

1 Assessing the potential of environmental DNA metabarcoding for monitoring  
2 Neotropical mammals: a case study in the Amazon and Atlantic Forest, Brazil

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21

22 **Abstract**

23 The application of environmental DNA (eDNA) metabarcoding as a biomonitoring  
24 tool has greatly increased in the last decade. However, most studies have focused  
25 on aquatic macro-organisms in temperate areas (e.g., fishes). We apply eDNA  
26 metabarcoding to detect the mammalian community in two high-biodiversity regions  
27 of Brazil, the Amazon and Atlantic Forest. We identified critically endangered and  
28 endangered mammalian species in the Atlantic Forest and Amazon respectively and  
29 found congruence with species identified via camera trapping in the Atlantic Forest.  
30 In light of our results, we highlight the potential and challenges of eDNA monitoring  
31 for mammals in these high biodiverse areas.

32

33 **Keywords:** Camera traps; critically endangered; eDNA; river

## 34 **Introduction**

35 A quarter of mammal species are endangered according to the IUCN Red List of  
36 Threatened Species (IUCN 2019) and there is clearly a need for more cost-effective  
37 and rapid methods for long-term biomonitoring to be applied across different biomes  
38 and over large spatial and temporal scales (Sales et al. 2019a). In recent years,  
39 environmental DNA (eDNA) metabarcoding (the simultaneous identification via next-  
40 generation sequencing of multiple taxa using DNA extracted from environmental  
41 samples, e.g., water, soil) has delivered on its initial potential and is now  
42 revolutionizing how we monitor biodiversity (Deiner et al. 2017). The majority of  
43 eDNA metabarcoding applications have focused on monitoring fish and  
44 macroinvertebrates, with mammals being targeted in only 8% of vertebrate studies  
45 (Tsuji et al. 2019). However, with the development of universal primers for  
46 vertebrates and mammals specifically, there has been a recent surge in studies  
47 tailored to detect and/or monitor mammalian communities in terrestrial and  
48 freshwater environments (e.g. Ushio et al. 2017, Harper et al. 2019, Sales et al.  
49 2019a).

50 Several recent mammal-focused eDNA metabarcoding studies in temperate  
51 regions in the northern hemisphere have relied on well-studied systems with  
52 accompanying long-term or historical survey data to test the efficiency of this novel  
53 biomonitoring tool (e.g., Harper et al. 2019, Sales et al. 2019a). However, mammal  
54 conservation can be more challenging in biodiversity-rich countries as long-term  
55 monitoring systems are still scarce outside of Europe and North America (Proença et  
56 al. 2017) and ecological field studies, usually used to plug this gap, are often  
57 hindered due to difficulties of sampling over wide spatial scales. For effective

58 conservation action, adequate knowledge regarding the biodiversity components  
59 present in each area is of paramount importance.

60 Environmental DNA from lentic and lotic systems has been found to be  
61 effective in not just monitoring aquatic and semi-aquatic mammals, but also  
62 terrestrial species (Harper et al. 2019, Sales et al. 2019a). Here, we explore the  
63 application of eDNA metabarcoding for Neotropical mammals by verifying its ability  
64 to detect aquatic and terrestrial animals from rivers/streams in the highly biodiverse  
65 biomes of the Brazilian Amazon and Atlantic Forest. The Amazon is the largest  
66 tropical rainforest on Earth, encompassing at least 10% of the world's biodiversity.  
67 The Atlantic Forest, which is currently represented by only 11% of its original cover  
68 (Ribeiro et al. 2009), is the second most biodiverse biome in South America (WWF  
69 2018).

70

## 71 **Methods**

72 In the Amazon, water samples (500mL each, in three replicates) were obtained from  
73 six sites within three main areas (A-C; Fig. 1; Table S1). In the Atlantic Forest, water  
74 and sediment samples (500mL of water and 25mL of sediment, in three replicates)  
75 were obtained from eight sites located in two valleys of the Caparaó National Park  
76 (D-E; Fig. 1; Table S1). Temperature and pH were recorded at each site in the  
77 Amazon. Mammal-specific universal primers targeting the mitochondrial 12S rRNA  
78 gene were used (Ushio et al. 2017). The workflow was conducted following the  
79 protocol described in Sales et al. (2019a) and a more detailed description is included  
80 in the Supporting Information.

81 Additional data regarding species' distribution in the Atlantic Forest were  
82 obtained through camera-trap surveys. Both valleys in the Caparaó National Park

83 were surveyed with terrestrial and arboreal camera traps (Bushnell Trophy Cam™,  
84 USA; see Supporting Information).

85

## 86 **Results and Discussion**

87 A total of ~1.3 million mammal reads were obtained after all the bioinformatic filtering  
88 steps (Amazon – 833,623 reads; Caparaó – 109,233 reads for water samples and  
89 334,593 for sediment samples). Only reads recovered for wild mammals (919,910  
90 reads) were retained for downstream analyses.

91 Overall, we detected 28 Molecular Operational Taxonomic Units (MOTUs -  
92 Blaxter et al. 2005) from terrestrial and aquatic mammals, representing eight orders  
93 and 14 families (Table S2). Considering a threshold of >0.97 minimum identity, only  
94 13 MOTUs could be assigned to the species level (Table S2). In the Amazon, six  
95 species were recovered, with three currently listed as endangered by the IUCN's  
96 Red List (IUCN 2019) in different categories: the Endangered Amazon river dolphin  
97 (*Inia geoffrensis*), the Vulnerable giant anteater (*Myrmecophaga tridactyla*) and the  
98 Vulnerable lowland tapir (*Tapirus terrestris*). Three Least Concern species were also  
99 identified: *Thyroptera discifera* and *Rhynchonycteris naso* in the order Chiroptera  
100 and the rodent *Toromys rhipidurus*. Only one MOTU was detected for each family  
101 (Fig. 1).

102 In Caparaó National Park, nine families were detected using eDNA: five in the  
103 west side of the park (D) and nine in the east side (E; Fig. 1 and S1). Of these, only  
104 seven could be assigned to the species level (Table S2). Here, camera-trap surveys  
105 detected 17 species (and additional unidentified small mammal species),  
106 encompassing 12 families (Fig. S2; Table S3). Combining the two non-invasive  
107 techniques, 15 families were detected overall (Table 1), six of which by both

108 methods, three exclusively by eDNA metabarcoding and six solely by the camera  
109 traps. Although this study was not designed to provide a direct comparison between  
110 methods (e.g., Harper et al. 2019, Sales et al. 2019a), it highlights the potential of  
111 implementing multiple non-invasive approaches in providing an overview of the  
112 mammalian community composition in biodiversity rich areas.

113 More MOTUs were retrieved for the families detected in the Atlantic Forest,  
114 suggesting the occurrence of several species of the same family in this area. For  
115 example, three MOTUs were recovered in the east side and two from the west side  
116 of the Park for both Didelphidae and Cuniculidae. Camera trapping recorded three  
117 species of Didelphidae (*Caluromys philander*, *Didelphis* sp., *Philander frenatus*), in  
118 accordance with the eDNA data. Only one species from the Cuniculidae (*Cuniculus*  
119 *paca*) recorded by camera traps is known to occur in the Caparaó and the existence  
120 of three MOTUs for this family might be due to natural intraspecific genetic variability  
121 (Fig. 1). Cricetidae had three MOTUs in the west side of the Park: although this  
122 family was not identified by camera traps, several species are described for the  
123 Atlantic Forest, including endemic and recently described species (Gonçalves &  
124 Oliveira 2014). Furthermore, the Critically Endangered primate *Brachyteles*  
125 *hypoxanthus* was detected using eDNA, demonstrating its potential to detect  
126 arboreal mammals from water samples (Harper et al. 2019).

127 As a similar sampling effort was applied for both areas in this study, there is a  
128 need to consider what factors might explain the difference in the number of MOTUs  
129 recovered for each biome, particularly if we assume that mammalian alpha diversity  
130 should at least be as high in the Amazonian sampling sites as in the Caparaó forest  
131 site (see Costa et al. 2000). For example, all the families detected in the Atlantic  
132 forest that were not detected in the Amazonian samples are known to occur in Area

133 B of the Amazon (Mendes Pontes et al. 2008). DNA degradation in water is one of  
134 the main factors reducing detectability over time and limiting temporal inferences.  
135 The sampled black waters in the Amazon have low pH (ranging from 3.85 to 4.27),  
136 whereas in the Caparaó the reported values are above 6.5 (Rodrigues 2015). Acidic  
137 environments show higher eDNA decay and lower persistence rate due to the  
138 increased degradation of DNA via chemical hydrolysis (Seymour et al. 2018).  
139 Therefore, the eDNA recovered in the low pH waters of the Amazon might be  
140 derived from specimens which have had very recent contact with the water body.  
141 Mammal eDNA recovery depends not only on the species presence but also on the  
142 direct/indirect contact with the water system (Harper et al. 2019). The junction of the  
143 Negro and Amazon Rivers (area C) has an enormous volume of water and possibly  
144 much time had elapsed since it flowed under the forest canopy, but the other  
145 Amazonian streams (area B; Fig. 1) are similar in size to those in the Atlantic Forest.  
146 In the Amazon, all species/MOTUs were detected in a single replicate, except for the  
147 lowland tapir (detected in four replicates in three different streams). This species is  
148 known to defecate more frequently in water than on land (Tobler et al. 2010) so this  
149 may explain its higher rates of eDNA detection. In the Atlantic Forest, several  
150 MOTUs/species were recovered from multiple replicates/sites (Fig. S1), suggesting  
151 longer persistence of eDNA in this environment.

152         There is a clear limitation in terms of available DNA sequences in public  
153 databases (e.g., Genbank) to either match identified MOTUs to species, or to  
154 distinguish between closely related species within the same genus. This issue has  
155 been highlighted in previous Neotropical eDNA studies for other taxonomic groups  
156 (Cilleros et al. 2019, Sales et al. 2019b). A 12S reference database exists for 164  
157 Amazonian mammalian species in French Guiana (Kocher et al. 2017) and all

158 Amazonian MOTUs were identified to species level here. However, this was not the  
159 case for the Atlantic Forest. This biome hosts more than 300 mammalian species  
160 (more than 50% of medium/large species considered at least Vulnerable; Souza et  
161 al. 2019). Therefore, for eDNA monitoring to be implemented in this biome, there is a  
162 clear need to generate reference DNA barcodes of a large proportion of the  
163 mammalian communities present.

164 Here, we demonstrated the potential of applying a cutting-edge non-invasive  
165 and cost-effective molecular approach for biodiversity assessment and systematic  
166 monitoring scheme of Neotropical mammals, including highly threatened species.  
167 This is particularly relevant given the current political climate in Brazil, which is  
168 resulting in research funding and environmental crises. However, significant  
169 challenges remain to implement this method in the Neotropics, from a better  
170 understanding of the ecology of eDNA within these variable environments, to the  
171 current lack of appropriate reference sequences for species determination in these  
172 biodiversity-rich and anthropogenically-impacted biomes.

173

#### 174 **Acknowledgments**

175 This project was partially funded by the University of Salford Internal Research  
176 Award awarded to CB, ADM and IC. We are grateful to Vitor Borges for assistance in  
177 the field. The present study was carried out with all required permits (ICMBIO N.  
178 54795-2, DEFRA 126191/385550/0).

179

#### 180 **Author Contributions**

181 NGS, MDCK, ADM, CB, IC, JPB, WEM and MNFS conceived, and NGS, MDCK,  
182 ADM, IC and CB designed the study. NGS, MDCK, AH and CB carried out the eDNA



183 sampling. NGS, MDCK and JCP performed the laboratory work. NGS carried out the  
184 bioinformatic analyses. MDCK analysed the camera trap data. NGS, ADM and IC  
185 analysed the eDNA data. NGS, ADM and MDCK wrote the paper, with all authors  
186 contributing to editing and discussions.

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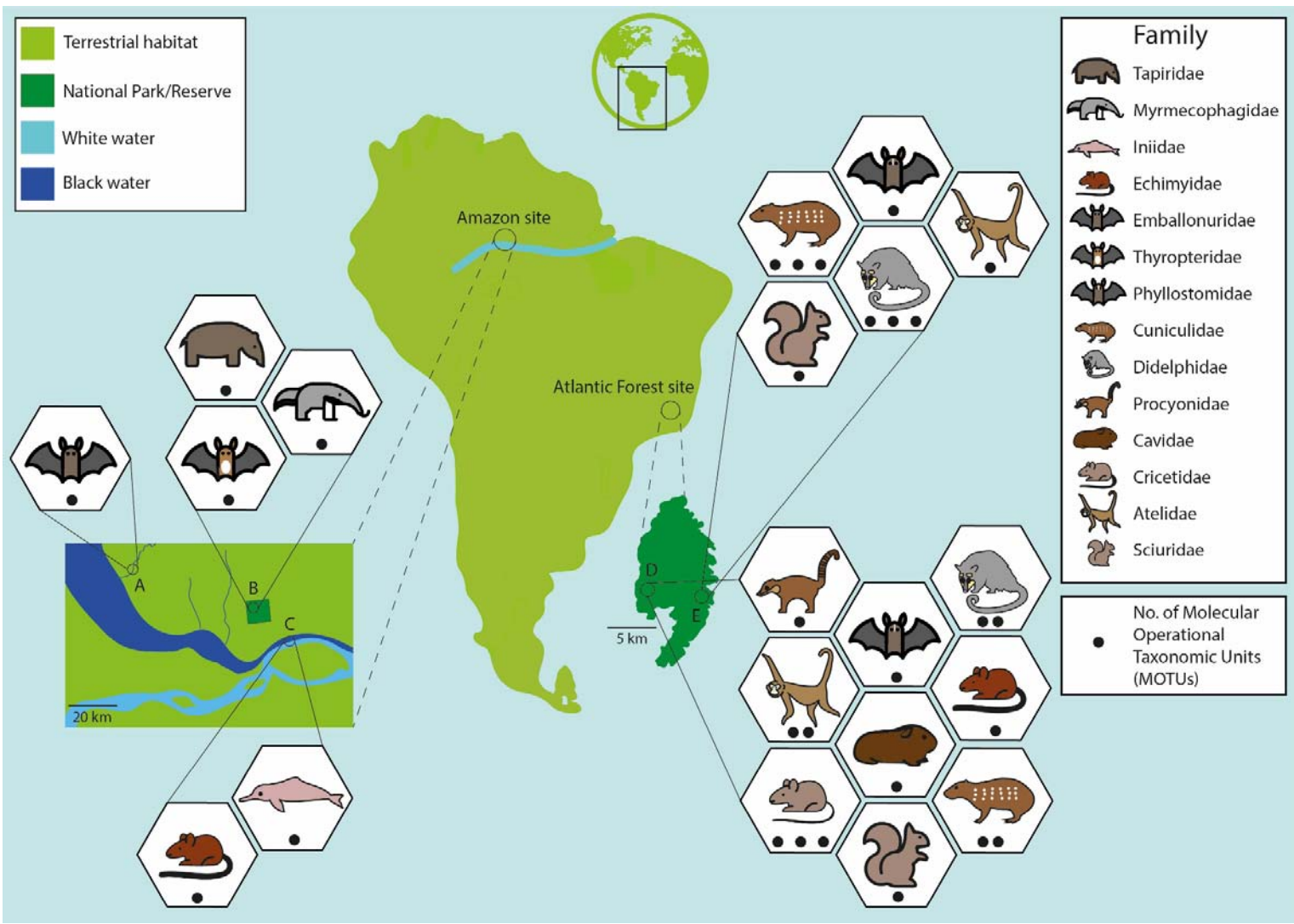
251 **Table 1.** Number (*n*) of species captured with camera traps and number of Molecular  
252 Operational Taxonomic Units (MOTUs) captured with environmental DNA (eDNA)  
253 metabarcoding for orders and families within Caparaó National Park, Atlantic Forest.  
254 See Tables S2 and S3 for a more extensive breakdown of camera trap and eDNA  
255 data, respectively.

Order	Family	Camera ( <i>n</i> species)	eDNA ( <i>n</i> MOTUs)
<b>Carnivora</b>	Felidae	1	-
	Mustelidae	1	-
	Procyonidae	2	1
<b>Chiroptera</b>	Phyllostomidae	-	2
<b>Didelphimorphia</b>	Didelphidae	3	3
<b>Pilosa</b>	Myrmecophagidae	1	-
<b>Primates</b>	Atelidae	1	2
	Callithrichidae	1	-
	Cebidae	1	-
<b>Rodentia</b>	Caviidae	1	1
	Cricetidae	-	3
	Cuniculidae	1	3
	Echimyidae	2	1
	Erethizontidae	2	-
	Sciuridae	1	1

256

257 **Figure legend**

258 Figure 1. Sampling areas for environmental DNA (eDNA) in the Amazon (A-C) and  
259 Atlantic Forest (D-E) biomes in Brazil. The families recovered from eDNA  
260 metabarcoding in each area are represented by stylized drawings and the number of  
261 Molecular Operational Taxonomic Units (MOTUs) recovered within each family is  
262 indicated.



263

264

Figure 1.