The Gut Microbiota composition of Feral and Tamworth Pigs determined using High-Throughput Culturomics and Metagenomics Reveals Compositional Variations When Compared to the Commercial Breeds.

Gavin J. Fenske[1,2], Sudeep Ghimire[1,2], Linto Antony[1,2], Jane Christopher-Hennings[1,2], and Joy Scaria[1,2*]

[1] Department of Veterinary and Biomedical Sciences, South Dakota State University, Brookings SD 57007, USA

[2]South Dakota Center for Biologics Research and Commercialization, SD, USA.

[*]Corresponding Author: Joy.Scaria@sdstate.edu

Abstract

Bacterial communities in the hindguts of pigs have a profound impact on health and disease. Yet very limited studies have been performed outside intensive swine farms to determine pig gut microbiome composition in natural populations. Feral pigs represent a unique situation where the microbiome structure can be observed outside the realm of modern agriculture. Additionally, Tamworth pigs that freely forage were included to characterize the microbiome structure of this rare breed. In this study, gut microbiome of feral and Tamworth pigs were determined using metagenomics and culturomics. Tamworth pigs are highly dominated by Bacteroidetes primarily composed of the genus *Prevotella* whereas feral samples were more diverse with almost equal proportions of Firmicutes and Bacteroidetes. In total, 46 distinct species were successfully isolated from 1000 colonies selected. The combination of metagenomics and culture techniques facilitated a greater retrieval of annotated genes than either method alone. Furthermore, the naturally raised Tamworth pig microbiome contained a higher number of antibiotic resistance genes when compared to the feral pig microbiome. The single medium based pig microbiota library we report is a resource to better understand pig gut microbial ecology and function by assembling simple to complex microbiota communities in bioreactors or germfree animal models.

Introduction

The microbiome in the hindgut of mammals has been associated with feed conversion efficiency (Singh, et al. 2014: 145-54), pathogen exclusion (Piewngam, et al. 2018: 532-7), and the production of metabolites that directly influence host signaling pathways (Byndloss, et al. 2017: 570-5). It has become clear in recent years, that the microbiome has a drastic impact on host health. Many current methods to study the swine microbiome, are based upon dietary intervention (Hedegaard, et al. 2016: e0147373, Metzler-Zebeli, et al. 2015: 8489). That is, a dietary substrate is introduced to the animal and an effect on microbiome composition, typically 16s rRNA analysis, is measured. Within the swine industry, there is an upswell of work devoted to increasing feed conversion rate ; feed alone accounts for nearly 60% of production costs (Jing, et al. 2015: 11953). While focusing on feed conversion efficiency makes economic sense, the process disregards the biological factors that shaped hindgut evolution, and thus the evolution of the microbiome in pigs. This would be similar to sampling traditional hunter-gatherers which has provided insight into the microbiome of humans outside the realms of modern dietary practices (Smits, et al. 2017: 802). Feral pigs in the American South and Tamworth pigs which are raised as “natural foragers” may provide models of the pig microbiome outside the realm of modern agricultural processes.

Currently there are an estimated 6 million feral pigs in the United States (USDA 2018). Feral pigs were first introduced in the early 1500s by Spanish settlers and cause significant ecological damage. It has been shown that feral pigs decrease the amount of plant litter and cover in areas where they feed (Siemann, et al. 2009: 546-53). Yet, with the ecological and economic toll feral pigs exert, little study has been conducted to elucidate the structure of their microbiome. Here, we used feral pigs as a case study to compare against Tamworth breed pigs. The Tamworth breed is thought to be descended from the Old English Forest pig and has not been crossed or improved with other breeds since the late 18th Century (British Pig Association n.d.). The breed is not a traditional animal used in high production agriculture, bred instead for its tolerance to cold weather and ability to forage. Given Tamworth's unique heritage, close relation to an indigenous pig species in the British Isles, and dietary habits closely matching wild pigs, we chose to include them as another model of the pig microbiome outside of the influence of modern agriculture. Additionally, the Tamworth breed is under watch by the Livestock Conservancy, after previously being designated as threatened, and the microbiome composition has yet to be characterized.

Here we attempt to characterize the microbiomes of Tamworth and feral pigs using metagenomic sequencing and high throughput culturomics on direct colon and cecum contents. To date, modern culturomic efforts have been reserved almost exclusively to human fecal samples and we look to extend such methodology to pigs. The culture strategy employs a single

79 medium with various selection screens to shift the taxa retrieved. A single medium isolation
80 strategy will facilitate downstream defined community studies. For example, simple to complex
81 bacterial communities can be assembled in bioreactors to study the mechanisctics of pig gut
82 microbiome succession(Auchtung, et al. 2015: 42). Similiarly, colonization of such defined
83 communities constituted from a well characterized gut microbiome library could reveal how gut
84 bacterial species or combinations impact gut development and immunity(Goodman, et al. 2011:
85 6252-7). We further characterized the representiative species genomes from our library by whole
86 genome sequencing. Availability of a well characterized strain library with genome information
87 will facilitate future studies to better understand the role of pig gut microbiome in health and
88 disease.

89 # Materials and Methods

90 **Sample Collection and Preparation**

91 Permission was granted from purchasers of three Tamworth pigs to obtain colon and cecum
92 samples immediately following slaughter. The pigs were raised and housed in the upper North
93 Plains of the United States. Small incisions were made into either the colon or cecum with a
94 sterile disposable scalpel. Lumen contents were gently squeezed into sterile 50 mL tubes, mixed
95 with an equal proportion of 40% anaerobic glycerol (final concentration 20% anaerobic
96 glycerol), and immediately snap frozen in liquid nitrogen. For culture preparation, samples were
97 pooled under anaerobic conditions in a vinyl chamber (Coy Labs, USA). Feral samples from four
98 animals were kindly provided by boar hunters in Texas, US. A similar procedure was followed

where colon and cecum samples were taken immediately following evisceration, mixed with anaerobic glycerol and frozen.

Metagenomics

DNA was extracted from gut samples using the DNeasy PowerSoil kit (Qiagen, Germany) following the provided kit protocol. After extraction, Microbial DNA was enriched with the NEBNext® Microbiome DNA Enrichment Kit (New England Biolabs, US) to remove host DNA present after DNA extraction. Metagenomic sequencing was conducted on the Illumina MiSeq platform utilizing V2 (250 bp) paired-end sequencing chemistry. Raw sequencing reads were quality controlled using the read-qc module in the software pipeline metaWRAP (Uritskiy, et al. 2018: 158). Briefly, reads are trimmed to PHRED score of > 20 and host reads not removed by enrichment were removed by read-mapping against a reference pig genome (GCF_000003025.6). Resultant reads from read-qc are hereby referred to as high-quality reads. High-quality reads were passed to Kaiju (Menzel, et al. 2016: 11257) for taxonomy annotation against the proGenomes database (http://progenomes.embl.de/, downloaded March 1, 2019). Kaiju was run in default greedy mode and resultant annotation files were parsed in R (R Core Team 2019). Mash (Ondov, et al. 2016: 132) was run to estimate the Jaccard distance between samples. 10,000 sketches were generated for each sample and the sketches were compared using the *dist* function provided in the Mash software.

Antimicrobial resistance (AMR) genes were predicted from metagenomics assemblies. High-quality sequencing reads were assembled into contigs using the assembly module in metaWRAP ; metaSPAdes (Nurk, et al. 2017: 824-34) was the chosen to assemble the reads: contigs greater than 1,000 bp were retained. Prodigal (Hyatt, et al. 2010: 119-) was run to predict open reading frames (ORF) using the metagenomic training set. Abricate (Seemann 2018) was

then run to annotate the ORF against the NCBI Bacterial Antimicrobial Resistance Reference Gene Database (https://www.ncbi.nlm.nih.gov/bioproject/PRJNA313047, downloaded April 22, 2019).

Contigs were gathered into bins using three methods: MetaBAT2 (Kang, et al. 2019: e27522v1), MaxBin2 (Wu, et al. 2016: 605-7), and CONCOCT (Alneberg, et al. 2014: 1144). Contig bins were kept if the contamination was less than 5% and bin completeness was greater than 85% as determined by CheckM (Parks, et al. 2015: 1043-55). Bins from the three methods used were refined into a coherent bin set using the bin_refinement module in metaWRAP. Refined bins were reassembled with a minimum contig length of 200 bp and the same contamination and completeness parameters as initial bin construction. Metagenomic bin and pure isolate phylogeny was generated using UBCG (Na, et al. 2018: 280-5) to identify and align 92 marker genes. Tree construction was conducted using RAxML (Stamatakis 2014: 1312-3) : GTR+G4 nucleotide model. To identify KEGG homologues, ORF were identified in metagenomic assemblies, bins, and culture genomes using Prodigal. The resultant ORF were annotated against the KEGG database using KofamKOALA (Aramaki, et al. 2019: 602110) run locally.

Culturomics

Colon and cecum samples were pooled respective to feral and Tamworth samples before culture experiments. All culture experiments, including pooling, were conducted under anaerobic conditions inside an anaerobic chamber (Coy Labs, USA). Samples were serially diluted in sterile anaerobic PBS and spread plated onto the media conditions listed in supplemental table 1. Plates were inoculated at 37°C for 48 hours before initial colony selection. 25 colonies were non-selectively sub-cultured from the initial plate to y?BHI plates. The procedure was repeated after

72 hours for a total of 50 colonies per media condition. Colonies were primarily identified using MALDI-TOF (Bruker, Germany). MALDI-TOF scores greater than 2.0 were considered a positive species identification. Scores between 1.7 - 2.0 were taken as positive genus identification. Colonies without a positive MALDI-TOF identification were identified by sequencing the 16s rRNA gene. Briefly, DNA was extracted from colonies using the DNeasy Blood and Tissue Kit (Qiagen, Germany) following the manufacturer's protocol. 16s rRNA sequence was amplified using 27F and 805R primers. The primer sequence is listed in supplemental table 1. Genomes of the selected strains were sequenced on the MiSeq platform utilizing paired-end v3 chemistry (300 bp). Sequencing reads from individual strains were assembled with Unicycler (Wick, et al. 2017: e1005595) : minimum contig length of 200 bp. The raw sequencing reads from the culture isolates and metagenomic samples are hosted at NCBI under the BioProject ID PRJNA555322.

Results

Tamworth and feral pigs harbor distinct microbiotas

We chose to examine Tamworth breed and feral pigs as their lifestyles differ from traditionally raised agricultural breeds. The Tamworth pigs sampled here were not given any antibiotics or growth promoters and could freely graze. We hypothesized that such raising would cultivate a microbiota that would be different to that of swine raised in intensive hog farms. To begin the investigation, colon and cecum samples were metagenomically sequenced from both breeds. Figure one shows the taxonomic annotation of the metagenomic reads respective to the source of isolation. Contradicting our hypothesis that Feral and Tamworth pigs would harbor similar microbiomes, Feral and Tamworth pigs have inverse Bacteroidetes to Firmicutes compositions. The phylum Bacteroidetes represents nearly 53% of all classified reads in Tamworth pigs

compared to 29% in feral pigs. The abundance of Firmicutes in Tamworth samples is lower than feral samples at 15% and 28% respectively. Additionally, nearly 10% more of the feral reads were unclassified compared to Tamworth (37%, 28%, respectively) indicating more of the diversity in feral pigs is not yet known in the proGenomes database. Turning to the genus level, the large increase of Bacteroidetes in Tamworth pigs is primarily composed of the genus *Prevotella*, Figure 1 (B) (38%, feral 11%). Remarkably, the genus *Bacteroides* showed almost identical distribution between the feral and Tamworth pigs (7.6% and 7.6% respectively). The increase of Firmicutes in feral samples is due to an increase in several genera such as *Ruminococcus, Clostridium*, and *Eubacterium* corresponding with significantly higher Shannon diversity index values compared to Tamworth ($p = 0.0024$, Wilcoxon rank-sum test). Full phylum and genus annotation tables are provided in supplemental table 2.

To better understand the distance within a sample source, Tamworth vs. Tamworth, and the difference between sources, Tamworth vs. feral, two clustering methods were employed. First, Mash (Ondov, et al. 2016: 132) was used to sketch the reads sets and compile a distance matrix (Figure 1C). Within the matrix, both Kmeans clustering and hierarchical clustering (average-linkage) separate the samples into Tamworth and feral clades. Mash provides a method to compare metagenomes that is not subject to annotation bias. Principal component analysis (PCA) of the OTU tables was the second method employed. Again, two distinct groups of Tamworth and feral samples are seen in the plot (figure 1D). Interestingly, the Tamworth samples are more homogenous in both the Mash and PCA methods. All pigs were taken from the same farm and this may account for the lower inter-animal microbiome divergence. Thus, the Tamworth and feral pigs examined here harbor distinct microbiotas. Tamworth samples are dominated by the phylum Bacteroidetes, which in turn is largely comprised of *Prevotella*. Feral

191 samples show a much more even distribution of Firmicutes and Bacteroidetes and are more
192 diverse in general.

193 Stated earlier, Tamworth pigs sampled were not given antibiotics in feed nor given any
194 growth promoters. We hypothesized that the lack of antimicrobial agents would correspond to a
195 relatively low number of AMR homologues in the Tamworth microbiota. Additionally, as feral
196 animals (we presume) do not uptake antimicrobials, their AMR number would be low as well.
197 Confoundingly, Tamworth pigs' microbiomes contain at least eight AMR homologues.
198 Additionally, all Tamworth samples yield more AMR homologues than feral samples (figure 2).
199 All Tamworth samples contain four putative AMR genes: *cfxA, lnu(AN2), mef(En2), and tet(40).*
200 No common pattern is apparent for Feral samples; *tet(Q)* is found in 5 of 9 feral samples. Thus,
201 in microbiome composition and AMR presence Tamworth pigs do not mirror feral pigs in
202 microbiome composition nor structure.

203 **Selective screens shift plating diversity**

204 High throughput culturomics was the second method employed to sample the two
205 microbiomes. We chose culture sampling, in addition to sequencing methods, as we believed that
206 many low abundance taxa could be retrieved through culture methods that would be lost in
207 metagenomics. Also, the generation of a culture library enables defined community experiments
208 in the future. The culture sampling strategy utilized is as follows: a base medium (yBHI, or close
209 derivatives) had various selective screens (antibiotics, heat, bile, etc.,) applied to it. A finite
210 growing surface is available for colonization and some species will grow more rapidly and
211 subsequently outcompete others. If appropriate selective pressure is applied, we hypothesized
212 that interspecies selection would decrease allowing for taxa not retrieved in plain medium
213 conditions to grow. The approach is similar to one previously used to culture strains from human

fecal samples (Rettedal, et al. 2014: 4714). One major difference is said work used multiple media compositions, rather than one as in our study. Ten media conditions were used for both Tamworth and feral samples and are listed in supplemental table 1. Twenty-five colonies were picked at 48- and 72-hours post inoculation, for a total of 50 colonies per condition. In total, 1000 colonies were selected from plates, of which 884 were successfully identified. Selective screens shifted the taxa retrieved (figures 3). Figure 3 depicts the number of isolates per media condition with a bar plot depicting the total number of isolates retrieved. *Lactobacillus sp*. was the most abundant organism retrieved (166 isolates) followed by *Escherichia coli* (86), *Lactobacillus mucosae* (74) and *Streptococcus hyointestinalis* (64). The top ten isolates cultured are listed in table 1. One case of selection completely changing plate diversity compared to plain media is that of heat shock treatment. As expected, many spore forming genera including *Bacillus* and *Clostridium* were only able to grow when the inoculum was heated to kill vegetative cells. The selective screens placed upon yBHI not only shifted the taxa retrieved from each plating condition as shown in Figure 3, but also shifted species richness and evenness (figure 4). The most diverse plating condition (Shannon Index) for both Tamworth and feral samples was obtained from plain yBHI : showing as a log-normal community distribution. Similar log-normal community structures are observed for BSM (Tamworth only), Erythromycin and heat shock treatments. Bile treatments and chlortetracycline exhibited strong selective pressure shown as geometric series in the species-rank abundance plots (figure 4). Most of the taxa retrieved from the bile condition were identified as Proteobacteria, indicating that the dosage of bile (1 g / L) was too high.

The culture strategy did not recapitulate the community in the inoculum as defined by metagenomics. In both Tamworth and feral samples, a high number of Firmicutes and

Proteobacteria were isolated, compared to the metagenomic sampling where Bacteroidetes was the most abundant phylum for both sources. If we disregard the bile conditions, which were dominated by Proteobacteria, yBHI clearly selects for common Firmicutes genera including: *Lactobacillus, Streptococcus,* and *Bacillus.* While the screens were successful in increasing the total number of species retrieved, no condition matched the inoculum in form. *Prevotella* for example, the most abundant genus in Tamworth pigs, was only retrieved seven times from 500 colonies. Taken together, the strategy was successful in gathering many isolates that can grow on a common medium but failed in that the most abundant taxa were not retrieved in proportion to the inoculum.

Culturing captures genomic information not captured in metagenomics

The sampling strategy employed did not recapitulate the inoculum community. However, one of the main reasons we chose to culture was that we believed rare taxa would provide information that would be lost to metagenomics. To examine this, we sequenced selected isolates and generated 81 high quality metagenomic bins (completeness > 85%, contamination < 5%). The phylogeny of the metagenomic bins and culture genomes was estimated (figure 5). Consistent with read taxonomy, many of the bins constructed from both Tamworth and feral samples were annotated to the phylum Bacteroidetes. The phyla Firmicutes, Proteobacteria and Actinobacteria were comprised almost entirely of isolate genomes. Isolate genomes not only populated clades of the tree missed by metagenomic bins, but provided genes not observed in metagenomic assemblies nor bins (figure 6). Open reading frames (ORF) were predicted from metagenomic assemblies, metagenomic bins, and culture isolate and were annotated against the KEGG database. Figure 6 shows the abundance (natural log) of KEGG homologues respective to the source of the ORF. Metagenomic bins contained less information than the metagenomic

assemblies. This is expected as the bins are derived from contigs in the assemblies and not all of the contigs will be gathered into bins. The isolates however provided KEGG homologues that were completely missed through culture-independent methods. Thus, culture and culture-independent methods can augment a microbiota analysis providing information that the other method cannot capture.

Discussion

Metagenomic and culture analysis revealed that despite the "organic" raising of the Tamworth pigs studied (ability to forage, no antimicrobials), their microbiome does not resemble that of feral pigs. While it is true that the two most abundant genera were the same for both sources, *Prevotella* and *Bacteroides*, the Tamworth pigs examined were rather homogenous in microbiome composition and were dominated by the genus *Prevotella*. An increase of *Prevotella* in human samples has been attributed to increase of dietary fiber (David, et al. 2014: 559-63, De Filippo, et al. 2010: 14691-6, Smith, et al. 2013: 548-54). It was noted that the Tamworth pigs were fed a high forage diet and were bedded on alfalfa straw. The high dietary fiber intake in Tamworth pigs may be responsible for the high levels of Prevotella and could account for the lower diversity values compared to feral pigs. The exact diet of the feral pigs is unknown, but it has been observed that a major portion of feral pigs diet in Texas is composed of vegetation (Taylor and Hellgren 1997: 33-9). Prevotella has been identified as the most abundant genus in the swine microbiome to date (Holman, et al. 2017: e00004-17) and the sample size is simply too small to discern whether the large dominance of *Prevotella* in Tamworth pigs is breed or diet specific in nature.

Despite the high abundance of *Prevotella* in both Tamworth and feral pigs, and being the most abundant genus in pigs, the sampling strategy we employed only isolated seven *Prevotella* isolates from Tamworth samples (7/500, 1.4%) and no *Prevotella* was isolated from the feral inoculum. In contrast, several genera including *Lactobacillus*, *Escherchia*, *Streptococcous*, and *Bifidobacterium* were overrepresented in culture samples compared to metagenomic sequencing. Our culture results align with an early culture examination of the pig microbiome where the two most abundant isolates cultured were gram-positive cocci and *Lactobacillus* (Russell 1978: 187-93). Both our work and the earlier work relied upon complex media derived largely of peptone digests. As Prevotella is associated with an increase of dietary fiber, work will be needed to develop a defined media that is not based upon peptides such as yBHI. Culturomic techniques have largely focused on human fecal samples. Such studies have been wildly successful in culturing many bacteria that were previously thought to be "unculturable" (Browne, et al. 2016: 543, Lagier, et al. 2016: 16203). Many of the techniques rely upon anaerobic plating onto multiple media formulations, selection of single colonies, and identification. While the multiple media approach generates a higher number of taxa, one study isolated over 1,300 species (Lagier, et al. 2016: 16203), creating multiple media formulations can be expensive and time-consuming. Additionally, bacteria isolated from different media may not grow together on a common media, forfeiting any combined in vitro experimentation. Given the importance of swine in global agriculture, coordinated culture efforts are needed to develop defined community models. Such reduced communities will help to uncover the impact of major ecological principles (drift, selection, speciation, dispersion) at work in the swine hindgut.

One of the original motivations for this work was to establish the microbiotas of pigs outside of traditional agricultural processes. Remarkably, it is the Tamworth pigs and not the

feral pigs that depart from the pig microbiota previously established (Holman, et al. 2017: e00004-17). The ratio of Firmicutes to Bacteroidetes is roughly equal in feral samples and that result aligns with agricultural animals. Tamworth at the phylum level is dominated by Bacteroidetes. Turning to the genus level, the top genus from both sources, *Prevotella*, aligns with the most abundant genus isolated from agricultural animals (Holman, et al. 2017: e00004-17). The genus *Bacteroides* is the second most abundant genus identified in both Tamworth and feral samples and is found in greater abundance than in conventionally-reared agricultural animals. The nearly identical distribution of *Bacteroides* between Tamworth and feral, and the discrepancy between conventionally-reared agricultural animals may indicate that traditional agricultural processes are negatively selecting for the genus. It has been shown that after weaning *Bacteroides* levels plummet in growing pigs and are supplanted by *Prevotella* (Frese, et al. 2015: 28-). Yet in our samples a stable population of *Bacteroides* has persisted. It should be noted that the discrepancy may be accounted for by differing identification methods, metagenomics vs amplicon sequencing, or could be an artifact of sampling size.

The Tamworth pigs harbored more AMR homologues than the feral pigs despite no antimicrobials being provided. It has been shown previously that organically raised pigs harbor significantly more chlortetracycline resistant isolates than feral pigs (Stanton, et al. 2011: 7167). The previous report and our findings indicate that feral pigs are not a significant reservoir of AMR genes. However, the presence of AMR genes in Tamworth pigs may be contributed to recombination. Previous work has established that AMR genes may cluster together with mobile genetic elements and that pigs typically harbor genes conferring resistance to agents not typically used on a particular farm (Johnson, et al. 2016: e02214-15).

Recent studies have proposed metagenomic binning as a culture-independent method to extract genomes from samples (Albertsen, et al. 2013: 533, Pasolli, et al. 2019: 649-62.e20, Tully, et al. 2018: 170203, Wang, et al. 2019: 48). However, one of the main pitfalls of metagenomic binning is that metagenomic assemblers struggle to assemble contigs of closely related taxa, especially if the organisms are found in low abundance (Ayling, et al. 2019). With knowledge now that strain-level variation occurs in species of the microbiome (Lloyd-Price, et al. 2017: 61-6), targeted culture efforts are needed to confirm that strain variation observed in metagenomic data is not simply due to assembler bias. Also, a large portion of genes were not annotated in metagenomic assemblies that were identified in culture isolates. We propose a wholistic approach where metagenomic sequencing coupled with high-throughput culture strategies can effectively cover the shortcomings of either technique, leading to a more complete method of microbiome sampling.

Acknowledgements

This work was in part supported by the grants from the South Dakota Governors Office of Economic Development (SD-GOED) and the United States Department of Agriculture (grant numbers SD00H532-14 and SD00R646-18) awarded to JS. We greatfully thank Mr. Tom Harrison, Jeff Hopper and Brett Grogan for their help in collecting feral pig gut microbiota samples. We are saddened by the untimely passing away of Tom Harrison during the course of this study. We dedicate this manuscript to his memory.

References

Albertsen M, Hugenholtz P, Skarshewski A *et al.* Genome sequences of rare, uncultured bacteria obtained by differential coverage binning of multiple metagenomes. *Nature Biotechnology* 2013;**31**: 533.

Alneberg J, Bjarnason BS, de Bruijn I *et al.* Binning metagenomic contigs by coverage and composition. *Nature Methods* 2014;**11**: 1144.

Aramaki T, Blanc-Mathieu R, Endo H *et al.* KofamKOALA: KEGG ortholog assignment based on profile HMM and adaptive score threshold. *bioRxiv* 2019, DOI 10.1101/602110: 602110.

Association BP. The Tamworth, n.d.

Auchtung JM, Robinson CD, Britton RA. Cultivation of stable, reproducible microbial communities from different fecal donors using minibioreactor arrays (MBRAs). *Microbiome* 2015;**3**: 42.

Ayling M, Clark MD, Leggett RM. New approaches for metagenome assembly with short reads. *Briefings in Bioinformatics* 2019, DOI 10.1093/bib/bbz020.

Browne HP, Forster SC, Anonye BO *et al.* Culturing of 'unculturable' human microbiota reveals novel taxa and extensive sporulation. *Nature* 2016;**533**: 543.

Byndloss MX, Olsan EE, Rivera-Chavez F *et al.* Microbiota-activated PPAR-gamma signaling inhibits dysbiotic Enterobacteriaceae expansion. *Science* 2017;**357**: 570-5.

David LA, Maurice CF, Carmody RN *et al.* Diet rapidly and reproducibly alters the human gut microbiome. *Nature* 2014;**505**: 559-63.

De Filippo C, Cavalieri D, Di Paola M *et al.* Impact of diet in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa. *Proc Natl Acad Sci U S A* 2010;**107**: 14691-6.

Frese SA, Parker K, Calvert CC *et al.* Diet shapes the gut microbiome of pigs during nursing and weaning. *Microbiome* 2015;**3**: 28-.

Goodman AL, Kallstrom G, Faith JJ *et al.* Extensive personal human gut microbiota culture collections characterized and manipulated in gnotobiotic mice. *Proc Natl Acad Sci U S A* 2011;**108**: 6252-7.

Hedegaard CJ, Strube ML, Hansen MB *et al.* Natural Pig Plasma Immunoglobulins Have Anti-Bacterial Effects: Potential for Use as Feed Supplement for Treatment of Intestinal Infections in Pigs. *PLOS ONE* 2016;**11**: e0147373.

Holman DB, Brunelle BW, Trachsel J *et al.* Meta-analysis To Define a Core Microbiota in the Swine Gut. *mSystems* 2017;**2**: e00004-17.

Hyatt D, Chen G-L, Locascio PF *et al.* Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC bioinformatics* 2010;**11**: 119-.

Jing L, Hou Y, Wu H *et al.* Transcriptome analysis of mRNA and miRNA in skeletal muscle indicates an important network for differential Residual Feed Intake in pigs. *Scientific Reports* 2015;**5**: 11953.

Johnson TA, Stedtfeld RD, Wang Q *et al.* Clusters of Antibiotic Resistance Genes Enriched Together Stay Together in Swine Agriculture. *mBio* 2016;**7**: e02214-15.

Kang D, Li F, Kirton ES *et al.* MetaBAT 2: an adaptive binning algorithm for robust and efficient genome reconstruction from metagenome assemblies. *PeerJ Preprints* 2019;**7**: e27522v1.

Lagier J-C, Khelaifia S, Alou MT *et al.* Culture of previously uncultured members of the human gut microbiota by culturomics. *Nature microbiology* 2016;**1**: 16203.

Lloyd-Price J, Mahurkar A, Rahnavard G *et al.* Strains, functions and dynamics in the expanded Human Microbiome Project. *Nature* 2017;**550**: 61-6.

Menzel P, Ng KL, Krogh A. Fast and sensitive taxonomic classification for metagenomics with Kaiju. *Nature Communications* 2016;**7**: 11257.

393 Metzler-Zebeli BU, Schmitz-Esser S, Mann E *et al.* Adaptation of the Cecal Bacterial Microbiome of
394 Growing Pigs in Response to Resistant Starch Type 4. *Applied and Environmental Microbiology*
395 2015;**81**: 8489.
396 Na S-I, Kim YO, Yoon S-H *et al.* UBCG: Up-to-date bacterial core gene set and pipeline for
397 phylogenomic tree reconstruction. *Journal of Microbiology* 2018;**56**: 280-5.
398 Nurk S, Meleshko D, Korobeynikov A *et al.* metaSPAdes: a new versatile metagenomic assembler.
399 *Genome research* 2017;**27**: 824-34.
400 Ondov BD, Treangen TJ, Melsted P *et al.* Mash: fast genome and metagenome distance estimation using
401 MinHash. *Genome Biology* 2016;**17**: 132.
402 Parks DH, Imelfort M, Skennerton CT *et al.* CheckM: assessing the quality of microbial genomes
403 recovered from isolates, single cells, and metagenomes. *Genome research* 2015;**25**: 1043-55.
404 Pasolli E, Asnicar F, Manara S *et al.* Extensive Unexplored Human Microbiome Diversity Revealed by
405 Over 150,000 Genomes from Metagenomes Spanning Age, Geography, and Lifestyle. *Cell*
406 2019;**176**: 649-62.e20.
407 Piewngam P, Zheng Y, Nguyen TH *et al.* Pathogen elimination by probiotic Bacillus via signalling
408 interference. *Nature* 2018;**562**: 532-7.
409 R Core Team. R: A Language and Environment for Statistical Computing, 2019.
410 Rettedal EA, Gumpert H, Sommer MOA. Cultivation-based multiplex phenotyping of human gut
411 microbiota allows targeted recovery of previously uncultured bacteria. *Nature Communications*
412 2014;**5**: 4714.
413 Russell EG. Types and Distribution of Anaerobic Bacteria in the Large Intestine of Pigs. *Applied and*
414 *Environmental Microbiology* 1978;**37**: 187-93.
415 Seemann T. ABRicate: 0.8.0 Edition. GitHub, 2018.
416 Siemann E, Carrillo JA, Gabler CA *et al.* Experimental test of the impacts of feral hogs on forest
417 dynamics and processes in the southeastern US. *Forest Ecology and Management* 2009;**258**: 546-
418 53.
419 Singh KM, Shah TM, Reddy B *et al.* Taxonomic and gene-centric metagenomics of the fecal microbiome
420 of low and high feed conversion ratio (FCR) broilers. *Journal of Applied Genetics* 2014;**55**: 145-
421 54.
422 Smith MI, Yatsunenko T, Manary MJ *et al.* Gut microbiomes of Malawian twin pairs discordant for
423 kwashiorkor. *Science* 2013;**339**: 548-54.
424 Smits SA, Leach J, Sonnenburg ED *et al.* Seasonal cycling in the gut microbiome of the Hadza hunter-
425 gatherers of Tanzania. *Science* 2017;**357**: 802.
426 Stamatakis A. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies.
427 *Bioinformatics (Oxford, England)* 2014;**30**: 1312-3.
428 Stanton TB, Humphrey SB, Stoffregen WC. Chlortetracycline-Resistant Intestinal Bacteria in Organically
429 Raised and Feral Swine. *Applied and Environmental Microbiology* 2011;**77**: 7167.
430 Taylor RB, Hellgren EC. Diet of Feral Hogs in the Western South Texas Plains. *The Southwestern*
431 *Naturalist* 1997;**42**: 33-9.
432 Tully BJ, Graham ED, Heidelberg JF. The reconstruction of 2,631 draft metagenome-assembled genomes
433 from the global oceans. *Scientific Data* 2018;**5**: 170203.
434 Uritskiy GV, DiRuggiero J, Taylor J. MetaWRAP—a flexible pipeline for genome-resolved metagenomic
435 data analysis. *Microbiome* 2018;**6**: 158.
436 USDA. History of Feral Swine in the Americas: USDA, 2018.
437 Wang W, Hu H, Zijlstra RT *et al.* Metagenomic reconstructions of gut microbial metabolism in weanling
438 pigs. *Microbiome* 2019;**7**: 48.
439 Wick RR, Judd LM, Gorrie CL *et al.* Unicycler: Resolving bacterial genome assemblies from short and
440 long sequencing reads. *PLOS Computational Biology* 2017;**13**: e1005595.
441 Wu YW, Simmons BA, Singer SW. MaxBin 2.0: an automated binning algorithm to recover genomes
442 from multiple metagenomic datasets. *Bioinformatics* 2016;**32**: 605-7.

Figures

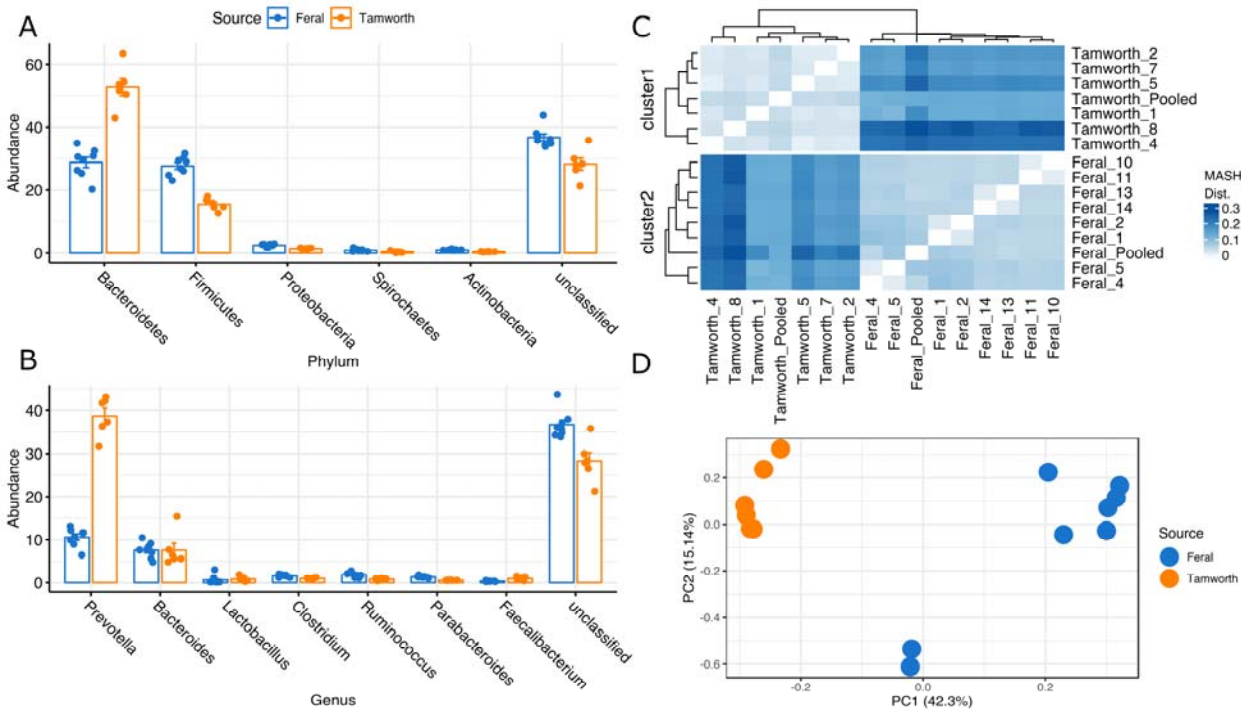


Fig 1. Metagenomic analysis of feral and Tamworth colon and cecum samples. (A)(B) Relative abundance of major phyla and genera annotated from sequencing reads respective to isolation source. (C) Triangle matrix depicting the MASH distance between feral and Tamworth samples. Clusters 1 and 2 are defined by kmeans clustering.

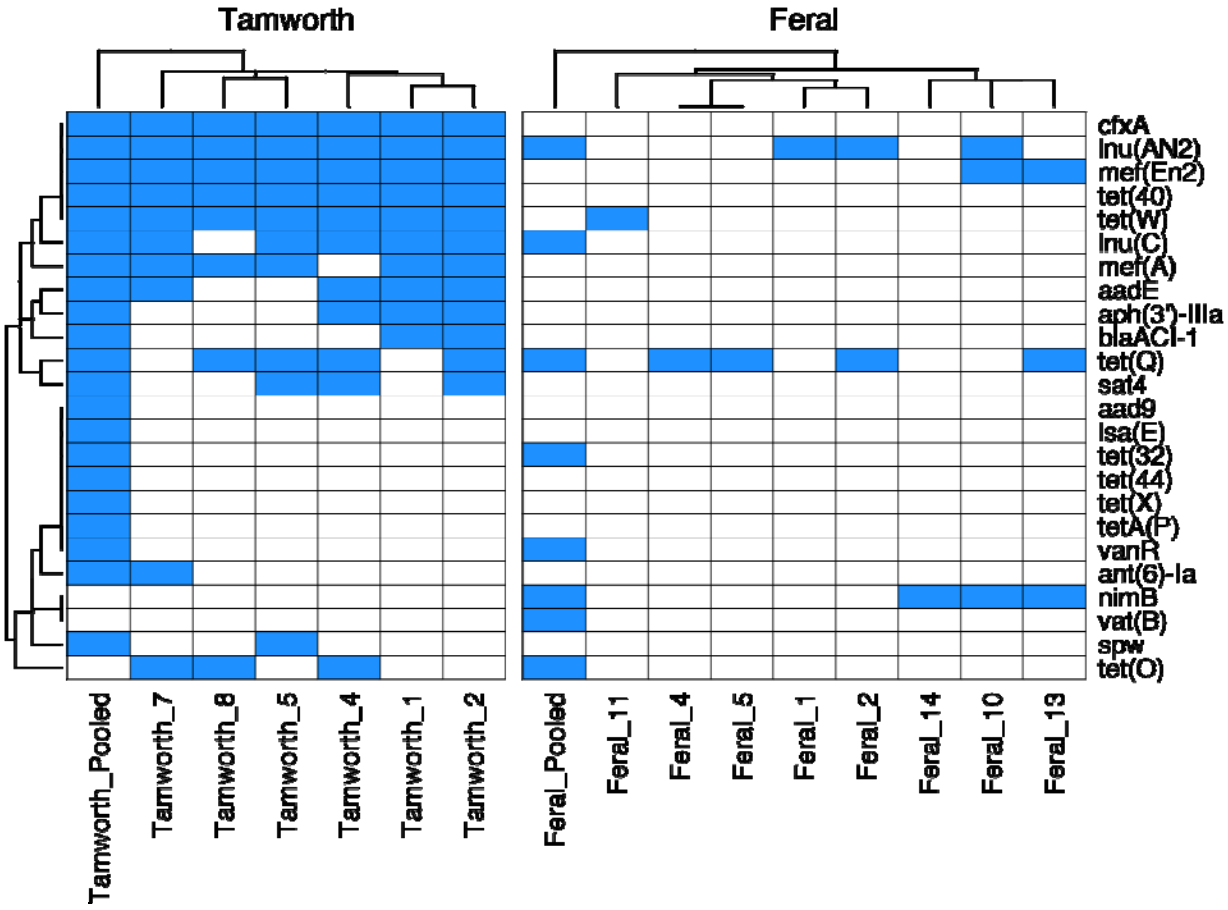


Fig 2. Antimicrobial resistance (AMR) homologues annotated from metagenomic samples. Columns depict individual samples and rows correspond to AMR homologues. Blue color depicts the presence and white color corresponds to absence. AMR homologues were considered present if the coverage value was greater than 90% and a percent homology greater than 70%.

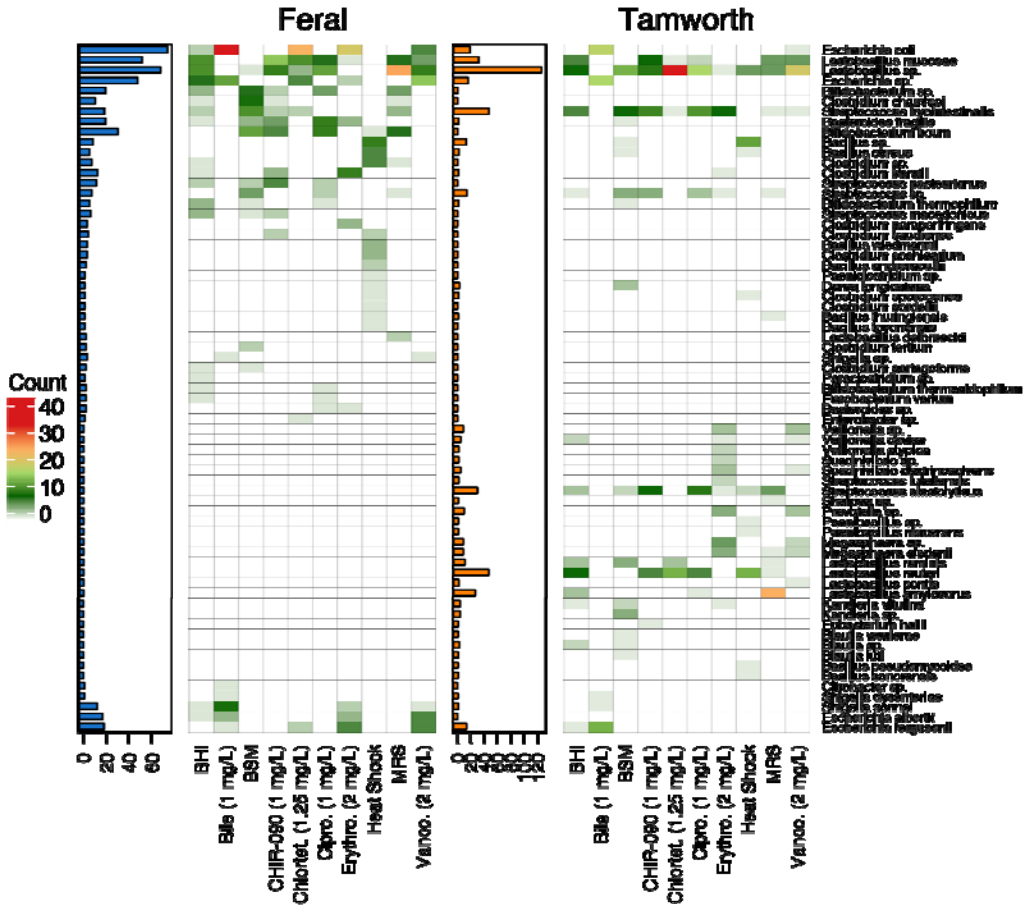
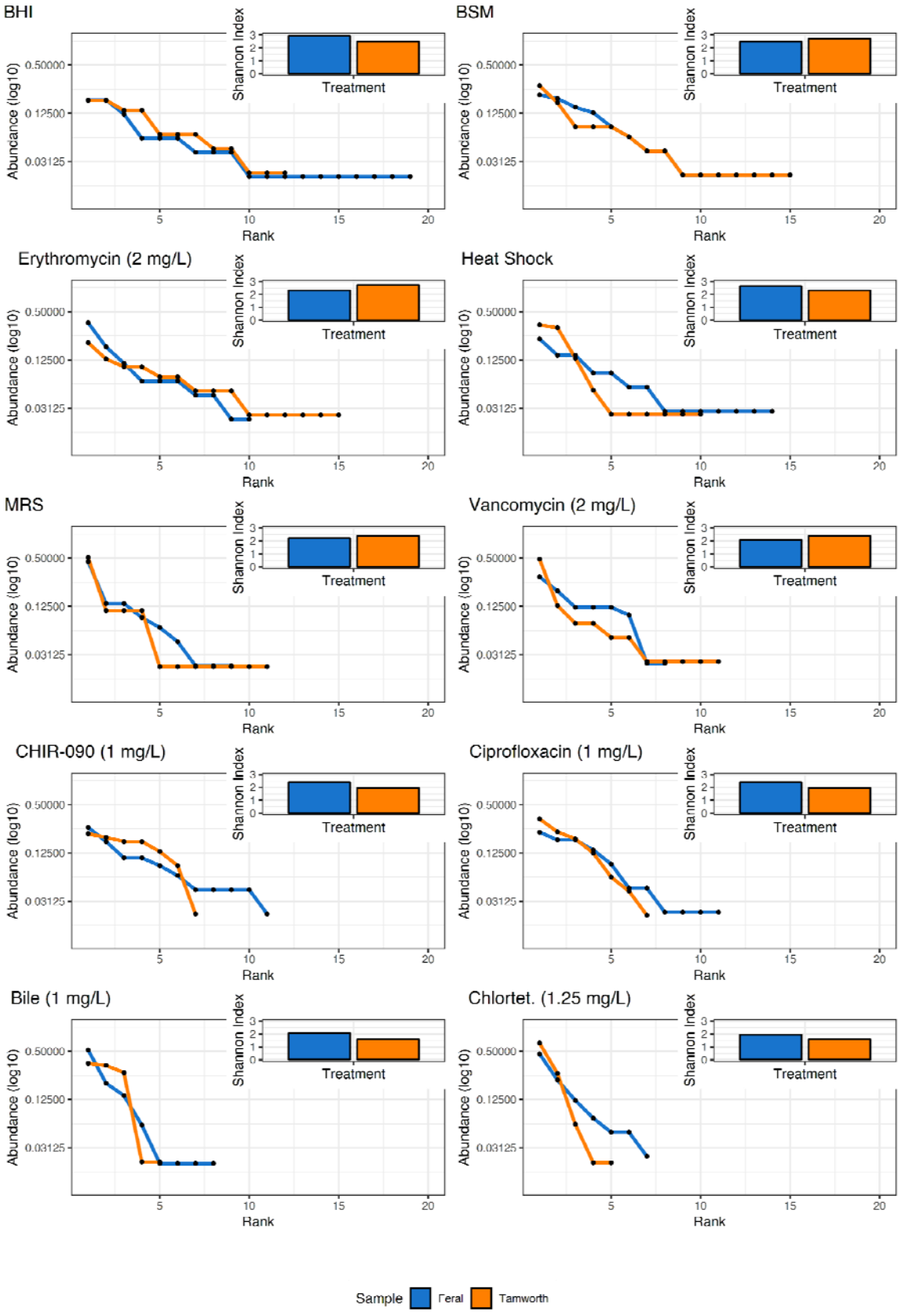


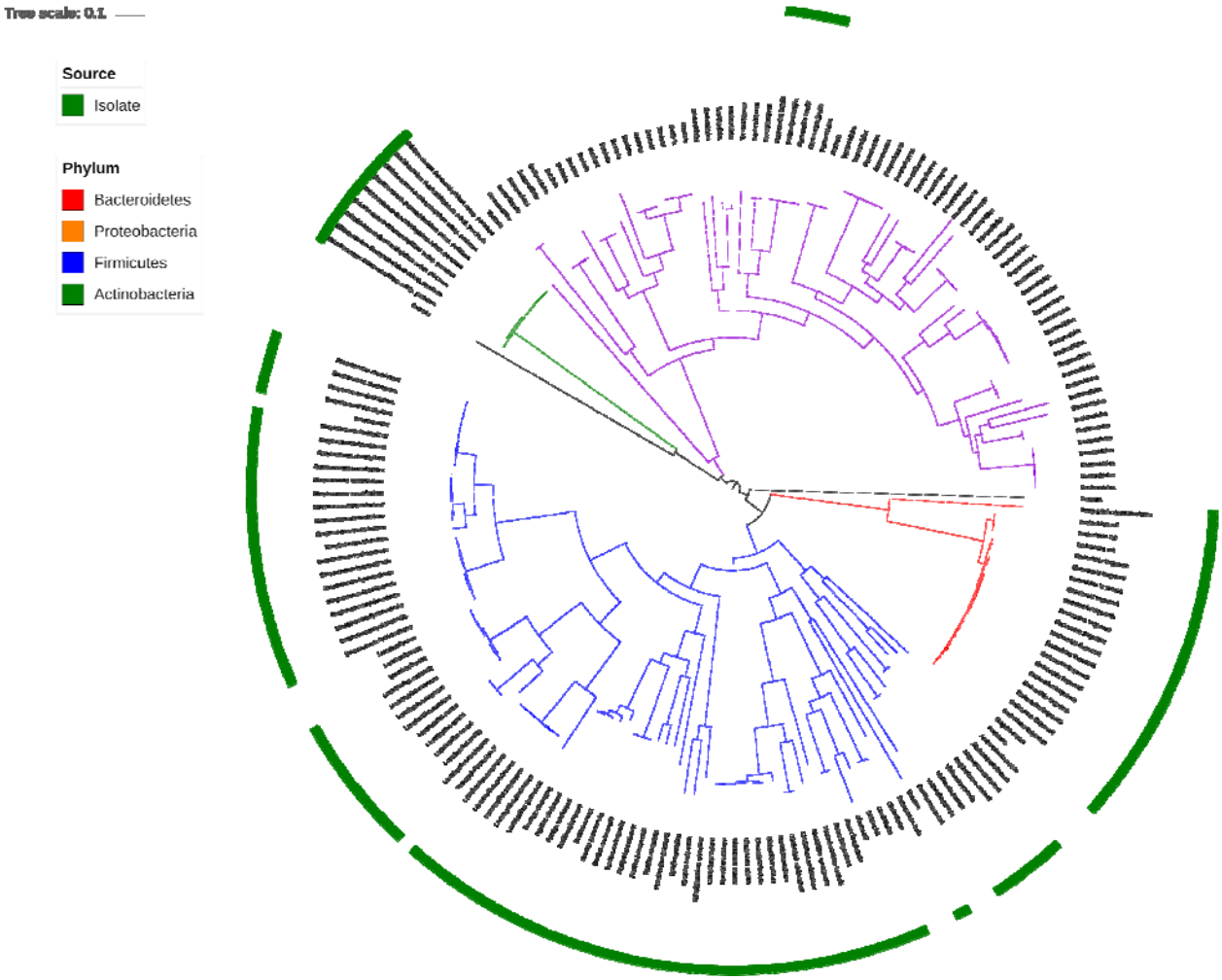
Fig 3. Bacteria isolated from various media conditions. Columns represent individual media conditions and row correspond to bacterial taxa retrieved, cells are colored respective to the number of isolates cultured per media condition. The corresponding bar plot to the left of the matrices shows the total number of isolates retrieved per isolation source.



485 **Fig 4. Rank abundance curves of the various media conditions**. The community evenness of
486 the various media conditions is shown respective to the isolation source. The inlay plot depicts
487 the Shannon Index respective to the isolation source.

488

489



490

491

492 **Fig 5. Maximum-likelihood tree of metagenomic bins and culture genomes.** Tree was
493 constructed from a nucleotide alignment of 92 single-marker genes. General time reversible
494 (GTR) was chosen as the substitution model in tree construction.

495

496

497

498

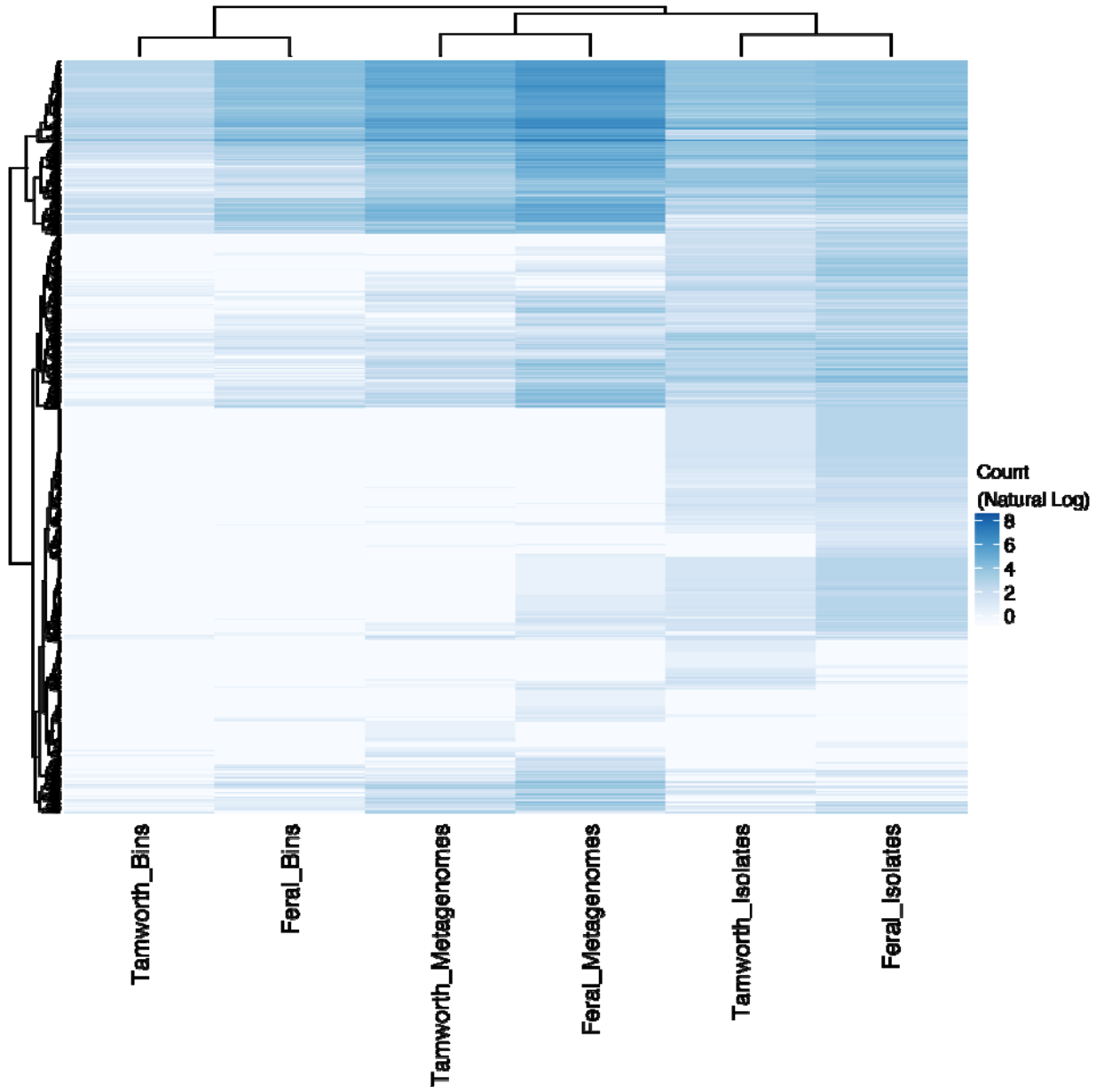


Fig 6. KEGG annotation of open frames from metagenomic assemblies, bins, and culture genomes. Rows and columns are clustered using an average linkage method. KEGG annotations counts are represented as the natural log to increase the clarity of the figure.