

1 Segregation of Familial Risk of Obesity in NHANES Cohort Supports a
2 Major Role for Large Genetic Effects in the Current Obesity Epidemic
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18

19 Abstract

20 The continuing increase in many countries in adult body mass index (BMI kg/m²) and its dispersion
21 is contributed to by interaction between genetic susceptibilities and an increasingly obesogenic
22 environment (OE). The determinants of OE-susceptibility are unresolved, due to uncertainty around
23 relevant genetic and environmental architecture. We aimed to test the multi-modal distributional
24 predictions of a Mendelian genetic architecture based on collectively common, but individually rare,
25 large-effect variants and their ability to account for current trends in a large population-based sample.
26 We studied publicly available adult BMI data (n = 9102) from 3 cycles of NHANES (1999, 2005,
27 2013). A first degree family history of diabetes served as a binary marker (FH₀/FH₁) of genetic
28 obesity susceptibility. We tested for multi-modal BMI distributions non-parametrically using kernel-
29 smoothing and conditional quantile regression (CQR), obtained parametric fits to a Mendelian model
30 in FH₁, and estimated FH x OE interactions in CQR models and ANCOVA models incorporating
31 secular time. Non-parametric distributional analyses were consistent with multi-modality and fits to a
32 Mendelian model in FH₁ reliably identified 3 modes. Mode separation accounted for ~40% of BMI
33 variance in FH₁ providing a lower bound for the contribution of large effects. CQR identified strong
34 FH x OE interactions and FH₁ accounted for ~60% of the secular trends in BMI and its SD in
35 ANCOVA models. Multimodality in the FH effect is inconsistent with a predominantly polygenic,
36 small effect architecture and we conclude that large genetic effects interacting with OE provide a
37 better quantitative explanation for current trends in BMI.

38 Introduction

39 The recent and continuing increase in the global mean adult BMI, first seen in high income countries,
40 is now seen in most countries across a wide range of ethnic composition and socio-economic
41 conditions (Di Cesare et al., 2016) and is accompanied by increases in measures of dispersion

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42 (Krishna et al., 2015; Silventoinen et al., 2017). Although BMI is known from family-based studies to
43 be under strong genetic influences (Loos, 2018) population genetic backgrounds have been
44 effectively constant over this time, implying that BMI trends are driven by change in environmental
45 factors (obesogenic environment, OE). Evidence from twin studies, which demonstrate increased
46 genetic variance over time, supports an important role for interactions between OE and genetic
47 susceptibility (G x OE) on both mean and dispersion of BMI (Rokholm et al., 2011; Silventoinen et
48 al., 2017), but how large a role is not yet known. Defining the role of G x OE in “epidemic” obesity,
49 and hence of genetic susceptibility itself, is hindered by problems of measurement and modeling of
50 interactions (Franks and McCarthy, 2016) and by uncertainty around both the genetic architecture
51 (Loos, 2018) and the exact nature of the environmental drivers (Hall, 2018). Whether population
52 susceptibility to OE is predominantly determined by a subgroup with high genetic susceptibility or is
53 more evenly spread within populations is unresolved despite important implications for the
54 management of obesity and related disorders at population and individual
55 levels (Kivimaki et al., 2015; Krishna et al., 2015; Jenkins and Campbell, 2015; Razak et al., 2015).
56
57 The genetic variants responsible for obesity susceptibility remain largely unknown. Genome-wide
58 association studies (GWAS) have identified significant associations with >200 markers with small
59 effects on BMI (polygenes), together explaining only approximately 3-4 % of total variance
60 compared to family-based heritabilities (h^2) of 50-75% (Speakman et al., 2018). Few causative
61 mechanisms responsible for these phenotypically weak associations are known (Loos, 2018). The
62 sources of the h^2 unaccounted for by GWAS are uncertain; suggestions include overestimation of h^2 ,
63 large numbers of common genetic variants with small, statistically insignificant effects on phenotypes
64 (Locke et al., 2015; Khera et al., 2019) and importantly, candidates not tested in most GWAS.
65 Among the latter are rare genetic variants with large phenotypic effects and G x OE interactions
66 (Loos, 2018). Recently significant G x OE interactions have been detected in individual GWAS loci

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67 and in composite genetic risk scores, which however explain little of the missing component of h^2

68 (Abadi et al., 2017; Nagpal et al., 2018).

69

70 A family history of diabetes (FH) is a potent, predominantly genetic (Hemminki et al., 2010;

71 Willemssen et al., 2015) risk factor for diabetes diagnosis (DM) and for obesity-related phenotypes

72 (Ghosh et al., 2010; Tirosh et al., 2011; Scott and Consortium, 2013; Jenkins et al., 2013) consistent

73 with the strong association between type 2 DM and overweight/obesity. Familial effects on obesity-

74 related phenotypes in adults are also predominantly genetic (Stryjecki et al., 2018; Silventoinen et al.,

75 2017), so to the extent that the DM generating FH is of type 2 (approximately 94% of DM in the US

76 population (Xu et al., 2018)), FH is a prevalent and readily obtained marker of genetic susceptibility

77 both to diabetes and to the obesity commonly preceding it. We have previously reported evidence

78 from a small sample of a multi-modal effect of FH on a composite adiposity index consistent with

79 segregation in families of discrete obesity risk (Jenkins et al., 2013). Polygenic risk scores (PRS)

80 based on large numbers of small effects are expected to be, and appear to be, unimodally-distributed

81 (Llewellyn et al., 2014; Rask-Andersen et al., 2017) and thus cannot account for familial segregation

82 of discrete risk. The present work is based on the alternative hypothesis that individually rare, but

83 collectively common, genetic variants with large phenotypic effects are the source of most of the

84 missing h^2 and of most of $G \times OE$, and that their effects can be detected through analyses of

85 phenotypic segregation in high-risk families (Jenkins and Campbell, 2014).

86

87 The Continuous National Health and Nutrition Examination Survey (NHANES) is a continuing

88 (1999-) large-scale population-based survey incorporating an index of adiposity (Body Mass Index,

89 BMI) and first-degree FH (FH_0/FH_1) together with potential covariates and confounders. Although

90 BMI has recognized limitations as an adiposity phenotype (Jenkins and Campbell, 2014; Müller et

91 al., 2018) it is the basis for most large-scale genetic studies and like other authors, we assume that a

92 large enough scale and appropriate modeling of covariates will reduce effects of imprecision and bias

93 (Speakman et al., 2018). We aimed to test in a large multi-cycle NHANES sample for the presence of
94 familial segregation of genetic risk and to estimate the contribution of FH, and by extension all
95 discrete genetic risk, to recent secular trends in adult BMI. The results support a predominant role for
96 large genetic effects interacting with OE in the obesity “epidemic”.

97

98 **Subjects and Methods**

99 **Subjects**

100 We used data from the 1999-2000, 2005-2006 and 2013-2014 cycles of NHANES
101 (<https://www.cdc.gov/nchs/nhanes/index.htm> accessed 25 Aug 2017). We extracted records for
102 participants age 20-65 years with non-missing gender, race/ethnicity, BMI, diabetes family history
103 (FH) and diabetes diagnosis data, and current smoking status if available. The definitions of two
104 fields changed over the sampling period: 1) Diabetes family history was defined in terms of 1° and 2°
105 relatives in 1999-2000 but by 1° relatives only in subsequent cycles. We recoded the 1999-2000
106 diabetes family history data to conform to the later definition using the separately collected data for
107 affected parents and siblings. 2) The self-identified race/ethnicity field (RIDRETH1) code was used
108 excluding other races (OR) to maintain consistency of categories across cycles (Supplementary
109 Methods). We excluded from the primary analyses subjects diagnosed with diabetes because of
110 possible confounding by effects of either diabetes or diabetes therapies on BMI. The resulting data
111 set is summarized in Table 1.

112 **Statistical analyses**

113 *Approach*

114 We treated the data as a convenience sample and took no account of the sampling weights provided
115 by NHANES to permit nationally representative estimates. Our results are not intended to be
116 representative of the US population.

117 Our primary analyses are based on non-parametric visualization (kernel-smoothing) and analyses
118 (conditional quantile regression, CQR) of distributions requiring no prior distributional assumptions.
119 Parametric fits to multimodal distributions were then used to quantify the contributions of the
120 predicted large genetic effects model. $FH_{(0.1)}$ is treated analytically as a binary genetic risk marker
121 but the distribution of its effect across quantiles is interpreted in a Mendelian model in which FH_1
122 represents enrichment of a mixture of single and double carriers of risk variants. Calendar time is
123 treated as a continuous surrogate of OE. Effects of OE interacting with FH were assessed in CQR
124 models, and also in least-squares ANCOVA models using bootstrap resampling to minimize
125 distributional assumptions in the calculation of effect size estimates and errors. All analyses were
126 performed using R 3.6.1 (R Development Core Team, 2016).

127

128 *Summary statistics*

129 Heterogeneity of the samples across cycles was assessed by Chi square test for categorical variables
130 and by one-way ANOVA for age. Effects on BMI were assessed by ANCOVA against continuous
131 time ($yr = \text{calendar start year} - 1999$). Effects on phenotype SD's were assessed by Bartlett's test in
132 one-way ANOVA models.

133

134 *Adjustment*

135 Prior to analysis BMI was adjusted for effects of age, gender and race/ethnicity in a linear model (age
136 + age² + race x gender). The adjustment model accounted for 4 % of the total variance in BMI
137 (Supplementary Table S1).

138

139 *Distributions*

140 Visualization

141 The effect of FH on the distribution of adjusted BMI was visualized using kernel-smoothed density
142 estimates by FH status (R base function density). The degree of smoothing is controlled by the
143 bandwidth parameter (bw) which was obtained in the full non-diabetic data set (bw = 0.99) from a
144 measure of the dispersion of the data (Sheather and Jones, 1991). This produces a continuous
145 distribution function and is widely used to visualise features of potential interest which may be
146 obscured in histograms. The credibility of the apparent effect of FH on the shape of the distribution
147 was assessed by post-hoc analysis of density ratios (FH₁ / FH₀) by quantiles (20) of the full sample.
148 Mean density ratios with SEM were obtained by quantile by bootstrap resampling with replacement
149 (1000 draws, stratified by FH status with resample sizes = strata sizes) and compared to the
150 predictions of normal and log-normal mixture distributions characterised by the proportions, means
151 and SD's in the sample stratified by FH status and cycle. Lack of fit to the mixture distributions was
152 assessed by X² tests in 1/variance-weighted linear regressions.

153

154 Conditional Quantile Regression (CQR)

155 Conditional quantile regression is a powerful tool for analyzing the effect of covariates on
156 distributions without assumptions of distributional shape. In contrast to ordinary least-squares (OLS)
157 regression which characterizes effects on global features of a distribution, CQR analyses local effects

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158 of covariates independently at any specified quantiles and can detect variations in covariate effects
159 across quantiles. Originating in econometrics (Koenker, 2017) it is now used in other areas including
160 genetics (Briollais and Durrieu, 2014). In the CQR framework developed by Abadi et al (Abadi et al.,
161 2017) for analysis of genomic markers, trends in effect sizes across quantiles represent interactions
162 between genetic effects and unobserved environmental and/or genetic factors. We treat FH as a
163 binary genetic risk marker (FH_0/FH_1) and a linear trend in quantile regression coefficients (β_{1i}) across
164 quantiles (τ_i) represents summed linear interactions of FH with unobserved factors. We analysed the
165 effects of FH on adjusted BMI by CQR using the R package quantreg. The effect of all interactions
166 on the FH effect was tested in a 2 parameter linear model:

167 for each quantile τ_i in y ,

$$168 \quad y(\tau_i | FH=fh) = \beta_{0i} + \beta_{1i} * FH$$

169 where $y(\tau_i | FH=fh)$ = the i th quantile of adjusted BMI conditional on the value of FH (0,1), the
170 intercept β_{0i} is the i th quantile value in FH_0 and β_{1i} is the FH effect size in quantile i .

171

172 The interaction between FH and continuous calendar time was estimated in the ANCOVA model:

$$173 \quad y(\tau_i | FH=fh) = \beta_{0i} + \beta_{1i} * FH + \beta_{2i} * yr + \beta_{3i} * FH * yr$$

174 where yr = cycle start year – 1999, β_{0i} is the i th quantile value in FH_0 at $yr = 0$ and β_{2i} and β_{3i} are CQR
175 coefficients for time and time*FH interaction effects in quantile i . The coefficients for the time-
176 related effects represent the effects in FH_0 (β_{2i}) and the additional effects in FH_1 (β_{3i}) so that the
177 coefficients for total time effects in $FH_1 = \beta_{2i} + \beta_{3i}$.

178

179 Equality of CQR parameter effect sizes across quantiles was tested using the `anova.rq` function in the
180 R package `quantreg`.

181

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182 The strengths of the CQR effects across quantiles were also assessed in linear meta-regression
183 analyses of relationships between quantile coefficients β_{1-3i} and quantiles using the R package
184 metafor (Abadi et al., 2017). Regression coefficients (β_{MR}) with SEM are reported in units of $\text{kg}\cdot\text{m}^{-2}$
185 over the full quantile scale (0-1). The structure of the FH effects in relation to the multi-modal
186 Mendelian hypothesis was assessed in an analysis of residuals from linear OLS fits of β_{1i} to β_{0i} with
187 Durbin-Watson tests for residual 1st order autocorrelation (function `durbinWatsonTest`
188 in the R package `car`).

189 Parametric fits

190 We obtained fits to a 3-component normal mixture distribution representing a simple Mendelian
191 model of fixed genetic effect using an expectation-maximization algorithm (`normalmixEM` function
192 in the R package `mixtools`). The models are characterised by the fitted means (μ_i), standard deviations
193 (σ_i) and mixing proportions (λ_i) of the three component distributions. Full model fits were obtained
194 in FH_1 but were not obtainable in FH_0 or DM_1 groups and we constrained μ_i in those groups to values
195 estimated in FH_1 in order to obtain comparable estimates of σ_i and λ_i . Risk allele frequencies (q)
196 under an additive Mendelian model of large genetic effects were calculated from the fitted λ_i :

197
$$q = 0.5*\lambda_2 + \lambda_3$$

198 where λ_2 and λ_3 represent the proportions of carriers of 1 and 2 risk alleles respectively. Within-
199 sample consistency of calculated q across the three groups analysed was assessed by comparing
200 fitted q_{FH_1} with the prediction from random mating of DM_1 into the full sample:

201
$$\text{predicted } q_{\text{FH}_1} = (q_{\text{DM}} + n\text{-weighted mean}(q_{\text{DM}}, q_{\text{FH}_1}, q_{\text{FH}_0}))/2$$

202 *Secular trends*

203 Effects of diabetes family history status (FH₀, FH₁) and continuous time (yr = calendar start year -
204 1999) on adjusted BMI and its standard deviation (SD) were assessed in ANCOVA models of the
205 form:

$$206 \quad y = \beta_0 + \beta_1 * FH + \beta_2 * yr + \beta_3 * FH * yr$$

207 where y = adjusted BMI mean or SD by FH status (0/1) and cycle, and yr = cycle start year – 1999.

208 Each OLS fit estimated 4 parameters from the 6 data points. Mean parameter estimates with 95% CI
209 were obtained by bootstrap resampling with replacement (1000 draws stratified by FH status and
210 cycle with resample sizes = strata sizes).

211

212 *Comparison of cross-sectional and secular trend effects*

213 Effects of FH on BMI distribution and on secular trends in BMI were compared by calculating the
214 contribution (%) of FH₁ to the effect in the full non-diabetic sample for calculated risk allele
215 frequency (q%) and to the slope (β%) of the relationships between time and BMI in ANCOVA model
216 described above. Mean (± SEM where possible) q% and β% were calculated in the relevant bootstrap
217 samples.

218

219 **Results**

220 **Participant characteristics**

221 Data from 9102 non-diabetic subjects met the inclusion criteria, approximately equally distributed
222 across the 3 cycles. Gender balance varied little but there was a cycle effect in race/ethnicity, most
223 obvious in the reduced representation of MA in the two later cycles. Average age varied across cycles
224 but not its SD, while adjusted BMI and its SD showed linear trends with cycle time. FH₁ prevalence

225 was higher in the two later cycles compared to 1999-2000 as was DM₁ prevalence. Current smoking
226 status was predominantly missing in the data (55%) and was not included in the BMI adjustment
227 model. However smoking status was not related to FH status whether analysed in the full data ($\chi^2 =$
228 2.80, 2 df, $p = 0.25$) or in those with non-missing smoking status ($\chi^2 = 0.43$, 1 df, $p = 0.51$), hence is
229 unlikely to confound analyses of FH effects. The mean age at diagnosis of DM (43.6 yr) is consistent
230 with predominant type 2 DM in the sample.

231 **Distributions**

232 Visualization

233 Adjusted BMI in the non-diabetic sample showed an apparently unimodal distribution, right-skewed
234 compared to a normal model and closer to a log-normal model (Fig 1A). When visualized by FH
235 status (Fig 1B) the predicted multimodality in FH₁ was indicated with modes in the normal weight,
236 overweight and obese regions of the BMI distribution. In contrast FH₀ showed an apparently
237 unimodal distribution. A difference in shape between the two groups was supported by the analysis of
238 density ratios (Fig 1B inset) in which models based on mixtures (FH x cycle) of either normal or log-
239 normal distributions did not provide adequate fits to the data ($p \leq 0.001$). BMI distribution in the
240 diabetic sample appeared to be depleted in the lower mode and enriched in the upper modes
241 compared to FH₁ (Fig 1C).

242

243 CQR

244 Analysis of the effect of FH status alone on the shape of the BMI distribution using CQR
245 demonstrated increasing FH effect size at higher levels of BMI ($\beta_{1MR} = 2.2 \pm 0.2$ (SEM) $\text{kg}\cdot\text{m}^{-2}$ Fig
246 2A, main panel), indicating strong interactions between FH₁ and other variables not included in the
247 model. FH₁ effect size ranged from $< 1 \text{ kg}/\text{m}^2$ in the lower quantiles to $\sim 3 \text{ kg}/\text{m}^2$ in the upper
248 quantiles, substantially different in both regions to the OLS estimate ($1.7 \text{ kg}/\text{m}^2$). Inclusion of

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249 calendar time in the two-way model weakened the trend in β_1 across quantiles ($\beta_{1MR} = 1.5 \pm 0.3$, Fig
250 2B main panel) and exposed significant OLS effects and trends across quantiles for main ($\beta_{2MR} =$
251 0.07 ± 0.02) and interaction ($\beta_{3MR} = 0.11 \pm 0.03$) effects of time (Fig 2C-D, main panels). The total
252 OLS and MR interaction effect sizes in FH₁ ($\beta_{2OLS} + \beta_{3OLS} = 0.11 \pm 0.03$ (SE), $\beta_{2MR} + \beta_{3MR} = 0.18 \pm$
253 0.04) were approximately double those in FH₀ (β_{2OLS} , β_{2MR} , Fig 2C). The analysis supports the
254 conclusion that calendar time is a strong surrogate of OE interacting with genetic factors as
255 represented by FH status, and that susceptibility to OE increases with increasing BMI in both groups
256 but more strongly in FH₁.

257 While the overall MR relationships between β_1 and quantiles in both models were approximately
258 linear (Fig 2A,B) there was strong evidence for additional non-linear structure in the OLS analysis of
259 β_{1i} against β_{0i} (insets Fig 2A,B). The linear models provided good fits (one-way: slope = $0.15 \pm$
260 0.003 (SE), $R^2 = 0.98$; two-way: slope = 0.10 ± 0.01 , $R^2 = 0.86$) but residual sequential structure was
261 apparent in both models, confirmed by tests of autocorrelation in residuals ($p_{DW} \leq 0.002$). The
262 pattern of residuals in the two-way model (Fig 2B inset), adjusted for time-related effects, shows
263 clear signs of discrete effects of FH₁ on the distribution of adjusted BMI with distinct peaks in the
264 lower, middle and upper regions of the distribution. This pattern in the conditional quantile
265 coefficients does not map directly onto the unconditional quantile plots in Fig 1B, but does highlight
266 similar regions in the distribution, and permits the conclusion that FH₁ has discrete, not continuous
267 effects on BMI.

268 Parametric analysis

269 The distribution of BMI in FH₁ (Fig 1B) and the discrete pattern in the FH₁ effect by CQR (Fig 2B
270 inset) appear consistent with a simple Mendelian model and fitting a 3-component normal
271 distribution model to the FH₁ data resulted in robust estimates of component means (Fig 3A) and
272 mixing coefficients and SDs (Table 2). Approximately 50% of FH₁ occupied the upper two modes
273 and separation between modes accounted for approximately 40% of the total variance in adjusted

274 BMI with the remainder assigned to dispersion within modes (Fig 2B). Under a Mendelian model the
275 variance due to mode separation represents a lower bound on the contribution of large effects as some
276 of the dispersion within modes represents variance in effect sizes of individual contributing causal
277 loci (see Discussion) which will contribute to the ~60% of variance assigned to within-modes.
278 Estimates of component SDs and mixing proportions with component means, constrained for FH_0 and
279 DM_1 to those identified in the FH_1 data (Table 2), support enrichment in the two upper components in
280 FH_1 compared to FH_0 (48% vs. 33%) and more strongly in DM_1 (72%). Predicted risk allele
281 frequencies in FH_1 (q - Table 2) express these distributional properties in Mendelian terms and show
282 within-sample consistency in that q_{FH_1} predicted from random mating of DM_1 (0.37) is not different
283 to the fitted estimate (0.30 ± 10).

284

285 **Secular trends**

286 Adjusted BMI mean (Fig 2A) and SD (Fig 2B) increased over the sampling period significantly faster
287 in FH_1 compared to FH_0 in the bootstrapped ANCOVA model, and estimates of β and $\Delta\beta$ in the mean
288 data were indistinguishable from the OLS estimates provided by the CQR analysis (Fig 2C,D).
289 Similar results were obtained with log-transformed BMI (Supplementary Fig S2). FH_1 accounted for
290 62% of the BMI mean trend and 60% of the BMI SD trend in this sample over the period 1999-2014,
291 effects similar in magnitude to the estimated FH_1 contribution to the sample risk allele frequency
292 (50%, Supplementary Table S2).

293

294 **Discussion**

295 **Summary**

296 We tested the prediction of segregation of discrete effects of FH on adult BMI (Jenkins et al., 2013),
297 modeled as modes of distribution, and estimated the contribution of FH_1 to recent trends in BMI

298 mean and dispersion in a large population-based sample. The results support a predominant role in
299 the recent obesity "epidemic" for rare genetic variants with large effects interacting with OE.

300

301 **Segregation of genetic susceptibility**

302 The non-parametric analysis provided evidence for a multi-modal distribution in the FH_1 group
303 consistent with the prediction of segregation of large genetic effects in families (Jenkins et al., 2013).
304 Multi-modality was supported by the analysis of density differences between FH_1 and FH_0 by
305 unconditional quantiles (Fig 1B) and by evidence of discrete signals in the OLS analysis of CQR
306 coefficients (Fig 2A&B insets). The two upper peaks in Fig 2B inset are consistent with the predicted
307 Mendelian effects of large effect variants on BMI but the potential lower peak is unexpected and may
308 reflect the presence of type 1 diabetes family history in FH_1 group. Individuals with type 1 diabetes
309 often present with BMI in the underweight (<18.5) – normal range (<25) (Manyanga et al., 2016) but
310 a genetic basis for this has not been established.

311 Polygenic risk scores (PRS) in population-based samples are expected to be unimodally-distributed,
312 and appear to be so (Llewellyn et al., 2014; Rask-Andersen et al., 2017). Any elevated polygenic
313 obesity risk in DM_1 will dilute into the mating population resulting in a right-shifted distribution in
314 FH_1 compared to FH_0 , not discrete effects. Alternative explanations for multi-modality might be
315 discrete stratification of OE components not captured by calendar time which seems unlikely, or un-
316 modeled interactions between FH and other covariates. Un-modeled interactions between FH and
317 stratified residual confounders may exist and contribute but we found no evidence for this in plots of
318 distributions by gender and race/ethnicity (Supplementary Fig S1) or in an analysis of smoking status
319 against FH. Discrete inheritance of genetic variants with large effects is the most likely explanation
320 for multi-modality in the FH effects on the BMI distribution .

321

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322 Approximately 40% of the adjusted BMI variance in FH_1 was accounted for by between-modes
323 variance (Fig 3B) but this represents a lower bound since the identified modes are likely to be
324 synthetic ie composed of a range of effect sizes due to rare variants at different loci. Indications of
325 fine structure within the broad central peak (Fig 2B inset) are suggestive. Examples of rare variants
326 with large effects on BMI in adults (β) are known from studies of candidate genes and monogenic
327 obesity loci (Jenkins and Campbell, 2014) while more recently a common variant in Samoans (EAF =
328 0.26, $\beta \approx 1.4 \text{ kg/m}^2$), very rare in other populations (Minster et al., 2016), and an African-specific
329 rare variant (EAF = 0.008, $\beta = 4.6 \text{ kg/m}^2$) undetected in Europeans and Asians (Chen et al., 2017)
330 have been identified by GWAS. Overall, β in these nine examples ranges from 1.4- 9 kg/m^2 and a
331 similar range in effect sizes in the NHANES sample would contribute substantially to the within-
332 mode variance estimated here. A combination of within-subject variance (~5% (Wormser et al.,
333 2013)) with polygenic variance (~ 5% (Loos, 2018)) sets a lower bound for within-modes variance
334 and hence the upper bound for between modes, implying that between 40% and 90% of total variance
335 in FH_1 may be attributed to large genetic effects.

336

337 **G x OE**

338 FH_1 is a prevalent (36%) and powerful determinant of the rate of change of mean BMI and its
339 dispersion over time in the ANCOVA models, accounting for 62% of the BMI trend and 60% of the
340 BMI SD trend in this sample over 1999-2014. Consistent results were obtained in the CQR models
341 with β_{OLS} and β_{MR} in FH_1 approximately double those in FH_0 . Under a polygenic model the familial
342 risk would be distributed normally over FH_1 which would then be a marker of a large fraction of the
343 at-risk population. However under the Mendelian model supported here genetic risk would segregate
344 in families and only approximately 50% of FH_1 would acquire the excess familial risk and only ~18%
345 of the sample would then account for ~60% of the trends. Individuals with a family history DM_1 must
346 represent a fraction of individuals with elevated genetic obesity risk and it is likely that the

347 remainder, particularly those with a family history of obesity without DM, would substantially
348 increase the genetic component of the trends consequent to the high heritability of BMI (Stryjecki et
349 al., 2018). This Mendelian model is internally consistent in estimates of risk allele frequencies (q) in
350 FH₁, FH₀ and DM₁ (Table 2) and in comparisons of FH₁ effect sizes in cross section (q , ~50%) and
351 over time (β , ~60%) (Supplementary Table S2). Our results support the proposition that the largest
352 part, and perhaps all, recent trends in mean and dispersion of BMI are due to a minor subset of
353 individuals with elevated genetic susceptibility to OE.

354

355 **Limitations**

356 The design and interpretation of fits to parametric mixture distribution models involves choices
357 concerning the number of components, parameter starting values and algorithms, and fit to a specific
358 model cannot be taken in isolation as support for its structural validity. We base our choice and
359 structural interpretation of 3-component normal mixture model fits and parameters on the *a priori*
360 hypothesis of Mendelian segregation of obesity risk in families (Jenkins et al., 2013) supported by the
361 non-parametric distributional plots (Fig 1B,C) and CQR analysis (Fig 2A,B insets). Like Abadi et al
362 (Abadi et al., 2017), we interpret interactions in the CQR analysis as predominantly G x OE although
363 a contribution from G x G interactions cannot be excluded. Our interpretation is supported by the
364 effects on the interaction of including calendar time in the CQR model (Fig 2B). Other limitations
365 discussed above include our inability to exclude discreet stratification of OE and the possible
366 influence of unmeasured/unknown confounders of FH effects.

367

368 **Conclusions**

369 We conclude that a Mendelian model of individually rare but collectively common genetic risk
370 variants with large effects interacting with OE provides a plausible quantitative explanation for recent

371 trends in obesity and should be favored over a predominantly polygenic model which does not. The
372 evidence for a predominant role for polygenes (Khera et al., 2019) can appear to be strong (eg
373 “Polygenic obesity is the most common form of obesity in modern societies...” (Albuquerque et al.,
374 2017)) but recent interpretations seek to explain the still missing heritability in obesity in terms of
375 unidentified large genetic effects and G x OE (Saeed et al., 2018; Loos, 2018) and recommend a
376 renewed focus on family-based designs and on specific populations in which large effect variants
377 may be enriched and identified (Minster et al., 2016; Chen et al., 2017). Our results strengthen that
378 view by showing that a model based on unidentified segregating variants with large effects
379 interacting with OE can account for the largest part of the secular trend in BMI and its dispersion in a
380 large population-based sample.

381

382 Conflict of Interest

383 The authors declare that the research was conducted in the absence of any commercial or
384 financial relationships that could be construed as a potential conflict of interest.

385

386 Author Contributions Statement

387 All authors contributed to the study design. AJ extracted and analysed the data in consultation
388 with MB. AJ and LC wrote the first draft of the manuscript. All authors contributed to manuscript
389 revision and editing.

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393 References

- 394 Abadi, A. et al. (2017) Penetrance of Polygenic Obesity Susceptibility Loci across the Body Mass
395 Index Distribution. *Am J Hum Gen*, **101**, 925-938.
- 396 Albuquerque, D., Nóbrega, C., Manco, L. & Padez, C. (2017) The contribution of genetics and
397 environment to obesity. *Br Med Bull*, **123**, 159-173.
- 398 Briollais, L. & Durrieu, G. (2014) Application of quantile regression to recent genetic and -omic
399 studies. *Hum Genet*, **133**, 951-966.
- 400 Chen, G. et al. (2017) Genome-wide analysis identifies an African-specific variant in SEMA4D
401 associated with body mass index. *Obesity*, **25**, 794-800.
- 402 Di Cesare, M. et al. (2016) Trends in adult body-mass index in 200 countries from 1975 to 2014: a
403 pooled analysis of 1698 population-based measurement studies with 19.2 million participants.
404 *Lancet*, **387**, 1377-1396.
- 405 Franks, P.W. & McCarthy, M.I. (2016) Exposing the exposures responsible for type 2 diabetes and
406 obesity. *Science*, **354**, 69-73.
- 407 Ghosh, A., Liu, T., Khoury, M.J. & Valdez, R. (2010) Family History of Diabetes and Prevalence of
408 the Metabolic Syndrome in U.S. Adults without Diabetes: 6-Year Results from the National
409 Health and Nutrition Examination Survey (1999-2004). *Public Health Genom*, **13**, 353-359.
- 410 Hall, K.D. (2018) Did the Food Environment Cause the Obesity Epidemic? *Obesity*, **26**, 11-13.
- 411 Hemminki, K., Li, X., Sundquist, K. & Sundquist, J. (2010) Familial risks for type 2 diabetes in
412 Sweden. *Diabetes Care*, **33**, 293-297.
- 413 Jenkins, A.B., Batterham, M., Samocha-Bonet, D., Tonks, K., Greenfield, J.R. & Campbell, L.V.
414 (2013) Segregation of a latent high adiposity phenotype in families with a history of type 2

- 415 diabetes mellitus implicates rare obesity-susceptibility genetic variants with large effects in
416 diabetes-related obesity. *PLoS One*, **8**, e70435.
- 417 Jenkins, A.B. & Campbell, L.V. (2014) Future management of human obesity: understanding the
418 meaning of genetic susceptibility. *Adv Genomics Genet*, **4**, 219-232.
- 419 Jenkins, A.B. & Campbell, L.V. (2015) Variation in Genetic Susceptibility Drives Increasing
420 Dispersion of Population BMI. *Am J Clin Nutr*, **101**, 1308.
- 421 Joyner, M.J., Paneth, N. & Ioannidis, J.P. (2016) What Happens When Underperforming Big Ideas in
422 Research Become Entrenched? *JAMA*, **316**, 1355-1356.
- 423 Khera, A.V. et al. (2019) Polygenic Prediction of Weight and Obesity Trajectories from Birth to
424 Adulthood. *Cell*, **177**, 587-596.e9.
- 425 Kivimaki, M., Stenholm, S. & Kawachi, I. (2015) The widening BMI distribution in the United
426 States. *Am J Clin Nutr*, **101**, 1307-1308.
- 427 Koenker, R. (2017) Quantile Regression: 40 Years On. *Ann Rev of Econ*, **9**, 155-176.
- 428 Krishna, A., Razak, F., Lebel, A., Davey Smith, G. & Subramanian, S.V. (2015) Trends in group
429 inequalities and interindividual inequalities in BMI in the United States, 1993–2012. *Am J Clin*
430 *Nutr*, **101**, 598-605.
- 431 Llewellyn, C.H., Trzaskowski, M., Plomin, R. & Wardle, J. (2014) From modeling to measurement:
432 developmental trends in genetic influence on adiposity in childhood. *Obesity*, **22**, 1756-1761.
- 433 Locke, A.E. et al. (2015) Genetic studies of body mass index yield new insights for obesity biology.
434 *Nature*, **518**, 197-206.
- 435 Loos, R.J. (2018) The genetics of adiposity. *Curr Opin Genet Dev*, **50**, 86-95.
- 436 Manyanga, T., Sellers, E.A.C., Wicklow, B.A., Doupe, M. & Fransoo, R. (2016) Is the change in
437 body mass index among children newly diagnosed with type 1 diabetes mellitus associated with
438 obesity at transition from pediatric to adult care. *Pediatric Diabetes*, **17**, 599-607.

- 439 Minster, R.L. et al. (2016) A thrifty variant in CREBRF strongly influences body mass index in
440 Samoans. *Nat Genet*, **48**, 1049-1054.
- 441 Müller, M.J. et al. (2018) The case of GWAS of obesity: does body weight control play by the rules.
442 *Int J Obes*,
- 443 Nagpal, S., Gibson, G. & Marigorta, U. (2018) Pervasive Modulation of Obesity Risk by the
444 Environment and Genomic Background. *Genes*, **9**, doi: 10.3390/genes9080411.
- 445 R Development Core Team (2016) R: A Language and Environment for Statistical Computing. R
446 Foundation for Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>,
- 447 Rask-Andersen, M., Karlsson, T., Ek, W.E. & Johansson, A. (2017) Gene-environment interaction
448 study for BMI reveals interactions between genetic factors and physical activity, alcohol
449 consumption and socioeconomic status. *PLoS Genet*, **13**, e1006977.
- 450 Razak, F., Smith, G.D., Krishna, A., Lebel, A. & Subramanian, S.V. (2015) Variation in genetic
451 susceptibility drives increasing dispersion of population BMI Reply. *Am J Clin Nutr*, **101**,
452 1308-1309.
- 453 Rokholm, B., Silventoinen, K., Ängquist, L., Skytthe, A., Kyvik, K.O. & Sørensen, T.I. (2011)
454 Increased genetic variance of BMI with a higher prevalence of obesity. *PLoS One*, **6**, e20816.
- 455 Saeed, S., Arslan, M. & Froguel, P. (2018) Genetics of Obesity in Consanguineous Populations:
456 Toward Precision Medicine and the Discovery of Novel Obesity Genes: Obesity and
457 Consanguinity. *Obesity*, **26**, 474-484.
- 458 Scott, R.A. & Consortium, I. (2013) The link between family history and risk of type 2 diabetes is not
459 explained by anthropometric, lifestyle or genetic risk factors: the EPIC-InterAct study.
460 *Diabetologia*, **56**, 60-69.
- 461 Sheather, S.J. & Jones, M.C. (1991) A Reliable Data-Based Bandwidth Selection Method for Kernel
462 Density Estimation. *J Roy Stat Soc B Met*, **53**, 683-690.

Segregation of Familial Obesity Risk

- 463 Silventoinen, K. et al. (2017) Differences in genetic and environmental variation in adult BMI by sex,
464 age, time period, and region: an individual-based pooled analysis of 40 twin cohorts. *Am J Clin*
465 *Nutr*, **106**, 457-466.
- 466 Speakman, J.R., Loos, R.J.F., O’Rahilly, S., Hirschhorn, J.N. & Allison, D.B. (2018) GWAS for
467 BMI: a treasure trove of fundamental insights into the genetic basis of obesity. *Int J Obes*, **42**,
468 1524-1531.
- 469 Stryjecki, C., Alyass, A. & Meyre, D. (2018) Ethnic and population differences in the genetic
470 predisposition to human obesity. *Obes Rev*, **19**, 62-80.
- 471 Tirosh, A. et al. (2011) Adolescent BMI trajectory and risk of diabetes versus coronary disease. *N*
472 *Engl J Med*, **364**, 1315-1325.
- 473 Willemssen, G. et al. (2015) The Concordance and Heritability of Type 2 Diabetes in 34,166 Twin
474 Pairs From International Twin Registers: The Discordant Twin (DISCOTWIN) Consortium.
475 *Twin Res Hum Genet*, **18**, 762-771.
- 476 Wormser, D., White, I.R., Thompson, S.G. & Wood, A.M. (2013) Within-person variability in
477 calculated risk factors: Comparing the aetiological association of adiposity ratios with risk of
478 coronary heart disease. *Int J Epidemiol*, **42**, 849-859.
- 479 Xu, G. et al. (2018) Prevalence of diagnosed type 1 and type 2 diabetes among US adults in 2016 and
480 2017: population based study. *BMJ*, **362**, k1497.

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484 Figure legends

485 Figure 1: Distribution of adjusted BMI in non-diabetic (DM_0) and diabetic (DM_1) participants in
486 combined NHANES 1999-2000, 2005-2006 and 2013-2014 cycles. A) Full non-diabetic sample
487 ($n=9102$) binned by quantiles ($n=20$) with superimposed kernel-smoothed and fitted densities in
488 normal and log-normal models. B) Main: kernel-smoothed adjusted BMI density by FH status. Inset:
489 density ratios (FH_1 / FH_0) \pm SEM obtained by bootstrap resampling by quantiles of the full sample
490 compared to predictions of normal (solid line) and log-normal (dotted line) mixture models with p
491 associated with lack-of-fit testing (p_{lof}). C) Kernel -smoothed adjusted BMI density in non-diabetic
492 FH_1 ($n=3297$) compared to diabetic participants ($n=793$).

493 Figure 2: Conditional quantile regression effects of diabetes family history (FH) on adjusted BMI in
494 non-diabetic participants in models consisting of FH alone (A) and in interaction with continuous
495 calendar time (B-D). The main panels show the effect sizes (β_{1-3}) by quantile with 95% CI (grey-
496 shaded area), the OLS estimate of the average effect (solid green line) with 95% CI (dotted green
497 lines) with p-values (p_{ANOVA}) from anova tests for equality of β_i across quantiles and the meta-
498 regression fits \pm 95% CI (magenta lines) with $\beta_{MR} \pm$ SEM. The insets in panels A and B show the
499 patterns of residuals ($\Delta\beta_{1i}$) \pm residual SEM from linear OLS fits of β_{1i} (FH_1) to β_{0i} (FH_0) with 95% CI
500 on the fits (dotted lines) around the lines $\Delta\beta_1 = 0$ representing perfect fits, and with the p value (p_{DW})
501 from a Durbin-Watson test of autocorrelation of residuals. CQR estimates of the SEM of β_{0i} are also
502 depicted but are mostly obscured by the point symbols.

503

504 Figure 3: A) Adjusted BMI density in FH_1 by quantile (grey bars) and kernel-smoothed (black line)
505 with fits to a three component normal mixture distribution. B) Estimated contributions in FH_1 of the
506 components of the mixture distribution to the prevalence (mixture coefficients, λ) and variance of
507 adjusted BMI.

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508 Figure 4: Effects of diabetes family history ($FH_{0/1}$) on linear secular trends in age-, gender- and
509 race/ethnicity-adjusted BMI mean \pm sem (A) and standard deviation \pm sem (B). Parameter estimates
510 with 95% CI were obtained in ANCOVA models by stratified bootstrap resampling of all non-
511 diabetic individuals (see Methods). Dotted lines enclose 95% CI on fitted values at each point; $\beta =$
512 regression slope vs. time (kg/m^2 per year); $\Delta\beta = \beta_{FH1} - \beta_{FH0}$ (95%CI).
513

514 Tables

Table 1: Participants by diabetes status (DM₀/DM₁)

	NHANES cycle				p [†]
	All cycles	1999-2000	2005-2006	2013-2014	
DM₀					
n	9102	2865	3076	3161	-
Gender (F%)	53	55	54	52	0.06
Race/Ethnicity (%) (MA/OH/NHW/NHB) [‡]	23/8/46/23	30/7/44/20	24/4/49/24	16/11/48/25	7.8 x 10 ⁻⁵⁶
Age (yr): Mean	40.5	40.7	39.4	41.5	4.5 x 10 ⁻⁹
SD	13.2	13.2	13	13.2	0.63
BMI (kg/m ²): Mean	28.7	28.3	28.6	29.2	4.6 x 10 ⁻⁹
SD	6.6	6.3	6.4	7.1	1.2 x 10 ⁻¹¹
Diabetes Family History (Y%)	36	29	42	38	4.1 x 10 ⁻²⁵
Current smoking (%, Y/N/missing)	25/20/55				-
DM₁					
n (%)	793 (8.0)	211 (6.9)	252 (7.6)	330 (9.5)	0.0003
Age at Diagnosis (yr)	43.6	44.3	42.9	43.6	0.47

515

516 † Cycle effects (p) by ANOVA (age), ANCOVA (BMI), Bartlett's test (SD's) and Chi-squared test for categorical

517 variables.

518 ‡ MA = Mexican American, OH = Other Hispanic, NHW = Non-Hispanic White, NHB = Non-Hispanic Black

519 * Adjusted for age, gender and race/ethnicity in a linear model (see Table S1).

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Table 2: Three-component normal mixture distribution fits to adjusted BMI by FH and DM status^Y

	Component	DM ₀		DM ₁
		FH ₀	FH ₁	
mean	1	*	25.8±1.0	*
	2	*	32.1±1.8	*
	3	*	40.6±2.5	*
sd	1	3.8	3.8±0.9	3.7
	2	5.7	5.1±1.2	4.5
	3	10	8.2±0.8	8.4
λ	1	0.67	0.52±0.16	0.28
	2	0.29	0.37±0.15	0.47
	3	0.04	0.11±0.06	0.25
q [†]	-	0.18	0.30±0.10 (0.37) [§]	0.49

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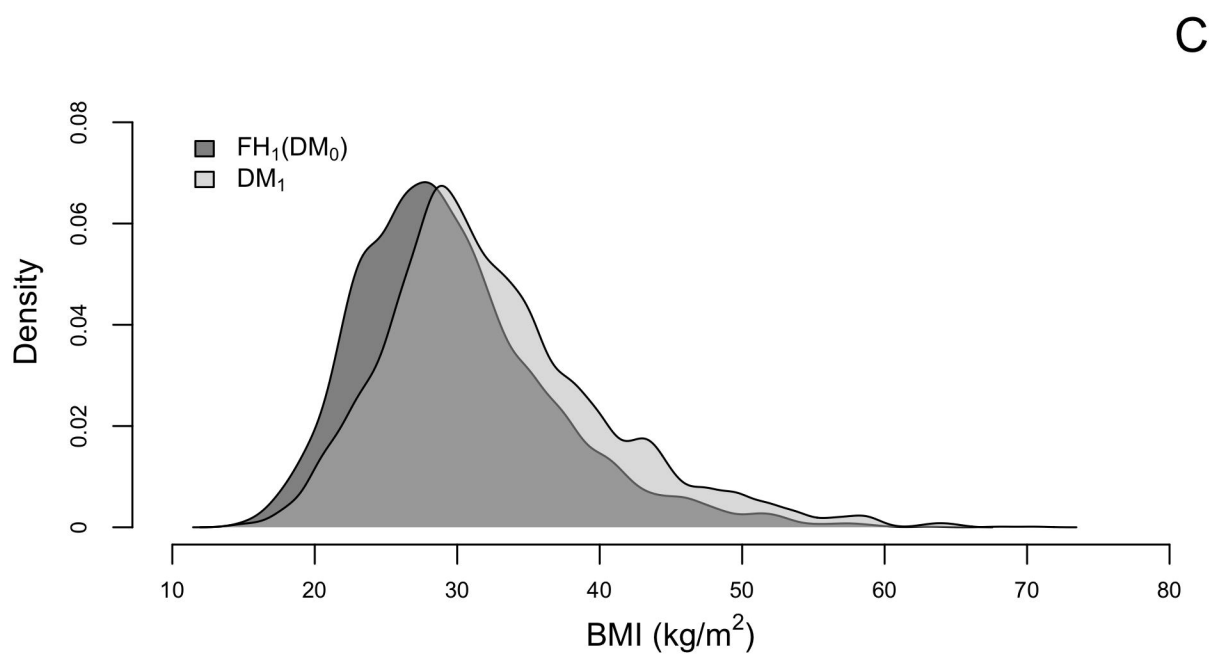
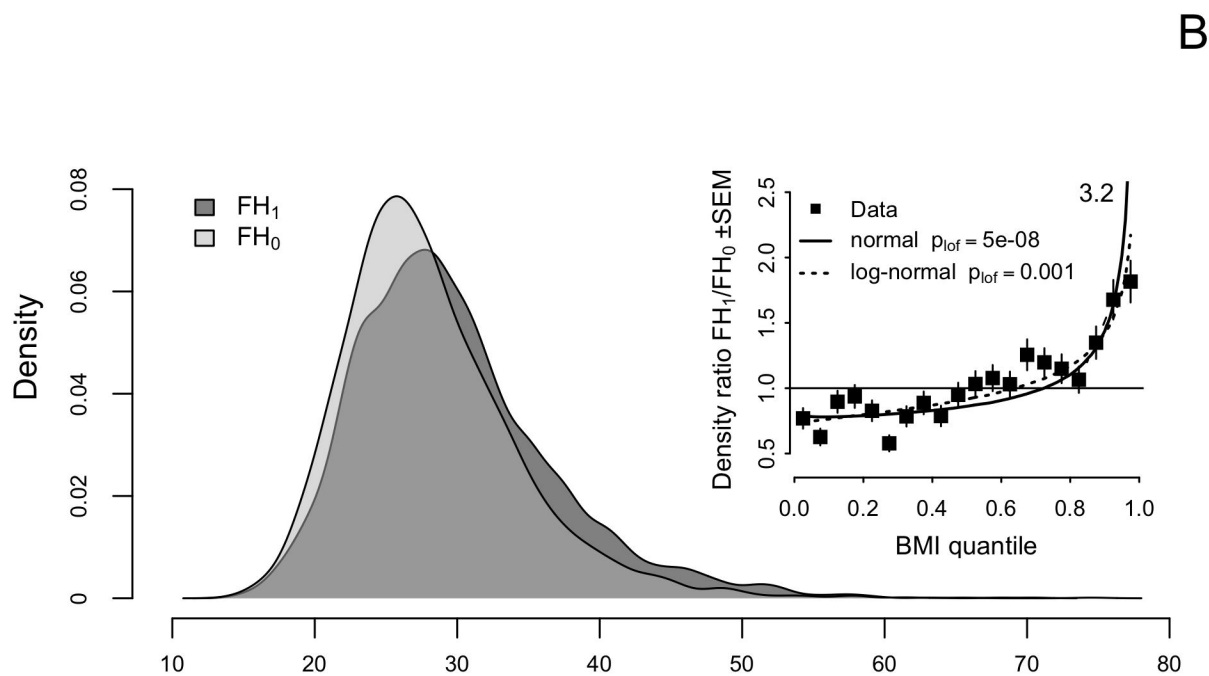
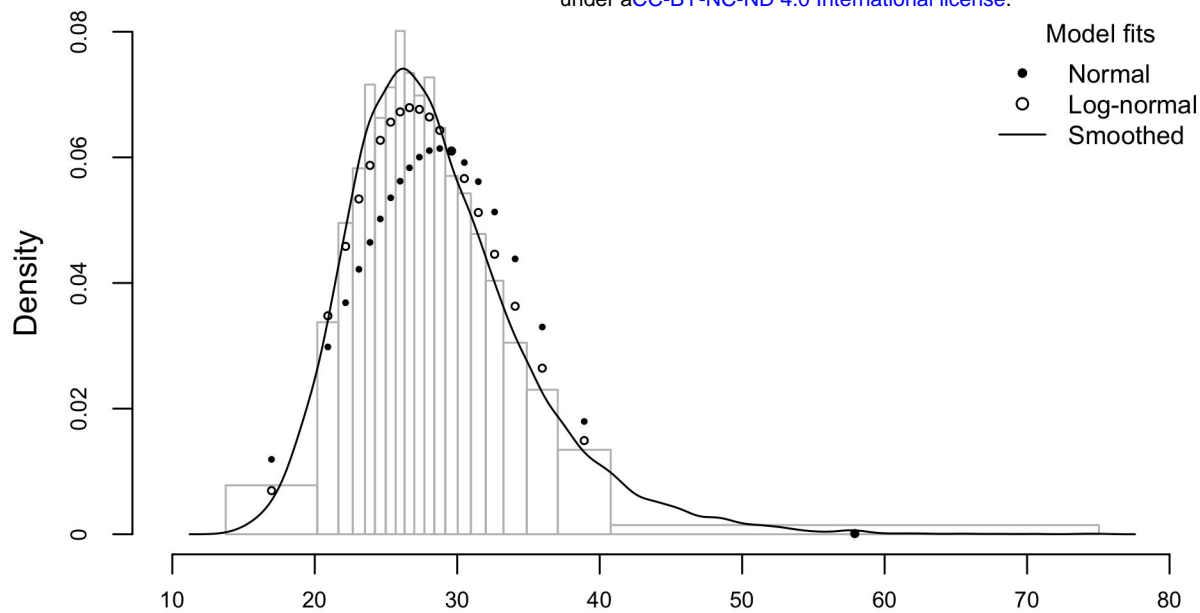
523 ^Y ± bootstrap standard error for FH₁ only

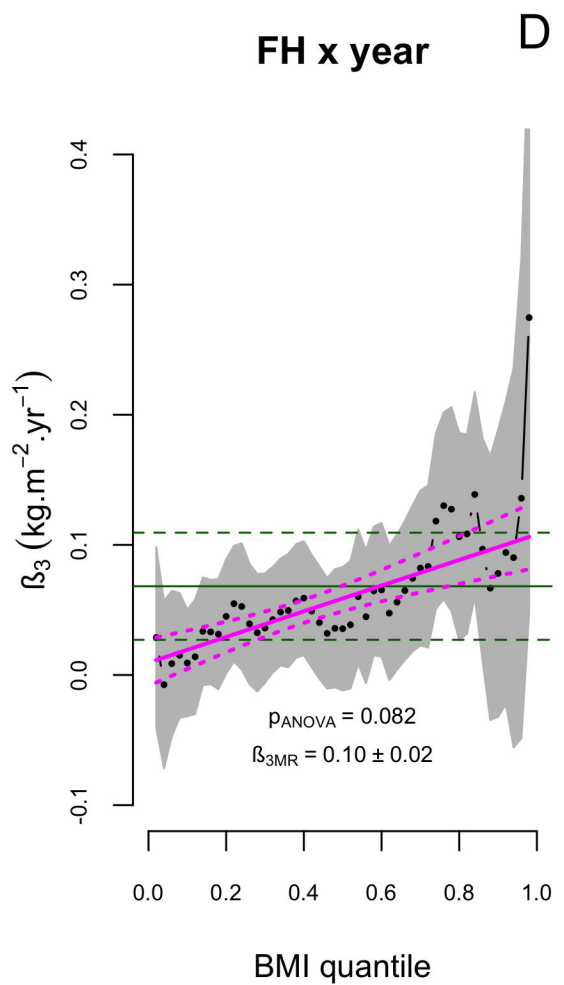
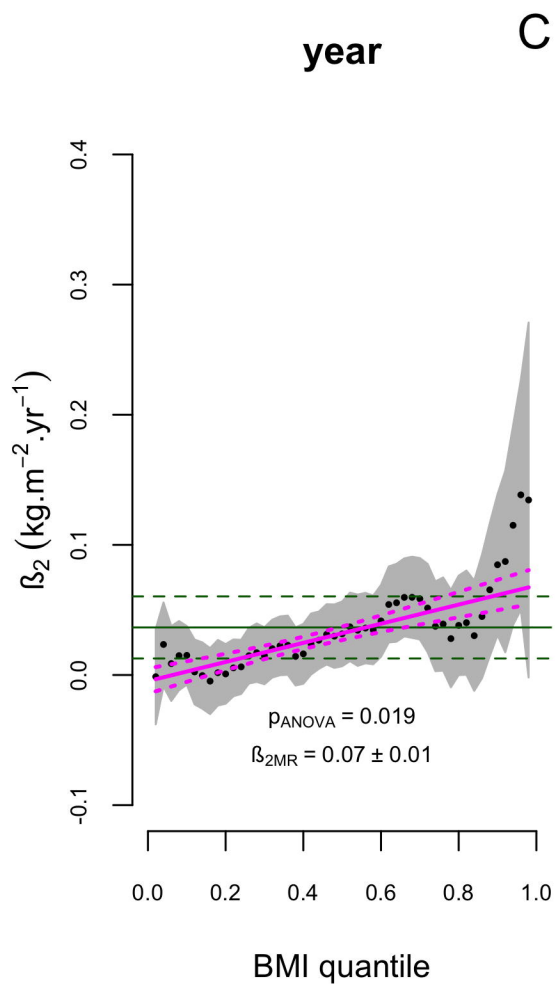
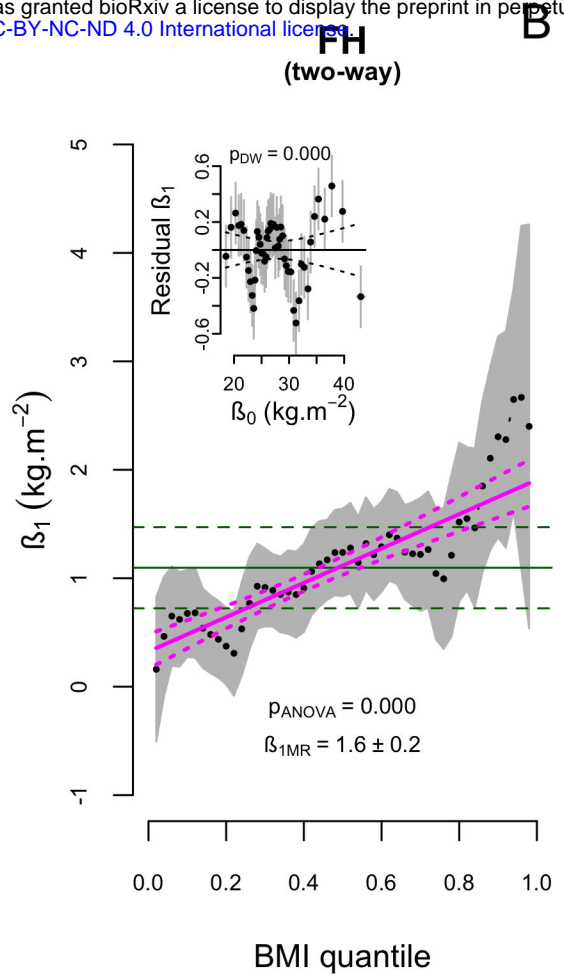
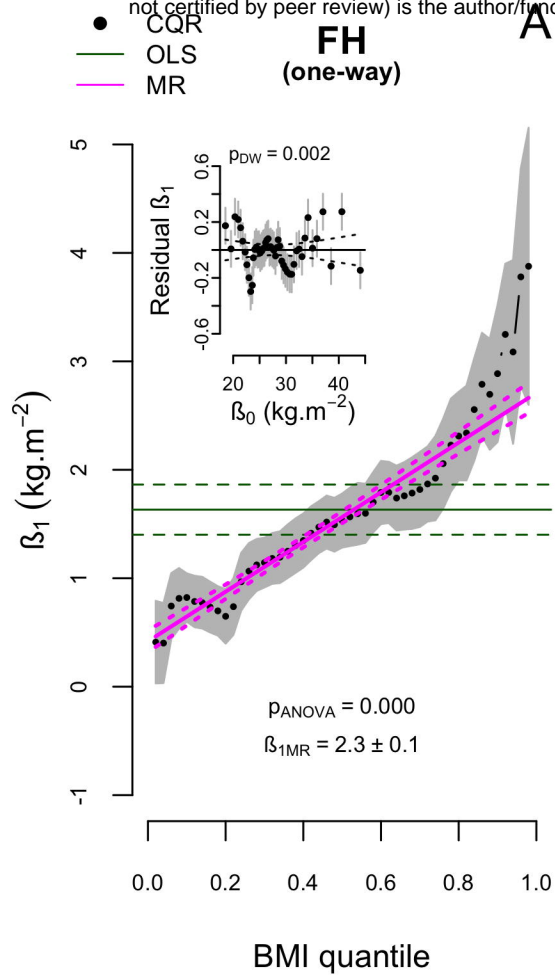
524 * Means in FH₀ and DM₁ constrained to fitted values in FH₁

525 [†] Calculated risk allele frequency in an additive Mendelian model of large effects

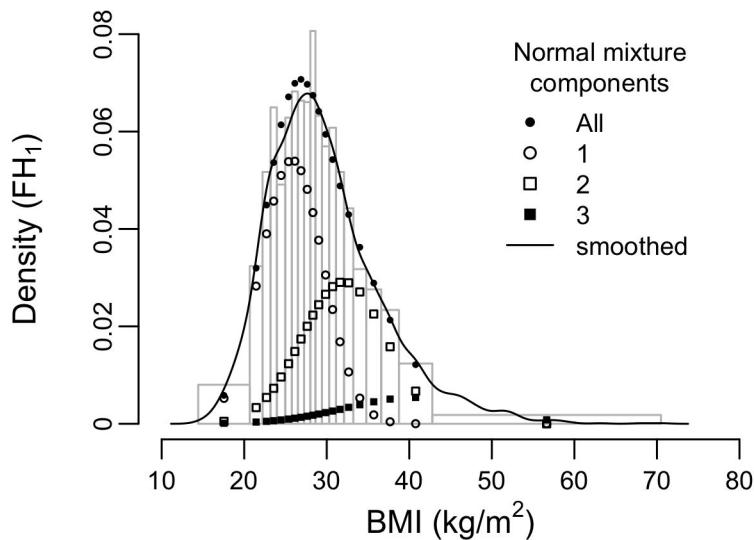
526 [§] Predicted from DM₁ mating randomly into the full sample.

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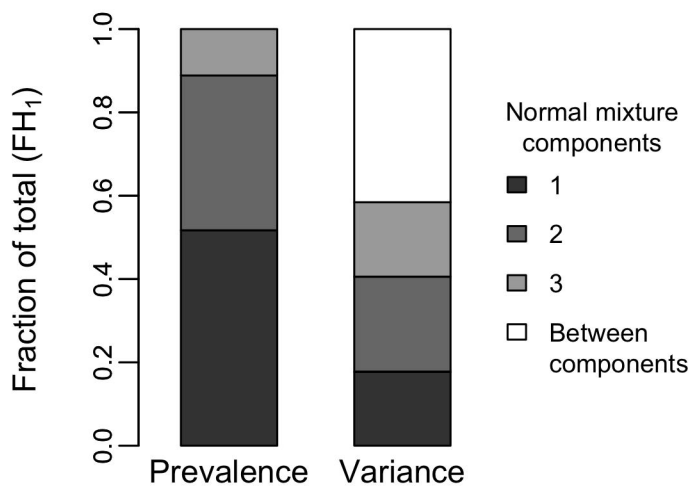




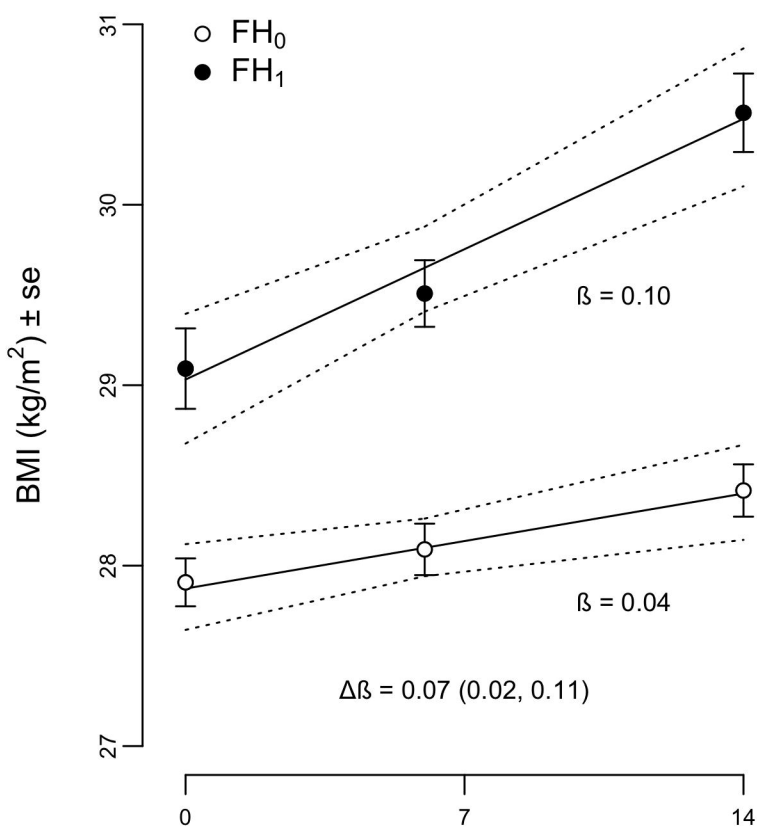
A



B



A



B

