Spatial and phylogenetic structure of Alpine stonefly community assemblages across seven habitats using DNA-species

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Abstract

1. Stream ecosystems are spatially heterogeneous, with many different habitat patches distributed within a small area. The influence of this heterogeneity on the biodiversity of benthic insect communities is well documented; however, studies of the role of habitat heterogeneity in species coexistence and community assembly remain limited. Here, we asked how habitat heterogeneity influences spatial structure (beta biodiversity) and phylogenetic structure (evolutionary processes) of benthic stonefly (Plecoptera, Insecta) communities.

2. We sampled 20 sites along two Alpine rivers, including seven habitats in four different reaches (headwaters, meandering, bar-braided floodplain, and lowland spring-fed). We identified 21 morphological species and delineated 52 DNA-species based on sequences from mitochondrial *cox1* and nuclear ITS markers. Using DNA-species, we first analysed the patterns of variation in richness, diversity, and composition by quantifing the contribution of each of the four reaches and seven habitats to the overall DNA-species diversity using an additive partition of species diversity analysis and distance-based redundancy analysis. Using gene-tree phylogenies, we assessed whether environmental filtering led to the co-occurrence of DNA-species using a two-step analysis to find a phylogenetic signal.

3. The four reaches significantly contributed to DNA-species richness; with the meandering section displaying the highest contribution. However, we found that habitats had an effect on DNA-species diversity, where glide, riffle, and pool influenced the spatial structure of stonefly communities possibly due to a high species turnover.

4. Among the habitats, the pool showed significant phylogenetic clustering, suggesting high levels of evolutionary adaptation and strong habitat filtering. This community structure may be caused by long-term stability of the habitat and the similar requirements for co-ocurring species.

5. Our study shows the importance of different habitats on the spatial and phylogenetic structure of stonefly community assemblies and sheds light on the habitat-specific diversity that may help improve conservation practices.

KEYWORDS

DNA-species, habitats, spatial structure, phylogenetic structure, Plecoptera

1. INTRODUCTION

Understanding species diversity patterns and processes governing species coexistence in a community is one of the fundamental research objectives in ecology and biodiversity. The development of effective and suitable biodiversity conservation strategies requires knowledge of the processes driving community variation (Socolar, Gilroy, Kunin, & Edwards, 2016) along temporal and spatial gradients (Anderson et al., 2011). Habitat specialisation is known to influence changes in community composition along temporal and spatial gradients (e.g., Wiens et al., 2010); however, this change depends on the degree of specialisation of the species within a community (Rádková et al. 2014).

Stream ecosystems provide one of the most heterogeneous landscapes, given the dynamic interaction between spatial elements (e.g., topography) and ecological process (e.g., hydrology) (Benda et al., 2004; Tockner & Stanford, 2002). This interaction creates a variety of different habitats throughout the longitudinal (upstream-downstream) gradient of the river, classified as lotic (running water) and lentic (standing water) (Dobson & Frid, 1998; Calow & Petts, 1996; Hauer & Lamberti, 1996). At the biogeographic scale, habitat heterogeneity in a river channel defines environmental patches that impact aquatic community composition and species distribution (Brasil, Da Silva, Batista, Olivera, & Ramos, 2017; Dias-Silva, Cabetter, Juen, & De Marco, 2013); while at the local scale, creates environmental filtering to sort species with similar requirements (Saito, Cianciaruso, Siqueira, Fonseca-Gessner, & Povoine, 2016; Mykra, Heino, &, Muotka, 2007; Webb, Ackerly, McPeek, & Donoghue, 2002). Aquatic insects are abundant, diverse, and broadly distributed (Lancaster & Downes, 2013). Many studies have reported a strong positive relationship between aquatic insects species diversity and spatial habitat heterogeneity (e.g. Astorga et al., 2014; Karaus, Larsen, Guillong, & Tockner, 2013; Arscott, Tockner, & Ward, 2005; Benda et al., 2004; Batista, Buss, Dorville, & Nessimian, 2001), however, these were only based on one spatial scale, where the relationship between assembly process on both biogeographical and local scales remains limited to a few studies. Thus, understanding local trends in biodiversity is crucial to describe the interaction between habitat and species diversity at different scales.

One of the challenges in biodiversity research is that of species delimitation. Due to limitations in taxonomic expertise and incomplete descriptions of taxonomic groups, many species may remain unidentified or unclassified (Bickford et al., 2007), making it difficult to describe local biodiversity. Genetic methods developed over the last decade have helped with species identifications and the clarification of species boundaries of a wide range of taxa, including aquatic insects (e.g., Gamboa & Arrivillaga-Henriquez, 2019; Serrana, Miyake, Gamboa, & Watanabe, 2019; Gattoliatt, Rutschmann, Monaghan, & Sartori, 2018). The Generalized Mixed Yule Coalescent model (GMYC; Pons et al., 2006) is a method commonly used to inditify putative DNA-species, by quantifying the transition between inter- and intra-species branching events on an ultrametric tree (i.e., where the distance from the root of every branch is equal), using molecular markers, such as mitochondrial DNA (Fujisawa &

Barraclough, 2013; Mynott, Webb, & Suter, 2011), or based on the congruency between mitochondrial and nuclear DNA (Rutschmann et al., 2016).

Stream ecologists have reported that local environmental heterogeneity may be the strongest driver of community assembly patterns using putative DNA-species diversity of aquatic insects along the river (Astorga et al., 2014; Baselga et al., 2013; Finn, Bonada, Múrria, & Hughes, 2011). However, the majority of these observations is based on DNA-species diversity changes along the river's longitudinal gradient (Finn, Zamora-Muñoz, Múrria, Sáinz-Bariáin, & Alba-Tercedor, 2013; Gill, Harrington, Kondratieff, Zamudio, Poff, & Funk, 2013; Jackson, Battle, White, Pilgrim, Stein, Miller, & Sweeney, 2013; Finn et al., 2011; Finn & LeRoy Poff, 2011; Hughes, Schmidt, & Finn, 2009), where the influence of other river dimensions (e.g., habitats), has only been described in few species, such as beetles (Ribera & Vogler, 2008) or caddisflies (Marten, Brandle, & Brandl, 2006). Therefore, investigating variations in spatial patterns of aquatic insects community composition using DNA-species in the river channel and their relationship with habitat heterogeneity will help to improve the understanding of the process of community assembly in aquatic ecosystems.

The understanding of the community assembly not only has focused on spatial patterns and composition changes analysis. The phylogenetic signal has been proposed as an additional insight to understand the drivers of community assembly (Violle et al., 2011; Webb et al., 2002). The basis of this approach is to compare ecological traits and phylogenetic structures in communities with those expected under null models (i.e., phylogenetic signal). Environmental filtering is thought to cause related species to co-occur (i.e., phylogenetic clustering) more than what is expected by chance, as closely-related species are assumed to share similar physiological requirements. By contrast, competitive exclusion causes the opposite pattern (phylogenetic overdispersion) as species compete for the same limiting resources (Cavender-Bares & Wilczek, 2003). Although phylogenetic clustering and overdispersion have been observed in aquatic insect community assemblies at spatial scales (Saito, Soininen, Fonseca-Gessner, & Siquiera, 2015a; Saito, Siquiera, & Fonseca-Gessner, 2015b; Saito et al., 2016) or based in specific trait-based approaches (e.g., respiration strategy; Buchwalter, Jenkins, & Curtis, 2002, and anthropogenic pollutants; Martin, Cain, Luoma, & Buchwalter, 2007), the assess of the phylogenetic structure in different habitats have not been explored.

This study aims to investigate the impact of habitat heterogeneity on changes in stonefly community composition in different riverine locations, by assessing the relative role of seven aquatic habitats on the beta biodiversity and phylogenetic signal on stonefly community composition using DNA-based species. Stoneflies (Plecoptera) are aquatic insects that show more complex responses to habitat change than other insects (Bojková, Rádková, Soldán, & Zahrádková, 2014), such as ecological (Lancaster & Downes, 2013) and evolutionary responses (Gamboa & Watanabe, 2019). We hypothesised that species distribution and community assembly are related to specific habitats and that habitat filtering acts on closely-related species with similar physiological requirements. By contrast, high levels of species

diversity are expected to limit similarities in community structure and lead to phylogenetic overdispersion. Using two molecular markers (*cox1* and ITS) to delineate putative species, we aimed to: (1) quantify the contribution of four reaches and seven habitats to the overall DNA-species diversity, (2) determine the influence of habitats on community assembly, and (3) compare the phylogenetic structure of stoneflies communities in each of the seven habitats.

2. METHODS

2.1 Study sites and sampling collection

We selected seven habitat types (Fig. 1), waterfall (vertical descent of watercourse), riffle (high turbulence water flow), glide (low turbulence water flow), pool (flowing water > 1 m deep), wando (small-sized water in the head or tail on a sandy bar; as described by Ishida, Abekura, & Takemon, 2005), side-channel (small-sized water pass flowing on a sandy bar), and isolated pond (isolated still water habitat on the sandy bar) over 20 sampling sites in two Alpine rivers in northeastern Italy. Fifteen sampling sites were located on the Tagliamento River (T01 to T15) and five were located in its major tributary, the Fella River (F01 to F05) (Fig. 2). Sampling was performed across four geomorphologically distinct reaches (4 to 1556 m a.s.l.): constrained headwaters, meandering (zigzagging river movement), bar-braided floodplain (convergent river bankfull width), and lowland spring-fed streams in the floodplain (Doering, Uehlinger, & Tockner, 2013). All seven habitat types were sampled at each sampling site. Habitat diversity per site was measured using Simpsons Diversity index with at least one individual per habitat and sampling site.

We collected qualitative samples of stonefly nymphs using D-frame nets (250 µm mesh), spending one hour per site sampled (total sampling time: 20h). Between one and five replicates per habitat per site were collected (n = 188), where each replicate represented a different random location within each habitat sampled. All samples collected in the field were immediately immersed in >99 % ethanol and rinsed twice with fresh >99 % ethanol after being transported to the laboratory. Morphological species identification was conducted using a stereoscopic microscope (80 X) following the classifications in Consiglio (1980) and Fochetti and Tierno de Figueroa (2008). Field sampling (all sites/habitats) was carried out twice in summer 2009 (10 July to 10 September) and once in early spring 2010 (24 March to 15 April).

2.2 DNA extraction and sequencing analysis

A total of 375 stonefly nymphs were collected (see Results). Genomic DNA was extracted from each nymph using DNeasy Blood and Tissue kits, according to the manufacturer's instructions (Qiagen GmbH, Hilden, Germany). In all individuals, a 658 bp fragment of mitochondrial *cox1* was amplified using LCO-1490 and HCO-2198 primers (Folmer,

Black, Hoeh, Lutz, & Vrijenhoek, 1994) with an annealing temperature of 48 °C and 30 PCR cycles. The internal transcribed spacer (ITS) nuclear marker (average 930 bp) was amplified in a subset of 116 individuals chosen to represent all morphological taxonomic groups using ITS1 and ITS2 primers (McLain, Wesson, Oliver, & Collins, 1995) with an annealing temperature of 58 °C and 40 PCR cycles. PCR products were purified using the QIAquick PCR Purification Kit (Qiagen) and sequenced in both directions using the aforementioned primers. *Cox1* sequences were analysed on a 3500xL automated sequencer (Applied Biosystems) and ITS sequences were analysed commercially by Eurofins – Operon, Tokyo. All sequencing data reported in this study have been deposited into GenBank.

Forward and reverse reads were assembled and edited using CodonCode Aligner v 3.5 (Codon Code Corporation, Dedham, USA). Multiple sequence alignment was then performed using ClustalW (align.genome.jp; Larkin et al., 2007). Highly variable regions of ITS were removed using the Indel Module of SeqFIRE v 1.0.1 (https://omicstools.com/seqfire-tool; Ajawatanawong, Atkinson, Watson-Haigh, MacKenzie, & Baldauf, 2012) with the default settings. This reduced the ITS alignment length from 915-1016 bp to 790 bp. For both markers, identical sequences were collapsed using CleanCollapse v. 1.0.5 (https://sray.med.som.jhmi.edu/SCRoftware/CleanCollapse/). Sequences of mtDNA and nDNA markers were compared to the NCBI nucleotide database using blastn queries (http://blast.ncbi.nlm.nih.gov) to verify species identification (>90 and 80 % similarity for mtDNA and nDNA, respectively).

Genetic diversity for each marker was calculated as the number of polymorphic sites, nucleotide diversity (Kimura 2-parameter model), and the number of haplotypes using DnaSp v. 5.10 (Librado & Rozas, 2009).

2.3 Data analysis

2.3.1 DNA-based species delimitation

Putative DNA-species were delineated using the General Mixed Yule Coalescent approach (GMYC; Fujisawa & Barraclough, 2013). First, ultrametric gene trees of *cox1* and ITS were constructed independently using BEAST v. 1.8.3 (Drummond, Suchard, Xie, & Rambaut, 2012) with a relaxed lognormal clock and a coalescent prior run for 50 million generations. The results were summarised with the Tree Annotator (BEAST package). A GMYC analysis was run using the splits package (Ezard, Fujisawa, & Barraclough, 2014) in R v. 3.3 (R Core Team, 2014). We used the single-threshold GMYC model based on Fujisawa and Barraclough (2013). The maximum likelihood of each GMYC model was tested using the likelihood ratio test against a one-species null model (where the entire tree is considered as a single coalescent). Outputs for *cox1* and ITS were compared and the results were combined to generate the list of DNA-species.

2.3.2 Spatial structure of stonefly community assemblies

DNA-species diversity per reach and per habitat were calculated using the Shannon-Weaver and Simpsons diversity indices using the vegan package (Oksanen et al., 2012) in R. The contribution of each of the four reaches and seven habitat types to overall DNA-species diversity was quantified using an additive partition of species diversity analysis (Lande, 1996). This analysis allows alpha and beta values to be simultaneously calculated using an abundance matrix organized hierarchically among habitats (alpha diversity, α) and reaches (beta diversity, β) (Gering, Crist, & Veech, 2003). Simpsons and Shannon-Weaver diversity were partitioned into alpha and beta diversity to observe whether the overall value was greater than the mean values (Lande, 1996). The statistical significance of each component was tested with a randomisation procedure (10,000 randomisations) using the adipart function of the vegan package in R. We then partitioned beta diversity into turnover (i.e., the replacement of species by other species in different habitats) and nestedness (i.e., species loss or gain between habitats) components (Baselga et al., 2013) using a Sorenson dissimilarity matrix and the betadisper function of the vegan package in R. We used a null model with 10,000 randomisations to test if the results of each component were greater than expected by chance.

2.3.3 Habitat influence on community assembly

We performed a distance-based redundancy analysis (db-RDA; Legendre & Anderson, 1999) to quantify the influence of habitats on spatial community assemblies between sampling sites. We first conducted Principal Coordinates Analysis (PCoA) using a community dissimilarity matrix (the pairwise distance of DNA-species within each sampling site) calculated as the Bray-Curtis index. Then, we used the resulting eigenvalues as the “response” variable in the db-RDA models, while geographical distance and habitat influence were used as explanatory variables. The geographical distance between each pair of sampling site was calculated as the Euclidean distance extracted from the geographical coordinates as proposed by Gamboa and Watanabe (2019) using the Vicenty Ellipsoid package (Kamey, 2013) in R. To analyse the contribution of the different habitats to community assembly we calculated a contribution diversity approach matrix proposed by Lu, Wagner & Chen (2007). This approach calculated a differentiation coefficient to evaluate the distribution of DNA-species diversity in a given habitat within the sampling sites. We obtained the best model by first including all explanatory variables (seven habitats and geographical distance) then running nested models using the vegdist and capscale functions in the vegan package in R. After identifying the best model, we calculated the percentage of explained variance obtained by the db-RDA as suggested by Peres et al. (2006) using the ordistep function in vegan. We performed a permutation test (1000 permutations) and used ANOVA to test each explanatory variable of the db-RDA model for statistical significance using the varpart function in vegan.

We analysed multivariate dispersion as a measure of beta diversity based on a community dissimilarity matrix using DNA-species to further investigate habitat influence on beta diversity. We calculated beta diversity as the distance to group centroid (the homogeneity of a community in a given habitat) based on a Bray-Curtis dissimilarity matrix. The community matrix consisted of DNA-species richness in each sampling site at both rivers. We also analysed an homogeneity of multivariate dispersions using a linear model in the betadisper function in the R package vegan with a permutation test (1000 permutations).

2.3.4 Phylogenetic structure of the stonefly community

The phylogenetic relationship among species was estimated using Maximum-Likelihood (ML) gene trees for *cox1* and ITS separately in PhyML 3.1 (Guindon & Gascuel, 2003), with the default settings under a GTR+I+G model as obtained in jModeltest 3.0 (Posada, 2008). Node support was determined by bootstrapping (10,000 times). We used sequences of Orthoptera sp. as outgroups in the *cox1* (HM381647) and ITS (KT440350) trees based on the reported phylogenetic relationship with stoneflies (Misof et al., 2014). Additional adult specimens (Gamboa & Monaghan, 2015) were added on the *cox1* tree to corroborate nymph species identification.

A congruency index (de Vienne, Giraud, & Martin, 2008) calculated using the web version of MAST (http://max2.ese.u-psud.fr/icong/index.help.html), indicated that *cox1* and ITS tree topologies were congruent (Icong = 1.9, $p<0.05$). Therefore, we concatenated the two markers using the FASconCAT-G perl script (https://github.com/PatrickKueck/FASconCAT-G). Using this matrix, we quantified multisite phylogenetic similarities (i.e., similarity measures comparing more than two sampling sites) as proposed by Leprieur et al. (2012). We quantified multisite phylogenetic turnover, nestedness and phylo-beta diversity index (known as Sorensen similarity index) for every community by calculating the sum of the branch length in a phylogeny connecting all species in a community. We then observed the proportion of shared branch lengths between two sampling sites. We performed the analysis using the phylo.beta.multi function in the R package betapart (Baselga & Orne, 2012). We also performed a two-step analysis to check for a phylogenetic signal in stonefly communities within each habitat. First, we estimated niche conservatism (i.e., closely related species are ecologically similar and thus share similar habitats; Wiens et al., 2010) using Blomberg's K-statistic (Blomberg, Garland, & Ives, 2003) and the phylogenetic independent contrast (PIC) test (Felsenstein, 1985) within each habitat. The Blomberg's K test and PIC were implemented using the phytools package (Revell, 2012) and picante package (Kembel et al., 2010) in R. Both tests were run for 10,000 iterations to obtain the null distribution. If the p-values of the observed versus random variance were <0.05, we interpreted them as evidence of niche conservatism. Second, we calculated the Net Relatedness Index (NRI) and the Nearest Taxon Index (NTI) to measure the degree of phylogenetic clustering at the community level (Webb et al., 2002) within each habitat. NRI measures the phylogenetic dispersion of a community assemblage (community associated with

the same habitat type) by comparing the observed mean pairwise phylogenetic distance between species in a community assemblage to the null model, while NTI measures the phylogenetic dispersion of a community assemblage by comparing the observed mean nearest phylogenetic neighbour distance between species in a community assemblage to the null model. Positive values of NRI and NTI indicate that community assemblages with similar habitat preferences are phylogenetically clustered (more closely related than expected), while negative values indicate phylogenetic overdispersion (more distantly related than expected). The NRI and NTI were calculated using the ses.mpd and ses.mntd functions, respectively, in R package picante, with 1000 replications.

3. RESULTS

We identified a total of 21 morphological species in 12 stonefly genera (Table S1) based on collections of 5-40 individuals (mean = 19) and 1-8 morphological species (mean = 4) from each of the 20 sampling sites (Table 1). There were 184 *cox1* haplotypes from the 375 sequenced individuals and 87 ITS genotypes from the subset of 116 sequenced individuals. Samples were dominated by *Leuctra major* (50 and 40 % of the total haplotypes (*cox1*) and genotypes (ITS), respectively; Table S1). For both markers, the log-likelihood of the GMYC model at the threshold (*cox1* = 2514.495; ITS = 2819.385) was significantly higher than the null model for a single coalescent (*cox1* = 2400.932; ITS = 2701.875), as shown by the likelihood ratio test ($p < 0.001$). Both genes delimited 52 individual putative species, 38-59 species (95% CI) for *cox1* and 40-64 species (95% CI) for ITS (Table S1). The *cox1* gene was composed of 42 clusters and ten singletons, while ITS was composed of 31 clusters and 21 singletons. Intraspecific nucleotide diversity for *cox1* ranged from 0-13 % among putative DNA-species, being the highest value for *L. major* (Table S1). Fourteen of the 21 putative morpho-species were split into multiple DNA-species. All DNA-species occurred in more than one habitat, sampling site, and reach.

3.1 Spatial structure of stonefly community assemblies

Among the four reaches, the meandering contained the highest number of DNA-species (n = 52) (Table 1) and the highest Shannon-Weaver (0.89) and Simpson (0.6) diversities (Table S2). The single most species-rich site was T08 in the meandering section (n = 15 DNA-species) (Table 1). The headwater section contained the highest number of cryptic species (25 of 31 total cryptic DNA-species). Among the seven habitats, riffles contained the highest number of DNA-species (n = 31) followed by glides (n = 25) (Table 2 and S2). Riffles was the most species-rich habitat in the meandering section.

The relative contribution of the four reaches and seven habitats to overall DNA-species diversity showed that reaches were the most important driver for richness (57 %, $p < 0.001$, Fig. 3), while habitats were the most important driver for species diversity (Shannon-Weaver

index; 88 %, $p < 0.001$). DNA-species turnover among habitats showed a significantly high species turnover (0.60) than species nestedness (0.16) to overall beta diversity (0.79) ($p = 0.0016$).

Spatial variation in DNA-species was only found to be significant in three habitats: riffle, glide, and pool ($p<0.05$, Fig. 4). Among the seven habitats, the riffle and glide had the highest diversity dissimilarity ($F = 1.43$, $p = 0$, Fig. 5), suggesting that the community in these two habitats were less similar among each other that in the remaining habitats.

3.2 Phylogenetic structure of stonefly community

The mtDNA phylogeny recovered all recognised morphospecies as clades; however, 11 morphospecies were composed of multiple lineages. These lineages likely reflect cryptic species, concurring with GMYC results. The nuclear ITS tree showed congruence with the *cox1* tree, except those of *Nemoura cinerea* and *N. mortini* which were located in the same clade as *Capnia nigra* and *C. vidua* (Fig. S1).

The stonefly community phylogenetic diversity among sampling sites showed that species turnover (0.927) contributed more than nestedness (0.055) to beta diversity (0.98) ($p < 0.005$). Pool habitats showed significant niche conservatism and phylogenetic clustering based on the two-step analysis of phylogenetic signal (Blomberg's K test, PIC, NRI, and NTI, all $p < 0.05$; Table 2), suggesting high similarity among closely-related species within this habitat. Additionally, significant negative values for NRI and NTI indices were reported ($p < 0.05$, Table 2) in the wando and side channel for NRI, and riffle for NTI, indicating that the community assemblage presents phylogenetic overdispersion.

4. DISCUSSION

Community assemblies within the stream are strongly related to habitat heterogeneity within a given area (Astorga et al., 2014). We observed that different habitat types influenced Alpine stoneflies DNA-species diversity, their spatial structure, and their evolution.

In this study, the habitats were the most important driver of DNA-species diversity (diversity partition analysis, Shannon index), concurring with previous studies of aquatic invertebrates (Astorga et al., 2014; Finn & LeRoy Poff, 2011; Finn, Theobald, Black, & Poff, 2006; Finn et al., 2011; Ribera & Vogler, 2008; Marten et al., 2006). Other than niche heterogeneity, one of the main determinants of beta diversity may be the dispersal ability of the organisms (Heino et al., 2015; Astorga et al., 2014; Finn & LeRoy Poff, 2011). A high species turnover is associated with poor dispersal ability (Thompson & Townsend, 2006). Our results showed a high species turnover among habitats (species turnover = 0.60), suggesting that non-random migration is likely to occur among habitats, probably caused by environmental filtering or spatial restrictions in their original habitat. Stoneflies have relatively poor dispersal abilities

within stream reaches (Consiglio, 1980; Fochetti & Tierno de Figueroa, 2008), and tend to avoid migration among habitats due to predator pressure (Tiziano, Fenoglio, Lopez-Rodriguez, Tierno de Figueroa, Grenna, & Cucco, 2010). The habitat heterogeneity along the Tagliamento and Fella rivers could potentially reduce the intra-stream stonefly dispersal, and might explain the large number of cryptic species found (GMYC and phylogenetic analyses). However, high species turnover is also associated with sampling bias. Non-common species among habitats could be a consequence of an un-equal sampling effort or habitat availability. In this study, habitat availability was similar across most sampling sites (average = 0.75, range = 0.55-0.84, Table 1) and despite a similar sampling effort at every site, some species were not detected. Therefore, future studies will need to increase sampling effort to estimate if sampling bias could impact our interpretations of species turnover.

Among the seven habitats, the riffle, glide, and pool had significant impacts on the stonefly community structure (according to the db-RDA analysis) and two habitats (riffle and glide) has significantly higher species diversity (according to the multivariate dispersion analysis). The riffle and glide habitats characterised by a high and low turbulence flow, respectively; both play an essential role in determining habitat suitability for many species of aquatic insects (Lancaster & Downes, 2013; Benda et al., 2004), particulary stoneflies (Lancaster & Downes, 2013; Batista et al., 2001). The dynamic nature of riffle and glide habitats influence density-dependent local competition and high organisms migration with their turbulent flow, leading to high levels of biodiversity (Hughes et al., 2009; Arscott et al., 2005; Batista et al., 2001). On the contrary, the pool (a deep-high river section with a slow/inexistent flow) is only weekly correlated with aquatic insect community diversity (Herrera-Vasquez, 2008) due to the specialised taxa living in this stable habitat (Pastuchova, Lehotsky, & Greeskova, 2008).

Surprisingly, we also found that the pool habitat led to a phylogenetic clustering of the stonefly community, according to the four indices used in this study (Blomberg's K, PIC, NRI, NTI). Long-term adaptation (Lososova et al., 2015), dispersal limitations (Saito et al., 2015a, 2015b), colonisation history (Aizen, Gleiser, Sabatino, Gilarranz, Bascompte, & Verdu, 2015), and low resource competition (Webb et al., 2002) are all associated with phylogenetic clustering, particulary in habitats linked with long-term stability. Pools are rarely affected by physical perturbations (Buffington, Lisle, Woodsmith, & Hilton, 2002), and they may persist despite high river flow (Calow & Petts, 1996), discharge (Rolls, Leigh, & Sheldon, 2012) and drought (Lake, 2003). Therefore, habitats with low levels of perturbation enhanced niche conservatism, clustering, and evolutionary adaptation of stonefly communities in Alpine rivers. However, a greater competitive asymmetry (i.e., an unequal resource division) among distant phylogenetic relatives and facilitation among close phylogenetic relatives can also cause phylogenetic clustering (Mayfield & Levine, 2010; Sargent & Ackerly, 2008). Future studies should address this concern by studying the distribution of resources (i.e., food) among habitats, to obtain a clear picture of the evolutionary of co-occurring species.

In riffle, wando and side channel habitats, a phylogenetic overdispersion pattern was detected, which is commonly observed in aquatic insects (Saito et al., 2016; Violle et al., 2011).

Efficient colonisation and high dispersal patterns are thought to be driving factors for the phylogenetic overdispersion of several species groups, such as birds (Sobral & Cianciaruso, 2016), mammals (Cardillo, Gittleman, & Purvis, 2008) and insects (Violle et al., 2011). These characteristics have been rarely documented in stoneflies (e.g., *Leutra ferruginea*, Macneale, Peckarsky, & Likens, 2005), however, stonefly nymph dispersal is driven by intra-stream drift (Stewart & Stark, 2008). Phylogenetic overdispersion is common in highly-disturbed habitats which tend to preserve high phylogenetic diversity (Xiu et al., 2017). Habitats with high-turbulent water flow (riffle) and high physical perturbation such as sandy bar habitats (wando and side-channel) in the Tagliamento river show a 62 % habitat turnover (Tockner et al., 2003) mainly driven by two annual flooding events (Ward et al., 1999). Therefore, the phylogenetic overdispersion of stonefly communities at riffle, wando, and side-channel habitats is likely driven by the high level of disturbance in these habitats.

Overall, the spatial and the phylogenetic structure of stonefly community assemblies in the Tagliamento and Fella rivers were promoted by the availability and stability of habitats. Among the four reaches, the meandering harboured the largest number of DNA-species (n = 52) and was highly diverse among habitats (Shannon-Weaver index: 0.89). Meanderings are highly complex morphodynamic sections of the river supporting high benthic invertebrate diversity, given the suitability and resilience of their habitats within fluvial ecosystems (Garcia, Schnauder, & Push, 2012). However, the headwaters exhibited a high number of cryptic species (78% of the total DNA-species richness), as previously shown by Jackson et al., 2013; Murria, Bonada, Arnedo, Prat, & Vogler, 2013; Finn et al., 2011; Hughes et al., 2009. This tendency is due to lower canalisation, loss of connectivity with the stream network, and low levels of anthropogenic disturbance (Finn & LeRoy Poff, 2011; Finn et al., 2011). Cryptic species are often observed in stoneflies (e.g., Viteck, Vincon, Graf, & Pauls, 2017), possibly due to introgression (Boumans, Hogner, Brittain, & Johnsen, 2017) or hybridisation (Young, Smith, Pilgrim, Fairchlid, & Schwartz, 2019) which remain unresolved species. Headwaters in the Tagliamento and Fella rivers have narrow valleys and gorges with low sinuosity and confinement (Tockner et al., 2003); therefore, this reach enhances intraspecific genetic differences among stoneflies species. Both meandering and headwater reaches strongly influenced the maintenance of stonefly biodiversity in alpine rivers; therefore, their environmental protection should be a priority in achieving conservation practices.

In conclusion, we demonstrated that different habitats played an essential role in the spatial and phylogenetic structure of stonefly community assemblies in alpine regions. Community diversity was linked to turbulent water flow, while evolutionary adaptation was linked with habitats with allow long-term stability. Our results highlight the importance of molecular studies on quantitative measures of biodiversity in given area and the essential role of habitats in a region, which is particulary important to conserve community assemblages with threatened species or within threatened habitats/landscapes (Esteban & Finlay, 2010).

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available form the corresponding author upon reasonable request.

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TABLE 1 Sampling site characteristics, along with morphological and DNA-species richness in the Tagliamento (sites T) and Fella (sites F) rivers.

Reaches	Sampling site	Altitude (m)	Habitat Simpsons diversity	Morpho-species	DNA species	Haplotypes
Headwater	T01	1174	0.77	2	3	3
	T02	1064	0.77	3	4	6
	T04	1556	0.76	3	5	5
	T06	1346	0.78	8	10	10
	F01	1042	0.80	4	9	9
	F03	978	0.80	3	4	15
Meandering	T05	486	0.76	6	10	12
	T08	625	0.81	5	15	12
	T13	183	0.80	5	10	13
	T15	5	0.81	2	5	2
	F02	769	0.80	4	10	13
	F04	488	0.76	2	2	2
Bar-braided (floodplain)	T03	661	0.81	6	10	13
	T07	524	0.78	3	6	10
	T09	393	0.76	3	11	11
	T10	287	0.78	5	8	13
	T12	131	0.84	5	9	16
	F05	298	0.82	1	1	1
Spring-fed	T11	170	0.55	4	11	15
	T14	4	0.55	1	1	3

TABLE 2 Phylogenetic signal of stonefly communities in the seven habitats sampled in this study. Bold values indicate statistical significance ($p<0.05$). Positive values indicate phylogenetic clustering and negative values indicate overdispersion. N = total number of DNA-species per habitat, K = Blomberg's K, PIC = phylogenetic independent contrasts, NRI = net relatedness index, NTI = nearest taxon index

	N	**K**	**PIC**	**NRI**	**NTI**
Waterfall	21	2.30	2.41	-1.29	-0.79
Riffle	31	2.30	41.73	1.52	**-1.85**
Glide	25	2.71	26.47	0.11	-0.87
Pool	10	**4.03**	**76.80**	**2.53**	**1.79**
Wando	6	1.73	10.89	**-6.44**	-1.26
Side channel	4	1.25	64.83	**-2.06**	-1.06
Isolated Pond	17	2.09	16.59	0.15	-0.82

Figure caption

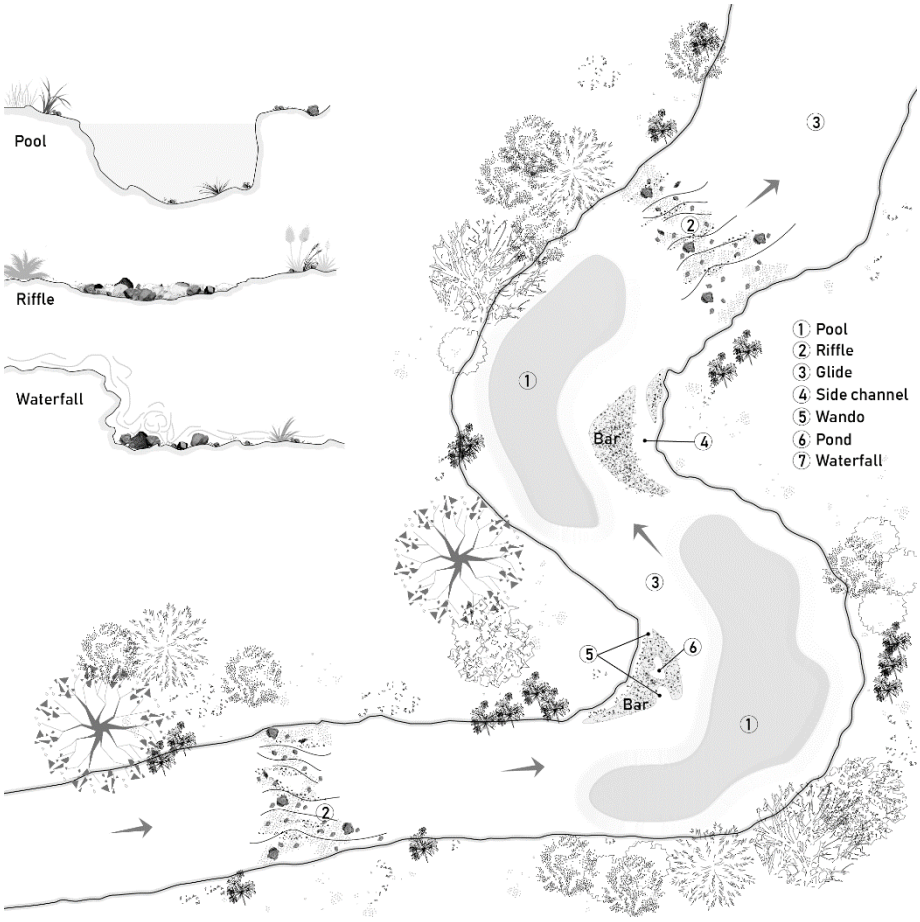
FIGURE 1 Habitats types sampled at each sampling site (Fig. 2) along the Tagliamento and Fella rivers. The arrows indicate the fow direction

FIGURE 2 Sampling sites along the Tagliamento and Fella rivers. The rivers are located in the northeast region of Italy and sampling sites were coded using T1-15 (Tagliamento) and F1-05 (Fella)

FIGURE 3 Additive diversity partition of species richness and Shannon-Wiener diversity (based on DNA-species) into observed and expected hierarchical components of reaches (light grey) and habitats (dark grey). Both partition analyses were significant (<0.001) based on 10,000 randomizations. Note that the plots have different scales

FIGURE 4 Distance based-redundancy analysis (dbRDA) of stonefly DNA-species among 20 sampling sites displaying the influence of habitats on community structure. T = Tagliamento river, F = Fella river. Three habitats (pool, glide, and riffle) explained the highest proportion of the total variation

FIGURE 5 Beta diversity of stonefly communities in each of the seven habitats, based on the distance of the values for each habitat to their centroid (community homogeneity in a given habitat, see methods section 2.3.3). The bold line represents the median, while the bottom and top of the box represent lower and upper quartiles, respectively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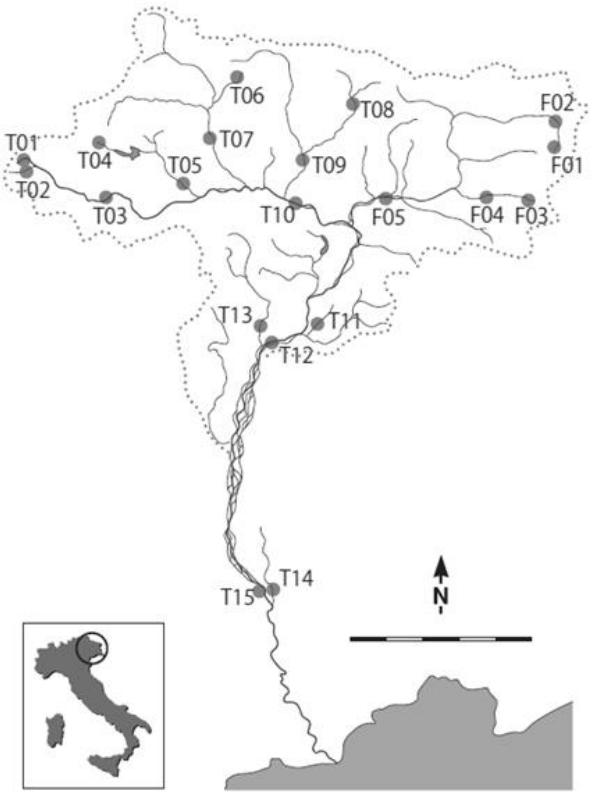
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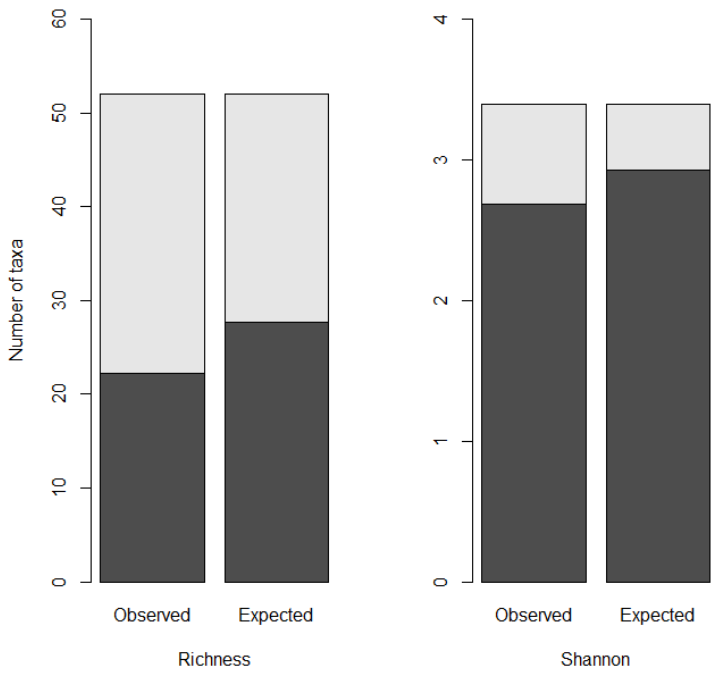
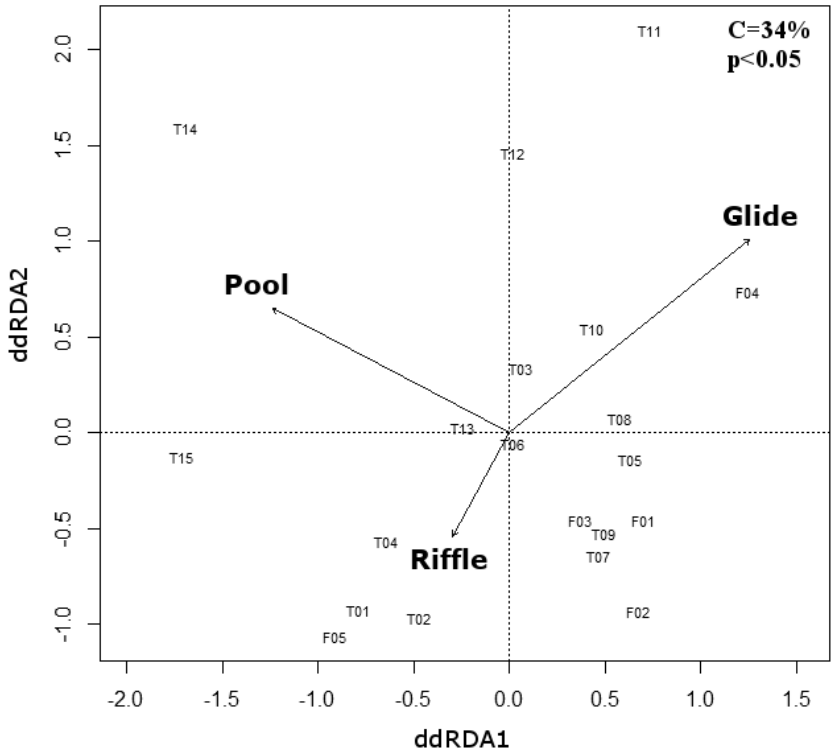
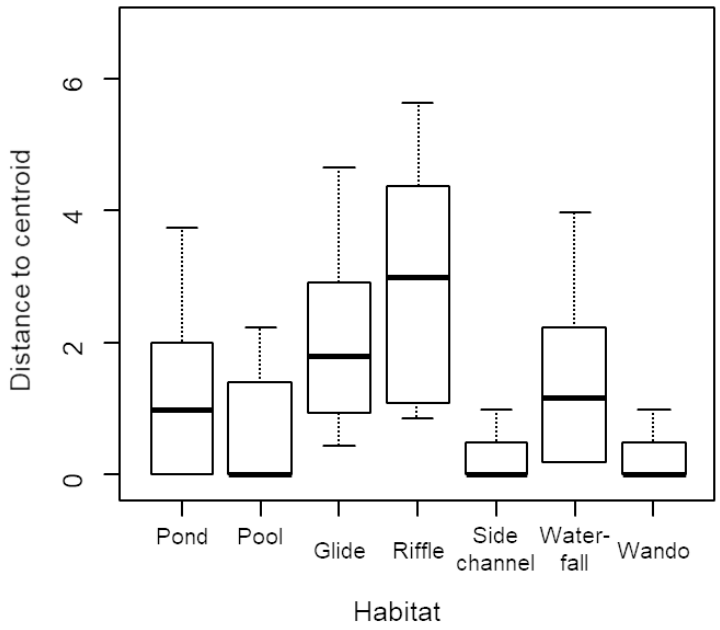
F05
N


Number of taxa
0
10
20
30
40
50
60
Observed
Expected
Richness
0
1
2
3
4
Observed
Expected
Shannon


C=34%
p<0.05
Glide
Pool
Riffle
T11
T14
T12
F04
T10
T03
T08
T13
T06
T15
T05
F03
F01
T09
T04
T07
T01
T02
F02
F05
ddRDA2
ddRDA1
-2.0
-1.5
-1.0
-0.5
0.0
0.5
1.0
1.5
2.0


Distance to centroid
0
2
4
6
Pond
Pool
Glide
Riffle
Side channel
Water-fall
Wando
Habitat

794

Supplementary Material

TABLE S1 DNA-species list for all sampling sites

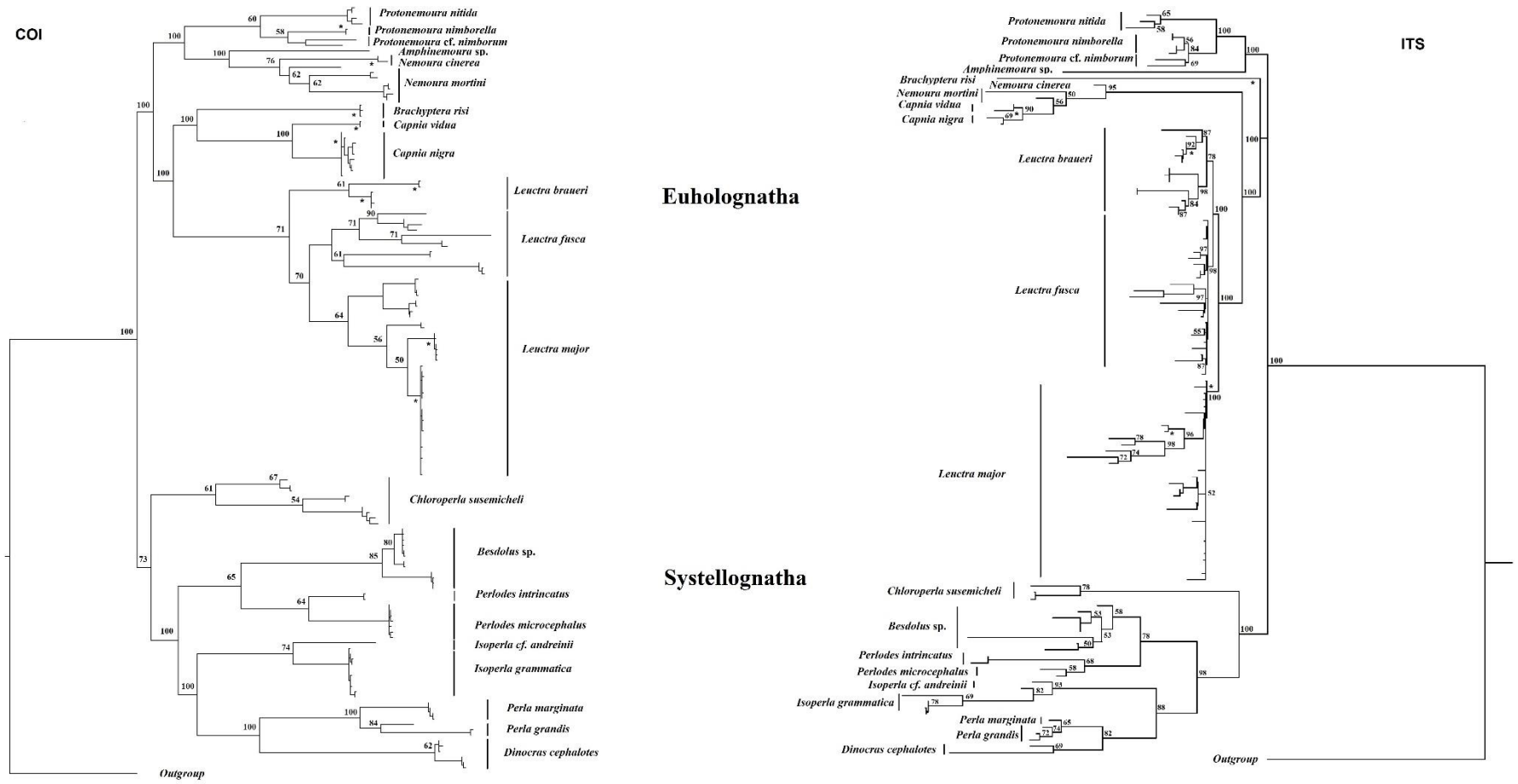
Morphospecies	Total n	DNA-species	Haplotypes	Present sampling sites	Present habitats	Genetic Diversity (%)	Altitude range (m)
Systellognatha							
Perlodidae							
Isoperla cf. *andreinii*	2	1	1	1	1	0	1346
Isoperla grammatica	15	2	9	5	4	2	486-1346
Besdolus sp.	28	3	14	7	5	3.5	183-1556
Perlodes microcephalus	16	2	10	6	2	2	287-769
Perlodes intrincatus	4	1	3	3	2	0	287-625
Perlidae							
Dinocras cephalotes	13	2	8	4	2	2.6	4-183
Perla marginata	24	2	15	6	5	6.5	131-661
Perla grandis	4	2	3	2	1	3	183-625
Chloroperlidae							
Chloroperla susemicheli	13	5	9	2	5	9.5	287-661
Euholognatha							
Capniidae							
Capnia nigra	17	3	9	2	2	7	131-170
Capnia vidua	8	1	2	1	2	0	170
Nemouroidae							
Nemoura mortini	8	3	5	4	4	8.2	486-1346
Nemoura cinerea	10	1	2	1	2	0	1346
Amphinemoura sp.	2	1	1	1	1	0	524
Protonemoura cf. *nimborum*	3	2	3	3	4	5.2	393-1174
Protonemoura nimborella	7	1	1	1	1	0	1556
Protonemoura nitida	6	2	4	1	2	1.3	1346
Leuctricidae							
Leuctra fusca	25	7	12	6	3	3.7	131-287
Leuctra braueri	10	2	6	2	3	4.2	978-1346
Leuctra major	91	8	64	18	7	13.1	5-1556
Taenioterygidae							
Brachyptera risi	6	1	3	1	2	0	131

TABLE S2 DNA-species richness among habitats per sampling site

Reaches	Sampling sites	Waterfall	Riffle	Glide	Pool	Wando	Side-channel	Isolated Pond	Shannon diversity	Simpson diversity
Headwater	T01	0	0	3	0	0	0	0	0.048602	7.68E-05
	T02	4	2	1	0	0	0	0	0.092222	0.000538
	T04	3	3	1	0	0	0	3	0.119007	0.001152
	T06	4	10	4	0	0	1	2	0.19427	0.005376
	F01	13	7	1	0	2	0	3	0.220693	0.008321
	F03	0	5	7	0	0	0	0	0.134995	0.00169
Meandering	T05	3	4	2	3	0	0	4	0.163554	0.003072
	T08	1	17	1	0	6	0	2	0.225542	0.008986
	T13	5	6	8	0	2	1	3	0.215707	0.00768
	T15	0	0	5	1	0	0	0	0.082351	0.000384
	F02	1	11	1	0	3	0	1	0.170096	0.003482
	F04	0	1	0	1	0	0	0	0.035297	2.56E-05
Bar-braided (floodplain)	T03	6	8	1	2	0	1	3	0.19427	0.005376
	T07	0	2	4	1	1	0	1	0.110493	0.000922
	T09	2	7	7	0	0	0	0	0.163554	0.003072
	T10	2	4	4	2	0	0	6	0.176427	0.003917
	T12	1	6	8	3	0	1	1	0.188504	0.004864
	F05	0	0	0	0	0	2	0	0.035297	2.56E-05
Spring-fed	T11	2	5	4	8	0	0	0	0.18256	0.004378
	T14	0	1	0	2	0	0	0	0.048602	7.68E-05
	Total	47	99	62	23	14	6	29	2.802043	0.936585

List of supplementary figures

FIGURE S1 Maximum likelihood phylogenetic trees of stonefly species located along theTagliamento and Fella rivers. The *cox1* (mtDNA) tree is shown on the left and ITS (nDNA) on the right. The numbers at each node indicate the likelihood bootstrap support values. Asterisks show the adult specimen position. A subdivision in two morphological groups was observed, Euholognatha and Systellogna


COI
Protonemoura nitida
Protonemoura nimborella
Protonemoura cf. nimborum
Amphinemoura sp.
Nemoura cinerea
Nemoura mortini
Brachyptera risi
Capnia vidua
Capnia nigra
Leuctra braueri
Leuctra fusca
Leuctra major
Chloroperla susemicheli
Besdolus sp.
Perlodes intrincatus
Perlodes microcephalus
Isoperla cf. andreinii
Isoperla grammatica
Perla marginata
Perla grandis
Dinocras cephalotes
Outgroup
Euholognatha
Systellognatha
ITS
Protonemoura nitida
Protonemoura nimborella
Protonemoura cf. nimborum
Amphinemoura sp.
Brachyptera risi
Nemoura mortini
Nemoura cinerea
Capnia vidua
Capnia nigra
Leuctra braueri
Leuctra fusca
Leuctra major
Chloroperla susemicheli
Besdolus sp.
Perlodes intrincatus
Perlodes microcephalus
Isoperla cf. andreinii
Isoperla grammatica
Perla marginata
Perla grandis
Dinocras cephalotes
Outgroup

809