1	Genetic variation in parental effects contribute to the evolutionary potential of prey
2	responses to predation risk
3	
4	Natasha Tigreros <sup>1*</sup>
5	Anurag A. Agrawal <sup>1,2</sup>
6	Jennifer S. Thaler <sup>1,2</sup>
7	
8	<sup>1</sup> Department of Entomology, Cornell University, Ithaca, New York
9	
10	<sup>2</sup> Department of Ecology and Evolutionary Biology, Cornell University, Ithaca, New York
11	
12	*CORRESPONDING AUTHOR: Natasha Tigreros <u>ntigreros@alumni.tufts.edu</u>
13	
14	
15	
16	
17	
18	COMPETING INTERESTS: We have no competing interests.
19	
20	
21	
22	

# 23 ABSTRACT

24	Despite the ubiquity of parental effects and their potential impact on evolutionary dynamics,
25	their contribution to the evolution of ecologically relevant adaptations remains poorly
26	understood. Using quantitative genetics, here we demonstrate that parental effects contribute
27	substantially to the evolutionary potential of larval antipredator responses in a leaf beetle
28	(Leptinotarsa decemlineata). Previous research showed that larger L. decemlineata larvae elicit
29	stronger antipredator responses, and mothers perceiving predators improved offspring responses
30	by increasing intraclutch cannibalism –an extreme form of offspring provisioning. We now
31	report substantial additive genetic variation underlying maternal ability to induce intraclutch
32	cannibalism, indicating the potential of this adaptive maternal effect to evolve by natural
33	selection. We also show that paternal size, a heritable trait, impacted larval responses to
34	predation risk, but that larval responses themselves had little additive genetic variation. Together,
35	these results demonstrate how larval responses to predation risk can evolve via two types of
36	parental effects, both of which provide indirect sources of genetic variation for offspring traits.
37	
38	
39	
40	KEYWORDS: transgenerational plasticity, maternal effects, inducible defenses, cannibalism,
41	predation risk
42	
43	
A A	

## 45 **1. INTRODUCTION**

Parental effects, which are widespread in animals and plants, provide an important source of 46 47 variation in offspring phenotype and fitness complementing that due to the direct inheritance of genes (Lande and Kirkpatrick 1990; Mousseau and Fox 1998; Wolf et al. 1998; Räsänen and 48 Kruuk 2007). From the provisioning of parental care to the transfer of hormones and antibodies 49 50 to young, parents alter the phenotype of their offspring, sometimes in an adaptive manner. Indeed, parental effects have been recognized as an important component of phenotypic variation 51 that may facilitate rapid evolutionary responses to a number of ecological stressors (Mousseau 52 and Fox 1998; Räsänen and Kruuk 2007; Donelson et al. 2018). Yet, predictions on the 53 evolutionary consequences of parental effects are complicated by the fact that parental effects 54 55 may be themselves shaped by the environmental conditions that parents experience, by genetic differences among parents, and by the interaction of these two (McAdam et al. 2014). While 56 environmental and genetic influences in the parental generation should strongly impact 57 58 evolutionary dynamics –increasing vs. decreasing trait response to selection-, little empirical 59 work to date has partitioned their contribution to variation in offspring traits (reviewed in 60 Räsänen & Kruuk 2007), especially those involving responses to natural ecological stressors. 61

Quantitative assessments of parental effects were initially included in quantitative genetic studies with the sole purpose of controlling for non-genetic sources of variation in offspring (Falconer and Mackay 1996; Lynch and Walsh 1998; Futuyma 2009). Nonetheless, it is now recognize that these effects often reflect genetic differences among parents, and therefore can evolve in response to selective forces occurring in both the parental and the offspring generation (Räsänen and Kruuk 2007). Importantly, genetic variation in parental effects provides an additional source of genetic variation that would facilitate evolution of offspring traits that hold little additivegenetic variation (Räsänen and Kruuk 2007).

70

71 Parental effects on offspring often reflect environmental hardships that the parents experienced, including limited food, extreme weather, and a high risk of predation (Mousseau and Dingle 72 1991; Mousseau and Fox 1998). The triggering of environmental parental effects may reflect a 73 passive consequence of stress or the resource environment that parents experience, or may 74 involve adaptive responses counter to those conditions. Research over the last two decades 75 76 indicates that parental effects can function as a form of adaptive transgenerational plasticity – commonly referred as "anticipatory parental effects" (Wade 1998; Agrawal et al. 1999; 77 Galloway and Etterson 2007; Marshall and Uller 2007; Love and Williams 2008; Sheriff and 78 Love 2013). Here, parents improve their offspring's fitness by matching the offspring's 79 phenotype to environmental challenges they will likely experience (e.g. Marshall & Uller 2007; 80 Sheriff & Love 2013). Despite growing evidence on the adaptive nature of parental effects on 81 82 offspring (Agrawal et al. 1999; Sheriff et al. 2010; Storm and Lima 2010; Jensen et al. 2014), evidence of genetic variation in anticipatory parental effects (i.e. maternal genotype by 83 environment interaction) is scarce in both animals (but see Fox et al. 1999) and plants (Galloway 84 2005). As a consequence, support for the evolutionary potential of anticipatory parental effects is 85 to date limited (reviewed in Wade 1998; Räsänen and Kruuk 2007; McAdam et al. 2014). 86 87

Parental effects have been found to be important determinants of traits associated with predator
avoidance in a number of species. For example, in a recent study in a leaf beetle (*Leptinotarsa decemlineata*), we demonstrated that variation in larval responses to predation risk was partially

determined by an anticipatory parental effect. When experiencing predation risk, larger
hatchlings were found to elicit stronger antipredator responses –measured as greater feeding
reductions in the presence of predators. Remarkably, mothers increased offspring provisioning
by inducing egg cannibalism within their clutches after detecting a high risk of predation. As a
consequence, cannibalistic offspring –being larger and in better nutritional condition than their
non-cannibal siblings– exhibited stronger responses to predation risk (Tigreros *et al.* 2017).

Here, we use classic quantitative genetics to examine the relative importance of parental effects 98 for traits associated to predator avoidance in L. decemlineata larvae, including decreased 99 foraging activity (leaf consumption) and increased assimilation efficiency. Specifically, we first 100 estimate the contributions of maternal effects  $(V_M)$ , relative to that of additive genetic  $(V_A)$  and 101 102 environmental effects (V<sub>E</sub>), for larval responses to predation risk. Second, we test if larval responses to predation risk –which are known to depend on the larva's initial size–, are 103 influenced by variation in maternal or paternal body size, a key trait known to impact offspring 104 105 phenotype in many systems (Fox 1994; Bernardo 1996a; Fox and Czesak 2000; Bennett and Murray 2014). Finally, we estimate the relative contribution of additive genetic and 106 environmental variances (V<sub>A</sub> and V<sub>E</sub>) of maternal responses to predation risk, including changes 107 in intraclutch cannibalism –a mechanism linked to an anticipatory maternal effect. 108

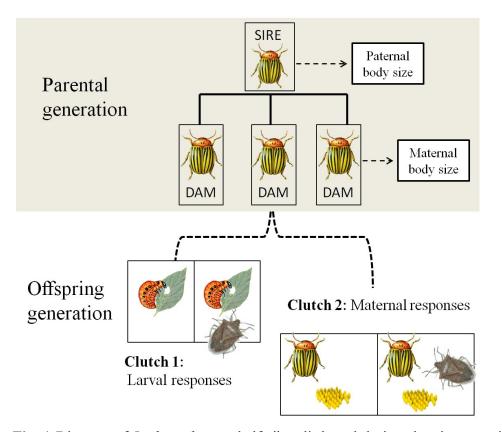
109

### 2. METHODS

# 110 (a) Breeding design

111 To estimate the relative contribution of parental effects to larval responses to predation risk we112 used a half-sib design (Falconer and Mackay 1996) (see Fig. 1). The experiment was initiated

with 25 females collected from a field population in Ithaca, NY, which were allowed to lay eggs 113 in the laboratory; their offspring, once mature, were considered the "parental generation" (Figure 114 1) from which sires and dams were selected. All sires, dams, and offspring from the different 115 116 sire by dam crosses were reared separately from birth –which minimizes common environmental effects- and were maintained in standardized conditions, fed with Solanum tuberosum L (Yukon 117 Gold variety) with 18-L : 6-D photoperiod and corresponding temperatures of 23 : 21 °C. The 118 119 half-sib families were initially established with 22 males (sires) each randomly mated to three unrelated females (dams). Sires that failed to inseminate at least two females were excluded and 120 final analyses included fewer sires and dams, which is specified in each section below. 121



**Fig. 1** Diagram of *L. decemlineata* half-sib split-brood design showing one sire family. Nineteen sires were each mated to three virgin females (dams). The offspring of these three dams were used to investigate parental effects on plastic responses to predation risk: predator-free (P-Free) vs. predation risk (P-Risk). First, we examined the overall contribution of maternal effects ( $V_M$ ), relative to the additive genetic and environmental components ( $V_A$  and  $V_E$ ), to larval anti-predator responses: feeding reductions and increases in assimilation efficiency. Second, we compared how paternal and maternal body size influenced such larval responses to predation risk. Finally, using adult-mated-females, (from Clutch 2), we examined if there was additive genetic variation ( $V_A$ ) underpinning maternal response to predation risk. Maternal responses included changes in clutch size, proportion of viable offspring, and levels of intraclutch cannibalism; the last of these (increases in intraclutch cannibalism), is known to act as an adaptive maternal effect by improving larval anti-predator responses (Tigreros et al. 2017).

123

# 124 (b) Maternal effects on larval responses to predation risk

In a first step, we followed a variance partitioning strategy to quantify the amount of variance in 125 126 offspring's traits -larval leaf consumption and assimilation efficiency- that is explained by maternal identity (V<sub>M</sub>), while accounting for the contributions of additive genetic inheritance 127 (V<sub>A</sub>) and environmental variances (V<sub>E</sub>) (McAdam et al. 2014). Final analysis included 19 sires 128 129 mated to 47 dams (with varying numbers of dams per sire). We estimated larval plastic responses to predation risk by measuring changes in larval feeding (leaf consumption) and assimilation 130 efficiency in response to predation risk. To do this, six eggs from each maternal family (within 131 the same clutch) were separated right before hatching and kept in individual 266 ml cups, which 132 prevented intraclutch cannibalism and its influences on larval responses (Tigreros et al. 2017). 133 134 Additionally, to control for potential effects due to hatching asynchrony within and among families, we recorded variation in egg pigment levels, Low, Medium, and High, as described in 135 Tigreros et al. (2017). 136

137

Hatchlings from half of each maternal family were kept with a sham predator ("predation risk" 138 environment), while the remaining siblings were kept without it, hence experiencing a "predator-139 free" environment (Fig. 1). Sham predators consisted of adult male *Podisus maculiventris* –a 140 generalist stink bug that commonly feeds on L. decemlineata larvae- whose stylet's terminal 141 segment had been removed. While these altered stink bugs are no longer able to kill the beetle 142 larvae, previous studies have shown that their behaviour and lifespan do not significantly differ 143 from that of unaltered predators (Griffin and Thaler 2006; Thaler et al. 2012; Kaplan et al. 2014). 144 145 Larvae were kept in the predator-free and predation risk environments for a total of three days

(4- day old larvae). Then, we measured leaf consumption as consumed leaf area (mm<sup>2</sup>) using
ImageJ software (version 1.45), and assimilation efficiency as the ratio of 4-d old larval mass
(measured to the nearest 0.1mg using a Mettler AT261 balance; Mettler Toledo, Columbus, OH,
USA) over amount of leaf consumed.

150

Statistical analysis, of maternal effects on larval responses to predation risk, involved the use of 151 152 linear mixed models with maximum likelihood estimation. For each trait -leaf consumption and 153 assimilation efficiency, we included the treatment effect as a fixed factor (predator-free vs. predation risk) while the sire, dam, sire-by-treatment interaction and dam-by-treatment 154 interaction (referred as GxE and MxE) were all included as random factors. Significance of 155 156 variance components for larval responses to predation risk were estimated using the REML 157 method (Proc MIXED) and likelihood ratio tests (Saxton and SAS Institute. 2004). Evidence of a significant GxE or MxE was investigated in more detail by testing the null hypothesis that 158 159 genetic correlations of traits associated with plastic responses to predation risk, measured across the predator-free and predation risk environment ( $r_A$ ) were = 1 (Lynch and Walsh 1998; Messina 160 and Fry 2003). Genetic correlations significantly less than 1 suggest the potential for independent 161 trait evolution in different predation-risk environments and thus would provide additional 162 support for GxE. Additionally, because expression of maternal and additive genetic effects is 163 164 expected to differ within different environments, we estimated the genetic variance components (V<sub>M</sub>, V<sub>A</sub>, V<sub>E</sub>) and associated genetic parameters (e.g. narrow sense heritabilities and genetic 165 coefficient of variation) for the larval traits within each environment -predator-free and 166 predation risk (Table 1S). Variance components were calculated assuming the dominance 167 variance to be zero:  $V_A = 4$  sire,  $V_M = dam - sire$ , and  $V_P = total phenotypic variance (Falconer$ 168

and Mackay 1996). Narrow sense heritabilities  $(h^2)$  of traits within a predator environment were then calculated as  $V_A / V_P$ , representing the proportion of the total phenotypic variance explained by the direct genetic variance (Falconer and Mackay 1996).

172

# 173 c) Parental size effects on larval responses to predation risk

In a second step, we followed a trait-based approach (McAdam et al. 2014) to investigate the
effects of both maternal and paternal body size on larval plastic responses to predation risk.

176 Here, the effects of specific parental traits, the body size of sires (fathers) and dams (mothers),

are modeled. Because our main interest was to investigate effects on larval plastic responses to

178 predation risk (rather than effects on the traits in each environment) we calculated larval plastic

179 responses using Cohen's D effect size 
$$=\frac{Mean_{Risk-free}-Mean_{Predation\,risk}}{SD_{pooled}}$$
 and used multiple

regression analysis to test for both maternal and paternal size effects (for a similar approach see 180 Bennett and Murray 2014). Given that previous studies have shown that larger hatchlings elicit 181 stronger antipredator responses, we included larval size (averaged for each family) as an 182 additional predictor. Additionally, we analyzed the effects of parental body size on larval 183 responses using a "hybrid approach" (McAdam et al. 2014), which included sire and dam 184 identities (as described in "maternal effects on larval responses to predation risk") plus maternal 185 (or paternal) body size and its interaction with predation risk treatment as fixed effects (Size<sub>sire</sub> x 186 187 E and Size<sub>dam</sub> x E). This approach allow us to model the effects of specific parental traits (here body size) while accounting for the remaining variation in maternal and sire effects (V<sub>M</sub> and V<sub>A</sub>). 188 However, these more complex models, especially when unbalanced, may bias estimates of 189 190 genetic parameters as well as inference for fixed effects (e.g. Kenward and Roger 1997; Stroup and Littell 2002). 191

192

### 193 (d) Maternal plastic responses to predation risk

In a last experiment, we measured the amount of genetic variance (and associate genetic 194 195 parameters) underlying maternal responses to predation risk including clutch size, levels of intraclutch cannibalism, and proportion of viable offspring within a clutch (Fig. 1). Note that 196 although intraclutch cannibalism and clutch size may influence progeny phenotype (e.g. progeny 197 198 size), these are here considered a maternal rather than an offspring trait (see Mousseau & Fox 1998). To measure maternal responses to predation risk, two females from each maternal family 199 (and the same clutch) were individually reared to adults and mated with a full sibling. Because 200 males in a number of insects can influence female reproduction (e.g. fecundity), mating full sibs 201 may reduce chances to introduce an additional source of variation to the estimates of additive 202 203 genetic variance in maternal traits. Importantly, mating full-sibs did not cause any apparent inbreeding effects in female reproduction: clutch size and offspring produced by full-sib pairs 204 were comparable to those observed in females that had mated with unrelated males (see 205 206 Inbreeding analysis in Supplemental information). After mating, half of the females were each kept with two sham predators (predation risk environment) in a 0.5 L cup, with abundant plant 207 foliage for feeding and oviposition. The rest of females were kept in similar conditions but 208 without the sham predators (predator-free environment). We collected the first two clutches that 209 females laid and stored them individually with a fresh leaflet (in 30 ml cups). Clutch size 210 (number of eggs) and offspring produced were measured in the same clutch while intraclutch 211 cannibalism, the proportion of eggs that were fully consumed by the new hatchlings, were 212 measured on a second clutch (Collie et al. 2013; Tigreros et al. 2017). 213

215 The statistical approach to examine maternal responses to predation risk was similar to that used

- for larval responses to predation risk but, because each dam included only one daughter per
- treatment, maternal effects on maternal responses to predation risk ( $V_M x E$ ) cannot be estimated.
- 218 Final analysis of maternal responses to predation risk included 15 sires mated to 32.
- Additionally, to estimate variance components for levels of intraclutch cannibalism we used a
- 220 generalized linear model using LAPLACE method (Proc GLIMMIX), which captured the
- binomial distribution of intraclutch cannibalism using a logit link function (Saxton and SAS
- Institute. 2004). Because the residual variance estimates of this type of model is correlated with
- the mean of the population, we estimated heritability as "latent-scale heritability", by adding the
- variance component related to the link function (Nakagawa and Schielzeth 2010; Calsbeek et al.

225 2015), which for binomial models is  $=\frac{V_A}{V_P + \pi^2/3}$ . All analyses were conducted using SAS 9.4

- 226 (SAS Institute Inc., Cary, NC, USA).
- 227

### 228 **3. RESULTS**

# 229 (a) Maternal effects on larval responses to predation risk

230 Larval plastic responses to predation risk were substantial, involving a 22% reduction in leaf

consumption ( $F_{1,18}$  =7.53, P = 0.01; Fig. 2A, Table 1A) and a 17% increase in assimilation

efficiency ( $F_{1,18} = 5.43$ , P = 0.03; Fig. 2B, Table 1 A). Yet, we did not detect a significant

- interaction between predation risk and sire or dam families influencing the larval traits (Table
- 1A: Sire x Risk and Dam x Risk), which indicates that there was substantial plasticity in larval
- responses to predation risk, but there was not a detectable genetic or maternal variance
- 236 underpinning larval responses.

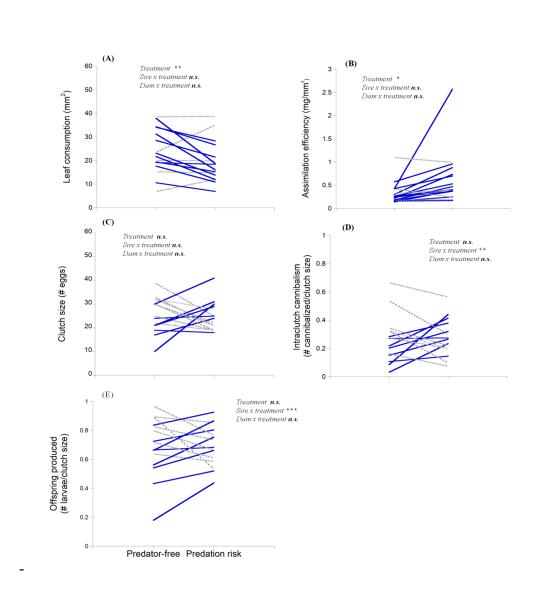


Figure 2. Reaction norms for *L. decemlineata* describing plastic responses to predation risk in terms of (A) Larval leaf consumption, (B) Larval assimilation efficiency, (C) Adult clutch size, (D) Intraclutch cannibalism and (E) Offspring number per clutch. Each line represents the mean score for each sire family. The slope of the line is a graphical representation of the strength and direction of plastic responses to predation risk shown by that family. Highlighted with solid blue lines (vs. doted grey lines) are the families that responded to predation risk in a direction that would typically improve offspring fitness (see Table 1 for further statistical elaboration).

#### (A) Larval responses

Leaf consumption		Fit statistic (-2 ResLogLikelihood): 2768.3		
-	Fix effects	F	df	Р
	Risk	7.53	1,18	0.013
	Egg-age	2.88	2, 229	0.058
	<b>Random Effects</b>	Estimate	SE	Р
	Sire	1 <b>6</b> .44	51.2	
	Dam(sire)	228.82	73.48	
	Sire x Risk	17.4 <b>6</b>	17.6	> 0.05 (2768.2)
	Dam x Risk	2.366	21.08	> 0.05 (2769.6)
	Error	269.02	25.1	
Assimilation		Fit statistic (	-2 ResLogL	ikelihood): 337.7
efficiency	Fix effects	F	df	Р
	Risk	5.43	1,18	0.03
	Egg-age	1.63	1,231	0.19
	<b>Random Effects</b>	Estimate	SE	Р
	Sire	0.0055	0.014	
	Dam(sire)	0.0356	0.017	
	Sire x Risk	0		> 0.05 <i>(337.7)</i>
	Dam x Risk	0		> 0.05 (337.7)
	Error	0.138	0.0117	
(B) Maternal responses				
(B) Maternal resp	oonses			

Clutch Size	Fit statistic (-2 ResLogLikelihood): 892.64			
	Fix effects	F	df	Р
	Risk	0.52	1,14	0.48
	<b>Random Effects</b>	Estimate	SE	Р
	Sire	0		
	Dam(sire)	40.5	17.35	
	Sire x Risk	6.25	11.3	> 0.05 (893.0I)
	Error	88.19	15.59	

Fit statistic (-2 ResLogLikelihood): 244.26 Intraclutch cannibalism Fix effects F df Ρ 1,13 Risk 0.37 0.5 Random Effects Estimate SE Р Sire 0.0066 0.209 Dam(sire) 0.296 0.16 Sire x Risk 0.205 <0.0001 (335.27) 0.389 Error 1.99 0.347 Proportion of Fit statistic (-2 ResLogLikelihood): -48.08 offspring **Fix effects** F df Р 0.81 Risk 0.06 1,13 **Random Effects** Estimate SE Р Sire 0.0089 0.0093 Dam(sire) 0.012 0.0064 Sire x Risk 0.0074 0.0053 0.023(-44.06) Error 0.0215 0.0037

Table 1. Tests on the statistical significance of the predation risk treatment (Risk) and its interaction with the dam (MxE) and sire (GxE) components. Interaction terms were tested using likelihood ratio test that compare the fit statistic,-2 ResLog likelihood fit, of the full model (showed on top row) with that of the model without the term of interest (showed in parenthesis after the P value).

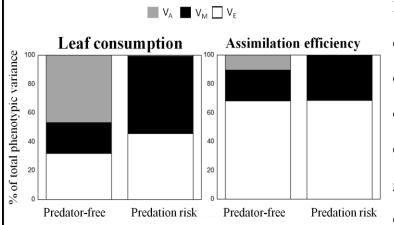


Fig. 3 Percent of phenotypic variation (for leaf consumption and assimilation efficiency) explained by the different genetic components of variation within each predator. Genetic components of variation include: additive genetic  $(V_A)$ , maternal  $(M_A)$ , and environmental  $(V_E)$  effects.

Partitioning of variance components of larval traits within each predator environment (predator-246 free and predation risk environment) showed, as expected, differences in the relative importance 247 248 of maternal  $(V_M)$  and additive genetic effects  $(V_A)$  (Fig. 3; Table 1S). While maternal effects 249 (V<sub>M</sub>) explained a large proportion of the phenotypic variance in both environments, this was 250 even stronger when in the predation risk environment, explaining over 50% of the variation in 251 leaf consumption and about 30% in assimilation efficiency (Fig. 3; Table S1 in Supplemental 252 information). In contrast, levels of additive genetic variation (V<sub>A</sub>) were not statistically 253 significant in either environment (Table S1). Note, however, that the number of sire families in our study was close to the minimum recommended to detect V<sub>A</sub> (Conner and Hartl 2004) and 254 255 therefore, finding no additive genetic variance in larval traits could be due to a low statistical power rather than an absolute lack of genetic variance. 256

257

## 258 (b) Parental size effects on larval responses to predation risk

To test parental size effects on larval responses, we first partitioned variation of adult body sizeinto additive genetic and environmental components. These results revealed a significant additive

261 genetic component (V<sub>A</sub> = 0.078 ± 0.056, p=0.025), with moderate heritability levels ( $h^2 = 0.2$ ), 262 underlying body size in adults.

264	Evaluation of parental size effects on larval responses provided similar results using both the
265	trait-based approach of regressing parents' body size on larval responses- and the hybrid
266	approach of including parent's body size as covariate plus Sire and Dam identities (McAdam et
267	al. 2014). Regression analyses (controlling offspring size), revealed that paternal body size, but
268	not maternal body size, influenced both the magnitude and direction of larval responses to
269	predation risk including leaf consumption ( $R^2 = 0.26$ : $\beta$ sire= -0.51, $p = 0.001$ ; $\beta$ dam= 0.04, $p =$
270	0.25; $\beta$ larvae= 0.06, $p$ = 0.4; Fig. 4A), and assimilation efficiency ( $R^2$ =0.23: $\beta$ sire= 0.48, $p$ =
271	0.003; $\beta$ dam=-0.05; $p$ = 0.73; $\beta$ larvae=0.04, $p$ = 0.77; Fig. 4B). Smaller sires producing larvae
272	with the strongest responses (reduced feeding and increased assimilation efficiency). Results
273	from models that included sire and dam identity corroborated the effect of sire size on larval
274	responses (Table S2 in Supplemental information).

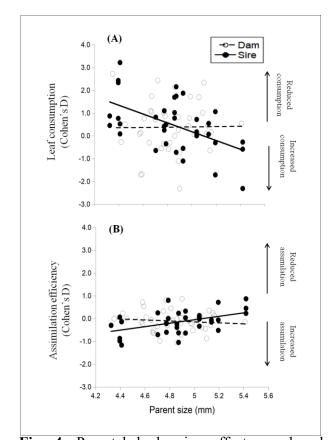


Fig. 4. Parental body size effects on larval responses to predation risk for (A) leaf consumption and (B) assimilation efficiency in L. decemlineata larvae. The magnitude and direction of larval plastic responses were calculated using Cohen's D effect size Mean<sub>Risk-free</sub>-Mean<sub>Predation</sub> risk Each ). SDpooled

observation represents the effect size and mean body size for each maternal family (full siblings).

# (b) Maternal response to predation risk

Analyses of maternal responses to predation risk indicated that there was not a main effect of predation risk treatment on clutch size, intraclutch cannibalism, or number of viable offspring produced per clutch (Table 1B; Fig. 2C-E). However, there was a significant sire by predation treatment interaction for proportion of intraclutch cannibalism and number of viable larvae produced, indicating that there is substantial genetic variation for these maternal traits, involving differences in the magnitude as well the direction of the response (Fig. 2 D, E). Further investigation of this GxE, by testing significance of genetic correlations across the predator-free and predation risk environments, indicated that these were significantly less than 1 (intraclutch cannibalism  $r_A = -0.01$ , se = 0.7, p = 0.03; proportion of offspring  $r_A = 0.74$ , se = 0.5, p < 0.0001). These results suggest the

potential for independent trait evolution in the different predation environments, providing
additional support for GxE in maternal responses to predation risk (Lynch and Walsh 1998).

301 DISCUSSION

Parental effects are important determinants of traits associated with predator avoidance in a number of species. As for all phenotypes, parental effects are shaped by the environment, the genotype, and their interaction. Only by disentangling the relative contribution of such different sources of parental effects –e.g., those owed to the environment vs. the parent's genes– we can understand their role in organisms' evolution.

307

As observed in previous studies of L. decemlineata (Hermann and Thaler 2014; Kaplan et al. 308 309 2014; Tigreros et al. 2017, 2018), we found strong plastic responses to predation risk, involving 28% reductions in leaf consumption and 15% increases in assimilation efficiency. Coupling of 310 feeding reductions with increased assimilation efficiency is critical to prey fitness, as this allows 311 312 prey to lower chances of predation while minimizing costs associated with reduced-food intake (Thaler et al. 2012; Kaplan et al. 2014). Based on 19 sires, we found little support of an additive 313 genetic or a maternal component underpinning larval responses to predation risk. However, 314 analyses that included size of the parents as covariates revealed that variation in responses –for 315 both leaf consumption and assimilation efficiency-were at least partially explained by paternal 316 size. Specifically, smaller fathers produced larvae with the greatest plasticity in response to 317 predation risk –strong feeding reductions and increases in assimilation efficiency. Given that 318 females often provide more reproductive investment than males, studies on parental effects often 319 320 rely on the notion that the maternal phenotype is the main influence on the offspring. However,

321 as we found in this study, male body size can be linked to parental performance (Eilertsen et al. 322 2009), perhaps through changes in the quality of their ejaculates (Gillott 2003), which is known to include accessory glands compounds in *L. decemlineata* beetles (Loof and Lagasse 1972). 323 324 Independent of the exact mechanism, because adult body size had a significant additive genetic component, size related paternal effects represented an indirect source of genetic variation 325 shaping larval plastic responses to predation risk. 326 327 Estimates of maternal variance  $(V_M)$  explained about half of the phenotypic variance in leaf 328 consumption when expressed under high risk of predation. These results are concordant with the 329 notion that the contribution of parental effects, relative to the environmental and additive genetic 330 variance components, is high for traits that are expressed early in life (Bernardo 1996b; 331 332 Lindholm et al. 2006; Wilson and Reale 2006; White and Wilson 2019) and under environmental stress (e.g. Rudin-Bitterli, Mitchell & Evans 2018). 333 334 335 In a previous study we demonstrated that larval responses to predation risk (including decreases in leaf consumption) was improved through an anticipatory parental effect: mothers experiencing 336 the risk of predation increased offspring provisioning by inducing intraclutch cannibalism 337 (Tigreros et al. 2017); cannibals, in better nutritional condition than non-cannibal siblings, 338 exhibited stronger antipredator behaviors. Here, we found that mothers indeed responded to 339 predation risk by altering levels of egg cannibalism within their clutches. However, such 340 responses varied in magnitude and direction, with some families increasing and others 341 decreasing cannibalism. In L. decemlineata, intraclutch cannibalism results in a classic life 342 343 history tradeoff between investment in offspring quality (cannibalistic offspring) and quantity

(cannibalized offspring). While fitness of the individual offspring may always improve with 344 cannibalism, optimal levels of cannibalism within a clutch should reflect the number of 345 cannibalistic offspring that would maximize female reproductive success in a given environment 346 347 (e.g. under predation risk). Finding that families with the highest and lowest levels of intraclutch cannibalism -in the predator-free environment- responded to predation risk by decreasing and 348 increasing cannibalism respectively, suggests that intermediate levels of intraclutch cannibalism 349 350 may be optimal under environments with high predation risk. Importantly, such variation in maternal responses to predation risk (changes in intraclutch cannibalism and offspring produced) 351 reflected genetic differences among the mothers (significant GxE) indicating the potential for 352 evolutionary change of a maternal effect, in response to predator. 353 354 355 The conditions under which organisms rely on parental effects vs. within-generation phenotypic plasticity remains an open question in evolutionary ecology. Using plasticity alone, L. 356 decemlineata larvae can achieve substantial feeding reductions (e.g.  $\sim 28\%$  in this study) when 357 358 facing predation risk. However, our previous work indicates that these responses are constrained by the larvae's nutritional state (Tigreros et al. 2017), and larvae feeding on lower quality host 359 plants have shown weaker feeding reductions in response to predation risk (Kaplan et al. 2014; 360 Tigreros et al. 2017). Through cannibalism, L. decemlineata appears to overcome such 361 nutritional constraints, and cannibals that experience predation risk are capable of reducing leaf 362 consumption, even on low quality host plants. Thus, this extreme type of maternal investment 363 should improve offspring fitness when facing highly stressful environments (Marshall et al. 364 2008; Olofsson et al. 2009). 365

Recent theoretical evidence suggests that evolution of parental effects is promoted when there is 367 strong selection on the phenotype and when within-generation plasticity is constrained (Auld et 368 al. 2010; Kuijper and Hoyle 2015). Predation is unquestionably one of the strongest selective 369 370 forces in nature, and is therefore thus likely to deplete genetic variation in traits associated with predator avoidance. In contrast, genetic variation underlying paternal and maternal effects is 371 expected to remain "protected" from the eroding effects of selection when carried by the other 372 sex (Wade 1998). Accordingly, results from this study indicate that even if larval responses to 373 predation risk were holding little or no genetic variation, parental effects -via induced intraclutch 374 cannibalism and paternal body size- should provide the necessary genetic variation for future 375 natural selection to act upon (Räsänen and Kruuk 2007). Additionally, because L. decemlineata 376 larval nutritional condition is a function of host plant quality as well as the degree of maternal 377 provisioning, evolution of paternal effects may be favored in environments where larval 378 plasticity is constrained by poor host plant quality. Together, our results show that the 379 evolutionary potential of predator avoidance in L. decemlineata, relies on at least two different 380 381 genetic parental effects, one linked to paternal body size and the other to maternal induction of intraclutch cannibalism. 382

383

384

#### 385 ACKNOWLEDGEMENTS

We thank Rachel H. Norris for help with collection of data. We also thank Kyle Benowitz and
Nicholas Aflitto, and two anonymous reviewers for their valuable comments on the manuscript.
This work was supported by USDA grants to JST: NIFA 2013-02649, Federal Capacity Funds
1397484; and AFRI 2018-67013-28068.

390

#### 391 REFERENCES

- 392 Agrawal, A. A, C. Laforsch, and R. Tollrian. 1999. Transgenerational induction of defences in
- animals and plants. Nature 401:60–63.
- Auld, J. R., A. A. Agrawal, and R. A. Relyea. 2010. Re-evaluating the costs and limits of
- adaptive phenotypic plasticity. Proceedings. Biological sciences 277:503–11.
- Bennett, A. M., and D. L. Murray. 2014. Maternal body condition influences magnitude of anti-
- 397 predator response in offspring. Proceedings of the Royal Society B: Biological Sciences
- **398** 281:20141806–20141806.
- Bernardo, J. 1996a. The particular maternal effect of propagule size, especially egg size:
- 400 patterns, models, quality of evidence and interpretations. American Zoologist 36:216–236.
- 401 Bernardo, J. 1996b. Maternal effects in animal ecology. American Zoologist 36:83–105.
- Calsbeek, R., M. C. Duryea, D. Goedert, P. Bergeron, and R. M. Cox. 2015. Intralocus sexual
  conflict, adaptive sex allocation, and the heritability of fitness. Journal of Evolutionary Biology
  28:1975–1985.
- Collie, K., S. J. Kim, and M. B. Baker. 2013. Fitness consequences of sibling egg cannibalism by
  neonates of the Colorado potato beetle, Leptinotarsa decemlineata. Animal Behaviour 85:329–
  338.
- 408 Conner, J. K., and D. L. Hartl. 2004. A Primer of Ecological Genetics. BioScience.
- 409 Donelson, J. M., S. Salinas, P. L. Munday, and L. N. S. Shama. 2018. Transgenerational

- 410 plasticity and climate change experiments: Where do we go from here? Global Change Biology.
- 411 Eilertsen, E. M. K., B. J. Bårdsen, S. Liljedal, G. Rudolfsen, and I. Folstad. 2009. Experimental
- 412 evidence for paternal effects on offspring growth rate in Arctic charr (Salvelinus alpinus).
- 413 Proceedings of the Royal Society B: Biological Sciences.
- 414 Falconer, D. S., and T. F. C. Mackay. 1996. Introduction to Quantitative Genetics (4th Edition).
- 415 Trends in Genetics (Vol. 12).
- 416 Fox, C. W. 1994. Maternal and genetic influences on egg size and larval performance in a seed
- 417 beetle (Callosobruchus maculatus): multigenerational transmission of a maternal effect? Heredity
- 418 73:509–517.
- 419 Fox, C. W., and M. E. Czesak. 2000. Evolutionary ecology of progeny size in arthropods.
- 420 Annual Review of Entomology 45:341–369.
- 421 Fox, C. W., M. E. Czesak, T. A. Mousseau, and D. A. Roff. 1999. The Evolutionary Genetics of
- an Adaptive Maternal Effect: Egg Size Plasticity in a Seed Beetle. Evolution 53:552.
- 423 Futuyma, D. J. 2009. Evolution. Evolution.
- Galloway, L. F. 2005. Maternal effects provide phenotypic adaptation to local environmentalconditions. New Phytologist.
- Galloway, L. F., and J. R. Etterson. 2007. Transgenerational Plasticity Is Adaptive in the Wild.
  Science 318:1134–1136.
- Gillott, C. 2003. Male accessory gland secretions: modulators of female reproductive physiologyand behavior. Annu Rev Entomol.

- 430 Griffin, C. A. M., and J. S. Thaler. 2006. Insect predators affect plant resistance via density- and
- 431 trait-mediated indirect interactions. Ecology Letters 9:338–346.
- 432 Hermann, S. L., and J. S. Thaler. 2014. Prey perception of predation risk: volatile chemical cues
- 433 mediate non-consumptive effects of a predator on a herbivorous insect. Oecologia 176:669–676.
- 434 Jensen, N., R. M. Allen, and D. J. Marshall. 2014. Adaptive maternal and paternal effects:
- 435 Gamete plasticity in response to parental stress. Functional Ecology.
- 436 Kaplan, I., S. H. McArt, and J. S. Thaler. 2014. Plant defenses and predation risk differentially
- 437 shape patterns of consumption, growth, and digestive efficiency in a guild of leaf-chewing
- 438 insects. PloS one 9:e93714.
- 439 Kenward, M. G., and J. H. Roger. 1997. Small Sample Inference for Fixed Effects from

440 Restricted Maximum Likelihood. Biometrics 53:983.

- 441 Kuijper, B., and R. B. Hoyle. 2015. When to rely on maternal effects and when on phenotypic
- 442 plasticity? Evolution; international journal of organic evolution 69:950–68.
- Lande, R., and M. Kirkpatrick. 1990. Selection response in traits with maternal inheritance.
  Genetical Research 55:189–197.
- Lindholm, A. K., J. Hunt, and R. Brooks. 2006. Where do all the maternal effects go? Variation
- in offspring body size through ontogeny in the live-bearing fish Poecilia parae. Biology Letters.
- Loof, A., and A. Lagasse. 1972. The ultrastructure of the male accessory reproductive glands of
- the colorado beetle. Zeitschrift for Zellforschung und Mikroskopische Anatomie 130:545–552.
- Love, O. P., and T. D. Williams. 2008. The adaptive value of stress-induced phenotypes: Effects

- 450 of maternally derived corticosterone on sex-biased investment, cost of reproduction, and
- 451 maternal fitness. American Naturalist 172:E135–E149.
- 452 Lynch, M., and B. Walsh. 1998. Genetics and analysis of quantitative traits. Sinauer Associates;
- 453 1 edition. 980p.
- 454 Marshall, D. J., R. Bonduriansky, and L. F. Bussière. 2008. Offspring size variation within
- 455 broods as a bet-hedging strategy in unpredictable environments. Ecology 89:2506–2517.
- 456 Marshall, D. J., and T. Uller. 2007. When is a maternal effect adaptive? Oikos.
- 457 McAdam, A. G., D. Garant, and A. J. Wilson. 2014. The effects of other's genes: maternal and
- 458 other indirect genetic effects. *in* A. Charmantier, D. Garant, and L. E. B. Kruuk, eds.
- 459 Quantitative genetics in the wild (First.). Oxford University Press, Oxford.
- 460 Messina, F. J., and J. D. Fry. 2003. Environment-dependent reversal of a life history trade-off in
- the seed beetle Callosobruchus maculatus. Journal of Evolutionary Biology 16:501–509.
- Mousseau, T. A., and H. Dingle. 1991. Maternal Effects in Insect Life Histories. Annual Review
  of Entomology 36:511–534.
- Mousseau, T. A., and C. W. Fox. 1998. The adaptive significance of maternal effects. Trends in
  Ecology & Evolution 13:403–406.
- 466 Nakagawa, S., and H. Schielzeth. 2010. Repeatability for Gaussian and non-Gaussian data: a
  467 practical guide for biologists. Biological Reviews 85:no-no.
- 468 Olofsson, H., J. Ripa, and N. Jonzén. 2009. Bet-hedging as an evolutionary game: the trade-off
- 469 between egg size and number. Proceedings of the Royal Society of London B: Biological

470 Sciences 276.

- 471 Räsänen, K., and L. E. B. Kruuk. 2007. Maternal effects and evolution at ecological time-scales.
- 472 Functional Ecology 21:408–421.
- 473 Rowiński, P. K., and B. Rogell. 2017. Environmental stress correlates with increases in both
- 474 genetic and residual variances: A meta-analysis of animal studies. Evolution.
- 475 Rudin-Bitterli, T. S., N. J. Mitchell, and J. P. Evans. 2018. Environmental Stress Increases the
- 476 Magnitude of Nonadditive Genetic Variation in Offspring Fitness in the Frog Crinia georgiana .
- 477 The American Naturalist.
- 478 Saxton, A. M., and SAS Institute. 2004. Genetic analysis of complex traits using SAS. SAS479 Institute.
- 480 Sheriff, M. J., C. J. Krebs, and R. Boonstra. 2010. The ghosts of predators past: Population
- 481 cycles and the role of maternal programming under fluctuating predation risk. Ecology 91:2983–
  482 2994.
- 483 Sheriff, M. J., and O. P. Love. 2013. Determining the adaptive potential of maternal stress.
  484 Ecology Letters.
- 485 Storm, J. J., and S. L. Lima. 2010. Mothers forewarn offspring about predators: a
- transgenerational maternal effect on behavior. The American naturalist 175:382–90.
- 487 Stroup, W., and R. Littell. 2002. Impact of variance component estimates on fixed effect
- 488 inference in unbalanced linear mixed models. Conference on Applied Statistics in Agriculture.
- 489 Thaler, J. S., S. H. McArt, and I. Kaplan. 2012. Compensatory mechanisms for ameliorating the

- 490 fundamental trade-off between predator avoidance and foraging. Proceedings of the National
- 491 Academy of Sciences of the United States of America 109:12075–80.
- 492 Tigreros, N., R. H. Norris, E. H. Wang, and J. S. Thaler. 2017. Maternally induced intraclutch
- 493 cannibalism: an adaptive response to predation risk? Ecology Letters 20:487–494.
- 494 Tigreros, N., E. H. Wang, and J. S. Thaler. 2018. Prey nutritional state drives divergent
- behavioural and physiological responses to predation risk. Functional Ecology 32:982–989.
- Wade, M. J. 1998. The evolutionary genetics of maternal effects. Pages 5–21 *in* Maternal Effects
- 497 as Adaptations.
- 498 White, S. J., and A. J. Wilson. 2019. Evolutionary genetics of personality in the Trinidadian
- 499 guppy I: maternal and additive genetic effects across ontogeny. Heredity.
- Wilson, A. J., and D. Reale. 2006. Ontogeny of additive and maternal genetic effects: lessonsfrom domestic mammals. The American naturalist.
- Wolf, J. B., E. D. Brodie III, J. M. Cheverud, A. J. Moore, and M. J. Wade. 1998. Evolutionary
  consequences of indirect genetic effects. Trends in Ecology & Evolution 13:64–69.
- 504
- 505
- 506
- 507