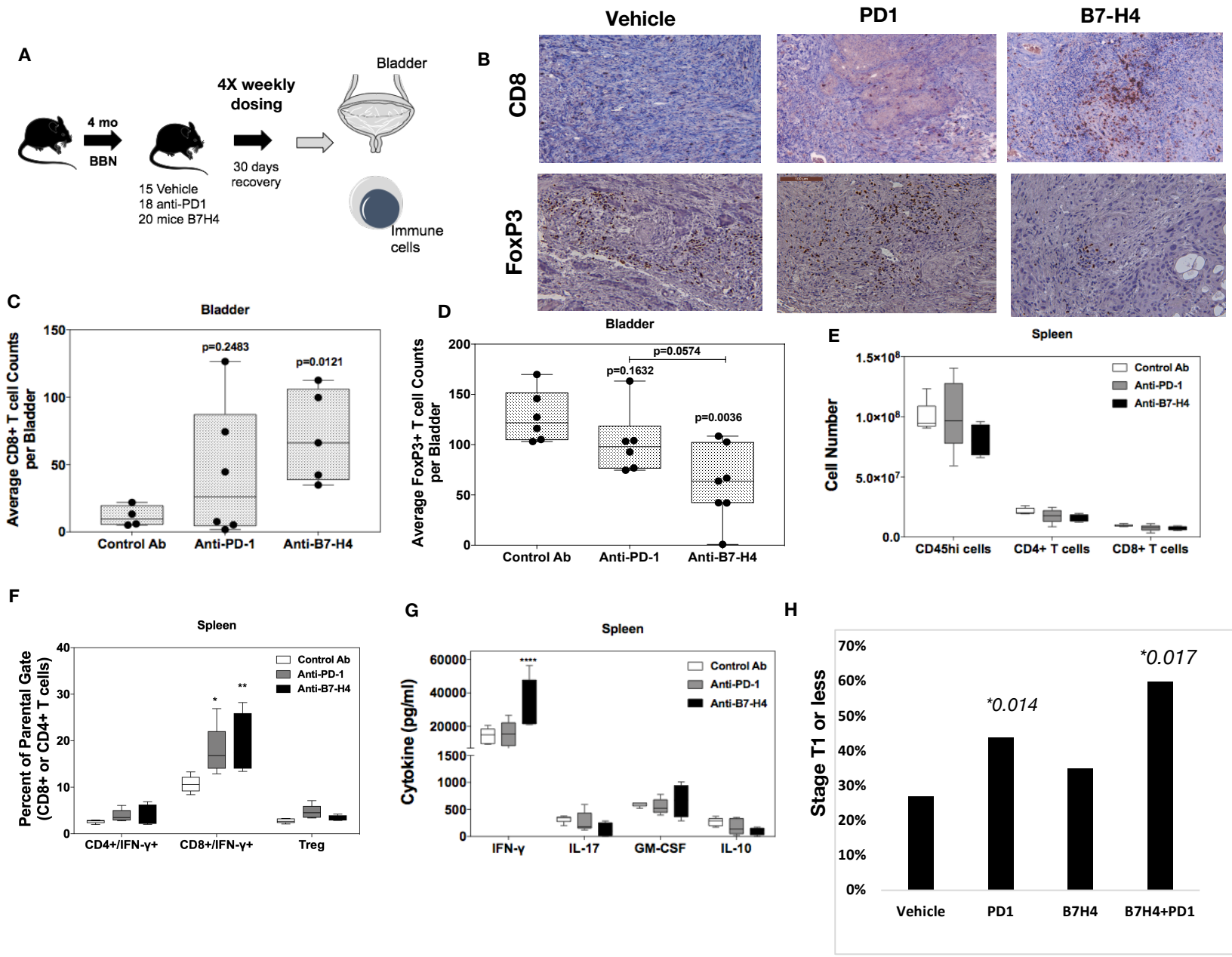


Figure 5. Anti-B7-h4 treatment increases CD8+ T cell function. (A) Treatment schema of mice for comparison of anti-tumor response of IgG, anti-PD1 and anti-B7-H4 antibodies. (B) Representative immunohistochemistry sections of BBN tumors for anti-CD8 (top) and anti-FoxP (bottom) staining. All images are at low-power (100X). Quantification of the average number of CD8+(C) and FoxP3 (D) immune cells per low power field. The number (E) and percentage (F) of singlet/live/CD45hi cell that were $CD4^+$/IFN-γ^+, $CD8^+$/IFN-γ^+, or $CD4^+$/$CD25^+$/$FoxP3^+$ present

within the spleens was assessed. The data are presented as the mean cell number or percentage of cells ± S.E.M. Total splenocytes were cultured in the presence of anti-CD3 (1μg/ml) for 3 days and the level of secreted cytokine assessed. The data are presented as the mean pg/ml of secreted cytokine (G). Asterisks (*, **, ****) indicates a statistically significant difference as compared to the Control Ab treated mice, $p < 0.05$, 0.01, 0.001 respectively. H) BBN mice treated with anti-PD1 and anti-B7H4 had lower rates of T2 or higher (detrusor invasion and greater) cancer compared to anti-PD1 or anti-B7-H4 alone.