Life history genomic regions explain differences in Atlantic salmon marine diet specialization

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Abstract

1. Animals employ various foraging strategies along their ontogeny to acquire energy, and with

varying degree of efficiencies, to support growth, maturation and subsequent reproduction events.

Individuals that can efficiently acquire energy early are more likely to mature at an earlier age, as a

result of faster energy gain which can fuel maturation and reproduction.

2. We aimed to test the hypothesis that heritable resource acquisition variation that co-varies with

efficiency along the ontogeny would influence maturation timing of individuals.

3. To test this hypothesis, we utilized Atlantic salmon as a model which exhibit a simple, hence

trackable, genetic control of maturation age. We then monitored the variation in diet acquisition

(quantified as the stomach fullness and composition) of individuals with different ages, and linked it

genomic regions (haploblocks) that were previously identified to be associated with age-at-maturity.

4. Consistent with the hypothesis, we demonstrated that one of the life history genomic regions tested

(*six6*) was indeed associated with age-dependent differences in stomach fullness. Prey composition was

marginally linked to both genomic regions (*six6* and *vgll3*). We further showed Atlantic salmon switched

to the so-called "feast and famine" strategy along the ontogeny, where older age groups exhibited heavier

stomach content, but that came at the expense of running on empty more often.

5. These results suggest genetic variation underlying resource utilization variation may explain the genetic

basis of age structure in Atlantic salmon. Given that ontogenetic diet has a genetic component and the

strong spatial diversity associated with these genomic regions, we predict populations with diverse

maturation age will have diverse evolutionary responses to future changes in marine food-web structures.

Keywords: *Diet specialization, ontogenetic diet shift, life history evolution, ontogenetic foraging variation,*

Atlantic salmon.

Introduction

Diet acquisition is a strong evolutionary force that can shape population demography and abundance,

and is an integral determinant of ecosystem functions (Engen and Stenseth 1989, Svanback and

Persson 2004, Bolnick and Araujo 2011). Individuals exhibit differences in prey preference and prey

acquisition efficiency, which, if heritable, may be a target of selection and ultimately promote

ecological specialization (Fox and Morrow 1981, Smith and Skulason 1996, Devictor et al. 2010,

Elmer et al. 2010, Machovsky-Capuska et al. 2016, Sexton et al. 2017). Large-scale disturbances in

community structure, e.g., as a result of climate change (Sydeman et al. 2015) alter food web

structures and the composition of available resources (Daufresne et al. 2009, Pershing et al. 2015,

Bentley et al. 2017), forcing species to rapidly adapt to new diet landscapes. Therefore, understanding

the underlying mechanisms shaping food acquisition strategies is fundamental to evolutionary

biology and vital for predicting species survival in a changing world.

If heritable, the inter-individual variation in resource acquisition strategies may have complex

evolutionary consequences mediated by trade-offs between energy gain and survival across density-

and frequency-dependent fitness landscapes (Mousseau et al. 2000, Reznick and Ghalambor 2001,

Reznick 2016, Sexton et al. 2017). For example, increased boldness to improve resource acquisition

success may come at the expense of higher predation risk, the fitness costs of which may be linked

to predator densities (Gotthard 2000, Carter et al. 2010, Bolnick et al. 2011). Likewise, the

composition and abundance of available resources may alter the demographic structure of a population (e.g., Heino and Kaitala 1999, Enberg et al. 2012). For example, fast growth at an early age, e.g. as a result of abundant food sources during the initial stages of life, may result in early maturation and hence a younger age at reproduction. In contrast, resource limitation due to high population densities results in increased allocation to somatic growth to improve size-dependent intrapecific competition (e.g., Reznick and Endler 1982).

Ontogenetic diet shifts in organisms may be viewed as a special type of resource acquisition strategy in which diet variation is expressed as a function of age. Ontogenetic diet shift is a significant source of variation in species' diet breadth, especially among size- and age-structured organisms, such as fishes. In general, relatively large and/or old individuals shift towards feeding at higher trophic levels and/or on larger prey items to maintain a positive energy balance (Werner and Gilliam 1984, Mittelbach and Persson 1998, Jensen et al. 2012). Under changing food-web dynamics, diet specialization among different age groups may substantially influence the demographic structure and life history diversity (Sanchez-Hernandez et al. 2019). For example, changes in resource composition that favour younger age groups would improve growth and subsequently increase the rate of maturation and the probability of survival at early ages. Ontogenetic diet shift is associated with a suite of changes in an individual's morphology, physiology and behaviour to maximize the efficiency of particular resources at a given ontogenic stage, perhaps at the expense of reduced efficiency at other stages (Claessen and Dieckmann 2002).

If ontogenetic diet variation has a genetic basis, then some individuals in a population may be selected for high prey acquisition efficiency early in their life history (via physiological or morphological trade-offs towards efficient exploitation at earlier stages), even if this may come at a cost of compromised energy acquisition at later stages in life (Claessen and Dieckmann 2002). We predict that such genetically driven trade-offs in resource acquisition efficiency between early and late stages

mediate the age structure (i.e. maturation timing) and abundance within and among populations and

maintain genetic variation in resource acquisition strategies, but such examples in the wild are rare.

Atlantic salmon (*Salmo salar*) is a fish species recognized as a diet generalist and an opportunistic

feeder with extensive ontogenetic and stage- and space-structured individual variation in diet breadth

(Erkinaro et al. 1997, Jacobsen and Hansen 2001, Haugland et al. 2006, Hvidsten et al. 2009,

Rikardsen and Dempson 2010, MacKenzie et al. 2012). At sea, where most growth occurs, salmon

increasingly feed on prey at higher trophic levels as they grow and age (Jacobsen and Hansen 2001,

Rikardsen and Dempson 2010). The time salmon spend at sea prior to maturation (sea age at maturity)

also varies greatly within and among populations (Friedland and Haas 1996). Although the functional

and physiological basis underlying age at maturity is not entirely known, it is considered to be a

threshold trait, whereby higher lipid deposition is associated with early maturation (Friedland and

Haas 1996, Jonsson et al. 1997, Thorpe et al. 1998, Taranger et al. 2010, Jonsson and Jonsson 2011).

Therefore, variation in resource acquisition may be a strong determinant of the life history variation

in salmon and a trait via which natural selection can act and result in adaptive genetic changes in

populations.

In Atlantic salmon, two genomic regions on chromosomes 9 and 25 have been identified to have a

disproportionate influence on life history strategy and population differentiation within and among

populations (Ayllon et al. 2015, Barson et al. 2015, Czorlich et al. 2018, Pritchard et al. 2018, Aykanat

et al. 2019). The so-called *vgll3* and *six6* genomic regions are named after the most prominent genes

in their respective haploblocks on chromosomes 25 and 9, respectively. The *vgll3* genomic region on

chromosome 25 has been shown to be associated with age at maturity (intially by GWAS, see: Ayllon

et al. 2015, Barson et al. 2015), iteroparity (Aykanat et al. 2019), and precocious male maturation in

Atlantic salmon (Lepais et al. 2017, Debes et al. 2019). This genomic region also exhibits strong

spatial divergence (Barson et al. 2015, Pritchard et al. 2018), and it has recently been shown to have

been affected by natural selection over the last 36 years (equivalent to 4-6 salmon generations) in

parallel to the changing age structure in a large salmon population (Czorlich et al. 2018). The *six6*

region on chromosome 9 is associated with sea age at maturity at the population level as a result of

the strong correlation between the average allele frequency and average maturation age of populations

(Barson et al. 2015). This region also exhibits the strongest signal of differentiation among European

populations (Barson et al. 2015) and Tana/Teno River populations (Pritchard et al. 2018) and is hence

distinguished as a critical genomic region for local adaptation. Genes found in these haploblocks

appear to have a role in adipose or energy metabolism regulation in other organisms. The *vgll3* gene

is an adipocyte inhibitor, the expression of which is correlated with body weight and gonadal adipose

content in mice (Halperin et al. 2013). Recently, a strong selective sweep near the *vgll3* gene was

postulated to be due to energy metabolism effects in humans in Mongolia (Nakayama et al. 2017). In

turn, genes in the *six6* genomic region are involved in cell growth, cell differentiation, apoptosis in

human cell lines (*PPM1A*, Lin et al. 2006), and myogenesis and skeletal muscle cell proliferation in

zebrafish (*six1b*, Ridgeway and Skerjanc 2001, Bessarab et al. 2004, O'Brien et al. 2014) and act as

an evolutionarily conserved regulator of eye development and the pituitary–hypothalamic axis (*six6*,

Serikaku and Otousa 1994, Toy et al. 1998, Gallardo et al. 1999). Collectively, this suggests that the

vgll3 and *six6* haploblocks might have broad-scale roles in reproductive and life-history strategies in

Atlantic salmon. However, how polymorphism in these regions may be translated to functional

differences expressed in the wild is unclear.

Here, our objective was to test whether age-dependent differences in food acquisition efficiency are

associated with the *vgll3* and *six6* genomic regions and discuss their role in explaining the genetic

variation in age structure. We achieved this goal by assessing stomach content data from adult

Atlantic salmon sampled along the coast during spawning migration and genotyping the same

individuals for the *vgll3* and *six6* genomic regions using a targeted sequencing approach. Using a

modelling framework that accounted for potentially confounding environmental and phenotypic

variables, we tested whether variation in diet and resource acquisition strategies had a genetic

component explained by the age at maturity-linked genomic regions. Elucidating the genetic interplay

between age at maturity and diet breadth is crucial to better understand the dynamics and evolution

of ecological specialization and to better predict future demographic changes in Atlantic salmon

populations under climate change.

Materials and Methods

Sample collection

As part of a larger effort within the project "Sea salmon fishery, resource and potential (KOLARCTIC)",

Atlantic salmon (*Salmo salar*), on their return migration to spawning grounds, were sampled and

stomachs were collected between mid-May and late July in 2008 by local sea fishers with bend nets

or bag nets along the Finnmark coast, northern Norway (Svenning et al. 2019, Figure 1). Sampled

fish were measured (fork length, cm) and weighed (g); their sex and maturity were identified, and

stomachs were frozen for later diet analysis. In addition, scales were sampled from all fish for sea age

determination, categorization as wild or farmed fish according to ICES guidelines (ICES 2011,

Svenning et al. 2019), and genetic analysis. The species composition of the diet was then identified

to species by visual inspection of the morphology of prey remains and otoliths which were compared

to a reference collection with known species identity, with uncertain cases further inspected using

guide books (Härkönen 1986, Pethon and Nyström 2005). All prey items, including unidentified

digested remains were weighted. The identifiable portion of the diet in the dataset was

overwhelmingly comprised of four fish species: sand eel (*Ammodytes* spp.), capelin (*Mallotus*

villosus), herring (*Clupea harengus*), and haddock (*Melanogrammus aeglefinus*, see Results section

for details). In the interest of analytical brevity, a few rare prey species were handled as follows: one

gadoid fish was grouped with haddocks, both of which belong to the Gadiformes order, and negligible

amounts of krill, other crustaceans, and Liparidae (0.2% of the total stomach weight) were categorized

together with the unidentified material.

Figure 1: Map of the study region including sampling locations and a description of age and stomach content distributions and sample sizes in each sampling location. See the inset key for a description of the parameters of the spatial distributions given on the map.

DNA extraction, microsatellite genotyping, and SNP genotyping by targeted sequencing

DNA was extracted from scales either using a QIAamp 96 DNA QIAcube HT Kit (Qiagen) following

the manufacturer's protocol or according to (Elphinstone et al. 2003). Microsatellite genotyping of 31

markers was performed as outlined in (Ozerov et al. 2017). Samples were further genotyped by

targeted sequencing at 173 SNP markers and the sex determination locus (sdy) using a GTSeq

approach (Campbell et al. 2015) as outlined in (Aykanat et al. 2016), with some modifications so the

genotyping panel was compatible with the Illumina platform. More specifically, 174 genomic regions

were first amplified in one multiplex PCR using locus-specific primers with truncated Illumina

adapter sequences and using primer concentrations re-optimized for the Illumina platform (Supp.

Table 1). The PCR products were then treated with Exonuclease I and FastAP Thermosensitive

Alkaline Phosphatase (Thermo Fisher) to remove unused primers and nucleotides. After the

treatment, the products were re-amplified with adapter-specific primers containing Illumina and

sample-specific dual-indexes. The index set was optimized using the BARCOSEL software

(Somervuo et al. 2018). The PCR products were then pooled, purified and quantified with a Qubit 2.0

fluorimeter (Thermo Fisher) and analysed on a fragment analyser (Agilent Technologies). The pooled

library was then size selected using BluePippin (Sage Sciences) to remove short unspecific products

and checked on a fragment analyser. Finally, samples were single-end sequenced using a 150-cycle

high-output sequencing kit on a NextSeq 500 Illumina Sequencer following the manufacturer's

guidelines. Loci with coverage over 12x were scored as in (Aykanat et al. 2016). To calculate

coverage for each SNP, raw genotype files (fastq) were scanned for every SNP, and the coverage

was determined by counting sequences that matched SNP's forward and reverse primer sequences,

and the 9 bp region around the SNP site. Finally, genotypes were scored based on coverage ratios

between alleles: $Ratio_{Cov(allele1/allele2)} > 10$ was assigned as homozygous for allele 1, $Ratio_{Cov(allele1/allele2)}$

$< 0{\cdot}1$ was assigned as homozygous for allele 2, $Ratio_{Cov(allele1/allele2)}$ between $0{\cdot}2$ and 5 was assigned

as heterozygous and any proportion in between was discarded (see also, Campbell et al. 2015).

Two focal SNPs used in the analyses were $vgll3_{TOP}$, which exhibits the strongest signal of association

with age at maturity in the $vgll3$ genomic region, and $six6_{TOP.LD}$ in the $six6$ haploblock on chromosome

9, the region that exhibited the second strongest association with sea age at maturity prior to

population structure correction and is 34.5 kb away from and in complete linkage disequilibrium with

the $six6_{TOP}$ SNP reported in Barson et al. (2015).

Genetic stock identification (GSI)

In total, 2023 samples that had greater than 80% success in regard to microsatellite genotyping were assigned to their population of origin with 31 microsatellite markers as described in (Svenning et al. 2019) using the Bayesian GSI methodology described in (Pella and Masuda 2001) and implemented in cBayes 5.0.1 (Neaves et al. 2005). In brief, the samples were allocated into 18 analysis groups, that is, the combination of two time periods (May-June and July) and 10 fisheries regions, with each group consisting of 30 to 288 samples for analysis. The GSI analyses were performed using five independent Monte Carlo Markov chains of 100K iterations starting from three random stocks (StartStock parameter), and the last 10K iterations of each chain were combined and used to assign individuals to their population of origin to remove the influence of initial starting values. The baseline population data for the GSI analysis included genetic information on 185 Atlantic salmon populations spanning from the Pechora River (Russia) in the east to the Beiarelva River (Norway) in the west (see details in Ozerov et al. 2017).

The probability (p) threshold for assignment of an individual to a population was set at ≥0.7 following Vähä et al. (2011, 2014) and Bradbury et al. (2015). The 30% of individuals assigned to a population with lower confidence were kept in the dataset with the highest ranked population assigned as the population of origin. Samples with no assignment due to low genotyping success with microsatellites (4%) were assigned a population of origin using genotype information from the SNP panel. In such cases, individuals were assigned to the population in which they exhibited the highest genetic similarity. This was measured according to the average genetic similarity of focal individuals to the individuals in each population (as inferred with the GSI analysis in the previous step) using the *A.mat* function in the rrBLUP package (Endelman 2011) A small subset individuals (N=16, < 1%) with poor genotyping success with SNPs (less than 50 SNPs with high quality genotypes) was randomly assigned to a population, in which population assignment probability is weighted over the total number of individuals that were assigned by GSI. The effect of including incomplete population

assignment was assessed for the main analysis by repeating the analysis but only including

confidently assigned individuals.

Missing data points for some variables were inferred from highly correlated variables. In that regard,

missing sea age information (i.e., due to unclear formation of sea annulus for detecting the correct

sea-age for some first time spawners) was inferred from length data for 15 (0.7%) individuals, where

the likelihood of age, given the length, was substantially higher (>20 times) for the inferred age group

than for other age groups. Additionally, for 21 (1.0%) individuals with missing length data, fit using

coefficients of log(weight) to log(length) regression (adjusted $R^2 = 0.94$) was used to estimate the

length information from the weight data. Finally, data for 33 (1.6%) individuals with missing $vgll3_{TOP}$

genotype scores were inferred from the genotype score of an adjacent SNP marker in the genotyping

panel, $vgll3_{Met54Thr}$, which is in close physical proximity to $vgll3_{TOP}$ with high linkage disequilibrium

($r^2 = 0.79$).

Genetic and ecological basis of the diet scope

Unless otherwise noted, all statistical analyses were performed in R software v.3.2.5 (R Core Team,

2018). Either a two-component hurdle model (with binomial and the conditional negative binomial

components) using the *glmmTMB* package (Brooks et al. 2017) or a binomial model (with a log link

function) using the *gamm* function in the *mgcv* package (Wood 2011) was employed as the statistical

model. In all models, population of origin was included as a random term to account for background

population effects. To control for spatio-temporal variation, sampling location (longitude) and the

day of sampling (Julian day, zero centred, and scaled to one standard deviation) were included as

smoother terms. Longitude, which explained 90.7% of the spatial variation (i.e., sampling locations

mostly occurred along a longitudinal axis (see Figure 1) and were included in the models as a

surrogate for the two-dimensional spatial distribution to decrease the parametrization of the model.

In addition to including the genetic variation in the *six6* and *vgll3* genomic regions additively in the

model (i.e., genotypes coded as a continuous factor with heterozygotes coded as the average of two

homozygotes), age at maturity (e.g. Fleming 1996) and residual length (log transformed total length

after controlling for age at maturity) were also included in the model as categorical and continuous

variables, respectively. All numeric variables were centred and scaled. For both genomic regions

(*six6* and *vgll3*), alleles associated with late and early age at maturity were labelled as *L* and *E*,

respectively.

The general model structure was as follows:

$Z = SA + resL_{SA} + V_{SA} + S_{SA} + s(D) + s(L) + p + e$

where Z is the vector of response variables given as a function of sea age (SA), scaled residual length

and *vgll3* and *six6* genotypes nested within sea age (resL$_{SA}$, V$_{SA}$ and S$_{SA,}$ respectively), the smoother

functions of sampling day (D) and location (L), and normally distributed random variance due to the

population (*p*) and individual (residual) effects. The genotypes are coded additively as 1, 2, and 3 (for

EE, *EL* and *LL*, respectively). In the model, the scaled genotypes and residual length were analysed

independently within sea age group (i.e., nested model), which provided a statistical framework

suitable for testing hypotheses related to ontogenetic diet structure. In this model, a small number of

4SW individuals (N = 15, 0.7%, SW denotes number of winters salmon spent at sea prior to

sampling.) was grouped and analysed within the 3SW for statistical coherence of the nested model.

Models also accounted for spatio-temporal variation in the diet with smoothing spline functions.

When using the *gamm* package, which provides a platform for generalized additive models, days and

location were modelled with a smooth function (*s()*). When using *glmmTMB* package, which provides

a platform for hurdle models but cannot directly model the smoother functions, an orthogonal spline

design matrix with a low-rank thin-plate function was generated using the *spl* function in the

MCMCglmm package (Hadfield 2010) in R and included in the model as fixed terms as a surrogate

for the spatio-temporal spline functions. The number of knot points (*k*, which defines the curvature

of the spline function) was set to five for both variables, but the results were robust to an increase in

the *k* value, which did not qualitatively change the results (data not shown).

A number of variables pertaining to the diet content data were used as response variables in this study.

Conceptually, these variables are linked to different aspects of diet acquisition mechanisms of this

species (e.g., Arrington et al. 2002) and may indicate different functional aspects associated with

performance and life history variation among individuals.

1)   *The presence and amount of total diet content in the stomach:* We used a two-component

hurdle model to simultaneously account for stomach content quantity and empty stomach probability.

By this, we tested for the prevalence of the "feast and famine" diet acquisition strategy in Atlantic

salmon as a function of ontogeny and genotype, whereby large piscivorous fish species are predicted

to experience prolonged periods with empty stomachs in the interest of acquiring a high quantity of

food (Arrington et al. 2002, Armstrong and Schindler 2011). Both components in the hurdle model

included the same set of covariates (as described above). A logistic model with a log link was used

to model the probability of the presence of a prey item, whereas a zero truncated negative binomial

model with a log link was utilized as the conditional component. In this analysis, the total stomach

weight, which had excess zero elements and a right-tailed continuous distribution (ranging from 0.5

to 393.3 g), was transformed to a discrete distribution by arbitrarily binning the total weight in 10 g

increments, with zero stomach content set as the first bin at a value of zero (Supp. Figure 1). This

transformation provided a distribution that can be modelled with the hurdle framework in the

glmmTMB package (Brooks et al. 2017). Finally, we also repeated the analysis by only including

confidently assigned indiviuals in order to assess the robustness of model to incomplete population

assignment.

2)   *Total number and average weight of prey items in the stomach:* An increase in the total prey

weight in the stomach can be explained either by an increase in prey number or an increase in the

average prey weight. Therefore, we next investigated the contribution of these two components in

terms of explaining the model using the same statistical framework as above. Similar to the total prey

weight, the average prey item weight (ranging between 0.2 and 300.2 g) was also transformed to

discrete units by arbitrarily binning the data at 5 g intervals, with zero stomach content set as the first

bin (Supp. Figure 1).

3)   *Relative prey composition:* Finally, to test whether sea age, size at age, or genotype is

associated with specific prey species, we modelled the prey composition, measured as the proportion

of a specific prey species contributing to the stomach content weight. The proportion of each of the

four prey species in the total prey weight was modelled as a response variable using binomial

regression in the mgcv package (Wood 2011)

Extensively digested, unidentified content in the stomach (4.5% of the total stomach weight) was not

treated as diet material in order to accurately reflect the recent feeding activity (e.g., Jacobsen and

Hansen 2001). For all models, the effect size and confidence intervals were calculated with 10,000

parametric permutations of the model coefficients. To account for potential spurious inflation

associated with genotype, i.e. due to cryptic family structure, the analytical pathway was repeated

using 168 independent and putatively neutral markers that are present in the SNP panel, and focal

SNPs were ranked across the background genetic effect (by comparing the genetic model and the null

model at each SNP marker).

Results

The final dataset contained 2121 individuals after excluding previously spawned and escaped farmed

salmon. In the final dataset, 93.3% of the samples had visibly detectable developing gonads,

confirming concordance between sea age and sea age at maturity. A total of 1372 (64.7%) individuals

were confidently assigned to a population of origin ($p > 0.7$), and (N=651) Individuals assigned to a

population with lower confidence ($p < 0.7$, 30.7%). A further 82 individuals (3.9%) with low

genotyping success with microsatellites were assigned a population of origin using SNP data and 16

individuals (< 1%) lacking reliable genotypes with either set of genetic markers were randomly

assigned to a population (see Materials and Methods for details).

Stomach content analysis

Out of the 2121 individuals examined in the final dataset, 992 individuals had identifiable prey items

in their stomachs (46.8%). Four fish species, sand eel (*Ammodytes* spp.), capelin (*Mallotus villosus*),

herring (*Clupea harengus*), and haddock (*Melanogrammus aeglefinus*), comprised the bulk of the diet

content, representing 42.2 kg of the 44.2 kg quantified diet content (95.5%). In total, there were 2843

identifiable prey items in the datasets, with sand eel being the most abundant and herring being the

largest percentage by weight (Supp. Figure 2). On average, prey weight significantly differed among

species, with haddock being the heaviest, followed by herring, capelin, and sand eel (Supp. Figure

2).

Prey probability and weight in the stomach as a function of sea age, size at age, and genetic variation

The two-component hurdle model revealed a striking negative relationship between the probability

of non-empty stomach (e.g. presence of identifiable prey item in the stomach) and prey weight (g) in

the stomachs of Atlantic salmon as a function of sea age (Figure 2a-b, Supp. Table 2). Young age

groups were more likely than older age groups to have prey in their stomach. 1SW individuals were

1.45 (1.07 - 1.95, 90% CI, $p = 0.020$) and 2.39 (1.73 - 3.30, 90% CI, $p < 0.001$) times more likely to

have any prey item in their stomachs than 2SW and 3SW individuals, respectively, and 2SW

individuals were 1.65 times more likely to have a prey item in their stomachs than 3SW fish (1.35 -

2.00, 90% CI, $p < 0.001$). The decrease in non-empty stomach was significantly associated with

residual size variation within the 2SW age group ($p = 0.027$, Figure 2e), with larger individuals having

empty stomachs more often than the smaller-sized fish in the same age group.

The conditional truncated negative binomial model suggested that young age groups had significantly

Figure 2: Changes in four stomach content variables in relation to sea age (a) and size at age (b) and associated 90% confidence intervals. Only p-values < 0.1 are given in the figures. Binned prey weights are given after back transformation to the original scale (i.e., grams). Note that estimates in the last three columns (stomach weight, average prey number and weight) are for non-empty stomachs only.

less prey in their stomachs than older age groups despite a higher likelihood of having non-empty stomachs (Figure 2b). The contrasting results between the zero-inflated and truncated negative binomial components suggested that resource acquisition strategies differed among age groups. The model estimated, on average, 9.9 g (7.0 - 13.9, 90% CI), 24.8 g (21.8 – 28.6, 90% CI), and 41.5 g (33.8 – 50.9, 90% CI) of prey items in the stomachs of 1SW, 2SW and 3SW fish, respectively, all of which were highly significantly different from one another (p < 0.001). Residual length at age also appeared to be a predictor of prey content, but only significantly so in the 2SW age group ($p = 0.036$, Figure 2b).

The *six6*L* allele, which has a higher frequency in populations with an older sea age at maturity (Barson et al. 2015), was associated with an increase in the probability of non-empty stomach in an age-dependent order, with a more pronounced effect in younger age groups (Figure 3a). Allelic substitution from *E* to *L* in the *six6* genomic region (i.e. change in effect size by changing one allele of the genotype) increased the probability of prey occurring in the stomach by 1.56 (1.06 – 2.30, 90% CI, $p = 0.057$) and 1.25 times (1.08 - 1.45, 90% CI, $p < 0.014$) in the 1SW and 2SW groups, respectively (Figure 3a). Both age groups exhibited significant or near significant age-dependent genotype effects relative to the 3SW group ($p = 0.036$ and 0.051, respectively, Figure 3b). Strikingly, the probability of prey in the stomach in relation to size at age (Figure 2e) was in the opposite direction to the *six6*L* effect (Figure 3a) despite the two (*six6* and size at age) exhibiting a significant positive correlation (Supp. Table 3), suggesting the occurrence of complex, contrasting effects of *six6* genetic variation across different phenotypic classes.

The conditional model suggested that the *six6*L* allele was also associated with increased total stomach weight in the young age groups (1SW and 2SW) but not in the 3SW group (Figure 3b). The allelic substitution effect from *E* to *L* was significant and associated with a 1.87-fold (1.29-2.73, 90% CI, $p = 0.006$) increase in prey weight in the 1SW group and was marginal in the 2SW group, associated with a 1.14 (1.00-1.30, 90% CI, $p = 0.099$) increase in prey weight (Figure 3b).

The *vgll3* genomic region was not associated with diet content variation, suggesting no causal link between the two (Figure 3e-h). However, selection on diet may still exert evolutionary change in the *vgll3* genomic region, via correlated response to selection (Lande and Arnold 1983), as a result of phenotypic covariation between diet and age at maturity and length at age. Accordingly, the effect of *vgll3* was significant when these co-varying phenotypes were not accounted for in the model (Supp. Table 4). Spatio-temporal variance in the dataset was substantial in explaining diet variation in both components of the hurdle model (Supp. Table 2, see also Supp. Figure 3). In general, diet presence and quantity were the highest at the westerly end of the distribution, with a gradual decrease towards

the east. At the temporal scale, sampling days in the middle of the sampling period were associated

with a higher presence and quantity of diet in the stomach (Supp. Figure 3).

Population of origin was not a significant source of diet variation and explained only a fraction of the

total variation in diet content (Supp. Table 2, ΔAIC = 3.91, $LRT_{2,2057}$ = 0.09, p = 0.96). When the

analysis was performed with samples assigned to a population of origin with high confidence (N =

1372), the variance due to population was similarly small (see Supp. Table 5 for total stomach weight

as the response variable) and the model was also less parsimonious than a model without the

population effect, as assessed by comparing the model fit by difference in their Akaike information

criterion and likelihood ratio test (ΔAIC = 3.42, $LRT_{2,1340}$ = 0.58, p = 0.75). Likewise, the relation

between total stomach weight and *six6* genetic variation was relatively robust for the full dataset

(N=2057, Supp. table 2) and dataset only including individuals with high population assignment

confidence (N=1372, Supp. table 5).

In our framework, digested, unidentified material in the stomach was not included in the analysis

(e.g., Jacobsen and Hansen 2001). However, the results were qualitatively similar when digested

material was included in the analysis (Supp. Table 6). A model including sex was less parsimonious

and the term was not included as a parameter in the model (ΔAIC = 0.75). Finally, when the fit of the

genetic models (*six6* and *vgll3*) was compared to the putatively neutral SNPs in the panel, *six6* ranked

first out 167, confirming its significance, while *vgll3* was only ranked 123rd (Supp. Figure 4).

In general, both the number of prey items and the increase in the individual prey weight contributed

to the variation in the total stomach weight (Figures 2c-d & 3c-d, Supp. Tables 7 & 8). The 3SW age

group was associated with significantly fewer prey items (0.49 prey items, 0.34-0.71, 90% CI) than

the 1SW (0.99 prey items, 0.65-1.52, 90% CI, $p = 0.004$) and 2SW age groups (1.32 prey items, 1.01-

1.71, 90% CI, $p < 0.001$), but the average prey weight was significantly heavier (27.40 g, 21.24-

35.35, 90% CI) than that in the 1SW (2.68 g, 1.79-4.06, 90% CI, $p < 0.001$) and 2SW (7.83 g, 6.45-

9.53, 90% CI, $p < 0.001$) salmon. The average prey weight, but not the prey number, was also

significantly different between the 2SW and 1SW age groups ($p < 0.001$, Figure 2c-d). Size within

Figure 3: Changes in four stomach content variables for differing sea age classes in relation to allelic substitution in *six6* (a) and *vgll3* (b) genomic regions and associated 90% confidence intervals. Only p-values < 0.1 are given in the figures. Note that estimates in the last three columns (stomach weight, average prey number and weight) are for non-empty stomachs only. P-values above the plots indicates comparative difference in genetic effects between sea age classes, and below indicates the significant genetic effect within age classes (no odds change in relation to allelic substitution). Same odds (odds ratio = 1) indicated by a dashed line.

age group also significantly influenced the number and size of prey. Larger fish within the 3SW age

group had fewer ($p < 0.001$) but heavier ($p < 0.001$) prey items in the diet than smaller fish, and larger

2SW individuals consumed smaller prey items ($p < 0.001$, Figure 2g-h, Supp. Tables 7 & 8).

The number and size of prey items was also explained by the *six6* genotype in an age-dependent

manner, with a more pronounced effect in the relatively young age groups. The *E* to *L* substitution in

six6 was associated with a 1.60-fold (1.08-2.36, 90% CI, $p = 0.048$) increase in prey number in the

1SW age group (Figure 3c). The allelic substitution was also associated with 1.65-fold (1.07-2.56,

90% CI, $p = 0.056$) and 1.22-fold (1.04-1.42, 90% CI, $p = 0.040$) increases in average prey weight in

the 1SW and 2SW age groups, respectively, which was significant compared to that observed in the

3SW age group ($p = 0.018$ and 0.030, respectively, Figure 3d). Genetic variation in *vgll3* was not

significantly associated with average individual prey weight or prey number after controlling for age

at maturity (Figure 3e-h).

Relative prey composition as a function of sea age, size, and genetic variation

Prey composition varied substantially across different age groups, suggesting a change in prey

composition as the fish grow older (Figure 4). In general, older age groups were more likely to prey

on herring and haddock (Figure 4a-b), while younger age groups preyed on capelin and sand eel

(Figure 4c-d). The same pattern was observed within age groups, (e.g. larger fish within an age group

had proportionally more herring and haddock than smaller fish in the same age group) albeit generally

not significantly (Supp. Figure 5), suggesting that size may be a contributing factor explaining prey

composition. In all analyses, spatio-temporal variation was a significant component explaining the

prey composition (Supp. Table 9).

Figure 4: Relative prey composition for the four key prey species, measured as the proportion of each prey species contributing to the stomach content weight, in relation to age at maturity. Note that estimates include only non-empty stomachs.

Genetic variation in *six6* and *vgll3* did not appear to be a strong predictor of prey composition, but

some notable associational trends existed for the two genomic regions (Supp. Figure 6, Supp. Table

9). Particularly, *E* to *L* substitution in *vgll3* is associated with 1.31 times (1.01-1.70, 90% CI, *p* =

0.089) and 1.49 times (1.03-2.16, 90% CI, *p* = 0.076) fewer capelin in the stomach relative to other

prey species in the 2SW and 3SW age groups, respectively (Supp. Figure 6). Additionally, there was

a significant age-dependent preference for capelin over herring associated with the *E* to *L* substitution

in the *six6* genomic region (Supp. Figure 6). When compared to putatively neutral SNPs in the

genotyping panel, capelin composition modelled with *vgll3* ranked 12th out of 164 SNPs (0.073, Supp.

Figure 7), a value that is consistent with the analytically inferred p-value. Finally, when sea age and

size at age were not controlled for, as expected, genetic variation in both the *vgll3* and *six6* genomic

regions explained a substantial portion of the variation also in the relative prey composition (Supp.

Table 10). This suggest the phenotypic covariance between and diet composition and age at maturity

may exert a correlated response to selection at the life history genomic region (i.e. *vgll3*), despite not

being causally linked to diet (i.e. Lande and Arnold 1983).

Discussion

Quantifying resource acquisition via stomach content analysis is an integral component of trophic

studies in ecology and evolution. In this study, we used stomach content information from a single

time point as a proxy for diet acquisition. The analysis of diet content may be difficult for a number

of reasons, such as the challenges in accounting for the diverse nature of diet content data, high

percentage of digested items, and the diversity of metrics for statistical analysis (e.g., Rice 1988,

Cortés 1997, de Crespin de Billy 2000, Baker et al. 2014). On the other hand, the relatively small

number of diet species in our dataset, all fishes, and the small proportion of undigested material

provided us with a robust quantification of the diet and a powerful statistical framework with

ecologically relevant response variables. The three most common prey species in our dataset (e.g.,

herring, capelin and sand eel) were from similar trophic levels independent of size or ontogeny

(Dommasnes et al. 2001, Bentley et al. 2017). In turn stable isotopes, which are sensitive to quantify

diet variation across trophic levels, would have been unable to tease the observed variation apart.

Although stable isotopes may effectively quantify diet variation at other life stages, or shed light on

long-term dietary patterns, stomach content analysis is a more robust approach when diet composition

is variable when considering similar trophic levels.

Our analyses indicate that diet acquisition strategies in the sea vary with sea age in Atlantic salmon

and that this variation is associated with genetic variation in key life history genomic regions,

particularly in the *six6* genomic region. The variation in diet explained by sea age and size at age was

mostly concordant, suggesting that size is the major driver of diet variation, influencing both the

quantity and species composition of prey (Figures 2 & 4). Atlantic salmon prey on heavier but fewer

prey as they grow older and larger, which seems to be a strategy that comes at the expense of a

reduced prey acquisition probability (Figure 2a-d, Huey et al. 2001, Arrington et al. 2002). This

pattern is consistent with the so-called "feast and famine" strategy observed among large piscivorous

fish species (Arrington et al. 2002). The feast and famine feeding strategy is suggested to be an

adaptation to maintain a positive energy balance at a large body size, especially when the acquisition

of energy-rich food sources is unpredictable (Armstrong and Schindler 2011). Large Atlantic salmon

appear to adopt this strategy, which is likely beneficial in terms of balancing the increase in energy

costs associated with a large body size. A suite of physiological adaptations and metabolic

adjustments, such as increased digestion capacity (Armstrong and Schindler 2011) and fat storage

(Bustard 1967), may be associated with this strategy (Wang et al. 2006). For example, it has been

shown that piscivorous species that adopt a feast and famine strategy maintain a large digestive tract,

which provides quick food utilization when abundant prey are encountered (Armstrong and Schindler

2011). This physiological trade-off seems to be evolutionarily favourable for large fish when the prey

distribution is stochastic despite the energetic costs of sustaining excess and energetically expensive

digestive tissue (Armstrong and Schindler 2011). The feast and famine strategy in large Atlantic

salmon may also be facilitated by other mechanistic processes, such as the trade-off of a lower success

rate linked to larger prey or a lower attack rate associated with increasing size. Nonetheless, variation

in foraging strategies among different age groups results in a large diet breadth, efficient resource

partitioning, and reduced intraspecific competition among age groups, which subsequently promotes

their co-existence (e.g., Polis 1984, Smith and Skulason 1996, Svanback and Bolnick 2007). It is

unclear what physiological or behavioural modifications are associated with the differential feeding

strategies among the various age groups in Atlantic salmon. Nonetheless, changes in marine food

webs may alter the density and composition of prey available to different age groups and hence alter

the age-dependent selection landscape, potentially leading to adaptive changes in age structure. Our

results confirm the value of Atlantic salmon as a model species to study the evolutionary physiology

of starvation and feeding in response to environmental changes in the wild.

Salmon are generally considered to be diet generalists. However, the association between the life

history genomic region *six6* and diet acquisition (as well as the suggestive acquisition with capelin

prey preference and *vgll3*, see, Supp. Table 6) indicates that genetically controlled intraspecific diet

specialization occurs in Atlantic salmon. This genetic variation may be linked to specialized dietary

adaptations (e.g., physiological, morphological or behavioural) allowing the efficient utilization of

diverse diet sources, resulting in increased niche breadth and reduced intraspecific competition

(Dalmo et al. 1997, Bolnick et al. 2003, Bolnick and Fitzpatrick 2007, Svanback and Bolnick 2007).

In particular, the *six6*L* allele was associated with increased content in the stomach, especially in

young age groups, which was explained by both an increase in the number of prey items and an

increase in the average prey weight in the stomach. Intriguingly, however, the *six6*L* allele was

associated with a larger fish size within all sea ages (Supp. Table 3, see also, Barson et al. 2015).

Given that larger size at a given sea age is indicative of higher performance and fitness in adult salmon

(e.g., Fleming 1996; Mobley et al. 2020), the *six6*L* allele (being linked to larger size) may be

expected to have a selective advantage over the *E* allele and is hence predicted to prevail within and

among populations as a result of directional selection. However, the *six6* genomic region is highly

variable within and among populations (Barson et al. 2015, Pritchard et al. 2018), and balancing

selection appears to be the pervasive mode of evolution, with both alleles exerting fitness advantage

with different life-history strategies (Barson et al. 2015). The results of this study suggest that the

genetic variation in *six6* locus could be explained by antagonistic pleiotropy along the life cycle of

an individual (i.e., fitness trade-offs within the lifetime of an individual associated with the *six6*

region) or fluctuating selection across generations. For example, genetic variation in *six6* may be

under balancing selection via antagonistic genetic correlations in diet acquisition over the life cycle.

Indeed, some results indicate that individuals having high freshwater growth may show poor seawater

growth (Einum et al. 2002), prescribing further hypothesis linking it to genetic variation in *six6*

genomic region.

Although our results suggest superior performance of the *six6*L* allele at the adult stage, the opposite

may be true at earlier life stages, when the prey species composition is substantially different, with

significantly more invertebrates in the diet (Jacobsen and Hansen 2001, Rikardsen et al. 2004,

Haugland et al. 2006), and different ecological drivers affect performance (Mittelbach et al. 2014,

Sanchez-Hernandez et al. 2019). For example, if increased metabolic costs are linked with increased

prey content in the stomach, a trait associated with *six6*L*, this may not be an optimal acquisition

strategy in younger years (i.e. juveniles prior to, or in the early phases of marine migration) when the

energy density of available prey cannot compensate for the greater effort (McNamara and Houston

1996, Enberg et al. 2012). Such antagonistic genetic correlations between early and late life histories

(i.e. genotype-environment interactions) in diet acquisition may maintain polymorphism in the

region. Alternatively, year-to-year variation in population-specific (e.g., density dependent) or

ecosystem-level processes (i.e., prey composition, food-web dynamics) may alter the adaptive

landscape of diet acquisition (Smith and Skulason 1996), resulting in fluctuating selection and a

change in the direction of selection among alleles, which would maintain the genetic variation in the

region. Indeed, in the year 2008, returning salmon had a notably older sea age at maturity in northern

Norway than that observed in more recent years (i.e., the oldest in the last 20 years, see (Anon 2014).

This is consistent with observed patterns that *six6*L* is linked with larger size, perhaps as a result of

providing a selective advantage for that particular year class. However, more research is required to

explicitly test this possibility. The *six6* genomic region is highly spatially differentiated among

populations (Barson et al. 2015, Pritchard et al. 2018). This suggests that any selection acting on *six6*

as a result of selection on diet acquisition would influence the fitness of populations differentially,

correlated with their average allele frequency. Hence, genetic variation in *six6* may be linked to the

differential survival of populations at sea and should be closely monitored in population management.

The genetic variation in the v*gll3* genomic region had no clear effect on diet quantity overall, but

there was a marginal ($p = 0.073$), albeit consistent, association with higher capelin content in the

stomach (Supp. Figures 6 & 7). Capelin is a key component in the Barents Sea ecosystem with critical

bottom-up effect (Gjøsæter et al. 2016), and both salmon post-smolts and returning adults utilize them

as a food resource (Rikardsen and Dempson 2010, this study) when they migrate through the coastal

regions (Gjøsæter et al. 2016). Individuals with the *vgll3*L* allele, which is associated with a later

sea-age at maturity (Barson et al. 2015), were marginally less likely to feed on capelin, particularly

in older age groups (2SW and 3SW). In addition, in contrast to herring, which was the most common

prey item in the stomachs of the individuals in the 2SW and 3SW age groups (Figure 4c), capelin is

likely a lower-energy prey item for Atlantic salmon (Elliott and Gaston 2008, Hedeholm et al. 2011,

Renkawitz et al. 2015). This supports the notion that *vgll3*L* may contribute to foraging adaptations

to support the high-energy demands associated with the larger body size of late maturing fish.

However, the mechanisms driving such a compositional difference have yet to be determined.

Overall, genetic variation in both life history genomic regions appears to have a role in intraspecific

diet specialization, but the mechanisms remain to be clarified. Although the underlying mechanism

of diet specialization is complex (e.g., Mittelbach et al. 2014) and challenging to disentangle,

performance trade-offs across different ecological settings and the life cycle are likely driving the

life-history variation associated with these genomic regions. Analyses of datasets collected in the

wild may suffer from confounding effects that co-vary with both the response variable and parameters

of interest and generate spurious associations if they are not accounted for. In this study, a highly

controlled linear model was employed to account for environmental and intrinsic parameters with

potential confounding effects. First, resource availability at sea, e.g., the prey species density and

distribution, is highly variable across time and space, even at small scales. Likewise, during their

return migration, Atlantic salmon may be non-randomly distributed in relation to their life history,

genotype, and population of origin (Svenning et al. 2019). For example, relatively early run timing is

linked to both later maturation (Jonsson and Jonsson 2011) and *six6*L* (Cauwelier et al. 2018,

Pritchard et al. 2018), a pattern that concordantly holds in our dataset (Supp. Table 11). Hence, the

analytical framework controlled for spatio-temporal variation and also accounted for non-linear

changes through space and time (Supp. Figure 3). Similarly, sea age at maturity and size within an

age group exhibited strong links with both diet and genetic variation in the life history genomic

regions. By accounting for these phenotypes in the model, we were able to exclude the possibility of

modelling the genetic variation via the effects of these intermediate phenotypes. Therefore, our

framework was rather robust to drawbacks related to confounding factors observed in wild settings.

Finally, variation in diet due to the population of origin was also accounted for in the model as a

random intercept but did not explain significant diet variation at sea (Supp. Tables 2 & 5).

On the other hand, the significance between diet variation and life history genomic regions were often

only marginally significant. Therefore, further research is warranted to validate these results. Further,

genetic variation outside these two major maturation-timing loci and potential pleiotropic effects

remain unexplained, which could be explored with a genome wide approach in future studies.

Likewise, similar set-ups at different life history stages, under different feeding regimes, or in

common garden conditions, would help to elucidate the effect of genotype-environment interactions

that may have been overlooked in this dataset.

Marine ecosystems, which are composed of mostly poikilothermic species, are sensitive and highly

responsive to temperature-driven changes (e.g., Clarke 2003, Sydeman et al. 2015). The Arctic region

is particularly sensitive to global climate change (Polyakov et al. 2010), with significant

anthropogenic effects further shaping the marine food webs in the region. Over the last 40 years, the

abundance of Atlantic salmon has been declining, and the age structure has been shifting towards a

younger age at maturity (Chaput 2012, Czorlich et al. 2018, Erkinaro et al. 2018). Such changes in

demography are likely the result of bottom-up changes in prey community structure, likely fuelled by

climate-induced changes to the ecosystem (Frederiksen et al. 2006, Todd et al. 2008).In this study,

we demonstrated that the inter-individual variation in diet specialization is linked with age structure

as well as the genetic variation in *six6* and *vgll3*, two genomic regions with substantial influence on

life history variation and population divergence. This heritable intraspecific variation in diet

specialization likely plays an important role in salmon life history by both promoting the niche

breadth of species and enabling evolutionary responses in populations to changes in food

composition. Given that both genomic regions are highly differentiated among populations,

evolutionary response and the resulting demographic trajectories likely differ among populations,

concordantly. Future work should focus on characterizing the underlying physiological and/or

behavioural mechanisms linking genetic variation with salmon diet acquisition to better predict the

evolutionary response of populations to changing environments.

Data Availability Statement

576 Data will be available to public in Dryad depository upon acceptance.

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References

Anon. 2014. Status of the River Tana salmon populations. Report of the working group on salmon monitoring and research in the Tana River system.

Armstrong, J. B., and D. E. Schindler. 2011. Excess digestive capacity in predators reflects a life of feast and famine. Nature **476**:84-87. doi: 10.1038/nature10240.

Arrington, D. A., K. O. Winemiller, W. F. Loftus, and S. Akin. 2002. How often do fishes "run on empty"? Ecology **83**:2145-2151. doi: 10.2307/3072046.

Aykanat, T., M. Lindqvist, V. L. Pritchard, and C. R. Primmer. 2016. From population genomics to conservation and management: a workflow for targeted analysis of markers identified using genome-wide approaches in Atlantic salmon Salmo salar. Journal of Fish Biology **89**:2658-2679. doi: 10.1111/jfb.13149.

Aykanat, T., M. Ozerov, J. P. Vaha, P. Orell, E. Niemela, J. Erkinaro, and C. R. Primmer. 2019. Co-inheritance of sea age at maturity and iteroparity in the Atlantic salmon vgll3 genomic region. Journal of Evolutionary Biology. doi: 10.1111/jeb.13418.

Ayllon, F., E. Kjaerner-Semb, T. Furmanek, V. Wennevik, M. F. Solberg, G. Dahle, G. L. Taranger, K. A. Glover, M. S. Almen, C. J. Rubin, R. B. Edvardsen, and A. Wargelius. 2015. The vgll3 Locus Controls Age at Maturity in Wild and Domesticated Atlantic Salmon (Salmo salar L.) Males. PLoS Genetics **11**:e1005628. doi: 10.1371/journal.pgen.1005628.

Baker, R., A. Buckland, and M. Sheaves. 2014. Fish gut content analysis: robust measures of diet composition. Fish and Fisheries **15**:170-177. doi: 10.1111/faf.12026.

Barson, N. J., T. Aykanat, K. Hindar, M. Baranski, G. H. Bolstad, P. Fiske, C. Jacq, A. J. Jensen, S. E. Johnston, S. Karlsson, M. Kent, T. Moen, E. Niemela, T. Nome, T. F. Naesje, P. Orell, A. Romakkaniemi, H. Saegrov, K. Urdal, J. Erkinaro, S. Lien, and C. R. Primmer. 2015. Sex-dependent dominance at a single locus maintains variation in age at maturity in salmon. Nature **528**:405-408. doi: 10.1038/nature16062.

Bentley, J. W., N. Serpetti, and J. J. Heymans. 2017. Investigating the potential impacts of ocean warming on the Norwegian and Barents Seas ecosystem using a time-dynamic food-web model. Ecological Modelling **360**:94-107. doi: 10.1016/j.ecolmodel.2017.07.002.

Bessarab, D. A., S. W. Chong, and V. Korzh. 2004. Expression of zebrafish six1 during sensory organ development and myogenesis. Developmental Dynamics **230**:781-786. doi: 10.1002/dvdy.20093.

Bolnick, D. I., P. Amarasekare, M. S. Araujo, R. Burger, J. M. Levine, M. Novak, V. H. Rudolf, S. J. Schreiber, M. C. Urban, and D. A. Vasseur. 2011. Why intraspecific trait variation matters in community ecology. Trends in Ecology & Evolution **26**:183-192. doi: 10.1016/j.tree.2011.01.009.

Bolnick, D. I., and M. S. Araujo. 2011. Partitioning the relative fitness effects of diet and trophic morphology in the threespine stickleback. Evolutionary Ecology Research **13**:439-459.

Bolnick, D. I., and B. M. Fitzpatrick. 2007. Sympatric Speciation: Models and Empirical Evidence. Annual Review of Ecology, Evolution, and Systematics **38**:459-487. doi: 10.1146/annurev.ecolsys.38.091206.095804.

Bolnick, D. I., R. Svanback, J. A. Fordyce, L. H. Yang, J. M. Davis, C. D. Hulsey, and M. L. Forister. 2003. The ecology of individuals: incidence and implications of individual specialization. American Naturalist **161**:1-28. doi: 10.1086/343878.

Bradbury, I. R., L. C. Hamilton, S. Rafferty, D. Meerburg, R. Poole, J. B. Dempson, M.J. Robertson, et al. 2015. Genetic evidence of local exploitation of Atlantic salmon in a coastal subsistence fishery in the Northwest Atlantic. Canadian Journal of Fisheries and Aquatic Sciences, **72**: 83–95. doi: 10.1139/cjfas-2014-0058

Brooks, M. E., K. Kristensen, K. J. van Benthem, A. Magnusson, C. W. Berg, A. Nielsen, H. J. Skaug, M. Machler, and B. M. Bolker. 2017. glmmTMB Balances Speed and Flexibility Among Packages for Zero-inflated Generalized Linear Mixed Modeling. R Journal **9**:378-400. doi: 10.32614/Rj-2017-066.

Bustard, H. R. 1967. Gekkonid lizards adapt fat storage to desert environments. Science **158**:1197-1198.

Carter, A. J., A. W. Goldizen, and S. A. Tromp. 2010. Agamas exhibit behavioral syndromes: bolder males bask and feed more but may suffer higher predation. Behavioral Ecology **21**:655-661. doi: 10.1093/beheco/arq036.

Cauwelier, E., J. Gilbey, J. Sampayo, L. Stradmeyer, and S. J. Middlemas. 2018. Identification of a single genomic region associated with seasonal river return timing in adult Scottish Atlantic salmon (Salmo salar), using a genome-wide association study. Canadian Journal of Fisheries and Aquatic Sciences **75**:1427-1435. doi: 10.1139/cjfas-2017-0293.

Chaput, G. 2012. Overview of the status of Atlantic salmon (Salmo salar) in the North Atlantic and trends in marine mortality. ICES Journal of Marine Science **69**:1538-1548. doi: 10.1093/icesjms/fss013.

Claessen, D., and U. Dieckmann. 2002. Ontogenetic niche shifts and evolutionary branching in size-structured populations. Evolutionary Ecology Research **4**:189-217.

Clarke, A. 2003. Costs and consequences of evolutionary temperature adaptation. Trends in Ecology & Evolution **18**:573-581. doi: 10.1016/j.tree.2003.08.007.

Cortés, E. 1997. A critical review of methods of studying fish feeding based on analysis of stomach contents: application to elasmobranch fishes. Canadian Journal of Fisheries and Aquatic Sciences **54**:726-738. doi: 10.1139/f96-316.

de Crespin de Billy, V. 2000. Biplot presentation of diet composition data: an alternative for fish stomach contents analysis. Journal of Fish Biology **56**:961-973. doi: 10.1111/j.1095-8649.2000.tb00885.x.Czorlich, Y., T. Aykanat, J. Erkinaro, P. Orell, and C. R. Primmer. 2018. Rapid sex-specific evolution of age at maturity is shaped by genetic architecture in Atlantic salmon. Nat Ecol Evol **2**:1800-1807. doi: 10.1038/s41559-018-0681-5.

Dalmo, R. A., K. Ingebrigtsen, and J. Bogwald. 1997. Non-specific defence mechanisms in fish, with particular reference to the reticuloendothelial system (RES). Journal of Fish Diseases **20**:241-273.

Daufresne, M., K. Lengfellner, and U. Sommer. 2009. Global warming benefits the small in aquatic ecosystems. Proc Natl Acad Sci U S A **106**:12788-12793. doi: 10.1073/pnas.0902080106.

Debes, P.V., N. Piavchenko, A. Ruokolainen, O. Ovaskainen, J.E. Moustakas-Verho, N. Parre, T. Aykanat, J. Erkinaro, C.R. Primmer. 2019. Large single-locus effects for maturation timing are mediated via condition variation in Atlantic salmon. bioRxiv, 780437

Devictor, V., J. Clavel, R. Julliard, S. Lavergne, D. Mouillot, W. Thuiller, P. Venail, S. Villéger, and N. Mouquet. 2010. Defining and measuring ecological specialization. Journal of Applied Ecology **47**:15-25. doi: 10.1111/j.1365-2664.2009.01744.x.

Dommasnes, A., V. Christensen, B. Ellertsen, C. Kvamme, W. Melle, L. Nøttestad, P. Torstein, S. Tjelmeland, and D. Zeller. 2001. An Ecopath model for the Norwegian and Barents Sea. Fisheries impacts on North Atlantic ecosystems: models and analyses. University of British Columbia Fisheries Center Research Report, **9**(4): 213e240

Einum, S., E. B. Thorstad, and T. F. Naesje. 2002. Growth rate correlations across life-stages in female Atlantic salmon. Journal of Fish Biology **60**:780-784. doi: PII 10.1006/jfbi.2002.1895.

Elliott, K. H., and A. J. Gaston. 2008. Mass-length relationships and energy content of fishes and invertebrates delivered to nestling Thick-billed Murres Uria lomvia in the Canadian Arctic, 1981–2007. Marine Ornithology **36**:25–34.

Elmer, K. R., T. K. Lehtonen, A. F. Kautt, C. Harrod, and A. Meyer. 2010. Rapid sympatric ecological
differentiation of crater lake cichlid fishes within historic times. BMC Biology **8**:60. doi:
10.1186/1741-7007-8-60.
Elphinstone, M. S., G. N. Hinten, M. J. Anderson, and C. J. Nock. 2003. An inexpensive and high-
throughput procedure to extract and purify total genomic DNA for population studies.
Molecular Ecology Notes **3**:317-320. doi: 10.1046/j.1471-8286.2003.00397.x.
Enberg, K., C. Jørgensen, E. S. Dunlop, Ø. Varpe, D. S. Boukal, L. Baulier, S. Eliassen, and M.
Heino. 2012. Fishing-induced evolution of growth: concepts, mechanisms and the empirical
evidence. Marine Ecology **33**:1-25. doi: 10.1111/j.1439-0485.2011.00460.x.
Endelman, J. B. 2011. Ridge Regression and Other Kernels for Genomic Selection with R Package
rrBLUP. Plant Genome **4**:250-255. doi: 10.3835/plantgenome2011.08.0024.
Engen, S., and N. C. Stenseth. 1989. Age-specific optimal diets and optimal foraging tactics: a life-
historic approach. Theoretical Population Biology **36**:281-295.
Erkinaro, J., Y. Czorlich, P. Orell, J. Kuusela, M. Falkegård, M. Länsman, H. Pulkkinen, C. R.
Primmer, and E. Niemelä. 2018. Life history variation across four decades in a diverse
population complex of Atlantic salmon in a large subarctic river. Canadian Journal of
Fisheries and Aquatic Sciences. doi: 10.1139/cjfas-2017-0343.
Erkinaro, J., J. B. Dempson, M. Julkunen, and E. Niemelä. 1997. Importance of ontogenetic habitat
shifts to juvenile output and life history of Atlantic salmon in a large subarctic river: an
approach based on analysis of scale characteristics. Journal of Fish Biology **51**:1174-1185.
doi: 10.1111/j.1095-8649.1997.tb01134.x.
Fleming, I. A. 1996. Reproductive strategies of Atlantic salmon: ecology and evolution. Reviews in
Fish Biology and Fisheries **6**:379-416. doi: 10.1007/bf00164323.
Fox, L. R., and P. A. Morrow. 1981. Specialization: species property or local phenomenon? Science
211:887-893. doi: 10.1126/science.211.4485.887.
Frederiksen, M., M. Edwards, A. J. Richardson, N. C. Halliday, and S. Wanless. 2006. From plankton
to top predators: bottom-up control of a marine food web across four trophic levels. Journal
of Animal Ecology **75**:1259-1268. doi: 10.1111/j.1365-2656.2006.01148.x.
Friedland, K. D., and R. E. Haas. 1996. Marine post-smolt growth and age at maturity of Atlantic
salmon. Journal of Fish Biology **48**:1-15. doi: 10.1111/j.1095-8649.1996.tb01414.x.
Gallardo, M. E., J. Lopez-Rios, I. Fernaud-Espinosa, B. Granadino, R. Sanz, C. Ramos, C. Ayuso,
M. J. Seller, H. G. Brunner, P. Bovolenta, and S. Rodriguez de Cordoba. 1999. Genomic
cloning and characterization of the human homeobox gene SIX6 reveals a cluster of SIX genes
in chromosome 14 and associates SIX6 hemizygosity with bilateral anophthalmia and
pituitary anomalies. Genomics **61**:82-91. doi: 10.1006/geno.1999.5916.
Gjøsæter, H., E. H. Hallfredsson, N. Mikkelsen, B. Bogstad, and T. Pedersen. 2016. Predation on
early life stages is decisive for year-class strength in the Barents Sea capelin (Mallotus
villosus) stock. ICES Journal of Marine Science: Journal du Conseil **73**:182-195. doi:
10.1093/icesjms/fsv177.
Gotthard, K. 2000. Increased risk of predation as a cost of high growth rate: an experimental test in a
butterfly. Journal of Animal Ecology **69**:896-902. doi: 10.1046/j.1365-2656.2000.00432.x.
Hadfield, J. D. 2010. MCMC Methods for Multi-Response Generalized Linear Mixed Models: The
MCMCglmm R Package. Journal of Statistical Software **33**:1-22.
Halperin, D. S., C. Pan, A. J. Lusis, and P. Tontonoz. 2013. Vestigial-like 3 is an inhibitor of adipocyte
differentiation. Journal of Lipid Research **54**:473-481. doi: 10.1194/jlr.M032755.
Haugland, M., J. Holst, M. Holm, and L. Hansen. 2006. Feeding of Atlantic salmon (Salmo salar L.)
post-smolts in the Northeast Atlantic. ICES Journal of Marine Science **63**:1488-1500. doi:
10.1016/j.icesjms.2006.06.004.

Hedeholm, R., P. Grønkjær, and S. Rysgaard. 2011. Energy content and fecundity of capelin (Mallotus villosus) along a 1,500-km latitudinal gradient. Marine Biology **158**:1319-1330. doi: 10.1007/s00227-011-1651-5.

Härkönen, T. (1986). Guide to the otoliths of the bony fishes of the Northeast Atlantic. Hellerup, Danbiu ApS.

Heino, M., and V. Kaitala. 1999. Evolution of resource allocation between growth and reproduction in animals with indeterminate growth. Journal of Evolutionary Biology **12**:423-429.

Huey, R. B., E. R. Pianka, and L. J. Vitt. 2001. How often do lizards "run on empty"? Ecology **82**:1-7.

Hvidsten, N. A., A. J. Jensen, A. H. Rikardsen, B. Finstad, J. Aure, S. Stefansson, P. Fiske, and B. O. Johnsen. 2009. Influence of sea temperature and initial marine feeding on survival of Atlantic salmon Salmo salar post-smolts from the Rivers Orkla and Hals, Norway. Journal of Fish Biology **74**:1532-1548. doi: 10.1111/j.1095-8649.2009.02219.x.

ICES. 2011. Report of the Workshop on Age Determination of Salmon (WKADS). Galway, Ireland.

Jacobsen, J. A., and L. P. Hansen. 2001. Feeding habits of wild and escaped farmed Atlantic salmon, Salmo salar L., in the Northeast Atlantic. ICES Journal of Marine Science **58**:916-933. doi: 10.1006/jmsc.2001.1084.

Jensen, H., M. Kiljunen, and P. A. Amundsen. 2012. Dietary ontogeny and niche shift to piscivory in lacustrine brown trout Salmo trutta revealed by stomach content and stable isotope analyses. Journal of Fish Biology **80**:2448-2462. doi: 10.1111/j.1095-8649.2012.03294.x.

Jonsson, B., and N. Jonsson. 2011. Ecology of Atlantic salmon and brown trout : habitat as a template for life histories. Springer, Dordrecht.

Jonsson, N., B. Jonsson, and L. P. Hansen. 1997. Changes in proximate composition and estimates of energetic costs during upstream migration and spawning in Atlantic salmon Salmo salar. Journal of Animal Ecology **66**:425-436. doi: 10.2307/5987.

Lande, R., and Arnold, S. J. 1983. The Measurement of Selection on Correlated Characters. Evolution 37: 1210-1226.

Lepais, O., A. Manicki, S. Glise, M. Buoro, and A. Bardonnet. 2017. Genetic architecture of threshold reaction norms for male alternative reproductive tactics in Atlantic salmon (Salmo salar L.). Sci Rep **7**:43552. doi: 10.1038/srep43552.

Lin, X., X. Duan, Y. Y. Liang, Y. Su, K. H. Wrighton, J. Long, M. Hu, C. M. Davis, J. Wang, F. C. Brunicardi, Y. Shi, Y. G. Chen, A. Meng, and X. H. Feng. 2006. PPM1A functions as a Smad phosphatase to terminate TGFbeta signaling. Cell **125**:915-928. doi: 10.1016/j.cell.2006.03.044.

Machovsky-Capuska, G. E., A. M. Senior, S. J. Simpson, and D. Raubenheimer. 2016. The Multidimensional Nutritional Niche. Trends in Ecology & Evolution **31**:355-365. doi: 10.1016/j.tree.2016.02.009.

MacKenzie, K. M., C. N. Trueman, M. R. Palmer, A. Moore, A. T. Ibbotson, W. R. C. Beaumont, and I. C. Davidson. 2012. Stable isotopes reveal age-dependent trophic level and spatial segregation during adult marine feeding in populations of salmon. ICES Journal of Marine Science **69**:1637-1645. doi: 10.1093/icesjms/fss074.

McNamara, J. M., and A. I. Houston. 1996. State-dependent life histories. Nature **380**:215-221. doi: 10.1038/380215a0.

Mittelbach, G. G., N. G. Ballew, M. K. Kjelvik, and D. Fraser. 2014. Fish behavioral types and their ecological consequences. Canadian Journal of Fisheries and Aquatic Sciences **71**:927-944. doi: 10.1139/cjfas-2013-0558.

Mittelbach, G. G., and L. Persson. 1998. The ontogeny of piscivory and its ecological consequences. Canadian Journal of Fisheries and Aquatic Sciences **55**:1454-1465. doi: 10.1139/f98-041.

Mobley, K. B., H. Granroth-Wilding, M. Ellmén, P. Orell, J. Erkinaro, and C.R. Primmer. 2020. Time spent in distinct life history stages has sex-specific effects on reproductive fitness in wild Atlantic salmon. Molecular Ecology, mec.15390. doi: 10.1111/mec.15390

Mousseau, T. A., B. Sinervo, and J. A. Endler. 2000. Adaptive genetic variation in the wild. Oxford University Press, New York.

Nakayama, K., J. Ohashi, K. Watanabe, L. Munkhtulga, and S. Iwamoto. 2017. Evidence for Very Recent Positive Selection in Mongolians. Molecular Biology and Evolution. doi: 10.1093/molbev/msx138.

Neaves, P. I., C. G. Wallace, J. R. Candy, and T. D. Beacham. 2005. CBayes: computer program for mixed-stock analysis of allelic data, version v5.01.

O'Brien, J. H., L. Hernandez-Lagunas, K. B. Artinger, and H. L. Ford. 2014. MicroRNA-30a regulates zebrafish myogenesis through targeting the transcription factor Six1. Journal of Cell Science **127**:2291-2301. doi: 10.1242/jcs.143677.

Ozerov, M., J.-P. Vähä, V. Wennevik, E. Niemelä, M.-A. Svenning, S. Prusov, R. Diaz Fernandez, L. Unneland, A. Vasemägi, M. Falkegård, T. Kalske, and B. Christiansen. 2017. Comprehensive microsatellite baseline for genetic stock identification of Atlantic salmon (Salmo salar L.) in northernmost Europe. ICES Journal of Marine Science. doi: 10.1093/icesjms/fsx041.

Pella, J., and M. Masuda. 2001. Bayesian methods for analysis of stock mixtures from genetic characters. Fishery Bulletin **99**:151-167.

Pershing, A. J., M. A. Alexander, C. M. Hernandez, L. A. Kerr, A. Le Bris, K. E. Mills, J. A. Nye, N. R. Record, H. A. Scannell, J. D. Scott, G. D. Sherwood, and A. C. Thomas. 2015. Slow adaptation in the face of rapid warming leads to collapse of the Gulf of Maine cod fishery. Science **350**:809-812. doi: 10.1126/science.aac9819.

Pethon, P. and B. O. Nyström (2005). Aschehougs store fiskebok : Norges fisker i farger. [Oslo], Aschehoug.

Polis, G. A. 1984. Age Structure Component of Niche Width and Intraspecific Resource Partitioning - Can Age-Groups Function as Ecological Species. American Naturalist **123**:541-564. doi: 10.1086/284221.

Polyakov, I. V., L. A. Timokhov, V. A. Alexeev, S. Bacon, I. A. Dmitrenko, L. Fortier, I. E. Frolov, J.-C. Gascard, E. Hansen, V. V. Ivanov, S. Laxon, C. Mauritzen, D. Perovich, K. Shimada, H. L. Simmons, V. T. Sokolov, M. Steele, and J. Toole. 2010. Arctic Ocean Warming Contributes to Reduced Polar Ice Cap. Journal of Physical Oceanography **40**:2743-2756. doi: 10.1175/2010jpo4339.1.

Pritchard, V. L., H. Makinen, J. P. Vaha, J. Erkinaro, P. Orell, and C. R. Primmer. 2018. Genomic signatures of fine-scale local selection in Atlantic salmon suggest involvement of sexual maturation, energy homeostasis and immune defence-related genes. Molecular Ecology **27**:2560-2575. doi: 10.1111/mec.14705.

Renkawitz, M. D., T. F. Sheehan, H. J. Dixon, and R. Nygaard. 2015. Changing trophic structure and energy dynamics in the Northwest Atlantic: implications for Atlantic salmon feeding at West Greenland. Marine Ecology Progress Series **538**:197-211. doi: 10.3354/meps11470.

Reznick, D. 2016. Hard and Soft Selection Revisited: How Evolution by Natural Selection Works in the Real World. Journal of Heredity **107**:3-14. doi: 10.1093/jhered/esv076.

Reznick, D., and J. A. Endler. 1982. The Impact of Predation on Life History Evolution in Trinidadian Guppies (Poecilia Reticulata). Evolution **36**:160-177. doi: 10.1111/j.1558-5646.1982.tb05021.x.

Reznick, D. N., and C. K. Ghalambor. 2001. The population ecology of contemporary adaptations: what empirical studies reveal about the conditions that promote adaptive evolution. Genetica **112**:183-198.

Rice, J. C. 1988. Repeated cluster analysis of stomach contents data: method and application to diet of cod in NAFO division 3L. Environmental Biology of Fishes **21**:263-277. doi: 10.1007/bf00000375.

Ridgeway, A. G., and I. S. Skerjanc. 2001. Pax3 is essential for skeletal myogenesis and the expression of Six1 and Eya2. Journal of Biological Chemistry **276**:19033-19039. doi: 10.1074/jbc.M011491200.

Rikardsen, A. H., and J. B. Dempson. 2010. Dietary Life-Support: The Food and Feeding of Atlantic Salmon at Sea. Pages 115-143 Atlantic Salmon Ecology. Wiley-Blackwell.

Rikardsen, A. H., M. Haugland, P. A. Bjorn, B. Finstad, R. Knudsen, J. B. Dempson, J. C. Holst, N. A. Hvidsten, and M. Holm. 2004. Geographical differences in marine feeding of Atlantic salmon post-smolts in Norwegian fjords. Journal of Fish Biology **64**:1655-1679. doi: 10.1111/j.1095-8649.2004.00425.x.

Sanchez-Hernandez, J., A. D. Nunn, C. E. Adams, and P. A. Amundsen. 2019. Causes and consequences of ontogenetic dietary shifts: a global synthesis using fish models. Biological Reviews of the Cambridge Philosophical Society **94**:539-554. doi: 10.1111/brv.12468.

Serikaku, M. A., and J. E. Otousa. 1994. Sine Oculis Is a Homeobox Gene Required for Drosophila Visual-System Development. Genetics **138**:1137-1150.

Sexton, J. P., J. Montiel, J. E. Shay, M. R. Stephens, and R. A. Slatyer. 2017. Evolution of Ecological Niche Breadth. Annual Review of Ecology, Evolution, and Systematics **48**:183-206. doi: 10.1146/annurev-ecolsys-110316-023003.

Smith, T. B., and S. Skulason. 1996. Evolutionary significance of resource polymorphisms in fishes, amphibians, and birds. Annual Review of Ecology and Systematics **27**:111-133. doi: 10.1146/annurev.ecolsys.27.1.111.

Somervuo, P., P. Koskinen, P. Mei, L. Holm, P. Auvinen, and L. Paulin. 2018. BARCOSEL: a tool for selecting an optimal barcode set for high-throughput sequencing. BMC Bioinformatics **19**:257. doi: 10.1186/s12859-018-2262-7.

Svanback, R., and D. I. Bolnick. 2007. Intraspecific competition drives increased resource use diversity within a natural population. Proc Biol Sci **274**:839-844. doi: 10.1098/rspb.2006.0198.

Svanback, R., and L. Persson. 2004. Individual diet specialization, niche width and population dynamics: implications for trophic polymorphisms. Journal of Animal Ecology **73**:973-982. doi: 10.1111/j.0021-8790.2004.00868.x.

Svenning, M.-A., M. Falkegård, E. Niemelä, J.-P. Vähä, V. Wennevik, M. Ozerov, S. Prusov, J. B. Dempson, M. Power, P. Fauchald, and D. Gomez-Uchida. 2019. Coastal migration patterns of the four largest Barents Sea Atlantic salmon stocks inferred using genetic stock identification methods. ICES Journal of Marine Science. doi: 10.1093/icesjms/fsz114.

Sydeman, W. J., E. Poloczanska, T. E. Reed, and S. A. Thompson. 2015. Climate change and marine vertebrates. Science **350**:772-777. doi: 10.1126/science.aac9874.

Taranger, G. L., M. Carrillo, R. W. Schulz, P. Fontaine, S. Zanuy, A. Felip, F. A. Weltzien, S. Dufour, O. Karlsen, B. Norberg, E. Andersson, and T. Hansen. 2010. Control of puberty in farmed fish. General and Comparative Endocrinology **165**:483-515. doi: 10.1016/j.ygcen.2009.05.004.

Thorpe, J. E., M. Mangel, N. B. Metcalfe, and F. A. Huntingford. 1998. Modelling the proximate basis of salmonid life-history variation, with application to Atlantic salmon, Salmo salar L. Evolutionary Ecology **12**:581-599. doi: 10.1023/A:1022351814644.

Todd, C. D., S. L. Hughes, C. T. Marshall, J. C. MacLean, M. E. Lonergan, and E. M. Biuw. 2008. Detrimental effects of recent ocean surface warming on growth condition of Atlantic salmon. Global Change Biology **14**:958-970. doi: 10.1111/j.1365-2486.2007.01522.x.

Toy, J., J. M. Yang, G. S. Leppert, and O. H. Sundin. 1998. The Optx2 homeobox gene is expressed
in early precursors of the eye and activates retina-specific genes. Proc Natl Acad Sci U S A
95:10643-10648. doi: 10.1073/pnas.95.18.10643.

Vähä, J.-P., J. Erkinaro, E. Niemelä, C. Primmer, I. Saloniemi, M. Falkegård, M.A. Svenning, and S.
Brørs. 2010. Temporally stable population-specific differences in run timing of one-sea-
winter Atlantic salmon returning to a large river system: Run timing of Atlantic salmon within
a river system. Evolutionary Applications **4**(1):39-53. 10.1111/j.1752-4571.2010.00131.x.

Vähä, J.-P., V. Wennevik, M. Ozerov, R. Diaz-Fernandez, L. Unneland, K. Haapanen, E. Niemelä,
M. Svenning, M. Falkegård, S. Prusov, I. Lyzhov, K. Rysakova, T. Kalske, B. Christiansen,
and G. Ustyuzhinsky. 2014. Genetic structure of Atlantic salmon in the Barents region and
genetic stock identification of coastal fishery catches from Norway and Russia [online].
Kolarctic ENPI CBC — Kolarctic salmon project (KO197) report. Available from
www.fylkesmannen.no/kolarcticsalmon.

Wang, T., C. C. Hung, and D. J. Randall. 2006. The comparative physiology of food deprivation:
from feast to famine. Annual Review of Physiology **68**:223-251. doi:
10.1146/annurev.physiol.68.040104.105739.

Werner, E. E., and J. F. Gilliam. 1984. The Ontogenetic Niche and Species Interactions in Size-
Structured Populations. Annual Review of Ecology and Systematics **15**:393-425. doi:
10.1146/annurev.es.15.110184.002141.

Wood, S. N. 2011. Fast stable restricted maximum likelihood and marginal likelihood estimation of
semiparametric generalized linear models. Journal of the Royal Statistical Society Series B-
Statistical Methodology **73**:3-36. doi: 10.1111/j.1467-9868.2010.00749.x.