

## Metal-Assisted and Microwave-Accelerated Germination

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### ABSTRACT

We report the proof-of-principle demonstration of a methodology, called Metal-Assisted and Microwave-Accelerated Germination, to modulate the germination of plant seeds and growth of plants using gold nanoparticles (Au NPs) and microwave heating. As a model plant seed, basil seeds were heated in a solution of 20 nm Au NPs using a microwave waveguide fiber connected to a solid-state microwave operating at 8 GHz at 20 W, which resulted in the development of longer basil gum as observed by optical microscopy. In control experiments, Au NPs or microwave heating was omitted to establish a baseline growth level under standard experimental conditions (no microwave heating or no Au NPs). Our results also show that hydroponic growth and soil growth of basil plants can be delayed with the use of 20 nm Au NPs at room temperature without microwave heating. The combined use of 20 nm Au NPs and microwave heating at 10 W for 6 minutes results in accelerated growth prolonged life of basil plants.

**Keywords:** Plant seeds, basil, germination, gold nanoparticles, microwave heating, plasma treatment of seeds.

## 32 INTRODUCTION

33 As humanity continue to face food shortage issues,(Perfecto and Vandermeer, 2010;  
34 Clay, 2011; Crist *et al.*, 2017) a number of methods are developed with the aim of greater food  
35 production(Townsend and Porder, 2012; Scott *et al.*, 2018; Jacobsen *et al.*, 2019; Tiberius *et al.*,  
36 2019). To increase the production of food from plants, their seeds are subjected to a number of  
37 physical methods, such as, ultraviolet light treatment,(Noble, 2002) magnetic field  
38 treatment,(Maffei, 2014) and hot water soaking,(Hsu *et al.*, 2003) and chemical  
39 methods(Farajollahi *et al.*, 2014) with the specific goal of reducing the natural germination times  
40 of seeds. The above-mentioned methods are either time-consuming, labor-intensive, or produce  
41 harmful by-product; thus, alternative techniques for the acceleration of the plant germination  
42 process are being pursued. Recently, emerging techniques, such as, cold plasma treatment,(Ling  
43 *et al.*, 2014; de Groot *et al.*, 2018) radio frequency,(Mildažienė *et al.*, 2019) or a combined use of  
44 the three(Mildaziene *et al.*, 2016) were developed for the potential rapid germination of plant  
45 seeds. Although these techniques enhance the germination process, several limitations including  
46 lack of reduction of germination time, inconsistent results, and unsuitable moisture around the  
47 seeds hinder their wide-spread application.

48 In addition, carbon-based and metallic nanoparticles (NPs) have been shown to enhance  
49 germination and seedling growth, physiological activities, nitrogen metabolism, mRNA  
50 expression and protein expression levels, and a few examples are summarized here. For example,  
51 in a study by Kole *et al.* (Kole *et al.*, 2013) fullerol was used to treat seeds to evaluate its effect  
52 on water uptake and fruit biomass, where an increase of up to 54% in biomass yield and 24% in  
53 water content was reported. Mahakham *et al.* (Mahakham *et al.*, 2017) primed rice seeds with  
54 phytosynthesized silver (Ag) NPs improved germination and seedling vigor when compared to  
55 an unprimed control, silver nitrate priming, and conventional hydropriming. A key observation  
56 of this study is the production of more reactive oxygen species in germinating seeds undergoing  
57 nano-priming treatment. Mahakham *et al.* (Mahakham *et al.*, 2017) proposed several  
58 mechanisms for germinations process, which includes creation of nanopores for water uptake,  
59 rebooting of ROS antioxidant systems in seeds, generation of hydroxyl radicals for the loosening  
60 of cell wall, and formation of nanocatalyst for starch hydrolysis fastening. Thuesombat *et al.*  
61 (Thuesombat *et al.*, 2014) investigated the possible effects of different sized Ag NPs (20 nm, 30-  
62 60 nm, 70-120 nm, and 150 nm in diameter) on jasmine rice (*Oryza sativa*) seed germination and

63 seedling growth, where Ag NPs > 150 nm in size were determined to have a more negative effect  
64 on rice seedling growth than smaller sizes. Greater permeation of Ag NPs is found in roots when  
65 smaller Ag NPs (<150 nm) are used, yet the smaller Ag NPs were transported to the shoots with  
66 less effectively than larger sizes. The use of Ag NPs yields inconsistent and negative results on  
67 the growth and development of plants and has the potential to increase environmental toxicity.  
68 The shortcomings of existing methods limit ability to accurately and reliably test with sensitivity,  
69 specificity, and rapidity. Efforts to address these inadequacies will require innovative techniques  
70 capable of rapid germination of seeds.

71 Our research group has been investigating the effect of combined use of metal NPs and  
72 microwave heating on crystallization of amino acids (Pinard and Aslan, 2010) and  
73 proteins,(Mauge-Lewis *et al.*, 2015) decrystallization of uric acid crystals,(Thompson *et al.*,  
74 2017; Boone-Kukoyi *et al.*, 2019) and biosensors,(Abel *et al.*, 2014; Mohammed *et al.*, 2014). In  
75 these studies, metal NPs are either immobilized on a planar surface to assist in the attachment of  
76 biological materials through chemisorption (for crystallization (Pinard and Aslan, 2010) and  
77 biosensors (Abel *et al.*, 2014; Mohammed *et al.*, 2014)) or in solution to function as “nano-  
78 bullets” in decrystallization of crystals on a planar surface (Thompson *et al.*, 2017; Boone-  
79 Kukoyi *et al.*, 2019). Based on our understanding of combined use of metal NPs and microwave  
80 heating to accelerate physical and biological interactions, we hypothesized that the germination  
81 of plant seeds and subsequent growth of plants can be modulated by the use of Au NPs and  
82 microwave heating.

83 In this proof-of-principle work, we report a methodology to modulate (i.e., delay or  
84 accelerate) the germination of basil seeds and subsequent growth of basil plant based on  
85 combined use of Au NPs and microwave heating, called Metal-Assisted and Microwave-  
86 Accelerated Germination (MAMAG). The crux of the MAMAG technique is depicted in **Figure**  
87 **1**. Citrate stabilized (negatively-charged) Au NPs in solution are accelerated by the incident  
88 microwave field towards the basil seeds. The increased kinetic energy of the Au NPs increases  
89 collisions and interactions between NPs and the basil seeds. Subsequently, the following series  
90 of events are thought to occur: the seed testae develop cracks and ridges due to collisions with  
91 Au NPs and microwave heating of water contained in the basil seeds (i.e., due to resistive  
92 losses), Au NPs bind to enzymes on the seed surface, enzyme activity on the basil seeds are  
93 modified by Au NPs and the germination process can be delayed or accelerated based on the

94 extent of enzyme activity that is directly correlated to the presence of Au NPs and microwave  
95 heating. We report that the germination of basil seeds, hydroponic growth and soil growth of  
96 basil seeds can be delayed with the use of 20 nm Au NPs without microwave heating and  
97 accelerated in combination with microwave heating at up to 20 W with a solid-state microwave  
98 source operating at 8 GHz. Since previous reports showed that larger Ag NPs had a negative  
99 effect on the growth of plants, we did not attempt to repeat these experiments with Au NPs in our  
100 study.

## 101 **MATERIALS and METHODS**

102 **Materials.** *Ocimum basilicum* (basil) seeds were purchased from a local seed store in  
103 Baltimore, Maryland, USA in 2014 and were kept in cool and dry place until use (i.e., basil seeds  
104 were 4 years old at the time of their use in 2018). Citrate-stabilized 20 nm Au NPs were  
105 purchased from Sigma Aldrich (Milwaukee, WI, USA) and stored at 4°C. 4-Methylumbelliferyl  
106 butyrate (product number: 19362-5G) was purchased from Sigma Aldrich (USA) and was used  
107 as fluorogenic substrates for esterase. HTS well plates containing 96, flat bottomed wells, was  
108 acquired from Sarstedt (Newton, NC, USA, product number: 82.1583.001).

109 **Instrumentation.** A Digi-microscope from Digitech Industries, Inc. (USA) was used to  
110 view basil seeds during microwave heating using the ISYS800 system (an 8 GHz solid-state  
111 microwave source with variable power levels from 0-20 W, from Emblation Ltd (Scotland, UK).  
112 Microwave heating of samples is accomplished using a ceramic applicator tip (diameter = 5 mm)  
113 and a coaxial cable (outer diameter = 6.7 mm and length = 1.3 m) that delivers microwaves from  
114 the source to the applicator tip. The natural luminescence of the seeds and the esterase activity  
115 was measured using a Cytation™ 5 Cell Imaging Multi-Mode Reader from BioTek Instruments,  
116 Inc. (Winooski, VT, USA). Changes in temperature of the seeds during microwave heating were  
117 measured using the ETS320 model thermal imaging camera from FLIR® Systems, Inc.  
118 (Wilsonville, OR, USA), which was placed directly under the optically clear HTS wells. A  
119 Phenom XL PhenomWorld Desktop Scanning Electron Microscope (SEM) equipped with  
120 elemental analysis capability (ThermoFisher Scientific, Waltham, MA, USA) was used to image  
121 the seeds before and after microwave heating.

122 Micro-seed hydroponic growth pads and pH Perfect Technology Grow nutrient prepared  
123 by Advanced Nutrients (Abbotsford, BC, Canada, optimum dilution 1:1000) were utilized in  
124 hydroponic studies. The soil used to fill growth pads was obtained Miracle Gro Seed Starting



125 Potting Mix (EcoGreenText, Inc. NY, USA). The greenhouse (dimensions 120 x 120 x 200 cm)  
126 was purchased from (Amazon.com) and a light source capable of reflecting all light was obtained  
127 from Amazon.com. All water used was purified using a Millipore Direct Q3 apparatus.

128 **Methods. Determination of seed population viability at room temperature.** One hundred  
129 basil seeds were completely covered with water in an open top pan at room temperature for 1  
130 hour. Then, basil seeds were placed on top of 0.22  $\mu\text{m}$  filter paper in a standard circular petri  
131 dish. Petri dish was closed with its original lid and were kept in the dark for 4 days during which  
132 the germination percentage was calculated daily. The percentage of basil seed germination was  
133 calculated using the following equation: ( $[\# \text{ of seeds germinated} / \text{total \# of seeds}] \times 100$ ) to  
134 estimate the viability of the seed population.

135 **Determination of the combined effect of gold nanoparticles and microwave heating on**  
136 **basil seeds.** Basil seeds were placed in either 50  $\mu\text{L}$  deionized water or 50  $\mu\text{L}$  20 nm gold  
137 nanoparticles (Au NPs) in a 96-well flat bottom plate. Each well of the HTS plates can hold up to  
138 100  $\mu\text{L}$  of solvent. Head space in the HTS wells after the addition of solutions to basil seeds was  
139 used to allow to help cope with the increase in water vapor pressure and to prevent spillage of  
140 water or 20 nm Au NPs from the wells during microwave heating. Dry basil seeds without  
141 solution were used as control seeds. All basil seeds (i.e., dry basil seeds, basil seeds in water, and  
142 basil seeds in 20 nm Au NPs) were treated with continuous microwave heating (2 W, 10 W, 20  
143 W) for 6 minutes. Before and after microwave heating, brightfield, fluorescent images, and SEM  
144 images were obtained to characterize the effect of microwave heating on basil seeds. Fluorescent  
145 images were obtained using a DAPI (blue) filter, a GFP (green) filter, and an RFP (red) filter  
146 setting of our Cytation 5 Multi-mode Cell Imager. Basil seed gum (mucilage) was characterized  
147 by SEM after microwave heating. We also measured temperature changes in the HTS wells with  
148 basil seeds during microwave heating using thermal imaging. Control experiments without  
149 microwave heating were performed at room temperature.

150 **Determination of gold nanoparticles and microwave heating effects on hydroponic**  
151 **growth.** We utilized a modified flood and drain hydroponic growth system to determine the  
152 combined effect of microwave heating and 20 nm Au NPs on hydroponic growth. In our  
153 hydroponic growth system, a polypropylene hydroponic growth pad was placed inside a petri  
154 dish. Three basil seeds kept in water at room temperature for 6 minutes (control experiment, no  
155 microwave heating), basil seeds kept in a solution of 20 nm Au NPs at room temperature for 6

156 minutes (no microwave heating), basil seeds in water with continuous microwave heating at 10  
157 W for 6 minutes, and basil seeds in a solution of 20 nm Au NPs with continuous microwave  
158 heating at 10 W for 6 minutes were placed in the hydroponic growth pad. Holes in the  
159 hydroponic growth pad were made as same depth as the basil seeds to prevent root entanglement.  
160 The hydroponic growth system was well hydrated without evaporation of water throughout basil  
161 seed germination process.

162 ***Determination of gold nanoparticles and microwave heating effects on growth of basil***  
163 ***seeds in soil in a greenhouse.*** Basil seeds were placed in either 50  $\mu$ L deionized water or 50  $\mu$ L  
164 solution of 20 nm Au NPs in a 96-well flat bottom plate. All basil seeds (i.e., basil seeds in water  
165 and basil seeds in 20 nm Au NPs) were treated with continuous microwave heating (2 W, 10 W,  
166 20 W) for 6 minutes. After microwave heating, basil seeds were placed in seedling starter trays.  
167 Seedling starting trays with basil seeds were placed in a greenhouse with LED light to monitor  
168 growth. Basil seeds kept at room temperature and in water at 35°C without microwave heating  
169 for 6 minutes were used as control seeds. All basil seeds were exposed to LED lighting system  
170 (Bloomspect, China, model BS300, 300 W) in 8-hour on and off cycles.

171 ***Thermal Imaging Analysis.*** A FLIR thermal imaging camera (model ETS320) was used  
172 to determine the changes in temperature in the solution containing basil seed during microwave  
173 heating and at room temperature (control experiment, no microwave heating). Basil seeds were  
174 exposed to the treatment of no solution (dry), water, or 20 nm Au NPs with no microwave  
175 treatment or 2 W, 10 W, or 20 W of microwave treatment for 6 minutes while simultaneously  
176 recording with the thermal imaging camera. Thermal images were taken every 30 seconds using  
177 FLIR<sup>®</sup> Systems software.

178 ***Scanning Electron Microscopy (SEM).*** The topography of basil seeds prior to and after  
179 exposure to experimental conditions described in the previous section was studied with SEM.  
180 Untreated basil seeds were placed on a piece of carbon tape and loaded into a Phenom XL SEM  
181 instrument. After focusing and setting the magnification to 300x, an image of the seed was  
182 captured. The same basil seeds were then subjected to the treatment of no solution (dry), water,  
183 or 20 nm Au NPs with no microwave heating or 2 W, 10 W, or 20 W of microwave heating for 6  
184 minutes. The seed was then reloaded into the SEM machine and another image was captured.  
185 Elemental analysis on the captured images were carried out to quantify the extent of 20 nm Au  
186 NPs on the basil seeds.

187            **Measurement of Electrical Conductivity of Basil Seeds.** Keithley source meter (model  
188 2450) was used to measure electrical conductivity of basil seeds soaked either in water or in a  
189 solution of 20 nm Au NPs. Dry basil seeds were used as control experiment. Using the graphic  
190 user interphase window of the instrument, basic source measure settings were set as follows;  
191 measurement type = ohmmeter, sense = 2-wire sense, range of resistance = auto, and current  
192 source parameters (range =auto, source = 1  $\mu$ A and voltage = 21 V). The two wire terminals  
193 from the source meter were pressed on each side of the seeds and the resistance read-out was  
194 recorded. Assuming that the seeds were elliptical in shape, the longer (1.1 mm) and shorter (0.11  
195 mm) lengths were measured, and the cross-sectional area determined. Resistivity values were  
196 calculated using resistance, area and length measurements. Electrical conductivity was calculated  
197 as a reciprocal of resistivity.

198            **Finite-Difference Time-Domain (FDTD) Simulations.** FDTD electromagnetic  
199 simulations were performed to determine the percentage of microwave absorption by each  
200 component of the system and to visualize the electric field propagation through the structure.  
201 MIT's open source MEEP FDTD software (Oskooi *et al.*, 2010) was utilized for the two-  
202 dimensional simulations. Dielectric constants of seeds at 8 GHz microwave frequency were used  
203 in all simulations.(Gabriel *et al.*, 1996) Basil seeds were modeled as an elliptical object (1.1 mm  
204 in length and 0.11 in height) placed in air or water. In the electric field visualization simulations,  
205 a monomode 8 GHz microwave radiation was modeled as a fixed frequency continuous source  
206 located on the top part of the simulation cell. As in the experiments, the microwave radiation was  
207 transmitted to the structure through a 5 mm diameter waveguide, enabling single mode  
208 transmission. The field images depicted the propagation of the microwave radiation through the  
209 structure.

210            Resistive losses within the basil seeds caused by the applied electromagnetic field were  
211 predicted using COMSOL software (Boston, MA, USA). In this regard, basil seeds were  
212 modeled as a 1.1 mm diameter round object placed in water (fully immersed) and were placed  
213 inside a structure with the size of a high-throughput screening well. A monomode microwave  
214 source operating at 8 GHz was placed on top of the well structure to completely cover the well  
215 similar to the experimental setup. Resistive losses (in  $W/m^3$ ) within the basil seeds were  
216 calculated and is shown in three or twelve slices starting from the bottom of the seeds to the top  
217 of the seeds. The number of basil seeds used in FDTD simulations (1, 3, 4, and 10 basil seeds)

218 were used to demonstrate the homogeneous heating of all seeds and were similar to that was  
219 obtained for a single basil seed.

220

## 221 **RESULTS AND DISCUSSION**

222 *Determination of Seed Population Viability.* To assess and minimize the potential  
223 experimental errors in germination due to inherent ability of basil seeds purchased from the  
224 vendor and the 4 years of storage, overall viability of basil seeds at room temperature was  
225 determined prior to the commencement of all experiments. In this regard, the percentage of  
226 viable basil seeds in 100 basil seeds was determined for up to 4 days (4-7 days are required for  
227 germination of basil seeds at room temperature). Fifteen percent ( $n = 15$ ) of the basil seeds  
228 germinated at day 2. At day 3, 85% ( $n = 85$ ) of the basil seeds sprouted. Ninety-eight percent ( $n$   
229  $= 98$ ) of the basil seeds germinated within 4 days. and only 2% of the basil seeds were  
230 determined nonviable after the 4-day period (**Figure S1, Supporting Information**). These  
231 results imply that 98% of the basil seeds used in this study can be germinated and accurate  
232 comparison of the differences in basil seed germination due to the experimental parameters can  
233 be investigated.

234 *Determination of the combined effect of Au NPs and microwave heating on basil seeds*  
235 *using optical microscopy and scanning electron microscopy.* To investigate the effect of using  
236 20 nm Au NPs and microwave heating on the basil seeds as compared to control samples (dry  
237 basil seeds and basil seeds in water without 20nm Au NPs/microwave heating), optical  
238 microscope images that show the top view of the whole basil seeds were taken before and after  
239 microwave heating or at room temperature without microwave heating (control experiment)  
240 (**Figure 2**). In this regard, basil seeds were exposed to the various conditions as described in the  
241 experimental section: dry basil seeds (no water, no Au NPs), basil seeds in water, in a solution of  
242 20 nm Au NPs were kept at room temperature without microwave heating for 6 minutes and  
243 were exposed to continuous microwave heating at 2 W, 10 W and 20 W (maximum power of the  
244 microwave source at 8 GHz) for 6 minutes.

245 It is important to discuss the reasons for the selection of experimental conditions prior to  
246 the presentation of our observations. The choice for volume of water and 20 nm Au NPs solution  
247 (i.e., 50  $\mu$ L sufficient to fully immerse up to 10 basil seeds) was due to the type of HTS well  
248 plates used (96 wells, flat-bottom for easy imaging from the bottom during microwave heating,

249 maximum capacity of 100  $\mu$ L). The choice for the type of metal nanoparticles (Au NPs, diameter  
250 = 20 nm) was due to the following factors: Au NPs are: 1) very stable in water, 2) increase  
251 enzyme activity, 3) act as “nano-bullets” in a microwave energy field, 4) easily observed  
252 individually under electron microscopy. The continuous microwave heating time of 6 minutes  
253 was chosen to minimize the changes to basil seeds in water at room temperature within 6 minutes  
254 observed using optical microscopy, therefore, the effect of 20 nm Au NPs and microwave  
255 heating on basil seeds can be clearly discerned while minimizing the time of exposure of basil  
256 seeds to microwave heating. Microwave heating of basil seeds longer than 6 minutes can  
257 potentially result in denaturation of enzymes in basil seeds due to increase in seed temperature  
258  $>40^{\circ}\text{C}$  and cause undesired structural damage to plant enzymes and germination process can be  
259 negatively affected.

260 *Optical Microscopy.* **Figure 2** shows the overall structural changes to the basil seeds  
261 observed using optical microscopy. In a control experiment, which was carried out to  
262 demonstrate that water is required to germinate basil seeds and that microwave heating of basil  
263 alone in the absence of water do not cause structural damage, no changes in the overall structure  
264 of the dry basil seeds was observed after 6 minutes at room temperature without microwave  
265 heating or with microwave heating. When kept in water and a solution of 20 nm Au NPs for 6  
266 minutes at room temperature (without microwave heating), basil seeds developed a yellowish  
267 gum around the outer layer indicating the commencement of the germination process. Basil seeds  
268 kept in a solution of 20 nm Au NPs without microwave heating for 6 minutes developed longer  
269 whitish gum with red tint due to the presence of 20 nm Au NPs (solution of 20 nm Au NPs is red  
270 in color due their high scattering coefficients above 500 nm).

271 Microwave heating of basil seeds in water or in a solution of 20 nm Au NPs resulted in  
272 the growth of longer gum around the basil seeds. As the microwave power was increased from 2  
273 W to 20 W, basil seed gum in water appeared to increase in length of and became dense and  
274 cloudier, which implies that microwave heating of basil seeds in water alone can increase the  
275 germination speed of basil seeds. Microwave heating of basil seeds in a solution of 20 nm Au  
276 NPs also resulted in the increase in length and change in the color of basil seed gum. These  
277 observations implied that Au NPs and microwave heating can be also used in increasing the  
278 speed of germination of basil seeds. Since microwave heating of dry basil seeds did not result in

279 the growth of basil seed gum, the remainder of the experiments were carried out in water or in a  
280 solution of 20 nm Au NPs.

281 To investigate whether multiple basil seeds can be germinated at once using microwave  
282 heating, 10 basil seeds in water or in the presence of a solution of 20 nm Au NPs were placed in  
283 the same HTS wells (basil seeds were distributed at the bottom of the well without overlapping  
284 each other) and were exposed to continuous microwave heating at 2 W, 10 W and 20 W for 6  
285 minutes. In addition, to monitor the structural changes in basil seeds during continuous  
286 microwave heating and to ascertain the extent of length of basil seed gum, optical images of basil  
287 seeds were captured from the bottom of the HTS wells every minute (**Figure S2, Supporting**  
288 **Information**). These observations reveal that the length of basil gum increases up to the 4<sup>th</sup>  
289 minute of microwave heating and remains at the same length until the end of the experiment, that  
290 is, microwave heating of basil seed for 6 minutes is sufficient to initiate germination of basil  
291 seeds.

292 *Scanning Electron Microscopy (SEM)*. In addition to the optical microscopy images of  
293 basil seeds after microwave heating in the absence and presence of 20 nm Au NPs, extensive  
294 investigation of structural changes to the basil gum using SEM and elemental analysis was  
295 carried out (**Figure 3 and Figures S3-S12, Supporting Information**), which provided an  
296 evidence for the overall structural changes the basil seeds. **Figures S3** shows the overall  
297 structural changes in the basil gum before ( $t = 0$  min) and after each experimental condition ( $t =$   
298 6 minutes) observed by SEM (images acquired from four sides, top, left, right, bottom, for full  
299 view of the seeds). SEM images of dry basil seeds kept at room temperature without microwave  
300 heating show that seed surface is unchanged after 6 minutes. After 6 minutes of microwave  
301 heating of dry basil seeds, light-colored spots on the surface of seeds appeared due to microwave  
302 heating. Basil seeds placed in water without 20 nm Au NPs at room temperature without  
303 microwave heating for 6 minutes developed gum with a small gap on the top and bottom of the  
304 seeds. Microwave heating of basil seeds in water without 20 nm Au NPs for 6 minutes resulted  
305 in looser gum with large gaps around the seed surface, which indicates that microwave heating  
306 alone causes significant changes to basil seeds. Basil seeds placed in a solution of 20 nm Au NPs  
307 at room temperature without microwave heating resulted in observations similar to those  
308 observed for basil seeds kept at room temperature for 6 minutes. Microwave heating of basil  
309 seeds in a solution of 20 nm Au NPs for 6 minutes resulted in looser gum with large gaps around



310 the seed surface. It is important to note that basil seeds were removed from water and the  
311 solution of 20 nm AuNPs to acquire the SEM images, therefore basil gum appears as wrapped  
312 around the seed rather than extending into the solution as seen in optical microscopy images.

313 To gain further insight into the structural changes in basil seeds and discern the  
314 differences in the basil gum developed around the basil seeds in our experiments, SEM images of  
315 the basil gum after each experimental condition (except dry basil seeds, which lacked basil gum)  
316 were collected and compared. **Figure 3** (also **Figure S5 and S9, Supporting Information**)  
317 show that basil gum appears more spread out in a solution of 20 nm Au NPs as compared to  
318 those kept in water. Microwave heating of basil seeds for 6 minutes results in a distribution of  
319 basil gum around the basil seeds depending on: 1) the level of microwave power and 2) the  
320 absence or presence of 20 nm Au NPs. Basil gum appears more spread out in basil seeds  
321 microwave heated as compared to basil gum kept in a solution of 20 nm AuNPs or water at room  
322 temperature without microwave heating. Similar observations are made as the microwave power  
323 is increased from 2 W to 20 W: basil gum appeared to spread out the most at 20 W, and the  
324 presence of 20 nm Au NPs increases the spreading of the basil gum. The observations discussed  
325 in the text so far imply the following: i) basil seeds develop basil gum faster in the presence of  
326 20 nm Au NPs and microwave heating, and can germinate faster due to increased enzyme  
327 activity, or ii) basil gum is destroyed due to microwave heating, enzymes are denatured and basil  
328 seeds cannot germinate as compared to the control basil seeds (at room temperature, no Au NPs,  
329 no microwave heating).

330 In addition, circular structures *ca.* 5  $\mu\text{m}$  in size that are part of the basil gum are seen  
331 around the basil seeds kept in water. Although 20 nm Au NPs are significantly smaller than these  
332 circular structures, 20 nm Au NPs are brighter due to their ability to scatter electrons more  
333 efficiently than the seed components. Individual 20 nm Au NPs are clearly visible and appeared  
334 to be distributed along the basil gum without aggregation as seen in SEM images, which can be  
335 attributed to the presence of enzymes (and other biological materials) in the basil seed and basil  
336 gum. Quantitative elemental analysis of the basil seeds after 6 minutes incubation in a solution of  
337 20 nm Au NPs at room temperature or microwave heating provides evidence for the spatial  
338 distribution of 20 nm Au NPs around the basil gum (**Figure S5 – S12, Supporting**  
339 **Information**). Since enzymes contain primary amine and thiol functional groups, Au NPs can

340 chemisorb on to the enzymes through these functional groups. The effect of chemisorption of Au  
341 NPs on the esterase on the basil seeds is discussed in the **Supporting Information**.

342         Microwave heating of basil seeds in the presence of a solution containing 20 nm Au NPs  
343 at 2 W, 10 W and 20 W resulted in an increase of esterase activity as compared to esterase  
344 activity for basil seeds in water (**Figure S15, Supporting Information**). These observations can  
345 be explained within the context of the combined effect of microwave heating on the basil gum  
346 and Au NPs-catalyzed esterase activity. (Deka *et al.*, 2012; Arsalan and Younus, 2018) As  
347 discussed in the previous paragraphs, microwave heating of basil seeds causes the basil gum to  
348 be looser as the microwave power is increased from 2 W to 20 W, and enzymatic activity is  
349 increased due to the availability of esterase around basil seed. In addition, microwave heating of  
350 basil seeds in the presence of 20 nm Au NPs changes the movement and distribution of 20 nm  
351 Au NPs around the basil gum, which in turn can affect the extent of interactions of 20 nm Au  
352 NPs with esterase in the basil gum. Our research group has previously shown that microwave  
353 heating of Au NPs in solution increases the diffusivity of Au NPs due to selective heating of  
354 water molecules in bulk and around the Au NPs.(Aslan and Geddes, 2007) Since the size of 20  
355 nm Au NPs is *ca.*  $1.88 \times 10^8$  times smaller than the wavelength of microwaves at 8 GHz (3.75 cm)  
356 and the 20 nm Au NPs are negatively charged, 20 nm Au NPs are moved within microwave field  
357 without absorbing the microwave energy, while the temperature of water molecules around 20  
358 nm Au NPs and in bulk is increased due to molecular friction, and subsequently, water is  
359 selectively heated. As a result of selective heating of water and coupling of negatively-charged  
360 20 nm Au NPs with electromagnetic field, 20 nm Au NPs move about the basil seed faster and  
361 chemisorption events between 20 nm Au NPs and esterase are accelerated, which affects the  
362 distribution of Au NPs on the basil gum.

363         In this regard, the extent of temperature changes in bulk during microwave heating of  
364 basil seeds in water and 20 nm Au NPs solutions for 6 minutes and subsequent cooling period of  
365 20 min taken to monitor enzymatic reactions was measured (**Figure S17, Supporting**  
366 **Information**). A maximum increase of *ca.* 11°C in temperature from room temperature (25°C)  
367 to 36°C was observed during microwave heating of basil seeds in water at 20 W. Microwave  
368 heating of basil seeds in a solution of 20 nm Au NPs at 20 W resulted in increase of temperature  
369 from room temperature to 34°C. Microwave heating of basil seeds in all solutions at 2 W and 10  
370 W yielded high temperature in the range of 27°C-30°C, respectively. These observations imply

371 that solutions are incrementally heated for 6 minutes below 37°C, where enzymes are not  
372 expected to denature due to microwave heating.

373 It is important to note that our experimental setup (**Figure S1, Supporting Information**)  
374 lacked the ability of simultaneous measurement of temperature (using a FLIR thermal camera)  
375 and enzyme activity (using a luminescence imaging instrument) during microwave heating of  
376 basil seeds. Therefore, enzymatic activity measurements were carried out immediately after 6  
377 minutes of microwave heating of basil seeds for an additional 20 minutes. In addition, changes in  
378 temperature of solution after microwave heating was stopped for additional 20 minutes was  
379 measured to simulate the experimental conditions during enzymatic activity measurements. The  
380 temperature of all solutions returned to their initial values within 10 minutes. It is also important  
381 to note that after microwave heating for 6 minutes basil seeds were mixed with an equal volume  
382 of substrate solution kept at room temperature or directly transferred to a filter paper for  
383 hydroponic growth. The temperature of basil seeds returned to room temperature within 2  
384 minutes after microwave heating and temperature does not affect enzyme activity after  
385 microwave heating was stopped. Therefore, we can conclude that esterase activity on basil seeds  
386 is affected mainly by microwave heating alone and microwave heating in the presence of 20 nm  
387 Au NPs for 6 minutes. Subsequently, microwave heating of basil seeds at 10 W was selected for  
388 hydroponic growth and soil growth experiments based on the following observations: i)  
389 temperature of the solution with basil seeds reaches a maximum value of 30°C when microwave  
390 heated at 10 W and ii) esterase activity can be modulated by using 20 nm Au NPs.

391 ***Finite-Difference Time-Domain (FDTD) simulations and electrical conductivity***  
392 ***measurements for basil seeds.*** In addition to experimental work to determine the effects of  
393 microwave heating of basil seeds, computational simulations were carried out to determine the  
394 percentage of microwave absorption by each component of the basil seed/water system and to  
395 visualize the electric field propagation through the system. **Figure S18 (Supporting**  
396 **Information)** shows the simulated electric field distribution around basil seeds with and without  
397 water (dry) exposed to monomode microwave point source operating at 8 GHz and 10 W. These  
398 simulations show that basil seeds in bulk water is predicted to absorb 9.1% of the homogenous  
399 electromagnetic energy. Conversely, dry basil seeds (no bulk water) are predicted to absorb a  
400 0.7% extent of the electromagnetic energy due to the presence of water in basil seeds. The

401 electric field propagation images also imply that the energy is absorbed by the basil seeds  
402 submerged in water, while the dry seeds transmit most of the incoming radiation.

403 To gain further insight to microwave heating of basil seeds through computational  
404 simulations of resistive losses of basil seeds (1, 3, 4, and 10 seeds) were carried out (**Figures 4**  
405 **and S19-S20, Supporting Information**). **Figure 4** shows the simulated resistive losses in terms  
406 of 12 cross-sections (in the xz-plane and y-direction) from four basil seeds in water exposed to  
407 monomode microwave point source operating at 8 GHz and 10 W using COMSOL software.  
408 Resistive losses are predicted to be similar for all four seeds and vary throughout all cross-  
409 sections of an individual basil seed. Since the microwave source is placed on top of the cavity,  
410 resistive losses from the top sections (slices 9-12) of the basil seeds are predicted to be the  
411 largest and the least resistive losses are predicted from the bottom sections (slices 1-4) of the  
412 basil seeds, while middle sections (slices 5-8) of the basil seeds show similar resistive losses. In  
413 addition, resistive losses from the top sections of the four basil seeds show the largest extent of  
414 losses occur from the sides of the basil seeds facing each other. In the middle and bottom  
415 sections of the basil seeds, resistive losses occur from the edges and opposite sides of basil seeds,  
416 respectively, which implies that the entire basil seed is heated. Therefore, basil gum is expected  
417 to develop from all sides of the basil seeds. Based on these computational simulations, resistive  
418 losses for a single seed, three seeds and ten seeds during microwave heating were also calculated  
419 (**Figures S19-S20, Supporting Information**), which predict similar patterns for resistive losses  
420 as described for four basil seeds.

421 In addition to theoretical calculations, to further our understanding of the effect of  
422 microwave heating on basil seeds, electrical conductivity of basil seeds immersed in water and in  
423 a solution of 20 nm Au NPs for 6 minutes was measured to be: 0.140 S/(cm.g) for basil seeds in  
424 water and 0.156 S/(cm.g) for basil seeds in 20 nm Au NPs. Since the resistivity of dry basil is  
425 extremely high, no electrical conductivity values were measured. These experimental  
426 measurements show that basil seeds in water and in a solution of 20 nm Au NPs have finite  
427 electrical conductivity, and electrical component of microwave energy can couple to the basil  
428 seeds and be converted into heat through resistive losses in the basil seeds. Since the bulk  
429 temperature of solutions containing basil seeds only reach a maximum value of 30°C when  
430 microwave heated at 10 W, we can conclude that the development of basil gum is accelerated  
431 due to bulk heating and resistive losses in the seeds while enzyme activity is maintained. In the

432 absence of microwave heating, development of basil gum in water is slower at room temperature,  
433 as shown in optical images in this study.

434 ***Effect of gold nanoparticles and microwave heating on hydroponic growth of basil***  
435 ***seeds.*** Based on our ability to control the development of basil gum using 20 nm Au NPs and  
436 microwave heating as described above, we investigated the effect of 20 nm Au NPs and  
437 microwave heating on hydroponic growth of basil seeds to answer the question of “*Can the use*  
438 *of 20 nm Au NPs and microwave heating result in modulation of hydroponic growth of basil*  
439 *seeds?*”. In this regard, basil seeds immersed in water or in a solution 20 nm Au NPs were  
440 microwave heated at 10 W for 6 minutes and placed in a hydroponic growth setup as described  
441 in the experimental section of this study. In addition, control experiments, where basil seeds  
442 were kept at room temperature without microwave heating in water and a solution of 20 nm Au  
443 NPs, were also carried out to discern the differences in using microwave heating versus basil  
444 seed growth at room temperature without microwave heating.

445 To establish a reference point for the hydroponic growth experiments with microwave  
446 heating treated basil seeds, we first discuss hydroponic growth from basil seeds kept at room  
447 temperature without microwave heating. **Figure 5** (see also **Figure S21, Supporting**  
448 **Information**), shows that basil seeds kept in water at room temperature for 6 minutes (i.e.  
449 control sample) developed long stems at 72 hours and grow to full reference length with leaves at  
450 128 hours. At 72 hours, basil seeds kept in a solution of 20 nm Au NPs at room temperature  
451 grew shorter stem as compared to control sample, which implies that the development of basil  
452 plant is delayed in the presence of 20 nm Au NPs. At 72 hours, basil seeds exposed to  
453 microwave heating in the presence of 20 nm Au NPs (i.e., sample labeled as MAMAG)  
454 developed longer stem and leaves as compared to the other samples, which implies that the  
455 development of basil plant is accelerated with the combined use of 20 nm Au NPs and  
456 microwave heating. At 128 hours, basil plants grown from basil seeds treated with the MAMAG  
457 technique (i.e., combined used of Au NPs and microwave heating) displayed the largest leaves  
458 and longest stem, while the growth of basil plants grown from basil seeds kept in the solution of  
459 20 nm Au NPs remained delayed and the control sample reached its peak value in stem length  
460 and leaf size. At 224 hours, two of the three basil plants grown from basil seeds treated with the  
461 MAMAG technique were still viable without any sign of deterioration of leaves or the plant  
462 stem. At 224 hours, control sample showed significant deterioration of leaves and the plant stem

463 and one of the three basil plants grown from basil seeds kept in the solution of 20 nm Au NPs  
464 remained viable. At the end of our observations of basil plant growth at 328 hours, all basil  
465 plants deteriorated completely. These observations clearly demonstrated that the hydroponic  
466 growth of basil plants can be delayed with the use of 20 nm Au NPs without microwave heating  
467 and accelerated and extended with the combined use of 20 nm Au NPs and microwave heating.

468 *Effect of gold nanoparticles and microwave heating on growth of basil seeds in soil.* In  
469 the final step of this study, we investigated the effect of 20 nm Au NPs alone and combined with  
470 microwave heating on the growth of basil seeds to find out whether the growth of basil seeds in  
471 soil can be delayed or accelerated on demand. In this regard, after incubation for 6 minutes at  
472 room temperature in water or 20 nm Au NPs or in 20 nm Au NPs with microwave heating (at 2  
473 W, 10 W, and 20 W), basil seeds were placed in seedling starter trays to monitor growth in soil  
474 for up to 73 days in identical conditions. After 5 days of planting, basil seeds kept in water at  
475 room temperature for 6 minutes (i.e., no microwave heating) developed two-leaves, which was  
476 used as a reference point (control sample) for the growth of basil seeds in the presence of 20 nm  
477 Au NPs and microwave heating (**Figure 6**). Basil seeds kept in in 20 nm Au NPs at room  
478 temperature without microwave heating developed smaller leaves and the height of the basil  
479 plants were shorter than those observed for the control sample, which implied that the growth of  
480 basil plants from their seeds treated with 20 nm Au NPs at room temperature was delayed in soil.  
481 Microwave heating of basil seeds (at 2 W, 10 W, and 20 W) in the presence of 20 nm Au NPs for  
482 6 minutes resulted in taller basil plants with wider leaves after 5 days of planting in soil, which  
483 implied that the growth of basil plants from their seeds microwave heated in the presence of Au  
484 NPs was accelerated in soil. Furthermore, after 73 days planting of basil seeds, significant  
485 differences between basil plants were observed: while basil plants grown from basil seeds kept in  
486 water at room temperature (i.e., no microwave heating) for 6 minutes wilted, basil plants grown  
487 from basil seeds kept in a solution of 20 nm Au NPs at room temperature and basil seeds  
488 microwave heated in a solution of 20 nm Au NPs were still thriving 73 days after planting in soil  
489 (**Figure 6**). It is important to note all basil plants were exposed to identical experimental  
490 conditions. Moreover, the height of the basil plants grown from basil seeds kept in a solution of  
491 20 nm Au NPs at room temperature were taller than any basil plant grown under other  
492 conditions.



493           Since microwave heating of basil seeds in water and in 20 nm Au NPs results in an  
494 increase in temperature up to 36°C, we investigated the effect of incubation of basil seeds in pre-  
495 heated water without microwave heating to compare these results with the results obtained using  
496 basil seeds treated with microwave heating. **Figures S22 (Supporting Information)** show that  
497 the development of basil gum, enzymatic activity and soil growth in 17 days were similar to the  
498 results observed for basil seeds kept at room temperature without microwave heating, and the  
499 MAMAG technique, based on combined use of microwave heating and 20 nm Au NPs, for the  
500 pre-treatment of basil seeds offer a superior alternative to simply soaking basil seeds in pre-  
501 heated water. These observations imply that the growth of basil seeds in soil can be delayed or  
502 accelerated on demand with 20 nm Au NPs and microwave heating.

503

## 504 **CONCLUSIONS**

505           In this work, we demonstrated that a solution of 20 nm Au NPs and microwave heating  
506 from a monomode microwave source can be used as new approach to modulate the germination  
507 of basil seeds and subsequent hydroponic growth and soil growth of basil plants. In this regard,  
508 single basil seeds or 10 basil seeds were placed in a 50 µL solution of 20 nm Au NPs in a HTS  
509 well plates and were exposed to 6 minutes of continuous microwave heating using a solid-state 8  
510 GHz microwave generator operating at 2 W, 10 W and 20 W microwave power. In control  
511 experiments, either 20 nm Au NPs or microwave heating was omitted to determine the effect of  
512 each of 20 nm Au NPs alone (at room temperature without microwave heating) or microwave  
513 heating alone (in water without 20 nm Au NPs). We have made the following conclusions from  
514 our experimental observations and computational simulations:

- 515 1) Optical microscopy was used to assess the macro-scale changes in basil seeds and showed  
516 that
- 517 a. dry basil seeds did not germinate with or without microwave heating, and
  - 518 b. basil in water or in a solution of 20 nm Au NPs in the presence of microwave heating  
519 resulted in longer basil gum as compared to basil seeds kept in room temperature without  
520 microwave heating.
- 521 2) SEM was used to assess the micro-scale changes in basil seed gum and showed that

- 522 a. basil seed gum is more spread out due to microwave heating and in the presence of 20 nm  
523 Au NPs, and
- 524 b. 20 nm Au NPs are distributed throughout the surface of the basil seeds without  
525 significant aggregation.
- 526 3) Luminescence kinetic analysis of enzymatic activity was used to assess the effect of 20 nm  
527 Au NPs and microwave heating on esterase, which is a key enzyme for germination of basil  
528 seeds. Esterase activity was monitored for an additional 20 minutes after the initial 6 minutes  
529 incubation of basil seeds,
- 530 a. esterase activity in basil seeds in water and in a solution of 20 nm Au NPs without  
531 microwave heating was similar, and
- 532 b. esterase activity in basil seeds in 20 nm Au NPs was up to 2-fold higher than esterase  
533 activity in water with microwave heating.
- 534 4) Maximum actual average temperature of water and 20 nm Au NPs containing basil seeds  
535 during microwave heating at 20 W was measured to be 36°C and 34°C, respectively.  
536 Enzymes in basil seeds are not expected to denature due to microwave heating of basil seeds  
537 in water and in a solution of 20 nm Au NPs.
- 538 5) FDTD simulations predict that
- 539 a. basil seeds in water absorbs 9.1%, transmits 85.4%, and reflects 5.5% of the microwave  
540 radiation.
- 541 b. Resistive losses from the top sections of the basil seeds show the largest extent of losses  
542 occur from the sides of the basil seeds facing each other. Resistive losses occur from the  
543 edges and opposite sides of basil seeds in the middle and bottom sections of the basil  
544 seeds, respectively.
- 545 c. The entire basil seed is heated during microwave heating, and therefore basil gum is  
546 expected to develop from all sides of the basil seeds after microwave heating.
- 547 6) Hydroponic growth of basil plants can be
- 548 a. delayed with the use of 20 nm Au NPs at room temperature without microwave heating,  
549 or
- 550 b. accelerated and life of the basil plant can be extended with the combined use of 20 nm  
551 Au NPs and microwave heating.
- 552 7) Growth of basil seeds in soil can be

- 553 a. delayed by pre-treatment of basil seeds in a solution of 20 nm Au NPs, or  
554 b. accelerated after pre-treatment of basil seeds with microwave heating and a solution of 20  
555 nm Au NPs.

## 556 **ACKNOWLEDGMENTS**

557 Authors greatly appreciate partial financial support from Morgan State University Innovation  
558 Works I-GAP Program.

## 559 **SUPPORTING INFORMATION**

560 Additional SEM images, luminescence microscopy images of basil seeds, detailed hydroponic  
561 growth results, real-time temperature measurements of the well with a single basil seed at room  
562 temperature and FDTD simulation of resistive losses are presented.

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659 **FIGURE LEGENDS**

660 **Figure 1.** The crux of the MAMAG technique: citrate-stabilized (negatively charged) gold  
661 nanoparticles (Au NPs) are accelerated by the electromagnetic field. The increased kinetic  
662 energy of Au NPs increases collisions between nanoparticles and basil seeds. Seed testae develop  
663 cracks and ridges due to collisions of Au NPs with the seed surface and resistive losses due to  
664 microwave heating of water within the seeds, leading to controlled germination of seeds.  
665 Esterase activity on the seed surface is also increased due to the presence of Au NPs and  
666 microwave heating. Gold nanoparticles bind to the enzymes/proteins via Au-thiol and  
667 Au/primary amine bonds and electrostatic interactions. Wavelength of monomode  
668 electromagnetic field at 8 GHz is ca. 3.75 cm.

669

670 **Figure 2.** Bright-field images of basil seeds before and after application of microwave heating.  
671 Dry basil seeds, basil seeds in water (no Au NPs), or basil seeds in 20 nm Au NPs were  
672 microwaved at 2 W, 10 W, and 20 W of microwave power. The presence of Au NPs and



673 microwave heating causes the seeds to develop longer gum within 6 minutes. Each basil seed is  
674 ca. 1.1 mm in length in the longest axis.

675

676 **Figure 3.** Scanning electron microscope images of basil seed gum after continuous microwave  
677 heating (2 W, 10 W, or 20 W) for 6 minutes. Control seeds were kept at room temperature for 6  
678 minutes in either water or 20 nm Au NPs without microwave heating. Basil seed gum appears  
679 more spread out due to microwave heating and 20 nm Au NPs. Circular structures seen in basil  
680 in water is in 5 mm in size and part of the basil gum. Although Au NPs are significantly smaller  
681 than these circular structures, Au NPs are brighter due to their ability to scatter light more  
682 efficiently than the seed components. Scale bars = 50 mm and 5 mm (images on the right).

683

684 **Figure 4.** FDTD simulation of resistive losses from basil seeds fully immersed in water exposed  
685 to monomode microwave point source operating at 8 GHz and 10 W using COMSOL software.  
686 Top and side view of the microwave cavity and seeds are shown on the left. Resistive losses  
687 from the seeds are shown as slices starting from the layer bottom of the seeds up to the layer  
688 corresponding to the plane on top of the seeds.

689

690 **Figure 5.** Basil seeds in hydroponic growth. Three basil seeds were kept in water without Au  
691 NPs at room temperature (No MW/RT), or kept in a solution of 20 nm Au NPs at room  
692 temperature, or microwave heated (MW) in in a solution of 20 nm Au NPs for 6 min at 10 W and  
693 placed into a modified growth system. Basil seeds in water at room temperature (no microwave  
694 heating) are used as a control sample to monitor the normal hydroponic growth.

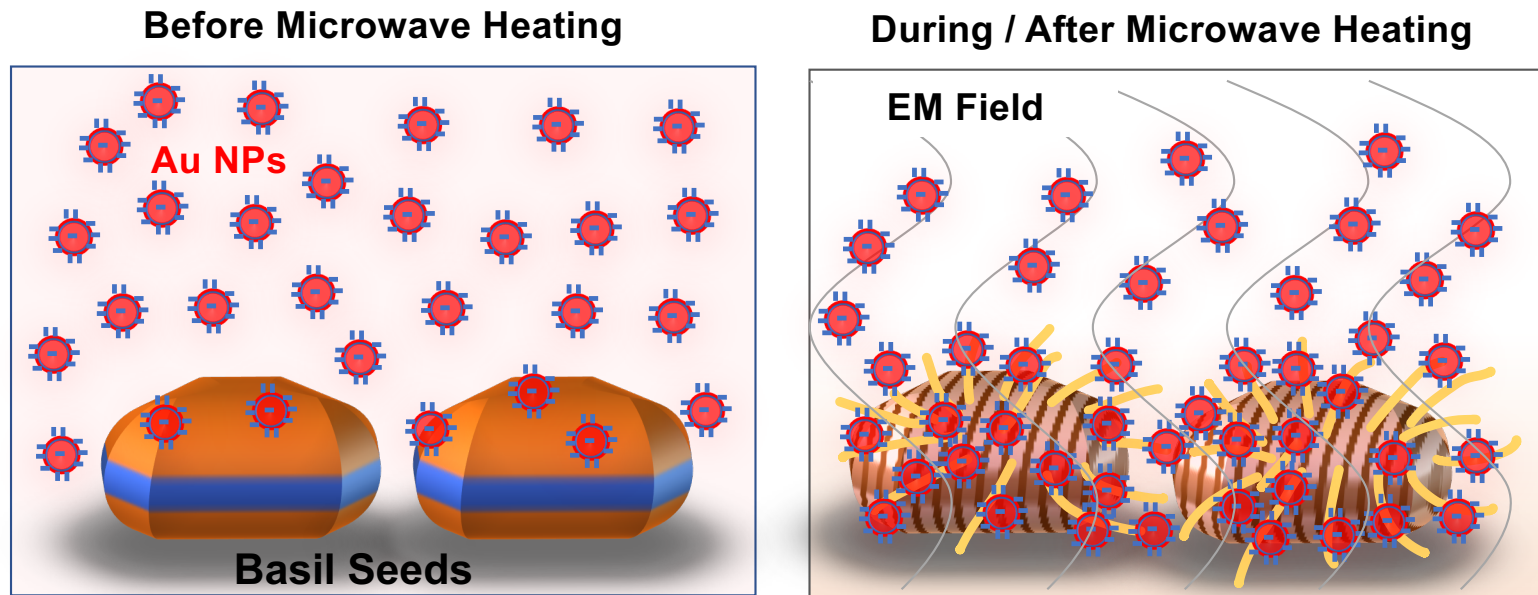
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696 **Figure 6.** Basil seed growth in soil in a greenhouse. Basil seeds were microwaved continuously  
697 for 6 minutes at 2 W, 10 W, and 20 W. Basil seeds in water only (no microwave heating) and  
698 basil seeds in 20 nm Au NPs only (no microwave) were used as control basil seeds. All pictures  
699 were taken together with the same camera. Vertical white line shows the length of basil plants =  
700 7 cm.

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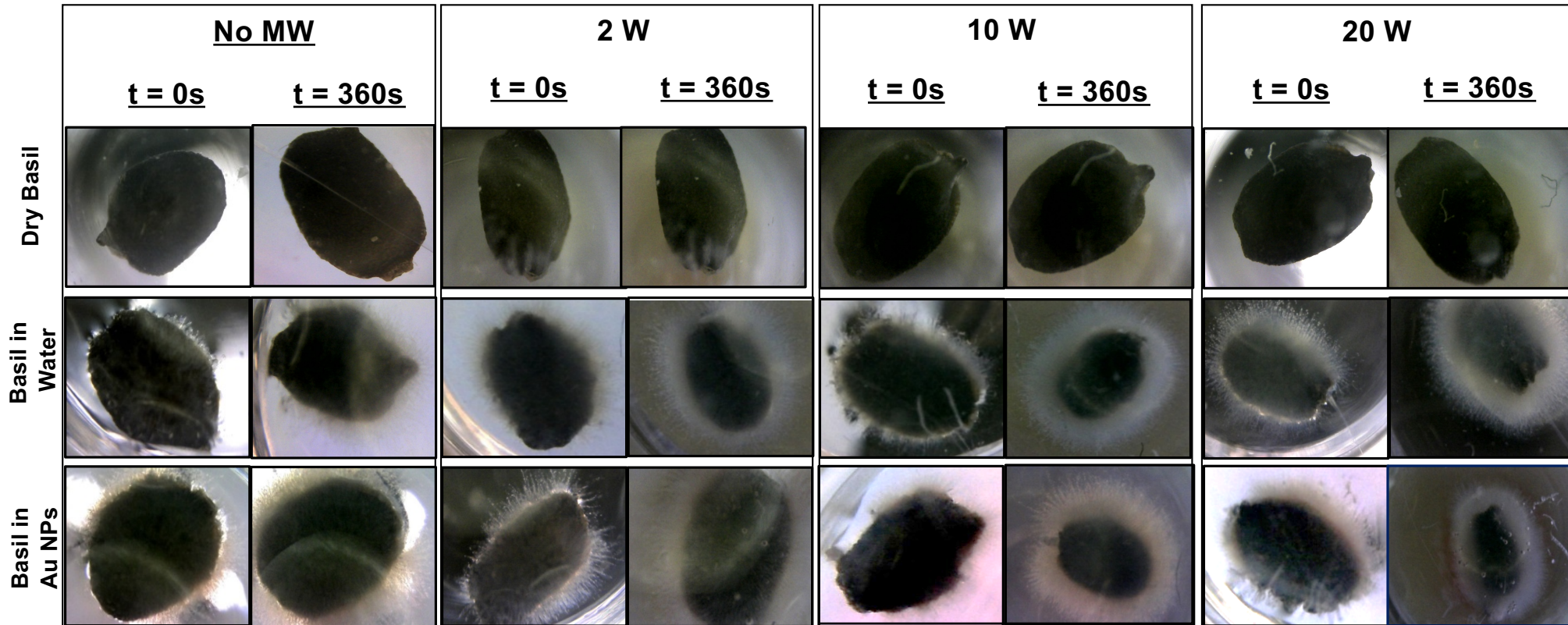
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## The crux of metal-assisted and microwave-accelerated germination of seeds



**Figure 1.** The crux of the MAMAG technique: citrate-stabilized (negatively charged) gold nanoparticles (Au NPs) are accelerated by the electromagnetic field. The increased kinetic energy of Au NPs increases collisions between nanoparticles and basil seeds. Seed testae develop cracks and ridges due to collisions of Au NPs with the seed surface and resistive losses due to microwave heating of water within the seeds, leading to controlled germination of seeds. Esterase activity on the seed surface is also increased due to the presence of Au NPs and microwave heating. Gold nanoparticles bind to the enzymes/proteins via Au-thiol and Au/primary amine bonds and electrostatic interactions. Wavelength of monomode electromagnetic field at 8 GHz is ca. 3.75 cm.

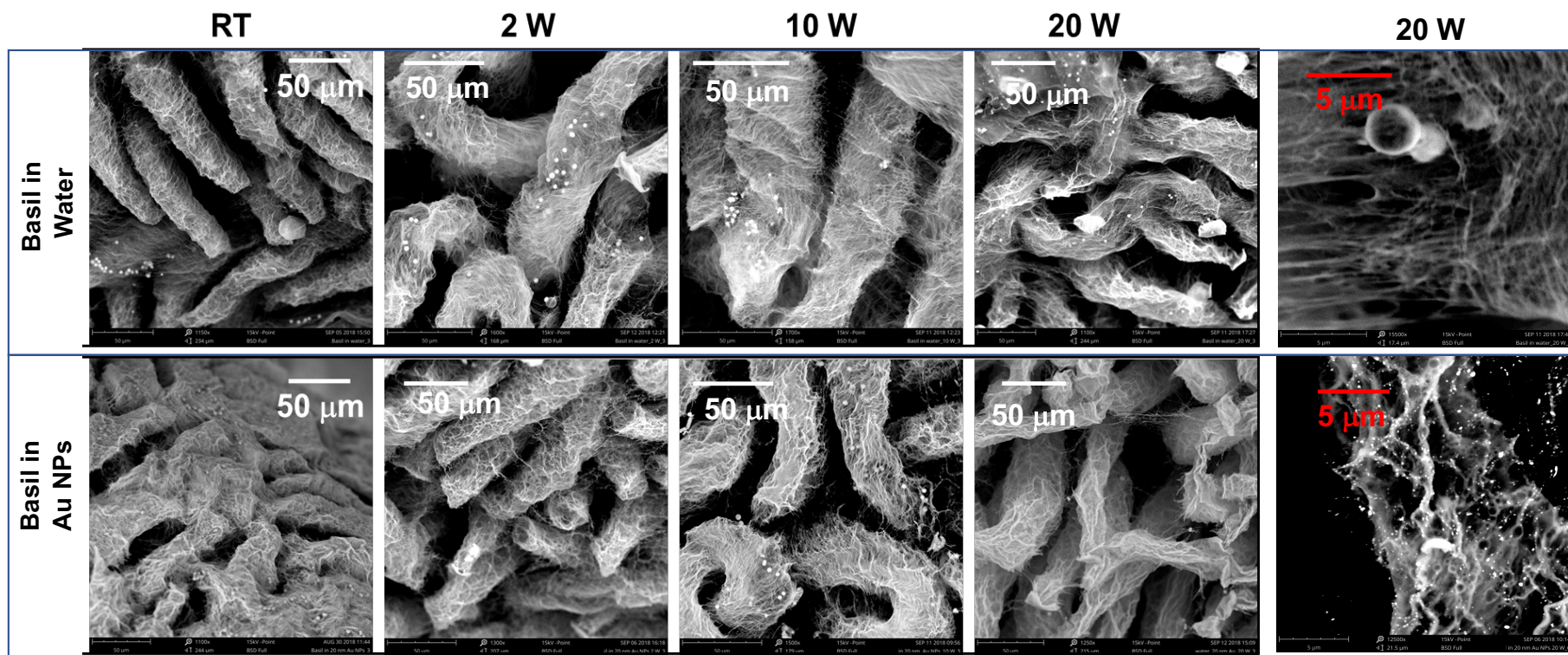
### Optical images of basil seeds



**Figure 2.** Bright-field images of basil seeds before and after application of microwave heating. Dry basil seeds, basil seeds in water (no Au NPs), or basil seeds in 20 nm Au NPs were microwaved at 2 W, 10 W, and 20 W of microwave power. The presence of Au NPs and microwave heating causes the seeds to develop longer gum within 6 minutes. Each basil seed is ca. 1.1 mm in length in the longest axis.

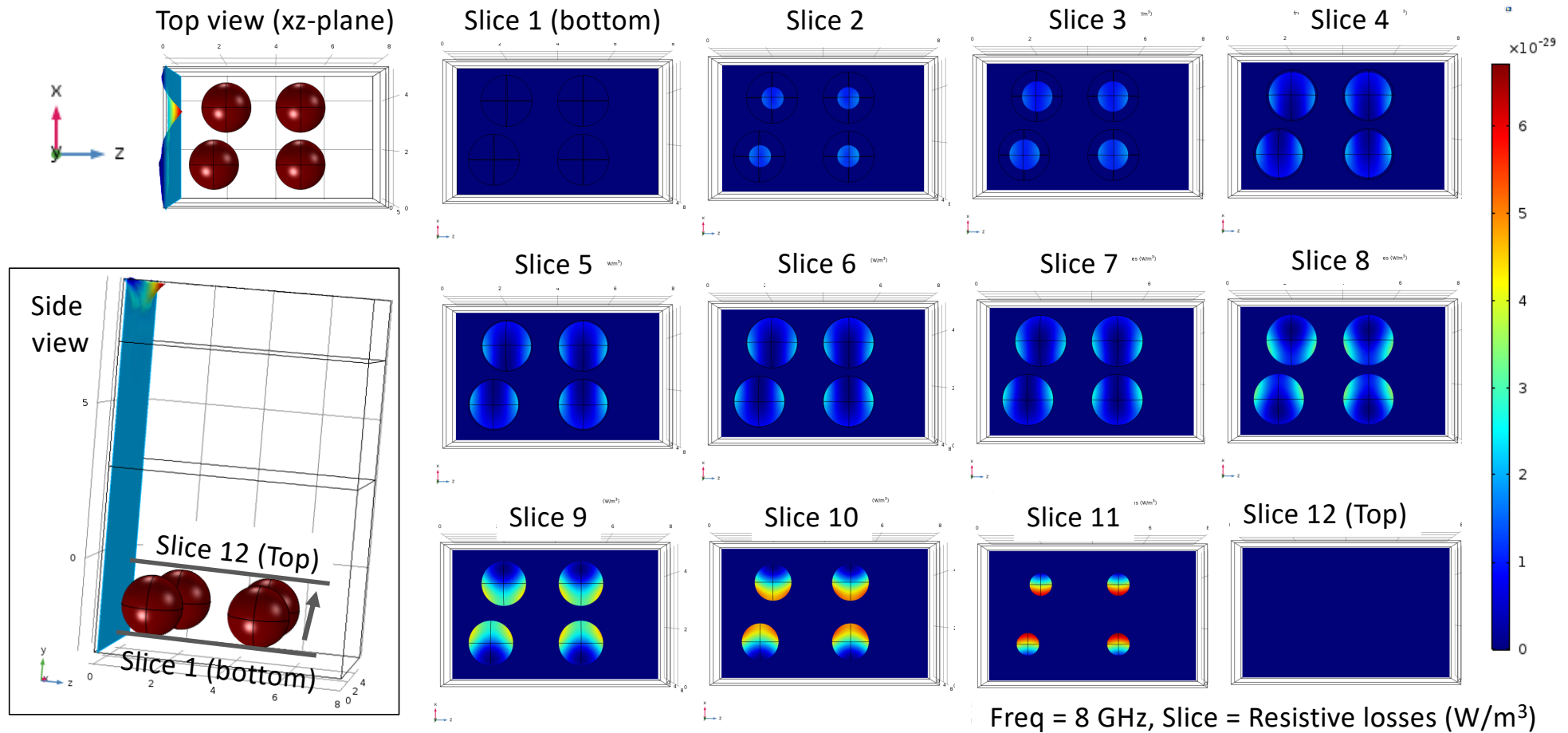


### SEM images of basil seeds after 6 minutes



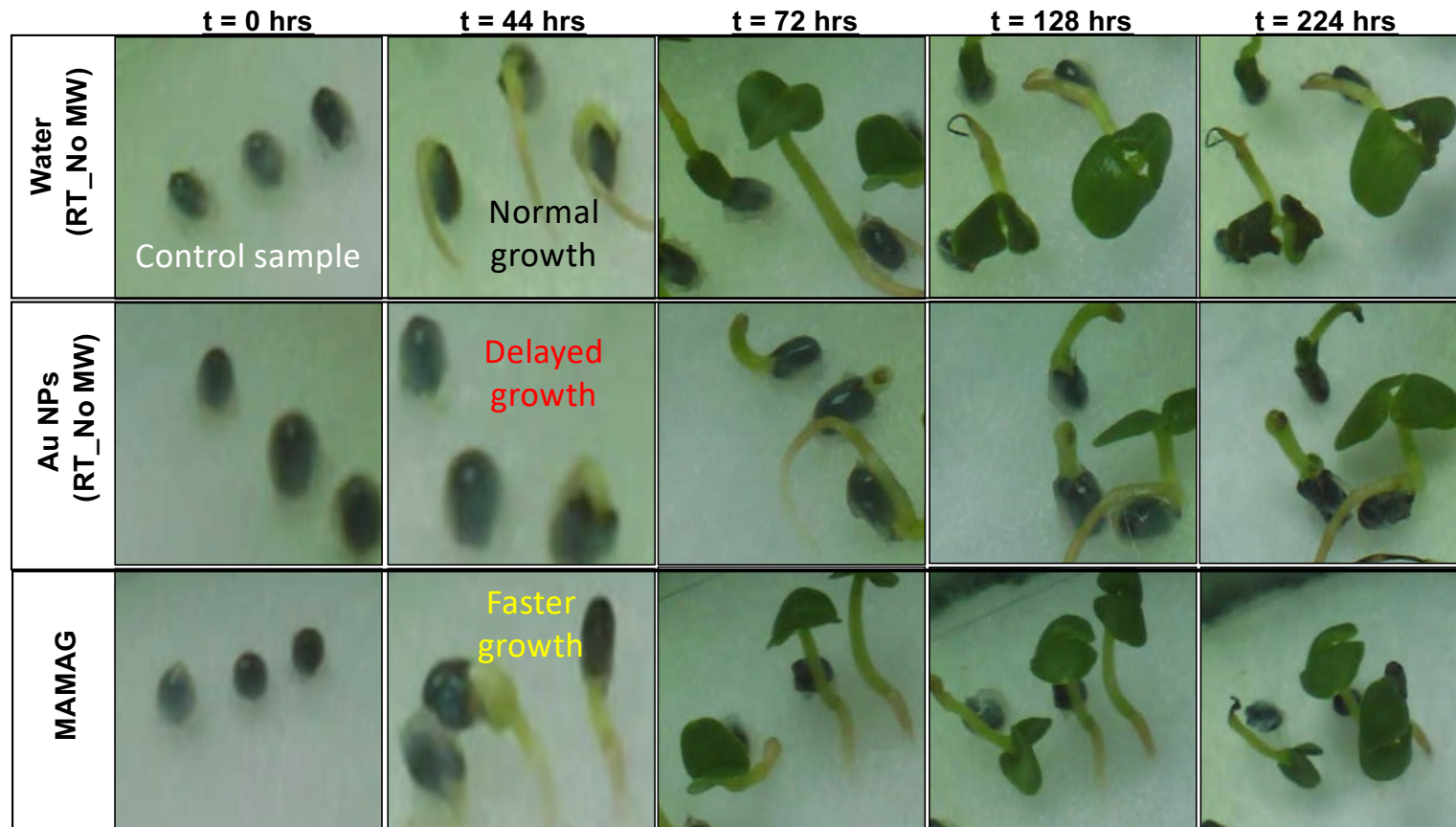
**Figure 3.** Scanning electron microscope images of basil seed gum after continuous microwave heating (2 W, 10 W, or 20 W) for 6 minutes. Control seeds were kept at room temperature for 6 minutes in either water or 20 nm Au NPs without microwave heating. Basil seed gum appears more spread out due to microwave heating and 20 nm Au NPs. Circular structures seen in basil in water is in 5 μm in size and part of the basil gum. Although Au NPs are significantly smaller than these circular structures, Au NPs are brighter due to their ability to scatter light more efficiently than the seed components. Scale bars = 50 μm and 5 μm (images on the right).

## Theoretical Simulations for Resistive Losses



**Figure 4.** FDTD simulation of resistive losses from basil seeds fully immersed in water exposed to monomode microwave point source operating at 8 GHz and 10 W using COMSOL software. Top and side view of the microwave cavity and seeds are shown on the left. Resistive losses from the seeds are shown as slices starting from the layer bottom of the seeds up to the layer corresponding to the plane on top of the seeds.

## Basil Seed Hydroponic Growth

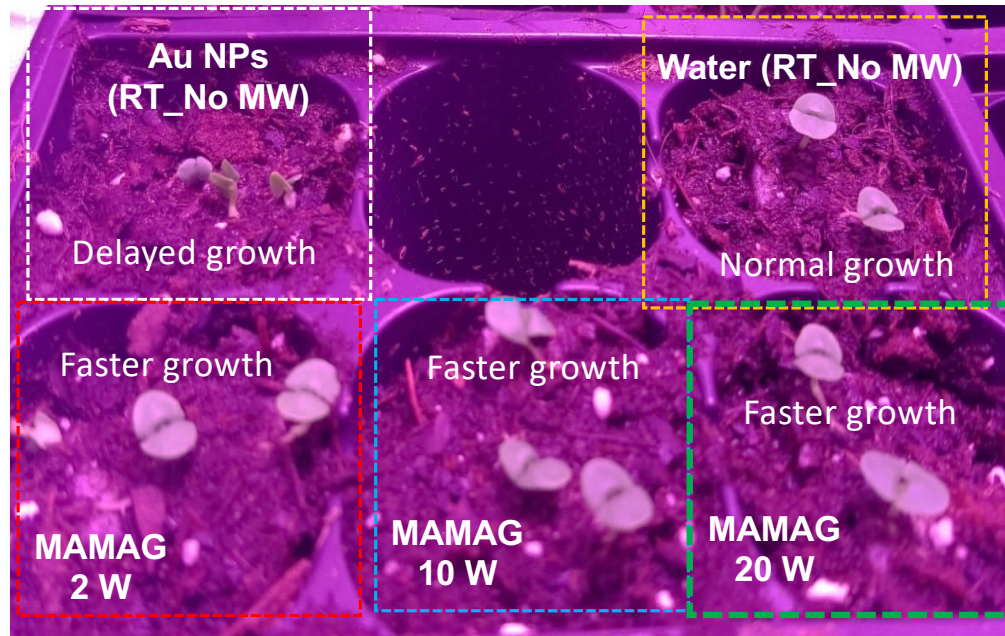


**Figure 5.** Basil seeds in hydroponic growth. Three basil seeds were kept in water without Au NPs at room temperature (No MW/RT), or kept in a solution of 20 nm Au NPs at room temperature, or microwave heated (MW) in in a solution of 20 nm Au NPs for 6 min at 10 W and placed into a modified growth system. Basil seeds in water at room temperature (no microwave heating) are used as a control sample to monitor the normal hydroponic growth.

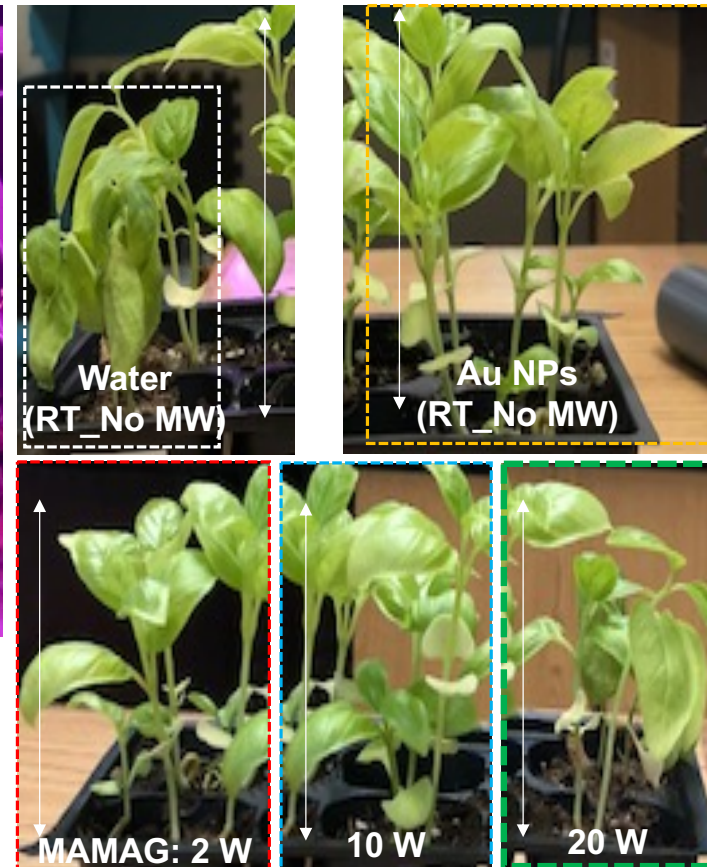


## Basil Seed Growth in Greenhouse

5 Days after planting in soil



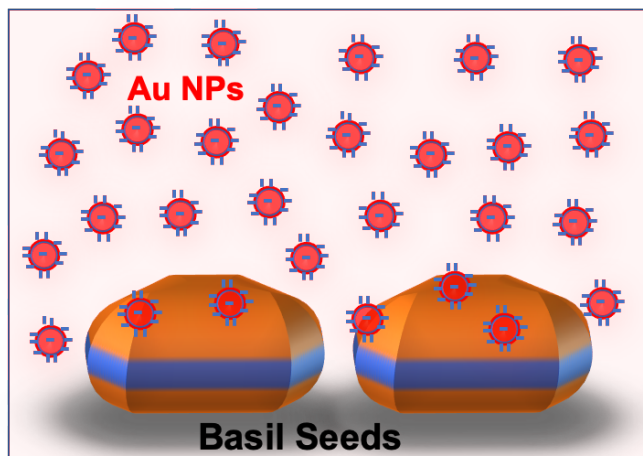
73 Days after planting in soil



**Figure 6.** Basil seed growth in soil in a greenhouse. Basil seeds were microwaved continuously for 6 minutes at 2 W, 10 W, and 20 W. Basil seeds in water only (no microwave heating) and basil seeds in 20 nm Au NPs only (no microwave) were used as control basil seeds. All pictures were taken together with the same camera. Vertical white line shows the length of basil plants = 7 cm.

TOC

**Before Microwave Heating**



**During / After Microwave Heating**

