On the relative ease of speciation with periodic gene flow

Ethan Linck[1,*] and C.J. Battey[2]

[1] *Department of Biology & Burke Museum of Natural History & Culture, University of Washington, Seattle, WA 98195 USA*
[2] *Institute of Ecology and Evolution, University of Oregon, Eugene, OR 97402*

*Corresponding Author: ethanblinck@gmail.com

ABSTRACT

Common models of speciation with gene flow consider constant migration or admixture on secondary contact, but earth's recent climatic history suggests many populations have experienced cycles of isolation and contact over the last million years. How does this process impact the rate of speciation, and how much can we learn about its dynamics by analyzing the genomes of modern populations? Here we develop a simple model of speciation through Bateson-Dobzhansky-Muller incompatibilities in the face of periodic gene flow and validate our model with forward time simulations. We then use empirical atmospheric CO_2 concentration data from the Vostok Ice Cores to simulate cycles of isolation and secondary contact in a tropical montane landscape, and ask whether they can be distinguished from a standard isolation-with-migration model by summary statistics or joint site frequency spectrum-based demographic inference. We find speciation occurs much faster under periodic than constant gene flow with equivalent effective migration rates (Nm). These processes can be distinguished through combinations of summary statistics or demographic inference from the site frequency spectrum, but parameter estimates appear to have little resolution beyond the most recent cycle of isolation and migration. Our results suggest speciation with periodic gene flow is a common force in generating species diversity through Pleistocene climate cycles, and highlight the limits of current inference techniques for demographic models mimicking the complexity of earth's recent climatic history.

INTRODUCTION

Eight decades after the modern synthesis the relative roles of gene flow, geography, and natural selection in speciation elude easy synthesis. For most of the 20th century, divergence in strict allopatry was assumed to be the dominant, if not the only, mode of speciation in most taxa, leading to an emphasis on the geography of diverging populations and an implicit suggestion of a major role for genetic drift (Dobzhansky, 1937; Mayr, 1942,

1963; Coyne and Orr, 2004; Provine, 2004). While a burgeoning body of theory has

demonstrated the plausibility of speciation with gene flow in a range of circumstances

(Smith, 1966; Felsenstein, 1981; Gavrilets, 2003), a lack of clear empirical examples and

the related difficulty of disproving a null hypothesis of allopatric speciation did little to

contest this paradigm.

More recently, a renewed emphasis on the role of natural selection in speciation in

the context of adaptive radiations and "ecological" speciation has questioned the

requirement of strict isolation (Schluter, 2001; Via, 2009; Nosil, 2012). The rise of genomic

datasets from a wide array of model and nonmodel organisms and the concordant

development of sophisticated statistical tools for demographic inference has further

complicated previous assumptions (Nosil, 2008). Coalescent theory and genome-wide

genetic markers provide the basis for simultaneous model selection and inference of

parameter values, now implemented in a range of computational tools (Nielsen and

Wakeley, 2001; Hey and Nielsen, 2007; Gutenkunst et al., 2009; Hey, 2010; Jouganous

et al., 2017). As much as an early consensus from the current proliferation of studies

applying these methods can be gleaned, it appears that gene flow between lineages at

various points in the speciation continuum is more common than previously assumed

(Mallet, 2008; Nosil, 2008; Strasburg and Rieseberg, 2008; Cui et al., 2013; Martin et al.,

2013; Kumar et al., 2017; Linck et al., 2019).

While more complex models of speciation dynamics can be applied using these

methods, e.g., by allowing parameter values to vary at different loci across the genome

(Rougeux et al., 2017), model selection has largely treated migration (m; the probability a

given allele is an immigrant) as an average rate in one of a maximum of two time periods

(Figure 1) (Sousa and Hey, 2013). Traditional isolation with migration models are

therefore distinguished from secondary contact by the presence of a time period (T_1 to T_2)

where $m=0$. Even this relatively sophisticated model is clearly a dramatic simplification of

speciation dynamics in most systems, however, where alternative assumptions of

continuous gene flow (recurrent migration every generation) or strict isolation are unrealistic in most cases. For example, divergence in allopatry is widely believed to be the dominant mode of speciation in birds (Mayr, 1963; Price, 2008). Yet the extreme vagility of many bird taxa poses the question of how readily geographic classifications of speciation map on to realized rates of gene flow, leading some researchers to strongly advocate for strictly process-based definitions (Fitzpatrick et al., 2008; Mallet, 2008). Further complications arise from the imprecise, redundant language associated with the study of speciation itself (Harrison, 2012).

An alternative to either divergence in strict isolation or in the face of recurrent migration involves alternating phases of interbreeding and isolation, i.e., speciation with intermittent (or "cyclical" / "periodic") gene flow. Cyclical processes are common over geologic, evolutionary, and ecological time scales, and include predator-prey dynamics (Elton and Nicholson, 1942), glacial cycling (Roy et al., 1996), ecological succession (Levin and Paine, 1974), and fluctuating population sizes (Vucetich et al., 1997). Yet the role of cyclical processes in generating or retarding evolutionary change has seen relatively little attention, despite evidence of unique dynamics from a handful of empirical and theoretical studies (Duckworth and Semenov, 2017; Alcala and Vuilleumier, 2014; Otto and Whitlock, 1997). Ehrendorfer (1959) proposed that "differentiation-hybridization" cycles were responsible for evolutionary patterns in the flowering plant genus Achillea, viewing hybridization as a strictly homogenizing force, an idea further developed by Rattenbury (1962) as an explanation for the diversity and persistence of the New Zealand flora. In phylogeography, Pleistocene glacial cycles in Boreal regions and the Amazon have been invoked to explain both diversification (Lovette, 2005) and patterns of genetic diversity and population genetic structure (Klicka and Zink, 1999), though the link between population subdivision and speciation remains unclear (Harvey et al., 2017).

Recently, He et al. (2019) proposed that cycles of gene flow and isolation constituted a new model of speciation, which they term mixing-isolation-mixing (or MIM), and suggest

it can lead to accelerated rates of diversification. Given its relaxed requirements and flexible application to a range of potentially stochastic histories, speciation with periodic gene flow is plausibly more common than either speciation in strict allopatry or with continuous gene flow. Furthermore, recent studies on hybridization as a generative force in evolution, through either homoploid hybrid speciation (Gompert et al., 2006; Brelsford et al., 2011; Schumer et al., 2014, 2018) or adaptive introgression (Delmore et al., 2015; Norris et al., 2015; Racimo et al., 2015; Irwin et al., 2018) suggest the interaction of gene flow and isolation could promote accelerated speciation under some circumstances.

If the MIM model (hereafter "speciation with periodic gene flow") is common and a realistic description of speciation dynamics in many circumstances, a more explicit description of its features and assessment of its consequences and genomic signature will be useful for a broad range of evolutionary biologists. As "pulse" models of admixture are widely applied in contemporary population genomics (Loh et al., 2013; Medina et al., 2018), these results may also inform our understanding of complex demographic histories below the species level. Here, we use theory and simulations to ask 1) if speciation is indeed easier to achieve under periodic migration rather than continuous migration given equivalent effective migration (Nm); 2) how time between migration pulses scales with waiting time to speciation; 3) whether periodic gene flow determined by a realistic scenario of glacial cycling leaves a distinct signature in commonly applied summary statistics; and 4) if joint site frequency-based approaches to demographic inference can distinguish divergence with periodic migration from traditional models of speciation with gene flow.

Methods

Analytical model

To evaluate the relative ease of speciation with periodic gene flow compared to speciation with continuous gene flow, we compare waiting times to speciation (average time until reproductive isolation develops between two diverging populations) using simple

Fig. 1. **A simple model of speciation with periodic gene flow**. Parameter values: m=probability an allele represents a migrant in a given generation; α= total time spent in isolation; $1 - \alpha$= total time spent exchanging migrants; n=total number of time periods; λ^1= speciation rate in isolation; λ^2= speciation rate with gene flow.

analytical models. First, we develop a general expectation of average waiting time given two constant speciation rates. Consider a scenario of alternating periods of migration (at a constant rate $m>0$) and isolation ($m = 0$) between populations K_1 and K_2, which we model as n alternating states of equal length τ, characterized by their different expected times to speciation (Figure 1). Let the rate of speciation (i.e., the probability of speciation occurring in a given time period τ) equal the reciprocal of the waiting time to speciation (either T^a, meaning the waiting time to speciation in isolation, or T^p, meaning the waiting time to speciation with recurrent migration), which we denote λ^1 and λ^2,respectively. The mean total expected waiting time to speciation (T^t) is the reciprocal of the weighted sum of the speciation rates under isolation and under migration:

$$T^t = \frac{n}{\sum_{i=1}^{n} \tau\lambda^1 + \tau\lambda^2} \ (1)$$

To explore the behavior of Equation 1 under different regimes of migration and isolation, we input speciation rates determined by Gavrilets' neutral Bateson-Dobzhansky-Muller icompatibility (BDMI) model of the evolution of genetic

incompatibilities on a holey adaptive landscape (Dobzhansky, 1936; Gavrilets, 2003). Full

formulae and justification are provided elsewhere (Gavrilets et al., 1998; Gavrilets, 1999,

2003); a brief description of the model follows:

Consider a panmictic ancestral population of diploid organisms fixed for genotype

$AABB$ without recombination and then split into two completely isolated daughter

populations, K_1 and K_2. In K_1, let A mutate to a and B mutate to b unidirectionally at an

equal rate of μ per generation. The combination of alleles a and B result in a single

genotype is lethal or otherwise precludes hybrid viability; A and b remain compatible.

Speciation is inevitably complete with when the genotype $aabb$ is fixed in the only

population with mutation (by drift alone). We define T as the time to speciation, or

average waiting period in generations to go from the ancestral state to complete

reproductive isolation. In the absence of positive selection or gene flow, i.e., a scenario of

allopatric speciation by mutation and drift alone, the average waiting time to speciation

will be the time to fixation of two neutral alleles (a and b), which is approximately two

times the reciprocal of its mutation rate and unrelated to effective population size (Nei,

1976):

$$T^a = 2 * \frac{1}{\mu} = \frac{2}{\mu} \ (2)$$

We next use Gavrilet's (2003) extension of this model to describe parapatric

speciation, i.e., speciation with limited gene flow ($0<m<0.5$) between two demes. Following

subdivision of K_1 and K_2, a portion m of K_1 each generation is supplied by immigrants

from K_2 with ancestral genotype $AABB$. Again assuming divergence in K_1 is driven only

by neutral mutation at a per generation rate of μ for both alleles, and assuming that m is

much larger than the mutation rate, the average waiting time to speciation is given by:

$$T^p = \left(2 + \frac{m}{\mu}\right)\frac{1}{\mu} \approx \frac{m}{\mu^2} \quad (3)$$

142 Given equal time is spent exchanging migrants as in allopatry, we calculate the

¹⁴³ arithmetic mean of these speciation rates (λ^t) :

$$\lambda^t = \frac{1}{2} \times \frac{\mu}{2} + \frac{1}{2} \times \frac{\mu^2}{m} = \frac{\mu}{4}\left(\frac{m}{m}\right) + \frac{\mu^2}{2m} = \frac{2\mu m + 4\mu^2}{8m} = \frac{2\mu(m + 2\mu)}{8m} \quad (4)$$

¹⁴⁴   The average waiting time across all time periods is given by its reciprocal:

$$T^t = \frac{8m}{2\mu(m + 2\mu)} \quad (5)$$

¹⁴⁵   We now consider speciation with periodic gene flow if n time periods τ are unequal.
¹⁴⁶ Let α equal the proportion of time spent in isolation and $1 - \alpha$ equal the proportion of
¹⁴⁷ time exchanging migrants at a constant rate $m{>}0$, and let the rate of speciation equal the
¹⁴⁸ reciprocal of the expected times to speciation T^a and T^a, given by λ^1 and λ^2. The total
¹⁴⁹ expected waiting time to speciation is again simply the reciprocal of the weighted sum of
¹⁵⁰ the speciation rates:

$$T^t = \frac{n}{\sum_{i=1}^{n} \alpha\lambda^1 + (1 - \alpha)\lambda^2} \quad (6)$$

¹⁵¹   Given Equations 2 and 3, total expected waiting time to speciation can be
¹⁵² calculated by:

$$T^t = \frac{n}{\sum_{i=1}^{n} \alpha\left(\frac{\mu}{2}\right) + (1 - \alpha)\left(\frac{\mu^2}{m}\right)} \quad (7).$$

¹⁵³   *Simulating cyclical speciation*

¹⁵⁴   We assessed the performance of our simple models using forward-time evolutionary
¹⁵⁵ simulations implemented in SLiM v. 3.1 (Haller and Messer, 2019), with diploid genotypes,
¹⁵⁶ constant population size, and non-overlapping generations. All parameters were chosen to
¹⁵⁷ closely approximate the model in Gavrilets (2003). For all simulations, we set a mutation
¹⁵⁸ rate of $1x10^{-4}$ and allowed no recombination. We modeled BDMI loci as two adjacent sites
¹⁵⁹ 1 bp in length; both of the genomic "regions" (BDMI locus A and BDMI locus B) on this

chromosome had a specific mutation type with no fitness effect. We considered speciation

complete when mutations were fixed in both BDMI loci in population K_2, and did not end

the simulation until this occurs; mutations were prevented from occurring on BDMI loci in

population K_1, which solely used as a donor of ancestral alleles in simulations involving

gene flow. Outputting the final generation (i.e., waiting time to speciation) each time, we

ran 100 replicates of all models. To simulate allopatric speciation, we used a single

population of size $n=100$. To simulate parapatric speciation, we established two

populations of size $n=100$ each, and set unidirectional migration from population K_1 to

population K_2. We simulated speciation under periodic gene flow using three different

values of α (the proportion of time spent in allopatry): 0.50, 0.1, and 0.01. We modified the

migration rate parameter m across each value of α to fix the total effective migration (Nm)

across the entire simulation, and initiated periods of allopatry every other generation, every

10 generations, and every 100 generations respectively. We again considered speciation

complete with the fixation of both BDMI loci in K_1. SLiM scripts for all simulations are

available at `https://github.com/elinck/speciation_periodic_migration`.

<center>*Simulating glacial cycles of isolation and contact*</center>

As a distinct goal from our analytic model of speciation and our forward time

simulations to validate it, we were interested in whether cycles of gene flow and isolation

between diverging populations would leave unique genomic signatures given biologically

realistic parameter values. To test this, we simulated population divergence and secondary

contact both 1) mediated by Pleistocene glacial cycles and 2) under a standard

isolation-with-migration model using the coalescent simulator msprime v. 0.7.1 (Kelleher

et al., 2016). We determined the timing and duration of gene flow between populations

using empirical atmospheric CO_2 concentration values from the Vostok Ice Core dataset

(Petit et al., 1999). A collaborative drilling project in East Antarctica, the Vostok Ice

Cores provide direct records of historical variation in atmospheric trace-gas composition

Fig. 2. Atmospheric CO_2 concentration data from the Vostok Ice Cores, color coded by migration rate in msprime simulations of populations diverging with periodic gene flow.

through entrapped air inclusions, and cover four glacial cycles from 417,160 to 2,342 years before present (Petit et al., 1999).

We simulated a $1x10^7$ bp region for two populations of size $n = 1x10^5$, using a recombination rate of $1x10^{-8}$, mutation rate $2.3x10^{-9}$, and allowing 10 migrants per generation (i.e., m=0.0001) when atmospheric CO_2 fell below 250 ppm (Figure 2). This configuration effectively mimics connectivity dynamics among populations of a species distributed across a tropical montane landscape, where cooler temperatures allow the expansion of forest habitat between adjacent peaks. We started and concluded the simulation with periods of isolation, allowing 4 major periods of migration of varying duration. After completing each simulation, we calculated Wright's F_{ST} between populations, and then used the approximation $F_{ST} \approx \frac{1}{4mN\left(\frac{n}{n-1}\right)^2+1}$ (Wang, 2004) to determine a constant migration rate resulting in equivalent equilibrium F_{ST} for a paired simulation under a standard isolation with migration model (i.e., a model with continuous gene flow). We otherwise used identical parameters as our periodic migration simulation. In total, we ran 100 replicates of each scenario. We compared patterns of genetic variation between scenarios by plotting the distribution of standard summary statistics calculated both between and within populations using scikit-allel (Miles et al., 2019) implemented in Python (scripts available at

Fig. 3. Demographic model parameters implemented in moments for A) periodic gene flow and B) constant gene flow (i.e., isolation-with-migration). Nu_1 and Nu_2 are parameters for the effective population size of populations 1 and 2, respectively; m is the symmetrical migration rate during a given time period T_0 through T_7.

). These statistics are:

the number of segregating sites (SNPs); mean pairwise genetic distance (π); Tajima's D;

Watterson's θ; observed heterozygosity; Weir & Cockerham's F_{ST}; D_{XY}, and the average

length and skew of identical-by-state haplotype blocks, a promising source of information

on historical demography and dispersal (Harris and Nielsen, 2013; Ringbauer et al., 2017).

Demographic inference

As speciation processes are often studied using joint site frequency spectrum-based

(JSFS) demographic inference, we used our coalescent simulations to see whether this

family of methods could distinguish between a model of periodic gene flow (PM) and a

standard isolation-with-migration model (IM) using moments v. 1.0.0 (Jouganous et al.,

2017). To model the evolution of allele frequencies under different demographic scenarios,

moments analytically solves for the expected site frequency spectrum using numerical

solutions of an appropriate diffusion approximation of allele frequencies (Jouganous et al.,

2017). We defined two models to describe the demographic history of our simulations: a

periodic migration model with four alternating periods of isolation and migration; and an

isolation with migration model with continuous migration to the present (Figure 3). Both

models featured symmetrical migration and independent effective population sizes

(parameters m, $Nu1$, and $Nu2$). We ran 20 optimizations from random starting

parameters for each model across 100 simulations using the "optimize_log" method. For

each simulation and model we used only the parameter set with the highest likelihood after

optimization for downstream analysis. We then identified the model with the lowest AIC

for each simulation (to correct for the higher number of parameters in periodic migration

models) and examined the distribution of parameter estimates across simulations.

RESULTS

Analytical model

We plot T^t against μ for different values of α in Figure 4, generated by Gavrilet's

models of speciation by BDM incompatibilities. Though simplified, these equations capture

the intuitive relationship between allopatric and parapatric speciation, e.g., that moderate

gene flow erodes differences between diverging populations and results in a longer average

waiting time to speciation. For example, assuming a mutation rate of $\mu = 1x10^{-8}$ and a

migration rate of $m = 0.1$, $T^a = 2x10^8$ generations under a model of allopatric speciation

and $T^p = 1x10^{15}$ in a under a model of parapatric speciation—i.e., parapatric speciation

takes five million times longer. (In the absence of positive selection, speciation is clearly

unlikely in both scenarios.) Notably, speciation is more likely under a model of periodic

gene flow than continuous gene flow given a constant number of migrants across the entire

divergence period for all three values of α. Further, as α approaches $1 - \alpha$ in value, the

Fig. 4. Analytical results for the relationship between BDM loci mutation rate and waiting time to speciation under alternate models of speciation, by proportion of total divergence time spent in allopatry (α).

240   average waiting time to speciation with period migration approaches its value under strict

241   isolation.

242   *Simulating cyclical speciation*

Our simulation of speciation by BDM incompatibilities broadly supported the

predictions of our analytical model. The median time to speciation was lowest in isolation,

highest under constant migration, and intermediate under periodic migration becoming

increasingly longer as the proportion of time spent in allopatry was decreased (Figure 5).

Time to speciation under periodic gene flow is bimodally distributed at all values of $\alpha = 1$,

and had the greatest interquartile range (IQR) when $\alpha = 0.01$ and lowest when $\alpha = 0.01$.

Interestingly, speciation ocurred more rapidly than theoretical expectations in all models,

with median time to speciation ranging from 18.37% to 69.99% of the expected number of

generations predicted by Gavrilets (2003).

Fig. 5. Forward time simulations of the waiting time to speciation by BDM incompatibilities under alternate models of speciation and different proportions of total divergence time spent in allopatry (α) Theoretical expectations of time to speciation from Gavrilets (2003) and our adapted analytical model are overlaid on the data, with the dashed line indicating time to speciation in isolation, the dotted line indicating time to speciation under periodic gene flow, and the solid line indicating time to speciation under continuous gene flow.

²⁵²   *Simulating glacial cycles of isolation and contact*

²⁵³   Despite controlling for equilibrium F_{ST} (which was not significantly differed

²⁵⁴ between periodic and continuous migration scenarios; Wilcox-Mann-Whitney U test,

²⁵⁵ $p = 0.89$), summary statistics could easily distinguish simulations of periodic and

²⁵⁶ continuous migration (Figure 6). The total number of SNPs, π, Watterson's θ, Tajima's D,

²⁵⁷ observed heterozygosity, and D_{XY} were all significantly higher in continuous migration

²⁵⁸ simulations (Wilcox-Mann-Whitney U test, $p < 2.2x10^{-16}$). Identical-by-state haplotype

²⁵⁹ blocks both between and within populations were significantly longer under periodic

²⁶⁰ migration ($p < 2.2x10^{-16}$). The skew of identical-by-state haplotype blocks was greater

²⁶¹ within populations under periodic migration ($p < 2.2x10^{-16}$) and between populations

²⁶² under continuous migration, though distributions were broadly overlapping ($p = 0.04978$).

²⁶³ There was no difference in the distribution of identical-by-state haplotype blocks over

²⁶⁴ 100,000 bp in length ($p > 0.05$).

Fig. 6. The distribution of summary statistics from simulations of periodic gene flow determined by Vostok Ice Core atmospheric CO_2 concentrations ("periodic") and a standard isolation-with-migration model ("continuous").

Demographic inference

Periodic migration models had lower AIC in 72.7% of simulations, with a median relative to continuous-migration models of -178.05 suggesting strong support for the periodic model (Figure 7). In most cases when continuous-migration models were preferred the corresponding periodic-migration likelihood was much lower than the median across simulations, suggesting that optimization had failed to find the global maximum. The top 5 periodic migration models accurately estimated population sizes (Figure S2) and the timing of the end of the last migration period, but appeared to have little power to estimate migration rates or the timing of earlier episodes of migration (Figure 7).

For continuous models, best-fit parameter sets were split between two regions of parameter space – high migration and ancient divergence or low migration and recent divergence (Figure S1). Parameters for the high-migration/ancient divergence regime were similar to the continuous-migration models we simulated to generate equivalent final-generation F_{st}, while the low-migration/recent divergence parameters appeared to

Fig. 7. Left: AIC for continuous and periodic migration models fit to simulations with $m = 10^{-4}$ when $CO_2 < 250ppm$. Grey lines connect the maximum-likelihood parameter set for each simulation across 20 optimizations. Right: Simulated and inferred migration rates for the top 5 periodic migration model fits.

capture events since the end of the last period of migration c. 10kya. In all cases the

low-migration/low-divergence-time regime had higher likelihood.

DISCUSSION

Speciation with periodic gene flow

Since the inception of the field, speciation research has grappled with the question

of whether reproductive isolation can develop while diverging populations exchange

migrants (Darwin, 1859; Wagner, 1873; Mayr, 1963; Smith, 1966; Coyne and Orr, 2004;

Fitzpatrick et al., 2008; Mallet, 2010). The inherent limitations of biogeographic definitions

of speciation have lead to newer, more precise models, based on underlying processes

(Schluter, 2009; Nosil, 2012) and / or the presence or absence of gene flow (Pinho and Hey,

2010). Our study of divergence with periodic gene flow found that genetic isolation is

relatively more likely to occur compared to a model of continuous migration. These results

suggest periodic gene flow between isolated populations may be an underappreciated force

in generating species diversity under realistic conditions, and indicate gene flow-based

₂₉₃ definitions are themselves a broad category containing related but distinct speciation

₂₉₄ processes.

₂₉₅   Our analytical model describes the expected waiting time to speciation given

₂₉₆ alternating periods of isolation and migration of specific durations and specific speciation

₂₉₇ rates (Figure 1; Equations 2 and 6). By using speciation rates determined by Gavrilets'

₂₉₈ models of allopatric and parapatric speciation by BDM incompatibilities (Gavrilets et al.,

₂₉₉ 1998; Gavrilets, 1999, 2003), we demonstrate that speciation with periodic gene flow can

₃₀₀ be nearly as likely (i.e., has a similar expected waiting time) as allopatric speciation when

₃₀₁ an equal amount of time is spent in isolation as exchanging migrants (Figure 4). Further,

₃₀₂ speciation with periodic gene flow remains substantially easier to achieve than speciation

₃₀₃ with constant migration even when as little as 1% of the total period of divergence is spent

₃₀₄ in isolation (Figure 4). This reflects an important property of our model, which is that

₃₀₅ total expected waiting time to speciation is equivalent to the harmonic mean of expected

₃₀₆ waiting times under alternate migration regimes. We thus know the average waiting time

₃₀₇ to speciation given periodic gene flow will always be less than or equal to its value given

₃₀₈ constant migration with equivalent effective migration (Nm). Intuitively, this application

₃₀₉ of the harmonic mean mirrors its use in estimating effective population size in the face of

₃₁₀ recurrent bottlenecks (Vucetich et al., 1997), and highlights the relationship between

₃₁₁ effective population size and effective migration rate ($4Nm$). These parameters together

₃₁₂ shape the time-averaged coalescent rate.

₃₁₃   As it represents a specialized case of speciation in the face of periodic gene flow, we

₃₁₄ highlight several potential limitations of our simple model. First, while our validations

₃₁₅ with forward time simulations appear to accurately reflect the expected relative median

₃₁₆ waiting time to speciation for different modes of speciation and different values of α, they

₃₁₇ are systematically lower than the theoretical predictions of Gavrilets (2003). This

₃₁₈ surprising result defies easy explanation, but we suspect it may result from a combination

₃₁₉ of the limitations of the Wright-Fisher model underlying our simulations and the

approximations underlying the Gavrilets (2003) equations for expected waiting time to speciation. Second, we model alternating periods of isolation and gene flow as independent, "memoryless" units: partial reproductive isolation developing in one time period as one or more BDMI loci drift closer to fixation is unrelated to the probability of reproductive isolation developing in the next period, instead contributing to an average across the entire history of divergence. This might alternately lead to an underestimate (if migration substantially reduces the frequency of a BDM locus allele) or overestimate (if the frequency of the BDM locus allele is little reduced by migration after progressing substantially towards fixation) of waiting time to speciation, a result potential reflected in the bimodal distribution of time to speciation in simulations with periodic gene flow (Figure 5). Third, we ignore the possibility that reinforcement, adaptive introgression, or many other evolutionary processes might interact with modeled dynamics (though these will be discussed below). Finally, we explore only a single model for evolving reproductive isolation between diverging populations. Could other isolating barriers—e.g., the evolution of a phenotype used in mate choice—have unique behavior under periodic gene flow? We encourage others to explore this interesting question with further theory and simulations.

Cycles of isolation and contact leave distinct genomic signatures

Repeated periods of isolation and migration between diverging populations are commonly cited in discussions of the impact of Pleistocene glacial cycles on extant biodiversity (Klicka and Zink, 1997; Avise and Walker, 1998; Klicka and Zink, 1999; Ayoub and Riechert, 2004; Lovette, 2005; He et al., 2019). Pleistocene glacial cycles and their effects on global climate (e.g., forest refugia in a drier Amazon) have been invoked as a mechanism for explaining both extant species richness and phylogeographic diversity in birds (Klicka and Zink, 1997, 1999; Weir and Schluter, 2004; Lovette, 2005; Lamb et al., 2019), plants (Vuilleumier, 1971; Bonaccorso et al., 2006; He et al., 2019), insects (Knowles, 2000; Ayoub and Riechert, 2004), fish (April et al., 2013), and mammals

(Hundertmark et al., 2002). These studies have primarily evaluated a hypothesis of a

Pleistocene effect by comparing molecular clock divergence time estimates between

phylogroups to the timing of the last glacial maximum, sometimes to the point of

controversy (Arbogast and Slowinski, 1998).

Our simulations using empirical atmospheric CO_2 concentration data from the

Vostok Ice Cores provide data that may be useful in more rigorously evaluating hypotheses

of the role of glacial cycles. We were surprised to find that periodic gene flow was readily

distinguishable from continuous gene flow by multiple summary statistics commonly

applied to genomic data (Figure 6). These data suggest that neutral evolutionary processes

have profoundly different effects in two scenarios that are often treated under the umbrella

of "divergence with gene flow." For example, metrics reflecting genetic diversity and

effective population size (e.g., D_{XY}, observed heterozygosity, π, Watterson's θ) are reduced

under periodic migration given equivalent equilibruim F_{ST}, reflecting the increased

strength of genetic drift during periods of isolation when the effective population size of

both lineages is reduced in the absence of migration. For similar reasons, the mean length

of identical-by-state haplotype blocks is greater under periodic migration: the average

population-scaled recombination rate ($\rho = 4Ner$) is reduced across the entire timeline of

divergence, preventing rapid breakdown of uniparentally inherited chromosomal regions.

While our study necessarily reflects a narrow window of parameter space, we believe these

statistics provide a useful starting point for detecting gene flow pulses at deeper time

scales (Harris and Nielsen, 2013).

367   *Inferring speciation with demographic modeling*

368   Joint site frequency spectrum based demographic model testing–conducted by

369 comparing empirical data with the expected JSFS, as calculated by diffusion

370 approximation (Gutenkunst et al., 2009), differential equations (Jouganous et al., 2017), or

371 other approaches (Gronau et al., 2011)—is widely used to evaluate alternate speciation

hypotheses (Hey, 2010; Nosil, 2012; Rougeux et al., 2017). While these studies have provided much of the recent evidence for speciation with gene flow (Butlin et al., 2008; Jónsson et al., 2013; Filatov et al., 2016; Rougeux et al., 2017) and led Nosil to suggest the process might be common (Nosil, 2008), they have typically employed a binary approach, contrasting models of either continuous gene flow or long periods of isolation.

Our finding that a realistic model of periodic gene flow is distinguishable from an isolation with migration model using these approaches suggests parameter-heavy models can in some circumstances be effectively applied to demographic inference. However finding the global optimum for both continuous- and periodic-migration models required starting many independent optimization runs from random starting parameters. In our analysis running 20 optimizations resulted in selecting the "correct" model in just 72% of simulations and in many cases optimization appeared to have trouble locating a global optimum. In addition, parameter estimates from these models were generally accurate for only the most recent cycle of isolation and migration. When standard IM models are fit to populations that have evolved under periodic migration, picking parameter sets with the highest likelihood tends to overestimate contemporary migration rates and underestimate divergence dates—reflecting dynamics since the end of the last period of migration rather than the full population history.

In light of these continued challenges and the reduced time to speciation under periodic gene flow compared to continuous gene flow (Figure 4), we suggest studies claiming that initial population divergence occurred in the face of gene flow argue for its plausible selective driver rather than relying on imprecise estimates of the timing of migration alone (Strasburg and Rieseberg, 2011). However, we highlight that many studies that use "speciation with gene flow" apply it broadly, including cases of secondary contact prior to the development of complete reproductive isolation (Rougeux et al., 2017). While we remain agnostic to its appropriate context, this ambiguity seems to be a continuation of a problem of language usage in speciation research more broadly (Harrison, 2012).

Complexity in speciation and future directions

⁠399 The advent of affordable genome-wide DNA sequence data for nonmodel organisms ⁠400 and concurrent leaps in theory and computational methods have spurred a renaissance in ⁠401 speciation research (Nielsen and Wakeley, 2001; Fitzpatrick et al., 2008; Mallet, 2008; ⁠402 Nosil, 2008; Rougeux et al., 2017). This new work has transformed our understanding of ⁠403 speciation, suggesting it is profoundly shaped by selection and frequently characterized by ⁠404 extensive introgression between diverging lineages (Mallet, 2008; Cui et al., 2013; Toews ⁠405 et al., 2016; Kumar et al., 2017). Our study highlights another dimension of complexity in ⁠406 speciation theory and research, and indicate temporal heterogeneity in rates of gene flow ⁠407 deserves increased scrutiny.

⁠408 Beyond the findings reported in this paper, we believe several aspects of speciation ⁠409 with periodic gene flow provide promising research directions. First, hybridization is ⁠410 increasingly recognized as a generative force in evolution; adaptive introgression can ⁠411 increase fitness across population and species boundaries (Norris et al., 2015; Racimo ⁠412 et al., 2015), and occasionally lead to the evolution of reproductive isolation itself ⁠413 (Gompert et al., 2006; Brelsford et al., 2011; Schumer et al., 2014, 2018; Barrera-Guzmn ⁠414 et al., 2018). Periodic gene flow could provide raw material for these processes, increase the ⁠415 efficacy of selection by boosting Ne, or facilitate reinforcement (Coyne and Orr, 2004), ⁠416 accelerating speciation. Second, while the constancy and timing of gene flow between ⁠417 diverging populations is an important parameter and connects the historical emphasis on ⁠418 biogeography with the population genomic era, the selective pressures driving speciation ⁠419 are likely more useful for the long-term goal of developing general predictions for when and ⁠420 where it will occur (Kumar et al., 2017). Though difficult to identify from genomic data ⁠421 alone, we believe the underlying mechanisms generating reproductive isolation form the ⁠422 most useful framework for classifying modes of speciation, and encourage the development ⁠423 of new methods to identify them (Schluter, 2009). Finally, demographic inference based on ⁠424 the site frequency spectrum will likely remain an important tool in speciation research, but

it is limited by its lack of haplotype information. Machine learning approaches approaches

based on realistic forward-time simulations and multiple summaries of genetic data (e.g.,

Schrider and Kern (2018)) or even raw genotype matrices (Flagel et al., 2019) represent a

largely untapped frontier in our quest to understand the mechanisms underlying biological

diversity.

Acknowledgements

We thank Kameron Harris for his helpful discussions on modelling. Murillo
Rodrigues and Peter Ralph provided help with SLiM, and Andy Kern, Lorenz Hauser, and
John Klicka provided useful feedback on the manuscript. Thanks to Nicholas Bierne for
helpful comments on the first version of the preprint. This research was supported by NSF
Doctoral Dissertation Improvement Grant 1701224 to J. Klicka and E.B.L, and a NDSEG
Fellowship to E.B.L.

Data availability

All code and processed data for this study are available at
https://github.com/elinck/speciation_periodic_migration.

References

Alcala, N. and S. Vuilleumier, 2014. Turnover and accumulation of genetic diversity across
    large time-scale cycles of isolation and connection of populations. Proceedings of the
    Royal Society B: Biological Sciences 281:20141369. URL
    https://royalsocietypublishing.org/doi/10.1098/rspb.2014.1369.

April, J., R. H. Hanner, A.-M. Dion-Côté, and L. Bernatchez, 2013. Glacial cycles as an
    allopatric speciation pump in north-eastern American freshwater fishes. Molecular
    Ecology 22:409–422.

Arbogast, B. S. and J. B. Slowinski, 1998. Pleistocene Speciation and the Mitochondrial
    DNA Clock. Science 282:1955–1955.

Avise, J. C. and D. Walker, 1998. Pleistocene phylogeographic effects on avian populations
    and the speciation process. Proceedings of the Royal Society B: Biological Sciences
    265:457–463.

Ayoub, N. A. and S. E. Riechert, 2004. Molecular evidence for Pleistocene glacial cycles
    driving diversification of a North American desert spider, Agelenopsis aperta. Molecular
    Ecology 13:3453–3465.

Barrera-Guzmn, A. O., A. Aleixo, M. D. Shawkey, and J. T. Weir, 2018. Hybrid speciation
    leads to novel male secondary sexual ornamentation of an Amazonian bird. Proceedings
    of the National Academy of Sciences 115:E218–E225.

Bonaccorso, E., I. Koch, and A. T. Peterson, 2006. Pleistocene fragmentation of Amazon
    species ranges. Diversity and Distributions 12:157–164.

Brelsford, A., B. Milá, and D. E. Irwin, 2011. Hybrid origin of audubon's warbler.
    Molecular Ecology 20:2380–2389.

Butlin, R., J. Galindo, and J. W. Grahame, 2008. Sympatric, parapatric or allopatric: the
    most important way to classify speciation? Philosophical Transactions of the Royal
    Society B: Biological Sciences 363:2997–3007.

Coyne, J. A. and H. A. Orr, 2004. Speciation. Sinauer.

Cui, R., M. Schumer, K. Kruesi, R. Walter, P. Andolfatto, and G. G. Rosenthal, 2013.
    Phylogenomics Reveals Extensive Reticulate Evolution in Xiphophorus Fishes.
    Evolution 67:2166–2179.

Darwin, C., 1859. On the origin of species by means of natural selection, or, the
    preservation of favoured races in the struggle for life. John Murray, London.

Delmore, K. E., S. Hübner, N. C. Kane, R. Schuster, R. L. Andrew, F. Cámara, R. Guigá, and D. E. Irwin, 2015. Genomic analysis of a migratory divide reveals candidate genes for migration and implicates selective sweeps in generating islands of differentiation. Molecular Ecology 24:1873–1888.

Dobzhansky, T., 1936. Studies on Hybrid Sterility. II. Localization of Sterility Factors in Drosophila Pseudoobscura Hybrids. Genetics 21:113–135.

———, 1937. Genetics and the Origin of Species. Columbia University Press.

Duckworth, R. A. and G. A. Semenov, 2017. Hybridization Associated with Cycles of Ecological Succession in a Passerine Bird. The American Naturalist 190:E94–E105.

Ehrendorfer, F., 1959. Differentiation-Hybridization Cycles and Polyploidy in Achillea. Cold Spring Harbor Symposia on Quantitative Biology 24:141–152.

Elton, C. and M. Nicholson, 1942. The Ten-Year Cycle in Numbers of the Lynx in Canada. Journal of Animal Ecology 11:215–244.

Felsenstein, J., 1981. Skepticism Towards Santa Rosalia, or Why Are There so Few Kinds of Animals? Evolution 35:124–138.

Filatov, D. A., O. G. Osborne, and A. S. T. Papadopulos, 2016. Demographic history of speciation in a Senecio altitudinal hybrid zone on Mt. Etna. Molecular Ecology 25:2467–2481.

Fitzpatrick, B. M., J. A. Fordyce, and S. Gavrilets, 2008. What, if anything, is sympatric speciation? Journal of Evolutionary Biology 21:1452–1459.

Flagel, L., Y. Brandvain, and D. R. Schrider, 2019. The Unreasonable Effectiveness of Convolutional Neural Networks in Population Genetic Inference. Molecular Biology and Evolution 36:220–238.

Gavrilets, S., 1999. A Dynamical Theory of Speciation on Holey Adaptive Landscapes. The American Naturalist 154:1–22.

———, 2003. Perspective: Models of Speciation: What Have We Learned in 40 Years? Evolution 57:2197–2215.

Gavrilets, S., H. Li, and M. D. Vose, 1998. Rapid parapatric speciation on holey adaptive landscapes. Proceedings of the Royal Society B: Biological Sciences 265:1483–1489.

Gompert, Z., J. A. Fordyce, M. L. Forister, A. M. Shapiro, and C. C. Nice, 2006. Homoploid Hybrid Speciation in an Extreme Habitat. Science 314:1923–1925.

Gronau, I., M. J. Hubisz, B. Gulko, C. G. Danko, and A. Siepel, 2011. Bayesian inference of ancient human demography from individual genome sequences. Nature Genetics 43:1031–1034.

Gutenkunst, R. N., R. D. Hernandez, S. H. Williamson, and C. D. Bustamante, 2009. Inferring the Joint Demographic History of Multiple Populations from Multidimensional SNP Frequency Data. PLOS Genetics 5:e1000695.

Haller, B. C. and P. W. Messer, 2019. SLiM 3: Forward Genetic Simulations Beyond the Wright-Fisher Model. Molecular Biology and Evolution 36:632–637.

Harris, K. and R. Nielsen, 2013. Inferring Demographic History from a Spectrum of Shared Haplotype Lengths. PLOS Genetics 9:e1003521.

Harrison, R. G., 2012. The Language of Speciation. Evolution 66:3643–3657.

Harvey, M. G., G. F. Seeholzer, B. T. Smith, D. L. Rabosky, A. M. Cuervo, and R. T. Brumfield, 2017. Positive association between population genetic differentiation and speciation rates in New World birds. Proceedings of the National Academy of Sciences 114:6328–6333.

He, Z., X. Li, M. Yang, X. Wang, C. Zhong, N. C. Duke, S. Shi, and C.-I. Wu, 2019. Speciation with gene flow via cycles of isolation and migration: Insights from multiple mangrove taxa. National Science Review 6:275–288.

Hey, J., 2010. Isolation with Migration Models for More Than Two Populations. Molecular Biology and Evolution 27:905–920.

Hey, J. and R. Nielsen, 2007. Integration within the Felsenstein equation for improved Markov chain Monte Carlo methods in population genetics. Proceedings of the National Academy of Sciences 104:2785–2790.

Hundertmark, K. J., G. F. Shields, I. G. Udina, R. T. Bowyer, A. A. Danilkin, and C. C. Schwartz, 2002. Mitochondrial Phylogeography of Moose (Alces alces): Late Pleistocene Divergence and Population Expansion. Molecular Phylogenetics and Evolution 22:375–387.

Irwin, D. E., B. Milá, D. P. L. Toews, A. Brelsford, H. L. Kenyon, A. N. Porter, C. Grossen, K. E. Delmore, M. Alcaide, and J. H. Irwin, 2018. A comparison of genomic islands of differentiation across three young avian species pairs. Molecular Ecology 27:4839–4855.

Jónsson, H., A. Ginolhac, M. Schubert, P. L. F. Johnson, and L. Orlando, 2013. mapDamage2.0: fast approximate Bayesian estimates of ancient DNA damage parameters. Bioinformatics 29:1682–1684.

Jouganous, J., W. Long, A. P. Ragsdale, and S. Gravel, 2017. Inferring the Joint Demographic History of Multiple Populations: Beyond the Diffusion Approximation. Genetics 206:1549–1567.

Kelleher, J., A. M. Etheridge, and G. McVean, 2016. Efficient Coalescent Simulation and Genealogical Analysis for Large Sample Sizes. PLOS Computational Biology 12:e1004842.

Klicka, J. and R. M. Zink, 1997. The Importance of Recent Ice Ages in Speciation: A Failed Paradigm. Science 277:1666–1669.

———, 1999. Pleistocene effects on North American songbird evolution. Proceedings of the Royal Society of London. Series B: Biological Sciences 266:695–700.

Knowles, L. L., 2000. Tests of Pleistocene Speciation in Montane Grasshoppers (genus Melanoplus) from the Sky Islands of Western North America. Evolution 54:1337–1348.

Kumar, V., F. Lammers, T. Bidon, M. Pfenninger, L. Kolter, M. A. Nilsson, and A. Janke, 2017. The evolutionary history of bears is characterized by gene flow across species. Scientific Reports 7:46487.

Lamb, A. M., A. G. d. S. Silva, L. Joseph, P. Sunnucks, and A. Pavlova, 2019. Pleistocene-dated biogeographic barriers drove divergence within the Australo-Papuan region in a sex-specific manner: an example in a widespread Australian songbird. Heredity P. 1.

Levin, S. A. and R. T. Paine, 1974. Disturbance, patch formation, and community structure. Proceedings of the National Academy of Sciences 71:2744–2747.

Linck, E., B. G. Freeman, and J. P. Dumbacher, 2019. Speciation with gene flow across an elevational gradient in New Guinea kingfishers. bioRxiv P. 589044.

Loh, P.-R., M. Lipson, N. Patterson, P. Moorjani, J. K. Pickrell, D. Reich, and B. Berger, 2013. Inferring Admixture Histories of Human Populations Using Linkage Disequilibrium. Genetics 193:1233–1254. URL https://www.genetics.org/content/193/4/1233.

Lovette, I. J., 2005. Glacial cycles and the tempo of avian speciation. Trends in Ecology & Evolution 20:57–59.

Mallet, J., 2008. Hybridization, ecological races and the nature of species: empirical evidence for the ease of speciation. Philosophical Transactions of the Royal Society B: Biological Sciences 363:2971–2986.

———, 2010. Why was Darwins view of species rejected by twentieth century biologists? Biology & Philosophy 25:497–527.

Martin, S. H., K. K. Dasmahapatra, N. J. Nadeau, C. Salazar, J. R. Walters, F. Simpson, M. Blaxter, A. Manica, J. Mallet, and C. D. Jiggins, 2013. Genome-wide evidence for speciation with gene flow in Heliconius butterflies. Genome Research 23:1817–1828.

Mayr, E., 1942. Systematics and the Origin of Species, from the Viewpoint of a Zoologist. Harvard University Press.

———, 1963. Animal species and evolution. Harvard University Press.

Medina, P., B. Thornlow, R. Nielsen, and R. Corbett-Detig, 2018. Estimating the Timing of Multiple Admixture Pulses During Local Ancestry Inference. Genetics 210:1089–1107.

Miles, A., P. Ralph, S. Rae, and R. Pisupati, 2019. scikit-allel: v1.2.0. Doi: 10.5281/zenodo.2652508.

Nei, M., 1976. Mathematical models of speciation and genetic distance. Pp. 723–766, *in* Population Genetics and Ecology. Academic Press, New York.

Nielsen, R. and J. Wakeley, 2001. Distinguishing migration from isolation: a Markov chain Monte Carlo approach. Genetics 158:885–896.

Norris, L. C., B. J. Main, Y. Lee, T. C. Collier, A. Fofana, A. J. Cornel, and G. C. Lanzaro, 2015. Adaptive introgression in an African malaria mosquito coincident with the increased usage of insecticide-treated bed nets. Proceedings of the National Academy of Sciences 112:815–820.

Nosil, P., 2008. Speciation with gene flow could be common. Molecular Ecology 17:2103–2106.

———, 2012. Ecological Speciation. Oxford Series in Ecology and Evolution. Oxford University Press, Oxford, New York.

Otto, S. P. and M. C. Whitlock, 1997. The probability of fixation in populations of changing size. Genetics 146:723–733.

Petit, J. R., J. Jouzel, D. Raynaud, N. I. Barkov, J.-M. Barnola, I. Basile, M. Bender, J. Chappellaz, M. Davis, G. Delaygue, M. Delmotte, V. M. Kotlyakov, M. Legrand, V. Y. Lipenkov, C. Lorius, L. Ppin, C. Ritz, E. Saltzman, and M. Stievenard, 1999. Climate and atmospheric history of the past 420,000 years from the Vostok ice core, Antarctica. Nature 399:429.

Pinho, C. and J. Hey, 2010. Divergence with Gene Flow: Models and Data. Annual Review of Ecology, Evolution, and Systematics 41:215–230.

Price, T., 2008. Speciation in Birds. Roberts and Company.

Provine, W. B., 2004. Ernst Mayr: Genetics and Speciation. Genetics 167:1041–1046.

Racimo, F., S. Sankararaman, R. Nielsen, and E. Huerta-Sánchez, 2015. Evidence for archaic adaptive introgression in humans. Nature Reviews Genetics 16:359–371.

Rattenbury, J. A., 1962. Cyclic Hybridization as a Survival Mechanism in the New Zealand Forest Flora. Evolution 16:348–363.

Ringbauer, H., G. Coop, and N. H. Barton, 2017. Inferring Recent Demography from Isolation by Distance of Long Shared Sequence Blocks. Genetics 205:1335–1351. URL https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5340342/.

Rougeux, C., L. Bernatchez, and P.-A. Gagnaire, 2017. Modeling the Multiple Facets of Speciation-with-Gene-Flow toward Inferring the Divergence History of Lake Whitefish Species Pairs (Coregonus clupeaformis). Genome Biology and Evolution 9:2057–2074.

Roy, K., J. W. Valentine, D. Jablonski, and S. M. Kidwell, 1996. Scales of climatic variability and time averaging in Pleistocene biotas: implications for ecology and evolution. Trends in Ecology & Evolution 11:458–463.

Schluter, D., 2001. Ecology and the origin of species. Trends in Ecology & Evolution 16:372–380.

———, 2009. Evidence for Ecological Speciation and Its Alternative. Science 323:737–741.

Schrider, D. R. and A. D. Kern, 2018. Supervised Machine Learning for Population
Genetics: A New Paradigm. Trends in Genetics 34:301–312.

Schumer, M., G. G. Rosenthal, and P. Andolfatto, 2014. How Common Is Homoploid
Hybrid Speciation? Evolution 68:1553–1560.

———, 2018. What do we mean when we talk about hybrid speciation? Heredity 120:379.

Smith, J. M., 1966. Sympatric Speciation. The American Naturalist 100:637–650.

Sousa, V. and J. Hey, 2013. Understanding the origin of species with genome-scale data:
modelling gene flow. Nature Reviews Genetics 14:404–414.

Strasburg, J. L. and L. H. Rieseberg, 2008. Molecular Demographic History of the Annual
Sunflowers Helianthus Annuus and H. Petiolaris—Large Effective Population Sizes and
Rates of Long-Term Gene Flow. Evolution 62:1936–1950.

———, 2011. Interpreting the estimated timing of migration events between hybridizing
species. Molecular Ecology 20:2353–2366.

Toews, D. L., S. Taylor, R. Vallender, A. Brelsford, B. Butcher, P. Messer, and I. Lovette,
2016. Plumage Genes and Little Else Distinguish the Genomes of Hybridizing Warblers.
Current Biology 26:2313–2318.

Via, S., 2009. Natural selection in action during speciation. Proceedings of the National
Academy of Sciences 106:9939–9946.

Vucetich, J. A., T. A. Waite, and L. Nunney, 1997. Fluctuating Population Size and the
Ratio of Effective to Census Population Size. Evolution 51:2017–2021.

Vuilleumier, B. S., 1971. Pleistocene changes in the fauna and flora of South america.
Science 173:771–780.

Wagner, M., 1873. The Darwinian theory and the law of the migration of organismus:
Translated from the german of Moritz Wagner by James L. Laird. Edward Stanford.

Wang, J., 2004. Application of the One-Migrant-per-Generation Rule to Conservation and
Management. Conservation Biology 18:332–343.

Weir, J. T. and D. Schluter, 2004. Ice sheets promote speciation in boreal birds.
Proceedings of the Royal Society B: Biological Sciences 271:1881–1887.

Fig. S1. A: Best-fit parameters across 20 optimizations for each simulation. Runs with low maximum likelihoods appear to be stuck in a high-migration/high-divergence time region of parameter space. B: Summary of parameter fits for continuous-migration models fit to simulations with periodic migration. Arrows show the true population sizes.

Fig. S2. Summary of parameter fits for periodic-migration models fit to simulations with periodic migration.