Systematic Pan-Cancer Analysis Reveals the Functional Roles of Ferroptosis Across Cancers

Ze-Kun Liu[1, #], Qi Zhao[1, #], Zhi-Xiang Zuo[1, #], Kai Yu[1, #], Qing-Feng Zhang[1], Xiao-Long Zhang[1], Hui Sheng[1], Huai-Qiang Ju[1], Rui-Hua Xu[1, *], Ze-Xian Liu[1, *]

[1]State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Sun Yat-sen University Cancer Center, Guangzhou 510060, China

[#]These authors contributed equally to this work.

[*]To whom correspondence should be addressed:

Dr. Ze-Xian Liu as the Lead Contact: liuzx@sysucc.org.cn.

Dr. Rui-Hua Xu: xurh@sysucc.org.cn

Abstract

Ferroptosis is a newly discovered type of cell death that is closely related to cancer, however, the characteristics of ferroptosis and its regulators in cancers are still largely unknown. Based on the cancer omics data in The Cancer Genome Atlas, we comprehensively analyzed the alternations of ferroptosis regulator genes (FRGs) and computationally established the ferroptosis potential index (FPI) to reveal the functional roles of ferroptosis in cancer. The results showed that the mutation rates of FRGs were generally low, but most FRGs were differentially expressed in tumors among various cancer types. Frequent copy number alterations (CNA) and differential DNA methylation of FRGs contributed greatly to their aberrant expression. In most cancers, the FPIs were higher in tumors than in the adjacent normal tissues and were associated with histological types, molecular subtypes, and immunophenotype, etc. Furthermore, the FPIs could predict patient survival and were correlated with cancer drug sensitivity in cancer cell lines, and there were intensive interactions between FRGs and clinically actionable genes. Taken together, our study presents a systematical analysis of ferroptosis and its regulator genes across cancers, and highlights the potential of ferroptosis-based cancer therapy.

Keywords

ferroptosis; pan-cancer analysis; phenotype; survival; drug sensitivity

Introduction

As a newly discovered type of programmed cell death, ferroptosis results from the accumulation of iron-dependent lipid hydroperoxides and leads to cytological changes, for which the features and mechanisms are different compared to typical cell death such as apoptosis (Dixon et al., 2012). Previous studies demonstrated that during the process of ferroptosis, sufficient and approachable cellular iron was required (Dixon et al., 2012). Inhibition of system X_C^-, which is a membrane Na^+-dependent cysteine-glutamate exchange transporter, and GPX4 can disrupt the oxidation-reduction balance and cause overwhelming lipid peroxidation that ultimately results in cell death (Stockwell et al., 2017; Yang et al., 2014; Yu et al., 2017). The initiation and execution of ferroptosis are affected by multiple factors including amino acids, lipids, and iron metabolism, and regulated by various signaling pathways such as amino acid and glutathione metabolism, lipid metabolism, iron metabolism, and mevalonate pathway (Stockwell et al., 2017).

As the understanding of ferroptosis has increased, its complex biological function has been revealed (Matsushita et al., 2015; Yang and Stockwell, 2016). Furthermore, ferroptosis has been found to be closely related with various human diseases including periventricular leukomalacia, Huntington's disease and acute kidney injury (Friedmann Angeli et al., 2014; Inder et al., 2002; Linkermann et al., 2014; Skouta et al., 2014). Besides, the literatures have confirmed that ferroptosis could suppress tumor growth and kill tumor cells (Yu et al., 2017), and then play an important role in cancers such as renal cell carcinomas and liver cancer (Sun et al., 2016b; Yang et al., 2014). For example, erastin-induced ferroptosis decreases the growth of tumors formed from human CRC cells (Xie et al., 2017), and ductal pancreatic cancer cells with a mutant *KRAS* gene were more susceptible to ferroptosis than wild type (Eling et al., 2015; Yu et al., 2017). Conversely, inducing tumor cell ferroptosis by small molecules has become an important strategy for the treatment of many tumors, such as hepatocellular carcinoma, kidney cancer, and pancreatic cancer (Eling et al., 2015; Louandre et al., 2013; Louandre et al., 2015; Yang et al., 2014). In addition, changes in the gene expression of tumor cells also affect ferroptosis, and a number of genes have been confirmed to regulate ferroptosis. For example, *ACSL4* participates in the biosynthesis and remodeling of polyunsaturated fatty acid-Pes, and the downregulation of *ACSL4* increases the resistance to ferroptosis (Dixon et al., 2015; Doll et al., 2017). Furthermore, ferroptosis and its regulator genes (FRGs) were identified to be correlated with drug resistance (Lu et al., 2017). For example, it was previously reported that liver

cancer cells repressed ferroptosis by regulating the expression of *NRF2* or *MT-1G*, which promoted sorafenib resistance *in vitro* and in tumor xenograft models (Sun et al., 2016a; Sun et al., 2016b). Recently, Wang *et al*. reported that CD8+ T cells could promote tumor ferroptosis during cancer immunotherapy treatment (Wang et al., 2019). Thus, ferroptosis might play important roles during cancer progression and treatment, and a systematic study for ferroptosis and its aberrances across cancers will be helpful.

In the present study, for the first time, we performed a comprehensive analysis of genomic variations and expression profiles of the FRGs across 20 cancer types. Furthermore, we computationally modelled the ferroptosis level based on FRGs expression, and dissected the relations between ferroptosis and cancer clinical features. It was found that ferroptosis was associated with various cancer hallmarks, immune microenvironment, drug resistance and patient survival. These results highlighted the critical roles of ferroptosis in cancer and should be helpful for further investigations of ferroptosis-related molecular mechanisms and therapy development.

Materials and Methods

Datasets and Source

The mRNA expression data, copy number alteration thresholded data, masked copy number segmentation data, and DNA methylation 450K data of twenty cancers which had both tumors and normal samples were downloaded from Firehose (http://gdac.broadinstitute.org). Mutation data, miRNA-seq data, and clinical data were downloaded from the Xena Browser (https://xenabrowser.net/datapages/). Additional gene-centric RMA-normalized gene expression profiles and drug response data of over 1000 cancer cell lines were accessed from the Genomics of Drug Sensitivity in Cancer (GDSC) database (https://www.cancerrxgene.org/downloads) (Yang et al., 2013). Immune associated data, including immune cell type fractions and immunophenoscore were obtained from TCIA (https://tcia.at/home) (Charoentong et al., 2017). Integrated protein-protein interaction data was obtained from the Human Protein Reference Database (http://www.hprd.org/) and BioGRID (https://thebiogrid.org) (Keshava Prasad et al., 2009; Peri et al., 2003). We obtained clinically actionable genes (CAGs) from a previous study (Van Allen et al., 2014) (https://software.broadinstitute.org/cancer/cga/target).

Differential expression analysis of mRNA

To test genes differentially expressed in tumor and normal tissue, gene expression data for 20502 genes across 20 cancer types were downloaded from TCGA at FireBrowse (http://gdac.broadinstitute.org, 2016 January). Then, the fold change and adjusted *P*-value were calculated by the edgeR package (Robinson et al., 2010). We defined genes with an adjusted *P*-value less than 0.05 as the differential expression genes (DEGs).

Establishing the Ferroptosis Sensitive Index Model

The enrichment score (ES) of gene set that positively or negatively regulated ferroptosis was calculated using single sample gene set enrichment analysis (ssGSEA) in the R package ‘GSVA’ (Hanzelmann et al., 2013), and the normalized differences between the ES of the positive regulation gene set minus negative regulation gene set was defined as the ferroptosis sensitive index (FPI) to computationally dissect the ferroptosis levels/trends of the tissue samples.

Somatic Copy-number Alteration (SCNA) and Mutation Analysis

The heterozygosity and homozygosity of amplification and deletion were included to elevate the copy-number alteration of each gene, in which over five percent was regarded as high-frequency SCNA. Pearson's correlation between expression values and copy number segment values of each gene was calculated to evaluate the association between CNA and expression. The R package "DISCOVER" was employed to evaluate mutual exclusivity between FRGs and tumor suppressor genes or oncogenes across tumor samples in each cancer type (Canisius et al., 2016). The mutation and CNA events were integrated, while only homozygous amplification and deletion were included, and only protein-coding mutations were retained. For each cancer type, the genes were considered to be mutually exclusive if they had a q value of 0.05.

DNA methylation analysis

The R package "IlluminaHumanMethylation-450kanno.ilmn12.hg19" from Bioconductor was imported to annotate the methylation probe for the promoter of each gene. Differential methylation of each gene in tumor and normal samples was tested by the Wilcoxon signed rank test, and genes that were significantly hypomethylated or hypermethylated were identified using a *P*-value cutoff of 0.05. Pearson's correlation between the transcriptional expression of FRGs and the Beta value of the promoter DNA methylation were calculated and considered significant if the *P*-value < 0.05.

miRNA expression analysis

To investigate the mechanisms of dysregulation for ferroptosis regulator genes in cancer, we searched potential miRNAs which might regulate the FRGs based on miRNA-target intersections in starBase. The Spearman correlation between the expression of miRNA and FRGs was statistically evaluated (adjusted *P*-value < 0.1, rho < -0.1). Cytoscape software was used to visualize the high-frequency interaction networks among FRGs and miRNA (Shannon et al., 2003).

Multivariate Regression Analysis of Gene Expression

To assess which factors had significant effects on FRG expression, the expression of each FRG was modeled by linear regression as a function of the median miRNA expression, the median Beta value of promoter methylation, and the copy number of the genes.

Clinical Features Analysis

The R package “survival” was used to assess the prognosis potential of the FRGs and ferroptosis potential index among cancers. For survival analysis, the expression threshold was exhaustively tested and the one with most significant *P*-value was considered the best cut-off. To test the association between ferroptosis level and clinical features, the Pearson correlation was calculated between FPI and tumor stages, age, body mass index (BMI) and cigarette exposure per day, which were converted to numeric variables (“stage1” = 1, “stage2” = 2, etc.). Wilcoxon rank sum tests and Tukey’s tests were used to determine the impact on the ferroptosis potential index for other clinical characteristics including race, remission status, and alcohol.

Immune Features Analysis

To study the relationship of ferroptosis and immune microenvironments, we computed the Pearson correlation between FPI and immune parameters including immune cell fractions and 5 types of immunophenoscores, while the influence of different MSI statuses on IPS were also considered.

Identifying the FPI associated significant driver gene mutation

A total of 375 driver genes identified in previous pan-cancer research were included for analysis (Subramanian et al., 2005). To test whether a driver gene’s mutational status was significantly associated with ferroptosis among cancers, the rank-transformed FPI was modeled by linear regression as a function of the driver gene’s mutational status, ignoring the synonymous variant. To diminish the confounding effects, the rank-transformed count of the total nonsynonymous mutations and the tumor type, which were encoded as virtual variables, were included. To further characterize each cancer, the Benjamini-Hochberg method was used to correct the *P* values across 375 genes. Genes with an adjusted *P* value less than 0.05 for mutation status variable were significantly associated with FPI.

Gene Set Enrichment Analysis

To identify the pathways associated with ferroptosis, the samples of each tumor type were divided into two groups according to the FPI, consisting of the top 30% and bottom 30%. Then, the gene set

enrichment analysis (GSEA) was performed (Subramanian et al., 2005).

Correlation between Drug Sensitivity and FPI/FRG Expression

To test the correlation between small molecular drugs and FPI and FRGs, the Pearson correlation coefficients for FPI, the expression value of FRGs and the area under the dose-response curve (AUCs) values were calculated, the results with $|R| > 0.1$ and P-value < 0.05 were considered as significantly correlated. To further investigate the influence of drugs on ferroptosis, the significant association between the expression of clinically actionable genes and FRGs were finished across cancer cell lines, and the associations were filtered with PPIs. Then the drugs which target CAGs were selected according to DrugBank (Wishart et al., 2018) (https://www.drugbank.ca).

Results

Genetic alterations of ferroptosis regulator genes (FRGs) in cancers

In this study, we defined the 24 genes including *CDKN1A*, *HSPA5*, *TTC35*, *SLC7A11*, *NFE2L2*, *MT1G*, *HSPB1*, *GPX4*, *FANCD2*, *CISD1*, *FDFT1*, *SLC1A5*, *SAT1*, *TFRC*, *RPL8*, *NCOA4*, *LPCAT3*, *GLS2*, *DPP4*, *CS*, *CARS*, *ATP5G3*, *ALOX15*, and *ACSL4*, as ferroptosis regulator genes (FRGs), which were identified to play critical roles in positively or negatively regulating ferroptosis by previous studies (Stockwell et al., 2017). To determine the patterns for dysregulation of FRGs in cancer, we examined the genomic data, including genetic variation, somatic copy number alternation (SCNA), mRNA expression and DNA methylation data of tumor and normal tissues from 20 cancer types. The analysis of nonsynonymous mutations in 20 cancer types showed that the mutation frequencies for FRGs were generally low (Figure S1A), while 10 FRGs in UCEC had mutation frequencies greater than 5%. Furthermore, *NFE2L2*, which encoded NRF2 and played a master regulator role in anti-oxidant responses and has been shown to cause resistance to ferroptosis (Sun et al., 2016b), showed relatively high mutation frequencies in multiple cancers compared to other FRGs.

To further dissect the genetic aberrance of FRGs in cancer, the percentage of SCNA were examined and the results showed that in general SCNA occurred at high frequency (with over five percent of all samples) in most cancer types (Figure 1A), but all FRGs in THCA showed a low frequency of SCNA. It was observed that FRGs presented diverse SCNA profiles. For example, *TTC35*, *HSPB1*, *TFRC*, and *RPL8* were more prone to copy number gain than copy number loss in almost all tumors, but *SCL7A11* and *ALOX15* showed the opposite profile. Furthermore, we analyzed the co-occurrence of mutations and SCNA between FRGs and oncogenes/tumor suppressor genes (TSGs) previously identified by Bailey *et al.* (Bailey et al., 2018). The significance was calculated via the mutual exclusivity test by DISCOVER (Canisius et al., 2016), with a false discovery rate of 1%. A total of 28 oncogenes (Figure S1B) and 43 TSGs (Figure S1C) were found to be altered with FRGs in the mutually exclusive manner in certain cancer types, and *FDFT1*, *RPL8*, *TFRC*, and *NFE2L2* were frequently exclusive to oncogenes or TSGs (Figure S1B-C).

Aberrant expression of FRGs among cancer

Besides genetic alternations, we analyzed the gene expression of FRGs and found that all FRGs were differentially expressed in at least one cancer type. Several FRGs showed consistent expression

patterns in cross-cancer analysis. *SLC7A11*, *FANCD2*, *CARS*, *SLC1A5* and *RPL8* were significantly upregulated in 15, 18, 12, 15, 14 types of cancers, respectively, while *NCOA4* was downregulated in 15 cancers (Figure 1B). Additionally, several FRGs showed miscellaneous cancer type specificities which has not been well characterized previously. For example, *HSPA5* was upregulated in most cancer types including breast invasive carcinoma (BRCA) (fold change [FC] = 1.87, adjust *P*-value = 1.42×10^{-42}) and LUAD (FC = 1.90, adjust *P*-value = 2.45×10^{-29}) but was significantly downregulated in THCA (FC = 0.68, adjust *P*-value = 2.67×10^{-7}). *DPP4*, which appears to play a suppressor role in the development of cancer (Masur et al., 2006; Pro and Dang, 2004; Wesley et al., 2005), showed significant upregulation in KIRC, KIRP, ESCA, LUAD, GBM, and THCA, but was downregulated in CHOL, KICH, BRCA, LUSC, HNSC, and STAD. It is interesting that *DPP4* had opposed expression profiles in different subtypes of tumors in the lung and kidney. This demonstrated that FRGs might play different roles in different tumor environments.

Since SCNA in tumors plays a critical role in regulating gene expression, we evaluate the effects of SCNA on gene expression of FRGs. The Pearson correlation between gene expression and the copy number from masked copy number segment of TCGA was examined. The results showed that the expression of most FRGs was obviously correlated to SCNA in most tumors (Figure 1C). For example, the expression of citrate synthase (*CS*), which participates in oxidative metabolism, was significantly associated with SCNA in all cancer. This result indicates that the aberrance of copy number for FRGs is common in most cancers and can have an influence on gene expression.

Besides SCNA, the methylation of promoter can regulate gene expression, and aberrant DNA methylation of the promoter is associated with tumorigenesis (Shen and Laird, 2013). We observed that FRGs showed complex methylation patterns in the 20 cancer types (Figure 1D), only *CDKN1A* showed hypomethylation in 11 tumors consistently. For example, we observed that *DPP4* showed hypermethylation in seven cancer types and hypomethylation in four cancer types. Although there were differences in the pattern of methylation for FRGs, a negative relationship was observed between gene expression and DNA methylation overall (Figure 1E). This result demonstrated that promoter DNA methylation may regulate the expression of FRGs in tumors.

As we know, besides SCNA and DNA methylation, microRNAs (miRNAs) can regulate gene expression and become involved in cancer development (Macfarlane and Murphy, 2010). To determine which miRNAs are involved in regulating ferroptosis, we performed the analysis to reveal

the network of miRNA-FRGs. The starBase database was used to infer miRNA that potentially targeting FRGs, and those miRNAs that were significantly negatively correlated with gene expression were thought to be involved in the regulation of FRGs (Li et al., 2014). As the network showed (Figure 1F), all of the interactions between miRNA and FRGs occurred in more than six cancers. It is obvious that FRGs could be targeted by miRNA with high frequency, including *ACSL4*, *NCOA4*, and *CDKN1A* (Figure 1F). To further understand the aberrances of the miRNA-FRG network in tumors, we conducted differential expression analysis of miRNA and counted the expression aberrances of miRNA across cancer types (Figure S1D). It was found that several miRNAs had consistent expression trends in tumors, for example, has-miR-93 targeted *CDKN1A* and was downregulated in 11 cancers. On the other hand, miRNA had different expression trends in different tumors. For example, has-miR-375 targeted *ACSL4* in five tumors, but it showed upregulation in two cancers and downregulation in three cancers. This suggests that miRNA play a regulatory role in FRGs expression, and the aberrant expression of miRNA in the tumors could impact ferroptosis.

Since SCNA, DNA methylation, and miRNA all can regulate FRGs expression, but the contribution of a single factor on gene expression is not clear. The linear regression approach was applied to analyze the contribution of each factor and diminish the confounding effects. As the results showed (Figure S1E), SCNA was positively correlated with gene expression, while methylation and miRNA showed a negative correlation. We further observed multiple regulated patterns in FRGs, and the expression of several FRGs was only related to a single factor in several tumors, while others were related to multiple factors. For example, *ACSL4* expression was only associated with miRNA in seven cancers, including BLCA, CESC, KIRC, KIRP, LUAD, PRAD, and UCEC. SCNV played the only significant regulator role for *CARS* in CHOL, ESCA and colorectal cancer. However, the expression of *NCOA4* and *NFE2L2* was significantly regulated by three factors in seven and ten tumors, respectively. Thus, all the FRGs show diverse regulation patterns in different cancers. This suggests that the expression regulation patterns of all FRGs were tumor-specific.

Computational modeling the ferroptosis level among cancers

To further understand the role of ferroptosis in tumorigenesis and investigate the factors or biological processes associated with ferroptosis, the ferroptosis potential index (FPI) was modeled based on the enrichment score (ES) of positive regulatory components calculated by ssGSEA minus

that of negative regulatory components. Recently, Hassannia *et al.* identified that withaferin A (WA) was a ferroptosis-inducing agent in neuroblastoma (Hassannia et al., 2018). We calculated the FPI of neuroblastoma cells based on the gene expression dataset (GSE112384) in the study, and the results showed that WA increased the FPI remarkably (Figure S2A). Because the increases of the CHAC1, PTGS2 and ACSL4 proteins were associated with cells undergoing ferroptosis in some experiments (Stockwell et al., 2017), we examined the correlation between FPI and the expression of these three genes. As shown in Figure S2B-D, the expression of *ACSL4* was positively correlated to the FPI in the tumor samples for 11 cancers and normal samples for 3 cancers. However, *PTGS2* and *CHAC1* showed a weaker association with FPI than *ACSL4* across cancer types. Thus, the FPI could be used to represent the potential level of ferroptosis based on the transcriptome data.

Based on the computed marker of ferroptosis, we compared the differences of FPI between tumor and normal tissues with the cancer genome atlas (TCGA) data (Figure 2A). It is interesting that significant differences were found in most cancers such as lung cancer and gastrointestinal cancer, and higher FPI was observed in most tumors except for BRCA and UCEC. We also noticed that the FPI of normal tissues was higher than tumors in female cancers (BRCA, UCEC, and CESC). To further dissect the factors involved in these different FPI patterns, we examined the FPIs for different subtypes of BRCA. The results presented in Figure 2B showed that all the estrogen receptor (ER)-positive, progesterone receptor (PR) positive and HER2 positive patients had lower FPI than the negative patients, respectively. Furthermore, triple negative breast cancer (TNBC) samples showed higher ferroptosis level than non-TNBC samples (Figure 2C), which was consistent with previous study (Ma et al., 2016). Furthermore, among the different subtypes of kidney cancer, the FPI of tumor samples was higher than normal in KIRP and KIRC, while it was interesting that the FPI in tumor was lower in KICH (Figure 2A). In addition, to further dissect the variances of ferroptosis level in different histological types, the FPI in the esophagus squamous cell carcinoma (ESCC) and esophagus adenocarcinoma (ESAD) were compared. The results showed that FPI in ESCC was significantly higher than ESAD and normal samples, while ESAD was higher than normal but not significant (Figure 2D).

Besides the cancer types and subtypes, we analyzed the correlation between FPI and remarkable molecular features such as microsatellite instability (MSI) and driver gene mutations. FPI decreased obviously in tumors with high-level MSI compared to MSI stable tumors in COAD, READ,

ESCA and STAD (Figure 2E-F), and higher level MSI presented lower FPI in COAD and STAD. Furthermore, we analyzed the correlation between FPI and 375 driver genes that frequently mutated in tumors based on regression-based analysis. Through controlling for cancer type and tumor mutation burden (TMB), 81 genes were found to be correlated, and most of them were negatively associated with FPI (adj.p < 0.1, Figure S2E). Strikingly, somatic mutations, namely *NFE2L2*, *BRAF*, and *TP53*, were positively associated with FPI in pan-cancer associations, but a mutation in *TP53* was negatively correlated with ferroptosis in LUSC. This observation was verified by the comparison of FPI between the *TP53* mutant and wild type groups (Figure S3A), consistent with previous finding that showed that *TP53* played a dual role in regulating ferroptosis (Kang et al., 2018). The influences of *KRAS* mutations on FPI were also investigated, and the results showed that *KRAS* mutations might significantly inhibit ferroptosis in gastric tumors (Figure S3B), and it was interesting that *KRAS* mutations increased FPI in hepatocellular carcinoma (Figure S2E, S3B). Furthermore, the differential expression of FRGs between wild type and mutant tumors for *TP53* (Figure S3C) and (Figure S3D) were found to be ubiquitous in most cancers. Taken together, ferroptosis was closely related with remarkable molecular features in cancer.

Association between ferroptosis and pathways in cancer

To further elucidate the related cellular signaling of ferroptosis in cancer, gene set enrichment analysis (GSEA) was performed based on the transcriptome of two tumor groups with the top and bottom thirty percent of FPI. It was observed that metabolism related pathways in KEGG usually enriched in the tumor with lower FPI, and pathways frequently enriched (>6 cancers) were presented in Figure 3A. For example, terpenoid backbone biosynthesis and steroid biosynthesis were enriched in the low FPI group in 19 and 16 cancers, respectively (Figure 3A). Peroxisome, biosynthesis of unsaturated fatty acids, etc. were also significantly correlated with lower FPI in many cancer types (Figure 3A). Furthermore, the relations between cancer hallmarks and FPI were also analyzed, and the results showed that 15 hallmarks were frequently significantly correlated with FPI (Figure 3B). For example, KRAS signaling, epithelial-mesenchymal transition, IL6 JAK STAT3 signaling, WNT Beta-catenin signaling were enriched in the high FPI group, which indicated that ferroptosis was positively related with these oncogenic pathways (Figure 3B). Also, metabolism related hallmarks were observed to be negatively related with ferroptosis, which was consistent with the pathway analysis

(Figure 3B).

Since previous studies showed that ferroptosis was related to the immune response process (Matsushita et al., 2015), here we investigated the association between ferroptosis and immune microenvironment in tumor. The results showed that in several cancers FPI was weakly negatively correlated with immunophenoscore (IPS) (Figure 3C), which could predict the response of the immune checkpoint blockade in melanoma tissue (Charoentong et al., 2017). Furthermore, FPI was positively correlated with MHC and EC (effector cells) but negatively correlated with immunomodulators (CP) and suppressor cells (SC) in most cancers (Figure 3C). The associations between ferroptosis and immune cell types were further evaluated in detail, and the results showed that FPI showed a positive correlation with macrophages in most cancers and a negative association with the activated dendritic cells, activated mast cells, plasma cells and T follicular helper cells (Figure S4A). Thus, there might be close connections between ferroptosis and immune microenvironment, and more investigations were needed to reveal the details.

Clinical relevance of ferroptosis and its regulators

Since ferroptosis was involved in cellular metabolism and immune microenvironment, it was proposed that ferroptosis and its regulators should be correlated with cancer survival and other clinical characteristics. Based on the clinical data from TCGA, the survival analysis was performed and the results were miscellaneous since it is found that lower FPI predicted a better survival in most cancers including CHOL, GBM, LUAD, LIHC, KIRC, KIRP, and UCEC, but worse in BLCA, KICH, and PRAD (Figure 4). We further dissected the factors that might impact FPI including tumor stage, cigarette smoking, age, and gender. As the results shown (Figure S4B), tumor stages were positively correlated with FPI in LIHC, KIRC, KIRP, and COAD. Only the FPI in LUAD tumors showed a positive correlation with cigarettes smoked per day. Age was positively associated with FPI in GBM and PRAD but showed a negative correlation in seven cancers such as BRCA and LIHC. It was observed that FPI was remarkably different between genders (Figure S4C), FPI for females was higher than males in KIRC, LIHC, LUAD, and LUSC, but lower in STAD. Significant difference in FPI were also observed among different races in BLCA, BRCA and ESCA. Furthermore, we found that the FPI was correlated cancer metastasis, recurrence, primary and follow outcome in several cancers. In addition, we evaluated the association between FPI and other clinical characteristics including alcohol abuse, fatty

liver, hemochromatosis, and viral infection (HPV, HBC, HCV, and EBV), while no significant results were observed. These results indicate that ferroptosis might play critical roles in cancer survival, and clinical factors have tumor-specific effects on FPI.

To further dissect the clinical relevance of ferroptosis in cancer, the roles of FRGs in cancer survival were analyzed. The survival analysis showed that all FRGs were associated with overall survival in at least six cancer types (Figure S4D), but the correlations were miscellaneous since most FRGs could both serve as protective and risk factors in different cancers. For example, patients with higher expression of *FANCD2* showed better survival in kidney cancer, LIHC, and LUAD, but showed worse survival in READ, STAD, and BLCA (Figure S4D). However, high expression of *CARS* and *MT1G* showed consistent better survival among cancers including KICH, UCEC, PAAD, and HNSC (Figure S4D). Thus, the functional roles of FRGs in cancer survival should be further explored.

To study the potential clinical implications of ferroptosis, we examined the association between the gene expression of FRGs and 156 clinically actionable genes (CAGs), including 136 genes for targeted therapy and 20 genes for immunotherapy across cancer types (Mak et al., 2016; Van Allen et al., 2014). As a FRG, *CDKN1A* regulates iron uptake and *GPX4* abundance (Stockwell et al., 2017), was also a CAG targeted by anti-cancer drugs, such as the inorganic compound arsenic trioxide (Hassani et al., 2018). We found that all 136 CAGs and FRGs had significant co-expression relationships that were significantly different across cancers as shown in Figure S5A. The number of CAGs correlated with FRGs ranged from 45 in LUSC to 132 in THCA (Figure S5B). The FRG-CAG correlation pairs ranged from 69 in LUSC to 581 in THCA (Figure S5C). For example, *GPX4* showed a significantly positive correlation with *TNFRSF4*, which are targeted genes for immunotherapy. *ACSL4* is significantly correlated with 17 of 20 genes targeted for immunotherapy, suggesting that *ACSL4* has a potential effect on cancer immunotherapy (Figure S6A). Furthermore, besides co-expression, the protein-protein interactions among drug-targeted CAGs and FRGs were analyzed, and the results presented in Figure S6B showed that the interactions were intense. For example, *EGFR*, the target of lapatinib, gefitinib and afatinib, was positively associated with FRGs including *DPP4* (R = 0.15) and *HSPA5* (R = 0.31), and negatively correlated with *FDFT1* (R = -0.24), while there were protein-protein interactions between EGFR and DPP4, HSPA5, FDFT1 (Figure S6B). These results indicated that ferroptosis might be involved in drug effects by interacting with targeted clinically actionable genes. Taken together, our studies suggest that clinically actionable genes are

closely related to FRGs, which highlights the significance of ferroptosis in cancer treatment, including both immunotherapy and targeted therapy.

To further understand the correlation between ferroptosis and drug sensitivity, the area under the percent viability curve (AUC) approach was employed to evaluate drug sensitivity (Basu et al., 2013) and calculate the correlation between FPI and drug sensitivity across cancer cell lines. Drugs associated with FPI were also tested for their correlation with FRG expression. We identified that FPI was significantly associated with 64 drugs, including 12 that were negatively correlated and 52 that were positively correlated (Figure S6C). To explore the effects of each FRG on drug sensitivity, we also analyzed the associations between the drug sensitivity of 64 cancer drugs and the expression of FRGs, and found 521 significantly correlated pairs (Figure S6C). Among them, the expression of *FANCD2* was correlated with AUC in 57 drugs, while no significant relation was observed for *CISD1* (Figure S6C). Furthermore, the associations classified the FRGs into two group, one group included genes such as *FANCD2* and *ATP5G3*, which were positively associated AUC of docetaxel, trametinib, etc. (Figure S6C). The other group included *SLC7A11*, *SAT1*, *HSPA5*, etc., which were negatively associated (Figure S6C). Taken together, these results suggest that ferroptosis was correlated to the sensitivity of multiple drugs.

Discussion

Ferroptosis is driven by damage of cell membranes caused by the loss of activity of GPX4 (Feng and Stockwell, 2018). Researchers identified that ferroptosis is closely related to tumorigenesis and plays an important role in cancer treatment (Shen et al., 2018; Yang et al., 2014). However, there is a lack of systematic studies on ferroptosis and its regulator genes across cancer types. In this study, we employed multi-omics data and clinical data across 20 cancer types from TCGA and revealed the global alterations of ferroptosis regulator genes at genetic, epigenetic and transcriptional levels. We also used ssGSEA to process expression data to establish FPI to characterize ferroptosis and addressed which genetic and nongenetic factors (including drug and patient phenotype) were related with FPI. Different molecular types affected the ferroptosis in breast cancer and gastrointestinal cancer, which meant that the responses of different molecular subtypes to treatment may be related to ferroptosis. On the other hand, molecular subtypes need to be considered when ferroptosis is applied as a therapeutic strategy.

The mechanism by which ferroptosis regulates tumor cell growth and proliferation is still unclear, but the relationship we observed between FPI and hallmarks of cancer could improve the understanding the role of ferroptosis. The results of GSEA demonstrate that the level of ferroptosis is closely related to tumor-associated hallmarks in most cancers. Ferroptosis genes can play both oncogene and tumor suppressor roles in cancer (Kang et al., 2018), and the FPI acts as a protective or risk factor across cancer types. We also found that several common clinical factors affected ferroptosis, such as cigarette smoking, BMI and tumor stage. As we know, cigarette smoking is a risk factor for esophageal cancer, lung cancer and kidney cancer (Chow et al., 2010; Mao et al., 2011; Pesch et al., 2012). In our results, the number of cigarettes per day was positively correlated with the ferroptosis potential index in LUAD, which may be because the cigarette-induced oxidation reaction promoted lipid peroxidation (Barreiro et al., 2010; Guan et al., 2013; Louhelainen et al., 2008). FPI also varied between the sexes in several cancers, including LIHC, LUAD, LUSC, and STAD and between different races in BRCA, BLCA, and ESCA, which implied that gender and race need to be considered when using ferroptosis as a treatment strategy. We also noticed that better clinical outcomes or status in several cancer types also have higher FPI, which further confirmed the dual role of ferroptosis. Thus, a different strategy of regulating the ferroptosis of tumor cells may benefit patients and improve prognosis.

Furthermore, we showed that FRGs were co-expressed with most clinically actionable genes and interacted with genes modulated by drugs, suggesting that studying ferroptosis may improve the strategy for cancer therapy. We observed that the co-expression of FRGs and CAGs could be roughly divided into two groups, and drugs that target clinical genes could have complex regulatory effects on ferroptosis. Most strikingly, the expression of *GPX4* was associated with *MAP2K2* in 20 cancers, which indicated that *MAP2K2* may play an important role in ferroptosis. To further explore drug sensitivity and ferroptosis, we observed that the FPI could characterize the sensitivity of many drugs, the AUC of many drugs was inversely associated with the FPI in cancer cell lines, which implied that regulating the ferroptosis of tumor cells may improve the therapeutic effect of cancer treatments. However, opposite effects were observed regarding the effects of drugs on ferroptosis. Thus, further detailed studies should be carried out to determine the functions and mechanisms of ferroptosis in different cancers.

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Figures

Figure 1 – Differential expression of the ferroptosis regulator genes (FRGs). The somatic copy number alternations (A) and differential expression (B) of FRGs among cancers. (C) The correlation between and somatic copy number alternations and expression of FRGs. The differential methylation (D) and its correlation (E) with expression of FRGs. (F) The miRNA-mRNA network for FRGs.

Figure 2 – The ferroptosis potential index (FPI) and its relations with histological types and molecular subtypes among cancers. (A) The different FPI between tumor and adjacent normal tissues among cancers. (B-C) The FPI for different molecular subtypes of breast cancer. (D) The different FPI among different histological types of esophageal carcinoma. The FSI for different MSI status of overall (E) and detailed (E) digestive system neoplasm.

Figure 3 – Relations between the ferroptosis and signaling pathways and immunophenotypes. Enrichment analysis for metabolism pathway (A) and cancer signaling (B) between high- and low-FPI tumor tissues. NES is the normalized enrichment score in the GSEA algorithm. (C) Correlations between immunophenotypes and the FPI, and dots indicate statistically significant results (*P* value<0.05).

Figure 4 – Kaplan–Meier analysis of overall survival according to the FPI among cancers.

Figure S1 – Molecular alternations of FRGs. The mutation frequencies (A), mutual exclusivity between FRGs and oncogenes (B) /tumor suppressor genes (C) for FRGs among cancers. (D) The differential expressed FRGs-related miRNAs. The correlation between FRGs expression and somatic copy number alternation, DNA methylation and miRNAs.

Figure S2 – (A) The FPI between withaferin A (WA) induced ferroptosis and the control. The correlation between FPI and the expression of well-known FRGs including PTGS2 (B), ACSL4 (C), CHAC1 (D) and driver genes (E).

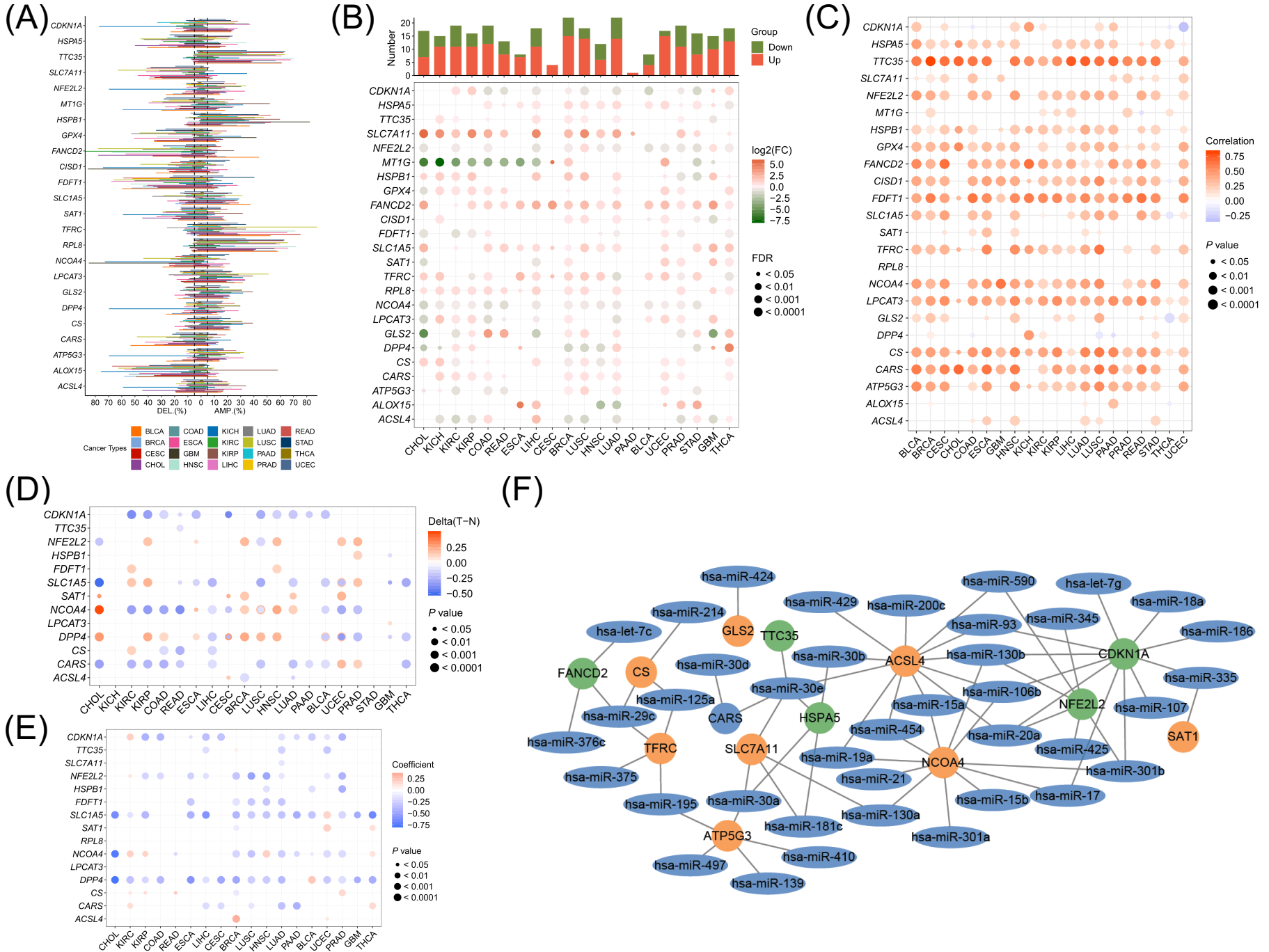
Figure S3 – The FPI between mutant and wild type tumors for *TP53* (A) and *KRAS* (B). The

differential expression of FRGs between mutant and wild type tumors for *TP53* (C) and *KRAS* (D).

Figure S4 – The correlation between ferroptosis and immune cells (A) and clinical characteristics (B-C). The overall prognosis abilities of FRGs (D).

Figure S5 – (A) The co-expression between FRGs and 136 genes for targeted therapy. (B) The number of CAGs correlated with FRGs. (C) The number of FRG-CAG correlation pairs.

Figure S6 – (A) The co-expression between FRGs and 20 genes for immunotherapy. (B) The interactions between drug, drug target and FRGs. (C) The correlation between expression of FRGs and the area under the dose-response curve (AUCs) for drugs.


(A)
CDKN1A
HSPA5
TTC35
SLC7A11
NFE2L2
MT1G
HSPB1
GPX4
FANCD2
CISD1
FDFT1
SLC1A5
SAT1
TFRC
RPL8
NCOA4
LPCAT3
GLS2
DPP4
CS
CARS
ATP5G3
ALOX15
ACSL4
80 70 60 50 40 30 20 10 0 10 20 30 40 50 60 70 80
DEL.(%)
AMP.(%)
Cancer Types
BLCA BRCA CESC CHOL COAD ESCA GBM HNSC KICH KIRC KIRP LIHC LUAD LUSC PAAD PRAD READ STAD THCA UCEC
(B)
Number
Group
Down
Up
log2(FC)
5.0
2.5
0.0
−2.5
−5.0
−7.5
FDR
< 0.05
< 0.01
< 0.001
< 0.0001
CHOL KICH KIRC KIRP COAD READ ESCA LIHC CESC BRCA LUSC HNSC LUAD PAAD BLCA UCEC PRAD STAD GBM THCA
(C)
Correlation
0.75
0.50
0.25
0.00
−0.25
P value
< 0.05
< 0.01
< 0.001
< 0.0001
(D)
Delta(T−N)
0.25
0.00
−0.25
−0.50
P value
< 0.05
< 0.01
< 0.001
< 0.0001
(E)
Coefficient
0.25
0.00
−0.25
−0.50
−0.75
P value
< 0.05
< 0.01
< 0.001
< 0.0001
(F)
hsa-miR-424
hsa-miR-590
hsa-let-7g
hsa-miR-18a
hsa-miR-429
hsa-miR-200c
hsa-miR-214
hsa-miR-93
hsa-miR-345
hsa-let-7c
GLS2
TTC35
hsa-miR-186
hsa-miR-30b
ACSL4
hsa-miR-130b
CDKN1A
FANCD2
CS
hsa-miR-30d
hsa-miR-335
hsa-miR-30e
hsa-miR-106b
hsa-miR-125a
hsa-miR-15a
NFE2L2
hsa-miR-107
hsa-miR-29c
CARS
HSPA5
hsa-miR-454
hsa-miR-20a
SAT1
hsa-miR-376c
TFRC
SLC7A11
hsa-miR-19a
hsa-miR-425
NCOA4
hsa-miR-375
hsa-miR-21
hsa-miR-301b
hsa-miR-195
hsa-miR-30a
hsa-miR-15b
hsa-miR-17
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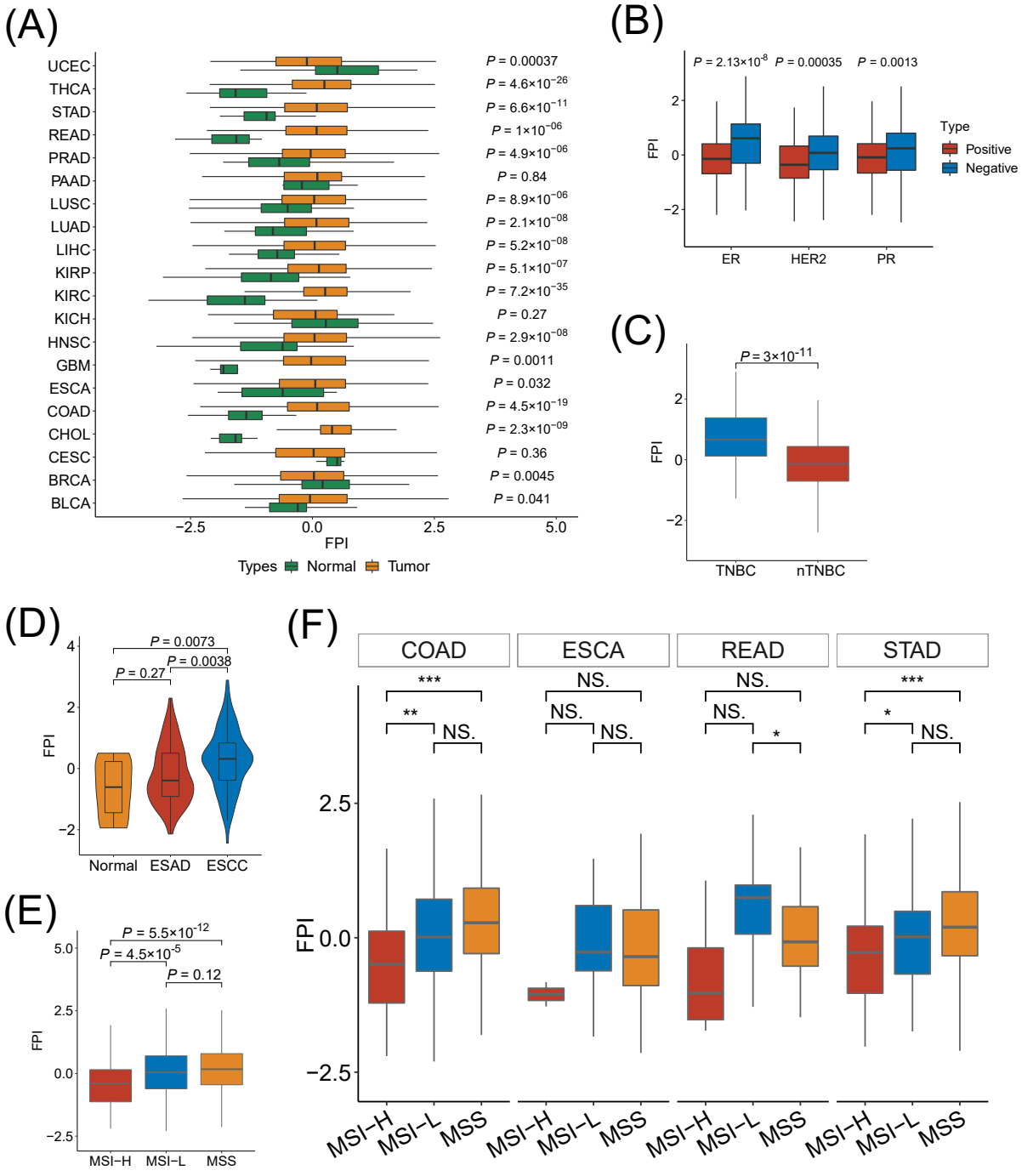
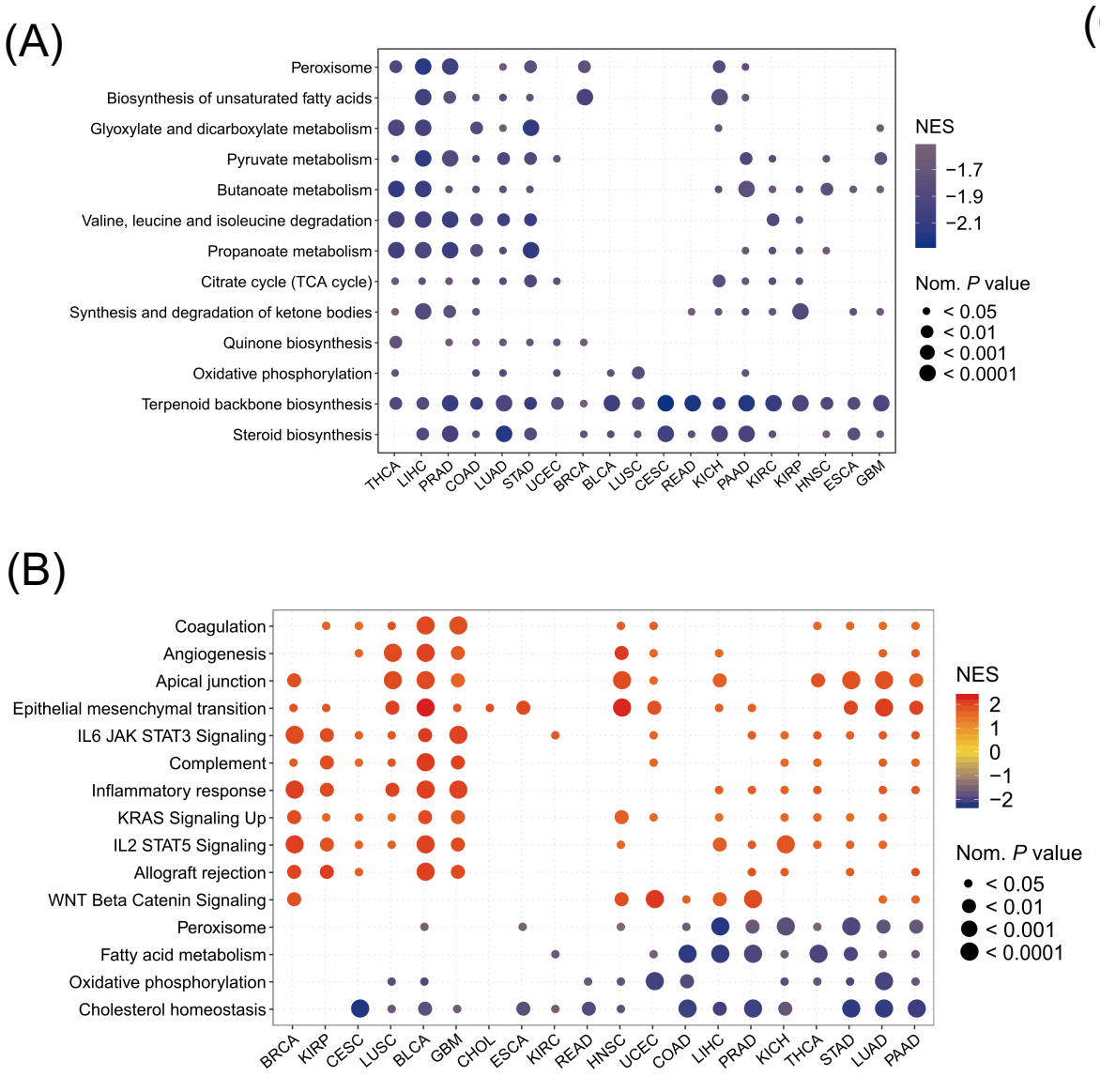
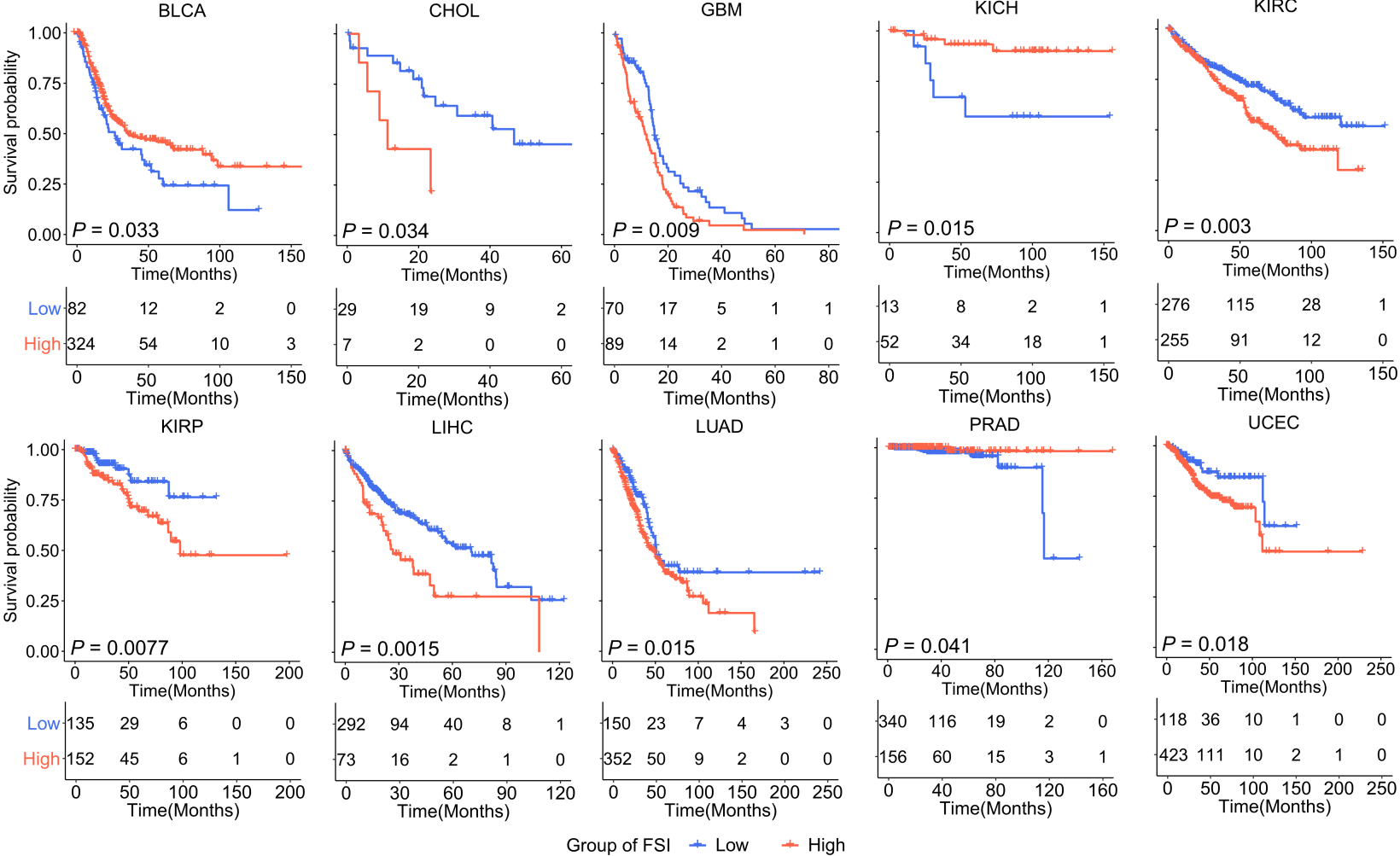

(A)
UCEC
THCA
STAD
READ
PRAD
PAAD
LUSC
LUAD
LIHC
KIRP
KIRC
KICH
HNSC
GBM
ESCA
COAD
CHOL
CESC
BRCA
BLCA
P = 0.00037
P = 4.6×10−26
P = 6.6×10−11
P = 1×10−06
P = 4.9×10−06
P = 0.84
P = 8.9×10−06
P = 2.1×10−08
P = 5.2×10−08
P = 5.1×10−07
P = 7.2×10−35
P = 0.27
P = 2.9×10−08
P = 0.0011
P = 0.032
P = 4.5×10−19
P = 2.3×10−09
P = 0.36
P = 0.0045
P = 0.041
−2.5
0.0
2.5
5.0
FPI
Types
Normal
Tumor
(B)
P = 2.13×10−8
P = 0.00035
P = 0.0013
FPI
2
0
−2
ER
HER2
PR
Type
Positive
Negative
(C)
P = 3×10−11
FPI
2
0
−2
TNBC
nTNBC
(D)
P = 0.0073
P = 0.0038
P = 0.27
FPI
4
2
0
−2
Normal
ESAD
ESCC
(E)
P = 5.5×10−12
P = 4.5×10−5
P = 0.12
FPI
5.0
2.5
0.0
−2.5
MSI−H
MSI−L
MSS
(F)
COAD
ESCA
READ
STAD
***
**
NS.
NS.
NS.
NS.
NS.
NS.
*
***
*
NS.
FPI
2.5
0.0
−2.5
MSI−H
MSI−L
MSS
MSI−H
MSI−L
MSS
MSI−H
MSI−L
MSS
MSI−H
MSI−L
MSS


(A)
Peroxisome
Biosynthesis of unsaturated fatty acids
Glyoxylate and dicarboxylate metabolism
Pyruvate metabolism
Butanoate metabolism
Valine, leucine and isoleucine degradation
Propanoate metabolism
Citrate cycle (TCA cycle)
Synthesis and degradation of ketone bodies
Quinone biosynthesis
Oxidative phosphorylation
Terpenoid backbone biosynthesis
Steroid biosynthesis
THCA LIHC PRAD COAD LUAD STAD UCEC BRCA BLCA LUSC CESC READ KICH PAAD KIRC KIRP HNSC ESCA GBM
NES
−1.7
−1.9
−2.1
Nom. P value
< 0.05
< 0.01
< 0.001
< 0.0001
(B)
Coagulation
Angiogenesis
Apical junction
Epithelial mesenchymal transition
IL6 JAK STAT3 Signaling
Complement
Inflammatory response
KRAS Signaling Up
IL2 STAT5 Signaling
Allograft rejection
WNT Beta Catenin Signaling
Peroxisome
Fatty acid metabolism
Oxidative phosphorylation
Cholesterol homeostasis
BRCA KIRP CESC LUSC BLCA GBM CHOL ESCA KIRC READ HNSC UCEC COAD LIHC PRAD KICH THCA STAD LUAD PAAD
NES
2
1
0
−1
−2
Nom. P value
< 0.05
< 0.01
< 0.001
< 0.0001
(C)
PRAD
LIHC
KICH
LUSC
LUAD
STAD
KIRC
BLCA
THCA
BRCA
KIRP
PAAD
GBM
UCEC
READ
CESC
HNSC
COAD
MHC EC CP SC IPS
Coefficient
−0.4 −0.2 0.0 0.2


BLCA
CHOL
GBM
KICH
KIRC
KIRP
LIHC
LUAD
PRAD
UCEC
Survival probability
Time(Months)
P = 0.033
P = 0.034
P = 0.009
P = 0.015
P = 0.003
P = 0.0077
P = 0.0015
P = 0.015
P = 0.041
P = 0.018
Low
High
Group of FSI