

1 **Demographic history, linked selection, and recombination shape the genomic landscape of a broadly**
2 **distributed Pacific salmon.**

3

4 Quentin Rougemont^{1*}, Jean-Sébastien Moore¹, Thibault Leroy², Eric Normandeau¹, Eric B. Rondeau^{3,4}, Ruth
5 E. Withler⁵, Donald M. Van Doornik⁶, Penelope A. Crane⁷, Kerry A. Naish⁸, John Carlos Garza⁹ Terry D.
6 Beacham⁵, Ben F. Koop^{3,4}, Louis Bernatchez¹

7

8 ¹ Département de Biologie, Institut de Biologie Intégrative et des Systèmes (IBIS), Université Laval, Québec,
9 Québec, Canada QC G1V OA6

10 ² ISEM, Univ. Montpellier. CNRS, EPHE, IRD, Montpellier, France

11 ³ Centre for Biomedical Research, University of Victoria, Victoria, BC, Canada V8P 5C2

12 ⁴ Department of Biology, University of Victoria, Victoria, BC, Canada V8P 5C2

13 ⁵ Department of Fisheries and Ocean, Pacific Biological Station, 3190 Hammond Bay Road, Nanaimo,
14 British Columbia, Canada V9R 5K6

15 ⁶ National Oceanic and Atmospheric Administration, National Marine Fisheries Service, Northwest Fisheries
16 Science Center, Manchester Research Station, 7305 Beach Drive East, Port Orchard, Washington 98366,
17 USA

18 ⁷ Conservation Genetics Laboratory, U.S. Fish and Wildlife Service, 1011 E. Tudor Road, Anchorage, Alaska
19 99503

20 ⁸ School of Aquatic and Fishery Sciences, University of Washington, 1122 NE Boat Street, Box 355020,
21 Seattle, WA 98195 5020, USA

22 ⁹ Fisheries Ecology Division, Southwest Fisheries Science Center, National Marine Fisheries Service and
23 Institute of Marine Sciences, University of California–Santa Cruz, 110 McAllister Way, Santa Cruz,
24 California 95060

25

26 *Corresponding author e-mail: quentinrougemont@orange.fr (QR)

27 **Abstract**

28 Understanding the impacts of current human activities on within-species genetic variation requires a
29 thorough description of the historical factors that have shaped the genomic and geographical distribution of
30 nucleotide diversity. Past and current conditions influencing effective population size have important
31 evolutionary implications for the efficacy of selection, increased accumulation of deleterious mutations, and
32 loss of adaptive potential under the nearly neutral theory. Here, we gather extensive genome-wide data that
33 represent the extant diversity of the Coho salmon (*Oncorhynchus kisutch*) to address three issues. First, we
34 demonstrate that a single glacial refugium is the source of the majority of present-day genetic diversity, with
35 minor but detectable inputs from secondary micro-refugia. We propose a scenario whereby several ancestral
36 populations located south of the ice sheets expanded in postglacial time, swamping out most of the diversity
37 from other putative micro-refugia. Following this expansion, we identify particular populations having
38 undergone continuous declines in population size (N_e). Second, we combine multiple evidence from
39 demographic modelling, analysis of recombination landscape, and genome-wide landscape of diversity to
40 demonstrate that selection at linked sites and Hill-Robertson interference played a major role in shaping
41 genetic diversity across the Coho salmon genome. Third, we demonstrate that this demographic history
42 generated subtle differences in the load of deleterious mutations among populations, a finding that mirrors
43 recent results from human populations. Taken together, we found considerable support for the joint
44 contributions of demographic history and linked selection in the load of deleterious mutations. We suggest
45 that these inferences should be better integrated in conservation genetics of managed fish species which
46 currently focuses largely on within-population adaptation.

47

48 **Author Summary**

49 Reconstruction of a species' past demographic history from genome-wide data allows understanding how
50 historical factors interact with intrinsic genomic properties to shape the distribution of genetic diversity along
51 its genome and its geographic range. Here, we combine genotyping-by-sequencing and whole genome
52 sequence data with demographic modelling to address these issues in the Coho salmon, a Pacific salmon
53 species with rapidly declining census size in some parts of its range, notably in the south. Our demographic
54 reconstructions indicate a linear decrease in genetic diversity towards the north of the species range,
55 supporting the hypothesis of a major southern refugia for the Coho salmon and a northern route of
56 postglacial recolonization. Accordingly, the number of candidate deleterious homozygous derived mutations
57 was higher in northern populations. Demographic modelling also suggested the existence of cryptic refugia
58 that may have been missed with the use of simpler summary statistics. We further showed that the species'
59 genome was shaped by linked selection and biased gene conversion. In particular, local variation in
60 recombination rates have modulated the efficacy of natural selection. These processes, together with a
61 complex demographic history, can contribute to the load of deleterious mutations – an effect we argue should
62 be taken into account more routinely in conservation genetics studies.

63 **Introduction**

64 Both plant and animal biodiversity are currently declining or disappearing at unprecedented rates due to
65 human activity [1]. This leads to population size reduction, reduced genetic diversity, and to the sixth mass
66 extinction [2]. Before humans became major drivers of changes in species distributions and abundance,
67 long-term climate change had a major influence [3]. The Pleistocene glaciations resulted in major
68 contractions in the geographical distributions of many species into refugia that persisted in unglaciated areas
69 [4]. Postglacial range expansions often led to contacts between ancestral populations previously segregated
70 in different refugia [4,5]. The effects of long-term climate change combined with recent human-induced
71 population declines can foster genetic changes including a loss of genetic diversity, increased inbreeding,
72 increased load of deleterious mutations, and a loss of local adaptation [6].

73 In this context, it becomes important to understand how demographic history interacts with past and
74 ongoing selection and recombination to shape genetic variation. By disentangling past and current drivers of
75 range-wide genomic diversity, this information can inform management and conservation decisions [7].
76 Beyond conservation implications, such context provides a unique opportunity to address outstanding
77 questions in evolutionary biology. In particular, what is the role of gene flow in shaping heterogeneous
78 differentiation landscape during population divergence [8,9]? What are the demographic conditions required
79 to generate substantial differences in deleterious load among populations [10]?

80 A major challenge to understanding drivers of genome-wide patterns of diversity is that different
81 demographic processes can lead to similar contemporary genomic footprints [11]. As populations diverge,
82 they accumulate genetic incompatibilities forming barriers to gene flow [12], while the rest of the genome
83 may continue to be freely exchanged. As a consequence, the genomic landscape of divergence is expected to
84 vary, with greater differences between populations at genomic barriers as compared to genomic regions
85 exhibiting ongoing gene flow. However, similar patterns of heterogeneous genome-wide divergence can be
86 due to genetic hitchhiking of neutral alleles linked to selective sweeps [13] or to background selection (BGS;
87 [14]). These combined effects, referred to as linked selection reduces polymorphism at sites closely linked to
88 advantageous or deleterious variants, and therefore reduces local effective population size (N_e) along the
89 genome. The intensity of selection on linked loci will be mostly modulated by variation in local
90 recombination rate and by gene density [15,16]. Under linked selection, diversity (π , D_{XY}) and differentiation
91 (F_{ST}) metrics are expected to be positively and negatively correlated with genome-wide variation in
92 recombination rate (ρ) respectively, and with the density of targets (e.g., genes, regulatory regions) that are
93 subject to selection [14–18]. It is now increasingly recognized that neglecting BGS can bias demographic
94 inferences [19,20] or lead to false adaptive interpretations [21].

95 An understanding of historical demography is also essential for a sound interpretation of patterns of
96 deleterious mutation load observed among contemporary populations [10,22]. Population bottlenecks are
97 predicted to reduce potential for local adaptation, but also to reduce standing genetic variation and the
98 efficacy of selection [23,24]. In turn, a reduced efficacy of purifying selection leads to an increase in the
99 number of deleterious variants segregating in a population. Moreover, intrinsic genome properties, in
100 particular local variation in recombination rate or background selection, can favour the accumulation of

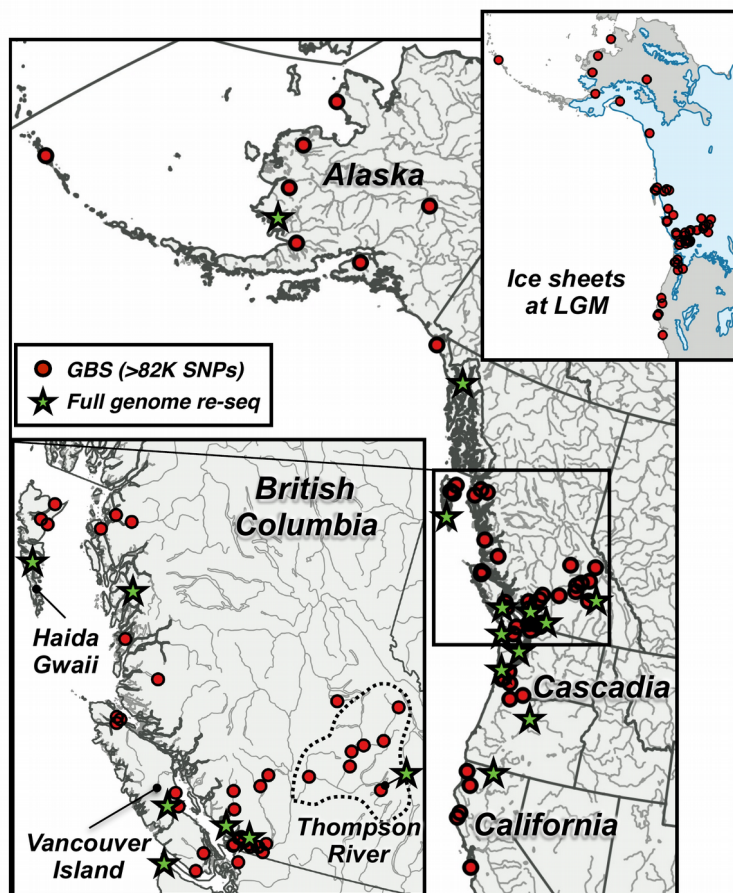
101 deleterious mutations [25]. From a conservation standpoint, populations harboring an elevated number of
102 deleterious variants might need to be monitored more closely.

103 Combining population genomics data with demographic modelling represents a powerful strategy to
104 test alternative hypotheses about historical drivers of existing genomic diversity. Previous studies employing
105 a similar approach have focussed mostly on species with a narrow geographic range, such as small islands
106 [26,27], which are on the verge of extinction [28–31] are strongly bottlenecked [32], shedding light on the
107 evolutionary consequences of small population size. Few studies, however, have investigated how historical
108 processes have shaped the geographical patterns in the distribution of genomic diversity in more broadly
109 distributed species, e.g., at the scale of a whole continent. An exception to this observation is the vast
110 literature on demographic reconstructions of human populations. Long-lasting debates in this literature
111 regarding the role of demography in generating mutation load differences among populations [22,33,34]
112 could benefit from studies of species displaying similarly complex demographic histories and broad
113 geographic distributions.

114 Salmonid fishes are economically important species that have suffered recent demographic declines
115 [35,36]. This is particularly the case for Coho salmon (*Oncorhynchus kisutch*), one of the five anadromous
116 species of Pacific salmon that supports important recreational and indigenous subsistence fisheries, which
117 has suffered dramatic population declines (> 90%) over the last three decades in parts of its range [36,37]. A
118 previous study investigated the range-wide population structure and demographic history of the species and
119 found a cline of decreasing diversity from south to north, as well as some endemic diversity in small putative
120 refugia [38] (see also [39]). This study indicated that Coho salmon may have survived the last glacial
121 maximum (LGM, i.e. the Fraser Glaciation in British Columbia, and the McConnell/McCauley Glaciation in
122 Yukon and Alaska; 23 to 18 Ky ago) in unglaciated areas of Haida Gwaii and Beringia in addition to areas
123 south of the ice sheets. This study, however, predates the genomic era and could not eliminate alternative
124 hypotheses regarding the origin and number of glacial refugia during the LGM. Most importantly, the
125 impacts of confounding factors such as background selection, recombination rate variation, and how these
126 factors may facilitate the accumulation of deleterious mutations could not be studied with the limited number
127 of genetic markers available at the time. In North America, the species is currently distributed from
128 California to Alaska [40]. Unglaciated areas that could potentially serve as glacial refugia persisted both
129 north (e.g. the Beringian refugium in Alaska, the Yukon Territory of Canada and areas of Asia and the Bering
130 Land Bridge) and south (e.g. all of the deglaciated area south of British Columbia, Canada) of the ice sheet
131 [40–42]. Other unglaciated areas (e.g. Haida Gwaii in British Columbia) could also have been micro-refugia
132 [43,44]. In this context, distinct demographic scenarios can be tested. Under a first scenario whereby
133 populations expanded north from a single southern refugium, we predict: *i*) a latitudinal decrease in genetic
134 diversity from south to north along with a pattern of IBD, and *ii*) ancestral populations located in areas south
135 of the ice sheets. Under a second scenario, populations expanded south from a single northern refugium, and
136 we predict the opposite geographic pattern. The third scenario corresponds to the survival of populations in
137 different refugia where we predict: *i*) the existence of clearly distinct genetic clusters, and *ii*) postglacial gene

138 flow with signatures of secondary contacts, with contact zones displaying higher genetic diversity through
139 postglacial admixture between different genomic backgrounds.

140 In order to test these alternative scenarios, we generated genome-wide data from nearly 2,000 Coho
141 salmon from California to Alaska, one of the most extensive genomic datasets for a non-model vertebrate
142 species to date. First, to resolve the species demographic history, we used a modelling approach that accounts
143 both for barriers to gene flow affecting migration locally, and for linked selection affecting the rate of drift.
144 Next, we tested the above predictions related to linked selection. Finally, we hypothesized that demographic
145 history and background selection shaped the pattern of deleterious mutation load, both within and among
146 populations. In particular, we hypothesized that postglacial re-colonisation influenced levels of standing
147 genetic variation and favoured the accumulation of deleterious mutations at the expansion front. In these
148 conditions, we predicted that genetic diversity should decrease as a function of the distance from the
149 ancestral populations, while the accumulation of putatively deleterious mutations should increase as a
150 function of the distance to the ancestral populations.



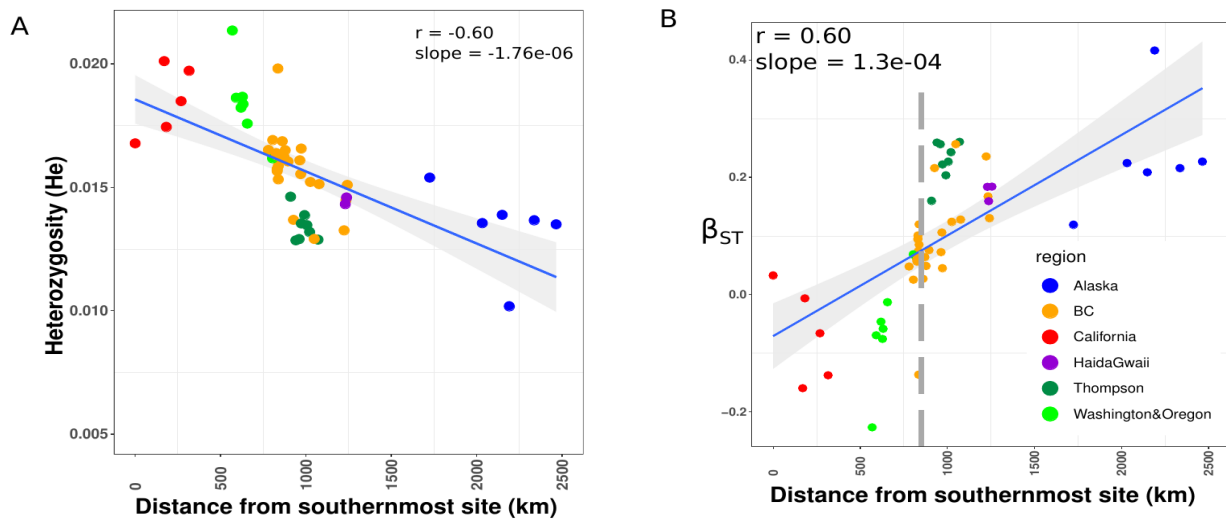
152 **Figure 1) Sampling design** A) Sampling locations of 58 Coho salmon populations distributed across the species' North
153 American range of distribution. Each dot represents a sampling location. Inset: Map showing the extend of ice-sheet
154 during the Last Glacial Maximum 13 KyA. Data modified from [115]

155 Results

156 Overall genetic diversity and population structure

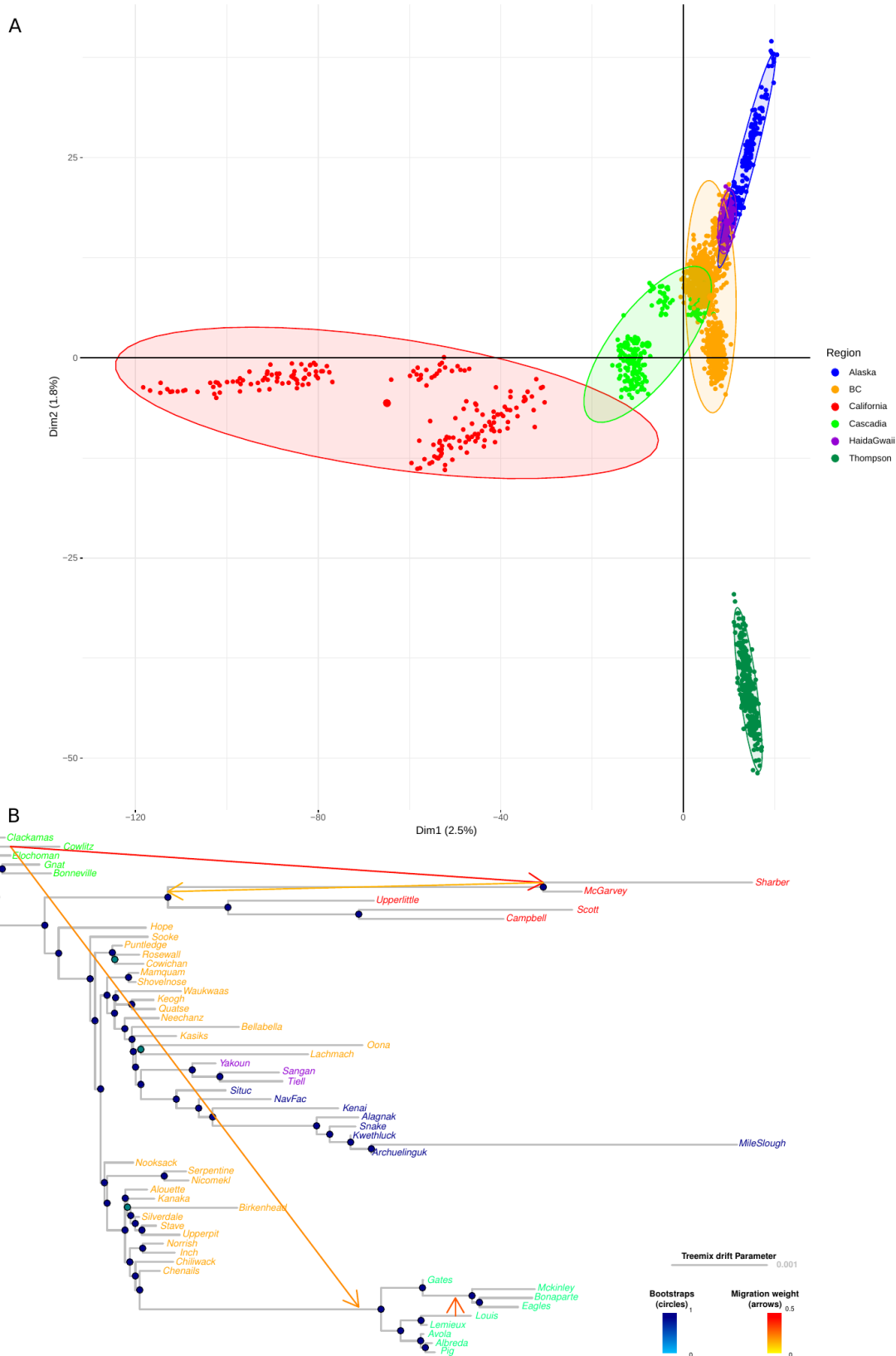
157 A total of 1,957 individuals was sampled from California to Alaska representing 58 sampling locations
158 (mean $n = 34$ fish per location, Fig 1a, S1 Table) and genotyped using a genotype by sequencing (GBS)
159 method that generated 82,772 high quality filtered single nucleotide polymorphisms (SNPs). Another set of
160 55 individuals representing 11 sampling locations from the same range (Fig 1a), were whole genome
161 sequenced (WGS) to $\sim 30X$ coverage, and used in specific analyses (S2 Table).

162 Levels of genetic diversity (observed and expected heterozygosity, π_{SNP}) were highest in formerly
163 deglaciated areas in the south (California, Cascadia, Fig 2a, Fig 1b) and decreased as a function of distance
164 from the southernmost site up to Alaska ($r = 0.64$, $p < 0.0001$, Fig 2a, S1 Fig). The Thompson River
165 watershed (Thompson R. hereafter) in southern British Columbia was an exception to this latitudinal pattern
166 and displayed the lowest average level of regional genetic diversity of all sampling locations which we
167 hypothesized to results from bottlenecks in this area. The remaining samples from British Columbia were
168 intermediate in genetic diversity.



170 Figure 2) Genetic diversity and differentiation

171 A) Linear relationship between expected heterozygosity and distance from the southernmost population located in
172 California. B) Linear increase in genetic differentiation as measured by β_{ST} as a function of the distance from the
173 southernmost population located in California. Negative values indicate the most likely ancestral population. The
174 relationship in A and B was tested using linear models. The grey vertical bar in panel B) indicates the approximate
175 location of the southern limit of the ice-sheet at the end of the last glacial maxima.



177 **Figure 3) Genetic structure and gene flow**

178 A) Principal Component Analysis (PCA) summarising population genetic structure among 1,957 individuals based on
 179 the principal component axes 1 and axes 2. Each point represents an individual and the colours represent the major
 180 regional groups. B) Inference of population splits and mixture by Treemix with 4 statistically significant migration
 181 events ($p < 0.0001$). Bootstrap supports and migration weights are indicated according to the colour scale.

182 The distribution of singletons provided further information regarding the most ancestral populations, with
183 older populations expected to have accumulated a higher density of singletons [45]. Counting the number of
184 singletons by sampling site and averaging by regional groups revealed the following differences: Cascadian
185 samples contained the highest number of singletons, with a mean of 1,263 singletons per site. Californian
186 samples had the fewest number of singletons ($n_{\text{MEAN}} = 55$) while Alaska harbored intermediate density (n_{MEAN}
187 $= 966$). Consistently, WGS data revealed 2.7 times more singletons in Cascadia (Tsoo-Yess River) as
188 compared to Alaska (Kwethluk River), whereas Thompson R. samples contained 6.7 times less singletons,
189 supporting the hypothesis of a pronounced bottleneck in these populations (S3 Table). Similarly, the
190 occurrence of private alleles was more prevalent among southern than northern populations, being nearly
191 twice as high in Cascadian ($n = 10,097$) than in Alaskan populations ($n = 5,270$). Again, populations from the
192 Thompson R. were an exception to this pattern with the lowest level of private polymorphism ($n = 1,479$, S2
193 Fig).

194 Comparing the decay of linkage disequilibrium (LD) across populations based on the WGS data
195 indicated rapid LD decay across all samples (S3 Fig). The Thompson R. populations again departed from the
196 general pattern with a much reduced LD decay. Indeed, r^2 values decreased to 0.4 at approximately 342 kb
197 for Thompson R. whereas this r^2 value was attained between 13 and 34 kb for all other populations.

198 Next we used the β_{ST} coefficient to identify ancestral population [46]. Unlike F_{ST} estimates [47], this
199 index can account for the non-independence among populations and negative values are indicative of
200 ancestral populations [46]. Here, β_{ST} indicated that ancestral populations were located in previously
201 unglaciated areas corresponding to Cascadia ($n = 5$ localities), California ($n = 3$ localities) as well as one site
202 from southern British Columbia (Fig 2b, S4 Table). A linear decrease in β_{ST} as a function of distance from the
203 southernmost site was observed ($r = 0.60$, slope = $1.03e-04$, $p < 0.0001$) as expected under IBD. Support for
204 this IBD pattern was also observed using F_{ST} (S4 Fig, $r = 0.66$, slope = $4e-05$, $p < 0.0001$) as well as Mantel
205 tests ($r = 0.64$; $p < 0.0001$, $r = 0.72$; $p < 0.0001$ when removing Thompson R. populations). Average pairwise
206 F_{ST} across all populations was 0.095 and varied from 0.002 to 0.334 (S5 Fig), indicating moderate population
207 structure, typical of anadromous species connected by gene flow [48].

208 Model-based analysis of population structure failed to reveal a clear number of distinct populations
209 (K value). Instead, K values ranging from 30 to 60 all fit the data well (S6 Fig), due to the confounding
210 effect of IBD. The first axis of a Principal Components Analysis (PCA, Fig 3a) revealed a separation of the
211 sample from South to North with the most divergent samples found in California. The second axis revealed
212 the divergence from East to West but with a strong separation of the Thompson R. populations. Along these
213 axes populations followed an IBD pattern. These results were also supported by an MDS analysis (S7 Fig).
214 The third and fourth axis did not yield further information (S8 Fig).

215 We then used Treemix [49] to infer population splits and gene flow (Fig 3b). A first tree assuming no
216 migration (i.e. drift only) explained 98.1% of the variance observed. Adding up to four significant migration
217 events ($p < 0.0001$) explained over 99.1% of variance, then the proportion of explained variance plateaued
218 (S9 Fig). Populations from Cascadia occupied basal positions. California populations displayed pronounced
219 genetic drift, corroborating the high divergence observed in the PCA. Populations from Alaska (MSL River)

220 and Thompson R. also displayed higher genetic drift, in line with evidence based on analyses of genetic
221 diversity and genetic structure. The two most supported migration events occurred from Cascadia south to
222 California and north to the Thompson R. We note that populations followed the south to north arrangement,
223 with the samples from Cascadia displaying less drift than those further north.

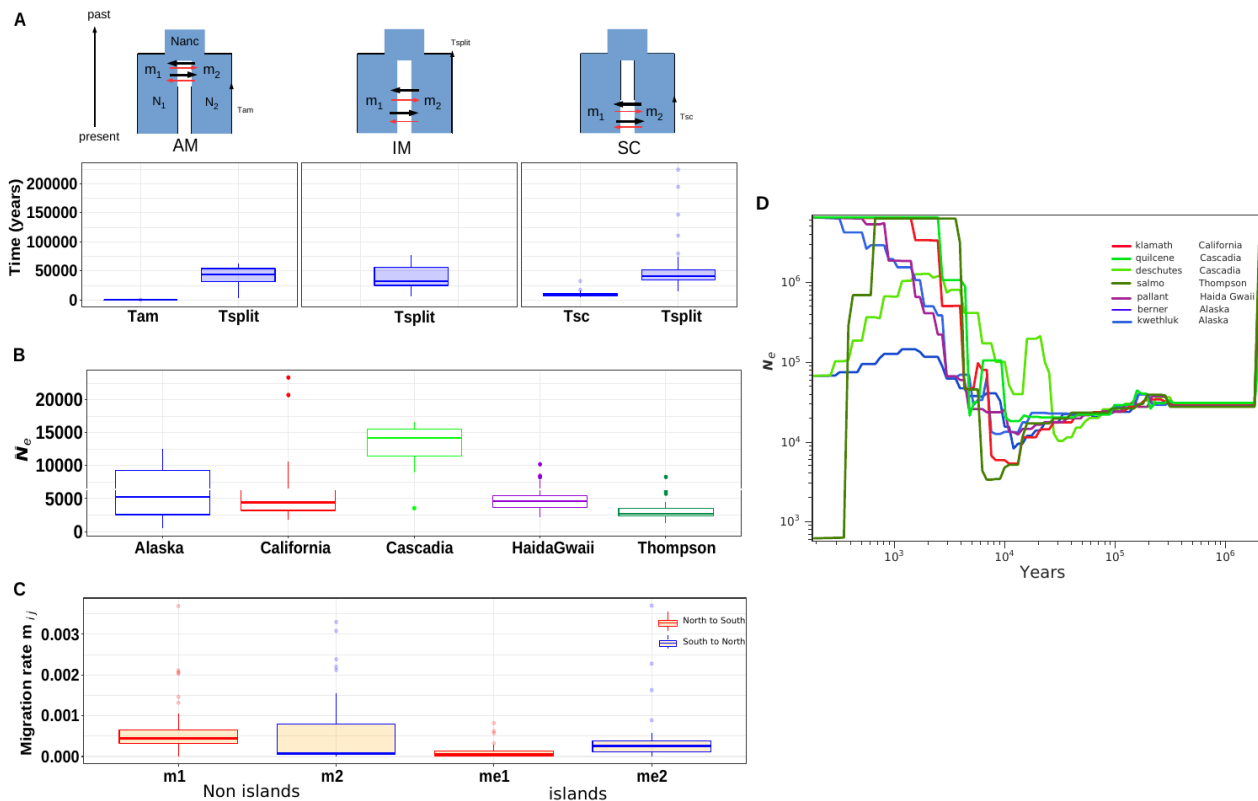
224

225 *Demographic history*

226 In order to assess more formally the occurrence of one or more refugial origins for contemporary
227 populations, we performed the following explicit model-based inferences of population divergence scenarios
228 using $\partial a \partial i$ [50]. Our models account for the confounding effects of selection at linked sites and that of the
229 accumulation of local barriers to gene flow in the genome [19,51]. Four major demographic models were
230 statistically compared using groups identified in the PCA and with a focus on previously hypothesized
231 refugia (i.e., Cascadia, California, Haida Gwaii, Alaska). The following models were tested: strict isolation
232 (SI model), divergence with ancient migration (AM model), divergence with continuous gene flow (IM
233 model) and secondary contact (SC model; S10 Fig). A total of 69 pairwise comparisons was performed with
234 $\partial a \partial i$. In each pairwise comparison one single representative population from a putative refugium was
235 compared against one population from another refugium. Due to the high local structure within groups, we
236 avoided pooling samples into higher order groups (e.g., regional groups) as this would unavoidably bias our
237 results. Models incorporating linked selection and restricted introgression along the genome always received
238 highest support (S5 Table). ΔAIC (minimum value > 5) confidently discriminated models in 87% of the $\partial a \partial i$
239 comparisons (S5 Table). The SC model received the highest support in 46% of the comparisons, the AM
240 model in 30%, and the IM model in 14%, with 13% remaining unclassified. Scenarios including periods of
241 gene flow clearly outperform scenarios assuming no gene flow. The fact that secondary contact was the best-
242 supported model suggests that more than one glacial refugia contributed to the recolonization of the
243 contemporary range occupied by Coho salmon.

244 Assuming a generation time of 3.5 years [52] and mutation rate of $8e^{-9}$ bp/generation revealed
245 similar divergence time estimates among models (i.e., 57 Kya under SC, 40 Kya under IM, and 42 Kya under
246 AM, S6 Table). The median time of secondary contact (SC) was 10 Kya [min = 4,800 – max = 32,900],
247 corresponding roughly to the onset of the last glacial retreat (Fig 4A). Parameter estimates under the AM
248 model supported a very recent reduction in gene flow (mean < 1 Kya), which therefore represents a
249 demographic model similar to an IM model. Pronounced variation in effective population size (N_e) was
250 observed with the highest values in Cascadia populations (mean 12,800; range [3,500 – 16,500]). On
251 average, the smallest N_e were observed in the Thompson R. populations (mean 3,000; range [1,300 – 8,300],
252 Fig 4B), again consistent with a population bottleneck. However, smaller N_e values were also observed in
253 the isolated Navfac Creek population in Alaska ($N_e = 500$) and the Scott Creek population in California (N_e
254 = 1,800), which are the most divergent populations identified in the PCA. The McGarvey (California) and
255 Clackamas (Cascadia) populations identified as “ancestral” according to the β_{ST} displayed the highest N_e
256 (23,000 and 16,500). We also note that incorporating linked selection and barriers to gene flow further
257 reduced N_e , regardless of the model (S11 Fig). Intrinsic barriers to gene flow reduced the estimated

258 migration rate by half of its value and affected 20 to 40% of the genome (Fig 4C, S6 Table). Similarly, the
 259 Hill-Robertson factor suggested that N_e was reduced to 37%-50% of its initial estimate and that
 260 approximately half of the genome was affected (S6 Table).



262 **Figure 4) Inferences of demographic history using $\partial\text{a}\partial\text{i}$ and $\text{smc}++$**

263 A) Estimates of divergence time (in years) between each major region as inferred by $\partial\text{a}\partial\text{i}$ under the best model
 264 (displayed in blue) based on the GBS SNP data set. B) Estimates of effective population size N_e for each major region
 265 as inferred by $\partial\text{a}\partial\text{i}$ under each best demographic model based on the GBS SNP data set. C) Estimates of migration rate
 266 among populations in neutral regions (m) of the genome and in areas of restricted recombination (me) based on the
 267 GBS SNP data set. D) Estimates of effective population size (N_e) change through time (in years) for the whole genome
 268 data using $\text{SMC}++$.

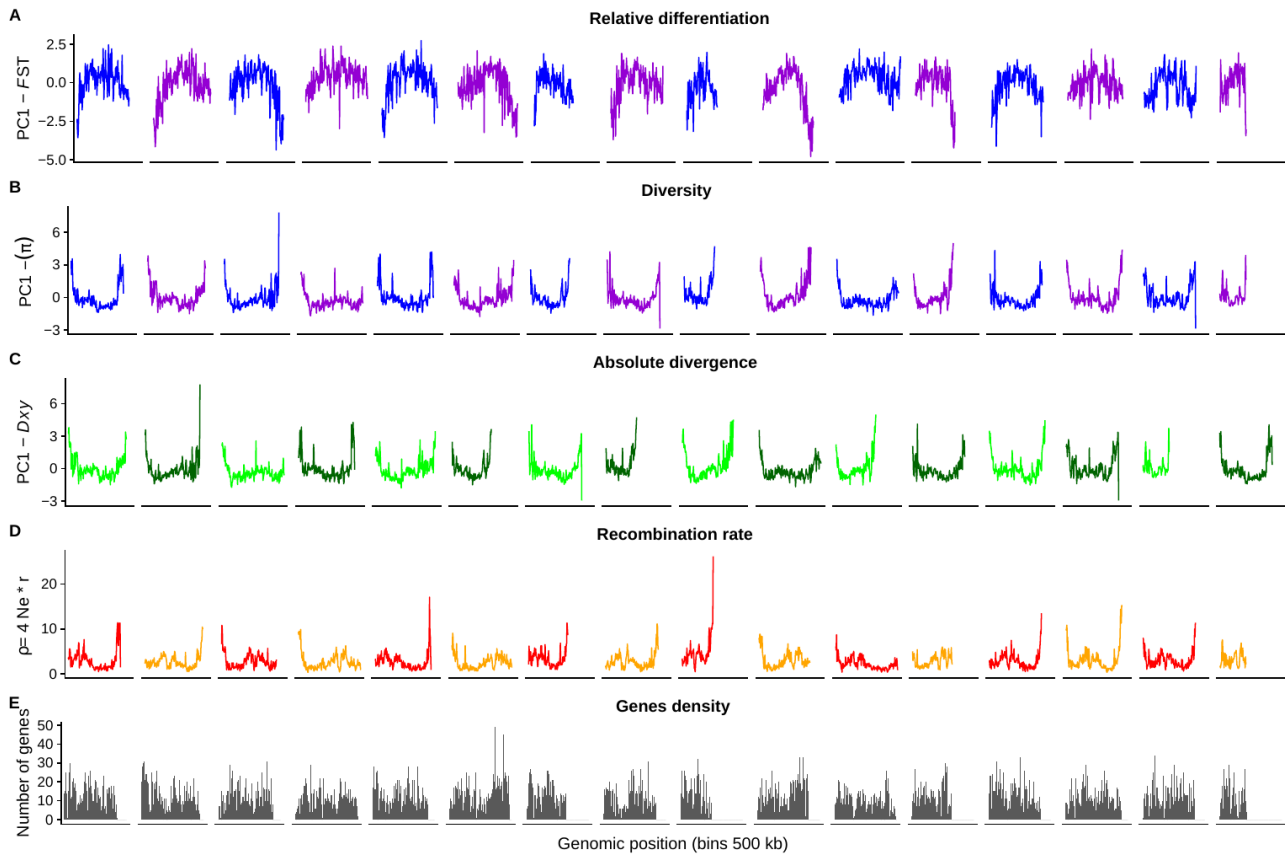
269
 270 We then investigated historical change in effective population size using the Sequentially Markovian
 271 Coalescent in $\text{SMC}++$ [54] and the 55 whole genome sequences, which are representative of 11 populations
 272 from California to Alaska (Fig 1, S2 Table). For ease of visualization, results for a subset of the populations
 273 are presented in Fig 4D and details for all populations are displayed in S12 Fig. This analysis revealed: i) an
 274 expansion of most populations approximately 12-20 KyA, concomitant with postglacial re-colonisation, ii) a
 275 slow and steady decline in the Thompson R. (Fig 4D, S12 Fig), and iii) a split time between all pairwise
 276 combinations of populations (median = 16,3 KyA, range = 6,7KyA - 61KyA, S13 Fig) compatible with the
 277 onset of postglacial population expansion (Fig 4D) which was accompanied by an increase in N_e across
 278 samples (Fig 4D). We note that using a different mutation rate ($1,25\text{e-}8$ mutations/bp/generations) yielded
 279 estimates of split time that were more in line with the estimates of postglacial expansion (median = 9,6 KyA,

280 S13 Fig) and with less variance among compared population (min = 5 KyA – max = 18 KyA). Overall,
281 SMC++ results are consistent with previous analyses indicating that until recently, all populations shared a
282 similar demographic history. We caution against a strict interpretation of the most recent time. Indeed,
283 reliable estimates for present time require a high number of samples, whereas we only had five samples per
284 populations [54]. Similarly, exact N_e estimates should be interpreted cautiously and only the trend should be
285 considered rather than the exact values.

286

287 *Linked selection shape the Coho salmon genomic landscape*

288



290 **Figure 5) Genome-wide landscapes in Coho salmon** with: A) landscape of differentiation, B) landscape of genetic
291 diversity C) divergence, D) recombination and E) gene density. Plots are averaged over 500-kb windows. Only the first
292 16 chromosomes are displayed for simplicity. The same figure with all chromosomes is available in Fig. S15.

293

294 We first described patterns of recombination and then tested correlations among genetic diversity (π),
295 divergence (D_{XY}), and differentiation (F_{ST}) parameters measured across the whole genome using 500 kb
296 sliding windows (Fig 5 A-C) and the population-scaled landscape of recombination (ρ) (Fig 5D) and gene
297 density (Fig 5E) to test our predictions using whole genome sequences.

298 This analysis first revealed a heterogeneous recombination landscape that varied both within and
299 among chromosomes. In particular, recombination rate was higher towards the ends of chromosomes (Fig
300 5D), as observed across many species [55] Second, we observed a negative correlation between
301 recombination rate and chromosome length ($R^2 = 0.481$, $p < 0.0001$, S14 Fig). To investigate the effects of

302 selection at linked sites, we summarised π (across all 11 WGS populations), D_{XY} and F_{ST} statistics (55
 303 pairwise comparisons), using the first axis of a PCA (PC1) to obtain the common variation in these metrics
 304 and reduce the dimensionality to single summary statistic (Fig 6, S15 Fig, S7 Table). PC1 captured 99% of
 305 variance in π and in D_{XY} , indicating that the PC1 was effective at summarizing information about diversity
 306 and divergence. All linear correlations among the tested variables were significant ($p < 0.0001$, S8A Table).
 307 We found a positive correlation between PC1- π and recombination ($r = 0.54$, S8A Table) and a negative
 308 correlation between PC1- π and gene density ($r = -0.20$) (Fig 6A-D). We also found a positive correlation
 309 between PC1- D_{XY} and recombination ($r = 0.55$) and a negative correlation between PC1- D_{XY} and gene
 310 density ($r = -0.21$). PC1- F_{ST} was negatively, albeit weakly correlated with both recombination ($r = -0.05$) and
 311 gene density ($r = -0.04$, Fig 6B-E and C-F). We observed a correlation of 0.999 between π and D_{XY} , as
 312 expected here because of a very recently shared common ancestor (Fig 6G). We also found a negative
 313 relationship between PC1- π and PC1- F_{ST} ($r = -0.44$, Fig 6H) as well as between PC1- D_{XY} and PC1- F_{ST} ($r =$
 314 -0.43 , Fig 6I). The smaller correlations between statistics involving F_{ST} can be explained by the modest
 315 variance in F_{ST} explained by PC1 (37%).

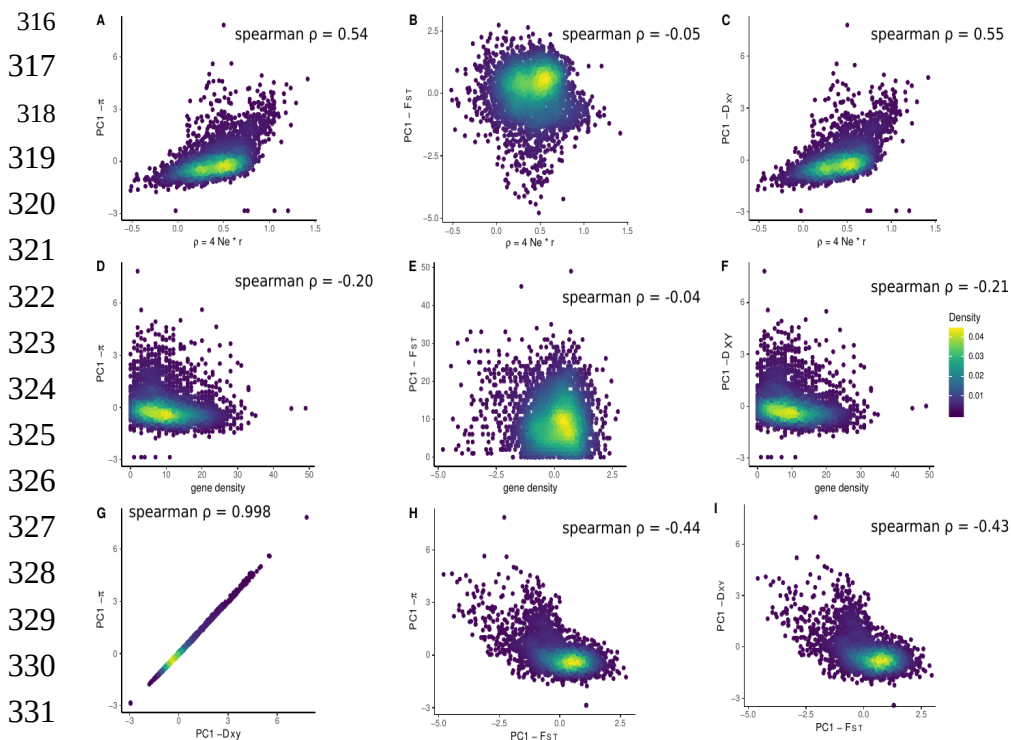


Figure 6) Correlations among different metrics.

Correlations between recombination rate ($\rho = 4Ne \cdot r$) genetic diversity (π), divergence (D_{xy}) and differentiation (F_{ST}) statistics (A-C), and correlation between gene density and π , D_{xy} , F_{ST} statistics (D-F). Correlations between π , D_{xy} and F_{ST} statistics (G-I). The Spearman correlation values are plotted but mixed linear models were fitted to further test interaction among explanatory variables (recombination rate and gene density).

334 In order to investigate further the effects of linked selection, we used mixed linear models that integrate
 335 interactions among tested variables (i.e. recombination, gene density). These revealed a significant effect of
 336 both recombination ($t = -41.17$, $p < 0.0001$, S8B Table) and gene density ($t = 5.72$, $p < 0.0001$) on PC1- D_{XY} (R^2
 337 $= 0.348$). The same was true when considering the correlation of PC1- π with recombination ($R^2 = 0.55$, Fig
 338 6A-D, $t = -40$, $p < 0.0001$) and gene density ($t = 5.22$, $p < 0.0001$). Significant effects were also found between
 339 PC1- F_{ST} and these variables (recombination: $t = 6.91$, $p < 0.0001$, gene density $t = 6.27$, $p < 0.0001$,
 340 interaction: $t = -4.073$, $p < 0.0001$, $R^2 = 0.025$, S8B Table). Similar correlations were found when analysing

341 populations separately (S9 Table). Therefore, our results suggest a role of linked selection in shaping the
 342 patterns of diversity along the Coho salmon genome.

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

373

374

375

376

377

378

379

Figure 7) Correlation

between π_N/π_S and

GC3 content for each

population separately

π_N/π_S is used here as a

proxy for the efficacy of

purifying selection with

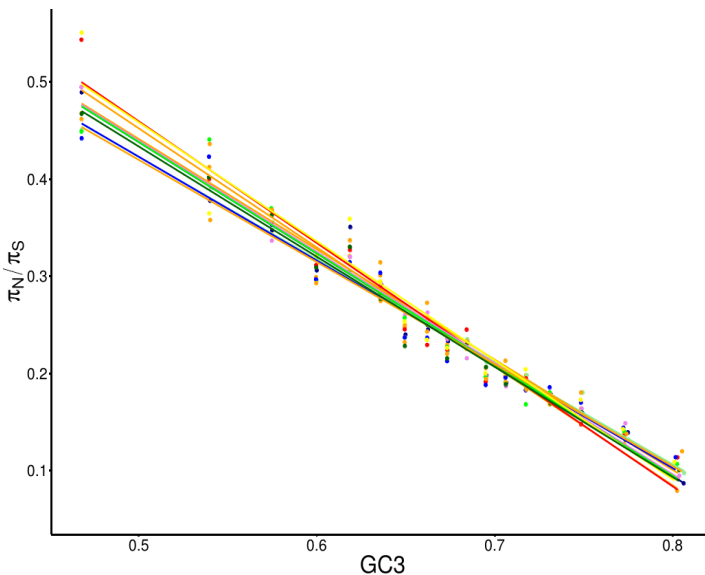
smaller values reflecting

more efficient purifying

selection.

All correlations are

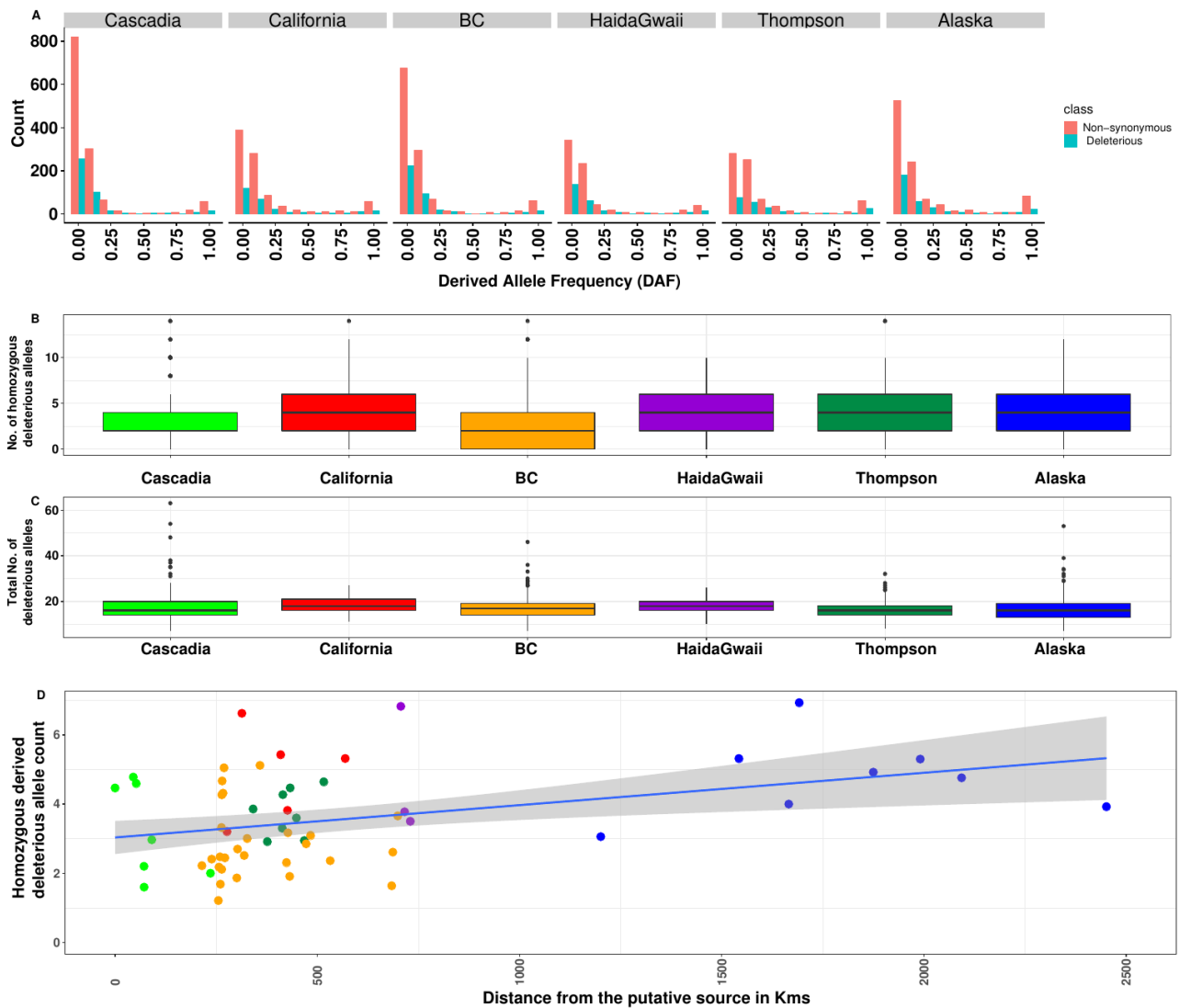
significant ($p < 0.001$).



We then tested whether the efficacy of selection also played a role in shaping patterns of diversity along the Coho salmon genome using the whole genome sequences. We measured ratios of non-synonymous to synonymous polymorphisms (π_N/π_S) as a measure of the efficacy of natural selection and GC content at the third codon position (GC3) as a proxy of local recombination rates. This metric was preferred as an indicator of localized recombination rate variation at the gene scale, as opposed to the large population scale estimates of recombination (ρ) over 250kb windows used above. Indeed, biased gene conversion (gBGC) occurs because base mismatch during homologous recombination is known to be biased toward G and C (conversion preferentially favours G+C over A+T bases) [56,57]. Here, we observed that the correlation between GC3 and ρ ranged from $r=0.3$ to 0.72 , depending on population. These modest correlations likely reflect the loss of information about fine scale recombination rate variation when using 250kb sliding windows, which also provides added justification to use the GC3 as a proxy. We found a strong correlation between π_N/π_S and GC3 for all populations (Fig 7, linear models all $R^2 > 0.9$, $p < 0.0001$, S10 Table). Next, we observed a negative correlation (linear model, $R^2 = 0.28$, $p = 0.051$) between historical N_e measured before the onset of population expansion $\sim 13,000$ years ago and π_N/π_S ratio (S16 Fig), in agreement with the nearly neutral theory of molecular evolution and indicating that our N_e estimates were good proxies of genetic diversity. Second, among-population differences in π_N/π_S across all genes were modest (mean = 0.252), with a minimum value of 0.248 observed in Tsoo-Yess R. (Cascadia) and Robertson Creek (BC) and maximum values of 0.257 and 0.256 observed in the Thompson R. and Inch Creek (BC) respectively. The π_N/π_S differences among populations were more contrasted in areas of low recombination of the genome (Fig 7) with lowest π_N/π_S values observed in the Kwethluck (Alaska, $\pi_N/\pi_S = 0.442$), Quilcene (BC, $\pi_N/\pi_S = 0.449$), and Deschutes (Cascadia, $\pi_N/\pi_S = 0.451$) river populations. The highest values were observed in the Thompson ($\pi_N/\pi_S = 0.551$), Klamath (California; $\pi_N/\pi_S = 0.543$) and Capilano (BC; $\pi_N/\pi_S = 0.495$) river populations.

380 *Spatial patterns of variation in deleterious mutations*

381



383 **Figure 8) Analysis of deleterious load in the Coho salmon genome**

384 A) Derived allele frequency spectra across major groups. The non-synonymous variants and putatively deleterious
 385 variants frequency are shown. Data are normalized for a sample of size $n = 102$ corresponding to the smallest size for
 386 the combined samples in Haida Gwaii. B) Distribution of the count of homozygous derived deleterious alleles in each
 387 major group. C) Distribution of the count of total derived deleterious alleles in each major group. D) Correlation ($p =$
 388 0.0034 , $R^2=0.13$) between geographic distance to the putative ancestral source and the distribution of homozygous
 389 derived putatively deleterious mutations.

390

391 Given the complex demographic history of population size changes and spatial expansions leading to
 392 secondary contacts in Coho salmon and the inference of linked selection, we predicted that deleterious
 393 mutations should segregate at higher frequencies at the edge of the norther expansion front in Alaska, but
 394 also in California likely representing a southern expansion from the Cascadian refugium, as well as in the
 395 bottlenecked Thompson R. populations.

396 To do so, we tested for an increase in the derived allele frequency (DAF), and homozygosity of
397 predicted deleterious mutations at non-synonymous sites. We estimated derived alleles using whole genome
398 information from three outgroups: 1) Chinook salmon (*Oncorhynchus tshawytscha*) [58], 2) Rainbow trout
399 (*Oncorhynchus mykiss*) [59] and 3) Atlantic salmon (*Salmo salar*) [60]. Out of 4,427 non-synonymous
400 mutations identified with the GBS dataset from all 58 populations, PROVEAN (63) predicted a total of 1,297
401 deleterious mutations in at least one of these populations and for which we were able to identify the derived
402 allele. Deleterious mutations were maintained at lower DAF than synonymous variants (Wilcoxon-tests, all p
403 < 0.001). The DAF spectrum (Fig 8A) were computed for each major region (using a sample of size $n = 100$)
404 and for each population separately (S17 Fig). DAF were significantly different, among region (Kruskal-
405 Wallis chi-squared = 100.57, $df = 5$, p -value $< 2.2e-16$) as well as among population (Kruskal-Wallis chi-
406 squared = 638.59, $df = 57$, p -value $< 2.2e-16$, S11 Table). They showed that Alaska, Thompson R. and
407 California populations displayed more fixed deleterious mutations ($n = 18, 12$ and 11 respectively) than those
408 from the Cascadia region ($n = 3$). Haida Gwaii and BC populations were intermediate with 7 and 6 fixed
409 deleterious mutations respectively (Fig 8A). The three former regions also displayed higher frequencies of
410 polymorphic derived deleterious mutation (S18 Fig) compared to British Columbia or Cascadia (Wilcoxon
411 test, $p < 0.001$).

412 Next, we examined the count of homozygous (proportional to the load under a recessive model
413 [33,34]), heterozygous and of total derived deleterious mutations (proportional to the load under an additive
414 model [33,34]). In particular, we expected that mutations in a heterozygous state should be more frequent
415 than in a homozygous state, especially in populations with higher effective sizes, where selection should be
416 more effective at purging these mutations [23]. We found that 77% of deleterious mutations were maintained
417 in heterozygous states across all samples. Also, fish from Alaska, Haida Gwaii and California populations
418 harbored a significantly higher number of deleterious mutations in a homozygous state when compared to
419 Cascadia or British-Columbia (Wilcoxon-test, $p < 0.01$) (Fig 8B, S12 Table). When considering the total load
420 of derived deleterious mutations, we found that, on average, there were significantly more putatively
421 deleterious variants per individual in California, Cascadia, and Haida Gwaii populations than in fish from
422 Alaska, British-Columbia or the Thompson R. watershed (S12 and S13 Table, Fig 8C, Wilcoxon-test, $p <$
423 0.01), although these differences were modest. Finally, we tested the prediction that distance from the likely
424 ancestral source predicts the deleterious load, as observed in human populations [33]. We found a nearly
425 linear relationship between the distance from the putative origin of ancestral populations (the site with the
426 lowest β_{ST} in Cascadia) and the number of derived, homozygous, putatively deleterious mutations (HDD;
427 linear models, $p < 0.0001$; $R^2 = 0.13$, Fig 8D). Under the hypothesis that higher recombination rate leads to
428 more efficient purging of deleterious mutations we also expected deleterious mutations to be preferentially
429 located in areas of low recombination (S19 Fig). As expected, the GLM revealed that the occurrence of
430 deleterious mutations decreased as recombination rate increased ($\chi^2 = 4.90$, $DF = 1$, $p = 0.027$). This effect
431 was stronger when considering non-synonymous variants instead of putatively deleterious variants ($\chi^2 =$
432 10.07 , $DF = 1$, $p = 0.0015$). Given the negative correlation of recombination with chromosome length, we also
433 found a positive correlation ($r = 0.76$, $p < 0.0001$) between the chromosome length and the number of

434 deleterious variants by chromosomes, as expected when purging is more efficient in areas of higher
435 recombination (S20 Fig). To further examine this, we classified regions by recombination rate into high (ρ
436 > 4.516 , see methods), low ($\rho < 1.411$) and intermediate classes and found that there was a significant
437 excess of candidate deleterious mutations in areas of low recombination and depletion of candidate
438 deleterious mutations in areas of high recombination ($\chi^2 = 13.33$, DF= 2, $p = 0.0012$).

439 **Discussion**

440

441 Coho salmon is an emblematic fish species that has undergone population declines in recent decades
442 throughout its North American range. We generated one of the largest collections of GBS and WGS data to
443 date for a non-model vertebrate species, which revealed: *i*) a complex demographic history involving
444 population splits, gene flow, and secondary contacts; *ii*) linked selection and Hill-Robertson interference
445 shaped genetic diversity along the genome, and *iii*) this demographic history has resulted in modest yet
446 detectable differences in the frequency of deleterious variants across regions and populations, which also
447 varied as a function of recombination rate along the genome. Together, these observations help illuminating
448 the drivers of variation in genetic diversity throughout the genome of a broadly distributed species of
449 economic and cultural importance.

450

451 **Expansion from a major southern refugium and secondary contact with micro-refugia**

452 Our results revealed the existence of a major ancestral refugium located south of the ice sheets in Cascadia
453 (i.e., Washington and Oregon) where contemporary populations contain most of the genomic diversity
454 present over the entire North American range of the species. This conclusion was supported by observations
455 of: *i*) a pronounced south-north gradual decrease of genetic diversity, singleton density, and private
456 polymorphism, *ii*) a pattern of IBD, and *iii*) the occurrence of ancestral populations south of formerly
457 glaciated areas. Therefore, although the so-called Beringian refugium that persisted north of the ice sheet
458 (mainly comprising Alaska and the Yukon Territory) was important for many temperate species (6), this was
459 not the case for Coho salmon. The gradual northward decrease in diversity also suggests that populations
460 subsequently expanded from Cascadia to British-Columbia and Alaska [4,5] and is indicative of a serial
461 founder effects due to small proportions of individuals issued from ancestral populations colonising new
462 locations, a process amply documented in humans (e.g., [61]). This expansion was likely postglacial,
463 although GBS-based parameter estimates of divergence time under AM, IM, and SC (~ 30 to ~ 45 KyA)
464 were not in full agreement with SMC++ estimates using WGS (~ 18 KyA). This, however, is expected given
465 the different assumptions made by the various models compared and that gene flow is expected to delay
466 divergence [62]. The last glacial period (Wisconsin Glaciation) which lasted from ~ 120 Ky to ~ 11 KyA was
467 interrupted by an interglacial period from 55 to 25 KyA ([40]). Therefore, our results indicate that neither
468 Beringia nor Haida Gwaii were refugia before the onset of the Wisconsin Glaciation. However, it is possible
469 that individuals from Cascadia already colonized and subsequently diverged in Alaska (Beringia) 55 to 25
470 KyA. If this was the case, however, the contribution of Beringia is likely minor as no strong footprint of
471 ancestral diversity and no signal of secondary contact was inferred in this area.

472 While a single major southern refugium hypothesis is well supported by our data, our analyses also
473 revealed patterns consistent with minor contributions of micro-refugia on extent genomic diversity. Indeed,
474 our demographic modelling supported a model of postglacial (10-20 KyA, S6 Table) secondary contact (SC)
475 detected in several pairwise comparisons among populations between California and Haida Gwaii ($n = 10$
476 pairwise comparisons), Cascadia and Haida Gwaii ($n = 2$), California and Thompson R. ($n = 11$), and
477 between Cascadia and Thompson R. ($n = 1$). These results are counter-intuitive, because contacts between
478 northern sites and California, but bypassing Cascadia, seem unlikely. A possible explanation is that our
479 statistical power to detect SC involving putative micro-refugia was reduced. Here, indeed, the SC period
480 represented a large proportion (22% on average) of the total divergence time across all models. However,
481 models of SC can easily be confounded with models of isolation with migration when SC represents a large
482 period of time ($> 10\%$) relative to the total period of divergence, as observed in our investigations [19,63].
483 Still, our inference of SC supports the hypothesis that smaller micro-refugia have persisted along the Pacific
484 Coast [43,64]. In particular, the Haida Gwaii archipelago is known for high endemism, and its role as a
485 refugium for mammals, invertebrates, and angiosperms is well established [64–66]. The divergence time
486 inferred between populations from California and Haida Gwaii was relatively recent (39 KyA), in the range
487 of inferred divergence times for bird species from the region [67] and in the time frame of the last
488 interglacial period 55 to 25 KyA. Our results also indicated a pronounced divergence between California and
489 Cascadia, suggesting that these were likely two separated refugia, thus supporting the hypothesis of “refugia
490 within refugia” [43,67]. However, California populations have suffered from strong census size reductions in
491 recent decades [68], possibly increasing genetic structure and lowering effective population size. Given our
492 estimates of split times and population sizes, the possible refugium in Haida Gwaii, and California were
493 demographically small and relatively recent, such that populations in these areas were unlikely to have
494 accumulated significant endemic genetic diversity. The case of the Thompson River, on the other hand, is
495 rather intriguing as this region was entirely covered by ice during the Pleistocene. A possible explanation is
496 that a strong and continuous bottleneck has led to a false signal of SC. The SC was the most parameterized
497 model and was therefore more likely to be falsely supported. A model including more drastic and continuous
498 post-divergence changes in population size may fit the data well but would likely be difficult to statistically
499 separate from a model of secondary contact. British-Columbia populations, located at intermediate latitudes,
500 also displayed intermediate levels of genetic diversity, with some populations displaying similar or higher
501 diversity than those located in Cascadia, further supporting the hypothesis of post-colonization admixture
502 between different refugia [69] Complex history of divergence in different refugia and possible mixing was
503 also inferred in the Chinook salmon along the Pacific Coast and was associated to different migratory
504 ecotypes [70]. A potential limitations of our current approach is that we did not incorporate samples from
505 Asia, where the Coho salmon also occurs [41]. Whether this would change our inferences remains an open
506 question. In summary, our modelling approach revealed the presence of multiple cryptic refugia that would
507 otherwise have been missed because their endemic genetic diversity appears to have been largely wiped out
508 by a major demographic expansion out-of-Cascadia, which has clearly been the primary contributor of extent

509 genomic diversity in North American Coho salmon. Knowledge of this non-equilibrium demography will
510 important to help interpret adaptive and deleterious genomics landscape in future studies.

511

512 **Selection at linked sites shape genome wide variation**

513 We also explored the role of gene flow and linked selection in generating a heterogeneous differentiation
514 landscape along the genome, a hotly debated topic in evolutionary genomics [8,9,71,72]. Disentangling the
515 two processes is of fundamental importance for correctly interpreting the origin of genomic islands of
516 divergence and genome scan results [73,74]. Although a positive relationship between absolute and relative
517 divergence may reveal the presence of barriers to gene flow [9], this is not relevant for early stages of
518 divergence [75], as in the present study. Instead, our modelling approach best supported a role of both gene
519 flow and linked selection. In particular, the role of linked selection is supported by the positive correlations
520 between π or D_{XY} and recombination rate, while F_{ST} was negatively correlated with recombination rate
521 [73,75]. The negative correlations between π or D_{XY} and gene density provided further evidence for the role
522 of linked selection [17]. Here, heterogeneous genomic divergence did not arise after speciation, as suggested
523 by Cruickshank & Hahn [9]. Instead, our within-lineage study indicated that linked selection arose within
524 structured populations, in line with recent findings in birds [73]. Similar effects of linked selection were
525 suggested between the more diverged ancestral lineages within Atlantic salmon (>1 MyA) [76]. Given that
526 correlated genomic landscapes have been reported among species diverged for over tens of millions of years
527 [77,78], it would be interesting to investigate how linked selection has also shaped genomic landscapes
528 within the entire radiation of salmonid fishes. Indeed, salmonid species have undergone a whole genome
529 duplication approximately 90 MyA [79] affecting recombination throughout their genome [57,80,81]. Precise
530 mapping of the genomic locations of duplicated regions will allow an improved understanding of how
531 recombination rate varies in these regions although we do not expect that will affect our main conclusion
532 related to linked selection. Salmonid genomes are also characterised by pronounced male heterochiasmy
533 [82,83] and here, we observed a significant correlation between recombination and chromosome length,
534 indicative of crossover interference [84]. Both heterochiasmy and crossover interference contribute to
535 heterogeneity in recombination rates and likely favour the effect of linked selection. Given that most non-
536 neutral mutations are deleterious [85], and given the impact of recombination across the genomes of fishes,
537 models of background selection could be the null model against which to test adaptive hypotheses [21]. Our
538 data revealed that the GC3 was a good proxy of recombination and that it was strikingly well correlated with
539 our proxy for the efficacy of natural selection (π_N/π_S). As expected, this metric revealed differences in the
540 most bottlenecked populations from the Thompson River drainage, where the severity of the bottleneck
541 mimics the effect of domestication bottleneck (e.g. [86]). Indeed, the Thompson R. populations displayed a
542 higher burden of non-synonymous mutations, and we suggest that the strong bottlenecks in these populations
543 had a dramatic impact on the efficacy of natural selection. When considering only genomic regions of low
544 recombination, however, we also found that some California populations also displayed an increased π_N/π_S
545 ratio. Given that these populations have recently undergone large declines in abundance [68], we expected a
546 lower efficacy of purifying selection in regions of low recombination where Hill-Robertson interference

547 should be high [82]. Furthermore, we suggest that the severe and recent demographic declines documented in
548 California populations compared to more long term and putatively less severe declines in the Thompson R.
549 may explain why differences were detected only in genomic regions of low recombination in the former but
550 genome-wide in the latter. Regardless, the non-equilibrium demography, and role of linked selection, in
551 particular background selection, are predicted to favour the accumulation of deleterious mutations, an
552 important finding to manage declining species.

553 **Accumulation of deleterious mutations under complex demography**

554 In the literature, contradictory results have been reported regarding the role of demographic history on the
555 load of deleterious mutations [10,33]. On the one hand, processes such as strong and repeated bottlenecks
556 [24,32], including domestication [86,88], large expansions [89,90] or postglacial colonization [91] can all
557 increase the deleterious load. On the other hand, empirical studies in human indicate that recent demography
558 should not have a strong impact on the load of deleterious variants over the long term [10]. However, other
559 studies indicate that strongly deleterious variants affecting fitness can still display increased in frequency due
560 to demographic growth [10]. Here, the inferred demographic history, together with support for linked
561 selection and gBGC, provides ideal conditions to favour the accumulation of deleterious mutations. In
562 particular, we found that populations displaying smaller effective population size (e.g. California), and those
563 at the extreme of the expansion front (Alaska), displayed deleterious mutations at higher derived allele
564 frequencies, and more frequently in homozygous state, in line with the nearly neutral theory. Similar findings
565 have been observed in domesticated species [86] and recently in Isle Royale wolves [26] where decreased
566 population sizes and inbreeding increased the frequency of deleterious recessive mutations. Overall, these
567 results are consistent with recent empirical findings in which small populations or populations at the
568 expansion front carry more homozygous derived deleterious mutations [26,27,86]. These deleterious
569 mutations in homozygous state are expected to be purged by purifying selection [23,92]. Populations from
570 Cascadia, with higher effective population sizes, contain a higher number of putatively deleterious variants
571 in heterozygous state, as expected from population genetics theory. We also found a nearly linear relationship
572 between the number of derived deleterious mutations in homozygous states and geographic distance from the
573 putative refugial source in Cascadia. This relationship mirrors the findings pertaining to the 'out-of-Africa'
574 expansion of human populations [33], although the maximum geographic distance in our study is an order of
575 magnitude smaller than in the human studies, and that the presence of multiple refugia (as opposed to the
576 sole African ancestral origin for humans) may have contributed to obscure somewhat the relationship.
577 Moreover, pronounced genetic drift or bottlenecks observed in some populations (e.g. Thompson R. and
578 California) may have contributed to reduce the efficacy of selection [24]. Finally, the lower prevalence of
579 deleterious mutation in fish from British-Columbia might also be explained by post-glacial admixture. In
580 some small populations it is possible that these mutation initially reached fixation but subsequently became
581 masked in heterozygous state due to gene-flow and introgression [93]. Similar effect could be due to
582 artificial supplementation programs, but the origin of source populations and intensity of stocking in our
583 study populations are poorly documented. Finally, we observed that recombination was significantly
584 correlated with the proportion of non-synonymous mutations and with the load of deleterious mutations, with

585 more deleterious mutations being found in genomic regions of low recombination. This is in line with
586 theoretical predictions (e.g. Hill-Robertson Interference discussed above), and also empirical studies in other
587 species [94,95]. Overall, our results suggest that Coho salmon populations with highly reduced population
588 sizes are exposed to higher inbreeding depression – a prediction with major conservation implications and
589 which should be investigated further in future studies.

590 **Conclusion**

591 In this study, we presented a rare combined assessment of the relative role of complex demographic history
592 (investigated both by empirical genomic data and modelling) and intrinsic genomic factors (recombination,
593 linked selection) in shaping drivers of the genomic landscape of a broadly distributed species. Complex
594 demographic processes including population expansion, isolation, and secondary contact were revealed
595 through the use of an extensive modelling framework. Moreover, our results highlighted the necessity of
596 accounting for local variation in recombination rate, a key driver of linked selection. Altogether, these
597 processes influence the efficacy of selection and can favour the accumulation of mutations affecting fitness.
598 Our findings suggest that such approaches offer enormous potential in the field of conservation genomics to
599 disentangle the impacts of historical vs. recent drivers of demographic declines and for assessing the
600 distribution of not only putatively beneficial, but also deleterious variants. We propose that future studies
601 should also integrate in-depth analysis of selective sweeps which will be necessary to investigate into more
602 details how linked selection, through background selection and hitchhiking, acts to maintain deleterious
603 mutations in the genome [96]. Finally, while it has become routine with new genome-wide datasets to focus
604 on the effect of positive selection and documenting patterns of local adaptation [7], this focus ignores the
605 fundamental prediction that most new mutations are likely to be deleterious [85]. An increased focus on
606 deleterious mutations would provide a more nuanced view of genomic evolution in wild species which
607 would benefit both the fields of evolutionary and conservation genetics

608

609 **Methods**

610 ***Genotyping By Sequencing***

611 A total of 2,088 individuals was collected from 58 sample sites located along the Pacific coast from
612 California to Alaska (S1 Table and Figure 1). DNA was extracted from all individuals and sequenced using a
613 GBS method (protocol detailed in [97]). Reads were aligned to the Coho salmon reference genome v1
614 (GCF_002021745.1) using bwa-mem 0.7.13 [102]. Samtools v1.7 was used to keep reads with a
615 mapping quality above 20, remove supplementary alignment and unmapped read. Variants were then
616 called with Stacks v1.46 [98]. To do so, the module “pstacks” was used with a minimum depth of 5, and up
617 to three mismatches were allowed in catalog assembly. The module “populations” was run to produce a vcf
618 file that was filtered with a custom python script. We performed stringent filtering to remove SNPs that were
619 1) genotyped in less than 60% of the individuals; 2) at a mean depth of sequencing below 7, and 3) with
620 observed heterozygosity above 0.60, thus resulting in 93,000 SNPs. The pipeline for SNP calling is available
621 on github at https://github.com/enormandeu/stacks_workflow/releases/tag/coho_demography_paper. Next,
622 we removed any individuals with more than 5% missing data and finally only kept SNPs present in at least
623 95% of the individuals yielding a total of 82,772 filtered SNPs for 1,957 individuals. Remaining filtration
624 was done according to the requirement of each analysis performed below.

625 ***Genetic diversity and ancestral populations***

626 For each sampling location we estimated the observed heterozygosity and π using vcftools 0.1.16 [99] and
627 hierfstat [100]. The most likely ancestral populations were identified using β_{ST} [46]. A total of 1,000
628 bootstraps was performed to obtain the 95% confidence intervals around the β_{ST} . Weir and Cockerham's F_{ST}
629 estimator θ [47] was computed in vcftools. We measured the relationship between observed heterozygosity,
630 β_{ST} , F_{ST} and the distance to the southernmost site using linear models. We also verified the relationship
631 between F_{ST} and the distance to the southernmost site using Mantel tests with 10,000 permutations. Vcftools
632 was also used to identify singletons (i.e. variants present in one single individual across the whole dataset).
633 Their distributions were then summed in each locality. We then computed the averaged (min, max and
634 median) number of singletons at the regional level. The scripts are available on github at [https://github.com/](https://github.com/QuentinRougemont/utility_scripts)
635 [QuentinRougemont/utility_scripts](https://github.com/QuentinRougemont/utility_scripts)

636

637 ***Population structure, admixture and gene flow***

638 Levels of ancestry and admixture proportions were inferred with the snmf function implemented in the R
639 package LEA [101]. We allowed less than 5% of missing data. We then kept a set of SNPs in approximate
640 linkage equilibrium by removing pairs of variants with r^2 greater than 0.2 (option --indep-pairwise 50 10 0.2)
641 resulting in 40,632 SNPs. K-values ranging between 1 and 60 were tested and cross-validations were
642 obtained using the cross-entropy criterion with 5% of masked genotypes. The default value for the
643 regularization parameter was used to avoid forcing individuals into groups and hence underestimating
644 admixture. Similar results were obtained from Admixture [102] and are not presented here. Genetic
645 relationship among all salmon was assessed using a PCA with the R package ade4 [103] based on the LD-
646 pruned dataset (40,632 SNPs). We used a 1% minor allele frequency (MAF) threshold and allowed less than
647 4% missing data. Formal tests of admixture were performed using Treemix [49] using the LD-pruned dataset
648 of 40,632 SNPs and without any MAF threshold. A MDS was also constructed using plink and plotted with
649 the ggplot2 [105] R package. We ran Treemix allowing up to 20 migration events and performed 500
650 bootstraps replicates of the best model to obtain robustness of the nodes. The “best” model was inferred after
651 inspecting the relevant migration edges by measuring the percentage of variance explained as migration edge
652 were added to the tree as well as by assessing the p-value associated to each migration edge. A total 500
653 bootstraps replicate was performed under the “best” model and under a model without migration to infer the
654 robustness of the nodes. The scripts are available on github at
655 https://github.com/QuentinRougemont/treemix_workflow

656

657 ***Explicit demographic inferences.***

658 We tested alternative hypotheses of secondary contact between major regional groups (Haida Gwaii,
659 California, Cascadia (Washington-Oregon), Alaska, British Columbia, Thompson R.) of populations by
660 comparing alternative divergence scenarios represent in Fig S12 and initially described in [19,76].
661 Alternative hypotheses of secondary contacts were tested between major groups by testing the significance
662 of alternative divergence scenarios. The four major models tested included a model of Secondary Contact

663 (SC), a model of Strict Isolation (SI), a model of Ancient Migration (AM) and a model of Isolation with
664 Migration (IM).

665 The models shared the following parameters: the ancestral populations of size N_{anc} , splits at time T_{split}
666 into two daughter populations of size N_1 and N_2 . Under the SI model, no gene flow occurs between the two
667 populations. Under AM, gene flow occurred between T_{split} and T_{am} and is followed by a period of strict
668 isolation. Under IM, gene flow occurs at a constant rate at each generation between the two populations.
669 Gene flow can be asymmetric, so that two independent migration rates m_{12} (from population 2 to 1) and m_{21}
670 (from population 1 to 2) were modeled. Under the SC model, the population evolved in strict isolation
671 between T_{split} and until T_{sc} where a secondary contact occurs continuously up to present time. Gene flow is
672 modeled as $M = 2N_{ref}m$. In $\partial a \partial i$, heterogeneous introgression was modeled using two categories of loci
673 occurring in proportions P (i.e., loci with a migration rates M_{12} and M_{21}) and $1-P$ (i.e., loci with a reduced
674 effective migration rates Me_{12} and Me_{21}) across the genome. The same procedure was used to account for
675 linked selection by defining two categories of loci with varying effective population sizes (proportion Q of
676 loci with a “neutral N_e ” and a proportion $1-Q$ of loci with a reduced effective population size due to either
677 linked or background selection). To quantify how linked selection affects reduced N_e , we used a Hill-
678 Robertson scaling factor (H_{rf}) to relate the effective population size of loci influenced by selection ($N_r = H_{rf}$
679 $\cdot N_e$) to that of neutral loci (N_e).

680 Models were fitted using the diffusion theory implemented in $\partial a \partial i$ [50] and also includes the effect of linked
681 selection and barriers to gene flow as detailed in [19,104]. $\partial a \partial i$ uses the SFS as a summary of the data. For a
682 given demographic model, the SFS is computed using diffusion approximation and compared to the
683 empirical SFS using AIC. Here, we started from the whole file containing 200,000 SNPs and used one single
684 SNP per GBS locus, filtered the data to minimize missing data. No MAF was used and singletons were kept
685 to obtain ascertainment-free estimates of demographic parameters. Ideally, no Hardy-Weinberg Equilibrium
686 (HWE) filter should be used for demographic inferences, as this also biases the distribution of allele
687 frequencies. However, to remove paralogs present in the Coho salmon genome, a permissive HWE filter
688 based on a p-value of 0.0001 was used. Here, a total of 69 pairwise comparisons between populations from
689 the major regional groups was performed in order to test for a prevailing pattern. Here, the regional groups
690 considered were all previously unglaciated areas during the LGM, namely California, Cascadia, Alaska
691 (although some samples were likely under ice at different time periods) and Haida Gwaii. For each model, 20
692 independent replicate runs were performed and only models with the lowest AIC and ΔAIC were kept. A
693 model was classified as “ambiguous” and not used for parameter estimation if ΔAIC between the best model
694 and second-best model was below 10. The whole pipeline is available at
695 <https://github.com/QuentinRougemont/DemographicInference>.

696

697 ***Analyses based on whole genome resequencing data.***

698 We used 55 individuals representing 11 populations from California to Alaska (S2 Table). Each individual
699 was sequenced on an Illumina platform using paired-end 150 bp reads. Reads were processed using fastp for
700 trimming [105], bwa mem v0.7.13 [106] for mapping, samtools v1.7 requiring a minimum quality of 10, and

701 picard to remove duplicates (<http://broadinstitute.github.io/picard/>). Then SNP calling was performed using
702 GATK [107]. Genotypes were filtered for depth between 10 and 100 reads to remove low confidence
703 genotypes including potential paralogs, displaying high coverage. Then following GATK Best Practices we
704 excluded all sites that did not match the following criterion: $MQ < 30$, $QD < 2$, $FS > 60$, $MQRankSum < -20$,
705 $ReadPosRankSum < 10$, $ReadPosRankSum > 10$. We also generated a vcf file using the samtools mpileup
706 pipeline, merging individuals with bcftools and performing the same stringent filtering as with the vcf
707 constructed with GATK. Finally, we also generated a separate vcf file using the emit-all-site option to call
708 variable and invariable sites across the whole genome. This vcf file was used in the sliding windows analysis
709 below to test the effect of linked selection. The whole pipeline is available on github
710 (https://github.com/QuentinRougemont/gatk_haplotype).

711 SMC++ [53] was used to infer changes in population size through time. SMC++ works similarly to
712 the PSMC model but takes advantage of information contained in the Site Frequency Spectrum (SFS) in
713 addition to linkage disequilibrium, and is particularly well suited to analyse large sample sizes. Estimates of
714 population size changes were performed for all 11 populations. Splitting time was estimated between all
715 pairs of samples based on the joint SFS. A generation time of 3.5 years and a mutation rate of $8e^{-9}$
716 mutation/bp/generation were applied. The pipeline to reproduce the analysis is available on github
717 (https://github.com/QuentinRougemont/smcpp_input)

718 We computed pairwise linkage disequilibrium using vcftools with the r^2 statistics calculated in all 11
719 populations separately. A window of 1,000,000 base pairs was used and all SNPs were included. To reduce
720 the number of SNPs, we allowed no missing data and used a MAF of 10% and a p-value of Hardy-Weinberg
721 disequilibrium of 0.05 in each population, keeping between 2 and 4 million SNPs depending on the
722 population. We then estimated LD decay by plotting LD against physical distance measured in base pairs and
723 using smoothing functions implemented in ggplot2 [108] package in R. vcftools was also used to identify
724 singletons, for which the distribution were counted by localities, as for the GBS dataset.

725 We used LDhat software [109] to estimate effective recombination rates ($\rho=4.Ne.r$ where r
726 represents the recombination rate per generation and Ne is the effective population size) along the genome.
727 Unphased genotypes were converted into LDhat format using vcftools with a minimum MAF of 10% since
728 only common variants are useful for such inferences [109]. Following the authors' guidelines, the genome
729 was split in chunks of 2,000 SNPs with overlapping windows of 500 SNPs to compute recombination rate
730 and data were then merged together. We measured recombination rates for each river as well as globally
731 including all populations except the population from the Thompson R. watershed that was too divergent from
732 the remaining samples. The pipeline to reproduce the analysis is available on github
733 (https://github.com/QuentinRougemont/LDhat_workflow).

734

735 ***Clarifying the role of linked selection***

736 The genetic landscape of divergence was measured by estimating levels of nucleotide diversity (π), gene
737 density, levels of genetic differentiation (F_{ST}), levels of divergence (D_{xy}), and scaled recombination rate
738 along the genome. Estimates of π , F_{ST} , and D_{xy} were computed into 500kb windows using Python scripts

739 available from [110]. Gene density was computed directly from the gff file of the Coho salmon genome v1.0
740 (NCBI ftp file: ftp://ftp-trace.ncbi.nih.gov/genomes/Oncorhynchus_kisutch/GFF/; Rondeau et al. In prep)
741 and measured in 500 kb windows. Recombination rates were averaged into 250 and 500-kb windows using a
742 Python script after being estimated with LDhat as described above. We used a PCA to obtain a synthetic
743 view across all 11 π estimates, and all pairwise 55 Dxy and 55 F_{ST} separately. This allowed capturing the
744 common variation affecting these three estimates. First, simple Spearman correlations based on linear models
745 testing each (z-transformed) variable projected on the PC1 axis separately were carried out to produce fig 5.
746 Then Mixed Linear Models were used to test for correlations between either π , Dxy, or F_{ST} and either
747 recombination landscape (ρ) or gene density windows as explanatory variables, with and without
748 interaction. Correlations were also calculated considering each π , Dxy, and F_{ST} by population separately
749 without PCA transformation (S9 Table) which returned patterns that were congruent with those observed
750 with PCA results.

751

752 **Measuring GC content, Non-Synonymous and Synonymous diversity**

753 The pipeline developed by [111] was used to compute Tajima's π estimator of nucleotide diversity and the
754 GC content over non-overlapping 10-kb windows. Then we concatenated the gene into different classes
755 according to their length and compute π_N and π_S over 4 mb windows, which allows circumventing the
756 problem associated to low π_S values. We then verified the correlation between populations scaled
757 recombination rate (ρ), measured over 250-kb windows and GC at the third codon position (approximately
758 neutral). To do so, we assigned a ρ value for each gene according to its position into each 250-kb windows.
759 Then we averaged ρ values over the same length class as GC3 classes and compared the median population
760 recombination rate estimates and GC3s over 4-mb windows. Finally, we measured the correlation between
761 GC3s and π_N/π_S using linear models.

762

763 **Genetic load estimation from the GBS data set**

764 *Estimating ancestral and derived alleles*

765 We used the derived allele count as an estimator of deleterious allele count. We used the genomes of three
766 outgroup species, the chinook salmon, the rainbow trout, and the Atlantic salmon, to classify SNPs as
767 ancestral or derived. Whole genome data for the chinook salmon ($n = 3$ individuals) were provided by one of
768 us (B. Koop, unpublished), whereas rainbow trout ($n = 5$) and Atlantic salmon ($n = 5$) data were downloaded
769 from NCBI Sequence Read Archive (rainbow trout, SRA, Bioproject : SRP117091; *Salmo salar* SRA
770 Bioproject: SRP059652). Each individual was aligned against the Coho salmon V1 reference genome
771 (GCF_002021745.1) using GATK UnifiedGenotyper and calling every SNP using the EMIT_ALL_SITES
772 modes. We then constructed a Python script and determined the ancestral state of the GBS SNPs if 1) the
773 SNP was homozygous in at least two of the three outgroups, and 2) match one of the two alleles identified in
774 Coho salmon. Otherwise, the site was inferred as missing and not used in subsequent analyses.

775 *Measuring damaging impact of non-synonymous alleles*

776 We tested differences in mutation load among populations as follows. The software Provean [112] was used
777 to predict whether a given non-synonymous mutation was deleterious with a threshold score of -2.5 or less
778 using the pipeline available at https://github.com/QuentinRougemont/gbs_synonymy_with_genome. We
779 analysed data in two ways: first we counted the total number of putative homozygous deleterious alleles per
780 individual as well as the total number of deleterious alleles (both in homozygous and heterozygous states)
781 using: $N_{total} = 2 \times N_{homo} + N_{hetero}$ [34]. These individual values were then averaged per population and major
782 regional groups (i.e., California, Cascadia, British Columbia, Haida Gwaii, Thompson, and Alaska). We then
783 computed derived allele frequencies (DAF) in all sampling locations and across mutation categories
784 (synonymous, non-synonymous, and non-synonymous deleterious) and tested for significant allele frequency
785 differences among populations in non-synonymous and non-synonymous deleterious mutations using
786 Wilcoxon rank sum tests. DAF spectra were constructed for all population separately. For the ease of
787 visualisation, we also constructed DAF spectra by region for a sample of size $n = 100$ individuals (Fig 7A).
788 This size was chosen according to the smallest sample size of the three combined Haida Gwaii populations.
789 Finally, we tested for a preferential enrichment of “hotspots or coldspots” of recombination in deleterious
790 mutations. To define a coldspot, we first computed a lower bound that we defined as the $mean_{RHO} -$
791 $5 \times \text{standard errors}$ for each chromosome separately. Similarly, “hotspots” of recombination were identified
792 using an upper bound defined as $mean_{RHO} + 5 \text{ standard errors}$. We then tested if the average recombination rate
793 of each of the 250 kb windows was falling below the threshold (for coldspot) or above it (for hotspots). We
794 then tested if recombination hotspots and coldspots contained more or less putatively deleterious mutations
795 than “normally” recombining regions using χ^2 tests. Linear mixed effects models were further used to test if
796 there was a relationship between recombination rate and the distribution of putatively deleterious mutations.
797 The response variable was the deleterious state considered binomial (0 = non deleterious, 1 = deleterious)
798 and the explanatory variable was the recombination rate. The chromosome identity was included as a random
799 effect. This model was compared against a null model excluding recombination rate. The analysis was
800 replicated but using the synonymous and non-synonymous state as the response variable instead of the
801 putatively deleterious state of the considered mutation. Models were carried out in R using the LME4
802 package [113]. Finally, we used SnpEff v3.4 [114] to obtain the functional annotations of the putatively
803 deleterious variants. The annotations were comprised of mis-sense variants, non-coding transcripts, 3' and 5'
804 untranslated regions, 5kb up- and down-stream variants, intergenic and intronic, splice acceptor and splice
805 region, stop gained and start loss (S14 Table). We found 35% of deleterious mutations to be missense
806 variants, 38% to be non-coding transcript and 22% to be either upstream or downstream gene variants, with
807 the remaining being spread over the categories.

808 **Ethical Statement**

809 A permit number SIRUL 111722 was obtained to work on DNA sequences.

810

811 Raw data will be deposited on NCBI together with Short Read Archive (SRA) accession number.

812 **Acknowledgements:**

813 We thank B. Bougas, A. Perrault-Payette, C. Hernandez for laboratory support and K. Wellband, H. Cayuela,
814 F. Hartmann for their constructive comments. TL and QR thank Benoit Nabholz for discussions regarding the
815 deleterious mutation loads and Hill-Robertson effects. Computations were performed on Colosse (Calcul
816 Quebec), Graham and Cedar (Compute Canada) servers. This research was carried out in conjunction with
817 EPIC4 (Enhanced Production in Coho: Culture, Community, Catch), a project supported by the government
818 of Canada through Genome Canada, Genome British Columbia, and Genome Quebec. The authors declare
819 no conflicts of interest.

820

821 **References:**

1. Allendorf FW. Evolution in a Toxic World. *BioScience*. 2017;67: 576–577. doi:10.1093/biosci/bix029
2. Ceballos G, Ehrlich PR, Dirzo R. Biological annihilation via the ongoing sixth mass extinction signaled by vertebrate population losses and declines. *Proc Natl Acad Sci*. 2017;114: E6089–E6096. doi:10.1073/pnas.1704949114
3. Provan J, Bennett KD. Phylogeographic insights into cryptic glacial refugia. *Trends Ecol Evol*. 2008;23: 564–571. doi:10.1016/j.tree.2008.06.010
4. Hewitt GM. Post-glacial re-colonization of European biota. *Biol J Linn Soc*. 1999;68: 87–112. doi:10.1006/bijl.1999.0332
5. Bernatchez L, Wilson CC. Comparative phylogeography of Nearctic and Palearctic fishes. *Mol Ecol*. 1998;7: 431–452. doi:10.1046/j.1365-294x.1998.00319.x
6. Frankham R, Ballou JD, Briscoe DA. Introduction to Conservation Genetics by Richard Frankham. In: Cambridge Core [Internet]. Jan 2010 [cited 2 Jul 2019]. doi:10.1017/CBO9780511809002
7. Funk WC, McKay JK, Hohenlohe PA, Allendorf FW. Harnessing genomics for delineating conservation units. *Trends Ecol Evol*. 2012;27: 489–496. doi:10.1016/j.tree.2012.05.012
8. Noor M a. F, Bennett SM. Islands of speciation or mirages in the desert? Examining the role of restricted recombination in maintaining species. *Heredity*. 2009;103: 439–444. doi:10.1038/hdy.2009.151
9. Cruickshank TE, Hahn MW. Reanalysis suggests that genomic islands of speciation are due to reduced diversity, not reduced gene flow. *Mol Ecol*. 2014;23: 3133–3157. doi:10.1111/mec.12796
10. Simons YB, Turchin MC, Pritchard JK, Sella G. The deleterious mutation load is insensitive to recent population history. *Nat Genet*. 2014;46: 220–224. doi:10.1038/ng.2896
11. Bierne N, Gagnaire P-A, David P. The geography of introgression in a patchy environment and the thorn in the side of ecological speciation. *Curr Zool*. 2013;59: 72–86. doi:10.1093/czoolo/59.1.72
12. Barton N, Bengtsson BO. The barrier to genetic exchange between hybridising populations. *Heredity*. 1986;57: 357. doi:10.1038/hdy.1986.135

13. Smith JM, Haigh J. The hitch-hiking effect of a favourable gene. *Genet Res.* 1974;23: 23–35.
14. Charlesworth B, Morgan MT, Charlesworth D. The effect of deleterious mutations on neutral molecular variation. *Genetics.* 1993;134: 1289–1303.
15. Kaplan NL, Hudson RR, Langley CH. The “hitchhiking effect” revisited. *Genetics.* 1989;123: 887–899.
16. Nordborg M, Charlesworth B, Charlesworth D. The effect of recombination on background selection. *Genet Res.* 1996;67: 159–174.
17. Payseur BA, Nachman MW. Natural selection at linked sites in humans. *Gene.* 2002;300: 31–42.
18. Hudson RR, Kaplan NL. Deleterious Background Selection with Recombination. *Genetics.* 1995;141: 1605–1617.
19. Roux C, Fraïsse C, Romiguier J, Anciaux Y, Galtier N, Bierne N. Shedding Light on the Grey Zone of Speciation along a Continuum of Genomic Divergence. *PLOS Biol.* 2016;14: e2000234. doi:10.1371/journal.pbio.2000234
20. Pouyet F, Aeschbacher S, Thiéry A, Excoffier L. Background selection and biased gene conversion affect more than 95% of the human genome and bias demographic inferences. Veeramah K, Wittkopp PJ, Gronau I, editors. *eLife.* 2018;7: e36317. doi:10.7554/eLife.36317
21. Comeron JM. Background selection as null hypothesis in population genomics: insights and challenges from *Drosophila* studies. *Philos Trans R Soc Lond B Biol Sci.* 2017;372. doi:10.1098/rstb.2016.0471
22. Simons YB, Sella G. The impact of recent population history on the deleterious mutation load in humans and close evolutionary relatives. *Curr Opin Genet Dev.* 2016;41: 150–158. doi:10.1016/j.gde.2016.09.006
23. Charlesworth D, Willis JH. The genetics of inbreeding depression. *Nat Rev Genet.* 2009;10: 783–796. doi:10.1038/nrg2664
24. Kirkpatrick M, Jarne P. The Effects of a Bottleneck on Inbreeding Depression and the Genetic Load. *Am Nat.* 2000;155: 154–167. doi:10.1086/303312
25. Charlesworth B. The Effects of Deleterious Mutations on Evolution at Linked Sites. *Genetics.* 2012;190: 5–22. doi:10.1534/genetics.111.134288
26. Robinson JA, Rääkkönen J, Vucetich LM, Vucetich JA, Peterson RO, Lohmueller KE, et al. Genomic signatures of extensive inbreeding in Isle Royale wolves, a population on the threshold of extinction. *Sci Adv.* 2019;5: eaau0757. doi:10.1126/sciadv.aau0757
27. Robinson JA, Ortega-Del Vecchyo D, Fan Z, Kim BY, vonHoldt BM, Marsden CD, et al. Genomic Flatlining in the Endangered Island Fox. *Curr Biol CB.* 2016;26: 1183–1189. doi:10.1016/j.cub.2016.02.062
28. Abascal F, Corvelo A, Cruz F, Villanueva-Cañas JL, Vlasova A, Marcet-Houben M, et al. Extreme genomic erosion after recurrent demographic bottlenecks in the highly endangered Iberian lynx. *Genome Biol.* 2016;17: 251. doi:10.1186/s13059-016-1090-1

29. Dobrynin P, Liu S, Tamazian G, Xiong Z, Yurchenko AA, Krasheninnikova K, et al. Genomic legacy of the African cheetah, *Acinonyx jubatus*. *Genome Biol.* 2015;16: 277. doi:10.1186/s13059-015-0837-4
30. Yang Y, Ma T, Wang Z, Lu Z, Li Y, Fu C, et al. Genomic effects of population collapse in a critically endangered ironwood tree *Ostrya rehderiana*. *Nat Commun.* 2018;9: 5449. doi:10.1038/s41467-018-07913-4
31. Xue Y, Prado-Martinez J, Sudmant PH, Narasimhan V, Ayub Q, Szpak M, et al. Mountain gorilla genomes reveal the impact of long-term population decline and inbreeding. *Science.* 2015;348: 242–245. doi:10.1126/science.aaa3952
32. Grossen C, Guillaume F, Keller LF, Croll D. Accumulation and purging of deleterious mutations through severe bottlenecks in ibex. *bioRxiv.* 2019; 605147. doi:10.1101/605147
33. Henn BM, Botigué LR, Peischl S, Dupanloup I, Lipatov M, Maples BK, et al. Distance from sub-Saharan Africa predicts mutational load in diverse human genomes. *Proc Natl Acad Sci.* 2016;113: E440–E449. doi:10.1073/pnas.1510805112
34. Henn BM, Botigué LR, Bustamante CD, Clark AG, Gravel S. Estimating the mutation load in human genomes. *Nat Rev Genet.* 2015;16: 333–343. doi:10.1038/nrg3931
35. Krkosek M, Ford JS, Morton A, Lele S, Myers RA, Lewis MA. Declining wild salmon populations in relation to parasites from farm salmon. *Science.* 2007;318: 1772–1775. doi:10.1126/science.1148744
36. Irvine JR, Fukuwaka M. Pacific salmon abundance trends and climate change. *ICES J Mar Sci.* 2011;68: 1122–1130. doi:10.1093/icesjms/fsq199
37. Gustafson RG, Waples RS, Myers JM, Weitkamp LA, Bryant GJ, Johnson OW, et al. Pacific Salmon Extinctions: Quantifying Lost and Remaining Diversity. *Conserv Biol.* 2007;21: 1009–1020. doi:10.1111/j.1523-1739.2007.00693.x
38. Smith CT, Nelson RJ, Wood CC, Koop BF. Glacial biogeography of North American coho salmon (*Oncorhynchus kisutch*). *Mol Ecol.* 2001;10: 2775–2785.
39. Beacham TD, Wetklo M, Deng L, MacConnachie C. Coho Salmon Population Structure in North America Determined from Microsatellites. *Trans Am Fish Soc.* 2011;140: 253–270. doi:10.1080/00028487.2011.558782
40. McPhail JD, Lindsey CC. Freshwater fishes of northwestern Canada and Alaska. Fisheries Research Board of Canada : available by mail from the Queen’s Printer; 1970.
41. UBC Press | Pacific Salmon Life Histories, By Cornelis Groot, Leo Margolis and Leo Margolis. In: UBC Press [Internet]. [cited 1 Jul 2019]. Available: <https://www.ubcpress.ca/pacific-salmon-life-histories>
42. Hocutt CH, Wiley EO, editors. *The Zoogeography of North American Freshwater Fishes.* 1 edition. New York: Wiley-Interscience; 1986.
43. Mee JA, Moore J-S. The ecological and evolutionary implications of microrefugia. *J Biogeogr.* 2014;41: 837–841. doi:10.1111/jbi.12254

44. Warner BG, Mathewes RW, Clague JJ. Ice-free conditions on the queen charlotte islands, british columbia, at the height of late wisconsin glaciation. *Science*. 1982;218: 675–677. doi:10.1126/science.218.4573.675
45. Cubry P, Vigouroux Y, François O. The Empirical Distribution of Singletons for Geographic Samples of DNA Sequences. *Front Genet*. 2017;8. doi:10.3389/fgene.2017.00139
46. Weir BS, Goudet J. A Unified Characterization of Population Structure and Relatedness. *Genetics*. 2017;206: 2085–2103. doi:10.1534/genetics.116.198424
47. Weir BS, Cockerham CC. Estimating F-Statistics for the Analysis of Population Structure. *Evolution*. 1984;38: 1358–1370. doi:10.2307/2408641
48. Ward RD, Woodwark M, Skibinski DOF. A comparison of genetic diversity levels in marine, freshwater, and anadromous fishes. *J Fish Biol*. 1994;44: 213–232. doi:10.1111/j.1095-8649.1994.tb01200.x
49. Pickrell JK, Pritchard JK. Inference of Population Splits and Mixtures from Genome-Wide Allele Frequency Data. *PLOS Genet*. 2012;8: e1002967. doi:10.1371/journal.pgen.1002967
50. Gutenkunst RN, Hernandez RD, Williamson SH, Bustamante CD. Inferring the Joint Demographic History of Multiple Populations from Multidimensional SNP Frequency Data. *PLOS Genet*. 2009;5: e1000695. doi:10.1371/journal.pgen.1000695
51. Roux C, Tsagkogeorga G, Bierne N, Galtier N. Crossing the species barrier: genomic hotspots of introgression between two highly divergent *Ciona intestinalis* species. *Mol Biol Evol*. 2013;30: 1574–1587. doi:10.1093/molbev/mst066
52. COSEWIC assessment and status report on the coho salmon *Oncorhynchus kisutch* (Interior Fraser population) in Canada - Species at Risk Public Registry [Internet]. [cited 1 Jul 2019]. Available: https://wildlife-species.canada.ca/species-risk-registry/document/default_e.cfm?documentID=105
53. Terhorst J, Kamm JA, Song YS. Robust and scalable inference of population history from hundreds of unphased whole genomes. *Nat Genet*. 2017;49: 303–309. doi:10.1038/ng.3748
54. Haenel Q, Laurentino TG, Roesti M, Berner D. Meta-analysis of chromosome-scale crossover rate variation in eukaryotes and its significance to evolutionary genomics. *Mol Ecol*. 2018;27: 2477–2497. doi:10.1111/mec.14699
55. Eyre-Walker Adam. Recombination and mammalian genome evolution. *Proc R Soc Lond B Biol Sci*. 1993;252: 237–243. doi:10.1098/rspb.1993.0071
56. Galtier N, Piganeau G, Mouchiroud D, Duret L. GC-Content Evolution in Mammalian Genomes: The Biased Gene Conversion Hypothesis. *Genetics*. 2001;159: 907–911.
57. Christensen KA, Leong JS, Sakhrani D, Biagi CA, Minkley DR, Withler RE, et al. Chinook salmon (*Oncorhynchus tshawytscha*) genome and transcriptome. *PLOS ONE*. 2018;13: e0195461. doi:10.1371/journal.pone.0195461
58. Gao L, Jia J, Kong X. A SNP-Based Molecular Barcode for Characterization of Common Wheat. *PLOS ONE*. 2016;11: e0150947. doi:10.1371/journal.pone.0150947

59. Yáñez JM, Naswa S, López ME, Bassini L, Correa K, Gilbey J, et al. Genomewide single nucleotide polymorphism discovery in Atlantic salmon (*Salmo salar*): validation in wild and farmed American and European populations. *Mol Ecol Resour.* 2016;16: 1002–1011. doi:10.1111/1755-0998.12503
60. Li JZ, Absher DM, Tang H, Southwick AM, Casto AM, Ramachandran S, et al. Worldwide human relationships inferred from genome-wide patterns of variation. *Science.* 2008;319: 1100–1104. doi:10.1126/science.1153717
61. Wakeley J. *Coalescent Theory: An Introduction.* 1st edition. Greenwood Village, Colo: W. H. Freeman; 2008.
62. Alcala Nicolas, Vuilleumier Séverine. Turnover and accumulation of genetic diversity across large time-scale cycles of isolation and connection of populations. *Proc R Soc B Biol Sci.* 2014;281: 20141369. doi:10.1098/rspb.2014.1369
63. Shafer ABA, Cullingham CI, Côté SD, Coltman DW. Of glaciers and refugia: a decade of study sheds new light on the phylogeography of northwestern North America. *Mol Ecol.* 2010;19: 4589–4621. doi:10.1111/j.1365-294X.2010.04828.x
64. O'Reilly P, Reimchen TE, Beech R, Strobeck C. MITOCHONDRIAL DNA IN GASTEROSTEUS AND PLEISTOCENE GLACIAL REFUGIUM ON THE QUEEN CHARLOTTE ISLANDS, BRITISH COLUMBIA. *Evol Int J Org Evol.* 1993;47: 678–684. doi:10.1111/j.1558-5646.1993.tb02122.x
65. Pruett CL, Topp CM, Maley JM, McCracken K, Rohwer S, Birks S, et al. Evidence from the genetics of landbirds for a forested pleistocene glacial refugium in the haida gwaii area. *Condor.* 2013;115: 725–737. doi:10.1525/cond.2013.120123
66. Geraldès A, Askelson KK, Nikelski E, Doyle FI, Harrower WL, Winker K, et al. Population genomic analyses reveal a highly differentiated and endangered genetic cluster of northern goshawks (*Accipiter gentilis laingi*) in Haida Gwaii. *Evol Appl.* 2019;12: 757–772. doi:10.1111/eva.12754
67. Gómez A, Lunt DH. Refugia within Refugia: Patterns of Phylogeographic Concordance in the Iberian Peninsula. In: Weiss S, Ferrand N, editors. *Phylogeography of Southern European Refugia: Evolutionary perspectives on the origins and conservation of European biodiversity.* Dordrecht: Springer Netherlands; 2007. pp. 155–188. doi:10.1007/1-4020-4904-8_5
68. Brown LR, Moyle PB, Yoshiyama RM. Historical Decline and Current Status of Coho Salmon in California. *North Am J Fish Manag.* 1994;14: 237–261. doi:10.1577/1548-8675(1994)014<0237:HDACSO>2.3.CO;2
69. Petit RJ, Aguinagalde I, Beaulieu J-L de, Bittkau C, Brewer S, Cheddadi R, et al. Glacial Refugia: Hotspots But Not Melting Pots of Genetic Diversity. *Science.* 2003;300: 1563–1565. doi:10.1126/science.1083264
70. Waples RS, Teel DJ, Myers JM, Marshall AR. Life-History Divergence in Chinook Salmon: Historic Contingency and Parallel Evolution. *Evolution.* 2004;58: 386–403. doi:10.1111/j.0014-3820.2004.tb01654.x
71. Burri R. Linked selection, demography and the evolution of correlated genomic landscapes in birds and beyond. *Mol Ecol.* 2017;26: 3853–3856. doi:10.1111/mec.14167

72. Stankowski S, Chase MA, Fuiten AM, Rodrigues MF, Ralph PL, Streisfeld MA. Widespread selection and gene flow shape the genomic landscape during a radiation of monkeyflowers. *bioRxiv*. 2019; 342352. doi:10.1101/342352
73. Burri R, Nater A, Kawakami T, Mugal CF, Olason PI, Smeds L, et al. Linked selection and recombination rate variation drive the evolution of the genomic landscape of differentiation across the speciation continuum of *Ficedula* flycatchers. *Genome Res*. 2015;25: 1656–1665. doi:10.1101/gr.196485.115
74. Ravinet M, Faria R, Butlin RK, Galindo J, Bierne N, Rafajlović M, et al. Interpreting the genomic landscape of speciation: a road map for finding barriers to gene flow. *J Evol Biol*. 2017;30: 1450–1477. doi:10.1111/jeb.13047
75. Burri R. Dissecting differentiation landscapes: a linked selection’s perspective. *J Evol Biol*. 2017;30: 1501–1505. doi:10.1111/jeb.13108
76. Rougemont Q, Bernatchez L. The demographic history of Atlantic salmon (*Salmo salar*) across its distribution range reconstructed from approximate Bayesian computations*. *Evolution*. 2018;72: 1261–1277. doi:10.1111/evo.13486
77. Vijay N, Bossu CM, Poelstra JW, Weissensteiner MH, Suh A, Kryukov AP, et al. Evolution of heterogeneous genome differentiation across multiple contact zones in a crow species complex. *Nat Commun*. 2016;7: 13195. doi:10.1038/ncomms13195
78. Van Doren BM, Campagna L, Helm B, Illera JC, Lovette IJ, Liedvogel M. Correlated patterns of genetic diversity and differentiation across an avian family. *Mol Ecol*. 2017;26: 3982–3997. doi:10.1111/mec.14083
79. Macqueen DJ, Johnston IA. A well-constrained estimate for the timing of the salmonid whole genome duplication reveals major decoupling from species diversification. *Proc R Soc B Biol Sci*. 2014;281. doi:10.1098/rspb.2013.2881
80. Kodama M, Briec MSO, Devlin RH, Hard JJ, Naish KA. Comparative mapping between Coho Salmon (*Oncorhynchus kisutch*) and three other salmonids suggests a role for chromosomal rearrangements in the retention of duplicated regions following a whole genome duplication event. *G3 Bethesda Md*. 2014;4: 1717–1730. doi:10.1534/g3.114.012294
81. Briec MSO, Ono K, Drinan DP, Naish KA. Integration of Random Forest with population-based outlier analyses provides insight on the genomic basis and evolution of run timing in Chinook salmon (*Oncorhynchus tshawytscha*). *Mol Ecol*. 2015;24: 2729–2746. doi:10.1111/mec.13211
82. Lien S, Koop BF, Sandve SR, Miller JR, Kent MP, Nome T, et al. The Atlantic salmon genome provides insights into rediploidization. *Nature*. 2016;533: 200–205. doi:10.1038/nature17164
83. Sutherland BJB, Rico C, Audet C, Bernatchez L. Sex Chromosome Evolution, Heterochiasmy, and Physiological QTL in the Salmonid Brook Charr *Salvelinus fontinalis*. *G3 Bethesda Md*. 2017;7: 2749–2762. doi:10.1534/g3.117.040915
84. Allendorf FW, Bassham S, Cresko WA, Limborg MT, Seeb LW, Seeb JE. Effects of crossovers between homeologs on inheritance and population genomics in polyploid-derived salmonid fishes. *J Hered*. 2015;106: 217–227. doi:10.1093/jhered/esv015

85. Eyre-Walker A, Keightley PD. The distribution of fitness effects of new mutations. *Nat Rev Genet.* 2007;8: 610–618. doi:10.1038/nrg2146
86. Marsden CD, Vecchyo DO-D, O'Brien DP, Taylor JF, Ramirez O, Vilà C, et al. Bottlenecks and selective sweeps during domestication have increased deleterious genetic variation in dogs. *Proc Natl Acad Sci.* 2016;113: 152–157. doi:10.1073/pnas.1512501113
87. Roze D, Barton NH. The Hill-Robertson effect and the evolution of recombination. *Genetics.* 2006;173: 1793–1811. doi:10.1534/genetics.106.058586
88. Zhou Y, Massonnet M, Sanjak JS, Cantu D, Gaut BS. Evolutionary genomics of grape (*Vitis vinifera* ssp. *vinifera*) domestication. *Proc Natl Acad Sci.* 2017;114: 11715–11720. doi:10.1073/pnas.1709257114
89. Peischl S, Dupanloup I, Kirkpatrick M, Excoffier L. On the accumulation of deleterious mutations during range expansions. *Mol Ecol.* 2013;22: 5972–5982. doi:10.1111/mec.12524
90. Peischl S, Dupanloup I, Foucal A, Jomphe M, Bruat V, Grenier J-C, et al. Relaxed Selection During a Recent Human Expansion. *Genetics.* 2018;208: 763–777. doi:10.1534/genetics.117.300551
91. Laenen B, Tedder A, Nowak MD, Toräng P, Wunder J, Wötzel S, et al. Demography and mating system shape the genome-wide impact of purifying selection in *Arabis alpina*. *Proc Natl Acad Sci.* 2018;115: 816–821. doi:10.1073/pnas.1707492115
92. Robinson JA, Brown C, Kim BY, Lohmueller KE, Wayne RK. Purging of Strongly Deleterious Mutations Explains Long-Term Persistence and Absence of Inbreeding Depression in Island Foxes. *Curr Biol CB.* 2018;28: 3487-3494.e4. doi:10.1016/j.cub.2018.08.066
93. Kim BY, Huber CD, Lohmueller KE. Deleterious variation shapes the genomic landscape of introgression. *PLOS Genet.* 2018;14: e1007741. doi:10.1371/journal.pgen.1007741
94. Haddrill PR, Halligan DL, Tomaras D, Charlesworth B. Reduced efficacy of selection in regions of the *Drosophila* genome that lack crossing over. *Genome Biol.* 2007;8: R18. doi:10.1186/gb-2007-8-2-r18
95. Rodgers-Melnick E, Bradbury PJ, Elshire RJ, Glaubitz JC, Acharya CB, Mitchell SE, et al. Recombination in diverse maize is stable, predictable, and associated with genetic load. *Proc Natl Acad Sci.* 2015;112: 3823–3828. doi:10.1073/pnas.1413864112
96. Torres R, Stetter MG, Hernandez RD, Ross-Ibarra J. The temporal dynamics of background selection in non-equilibrium populations. *bioRxiv.* 2019; 618389. doi:10.1101/618389
97. Moore J-S, Harris LN, Le Luyer J, Sutherland BJB, Rougemont Q, Tallman RF, et al. Genomics and telemetry suggest a role for migration harshness in determining overwintering habitat choice, but not gene flow, in anadromous Arctic Char. *Mol Ecol.* 2017;26: 6784–6800. doi:10.1111/mec.14393
98. Catchen JM, Hohenlohe PA, Bernatchez L, Funk WC, Andrews KR, Allendorf FW. Unbroken: RADseq remains a powerful tool for understanding the genetics of adaptation in natural populations. *Mol Ecol Resour.* 2017;17: 362–365. doi:10.1111/1755-0998.12669

99. Danecek P, Auton A, Abecasis G, Albers CA, Banks E, DePristo MA, et al. The variant call format and VCFtools. *Bioinforma Oxf Engl*. 2011;27: 2156–2158. doi:10.1093/bioinformatics/btr330
100. Goudet J. hierfstat, a package for r to compute and test hierarchical F-statistics. *Mol Ecol Notes*. 2005;5: 184–186. doi:10.1111/j.1471-8286.2004.00828.x
101. Frichot E, François O. LEA: An R package for landscape and ecological association studies. *Methods Ecol Evol*. 2015;6: 925–929. doi:10.1111/2041-210X.12382
102. Alexander DH, Novembre J, Lange K. Fast model-based estimation of ancestry in unrelated individuals. *Genome Res*. 2009;19: 1655–1664. doi:10.1101/gr.094052.109
103. Dray S, Dufour A-B. The ade4 Package: Implementing the Duality Diagram for Ecologists. *J Stat Softw*. 2007;22: 1–20.
104. Rougemont Q, Gagnaire P-A, Perrier C, Genthon C, Besnard A-L, Launey S, et al. Inferring the demographic history underlying parallel genomic divergence among pairs of parasitic and nonparasitic lamprey ecotypes. *Mol Ecol*. 2017;26: 142–162. doi:10.1111/mec.13664
105. Chen S, Zhou Y, Chen Y, Gu J. fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinforma Oxf Engl*. 2018;34: i884–i890. doi:10.1093/bioinformatics/bty560
106. Li H. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *ArXiv13033997 Q-Bio*. 2013; Available: <http://arxiv.org/abs/1303.3997>
107. DePristo MA, Banks E, Poplin R, Garimella KV, Maguire JR, Hartl C, et al. A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nat Genet*. 2011;43: 491. doi:10.1038/ng.806
108. Wickham H. ggplot2: Elegant Graphics for Data Analysis [Internet]. New York: Springer-Verlag; 2009. Available: <https://www.springer.com/gp/book/9780387981413>
109. McVean G, Awadalla P, Fearnhead P. A Coalescent-Based Method for Detecting and Estimating Recombination From Gene Sequences. *Genetics*. 2002;160: 1231–1241.
110. Martin S. https://github.com/simonhmartin/genomics_general.
111. Leroy T, Anselmetti Y, Tilak M-K, Bérard S, Csukonyi L, Gabrielli M, et al. A bird’s white-eye view on neosex chromosome evolution. *bioRxiv*. 2019; 505610. doi:10.1101/505610
112. Choi Y, Sims GE, Murphy S, Miller JR, Chan AP. Predicting the functional effect of amino acid substitutions and indels. *PloS One*. 2012;7: e46688. doi:10.1371/journal.pone.0046688
113. Bates D, Mächler M, Bolker B, Walker S. Fitting Linear Mixed-Effects Models Using lme4. *J Stat Softw*. 2015;67: 1–48. doi:10.18637/jss.v067.i01
114. Cingolani P, Platts A, Wang LL, Coon M, Nguyen T, Wang L, et al. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of *Drosophila melanogaster* strain w1118; iso-2; iso-3. *Fly (Austin)*. 2012;6: 80–92. doi:10.4161/fly.19695

115. Government of Canada NRC. GEOSCAN Search Results: Fastlink [Internet]. 7 Dec 2015 [cited 26 Jul 2019]. Available: <https://geoscan.nrcan.gc.ca/starweb/geoscan/servlet.starweb?path=geoscan/fulle.web&search1=R=214399>

822 **Supporting Information Legend**

823

824 **S1 Fig. Linear decrease in genetic diversity when considering π SNP as a function of the distance to the**
825 **southernmost sample site.** Each points represents a sample site and is coloured by region.

826

827 **S2 Fig. Network of shared and private polymorphisms.** The branch (grey) represent shared polymorphism
828 between sample site and are proportional to levels of sharing. Each point represent the number of private
829 polymorphisms and is coloured by region. Computation were based on a sample of size 100 in each region to
830 enable comparison.

831

832 **S3 Fig. Measure of linkage disequilibrium decay using whole genome sequencing data.** LD between
833 pairs of loci was measured as r^2 in windows of 1 million base pairs. Means smoothed value are display for
834 each whole genome samples.

835

836 **S4 Fig. Patterns of Isolation By Distance.** Increasing F_{ST} as a function of the distance to the southernmost
837 site. Each points represents a sample site and is coloured by region.

838

839 **S5 Fig. Summaries of F_{ST} values.**

840 A. F_{ST} -based Hierarchical tree depicting relationship among samples. Colors represent the major region.

841 B. Heatmap of F_{ST} values among samples ordered from North to South on the X and Y-axis.

842

843 **S6 Fig. Structure and Admixture inferences.**

844 A. Admixture Barplot obtained from LEA for various K-values.

845 B. Progressive decrease of LEA cross-entropy criterion. Lower cross-entropy values indicates the number of
846 cluster compatible with the data (here from 30 to 60).

847

848 **S7 Fig. Multidimensional Scaling (MDS) plot depicting relationship among individuals.** Each points
849 represents an individual site and is coloured by region.

850

851 **S8 Fig. Principal Component Analysis recapitulating the relationship among individuals.** The Axis 3
852 and axis 4 are displayed.

853

854 **S9 Fig. Treemix results**

855 A. Proportion of variance explained (y-axis) as a function of the number of migration edge (x-axis)

856 B. Treemix tree inferred without gene flow and residuals

857 C. Residuals for Treemix tree with four migration edges

858

859 **S10 Fig. Compared Demographic Models**

860 Strict Isolation (SI), Isolation with constant Migration (IM), Ancient Migration (AM) and Secondary
861 Contact (SC). The models shared the following parameters: T_{split} : number of generation of divergence
862 (backwards in time). N_{anc} , N_1 , N_2 : effective population size of the ancestral population, of the first and
863 second daughter population. M_1 and M_2 represent the effective migration rates per generation with m the
864 proportion of population made of migrants from the other population. T_{sc} is the number of generations since
865 gene flow started (secondary contact) after a period of isolation. T_{am} is the number of generations since the
866 two populations have diverged without gene flow. Each models are declined in alternative version allowing
867 homogeneous or heterogeneous gene flow and homogeneous and heterogeneous effective size to account for
868 the effect of linked selection (affecting N_e) and barrier to gene flow (affecting m).

869

870 **S11 Fig. Estimate of effective population size across models with linked selection (2N suffix) and**
871 **barriers to gene flow (2m suffix).** AM2m = Ancient Migration with heterogeneous migration.

872 AM2N = Ancient Migration with heterogeneous effective population size. IM2m = Isolation with Migration
873 with heterogeneous migration. IM2N2m = Isolation with Migration with heterogeneous migration and with

874 heterogeneous effective population size, SC2m = Secondary Contact with heterogeneous migration.

- 875 SC2N2m = Secondary Contact with heterogeneous migration and with heterogeneous effective population
876 size.
877
- 878 **S12 Fig. SMC++ estimates of effective population sizes for the 11 whole genome samples included.**
879
- 880 **S13 Fig. SMC++ estimates of divergence time between all possible pairs of joint Site Frequency
881 Spectra based on WGS data.** Two estimates of mutation rate were used. One standard estimate of $1.25e-8$
882 μ /bp/generation and a second based on unpublished data.
883
- 884 **S14 Fig. Negative Relationship between effective recombination rate (ρ) and length of chromosome
885 in bp.**
886
- 887 **S15 Fig. Genome-wide landscapes in coho salmon with (A) landscape of differentiation, B) landscape
888 of genetic diversity C) divergence, D) recombination E) gene density, plots are averaged over 500kb
889 windows.** The figure is the same as figure 5 but all the 30 chromosomes are displayed.
890
- 891 **S16 Fig. Correlation between π_N/π_S and effective population size (N_e) during postglacial time.** Each
892 point represent a sample locality and is color coded by region.
893
- 894 **S17 Fig. DAF spectrum of nonsynonymous and putatively deleterious mutation in each region for all
895 samples.**
896
- 897 **S18 Fig. Significant mean derived allele frequencies of deleterious mutation.** Displayed are the mean
898 derived allele frequencies of polymorphic deleterious sites in each regions +/- 2 standard deviation.
899 x-axis = chromosome position (in bp) and y-axis = effective recombination landscape. Each red dots
900 represents a candidate deleterious mutation.
901
- 902 **S19 Fig. Location of putatively deleterious mutation along the effective recombination landscape.**
903
- 904 **S20 Fig : Positive correlation between chromosome length and occurrence of deleterious variants along
905 chromosome.**
906
- 907 **Table Legend:**
908
- 909 **S1 Table:** Abbreviation, Region and coordinates (Longitude and Latitude) of each river used in the GBS data
910 with the number of individuals provided (nb. Inds).
911
- 912 **S2 Table:** River Name, region and coordinates (Longitude and Latitude) of each river used in the whole
913 genome resequencing data.
914
- 915 **S3 Table:** BST values along with 95% confidence intervals for each river from the GBS data (82 K SNPs).
916 95% confidence intervals obtained after 1000 bootstraps.
917
- 918 **S4 Table:** Distribution of singleton in the WGS data. The Thompson sample displays less singleton than
919 southern samples.
920
- 921 **S5 Table:** model choice results for dadi. AIC and deltaAIC are provided for each pairwise comparison and
922 model. AM = Ancient Migration, IM = Isolation with Migration, SI = Strict Isolation, SC = Secondary
923 Contact, the simplest models assume homogeneous migration and homogeneous effective population size.
924 2N suffix = heterogeneous effective population size, 2Msuffix = heterogeneous migration.
925 Model with both suffix assumes that both effective population size and migration are heterogeneous.
926 Model with a single suffix assumes that either migration or effective population size are heterogeneous.
927
- 928 **S6 Table :** Parameter estimates from GBS data obtained under the best demographic model with dadi.
929 N_{e1} and N_{e2} , effective population size of the compared pair. $m1 \leftarrow 2$ and $m2 \leftarrow 1$, migration from
930 population 2 to population 1 and migration from population 1 into population 2. me_{12} and me_{21} , effective
931 migration rate estimated in the most differentiated regions of the genome T_s : Split Time of the ancestral

932 population in two population; Tsc: duration of the secondary contact P: proportion of the genome freely
933 exchanged (1-P provides the proportion of the genome non-neutrally exchanged); Q: proportion of the
934 genome with a reduced effective population size due to selection at linked sites; hrf = Hill-Robertson factor
935 representing the reduction of Ne in the region Q with reduced Ne.

936

937 **S7 Table:** PCA loadings for Dxy, Fst and Pi.

938

939 **S8 Table:** A) Spearman correlation association to the comparison in Figure 6.

940 B) Linear models testing the combined effect of recombination (Rho) and gene density (Gene count).

941 Interaction terms were not significant for Pi and Dxy and were removed.

942 **S9 Table:** spearman correlation obtained between recombination and Dxy, Fst and Pi when considering each
943 possible pairs of river separately (n = 55) or each river independently (for Pi only).

944

945 **S10 Table:** Results of linear models testing the correlation between piN/piS and GC3.

946

947 **S11 Table:** Summary of deleterious variation by region.

948 1) Derived Allele Frequency (DAF) of deleterious mutation, after averaging by rivers and then by major
949 regional group. 2) Count of deleterious mutations in each rivers and then averaged by major regional group.

950 3) Number of homozygous derived deleterious mutations by individual, after averaging by rivers and then
951 by major regional group. 4) Number of heterozygous mutations by individuals, after averaging by rivers and

952 then by major regional group 5) Total load of derived deleterious mutations by individuals, after averaging
953 by rivers and then by major regional group.

954

955 **S12 Table:** Results of Wilcoxon test (Man-Whitney tests) for differences in derived allele frequencies
956 among major groups for a sample of size 100.

957

958 **S13 Table:** Results of Wilcoxon test (Man-Whitney tests) for differences in count of derived homozygous
959 variants and total load among individuals in each region.

960

961 **S14 Table:** SNPeff results classified by categories for the deleterious mutations identified in the GBS data.