

1 A comprehensive analysis of racial disparities in chemical biomarker concentrations in United States
2 women, 1999-2014

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25 Running Title: Racial disparities in chemical biomarkers in US women

26

27 **Abstract**

28 Background: Stark racial disparities in disease incidence among American women remains a persistent
29 public health challenge. These disparities likely result from complex interactions between genetic, social,
30 lifestyle, and environmental risk factors. The influence of environmental risk factors, such as chemical
31 exposure, however, may be substantial and is poorly understood.

32 Objectives: We quantitatively evaluated chemical-exposure disparities by race/ethnicity and age in United
33 States (US) women by using biomarker data for 143 chemicals from the National Health and Nutrition
34 Examination Survey (NHANES) 1999-2014.

35 Methods: We applied a series of survey-weighted, generalized linear models using data from the entire
36 NHANES women population and age-group stratified subpopulations. The outcome was chemical
37 biomarker concentration and the main predictor was race/ethnicity with adjustment for age, socioeconomic
38 status, smoking habits, and NHANES cycle.

39 Results: The highest disparities across non-Hispanic Black, Mexican American, Other Hispanic, and other
40 race/multiracial women were observed for pesticides and their metabolites, including 2,5-dichlorophenol,
41 o,p'-DDE, beta-hexachlorocyclohexane, and 2,4-dichlorophenol, along with personal care and consumer
42 product compounds. The latter included parabens, monoethyl phthalate, and several metals, such as mercury
43 and arsenic. Moreover, for Mexican American, Other Hispanic, and non-Hispanic black women, there were
44 several exposure disparities that persisted across age groups, such as higher 2,4- and 2,5-dichlorophenol
45 concentrations. Exposure differences for methyl and propyl parabens, however, were the starkest between
46 non-Hispanic black and non-Hispanic white children with average differences exceeding 4 folds.

47 Discussions: We systematically evaluated differences in chemical exposures across women of various
48 race/ethnic groups and across age groups. Our findings could help inform chemical prioritization in
49 designing epidemiological and toxicological studies. In addition, they could help guide public health
50 interventions to reduce environmental and health disparities across populations.

51

52 1. Introduction

53 The stark racial disparities in disease incidence and health outcomes among American women
54 remains a persistent public health challenge. For example, preterm birth incidence was found to be
55 approximately 60% higher in non-Hispanic Black women relative to non-Hispanic white women (Culhane
56 and Goldenberg 2011). Non-Hispanic Black and Hispanic women are at increased risk of being diagnosed
57 with developing dysglycemia (Marcinkevage et al. 2013) and diabetes (Cowie et al. 2009), relative to non-
58 Hispanic white women. Non-Hispanic Black women are also 2-3 times more likely to develop the most
59 aggressive subtype of breast cancer, triple negative, compared to non-Hispanic white women (Carey et al.
60 2006; Stark et al. 2010). Furthermore, relative to non-Hispanic white women, non-Hispanic Black women
61 are also 2.4 times more likely to die of breast cancer after being diagnosed with the pre-invasive lesion,
62 ductal carcinoma *in situ* (Narod et al. 2015).

63 Recent statistics from the American Cancer Society show variation in trends in breast cancer
64 incidence rates by race/ethnicity in US women from 2005-2014. Specifically, they show increasing trends
65 in breast cancer over time in Asian (1.7% per year), non-Hispanic Black (0.4% per year), and Hispanic
66 (0.3% per year) women, and stable trends in non-Hispanic white and American Indian/Alaska Native
67 women (DeSantis et al. 2017). Dementia rates also vary by race/ethnicity, with rates highest in non-
68 Hispanic black women, followed by American Indian/Alaskan native, Latina, Pacific Islander, non-
69 Hispanic white, and lowest in Asian American women (Mayeda et al. 2016). These rates vary 60% between
70 African American and Asian American women. Reproductive outcomes are also significantly different by
71 race/ethnicity, with studies reporting increased incidence of gestational diabetes in South and Central Asian
72 American women (Thorpe et al. 2005) and Black and Hispanic women (Tanaka et al. 2007). Collectively,
73 these findings suggest profound racial disparities in disease outcomes that manifest throughout the life
74 course. Understanding the etiological factors driving these health disparities is essential for informing
75 public health interventions seeking to promote health equity.

76 While health disparities are likely due to complex interactions between genetic, social, and lifestyle
77 factors, the impact of genetic factors on disease disparities appears to be minor (Braun 2007; Cooper et al.

2003; Diez Roux 2012). For example, a meta-analysis of genetic factors underlying racial disparities in cardiovascular disease failed to identify heterogeneity of genetic risk factors by race/ethnicity (Kaufman et al. 2015). These findings of a modest genetic impact on differential cardiovascular disease risk by race/ethnicity are consistent with genome-wide association studies. A study found that variants with the strongest association with blood pressure explain, in aggregate, less than 5% of the phenotypic variance (Ehret et al. 2011). Moreover, a meta-analysis of genetic risk factors and cancer disparities reported similar findings, with almost no heterogeneity in cancer risk alleles by race/ethnicity (Jing et al. 2014).

Environmental risk factors may be more influential in generating health disparities than other risk factors. For instance, estimates of environmental impacts on chronic disease suggest that 70-90% of risk is due to environmental exposures (Lim et al. 2012; Rappaport and Smith 2010). A mechanistic understanding of racial disparities in disease therefore requires a characterization of differences in environmental risk factors. In particular, differences in chemical exposures have been hypothesized to be important etiologic factors in racial disparities of disease rates (Hoover et al. 2012; Juarez and Matthews-Juarez 2018; Ruiz et al. 2018; Wang et al. 2016; Zota and Shamasunder 2017).

To investigate the influence of environmental risk factors on health disparities, the goal of this study was to conduct a comprehensive analysis of racial disparities in chemical biomarker concentrations among US women. For this, we leveraged data from the National Health and Nutrition Examination Survey (NHANES), an ongoing population-based health study conducted by the US Centers for Disease Control and Prevention (CDC). Additionally, we developed visuals to highlight differences in biomarker concentrations across races and age groups, by defining the relative magnitude of exposure disparities for individual chemicals and chemical families.

2. Methods

2.1 Study Population

NHANES is a cross-sectional study designed for collecting data on demographic, socioeconomic, dietary, and health-related characteristics in the non-institutionalized, civilian US population. For this analysis, we used the continuous NHANES data on chemical biomarkers and demographics, which were

104 collected from 1999-2014 with 82,091 participants initially. We excluded participants for not having any
105 data on chemical biomarkers ($N = 7,001$), resulting in a sample size of 75,090 study participants. Since this
106 analysis is focused on measuring chemical disparities in US women, we excluded male participants ($N =$
107 $37,010$), leading to a final sample size of 38,080 female participants. For a given chemical, we also excluded
108 participants with missing data on any of the following covariates: race/ethnicity, age, NHANES cycles,
109 poverty income ratio, cotinine levels, and urinary creatinine. Number of participants with complete data for
110 a given chemical and the listed covariates are tabulated in **Excel Table S1**. These exclusion and inclusion
111 criteria are delineated in **Figure 1**.

112 **2.2 Chemical Biomarker Measurements**

113 This section along with **Figure 1** delineate the curation process for selecting chemical biomarkers
114 to include for analysis. First, we excluded biomarkers that are not indicative of chemical exposures ($n =$
115 99). Next, we corrected for differences in chemical codenames by using a unique codename for each
116 biomarker ($n = 36$). We gave preference to lipid-adjusted data and therefore excluded non-lipid adjusted
117 chemical biomarkers ($n = 79$) when both types of data were provided for a given chemical. We replaced all
118 measurements below the limit of detection (LOD) with $LOD/\sqrt{2}$ as recommended by the CDC (CDC,
119 2009). This was to produce reasonably unbiased means and standard deviations (Hornung and Reed, 1990).
120 There were also instances in which urinary cadmium concentrations were recorded as 0 ng/mL due to
121 interference with molybdenum oxide (NCHS, 2005a, NCHS, 2005b). We replaced such values with
122 $LOD/\sqrt{2}$ if the participant's urinary cadmium level was under the LOD or otherwise excluded. We
123 calculated detection frequencies for each chemical biomarker (**Excel Table S2**) and excluded biomarkers
124 with detection frequencies of 50% or less ($n = 182$) across all study participants. Across the NHANES
125 cycles, improvements in laboratory technology can change the LOD and thus lead to differences in
126 detection frequencies by NHANES cycle (Nguyen et al. 2019). To limit bias from these changing LODs
127 over time, we calculated detection frequencies by NHANES cycle (**Excel Table S2**) for each chemical
128 biomarker and excluded measurements that showed drastic changes in the LOD (**Excel Table S3**) and

129 detection frequencies over time (Fig. 1). Measurements from given cycles for several PCBs, Dioxins,
130 Furans, Phytoestrogens, and VOCs along with Paranitrophenol, 2-naphthol, 1-pyrene and 9-pyrene
131 ($m = 449,396$) were therefore also excluded based on these criteria (**Excel Table S4**). The final dataset for
132 analysis consisted of 141 chemical biomarkers from 17 different chemical classes (**Excel Table S5**).

133 **2.3 Statistical Analysis**

134 We performed all analyses using R version 3.6.0. Given the NHANES complex sampling design,
135 we applied appropriate survey weights in our statistical models to produce estimates representative of the
136 non-institutionalized, civilian US population. To do this, we developed two databases. The first was a
137 database of codenames indicating the appropriate survey weights for each chemical biomarker and
138 NHANES cycle (**Excel Table S6**). For several of the Per- and Polyfluoroalkyl Substance (PFAS), there
139 were two different type of survey weights available within the same cycle (one for children aged 3-11 and
140 the other for participants aged 12 and older). Therefore, we developed another database of codenames
141 indicating which additional survey weights to use when generalizing these results for PFASs (**Excel Table**
142 **S7**).

143 Using multivariate regression models, we evaluated differences in biomarker concentrations in
144 blood and urine by race after log-transforming the data. We included log-transformed levels of cotinine as
145 a covariate to represent smoking (Benowitz, 1999), and creatinine levels to adjust for urine dilution and
146 flow differences (Barr et al., 2005). We modeled poverty income ratio (PIR) as a surrogate variable for
147 socioeconomic status. PIR is the ratio of household income and poverty threshold adjusted for family size
148 and inflation. First, we examined the racial differences in chemical biomarker levels by performing a series
149 of chemical-specific regression models with the main predictor being race/ethnicity (categorical), adjusting
150 for age (continuous), sex (categorical), NHANES cycle (continuous), PIR (continuous), and cotinine
151 (continuous) as described in Eq. (1):

152

$$\text{Log}_{10}(X_{\text{Chemical Concentration}}) = \beta_{\text{race/ethnicity},j}(X_{\text{race/ethnicity},j}) +$$

$$\begin{aligned} & \beta_{age}(X_{age}) + \\ & \beta_{cycle}(X_{cycle}) + \\ & \beta_{PIR}(X_{PIR}) + \\ & \beta_{cotinine}(X_{cotinine}) + \\ & \beta_{creatinine}(X_{creatinine}) + \\ & \alpha \end{aligned} \quad [1]$$

153

154 Here, $X_{Chemical\ Concentration}$ is the log-transformed, unadjusted chemical biomarker concentration for all
155 participants, X_i , where $i \in \{race/ethnicity, j, age, sex, cycle, PIR, cotinine, creatinine\}$, is the i
156 covariate for all participants, β_i is the linear regression coefficient for the i covariate, and α is the intercept.
157 $X_{race/ethnicity, j}$, where $j \in \{Mexican\ Americans, Other\ Hispanics, Non-Hispanic\ Black,$
158 $Other\ Race/Multiracial\}$ for 1999-2014, is the race covariate for comparing the j th race to the reference
159 group of Non-Hispanic Whites. For chemical biomarkers which were measured in urine, we further
160 corrected the regression models by adjusting for urinary creatinine levels (continuous). For the analyses
161 where cotinine concentration was the outcome, the regression models were not further corrected for
162 smoking. Prior to 2011, Asian Americans were categorized in Other Race/Multi-Racial category.
163 Accordingly, to evaluate chemical exposure disparities in Asian American women, we also applied Eq. 1
164 to the 2011-2014 data. Then to determine whether racial disparities are driven by differences in
165 socioeconomic status, we conducted a sensitivity analysis to observe how the race coefficients change with
166 and without adjustment for PIR in the regression models. The coefficient for j th race represents the
167 difference in log-transformed chemical biomarker concentration between the j th race and the reference
168 group of Non-Hispanic Whites. To account for multiple comparisons, we used a False Detection Rate
169 (FDR) method on the p-values of the linear regression race-coefficients (Benjamini and Hochberg, 1995).

170 To evaluate how these racial differences in chemical exposures differ by age group, we conducted
171 stratified analyses by age groups in the 1999-2014 data. We defined 4 age groups: 0-11, 12-25, 26-50, and

172 51-85. For each age group with chemical biomarker measurements, we performed a chemical specific linear
173 regression with the main predictor as race/ethnicity (categorical) and adjusted for age (continuous), sex
174 (categorical), NHANES cycle (continuous), PIR (continuous), and cotinine (continuous), stratified by age
175 group described in Eq. (2).

$$\begin{aligned} \text{Log}_{10}(X_{\text{Chemical Concentration}}[\text{age group} = k]) &= \beta_{\text{race/ethnicity},j,k}(X_{\text{race/ethnicity},j}[\text{age group} = k]) + \\ &\beta_{\text{age},k}(X_{\text{age}}[\text{age group} = k]) + \\ &\beta_{\text{cycle},k}(X_{\text{cycle}}[\text{age group} = k]) + \\ &\beta_{\text{PIR},k}(X_{\text{PIR}}[\text{age group} = k]) + \\ &\beta_{\text{cotinine},k}(X_{\text{cotinine}}[\text{age group} = k]) + \\ &\beta_{\text{creatinine},k}(X_{\text{creatinine}}[\text{age group} = k]) + \\ &\alpha \end{aligned} \quad [2]$$

177
178 Here, k is an available age group from the set of {0-11, 12-25, 26-50, 51-85},
179 $X_{\text{Chemical Concentration}}[\text{age group} = k]$ is the log-transformed, unadjusted chemical biomarker
180 concentration for all participants with ages in the k th age groups, $X_{i,k}[\text{age group} = k]$, where $i \in$
181 {*race/ethnicity, j, age, sex, cycle, PIR, cotinine, creatinine*}, is the i covariate for all participants
182 with ages with the k th age group, $\beta_{i,k}$ is the linear regression coefficient for the i covariate and k th age
183 group, and α is the intercept. $X_{\text{race/ethnicity},j,k}$, where $j \in$ {*Mexican Americans, Other Hispanics,*
184 *Non-Hispanic Black, Other Race/Multiracial*}, is the race covariate for comparing the j th race to the
185 reference group of Non-Hispanic Whites in the k th age group. To account for multiple comparisons, we
186 used a False Detection Rate (FDR) method on the p-values of the linear regression race-coefficients across
187 all age groups (Benjamini and Hochberg, 1995).

188 3. Results

189 **Table 1** displays demographic characteristics of the study population. The study population
190 includes 38,080 female study participants of ages 1-85 years, with a median age of 26. Using a series of
191 covariate adjusted regression models, we first calculated the fold-difference in chemical biomarker
192 concentrations by race across the entire study population. These regression results are presented in graphical
193 format in **Figure 2**, where the letters in the plot reflect the fold-difference in chemical biomarkers for each
194 race/ethnicity, relative to non-Hispanic white women, who made up the largest portion of the study
195 population. Full regression results for all covariates in the regression models for each covariate are
196 presented in **Excel Table S8**. Pesticides and pesticide metabolites, including 2,5-dichlorophenol, o,p'-DDE,
197 beta-hexachlorocyclohexane, and 2,4-dichlorophenol had amongst the highest average fold difference
198 across non-Hispanic Black, Mexican American, Other Hispanic, and other race/multiracial women. On
199 average, large differences by race are also apparent for personal care and consumer product compounds
200 including methyl paraben, propyl paraben, monoethyl phthalate and metals, such as mercury and arsenic.
201 Conversely, cotinine, PBDE-153, PBB-153, Equol, DEET, and bisphenol F were among the chemicals of
202 which non-Hispanic white women had the highest levels.

203 In order to more clearly visualize the differences in chemical biomarkers by race/ethnicity, we
204 generated volcano plots, which are displayed in **Figure 3**. The x-axis of these plots depicts the fold
205 difference in average chemical biomarker concentration between each race/ethnicity and non-Hispanic
206 white women. The y-axis depicts statistical significance, as reflected in the negative \log_{10} transformation of
207 the FDR-adjusted p-value from the regression analysis for that chemical biomarker, where chemicals with
208 larger values on the y-axis are more statistically significant. As shown in **Figure 3A**, non-Hispanic black
209 women have biomarker concentrations that are more than twice those of non-Hispanic white women for
210 multiple chemicals. These include 2,5-dichlorophenol, 1,4-dichlorobenzene, methyl paraben, monoethyl
211 phthalate, 2,4-dichlorophenol, and propyl paraben. The heavy metals, mercury (p-value = $1.39\text{E-}15$) and
212 lead (p-value = $1.85\text{E-}14$), are also significantly higher in non-Hispanic Black women. Conversely, levels
213 of benzophenone-3, a UV blocker used in sunscreen, are significantly higher in non-Hispanic white women
214 (p-value = $1.96\text{E-}15$). In general, concentrations of PCBs tend to be modestly elevated in non-Hispanic

215 Black women, while volatile organic compounds (VOCs) and phytoestrogen concentrations are higher in
216 non-Hispanic white women. **Figure 3B** shows relative differences in chemical biomarker concentrations
217 between Mexican American and non-Hispanic white women. Pesticides, including 2,5-dichlorophenol,
218 beta-hexachlorocyclohexane, and 2,4-dichlorophenol, along with the polycyclic aromatic hydrocarbon 2-
219 naphthol were on average higher in Mexican American women. Conversely, the smoking biomarker, cotinine
220 is significantly lower in Mexican American women (p-value = 8.23E-36). PCB levels, on average, are also
221 lower in Mexican American women, while heavy metal levels tended to be higher. Exposure patterns
222 comparing Other Hispanic and non-Hispanic white women, displayed in **Figure 3C**, showed some
223 similarities, with pesticides 2,5-dichlorophenol and p,p'-DDE elevated in Other Hispanic women. Multiple
224 PFASs, including PFOS, PFHxS, and 2-(N-methyl-PFOSA) acetate, as well as cotinine, are significantly
225 lower in Other Hispanic women. **Figure 3D** shows a distinct exposure pattern in women of other
226 race/ethnicity or multiracial women. Here, levels of heavy metals, including cadmium, mercury, and
227 multiple arsenic biomarkers, are significantly elevated relative to non-Hispanic white women. Conversely,
228 the smoking biomarkers, NNAL (p-value = 2.77E-07) and cotinine (p-value = 4.49E-4), are significantly
229 lower.

230 To understand whether socioeconomic status is a driver of racial disparities in chemical exposures,
231 we generated a series of correlation plots, comparing how the differences in chemical biomarker
232 concentrations by race/ethnicity change with the inclusion and exclusion of PIR in the regression models
233 (**Figure S1** and **Excel Table S9**). For many of the chemicals, the fold differences for comparing chemical
234 biomarker levels by race did not change drastically when including PIR as a covariate in the regression
235 models, implying that socioeconomic status is not the primary driver in explaining differences in chemical
236 exposures. However, for cotinine, PCB 194, and several chemicals used in personal care products, the
237 relative differences changed by greater than 25% when PIR was included as a covariate in the regression
238 models. This suggests that either exposure differences between races for these chemicals are mediated by
239 PIR, and/or exposure differences are explained by interactions between race and socioeconomic status. To
240 visualize differences in chemical biomarker concentrations by race across a gradient of income for a few

241 selected biomarkers, we generated violin plots of the chemical biomarker distribution stratified by
242 categories of PIR for each race/ethnicity (**Figure S2**). For benzophenone-3 and cotinine (**Figure S2A and**
243 **S2B**), the trends of biomarker concentrations across the PIR categories and the average concentrations
244 within the same PIR categories differ by race. This is similar for ethyl paraben (**Figure S2C**), but
245 differences are not as drastic. On the other hand, mercury (**Figure S2D**) along with other remaining
246 chemicals demonstrated a very different pattern from those of the previously mentioned substances. Across
247 all races, the trends across PIR categories are similar for mercury, but within the same PIR category, there
248 are differences in biomarker concentrations by race, suggesting that many chemical exposures disparities
249 by race are independent of PIR.

250 Starting in 2011, more detailed information on NHANES study participant race/ethnicity were
251 collected, including specifically identifying individuals who report Asian ethnicity. To understand whether
252 the results presented in **Figure 3D** predominantly reflect results in Asian women, who prior to 2011 were
253 categorized in other race/multi-racial category, we assessed exposure disparities specifically in the Asian
254 population. These results, presented in **Figure 4A**, show that, on average, multiple heavy metal biomarkers
255 are more than 2-fold higher relative to non-Hispanic white women, including cadmium, mercury, lead, and
256 arsenics. Additionally, the PFAS compound PFDA is significantly higher in Asian women (p-value =
257 3.82E-06), while cotinine (p-value = 1.88E-05) and biomarkers of phosphate flame retardants (Bis(1,3-
258 dichloro-2-propyl) phosphate p-value = 5.41E-3; Dibutyl phosphate p-value = 6.76E-4; Diphenyl phosphate
259 p-value = 3.27E-3) are significantly lower. We also calculated whether there were significant disparities in
260 chemical biomarker concentrations in women of other or multi-race after excluding Asian women. **Figure**
261 **4B** suggests relatively few differences in this regard, confirming that the other race effect in **Figure 3D** is
262 indeed associated with Asian women. Full regression results across all covariates for the 2011-2014 data
263 are presented in **Excel Table S10**.

264 We have previously shown dramatic differences in the chemical “exposome” by age in NHANES
265 study participants, not stratified by gender or race (Nguyen et al. 2019). Here, we tested for differences in
266 chemical biomarkers by race, after stratifying by age group. **Figure 5** displays these results across the entire

267 study population from 1999-2014. **Excel Tables S11-S14** includes the results for all regression analyses
268 stratified across each of the four age groups. Blue colors reflect chemicals where levels are higher in non-
269 Hispanic white women, while red colors reflect chemicals that are of higher concentration in women of the
270 labeled race/ethnicity. Here, there appear to be exposure disparity patterns that persist across age groups –
271 such as higher 2,4- and 2,5-dichlorophenol concentrations in Mexican American, Other Hispanic, and non-
272 Hispanic black women. Differences in 1,4-dichlorobenzene concentrations are consistent across age groups,
273 although this biomarker was not measured in the youngest individuals. Heavy metal concentrations are
274 elevated in women of other race across age groups. Some exposure patterns differ by age, however. For
275 example, differences in methyl and propyl paraben are most apparent between young non-Hispanic black
276 and non-Hispanic white women less than 12 years old. Increased levels of phosphate flame retardants and
277 the insect repellent DEET in non-Hispanic white women are the most evident in women less than 12 years
278 of age. Similarly, higher relative concentrations of benzophenone-3, bisphenol A, and bisphenol F occur in
279 non-Hispanic white women less than 12. Elevated PCB levels in non-Hispanic black women shown in
280 **Figure 3A** are most evident in women greater than 51 years of age. Overall, these results highlight racial
281 exposure disparities that are either stable or that vary across age groups.

282 **4. Discussion**

283 Based on population based chemical biomonitoring generated as part of the 1999-2014 NHANES,
284 we performed a comprehensive analysis of racial disparities in biomarker concentrations of 141 chemicals
285 in 38,080 participants. Specifically, we quantified the relative magnitude of racial disparities for individual
286 chemicals and chemical families while utilizing appropriate regression weightings. This helped ensure that
287 the results were as generalizable to the entire US population. These results highlighted striking differences
288 in chemical biomarker exposure patterns by race/ethnicity, independent of other demographic factors such
289 as socioeconomic status. In particular, exposure patterns of pesticides, heavy metals, tobacco smoke
290 associated compounds, and chemicals found in personal care products are found to be most disparate across
291 race/ethnic groups. Stratified analyses revealed exposure patterns that persisted across age groups. For
292 example, this was apparent in heavy metals exposure for women who identify as other race or multiracial,

293 as well as in age-specific exposure patterns, such as elevated PCB, dioxin, and dibenzofuran exposure in
294 older non-Hispanic black women. In some cases, average differences in chemical biomarker concentrations
295 between race/ethnic groups exceeded 400%, such as for urinary propyl or methylparaben concentrations
296 between the youngest non-Hispanic Black and non-Hispanic white women. These findings contextualize
297 racial disparities in chemical exposures across US women and highlight the vast differences in chemical
298 exposures between demographic groups with well characterized disparities in health outcomes.

299 Environmental injustice is the disproportionate exposure of individuals of color, lower
300 socioeconomic status, or other politically disadvantaged groups to toxic chemicals in food, air, consumer
301 products, at the workplace, or in their communities (Brulle and Pellow 2005). Disproportionate chemical
302 exposures have been hypothesized to be important drivers of health disparities, including obesity and
303 neurodevelopmental outcomes (Landrigan et al. 2010). While the primary goal of this study was to quantify
304 and compare chemical exposure disparities across racial/ethnic groups, independent of income, others have
305 evaluated combined income and race related disparities in exposure. For instance, one analysis compared
306 geometric mean concentrations of 228 chemical biomarkers between six groups stratified by income and
307 race in NHANES and identified 37 chemicals as likely contributing to environmental justice (Belova et al.
308 2013). Some of these chemicals, including cotinine, lead, 2,4- and 2,5-dichlorophenol, methyl paraben,
309 and propyl paraben, were associated with the highest disparities across race/ethnic group in the present
310 study. We also compared chemical exposures disparities across racial/ethnic groups with and without
311 adjustment for income and found that cotinine, PCB 194, methyl mercury, and chemicals used in personal
312 care products such as benzophenone-3, the parabens, and triclosan show disparities across both race and
313 socioeconomic status. However, for most of the studied chemicals, differences in chemical exposures were
314 not driven by socioeconomic status but were instead primarily associated with race/ethnicity. Furthermore,
315 a study of racial and social disparities in exposure to BPA and PFAS examined differences in biomarker
316 concentrations in NHANES study participants (Nelson et al. 2012). The concentrations of the four PFAS
317 chemicals examined, PFOA, PFOS, PFNA, and PFHxS, were inversely associated with household income,
318 while BPA concentrations were higher in individuals who reported low food security (Nelson et al. 2012).

319 Here, we identified that, independent of socioeconomic status, as assessed by poverty-income ratio, non-
320 Hispanic white women had the highest concentrations of PFOA, while non-Hispanic Black and other
321 race/multiracial women had the highest concentrations of PFDA. Major routes of exposure to PFAS
322 compounds include contaminated drinking water (Hu et al. 2016), diet (Schecter et al. 2010), and
323 occupational routes (Laitinen et al. 2014). BPA concentrations were not strikingly different by race in our
324 study, but non-Hispanic Black women had, on average, 93% higher BPS concentrations than non-Hispanic
325 white women. Common routes of exposure to BPA and other bisphenol analogues are diet, thermal paper,
326 and personal care products (Chen et al. 2016). Further research is necessary to identify the major routes of
327 exposure which are driving racial disparities in PFAS and bisphenol chemicals biomarker concentrations.

328 The findings of highly elevated monoethyl phthalate and methyl and propyl paraben concentrations
329 in the non-Hispanic Black women is consistent with a personal care product route of exposure. A study
330 assessing the chemical composition of hair products used by Black women consistently identified high
331 levels of cyclosiloxanes, parabens, and the fragrance carrier diethyl phthalate (Helm et al. 2018). In our
332 study, the concentrations of the diethyl phthalate metabolite monoethyl phthalate were approximately 78%
333 higher on average in non-Hispanic black women of all ages relative to non-Hispanic white women, and
334 122% higher in non-Hispanic black women less than 12 years of age. This is concerning, since urinary
335 concentrations of monoethyl phthalate have been positively associated with odds of developing breast
336 cancer in a case-control study of women from Northern Mexico (López-Carrillo et al. 2010). Differences
337 in concentrations of methyl and propyl paraben biomarkers were among the highest observed in this study,
338 particularly for the youngest non-Hispanic Black women. These chemicals have been used as preservatives
339 in personal care products, pharmaceuticals, and food additives, and have been found to promote cell growth
340 through multiple mechanisms, including estrogenicity (Gonzalez et al. 2018, 2019; Okubo et al. 2001) and
341 epidermal growth factor receptor signaling (Pan et al. 2016). Particularly relevant to our findings of the
342 greatest methyl and ethyl paraben disparities in the youngest non-Hispanic Black women was the finding
343 that early life paraben exposures can alter developing mammary gland morphology and induce gene
344 expression that resembles an early cancer-like state (Gopalakrishnan et al. 2017). Use of hair products has

345 been identified as a potential risk factor for breast cancer in non-Hispanic Black women (Stiel et al. 2016).
346 Further research is needed, however, to determine whether early-life exposure to potentially estrogenic
347 compounds, like parabens, can induce biological alterations that increase risk of estrogen receptor negative
348 breast cancers.

349 One of the most apparent disparities in chemical biomarker concentrations by race was with the
350 compounds 2,4-dichlorophenol, 2,5-dichlorophenol, and 1,4-dichlorobenzene. 1,4-dichlorobenzene is used
351 as a disinfectant, pesticide, and deodorant. 2,5-dichlorophenol is a metabolite of 1,4-dichlorobenzene, while
352 2,4-dichlorophenol is a metabolite of the antimicrobial triclosan or other pesticides. Elevated concentrations
353 of these chemicals in non-Hispanic Black individuals has been noted previously (Belova et al. 2013; Ye et
354 al. 2014) The concentrations of these three chemicals were up to 350% higher on average in non-Hispanic
355 Black women, relative to non-Hispanic white women, and also elevated in Mexican American and Other
356 Hispanic women. Importantly, these exposure disparities were consistent across all age groups. While 2,4-
357 dichlorophenol concentrations were significantly elevated in non-Hispanic Black and Hispanic women,
358 urinary triclosan levels were not significantly different by race/ethnicity. This suggests that either triclosan
359 is not the main chemical exposure that explains the differences in concentrations of 2,4-dichlorophenol or
360 that there are differences in metabolism and excretion rates by race, which is less likely. 1,4-
361 dichlorobenzene exposure has been associated with altered thyroid biomarkers in NHANES (Wei and Zhu
362 2016), altered immunologic and liver function parameters in occupationally exposed workers (Hsiao et al.
363 2009), and altered sperm production and increased prostate weight in exposed rats (Takahashi et al. 2011).
364 Understanding and mitigating exposure to these chemicals is therefore of importance to reduce disparate
365 risk of these health outcomes.

366 Heavy metals were among the chemicals most consistently different across racial/ethnic groups. In
367 particular, women who identified as other race or multiracial had the highest concentrations of multiple
368 metals, including cadmium, mercury, arsenics, lead, and manganese. Focusing on data from NHANES
369 2011-14, we identified that these elevated metals concentrations were restricted to women who identified
370 as Asian. This is consistent with a previous finding of increased concentrations of a subset of these metals

371 in Asian NHANES participants (Awata et al. 2017). Furthermore, elevated levels of mercury, lead, and
372 arsenics were also identified in non-Hispanic Black women, relative to non-Hispanic white women.
373 Mexican American women had elevated levels of uranium, lead, mercury, arsenics, and cadmium, while
374 Other Hispanic women had higher concentrations of mercury, arsenics, and cadmium than non-Hispanic
375 white women. Non-Hispanic white women, however, had higher concentrations of urinary barium. Previous
376 research has linked diet, occupation, education level, and smoking status to elevated metals exposure
377 (Awata et al. 2017), in addition to housing (Jacobs et al. 2013), air pollution (Suvarapu and Baek 2016),
378 and contaminated water (Pieper et al. 2017). The well characterized toxicity of heavy metals exposure, even
379 at low doses, make identifying and ameliorating heavy metal exposures a top priority for addressing
380 environmental health disparities.

381 The oldest non-Hispanic Black women in our study had consistently higher concentrations of
382 persistent organic pollutants, including dioxins, dibenzofurans, PCBs, and DDT metabolites. This is
383 consistent with a previous report of non-Hispanic black individuals having an increased risk of having
384 multiple persistent organic pollutants detectable their blood (Pumarega et al. 2016) or higher average levels
385 of PCBs (Xue et al. 2014). Biomarkers of persistent organic pollutants were quantified on an individual
386 (non-pooled) basis in the 1999-2004 NHANES cycles. Elevated concentrations of these pollutants, such as
387 the DDT metabolite, DDE, have been associated with an increased risk of breast cancer (Wolff et al. 1993).
388 A lack of disparities, and decreasing concentrations of these chemicals in younger individuals over time,
389 generally reflect a public health success in decreasing population exposures to these toxic compounds
390 (Nguyen et al. 2019). The long half-life of these chemicals suggests that the detected biomarkers
391 predominantly reflect historical exposures. This could, however, be of substantial importance for children
392 of non-Hispanic Black women, who could have been exposed to disproportionately high levels of these
393 chemicals in the womb or early in childhood. For example, *in utero* exposure to the pesticide, DDT, has
394 been associated with an increased risk of breast cancer in adulthood. Specifically, women in the highest
395 quartile of *in utero* DDT exposure were found to have a 3.7-fold increased risk of developing breast cancer
396 relative to women in the lowest quartile of exposure (Cohn et al. 2015). Prenatal exposure to organochlorine

397 compounds has also been associated with decreased lung function later in life (Hansen et al. 2016), risk of
398 infection in childhood (Dewailly et al. 2000), attention deficit hyperactivity disorder (Sagiv et al. 2010),
399 and obesity (Mendez et al. 2011). If these effects of elevated early life persistent organic pollutant exposure
400 last throughout the life course, there could be continued adverse health consequences that manifest in those
401 exposed for the foreseeable future.

402 Our study had important limitations. First, the cross-sectional nature of NHANES allows only a
403 single biomarker measurement per individual. Moreover, since the half-lives of the biomarkers assessed in
404 this study are highly variable (Nguyen et al. 2019), the precision of estimates of long-term exposure largely
405 varies across chemical family. Additionally, this study was not able to assess geographic variation in
406 exposure. Others have identified that persistent organic pollutant exposures in the NHANES cohort varies
407 geographically, with higher DDT metabolite concentrations in individuals residing in the West, and
408 elevated PCB concentrations in individuals residing in the Northeast (Wattigney et al. 2015). Future work
409 is needed to precisely characterize exposure “hot spots,” in order to design intervention studies to reduce
410 exposure disparities. Our study also focused on identifying average differences in biomarker
411 concentrations. By ignoring the extremes of these distributions, we have likely not considered individuals
412 at greatest risk of developing adverse health outcomes. Similarly, our analyses were limited by low
413 detection rates, with 182 chemicals not meeting our inclusion threshold of at least 50% detection in the
414 study population. A more in-depth analysis of differences in detection frequency by race/ethnicity could
415 identify additional chemicals with significant racial disparities. For chemical biomarkers measured in urine,
416 variations in the concentration of urinary creatinine, used as a correction factor for urine dilution, potentially
417 confounds our comparison of exposures between individuals of different races. This is because increased
418 average concentrations of urinary creatinine have been identified for non-Hispanic Black individuals,
419 relative to Mexican American and non-Hispanic white individuals (Barr et al. 2005). While we adjusted for
420 urinary creatinine as a covariate in our regression models, the still may be residual confounding. The large
421 number of chemicals assessed also precluded an in-depth characterization of the various routes of exposure
422 of individual chemicals – this is undoubtedly an essential future direction of research to develop strategies

423 to eliminate exposure disparities. Finally, while we performed analyzed all chemical biomarkers available
424 from NHANES 1999-2014, these chemicals only represent a small proportion of the over 80,000 chemicals
425 estimated to be used in commerce in the United States. Future studies could benefit from an unbiased
426 metabolomics approach to identify disparities in chemical exposures which are not captured in NHANES.

427 The persistent health disparities between women of different races/ethnicities makes understanding
428 the etiological drivers of these disparities a pressing public health issue. A recent commentary highlighted
429 a lack of knowledge regarding the molecular underpinnings of health disparities. It described how the vast
430 majority of genome sequencing data had been generated in populations of European ancestry (Sirugo et al.
431 2019). Environmental exposures, however, are hypothesized to be the major driving risk factors for a vast
432 suite of complex diseases (Rappaport and Smith 2010). Even when genetic data has been generated in an
433 equitable fashion, understanding gene-environment interactions and complex disease phenotypes will still
434 require in-depth quantification of environmental exposures. In this study, we have comprehensively
435 identified differences in biomarker of chemical exposure across women of various race/ethnic groups and
436 across age groups. These findings can guide future efforts to understand chemical impacts on health
437 disparities by helping to prioritize chemicals for assessment in epidemiological studies. Additionally,
438 chemicals as identified as highly disparate here can be further prioritized for toxicological assessment
439 relevant to disease outcomes of interest. Finally, these findings can inform public health interventions
440 designed to reduce chemical disparities and promote health equity across the population.

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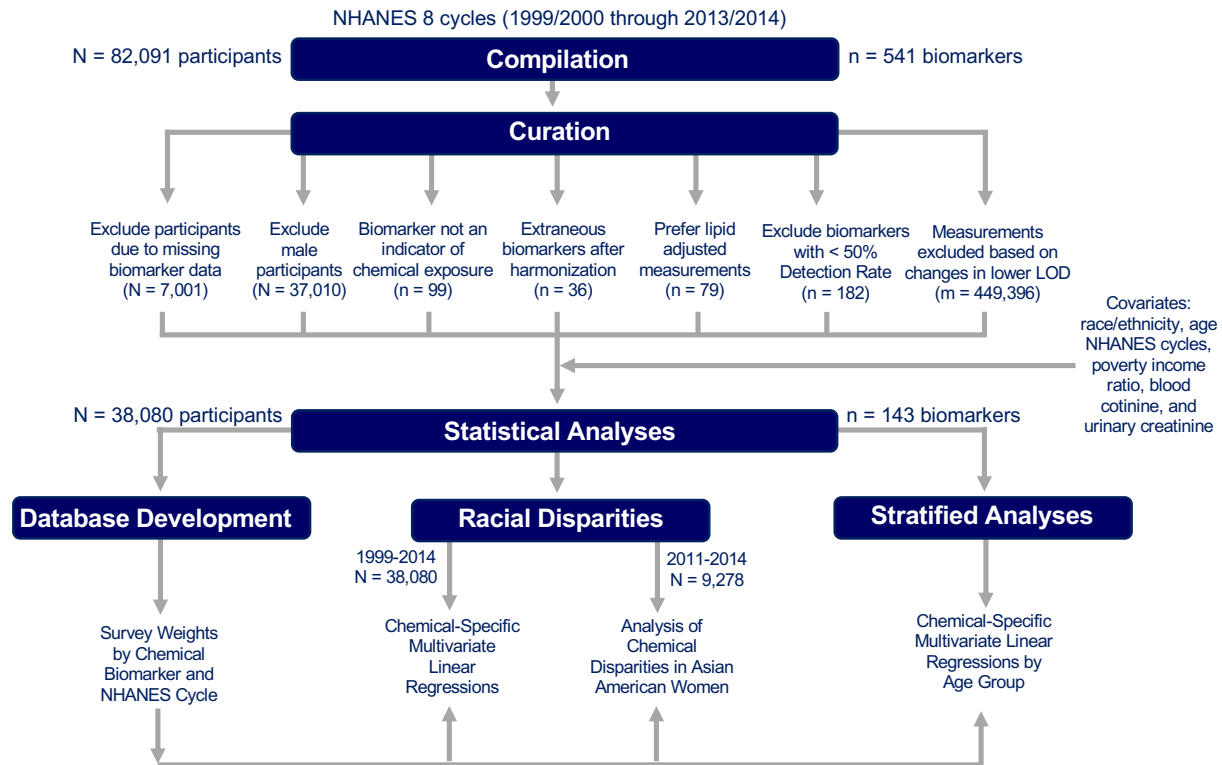
619 **Tables**
 620 **Table 1.** Demographic characteristics of the study population.
 621

CATEGORICAL					
Age	N (%)	Cycle	N (%)	Race/Ethnicity (%)	N (%)
0-11	9392 (24.66)	1999-2000 (Cycle 1)	4535 (11.91)	Mexican American	8760 (23.00)
12-25	9555 (25.09)	2001-2002 (Cycle 2)	5127 (13.46)	Other Hispanic	2949 (7.74)
26-50	9330 (24.50)	2003-2004 (Cycle 3)	4732 (12.43)	Non-Hispanic White	14384 (37.77)
51-85	9803 (25.74)	2005-2006 (Cycle 4)	4834 (12.69)	Non-Hispanic Black	9116 (23.94)
		2007-2008 (Cycle 5)	4628 (12.15)	Other Race	2871 (7.54)
		2009-2010 (Cycle 6)	4946 (12.99)		
		2011-2012 (Cycle 7)	4493 (11.80)		
		2013-2014 (Cycle 8)	4785 (12.57)		

CONTINUOUS					
	N measured (% of population)	5 th %tile	Median	Mean (SD)	95 th %tile
Age (years)	38080 (100)	2	26	32.1 (24.2)	77
PIR (-)	34968 (91.83)	0.29	1.73	2.2 (1.6)	5.00
Cotinine (ng/mL)	31699 (83.24)	0.011	0.045	29.9 (91.4)	245.00
Creatinine (mg/dL)	32314 (84.86)	22.00	102.00	115.9 (76.6)	263.00

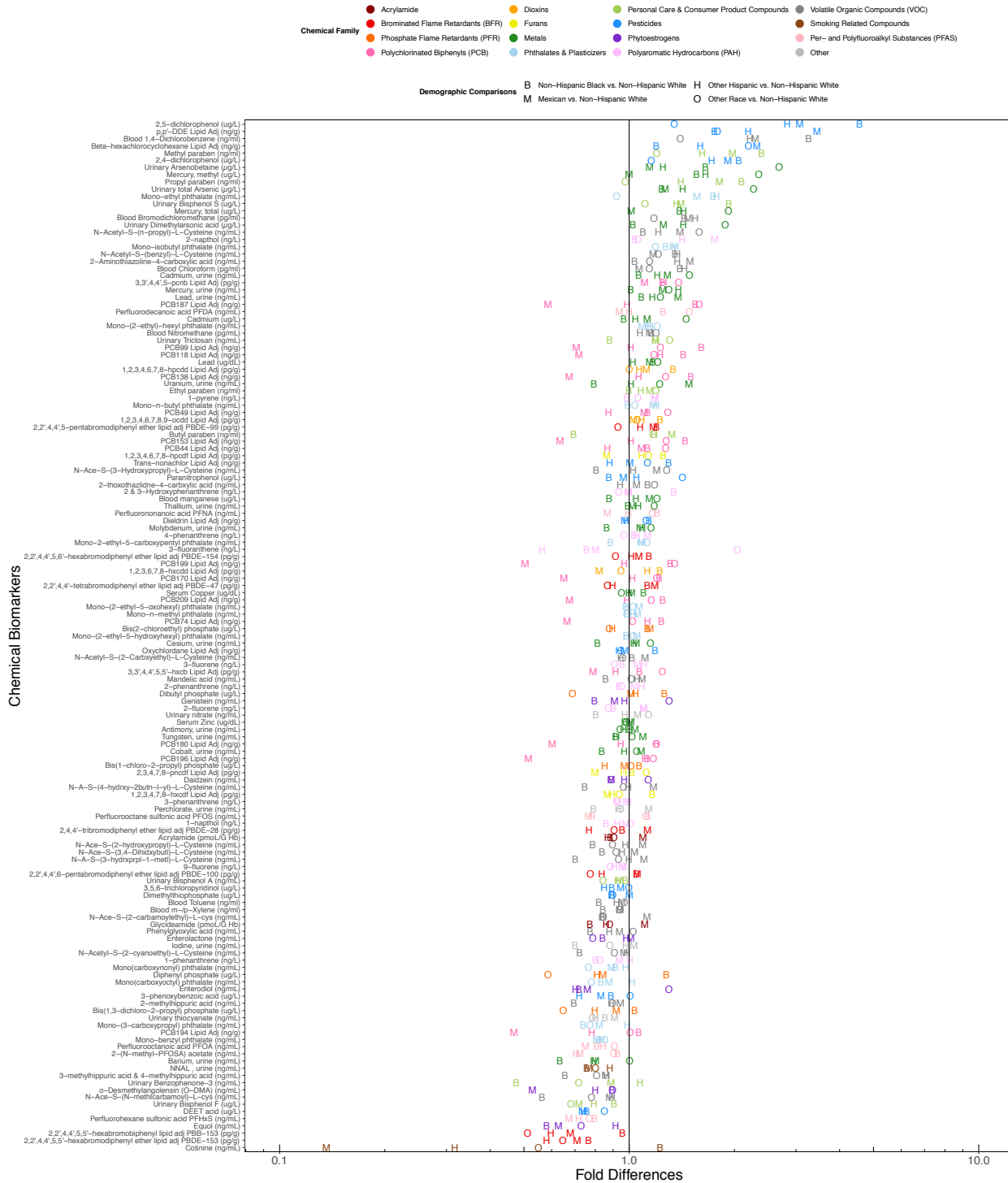
622 **Figure legends**

623 **Figure 1. Dataset compilation and cleaning workflow.**



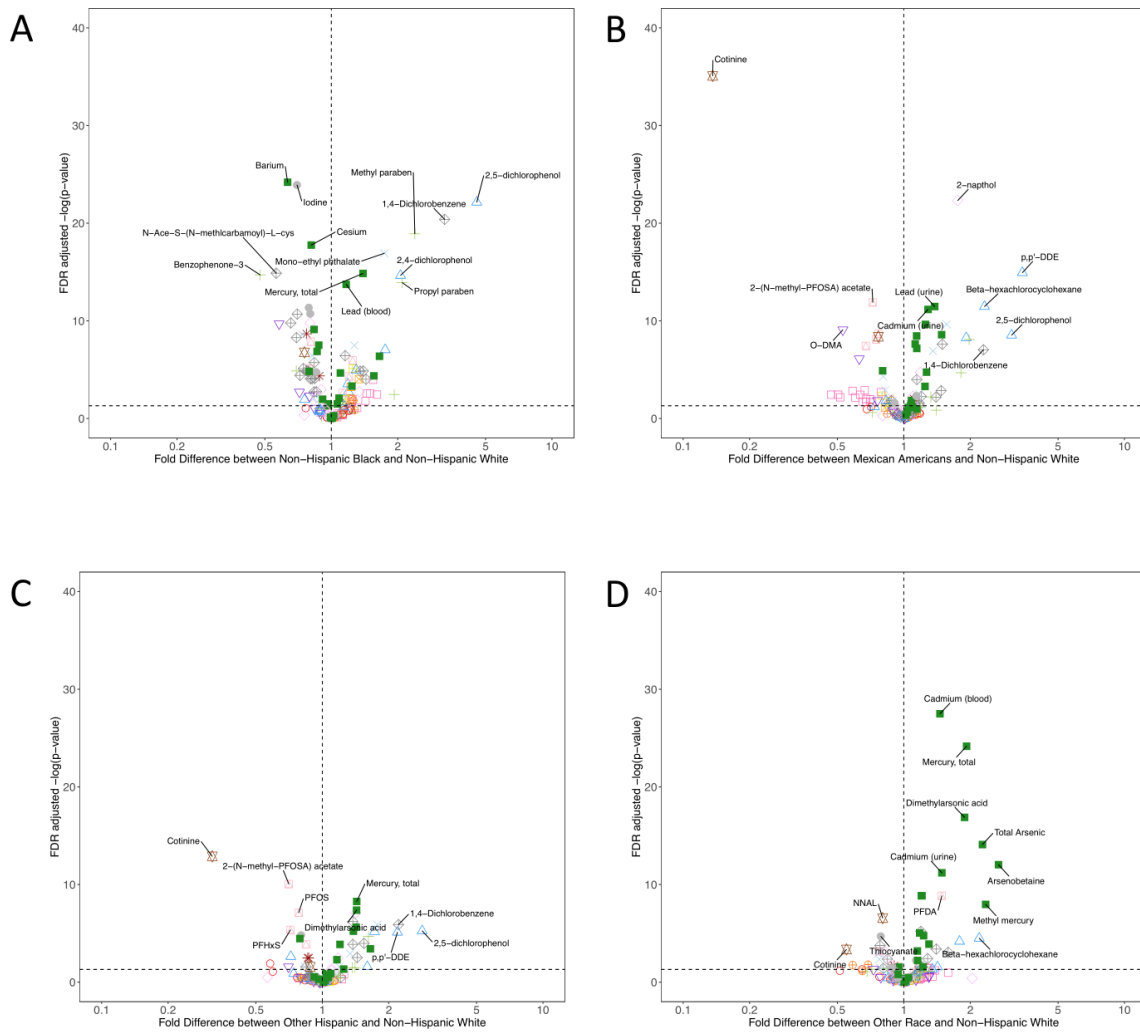
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625 **Figure 2.** Alphabet soup plot displaying the covariate adjusted fold differences in chemical biomarker
 626 concentration by race, ranked by the average difference with non-Hispanic White individuals. Colors
 627 represent the chemical families. Shapes represent the comparison between a given race and non-Hispanic
 628 White individuals.



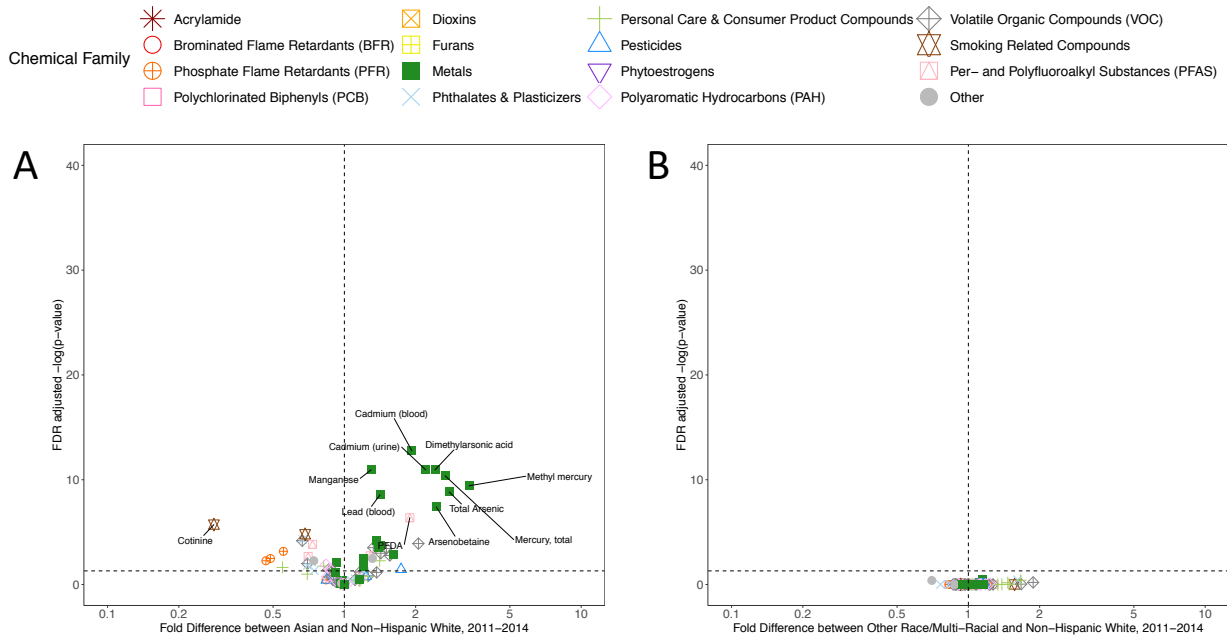
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631 **Figure 3.** Volcano plots representing the significance of the covariate-adjusted differences in chemical
 632 biomarker concentrations between non-Hispanic white women and (A) non-Hispanic Black women, (B)
 633 Mexican American women, (C) Other Hispanic women, and (D) Other race/multiracial women. Color and
 634 shapes represent the chemical families.



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 636

637 **Figure 4.** Volcano plots representing the significance of the covariate-adjusted differences in chemical
 638 biomarker concentrations between non-Hispanic white women and (A) Asian women, and (B) other race
 639 /multiracial women in NHANES 2011-2014. Colors and shapes represent the chemical families.



640
 641

642 **Figure 5.** Heatmap displaying covariate adjusted fold differences in chemical biomarker concentrations by
 643 race, relative to non-Hispanic white women, stratified by age group and chemical family. Color reflects the
 644 log₂ fold difference in chemical biomarker concentration. Biomarkers in grey color were not measured in
 645 that age group.

