1	Species and sex divergence in vocalizations between hybridizing role-reversed
2	shorebirds, Northern Jacana (Jacana spinosa) and Wattled Jacana (Jacana
3	jacana)
4	
5	Evan J. Buck ¹ , Toni Brown ² , Gina Zwicky ² , Elizabeth P. Derryberry ^{1,2} , Sara E. Lipshutz ^{1,2,3*}
6	¹ Department of Ecology and Evolutionary Biology, University of Tennessee, Knoxville, TN,
7	USA
8	² Department of Ecology and Evolutionary Biology, Tulane University, New Orleans, LA, USA
9	³ Department of Biology, Indiana University, Bloomington, IN, USA
10	* Corresponding author: slipshut@iu.edu
11	
12	ABSTRACT—Species-specific vocalizations can act as a reproductive isolating mechanism
13	between closely related populations. We analyzed vocal divergence between two hybridizing
14	species of sex-role reversed polyandrous shorebirds, the Northern Jacana (Jacana
15	spinosa) and Wattled Jacana (Jacana jacana). We found that J. spinosa calls have higher peak
16	frequency and fundamental frequency than J. jacana calls. We also compared calls
17	between males and females, as both jacana species are sex-role reversed and females compete for
18	male mates. Males produce calls with a higher peak frequency, exhibit shorter note lengths and
19	emit a greater number of notes within a calling bout than females, which could relate to mate
20	attraction. These results suggest that vocal divergence could act as a behavioral barrier to limit
21	hybridization between the species and vocalizations may function differently between male and
22	female jacanas.

23

- 24 Key words: call, hybridization, jacanas, sex differences, shorebird, vocal divergence
- 25

Resumen. Divergencia específica y sexual en las vocalizaciones de las aves costeras de roles sexuales invertidos Jacana Norteña (*Jacana spinosa*) y Jacana Carunculada (*Jacana*)

28 *jacana*).

29 Las vocalizaciones especie-específicas pueden actuar como mecanismos de aislamiento reproductivo entre poblaciones de especies estrechamente relacionadas. Analizamos la 30 31 divergencia en vocalizaciones entre dos especies de aves costeras poliándricas de rol sexual invertido, Jacana Norteña (Jacana spinosa) y Jacana Carunculada (Jacana jacana). Encontramos 32 que los llamados de J. spinosa contienen frecuencias pico y fundamental más altas que los 33 34 llamados de J. jacana. También comparamos los llamados entre machos y hembras en ambas especies, ya que ambas tiene el rol sexual invertido y las hembras compiten por parejas. Los 35 36 machos producen llamados con una frecuencia pico mayor, exhiben longitudes menores de notas y emiten un número mayor de notas dentro de un despliegue de vocalizaciones y producen notas 37 38 de menor duración que las hembras, lo que podría relacionarse con atracción de pareja. Estos 39 resultados sugieren que la divergencia en vocalizaciones podría actuar como barrera 40 comportamental para limitar la hibridación entre las especies y estas vocalizaciones pueden 41 funcionar distintamente entre machos y hembras de jacanas. Estudios futuros utilizando 42 experimentos de reproducción de audio podrían poner a prueba estas hipótesis. 43 Palabras clave: llamada, hibridación, jacanas, diferencias de sexo, ave costera, divergencia vocal 44

Between closely related species, divergence in mating signals can facilitate reproductive 45 isolation and drive the process of speciation (Irwin and Price 1999; Coyne and Orr 2004). 46 47 Whereas mating signals are used to attract and compete for mates within populations, divergence between populations can lead to a breakdown in communication such that individuals do not 48 recognize potential mates or rivals (Coyne and Orr 2004). Evidence from a wide range of taxa 49 50 suggests that the degree of divergence in mating signals influences the extent to which individuals discriminate between congeners, which ultimately shapes mating outcomes 51 (Andersson 1994). Therefore, it is important to understand how and why mating signals diverge 52 53 between populations. Hybrid zones – regions where distinct species come into contact and 54 interbreed – provide a natural experiment to examine the consequences of vocal divergence for 55 behavioral isolation (Hewitt 1988).

56 Divergent mating signals can serve to reproductively isolate species with otherwise incomplete barriers to gene flow (Grant and Grant 1997; Price 2008). Learned vocalizations, 57 58 such as oscine songs (Nottebohm 1972), have the potential to diverge rapidly via cultural 59 evolution (Mason et al. 2016) and are therefore important pre-mating barriers to gene flow 60 between hybridizing populations (Slabbekoorn and Smith 2002; Uy et al. 2018). In contrast, 61 innate vocalizations diverge more slowly than learned vocalizations, and there is mixed evidence 62 for the role of innate vocalizations as behavioral barriers to gene flow. For example, innate 63 vocalizations serve as behavioral barriers to hybridization in Alectoris partridges (Ceugniet and 64 Aubin 2001) and Streptopelia doves (De Kort et al. 2002), but not in Callipepla (Gee 2005), nor 65 Coturnix quails (Derégnaucourt and Guyomarc'h 2003). This leaves an important gap in our 66 understanding about innately derived vocalizations and how they vary between closely related 67 species that are hybridizing.

68	In many bird species, both males and females vocalize (Odom and Benedict 2018). Vocal
69	traits such as note length, complexity or production rate may differ between the sexes depending
70	on their function in courtship and other behavioral contexts (Appleby et al. 1999; Odom and
71	Mennill 2010; ten Cate 1997). For example, the sex that competes more for mates tends to
72	vocalize more often (Sordahl 1979; Sung et al. 2005). Whereas male competition for mates is
73	common across animals, females of some species are sex-role reversed, meaning they face
74	stronger competition for mates than males do (Emlen and Oring 1977). Currently, we know very
75	little about how female and male vocalizations compare in sex-role reversed species.
76	Sexually dimorphic vocalizations may also occur due to physical differences between
77	males and females. In many species the sexes diverge in body size. As a consequence,
78	morphological constraints on sound production can lead to distinctive spectral and temporal
79	characteristics between male and female vocalizations (Ryan and Brenowitz 1985; Ten Cate
80	1997). Consistent with signal design theory, larger body size is often associated with lower
81	sound frequencies both between and within sexes (Barbraud et al. 2000; Maurer et al. 2008).
82	Some taxa are exceptions to this rule; for example, many female owls are larger than males but
83	have higher frequency calls (Odom and Mennill 2010). Therefore, it is not clear whether signal
84	design theory will hold in sex-role reversed species, in which females are often larger than
85	males.
86	Jacanas are tropical, sex-role reversed shorebirds in which selection on females to

compete for mates is stronger than on males (Jenni 1974). The Northern Jacana (*Jacana spinosa*)
and Wattled Jacana (*J. jacana*) have been isolated for 700,000 years (Miller et al. 2014) and
hybridize in a narrow region in Panama (Lipshutz et al. 2019). It is unknown whether their
vocalizations are divergent, and what role their calls play in maintaining reproductive isolation

4

91	between the species. Here, we quantify variation in temporal and spectral characteristics
92	between the species and the sexes. We predict that vocalizations between J. spinosa and J.
93	jacana will be divergent, and that the larger-bodied J. spinosa will have lower frequency-related
94	characteristics. Second, we examine vocal divergence between males and females of both
95	species. Jacanas have extreme sexual dimorphism in size, and females weigh up to 60% more
96	than males (Emlen and Wrege 2004; Jenni and Collier 1972). Because female jacanas are larger
97	than males, we predict that female vocalizations will have lower frequency-related
98	characteristics. We also predicted that females should produce more calls than males, given that
99	these species are sex-role reversed.
100	
101	Methods
101	
102	Sound recordings
102 103	Sound recordings We recorded vocalizations from June-August 2015 and June-July 2018 at 9 different sites in
102 103 104	Sound recordings We recorded vocalizations from June-August 2015 and June-July 2018 at 9 different sites in Panama (Fig. 1). Across these sites we recorded a total of 12 individuals of each species and sex.
102 103 104 105	Sound recordings We recorded vocalizations from June-August 2015 and June-July 2018 at 9 different sites in Panama (Fig. 1). Across these sites we recorded a total of 12 individuals of each species and sex. Birds were either stimulated with playback and a taxidermic mount to elicit vocalizations, or in
102 103 104 105 106	Sound recordings We recorded vocalizations from June-August 2015 and June-July 2018 at 9 different sites in Panama (Fig. 1). Across these sites we recorded a total of 12 individuals of each species and sex. Birds were either stimulated with playback and a taxidermic mount to elicit vocalizations, or in some cases vocalizations were stimulated by the presence of the recordist near the bird's
102 103 104 105 106 107	Sound recordings We recorded vocalizations from June-August 2015 and June-July 2018 at 9 different sites in Panama (Fig. 1). Across these sites we recorded a total of 12 individuals of each species and sex. Birds were either stimulated with playback and a taxidermic mount to elicit vocalizations, or in some cases vocalizations were stimulated by the presence of the recordist near the bird's territory.
102 103 104 105 106 107 108	Sound recordings We recorded vocalizations from June-August 2015 and June-July 2018 at 9 different sites in Panama (Fig. 1). Across these sites we recorded a total of 12 individuals of each species and sex. Birds were either stimulated with playback and a taxidermic mount to elicit vocalizations, or in some cases vocalizations were stimulated by the presence of the recordist near the bird's territory. Recordings were made using a Marantz PMD661 MKII solid state digital recorder
102 103 104 105 106 107 108 109	Sound recordings We recorded vocalizations from June-August 2015 and June-July 2018 at 9 different sites in Panama (Fig. 1). Across these sites we recorded a total of 12 individuals of each species and sex. Birds were either stimulated with playback and a taxidermic mount to elicit vocalizations, or in some cases vocalizations were stimulated by the presence of the recordist near the bird's territory. Recordings were made using a Marantz PMD661 MKII solid state digital recorder (Marantz professional, Cumberland, Rhode Island, United States) set at 44.1 kHz sampling rate,
102 103 104 105 106 107 108 109 110	Sound recordings We recorded vocalizations from June-August 2015 and June-July 2018 at 9 different sites in Panama (Fig. 1). Across these sites we recorded a total of 12 individuals of each species and sex. Birds were either stimulated with playback and a taxidermic mount to elicit vocalizations, or in some cases vocalizations were stimulated by the presence of the recordist near the bird's territory. Recordings were made using a Marantz PMD661 MKII solid state digital recorder (Marantz professional, Cumberland, Rhode Island, United States) set at 44.1 kHz sampling rate, 16-bit, and WAV file type, and a Sennheiser K6 power module with a Sennheiser M67 shotgun
102 103 104 105 106 107 108 109 110 111	Sound recordings We recorded vocalizations from June-August 2015 and June-July 2018 at 9 different sites in Panama (Fig. 1). Across these sites we recorded a total of 12 individuals of each species and sex. Birds were either stimulated with playback and a taxidermic mount to elicit vocalizations, or in some cases vocalizations were stimulated by the presence of the recordist near the bird's territory. Recordings were made using a Marantz PMD661 MKII solid state digital recorder (Marantz professional, Cumberland, Rhode Island, United States) set at 44.1 kHz sampling rate, 16-bit, and WAV file type, and a Sennheiser K6 power module with a Sennheiser M67 shotgun microphone and windscreen (Sennheiser electronic corporation, Wedemark, Germany). We

1-32 call bouts per individual) in Audacity 2.1.2 (Audacity 2018). Call bouts are defined as a
series of evenly spaced notes less than 1 second apart.

115 Acoustic measurement

116 Jacana vocalizations contain harmonics covering a wide frequency bandwidth (Mace 1981). We

took measurements on one call type, repeated note calls (Jenni 1974), as these were consistently

found in recordings of both species and sexes (Fig. 2). We used the sound-analysis software

119 Luscinia (Lachlan 2007) to generate Fourier-based spectrograms. Calls were high pass filtered to

eliminate low frequency background noise below 200 Hz. We used the following settings to

measure call variation: FF jump suppression = 20, Max. Frequency (Hz) = 15,000, Frame length

122 (ms) = 5, Time step (ms) = 1, Spectrograph points = 221, Spectrogram Overlap % = 80, Dynamic

range (dB) = 50, Dynamic equalization (ms) = 0, Dynamic comp. % = 100, Dereverberation % =

124 200, Dereverberation range (ms) = 100, Windowing function = Gaussian, Frequency zoom % =

125 150, Time zoom % = varies, Noise removal (dB) = 0, NR range1 (ms) = 50, R range2 (ms) = 50.

126 We manually measured vocalizations in Luscinia (Lachlan 2007) by individually tracing

127 each note. We used the Luscinia software to automatically calculate three acoustic parameters:

note length (msec), peak frequency (frequency of the maximum amplitude, Hz), and fundamental

129 frequency (Hz). We averaged these parameters for each note within a call bout (range 2-70 notes,

130 mean 11.6) (Table 1). We also averaged the number of notes per bout for each individual to

131 calculate a fourth acoustic parameter, notes per bout. The four parameters were averaged in

132 Microsoft Excel after exporting from Luscinia.

133 Statistical analysis

134 We compared the four call parameters between the species and sexes using Student's *t*-tests or

135 Wilcoxon rank sum, depending on whether the parameters were normally distributed or not.

136	We also summarized the four acoustic parameters with a principal components analysis				
137	(PCA) using the prcomp function in R version 3.3.2 (R-Core-Team, 2015). Prior to the PCA, we				
138	log-transformed acoustic data to fulfill assumptions of multi-normality. We retained two PC				
139	scores that explained 60.6% and 19.7% of the variation in the acoustic parameters, totaling				
140	80.7% cumulative proportion of variation (Table S1). Frequency variables loaded negatively				
141	onto PC1 and positively onto PC2. Note length loaded positively on PC1 and PC2, and notes per				
142	bout loaded negatively onto PC1 and PC2. We compared vocalizations between the species an				
143	sexes using linear mixed effects models with the nmle package (Pinheiro et al. 2018) in R. We				
144	included vocalization PC1 and PC2 as separate response variables, species and sex as the fixed				
145	effects, and site as a random effect. We visually inspected residual plots to ensure they did not				
146	deviate from normality. We used a type III ANOVA to determine whether species and/or sex				
147	were significant predictors of variation in the PCs.				
148	We also conducted a discriminant function analysis (DFA) in R using the package				
149	flipMultivariates to assess whether spectral or temporal parameters could distinguish between				
150	species and sexes (https://github.com/Displayr/flipMultivariates/).				
151					
152	Results				
153	Species differences in vocalizations				
154	Vocalizations are different between these two jacana species, particularly regarding spectral				
155	characteristics (Table 1; Fig. 3). For both males and females, J. spinosa calls have significantly				
156	higher peak (t = -5.1, df = 41.8, $P < 0.001$) and fundamental frequencies (W = 42, $P < 0.001$)				
157	than J. jacana calls. Species was also a significant predictor both of vocalization PC1 ($F_{1,7}$ =				

of 48 (92%) individuals were classified to the correct species by a DFA. The best variables to distinguish the species were peak frequency ($r^2 = 0.36$, P < 0.001) and fundamental frequency (r^2 = 0.48, P < 0.001).

162 Sex differences in vocalizations

163 Vocalizations are also different between males and females of both species (Table 1; Fig. 3).

164 When species are combined, males have significantly more calls within a bout (W = 189.5, P =

165 0.043) and higher peak frequency calls (t = -2.7, df = 44.9, P = 0.009) than females. Females

- have longer calls than males (t = 3.1, df = 40.7, P = 0.003). Sex was also a significant predictor
- of vocalization PC1 ($F_{1,38} = 14.8$, P = < 0.001) but not of PC2 (Fig. S1). Using all acoustic

parameters, 45 out of 48 (73%) individuals were classified to the correct sex by a DFA. The best

variables to distinguish the sexes were call length ($r^2 = 0.17$, P = 0.005), peak frequency ($r^2 =$

170 0.14, P = 0.014), and calls within a bout ($r^2 = 0.12$, P = 0.028).

171 The sexes were more strongly differentiated by peak frequency in *J. jacana* (t = -3.0193,

172 df = 21.98, P = 0.006) than in J. spinosa (t = -1.9602, df = 21.999, P = 0.063). Similarly, males

and females differed more strongly in note length in *J. jacana* (t = 3.3939, df = 19.602, P =

174 0.003) than in *J. spinosa* (t = 1.6899, df = 19.318, P = 0.11). In contrast, the sexes differed more 175 strongly in calls within a bout for *J. spinosa* (W = 38.5, P = 0.057) than *J. jacana* (W = 53, P =

176 0.29).

- 177
- 178

Discussion

Jacana spinosa and *J. jacana* calls are different - *J. spinosa* calls are significantly higher
in peak and fundamental frequency. The sexes are consistently different across both species,

181 such that male calls have higher peak frequency and more notes than female calls, whereas182 female calls have longer note lengths.

183 Species Differences

184 Vocalizations of the two species of jacana have diverged spectrally but are similar temporally. In 185 other species of non-oscine birds that do not learn their songs, temporal traits change at slower 186 rates than frequency-related traits (Miller and Baker 2009, Seneviratne et al. 2012), which is consistent with our findings that spectral traits were more divergent than temporal traits between 187 these jacana species. Counter to the prediction that smaller-bodied species should have calls with 188 higher frequencies (Ryan and Brenowitz 1985), larger-bodied J. spinosa have higher peak and 189 190 fundamental frequency vocalizations than smaller-bodied J. jacana, for both sexes. One 191 hypothesis for this contradiction is that species divergence in vocalizations could relate to 192 differing environmental or habitat characteristics that have shaped their call frequencies (Morton 1975, Endler 1992). In a prior study, species distribution modeling indicated that J. spinosa 193 194 favors a warmer and wetter environment (Miller et al. 2014), suggesting that the species have 195 diverged in habitat preferences. Future work could compare the vegetative cover in the habitat of each species to determine whether the higher frequency vocalizations of J. spinosa relate to a 196 197 more open habitat. Another potential explanation for vocal divergence could be a difference in 198 the syringeal or bill structure between species (Seneviratne et al. 2012; Kingsley et al. 2018).

Sex Differences

We found that frequency differences in jacana vocalizations matched our predictions for females and males based on their body size dimorphism; males of both species produce higher frequency calls than the larger-bodied females. In many species with sexually dimorphic body size, the smaller of the two sexes produces higher frequency vocalizations (Maurer et al. 2008). Femalebiased size dimorphism is common in shorebirds other than jacanas, and larger females also have
lower frequency calls (Heidemann and Oring 1976; Douglas 1998). Contrary to our expectation,
male jacanas within our study also emitted more notes per calling bout than females. One likely
explanation for this is mate attraction: in the polyandrous Bronze-winged Jacana (*Metopidius indicus*), individual males that called more frequently received more copulations than other comates (Butchart et al. 1999). Calling rate could be a sexually selected trait in male Neotropical
jacanas, and this hypothesis should be tested using behavioral playback experiments.

211 Application and conclusion

212 We found that Neotropical jacanas diverged significantly in the peak and fundamental

213 frequencies of their vocalizations. Diverged vocal signals could promote reproductive isolation

between the two species when they come into contact. Furthermore, these spectral characteristics

differed between the species for both sexes, suggesting that both male and female signals could

facilitate species-specific discrimination in the hybrid zone. A phenotypic and genomic analysis

of the jacana hybrid zone found that species-specific traits such as plumage and facial

ornamentation were likely prezygotic barriers that maintain species boundaries (Lipshutz et al.

- 219 2019). Future playback studies could assess the relative role of visual and vocal signals as
- behavioral barriers to mating between the species. This phenotypic differentiation between *J*.
- spinosa and J. jacana likely contributes to the low occurrence of hybrids within the narrow

hybrid zone and may be one of the reasons for limited hybridization between the species.

223

224

Acknowledgments

This material is supported by the University of Tennessee's Ready for the World Chancellor's
 Honors Program and Summer Undergraduate Research Internship to EJB, National Science

227	Foundation Graduate Research Fellowship Grant No. 1154145, Doctoral Dissertation						
228	Improvement Grant IOS-1818235, and a Smithsonian Tropical Research Institute short-term						
229	reliowship to SEL, Tulane University's CELT summer research and faculty-student scholarly						
230	grants to GZ and a Louisiana Board of Personts NSE EDSCoP LINK Grant No. 177 to EDD						
221	Any oninion findings and conclusions or recommondations expressed in this material are those						
232 222	Any opinion, indings, and conclusions of recommendations expressed in this material are those of the authors(s) and do not necessarily reflect the views of the National Science Foundation						
235	of the authors(s) and do not necessarily reflect the views of the National Science Foundation.						
234	ΔNAM) Panama's environmental authority (nermit numbers: SE/A-45-12, SE/A-46-14, SE/A-						
235	17.18) and recording techniques were approved by the Institutional Animal Care and Use						
230	Committee of the Smithsonian Tropical Research Institute (IACUC permits: 2012-0315-2015						
237	2018 0116 2021) Tulana University (IACUC permit: 0446P) and the University of Tennessee						
200 220	(IACUC permit: 2573) We thank the STRI Bird Collection for preparing taxidermic mounts						
239	Daniel Ernesto Buitrago and Marcelo Araya Salas assisted with translating the abstract into						
240	Snanish Jacana illustrations by Stephanie McClelland						
242	Spanish. sucula mastations by Stephanic Weelenand.						
243	Data accessibility						
244	Recordings are available on xeno canto. Jacana spinosa: https://www.xeno-canto.org/set/4909						
245							
246							
247	Literature Cited						
248	Andersson M. 1994. Sexual selection (Monographs in behavior and ecology). Princeton (NJ):						
249	Princeton University Press.						
250	Appleby BM, Yamaguchi N, Johnson PJ, MacDonald DW. 1999. Sex-specific territorial						
251	responses in Tawny Owls Strix aluco. Ibis. 141:91–99.						
252	Audacity. 2018. Audacity ® Free, open source, cross-platform audio software for multi-track						
253	recording and editing. Audacity. doi: ISSN 1980-4431.						
<u></u>							
254	Baker MC, Boylan J1. 1999. Singing Behavior, Mating Associations and Reproductive Success						
255	in a Population of Hybridizing Lazuli and Indigo Buntings. The Condor. 101:493-504.						

256 Barbraud C, Mariani A, Jouventin P. 2000. Variation in call properties of the sn
--

257 Pagodroma nivea, in relation to sex and body size. Australian Journal of Zoology.

48:421–430.

259 Butchart S, Seddon N, Ekstrom JMM. 1999. Yelling for sex: Harem males compete for female

access in bronze-winged jacanas. Animal Behaviour. 57:637–646.

261 Catchpole CK, Slater PJB. 2008. Bird song: Biological themes and variations, second edition.

262 Cambridge (UK): Cambridge University Press.

263 Ceugniet M, Aubin T. 2001. The rally call recognition in males of two hybridizing partridge

species, red-legged (Alectoris rufa) and rock (A. graeca) partridges. Behavioural

265 Processes. 55:1–12.

- 266 Coyne JA, Orr AH. 2004. Speciation. Sunderland (MA): Sinauer Associates.
- 267 De Kort SR, Den Hartog PM, Ten Cate C. 2002. Vocal signals, isolation and hybridization in the
- vinaceous dove (Streptopelia vinacea) and the ring-necked dove (S. capicola). Behavioral
 Ecology and Sociobiology. 51:378–385.

270 Derégnaucourt S, Guyomarch JC. 2003. Mating call discrimination in female European

271 (Coturnix c. coturnix) and Japanese quail (Coturnix c. japonica). Ethology. 109:107–119.

272 Douglass HD. 1998. Response of Eastern Willets (Catoptrophorus s. semipalmatus) to

Vocalizations of Eastern and Western (C. s. inornatus) Willets. The Auk. 115:514–518.

Emlen ST, Oring LW. 1977. Ecology, sexual selection, and the evolution of mating

systems. Science. 197:215–223.

Emlen ST, Wrege PH. 2004. Size Dimorphism, Intrasexual Competition, and Sexual Section in

277 Wattled Jacana (Jacana jacana), a Sex-Role-Reversed Shorebird in Panama. The Auk.

278 121:391–403.

- Endler J. 1992. Signals, Signal Conditions, and the Direction of Evolution. The American
 Naturalist 139:S125.
- 281 Gee JM. 2005. No species barrier by call in an avian hybrid zone between California and
- Gambels quail (Callipepla californica and C. gambelii). Biological Journal of the Linnean
- 283 Society. 86:253–264.
- Grant PR, Grant BR. 1997. Genetics and the origin of bird species. Proceedings of the National
 Academy of Sciences. 94:7768–7775.
- Hunt J, Breuker CJ, Sadowski JA, Moore AJ. 2009. Male-male competition, female mate choice
- and their interaction: Determining total sexual selection. Journal of Evolutionary Biology.
 22:13–26.
- Heidemann MK, Oring LW. 1976. Functional Analysis of Spotted Sandpiper (Actitis macularia)
 Song. Behaviour. 56:181–193.
- Irwin DE, Price T. 1999. Sexual imprinting, learning and speciation, Heredity. 82:347–354.
- Jenni DA. 1974. Evolution of polyandry in birds. Integrative and Comparative Biology. 14:129–
 144.
- Jenni DA, Collier G. 1972. Polyandry in the American Jaçana (Jacana spinosa). The Auk.
 89:743–765.
- 296 Kingsley EP, Eliason CM, Riede T, Li Z, Hiscock TW, Farnsworth M, Thomson SL, Goller F,
- Tabin CJ, Clarke JA. 2018. Identity and novelty in the avian syrinx. Proceedings of the
 National Academy of Sciences. 115:10209-10217.
- Konishi M. 1963. The Role of Auditory Feedback in the Vocal Behavior of the Domestic Fowl.
 Zeitschrift für Tierpsychologie. 20:349–367.
- 301 Lachlan RF. 2007. Luscinia: a bioacoustics analysis computer program.

302	Lipshutz SE	Meier JI.	Derryberry	v GE, Miller N	/IJ. Seehausen	O. Derry	berry EP. 2	2019.
		, ,		/ ,		-,,		

- 303 Differential introgression of a female competitive trait in a hybrid zone between sex-role
 304 reversed species. Evolution. 73:188–201.
- Lynch A. 1996. The population memetics of birdsong. In: Kroodsma DE, Miller EH, editors.
- 306 Ecology and evolution of acoustic communication in birds. Ithaca (NY): Comstock
- 307 Publishers; p.181–197.
- Mace TR. 1981. Causation, function, and variation of the vocalizations of the Northern Jacana,
 Jacana spinosa. [dissertation]. Missoula (MT): University of Montana.
- 310 Mason NA, Burns KJ, Tobias JA, Claramunt S, Seddon N, Derryberry EP. 2016. Song evolution,

speciation, and vocal learning in passerine birds. Evolution. 71:786–796.

- Maurer G, Smith C, Süsser M, Magrath RD. 2008. Solo and duet calling in the pheasant coucal:
- 313 Sex and individual call differences in a nesting cuckoo with reversed size dimorphism.
- Australian Journal of Zoology. 56:143–149.
- Miller EH, Baker AJ. 2009. Antiquity of Shorebird Acoustic Displays. The Auk. 126:454–459.
- 316 Miller MJ, Lipshutz SE, Smith NG, Bermingham E. 2014. Genetic and phenotypic
- 317 characterization of a hybrid zone between polyandrous Northern and Wattled Jacanas in
- 318 Western Panama. BMC Evolutionary Biology. 14:227.
- Nottebohm F. 1972. The Origins of Vocal Learning. The American Naturalist. 106:116–140.
- 320 Odom, KJ, Benedict L. 2018. A call to document female bird songs: Applications for diverse
- 321 fields. The Auk 135:314–325.
- 322 Odom KJ, Mennill DJ. 2010. A Quantitative Description of the Vocalizations and Vocal
- Activity of the Barred Owl. The Condor. 112:549–560.

- 324 Pinheiro J, Bates D, DebRoy S, Sarkar D, Heisterkamp S, Van Willigen B. 2018. nlme: Linear
- and Nonlinear Mixed Effects Models. R package version 3.1-137, Retrieved from
- 326 https://cran.r-project.org/package=nlme.
- 327 Price T. 2008. Speciation in Birds. Greenwood Village (CO): Roberts and Company Publishers.
- 328 R-Core-Team. 2015. R: A language and environment for statistical computing. Vienna, Austria:
- 329 R Foundation for Statistical Computing.
- Ryan MJ, Brenowitz EA. 1985. The Role of Body Size, Phylogeny, and Ambient Noise in the
 Evolution of Bird Song. The American Naturalist. 126:87–100.
- 332 Seneviratne SS, Jones IL, Carr SM. 2012. Patterns of vocal divergence in a group of non-oscine
- birds (auklets; Alcidae, Charadriiformes), Evolutionary Ecology Research. 1:95–112.
- Slabbekoorn H, Smith TB. 2002. Bird song, ecology and speciation, Philosophical Transactions
 of the Royal Society B: Biological Sciences. 357:493–503.
- 336 Sordahl TA. 1979. Vocalizations and behavior of the Willet Catoptrophorus semipalmatus.
- 337 Wilson Bulletin. 91:551–574.
- 338 Sung H-C, Miller EH, Flemming SP. 2005. Breeding vocalizations of the piping plover
- 339 (Charadrius melodus): structure, diversity, and repertoire organization. Canadian Journal
 340 of Zoology. 83.579–595.
- 341 Taoka M, Sato T, Kamada T, Okumura H, 1989. Sexual Dimorphism of Chatter-Calls and Vocal
- 342 Sex Recognition in Leachs Storm-Petrels (Oceanodroma leucorhoa). The Auk:
- 343 Ornithological Advances. 106:498–501.
- Ten Cate C. 1997. Sex Differences in the Vocalizations and Syrinx of the Collared Dove
 (Streptopelia decaocto). The Auk. 114:22–39.

- 346 Uy JAC, Irwin DE, Webster MS. 2018. Behavioral Isolation and Incipient Speciation in Birds.
- Annual Review of Ecology, Evolution, and Systematics. 49:1–24.
- 348





Figure 1. Sampling map of *Jacana spinosa* (yellow) and *J. jacana* (red) vocalizations recorded

across the hybrid zone Panama. Circle size represents sample size (minimum 1, maximum 19).



Figure 2. Spectrograms of vocalizations for a) female *J. spinosa*, b) male *J. spinosa*, c) female *J. jacana*, d) male *J. jacana*

352





357 (light) and female (dark) *Jacana spinosa* (yellow) and *J. jacana* (red).

355

from male and female Jacana spinosa and J. jacana.						
	Jacana spinosa		Jacana jacana			
	Male	Female	Male	Female		
Number of Individuals	12	12	12	12		
Notes per Bout	15.97 ± 2.36	9.75 ± 1.55	14.03 ± 2.42	10.32 ± 1.66		
Range of Notes per Bout	2 - 45	2 - 33	3 - 70	2 - 68		
Note Length (ms)	70.17 ± 5.09	85.51 ± 7.52	78.12 ± 2.98	95.87 ± 4.3		
Peak Frequency (Hz)	2898.36 ± 129.97	2539.14 ± 129.2	2221.52 ± 166.34	1521.63 ± 161.45		
Fundamental Frequency (Hz)	1346.54 ± 112.46	1099.14 ± 99.4	731.83 ± 24.71	663.84 ± 23.46		

Table 1. Sampling information and spectral and temporal characteristics (mean \pm SE) for vocalizations recorded from male and female *Jacana spinosa* and *J. jacana*.

³⁵⁸





361 Supplemental Figure 1. Principal components analysis of *Jacana spinosa* (yellow) and *Jacana*

362 *jacana* (red) males (light) and female (dark) vocalizations

Component Analysis		
Vocal Parameter	PC1	PC2
Eigenvalue	1.56	0.89
Proportion of variance	60.60%	19.70%
Note per bout	-0.47	-0.58
Note length (ms)	0.49	0.45
Peak frequency (Hz)	-0.53	0.45
Fundamental frequency (Hz)	-0.52	0.5

Supplemental Table 1. Loadings for Principal

363

360